

Electronic Supplementary Information

Clinical blood sampling for oxylipin analysis

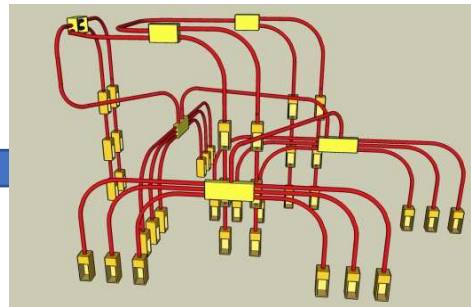
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Effect of storage and pneumatic tube transport of blood on free
and total oxylipin profile in human plasma and serum

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Sebastian Schott, Ralf Lichtinghagen, Faikah Gueler,
Nils Helge Schebb



Transfer of samples
at the ward to the
pneumatic tube
system



Pneumatic tube
system transport



„24/7“ sample
processing at
clinical chemistry
laboratory

Fig. S1: Overview of the pneumatic tube system transport at the Hannover Medical School. Samples collected on the ward or the intensive care unit are packed in tubes and transferred via the pneumatic tube system transport which covers in total about 50 km pipe distance to the clinical chemistry laboratory where continuous (24/7) sample processing is carried out.

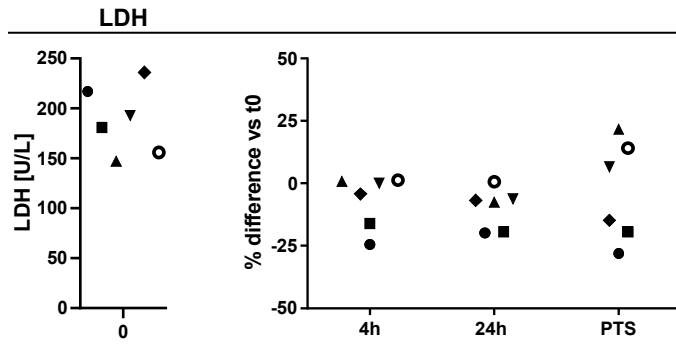


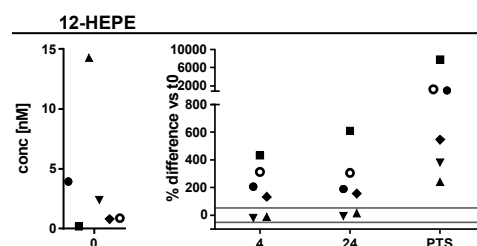
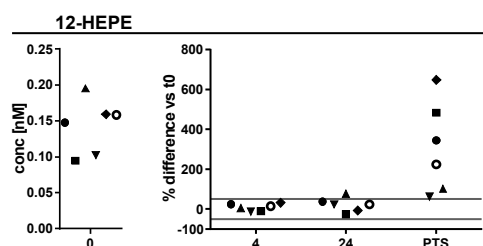
Fig. S2: Lactate dehydrogenase (LDH) in plasma. Shown are individual levels (n=6) of LDH at *t0* (immediate processing) and % difference vs. *t0* after storage for 4 h at 4 °C and 24 h at 4 °C or after pneumatic tube system transport before centrifugation to generate plasma. The different symbols represent samples from different individual human subjects.

LDH was analyzed by an Olympus analyzer (AU400) in an automated fashion according to the manufacturer's instruction.

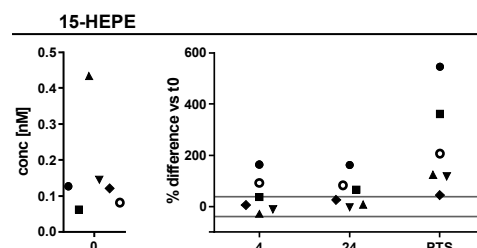
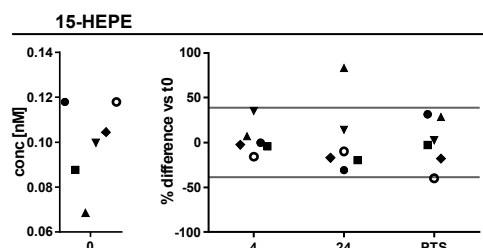
(A) Free oxylipins in plasma

(B) Free oxylipins in serum

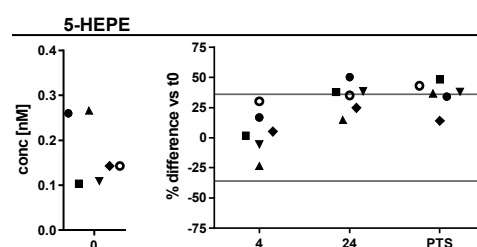
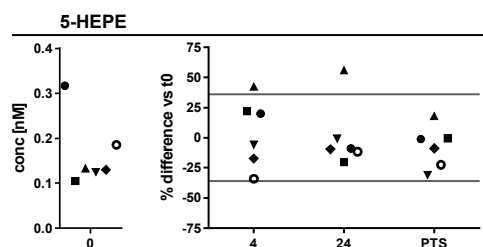
12-LOX



15-LOX/COX



5-LOX



CYP

17(18)-EpETE

< LLOQ

17(18)-EpETE

< LLOQ

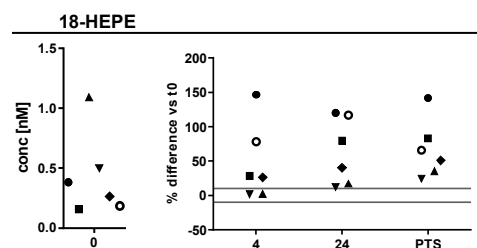
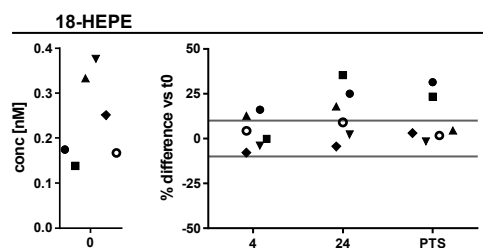
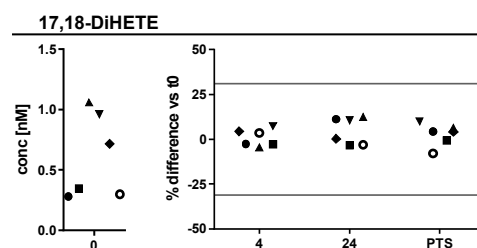
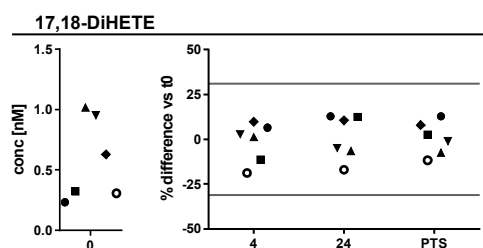
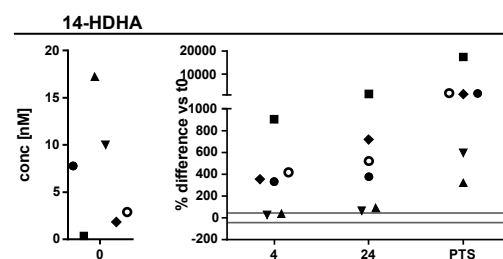
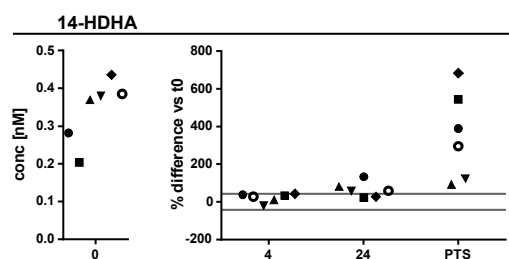


Fig. S3: EPA derived free oxylipins in (A) plasma and (B) serum. Shown are individual concentrations (n=6) of selected oxylipins from major formation pathways at *t*₀ (immediate processing) and % difference vs. *t*₀ after storage for 4 h at 4 °C and 24 h at 4 °C or after pneumatic tube system transport before centrifugation to generate plasma or serum. The different symbols represent samples from different individual human subjects. The grey lines indicate acceptable change limits calculated based on relative SD of quality control plasma (summarized in Tab.1).

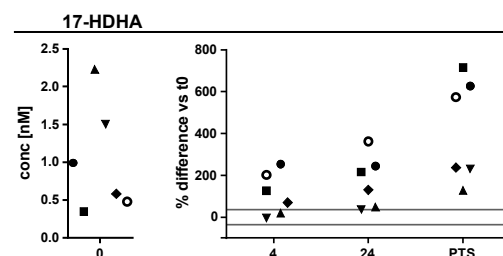
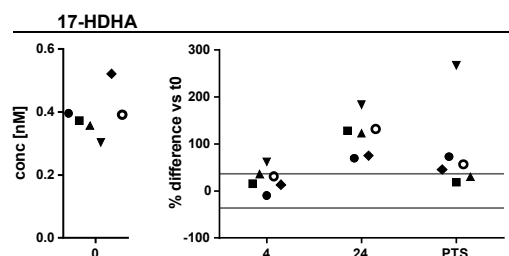
(A) Free oxylipins in plasma

(B) Free oxylipins in serum

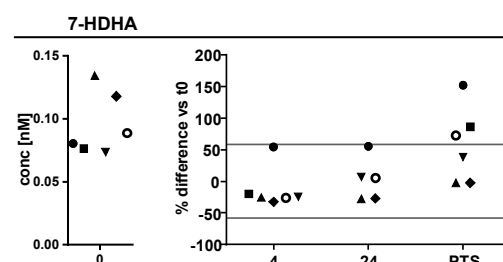
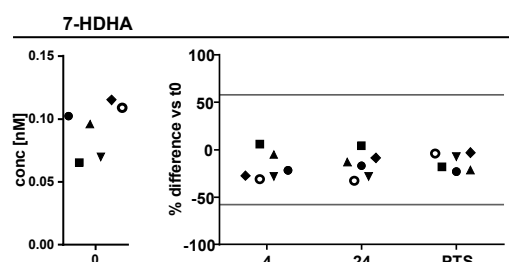
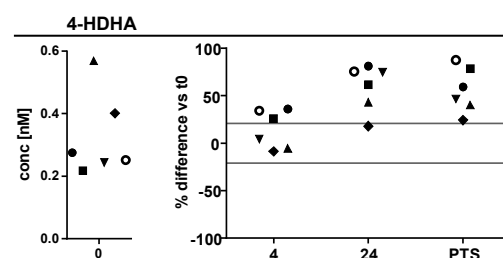
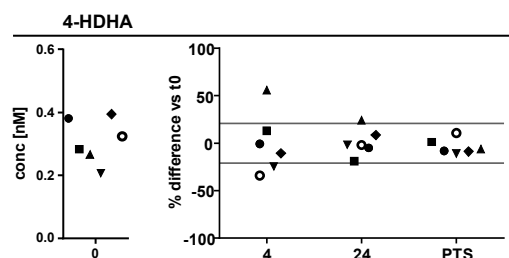
12-LOX



15-LOX



5-LOX



CYP

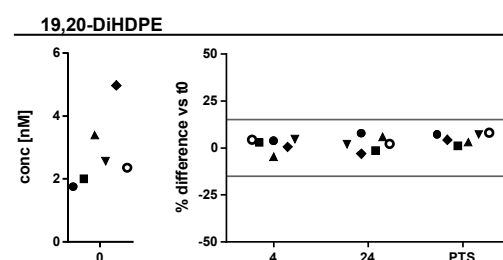
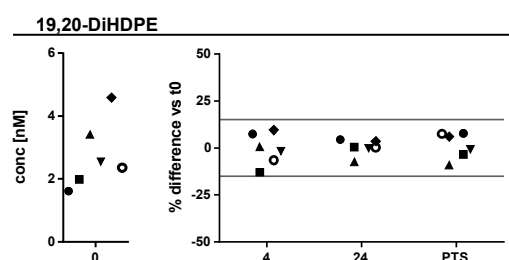
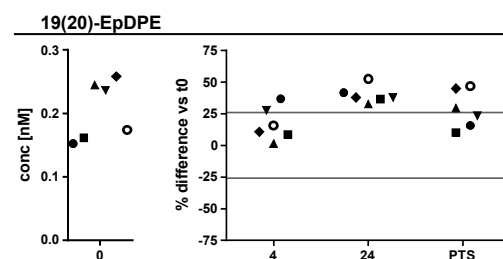
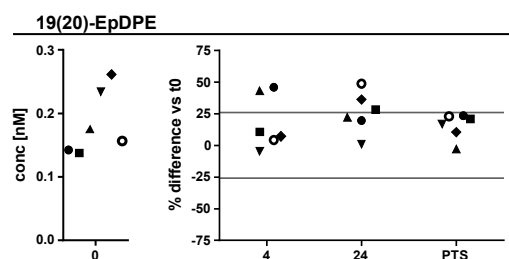


Fig. S4: DHA derived free oxylipins in (A) plasma and (B) serum. Shown are individual concentrations ($n=6$) of selected oxylipins from major formation pathways at t_0 (immediate processing) and % difference vs. t_0 after storage for 4 h at 4 °C and 24 h at 4 °C or after pneumatic tube system transport before centrifugation to generate plasma or serum. The different symbols represent samples from different individual human subjects. The grey lines indicate acceptable change limits calculated based on relative SD of quality control plasma (summarized in Tab.1).

(A) Total oxylipins in plasma

(B) Total oxylipins in serum

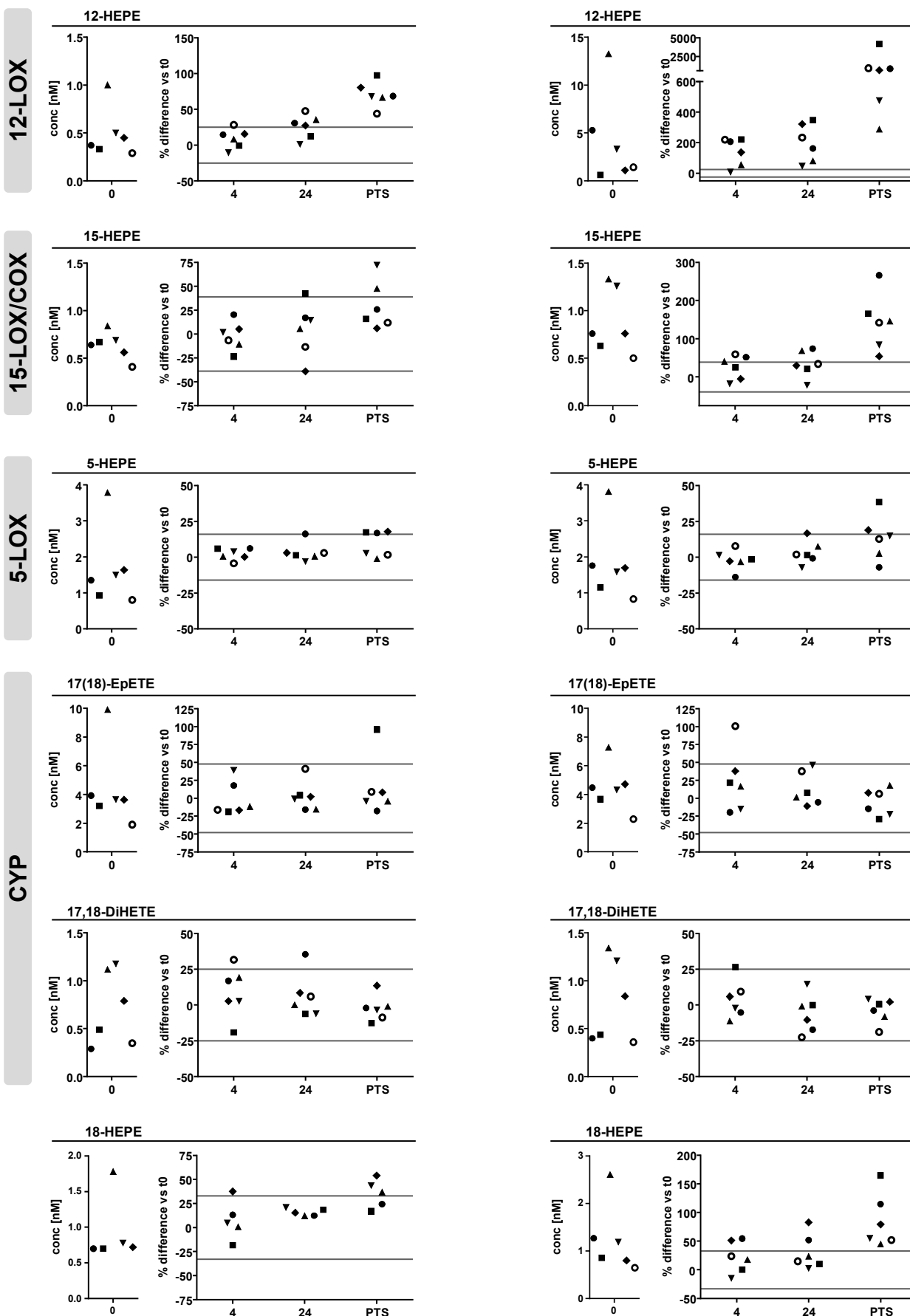
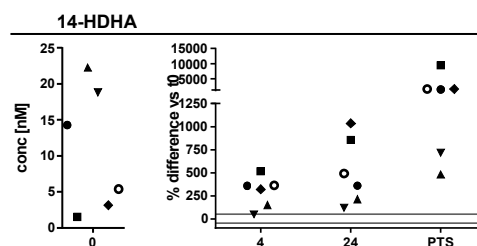
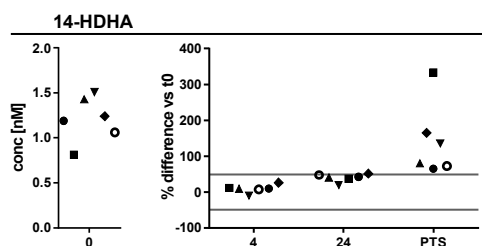


Fig. S5: EPA derived total (i.e. sum of free and esterified) oxylipins in (A) plasma and (B) serum. Shown are individual concentrations ($n=6$) of selected oxylipins from major formation pathways at t_0 (immediate processing) and % difference vs. t_0 after storage for 4 h at 4 °C and 24 h at 4 °C or after pneumatic tube system transport before centrifugation to generate plasma or serum. The different symbols represent samples from different individual human subjects. The grey lines indicate acceptable change limits calculated based on relative SD of quality control plasma (summarized in Tab.1).

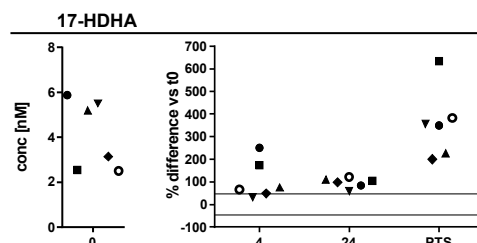
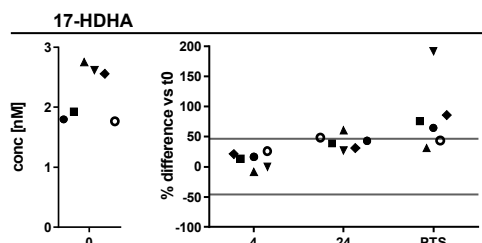
(A) Total oxylipins in plasma

(B) Total oxylipins in serum

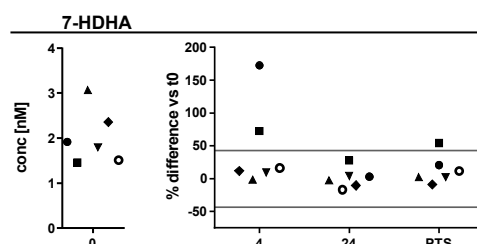
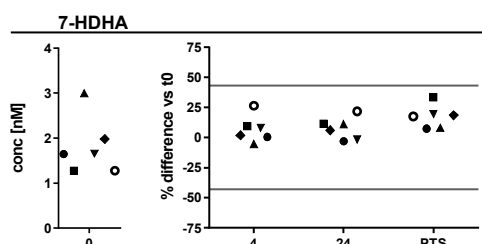
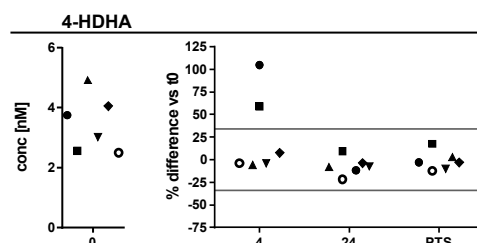
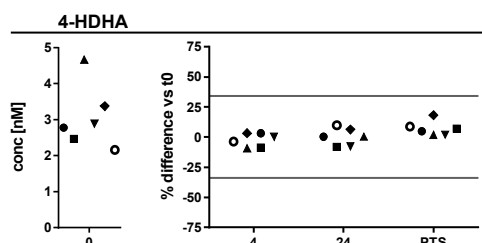
12-LOX



15-LOX



5-LOX



CYP

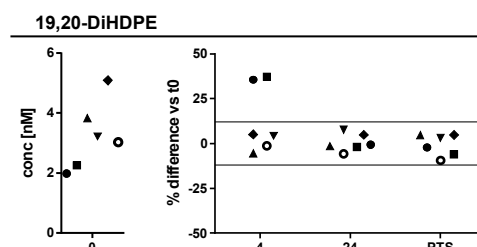
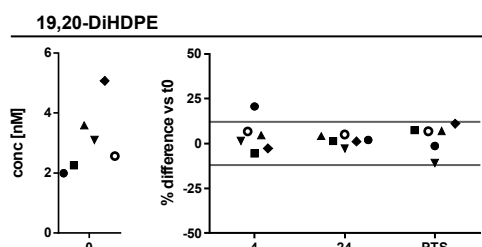
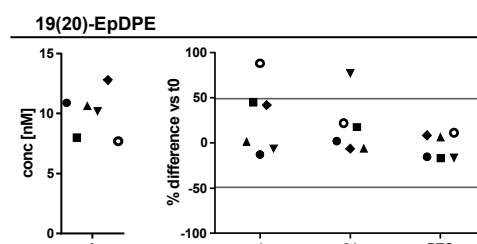
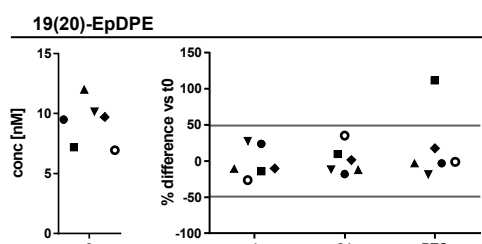
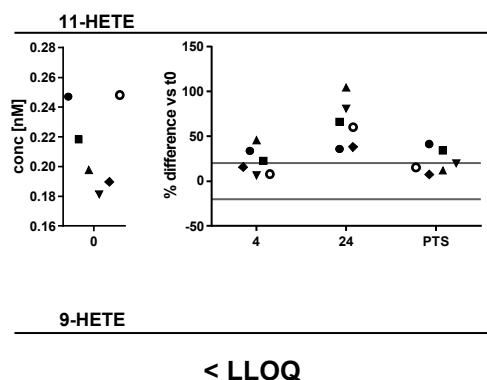
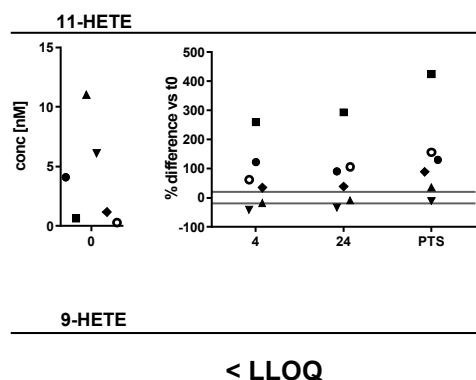


Fig. S6: DHA derived total oxylipins in **(A)** plasma and **(B)** serum. Shown are individual concentrations ($n=6$) of selected oxylipins from major formation pathways at t_0 (immediate processing) and % difference vs. t_0 after storage for 4 h at 4 °C and 24 h at 4 °C or after pneumatic tube system transport before centrifugation to generate plasma or serum. The different symbols represent samples from different individual human subjects. The grey lines indicate acceptable change limits calculated based on relative SD of quality control plasma (summarized in Tab.1).

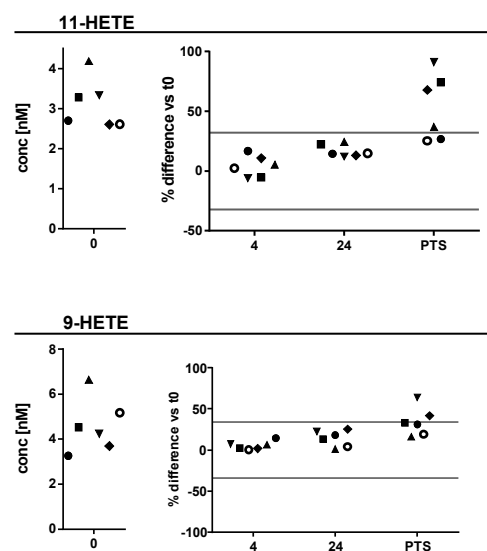
(A) Free oxylipins in plasma



(B) Free oxylipins in serum



(C) Total oxylipins in plasma



(D) Total oxylipins in serum

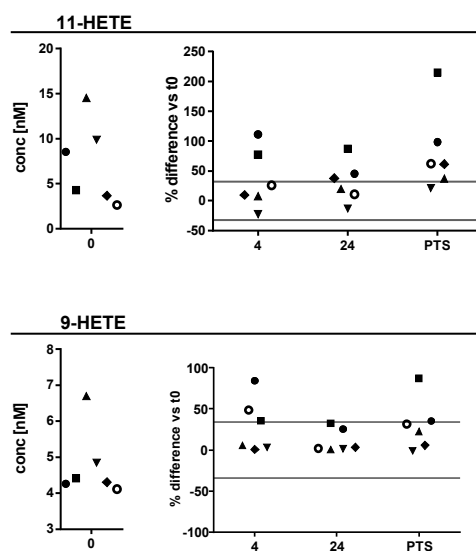


Fig. S7: ARA derived 11-HETE and 9-HETE in **(A+C)** plasma and **(B+D)** serum. Shown are individual concentrations ($n=6$) of **(A+B)** free or **(C+D)** total (comprising free and esterified) mediators at t_0 (immediate processing) and % difference vs. t_0 after storage for 4 h at 4 °C and 24 h at 4 °C or after pneumatic tube system transport before centrifugation to generate plasma or serum. The different symbols represent samples from different individual human subjects. The grey lines indicate acceptable change limits calculated based on relative SD of quality control plasma (summarized in Tab.1).

Tab. S1: Median concentrations with interquartil range (25% percentil, 75% percentil) [nM] of representative free oxylipins in plasma and serum following direct processing, after storage for 4 h or 24 h at 4 °C, or after pneumatic tube system transport (PTS) prior to centrifugation (n =6).

	plasma median (25% percentil, 75% percentil) [nM]				serum median (25% percentil, 75% percentil) [nM]			
	direct	4 h	24 h	PTS	direct	4 h	24 h	PTS
PGE ₂	0.010 (0.010, 0.039)	0.015 (0.014, 0.036)	0.022 (0.017, 0.038)	0.040 (0.033, 0.068)	1.3 (0.27, 1.8)	0.98 (0.50, 2.0)	0.98 (0.43, 1.6)	1.2 (0.78, 2.4)
TxB ₂	0.13 (0.078, 0.24)	0.21 (0.11, 0.32)	0.29 (0.18, 0.39)	0.56 (0.39, 0.78)	53 (9.1, 142)	69 (21, 168)	62 (21, 145)	73 (32, 167)
12-HHT	0.18 (0.15, 0.29)	0.24 (0.18, 0.35)	0.40 (0.35, 0.60)	0.65 (0.46, 0.79)	50 (8.6, 104)	53 (21, 133)	59 (20, 142)	67 (29, 158)
12-HETE	2.3 (1.1, 3.5)	2.5 (2.2, 4.7)	5.4 (2.1, 7.0)	5.8 (5.6, 7.8)	97 (22, 209)	114 (67, 273)	116 (74, 259)	312 (224, 538)
12-HEPE	0.15 (0.10, 0.17)	0.18 (0.087, 0.20)	0.17 (0.11, 0.24)	0.53 (0.34, 0.79)	1.6 (0.66, 6.5)	2.8 (1.7, 12)	2.9 (2.1, 13)	14 (10, 44)
14-HDHA	0.37 (0.26, 0.40)	0.40 (0.30, 0.52)	0.61 (0.48, 0.66)	1.3 (0.82, 2.0)	5.3 (1.5, 12)	14 (7.2, 26)	17 (13, 34)	68 (47, 88)
15-HETE	0.71 (0.66, 0.90)	0.80 (0.77, 1.0)	0.97 (0.77, 1.1)	0.88 (0.73, 1.2)	3.7 (1.2, 10)	4.5 (2.8, 13)	5.5 (2.9, 14)	9.1 (4.4, 19)
15-HEPE	0.10 (0.083, 0.12)	0.10 (0.081, 0.12)	0.097 (0.079, 0.12)	0.087 (0.082, 0.012)	0.12 (0.077, 0.22)	0.14 (0.12, 0.32)	0.17 (0.15, 0.37)	0.30 (0.23, 0.86)
17-HDHA	0.38 (0.34, 0.43)	0.49 (0.41, 0.53)	0.85 (0.76, 0.91)	0.65 (0.46, 0.85)	0.79 (0.45, 1.7)	1.4 (0.94, 2.9)	2.1 (1.2, 3.3)	4.1 (2.6, 5.6)
5-HETE	0.58 (0.46, 0.91)	0.65 (0.52, 1.86)	0.72 (0.63, 0.92)	0.57 (0.53, 0.89)	1.4 (1.1, 2.0)	1.5 (1.2, 2.0)	2.0 (1.5, 2.5)	2.1 (1.8, 3.7)
5-HEPE	0.13 (0.12, 0.22)	0.13 (0.12, 0.24)	0.14 (0.11, 0.23)	0.13 (0.10, 0.20)	0.14 (0.11, 0.26)	0.17 (0.11, 0.23)	0.19 (0.15, 0.33)	0.18 (0.15, 0.35)
4-HDHA	0.30 (0.25, 0.38)	0.34 (0.20, 0.39)	0.33 (0.22, 0.38)	0.32 (0.23, 0.36)	0.26 (0.24, 0.44)	0.35 (0.27, 0.42)	0.46 (0.36, 0.58)	0.46 (0.38, 0.57)
7-HDHA	0.099 (0.069, 0.11)	0.078 (0.064, 0.086)	0.078 (0.064, 0.090)	0.077 (0.062, 0.11)	0.084 (0.076, 0.12)	0.073 (0.060, 0.11)	0.090 (0.071, 0.10)	0.14 (0.11, 0.17)
14(15)-EpETrE	0.058 (0.048, 0.067)	0.063 (0.055, 0.070)	0.072 (0.045, 0.079)	0.059 (0.047, 0.065)	0.079 (0.071, 0.097)	0.090 (0.071, 0.11)	0.087 (0.072, 0.15)	0.090 (0.080, 0.093)
17(18)-EpETE	< LLOQ				< LLOQ			
19(20)-EpDPE	0.17 (0.14, 0.24)	0.22 (0.16, 0.26)	0.22 (0.18, 0.27)	0.18 (0.17, 0.28)	0.21 (0.16, 0.25)	0.23 (0.20, 0.29)	0.30 (0.20, 0.33)	0.27 (0.18, 0.33)
14,15-DiHETrE	0.59 (0.54, 0.64)	0.60 (0.51, 0.65)	0.58 (0.54, 0.67)	0.63 (0.54, 0.68)	0.60 (0.56, 0.71)	0.61 (0.57, 0.73)	0.62 (0.56, 0.72)	0.64 (0.59, 0.72)
17,18-DiHETE	0.48 (0.29, 0.97)	0.49 (0.25, 0.99)	0.53 (0.26, 0.92)	0.51 (0.27, 0.94)	0.53 (0.29, 0.99)	0.54 (0.30, 1.0)	0.59 (0.31, 1.1)	0.55 (0.29, 1.1)
19,20-DiHDPE	2.5 (1.9, 3.7)	2.4 (1.7, 3.8)	2.5 (1.9, 3.6)	2.5 (1.9, 3.5)	2.5 (1.9, 3.8)	2.6 (2.0, 3.7)	2.5 (1.9, 3.9)	2.7 (2.0, 3.9)
5(R,S)-5-F _{2t} -IsoP	0.092 (0.072, 0.11)	0.086 (0.064, 0.11)	0.097 (0.085, 0.13)	0.088 (0.071, 0.097)	0.094 (0.082, 0.11)	0.099 (0.086, 0.12)	0.11 (0.10, 0.16)	0.11 (0.096, 0.13)
9-HETE	< LLOQ				< LLOQ			
11-HETE	0.21 (0.19, 0.25)	0.27 (0.21, 0.30)	0.35 (0.31, 0.40)	0.25 (0.21, 0.31)	2.6 (0.56, 7.4)	3.0 (1.3, 9.1)	3.5 (1.4, 8.4)	4.4 (1.9, 11)
18-HEPE	0.21 (0.16, 0.34)	0.22 (0.17, 0.37)	0.23 (0.19, 0.39)	0.24 (0.17, 0.35)	0.32 (0.18, 0.65)	0.42 (0.29, 0.98)	0.48 (0.33, 0.95)	0.51 (0.30, 1.1)

Tab. S2: Median concentrations with interquartil range (25% percentil, 75% percentil) [nM] of representative total, i.e. sum of free and esterified oxylipins in plasma and serum following direct processing, after storage for 4 h or 24 h at 4 °C, or after pneumatic tube system transport (PTS) prior to centrifugation (n =6).

	plasma				serum			
	median (25% percentil, 75% percentil) [nM]				median (25% percentil, 75% percentil) [nM]			
	direct	4 h	24 h	PTS	direct	4 h	24 h	PTS
PGE ₂ *	-	-	-	-	-	-	-	-
TxB ₂ *	-	-	-	-	-	-	-	-
12-HHT	0.43 (0.37, 0.55)	0.51 (0.39, 0.64)	0.66 (0.56, 0.77)	1.1 (0.71, 1.2)	82 (14, 187)	87 (28, 243)	85 (31, 209)	105 (44, 234)
12-HETE	5.4 (4.1, 7.9)	5.9 (5.2, 9.5)	9.0 (6.1, 12)	12 (11, 13)	134 (25, 255)	159 (88, 451)	176 (100, 396)	644 (336, 892)
12-HEPE	0.41 (0.32, 0.63)	0.44 (0.36, 0.66)	0.50 (0.41, 0.77)	0.73 (0.57, 1.1)	2.4 (0.97, 7.3)	4.1 (2.4, 17)	4.8 (4.1, 16)	22 (13, 52)
14-HDHA	1.2 (1.0, 1.5)	1.3 (1.1, 1.6)	1.8 (1.5, 1.9)	2.9 (1.9, 3.5)	9.8 (2.7, 20)	26 (12, 58)	39 (27, 66)	138 (84, 174)
15-HETE	6.2 (5.6, 7.1)	6.1 (5.6, 7.4)	6.9 (6.1, 7.9)	8.9 (7.1, 11)	11 (7.1, 19)	15 (9.3, 27)	17 (10, 26)	39 (19, 44)
15-HEPE	0.65 (0.52, 0.73)	0.64 (0.48, 0.76)	0.77 (0.35, 0.90)	0.79 (0.56, 1.2)	0.76 (0.60, 1.3)	0.92 (0.78, 1.3)	0.99 (0.74, 1.6)	2.0 (1.2, 2.9)
17-HDHA	2.2 (1.8, 2.7)	2.4 (2.2, 2.7)	3.0 (2.6, 3.6)	3.5 (2.9, 5.5)	4.2 (2.5, 5.6)	7.1 (4.6, 12)	7.5 (5.5, 11)	18 (11, 25)
5-HETE	14 (13, 16)	13 (13, 16)	13 (13, 16)	15 (13, 18)	16 (15, 18)	19 (16, 21)	15 (14, 20)	17 (16, 22)
5-HEPE	1.4 (0.90, 2.2)	1.5 (0.93, 2.2)	1.5 (0.92, 2.2)	1.6 (1.0, 2.4)	1.6 (1.1, 2.3)	1.6 (1.1, 2.2)	1.6 (1.1, 2.5)	1.7 (1.4, 2.5)
4-HDHA	2.8 (2.4, 3.7)	2.9 (2.2, 3.7)	2.7 (2.3, 3.9)	2.9 (2.6, 4.2)	3.4 (2.5, 4.3)	4.2 (2.8, 5.4)	3.1 (2.6, 4.1)	3.3 (2.6, 4.2)
7-HDHA	1.7 (1.3, 2.2)	1.7 (1.6, 2.2)	1.6 (1.5, 2.4)	1.9 (1.7, 2.6)	1.9 (1.5, 2.5)	2.6 (1.9, 3.6)	1.9 (1.7, 2.3)	2.2 (1.8, 2.5)
14(15)-EpETrE	45 (38, 58)	51 (27, 55)	44 (37, 53)	40 (37, 68)	44 (41, 64)	53 (44, 76)	59 (49, 68)	46 (42, 52)
17(18)-EpETE	3.6 (2.9, 5.4)	3.8 (2.4, 6.0)	3.5 (3.1, 4.9)	3.7 (2.9, 7.1)	4.4 (3.3, 5.4)	4.5 (3.7, 7.0)	4.2 (3.8, 6.6)	3.6 (2.6, 6.0)
19(20)-EpDPE	9.6 (7.1, 11)	9.7 (5.9, 12)	9.2 (7.9, 10)	10 (8.0, 13)	10 (7.9, 11)	11 (9.6, 15)	11 (9.4, 14)	8.9 (8.1, 12)
14,15-DiHETrE	1.3 (1.2, 1.5)	1.3 (1.1, 1.6)	1.3 (1.2, 1.4)	1.3 (1.2, 1.7)	1.4 (1.2, 1.6)	1.6 (1.3, 2.3)	1.4 (1.2, 1.9)	1.4 (1.2, 1.5)
17,18-DiHETE	0.64 (0.34, 1.1)	0.64 (0.38, 1.2)	0.66 (0.39, 1.1)	0.66 (0.31, 1.1)	0.64 (0.39, 1.2)	0.72 (0.39, 1.2)	0.60 (0.32, 1.3)	0.65 (0.36, 1.2)
19,20-DiHDPE	2.8 (2.2, 4.0)	3.0 (2.3, 4.1)	2.9 (2.2, 4.1)	2.8 (2.3, 4.3)	3.1 (2.2, 4.1)	3.2 (2.9, 4.1)	3.2 (2.1, 4.2)	3.0 (2.1, 4.3)
5(R,S)-5-F _{2t} -IsoP	0.37 (0.32, 0.41)	0.40 (0.26, 0.48)	0.33 (0.22, 0.41)	0.37 (0.33, 0.46)	0.37 (0.31, 0.47)	0.36 (0.32, 0.54)	0.42 (0.36, 0.54)	0.44 (0.38, 0.57)
9-HETE	4.4 (3.6, 5.5)	4.6 (3.8, 5.7)	5.2 (4.4, 5.7)	6.1 (5.0, 7.1)	4.4 (4.2, 5.3)	6.0 (4.8, 7.3)	5.1 (4.4, 6.1)	5.6 (4.7, 8.2)
11-HETE	3.0 (2.6, 3.6)	3.1 (2.8, 3.5)	3.4 (3.0, 4.3)	5.0 (3.4, 5.9)	6.4 (3.4, 11)	7.7 (3.8, 16)	8.3 (4.5, 14)	13 (5.5, 18)
18-HEPE	0.72 (0.70, 1.3)	0.82 (0.68, 1.4)	0.83 (0.81, 1.5)	1.1 (0.84, 1.8)	1.0 (0.76, 1.6)	1.1 (0.85, 2.2)	1.3 (0.90, 2.3)	2.1 (1.3, 3.0)

* PGE₂ and TxB₂ are degraded during alkaline hydrolysis