Supporting Information

Imaging Stressed Organelle via Sugar-conjugated Color-Switchable pH Sensors

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Contents

Fig. S1 Acidity-independent retention of Sia-FR in lysosomes	3
Fig. S2 Acidity-independent retention of M6C-FR and Sia-FR in di	fferent cell line.
Fig. S3 Cytotoxicity of M6C-FR and Sia-FR	4
Fig. S5 ¹ H NMR spectrum of ^{Az} M6C	5
Fig. S6 ¹³ C NMR spectrum of ^{Az} M6C	5
Fig. S7 ¹ H NMR spectrum of S1	6
Fig. S8 ¹³ C NMR spectrum of S1	6
Fig. S9 ¹ H NMR spectrum of S5	7
Fig. S10 ¹³ C NMR spectrum of S5	7
Fig. S11 ¹ H NMR spectrum of S6	8
Fig. S12 ¹³ C NMR spectrum of S6	8
Fig. S13 ¹ H NMR spectrum of S7	9
Fig. S14 ¹³ C NMR spectrum of S7	9
Fig. S15 HPLC and HRMS analysis of M6C-FR	10
Fig. S16 HPLC and HRMS analysis of Sia-FR	10
Fig. S17 FRET between fluorescein and rhodamine	11



Fig. S1 Acidity-independent retention of Sia-FR in lysosomes. (A) HeLa cells prestained with Sia-FR (20 μ M) or LysoTracker Red (1 μ M) were further incubated in fresh DMEM for 24 h before analysis. (B) HeLa cells prestained with Sia-FR (20 μ M) or LysoTracker Red (1 μ M) were further incubated in DMEM spiked with or without Baf-A1 (20 nM) for 24 h prior to confocal fluorescence microscopy analysis. Scale bars, 10 μ m.



Fig. S2 Acidity-independent retention of M6C-FR (A) and Sia-FR (B) in different cell lines. HeLa cells prestained with M6C-FR or Sia-FR (20 μ M) were further incubated in DMEM spiked with or without Baf-A1 (20 nM) for 24 h prior to confocal fluorescence microscopy analysis. Scale bars, 10 μ m.



Fig. S3 Cytotoxicity of M6C-FR and Sia-FR. HeLa cells prestained with M6C-FR (A) or Sia-FR (B) (0-40 μ M) for 0-48 h. At indicated time points, the cells were determined for cell viability by MTT assay.



Fig. S5 ¹H NMR spectrum of ^{Az}M6C (CDCl₃).



Fig. S6 13 C NMR spectrum of Az M6C (CDCl₃).



Fig. S7 ¹H NMR spectrum of S1 (CDCl₃).



Fig. S8 13 C NMR spectrum of S1 (CDCl₃).







Fig. S10 ¹³C NMR spectrum of S5 (DMSO-d₆).







Fig. S13 ¹H NMR spectrum of S8 (CDCl₃).



Fig. S14 ¹³C NMR spectrum of S8 (CDCl₃).



Fig. S15 HPLC and HRMS analysis of M6C-FR.



Fig. S16 HPLC and HRMS analysis of Sia-FR.



Fig. S17 FRET between fluorescein and rhodamine. The green fluorescence emission at 515 nm of 5-FAM and M6C-FR and was determined as a function of buffer pH. The lower lelvels of fluorescence of fluorescein over M6C-FR at pH 9.0-6.5 is due to intramolecular effects of rhodamine-lactam on fluorscein moiety, because nonfluorescent rhodamine-lactam was present at neutral to alkaline pH. The assay also suggests insignificant FRET between fluorescein and rhodamine-amide moiety at pH 4.0-5.5.