Supporting Information

Rational design of a novel turn-on fluorescent probe for detecting

hydrazine with barbituric acid as recognition group and bioimaging

Junli Du^{a,b}, Xiaolu Li^{a,b}, Songsong Ruan^{a,b}, Yingchun Li^a, Fan Ren^{a,b}, Yanjun Cao^a, Xiaoqing Wang^c, Yongmin Zhang^{a,b,d}, Shaoping Wu^{a,b*}, Jianli Li^c

^a School of Pharmacy; Key Laboratory of Resource Biology and Biotechnology in Western China (Northwest University), Ministry of Education; Biomedicine Key Laboratory of Shaanxi Province, Northwest University, Xi'an 710069, China

^b Joint International Laboratory of Glycobiology and Medicinal Chemistry, Northwest University, Xi'an, Shaanxi 710069, China

^c Key Laboratory of Synthetic and Natural Functional Molecule Chemistry of Ministry of Education, College of Chemistry & Materials Science, Northwest University, Xi'an, Shaanxi 710127, P. R. China

^d Sorbonne Université, Institut Parisien de Chimie Moléculaire, CNRS UMR 8232, 4 place Jussieu, 75005 Paris, France

* Tel.: +86 029 88304569; Fax: +86 029 88304569. E-mail: <u>wushaoping@nwu.edu.cn</u>

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|------|---------------------------------------|-------------------|-------------------|--|-------------------------|------------------|--------------------------------------|
| Ref | Probe structures | λ_{ex} | λ_{em} | LOD(M) | Sensing pH ranges | Response Time | Biological imaging application |
| [1] | C C C C C C C C C C C C C C C C C C C | 415 nm | 458 nm, 562 nm | 7.4 × 10 ⁻⁸ M | 7-10 | 30 min | HeLa cells |
| [2] | Br, O O O O | 560 nm | 584 nm | 2 × 10 ⁻⁶ M | 7-10 | 60 min | Chinese hamster ovary (CHO) cells |
| [3] | | 440 nm | 517 nm | $1.4 \times 10^{-7} \text{ M}$ | 4-10 | 8 min | HeLa cells |
| [4] | | 685 nm | 715 nm | 1.6 × 10 ⁻⁷ M 5.09 ppb | 6-8 | 30 min | HeLa cells and mice |
| [5] | Br, O, O, O CH ₃ | 365 nm | 450 nm | $7 \times 10^{-8} \mathrm{M}$ | 7-9 | 30 min | HeLa cells |
| [6] | | 450 nm | 510 nm, 660 nm | $5.6 \times 10^{-7} \mathrm{M}$ | 4-10 | 24 h | - |
| [7] | ***= ***= | 675 nm | 706 nm | $1.7 \times 10^{-7} \text{ M}$ | 4-10 | 10 min | HeLa cells and Kunming mouse |
| [8] | | 580 nm, 780 nm | 627 nm, 814 nm | 1.2 × 10 ⁻⁸ M 0.38 ppb | 5-8 | 17 min | HeLa cells and mice |
| [9] | ОН О ОН | 390 nm | 560 nm | 8.42 × 10 ⁻⁸ M (2.7 ppb) | 4-10 | 7 min | HeLa cells |
| [10] | | 405 nm | 590 nm | 2.1 × 10 ⁻⁶ M | 5-10 | 4 min | HepG ₂ cells |
| [11] | | 460 nm | 532 nm, 660 nm | 7.71 × 10 ⁻⁹ M | 3-10 | 60 min | MCF-7 cells |

1. Recent progress in the development of fluorescent probes for the detection of N_2H_4 · H_2O

| Our | O ∭NH | 380 nm | 527 nm | $5 \times 10^{-8} \mathrm{M}$ | 3-12 | 10 min | SH-SY5Y cells |
|------|----------|--------|--------|-------------------------------|------|--------|---------------|
| work | s-(-)-NH | | | | | | |
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| | | | | | | | |

2. Calculation of the detection limit of probe DPT

$$LOD = \frac{3\sigma}{k}$$

Where σ is the standard deviation of the blank measurement, k is the slope between the fluorescence intensity and concentration N₂H₄·H₂O (0~26 μ M). The fluorescence intensity of DPT was measured by eleven times and the standard deviation of the blank measurement was achieved. The calculated LOD of probe DPT are showed in Table S1.

Table S1. Calculated detection limit for probe DPT.

| Probe | DPT |
|--------|-----|
| LOD/nM | 50 |

3. Calculation of fluorescence quantum yield about probe DPT

The quantum yield ($\boldsymbol{\Phi}$) of **DPT** denotes the fluorescence quantum yield. It was measured at room temperature referenced to fluorescein in aqueous solution of 0.1 M sodium hydroxide, which has a quantum yield of 0.92. We calculated the fluorescence quantum yield of **DPT** in different organic solvents. The results were shown in **Table S2**.

Table S2. Calculated the fluorescence quantum yields for DPT in different organic solvents.

| Solvent | МеОН | EtOH | ACN | DMK | DMSO | DMF | ТСМ | DCM | EtOAc | H ₂ O | THF |
|---------------------|------|------|------|------|------|------|------|------|-------|------------------|------|
| Φ_{f} | 0.12 | 0.26 | 0.30 | 0.52 | 0.41 | 0.38 | 0.19 | 0.56 | 0.54 | 0.07 | 0.49 |

 Φ_s was quantum yield of fluorescein; A_x and A_s indicated the absorption intensity of the sample and the standard at the excitation wavelength, respectively; F_x and F_s for the sample and the standard fluorescence integral area; **n** was refractive index of the solvent; Subscript **s** and **x** was the standard and unknown samples, respectively.

$$\Phi_{F(X)} = \Phi_{F(S)} \times \left(\frac{A_S \times F_X}{A_X \times F_S}\right) \left(\frac{n_X}{n_S}\right)^2$$

4. Synthesis of compound 1 and 2



Scheme S1. Reagents and conditions: (a) CH₃I, K_2CO_3 , DMF, 50 °C, 24 h, 48.2%; (b) POCl₃, DMF, 90 °C, 5 h; (c) Na₂S·9H₂O, C₂H₃ClO, DMF, 60 °C, 5 h, 52.1% for two steps.

1) Synthesis of compound 6-(dimethylamino)-3,4-dihydronaphthalen-1(2H)-one (1)

CH₃I (0.42 mL, 6.82 mmol, 5.5 equiv.) was added to a mixture of 6-amino-3,4dihydronaphthalen-1(2H)-one (200 mg, 1.24 mmol, 1.0 equiv.) and K₂CO₃ (377 mg, 2.73 mmol, 2.2 equiv.) in DMF (18 mL) and the mixture was protected from light and stirred for 24 h at 50 °C. After cooling to room temperature, water (10 mL) was added and the solution was extracted with EtOAc and washed with water and brine. The organic layer was combined and dried with anhydrous Na₂SO₄ and evaporated under reduced pressure. The obtained residue was dried and purified by column chromatography on silica gel (petroleum ether/EtOAc, 2:1 v/v) to give compound **1** as a light ginger yellow solid (yield: 48.2 %, R_f = 0.60).

2) Synthesis of compound 7-(dimethylamino)-4,5-dihydronaphtho[1,2-b] thiophene 2-carbaldehyde (2)

POCl₃ (88.5 μ L, 0.95 mmol, 1.3 equiv.) was added dropwise to a flask under nitrogen containing DMF (4 mL) with stirring at 0 °C over 30 min. Compound **1** (138 mg, 0.73 mmol, 1.0 equiv.) in DMF was added slowly with stirring and the mixture was heated for 5 h at 90 °C. Then the mixture was poured to ice water and the resulting precipitate was filtered off and washed with cold water to afford the 1-chloro-6-(dimethylamino)-3,4-dihydronaphthalene-2-carbaldehyde as yellow solid, which was utilized in the next reaction without further purification.

To a solution of $Na_2S \cdot 9H_2O$ (72 mg, 0.30 mmol, 1.1 equiv.) and DMF (5 mL) was added 1-chloro-6-(dimethylamino)-3,4-dihydronaphthalene-2-carbaldehyde (63.4 mmol, 0.27 mmol, 1.0 equiv.). The mixture was stirred at 60 °C during 2 h. Chloroacetaldehyde (50 μ L, 0.30 mmol, 1.1 equiv.) was added rapidly and the reaction was stirred during 3 h at 60 °C. K₂CO₃ (41.5 mg, 0.30 mmol, 1.1 equiv.) was dissolved in water (1.0 mL) and added to the reaction. The mixture was stirred during 10 min at 60 °C, cooled at room temperature and quenched in water. The resulting mixture was extracted with ethyl acetate. The organic layer was combined and dried with anhydrous Na₂SO₄ and evaporated under reduced pressure. The obtained residue was dried and purified by column chromatography on silica gel (petroleum ether/EtOAc, 3:1 v/v) to give compound **2** as a brown solid (yield: 52.1% in two steps, R_f = 0.61).

5. Optical properties of DPT in different organic solvents

The optical properties of **DPT** in different organic solvents included the maximum absorption wavelength (λ_{abs}) , Molar absorption coefficient (ϵ_{max}) , Emission wavelength, Stokes shift. The calculation results are shown in **Table S3**. We could get that there was a large Stokes shift for **DPT**.

| Solvent | $\lambda_{abs}\!/nm$ | $\epsilon_{max}/M^{-1} \cdot cm^{-1}$ | $\lambda_{em}\!/\!nm^a$ | $\Phi_{\mathrm{F}(\mathrm{X})}$ | Stoke | es shift |
|---------|----------------------|---------------------------------------|-------------------------|---------------------------------|-------|------------------|
| | | | | | nm | cm ⁻¹ |
| MeOH | 558 | 27867 | 547 | 0.12 | 167 | 8034 |
| EtOH | 558 | 31333 | 507 | 0.26 | 127 | 6573 |
| ACN | 549 | 16667 | 500 | 0.30 | 120 | 6296 |
| DMK | 543 | 28800 | 495 | 0.52 | 115 | 6114 |
| DMSO | 554 | 43867 | 495 | 0.41 | 115 | 6093 |
| DMF | 549 | 47867 | 494 | 0.38 | 114 | 6052 |
| TCM | 572 | 42267 | 493 | 0.19 | 113 | 6032 |
| DCM | 566 | 36400 | 491 | 0.56 | 111 | 5949 |
| EtOAc | 536 | 35067 | 490 | 0.54 | 110 | 5887 |
| H_2O | 547 | 6133 | 490 | 0.07 | 110 | 5887 |
| THF | 538 | 50000 | 484 | 0.49 | 104 | 5633 |

 Table S3. Optical properties of DPT in different organic solvents.

a: $\lambda_{ex} = 380$ nm.



Fig. S1. (a) Fluorescence spectra of **DPT** (10 μ M) in different organic solvents with N₂H₄·H₂O (20 μ M); (b) Fluorescence spectra of **DPT** (10 μ M) in various volumes of DMSO with N₂H₄·H₂O (20 μ M). $\lambda_{ex} = 380$ nm; slits: (5 nm/5 nm).



Fig. S2. (a) Fluorescence spectra of **DPT** (10 μ M) in different buffers with N₂H₄·H₂O (20 μ M); (b) Fluorescence spectra of **DPT** (10 μ M) in various volumes of PBS buffer with N₂H₄·H₂O (20 μ M). $\lambda_{ex} = 380$ nm; slits: (5 nm/5 nm).

6. Fluorescence spectra of probe DPT reaction with N₂H₄·H₂O



Fig. S3. Fluorescence spectra of probe DPT reaction with N_2H_4 · H_2O .

7. Characterization data of the intermediate and probe DPT



Fig. S4. ¹H-NMR spectrum of probe DPT in TFA- d_1 .



Fig. S5. ¹³C-NMR spectrum of probe **DPT** in TFA- d_1 .



Fig. S6. HRMS of probe DPT.

8. Proposed reaction mechanism of detecting N₂H₄·H₂O



Fig. S7. Proposed reaction mechanism of detecting N_2H_4 · H_2O .

9. DFT optimized structures of probe DPT and $DPT-N_2H_4$



Fig. S8. Density functional theory optimized structures of DPT and compound DPT-N₂H₄.

10. Cytotoxicity experiment of probe DPT

The cell viability (%) was assessed using the following equation:

Cell viability (%) = T/C × 100%, where T is the OD₄₅₀ value of experience group and C is the control group of OD₄₅₀ (*optical density*) value.



 OD_{450} value of each repeated wells are given as mean \pm standard deviation (SD)

Fig. S9. Cytotoxicity assays of probe DPT at different concentrations (0 μ M, 5 μ M, 10 μ M, 15 μ M, 20 μ M, 25 μ M) for SH-SY5Y neuroblastoma cells.

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