## **Supporting Information**

- 2 3 Detection of circulating tumor cells based on improved SERS-active magnetic nanoparticles 4 Ting Xue<sup>1,2</sup>, Siqi Wang<sup>3,4</sup>, Guoyu Ou<sup>3</sup>, Yong Li<sup>5</sup>, Huimin Ruan<sup>1</sup>, Zihou Li<sup>1</sup>, YuanYuan Ma<sup>1</sup>, Ruifen 5 Zou<sup>1</sup>, Jiaoyan Qiu<sup>1</sup>, Zheyu Shen<sup>1,\*</sup>, Aiguo Wu<sup>1,\*</sup> 6 7 1. Cixi Institute of Biomedical Engineering, CAS Key Laboratory of Magnetic Materials and 8 Devices, & Key Laboratory of Additive Manufacturing Materials of Zhejiang Province, & Division 9 of Functional Materials and Nanodevices, Ningbo Institute of Materials Technology and 10 Engineering, Chinese Academy of Sciences, 1219 Zhong-guan West Road, Ning-bo, Zhe-jiang 11 315201, China. 12 2. University of Chinese Academy of Sciences, 19 A Yu-quan Road, Shi-jing-shan District, Bei-jing 13 100049, China. 14 3. Department of Radiology, The Affiliated Hospital of Medical School of Ningbo University, 15 16 Ningbo University School of Medicine, Ning-bo, Zhe-jiang 315020, China. 4. Department of Nuclear Medicine, Sir Run Run Shaw Hospital, Zhejiang University, Hang-zhou, 17 Zhe-jiang 310020, China. 18 5. Analysis and Test Center, Ningbo Institute of Materials Technology and Engineering, Chinese 19 Academy of Sciences, 1219 Zhong-guan West Road, Ning-bo, Zhe-jiang 315201, China. 20 21 **Corresponding Authors** 22 \*E-mail: shenzhevu@nimte.ac.cn; Tel: +86 574 87617278 23 \*E-mail: aiguo@nimte.ac.cn; Tel: +86 574 86685163 24
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## 1 Materials

Ethylene glycol ( $C_2H_6O_2$ ), sodium acetate anhydrous ( $C_2H_3NaO_2$ ), sodium borohydride (NaBH<sub>4</sub>), 2 folic acid (C<sub>19</sub>H<sub>19</sub>N<sub>7</sub>O<sub>6</sub>), albumin from bovine serum (BSA), 1-ethyl-3-[3-(dimethyllamino)propyl] 3 carbodiimide hydrochloride (EDC·HCl) as well as N-hydroxysuccinimide (NHS) were ordered from 4 Aladdin Reagent Co. Ltd. (Shanghai, China). Iron chloride hexahydrate (FeCl<sub>3</sub>·6H<sub>2</sub>O) was purchased 5 from Alfa Aesar. Tetrachloroauric(III) acid tetrahydrate (HAuCl<sub>4</sub>·4H<sub>2</sub>O), trisodium citrate dehydrate 6 (C<sub>6</sub>H<sub>5</sub>Na<sub>3</sub>O<sub>7</sub>·2H<sub>2</sub>O) and Hoechst were purchased from Sinopharm Chemical Reagent Co. Ltd. 7 (Shanghai, China). 4-mercaptobenzoic acid (MBA), polyethylenimine (Mw~25000), DMSO and MTT 8 was ordered from Sigma-Aldrich. Lymphocyte isolation was ordered from Slolarbio Life Science Co. 9 Ltd. (Beijing, China). Fetal bovine serum (FBS), incomplete DMEM (high glucose) as well as trypsin-10 EDTA were ordered from KeyGen BioTech. Anti-CD45 antibody [F10-89-4] (Alexa Fluor® 488) 11 Alexa and Anti-Cytokeratin 8 antibody [EP1628Y] (Alexa Fluor® 647) were purchased from Abcam. 12 (Zhejiang, China). 13

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## 15 Instrumentation.

The nanoparticles were characterized by transmission electron microscopy (TEM, JEOL. 2100, 16 Tokyo, Japan), and UV-vis spectroscopy (T10CS, Beijing Purkinje General Instrument Co., Ltd., 17 China). The Raman spectra were observed on a confocal microprobe Raman system (Renishaw inVia 18 Reflex, Wolton-under-Edge, U.K.). The laser wavelength was fixed at 785 nm. The range of the 19 scattering spectra was set from 400 to 1500 cm<sup>-1</sup>, the time of data acquisition was set to 3.0 s, and the 20 laser power was 280 mW. The SERS spectra were observed from liquid samples with homogeneous 21 SERS hotspot. The diameter of the laser beam was tuned to be  $\sim 1.0$  mm to capture lots of SERS 22 hotspot at the same time. The particle size and size distribution of the nanoparticles were measured at 23 room temperature by dynamic light scattering (DLS) using a zeta particle size analyzer (Malvern, 24 England) with a detection angle of scattered light at 173°. Magnetic properties of the nanoparticles 25

- 1 were characterized using a Quantum Design Model-9 PPMS (Quantum Design, USA) by measuring
- 2 the applied field dependence of magnetization between -30 and 30 kOe at 300 K.

Table S1. Synthesis conditions and characterization results of SPION-PEI@AuNPs-MBA and
 SPION-PEI@AuNPs-MBA-rBSA-FA.

Nomenclature	C <sub>Au</sub> (mg/mL)	C <sub>Fe</sub> (mg/mL)	C <sub>MBA</sub> (µM)	C <sub>rBSA-FA</sub> (µg/mL)	SERS intensity
SPION-PEI@AuNPs-MBA1	0.17	0.15	2.5	-	5890±230
SPION-PEI@AuNPs-MBA2	0.17	0.15	5	-	9220±400
SPION-PEI@AuNPs-MBA3	0.17	0.15	10	-	14690±360
SPION-PEI@AuNPs-MBA4	0.17	0.15	25	-	5480±350
SPION-PEI@AuNPs-MBA3-rBSA-FA1	0.17	0.15	10	11	11440±220
SPION-PEI@AuNPs-MBA3-rBSA-FA2	0.17	0.15	10	25	10190±440
SPION-PEI@AuNPs-MBA3-rBSA-FA3	0.17	0.15	10	45	14760±960
SPION-PEI@AuNPs-MBA3-rBSA-FA4	0.17	0.15	10	60	11880±1010

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3 Fig. S1 X-ray diffraction (XRD) spectrum of SPION-PEI.

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2 Fig. S2 (a) Size distribution of SPION-PEI@AuNPs-MBA3-rBSA-FA3, SPION-PEI and AuNPs. (b)

3 Zeta potential of SPION-PEI@AuNPs-MBA3-rBSA-FA3, SPION-PEI@AuNPs-MBA3, SPION-

4 PEI@AuNPs, SPION-PEI and AuNPs.

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Fig. S3 (a) UV-visible spectroscopy of FA standard solutions with different concentrations. (b) A
calibration curve constructed from the spectroscopy of FA standard solutions with different
concentrations. (c) UV-visible spectroscopy of the supernatant during the purification of rBSA-FA.
FA conjugated content (FCC) = mass of FA conjugated to rBSA / mass of rBSA-FA × 100 %.



2 Fig. S4 Microscope images of (a) unseparated HeLa from WBCs, and (b-d) captured HeLa cells from

3 blood.