## **Electronic Supplementary Information (ESI)**

# Mitochondria-nucleolus migration fluorescent probe for monitoring of mitochondrial membrane potential and identification of cell apoptosis

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#### **Instruments and Materials**

UV-vis absorption spectra were measured in 1 cm quartz cells with a TU-1901 (PERSEE) UV-vis spectrophotometer. A F7000 fluorescence spectrophotometer (HITACHI) at 298 K was used for fluorescence measurements with a 600 V PMT voltage.<sup>1</sup>H NMR and <sup>13</sup>C NMR were measured on an Agilent AM400 NMR spectrometer using TMS as internal standard. High resolution electrospray ionization mass spectra (HR-ESI-MS) were recorded on a BrukerDaltonics Bio TOF mass spectrometer. Fluorescence imaging was conducted on a ZEISS LSM 780 confocal laser scanning microscope.

Unless otherwise noted, materials were obtained from commercial suppliers and were used without further purification. All of solvents used were analytical-reagent grade in synthesis and HPLC in the optical spectroscopic studies.

#### **DNase and RNase Treatment**

HeLa cells were fixed at -20 °C for 10 min in a cold MeOH solution and then washed with PBS three times. DNase (20 µg/mL) or RNase (25 µg/mL) solutions were added into the samples and incubated for 3 h in the cell culture incubator. Then cell samples were incubated with 5 µM **Cz-Bz-1** for another 30 min. The cells were washed with PBS three times and imaged.

#### **Synthesis**

Synthesis of **Cz-Bz-1**: Compound **3**<sup>1</sup> (2.0 mmol, 0.47 g) and **2**<sup>2</sup> (5 mmol, 1.76 g) were dissolved in 50 mL EtOH. 1 drop of piperidine was added and the mixture was refluxed for 12 hours. After cooling to room temperature, the solvent was removed under reduced pressure; the crude product was purified by silica gel column (DCM:MeOH 100:1) to give the product as a red solid (0.54 g) in 30% yield. <sup>1</sup>H NMR (400 MHz, DMSO-*d6*)  $\delta$  9.39 (s, 2H), 8.73 (d, *J* = 16.0 Hz, 2H), 8.49 (t, *J* = 9.4 Hz, 4H), 8.31 (d, *J* = 8.8 Hz, 2H), 8.23 (d, *J* = 8.0 Hz, 2H), 8.14 (d, *J* = 8.8 Hz, 2H), 7.95 (d, *J* = 8.4 Hz, 2H), 7.91-7.80 (m, 4H), 7.73 (t, *J* = 7.0 Hz, 2H), 4.37 (s, 6H), 4.09 (s, 3H), 2.10 (s, 12H). <sup>13</sup>C NMR (100 MHz, DMSO-*d6*)  $\delta$  182.0 (s), 153.3 (s), 144.5 (s), 139.6 (s), 137.6 (s), 133.0 (s), 130.8 (s), 130.1 (s), 128.4 (s), 127.2 (s), 127.0 (s),126.7 (s), 124.6 (s), 123.2 (s), 113.3 (s), 111.3 (s), 110.1 (s), 53.5 (s), 35.3 (s), 30.1 (s), 25.5 (s). HRMS (m/z): [M-2I]<sup>2+</sup> calcd. for C<sub>47</sub>H<sub>43</sub>N<sub>3</sub><sup>2+</sup>, 324.6723, found 324.6707.



Fig. S1 Normalized absorption (A) and fluorescence spectra (B) of Cz-Bz-1 (2  $\mu$ M) in different solvents.  $\lambda_{ex} = 460$  nm.

Table S1: Photophysical properties of Cz-Bz-1 in different solvents.

Solvents	Maxima $\lambda_{ab}$	Maxima λ <sub>em</sub>	ε (M <sup>-1</sup> · cm <sup>-1</sup> )	Φ(%)
PBS	460	640	43000	1.5
DCM	572	617	49000	4.8
THF	534	628	43500	2.1
ACN	501	614	37500	
MeOH	519	619	58500	4.1
DMSO	524	628	56500	5.8

 $\epsilon$  (M<sup>-1</sup> cm<sup>-1</sup>): molar absorption coefficients at maximum absorption wavelengths,  $\Phi$ :absolute fluorescence quantum yields in different solvents.



Fig. S2 Fluorescence intensity ( $\lambda_{em}$ = 640 nm) of Cz-Bz-1 (2  $\mu$ M) at different pH in PBS.  $\lambda_{ex}$  = 460 nm.



**Fig. S3** Absorption titrations of **Cz-Bz-1** (2 μM) in PBS with increasing concentrations of DNA (A) and RNA (B). DNA and RNA concentrations: 0-300 μg/mL.



Fig. S4 Fluorescence titrations of Cz-Bz-1 (2  $\mu$ M) in PBS with increasing concentrations of DNA (A) and RNA

(B). DNA and RNA concentrations: 0-300  $\mu g/mL,\,\lambda_{ex}\!=\!460$  nm.



Fig. S5 Fluorescent images of DNase and RNase digest experiments for Cz-Bz-1 (5  $\mu$ M).  $\lambda_{ex} = 488$  nm,  $\lambda_{em} = 600-650$  nm; scale bar = 10  $\mu$ m.



Fig. S6 MTS (A) and ATP detection (B) results of HeLa cell viabilities after incubation with Cz-Bz-1 for 24 h at different incubation concentrations. The viability of cells without Cz-Bz-1 is defined as 100%. The results are expressed as the mean  $\pm$  SD of three separate measurements.



Fig. S7 <sup>1</sup>H NMR spectrum of Cz-Bz-1.



Fig. S8 <sup>13</sup>C NMR spectrum of Cz-Bz-1.



Fig. S9 HRMS spectrum of Cz-Bz-1.

### Reference

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- 2. N. Narayanan, G. Patonay, J. Org. Chem., 1995, 60, 2391-2395.