

Electronic supplementary information

Tuning the matrix metalloproteinase-1 degradability of peptide amphiphile nanofibers through supramolecular engineering

Yejiào Shi,^a Daniela S. Ferreira,^a Jayati Banerjee,^a Andrew R. Pickford^b and Helena S. Azevedo^{a*}

^a School of Engineering and Materials Science & Institute of Bioengineering, Queen Mary, University of London, E1 4NS, UK, E-mail: h.azevedo@qmul.ac.uk

^b Centre for Enzyme Innovation & School of Biological Sciences, University of Portsmouth, PO1 2DY, UK

* E-mail: h.azevedo@qmul.ac.uk

Contents

Fig. S1	Templates for modelling of the MMP-1:GPQGIWGQ peptide complex	S2
Fig. S2	Characterization of PS (CH ₃ CONH-GPQGIWGQKKK-CONH ₂)	S3
Fig. S3	Characterization of PA1 (C ₁₅ H ₃₁ CONH-GPQGIWGQKKK-CONH ₂)	S3
Fig. S4	Characterization of PA2 (C ₁₅ H ₃₁ CONH-AAAAAGPQGIWGQKKK-CONH ₂)	S4
Fig. S5	Characterization of PA3 (C ₁₅ H ₃₁ CONH-VVVAAGPQGIWGQKKK-CONH ₂)	S4
Fig. S6	Characterization of PA4 (C ₁₅ H ₃₁ CONH-GPQGIWGQVVVAAKKK-CONH ₂)	S5
Fig. S7	Characterization of PF (NH ₂ -IWGQKKK-CONH ₂)	S5
Fig. S8	CAC determination of PS and PAs	S6
Fig. S9	Identification of the peptide fragment from MMP-1 degradation	S7
Fig. S10	MMP-1 degradation of PS nanofibers	S7
Fig. S11	MMP-1 degradation of PA3 and PA4 nanofibers	S8
Fig. S12	Standard curve of PF	S9

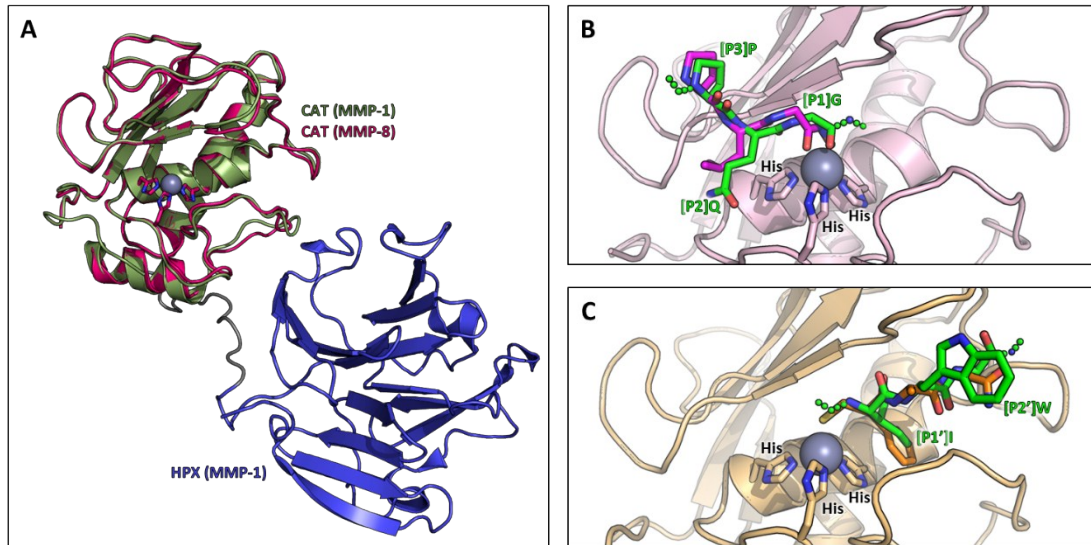


Fig. S1 Templates for modelling of the MMP-1:GPQGIWGQ peptide complex: (A) structural homology between the MMP-8 catalytic (CAT) domain (PDB accession code 1JAO) and that of MMP-1 (2CLT). Note that the MMP-8 crystal structure lacks the C-terminal hemopexin (HPX) domain; (B) modelling of the [P3]Pro-[P2]Gln-[P1]Gly tripeptide fragment (green) on the crystal structure of MMP-8 (pale pink) in complex with the inhibitor Pro-Leu-Gly-hydroxamate (magenta; 1JAP). The slight displacement of the peptide backbone arises from differential interactions of the active site zinc (grey sphere). In the MMP-8:inhibitor complex, this ion is chelated by the inhibitor's hydroxamate group, but in our model it is chelated by the hydrolyzing water molecule (not shown) that performs a nucleophilic attack on the carbonyl carbon of the scissile peptide bond; (C) Modelling of the [P1']Ile-[P2']Trp-[P3']Gly tripeptide fragment (green) on the crystal structure of MMP-8 (pale orange) in complex with the inhibitor 3-mercapto-2-benzylpropanoyl-Ala-Gly-NH₂ (orange; 1JAO). The backbone atoms of [P3']Gly lie directly behind the sidechain of [P2']Trp and, hence, are not labelled.

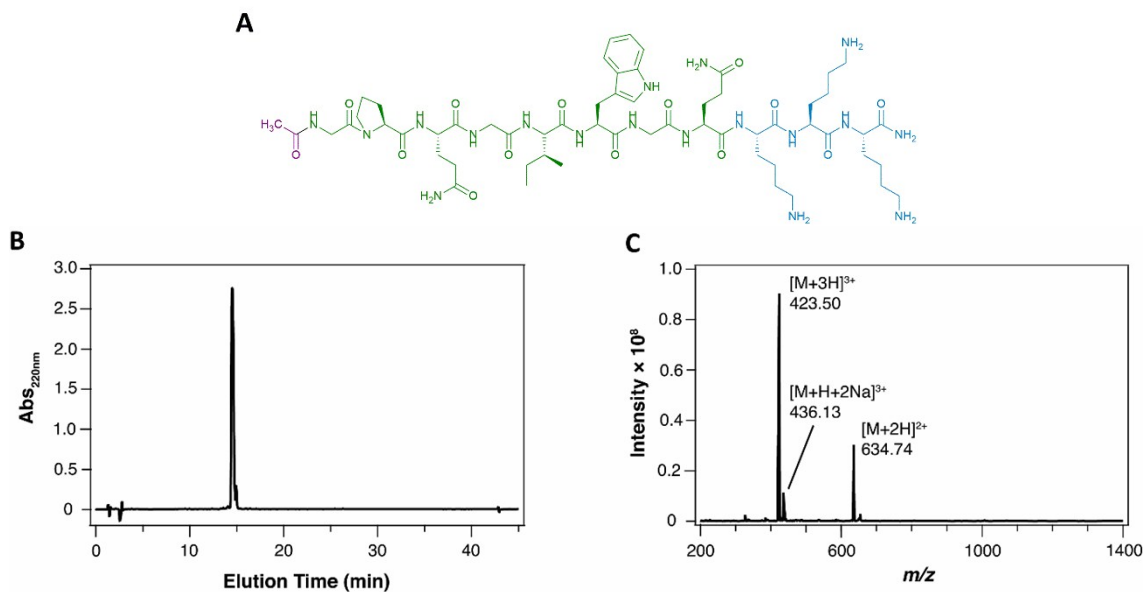


Fig. S2 Characterization of PS ($\text{CH}_3\text{CONH-GPQGIWGQKKK-CONH}_2$): (A) chemical structure; (B) analytical RP-HPLC chromatogram under the gradient of 98% to 0% H_2O (2% to 100% ACN) with 0.1% TFA from 5 to 35 min showing high purity; (C) ESI-MS spectrum showing the expected molecular mass ($\text{C}_{58}\text{H}_{94}\text{N}_{18}\text{O}_{14}$, Mw: 1267.49 g/mol).

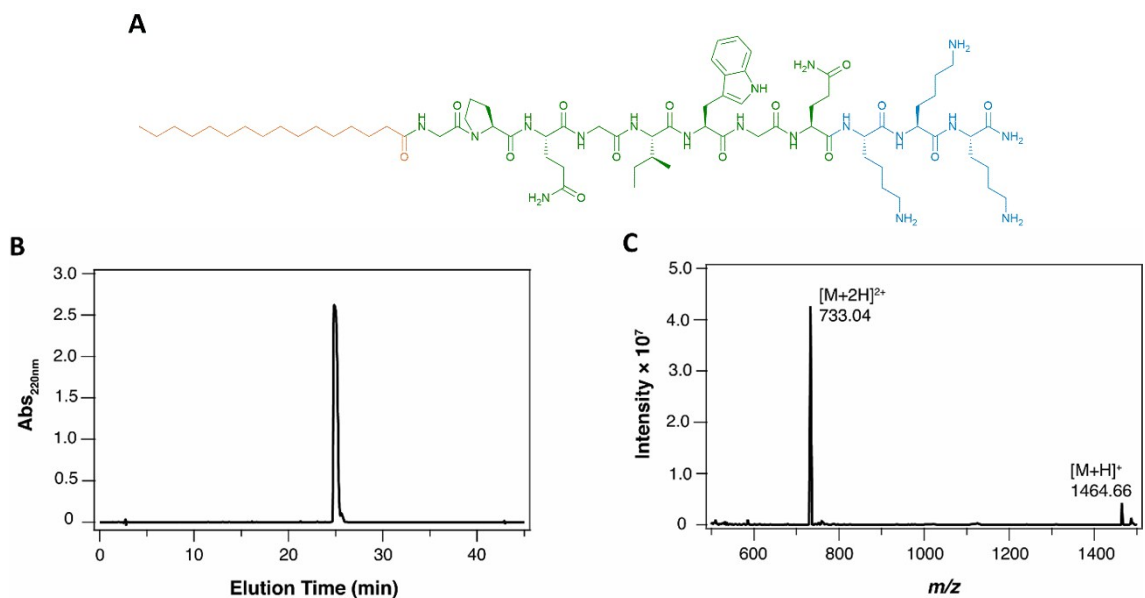


Fig. S3 Characterization of PA1 ($\text{C}_{15}\text{H}_{31}\text{CONH-GPQGIWGQKKK-CONH}_2$): (A) chemical structure; (B) analytical RP-HPLC chromatogram under the gradient of 98% to 0% H_2O (2% to 100% ACN) with 0.1% TFA from 5 to 35 min showing high purity; (C) ESI-MS spectrum showing the expected molecular mass ($\text{C}_{72}\text{H}_{122}\text{N}_{18}\text{O}_{14}$, Mw: 1463.86 g/mol).

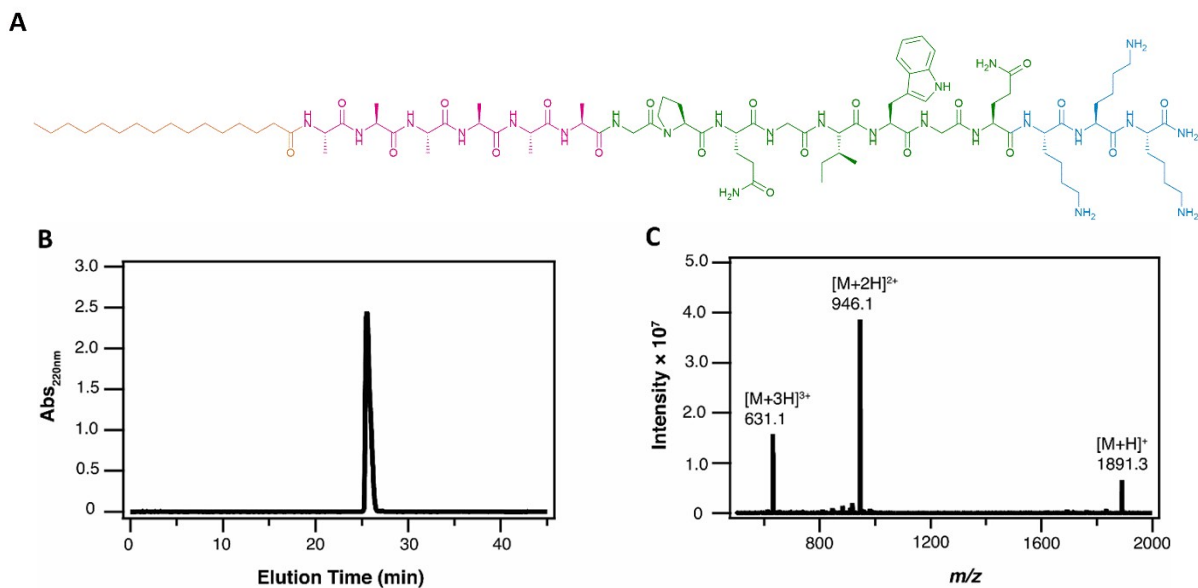


Fig. S4 Characterization of PA2 (C₁₅H₃₁CONH-AAAAAGPQGIWGQKKK-CONH₂): (A) chemical structure; (B) analytical RP-HPLC chromatogram under the gradient of 98% to 0% H₂O (2% to 100% ACN) with 0.1% TFA from 5 to 35 min showing high purity; (C) ESI-MS spectrum showing the expected molecular mass (C₉₀H₁₅₂N₂₄O₂₀, Mw: 1890.33 g/mol).

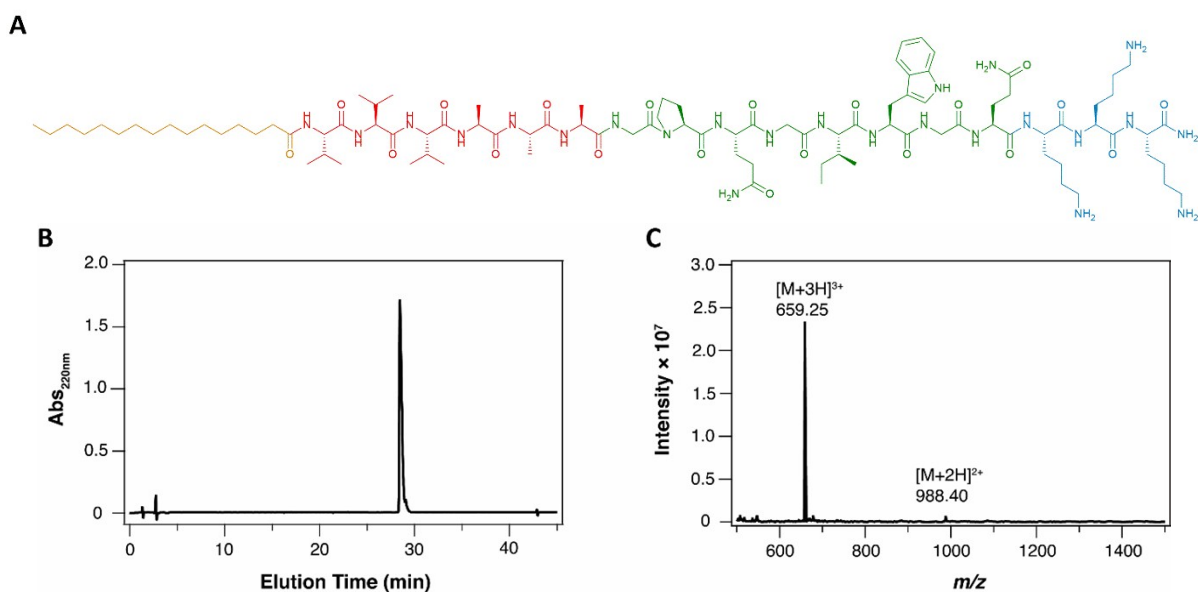


Fig. S5 Characterization of PA3 (C₁₅H₃₁CONH-VVVAAGPQGIWGQKKK-CONH₂): (A) chemical structure; (B) analytical RP-HPLC chromatogram under the gradient of 98% to 0% H₂O (2% to 100% ACN) with 0.1% TFA from 5 to 35 min showing high purity; (C) ESI-MS spectrum showing the expected molecular mass (C₉₆H₁₆₄N₂₄O₂₀, Mw: 1974.49 g/mol).

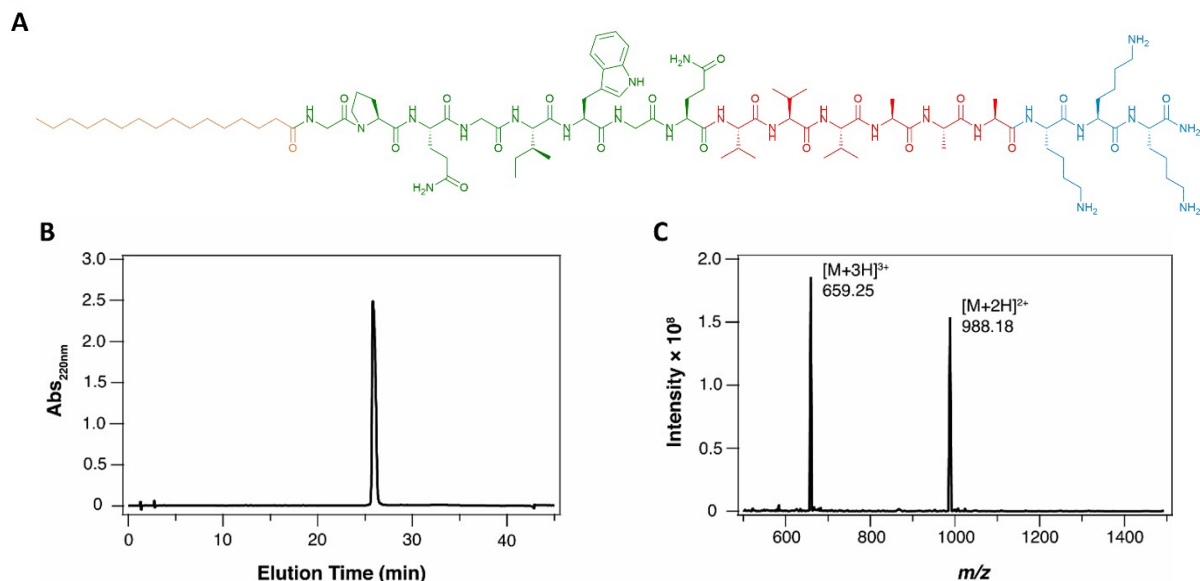


Fig. S6 Characterization of PA4 ($C_{15}H_{31}CONH-GPQGIWGQVVVAAAKK-CONH_2$): (A) chemical structure; (B) analytical RP-HPLC chromatogram under the gradient of 98% to 0% H_2O (2% to 100% ACN) with 0.1% TFA from 5 to 35 min showing high purity; (C) ESI-MS spectrum showing the expected molecular mass ($C_{96}H_{164}N_{24}O_{20}$, Mw: 1974.49 g/mol).

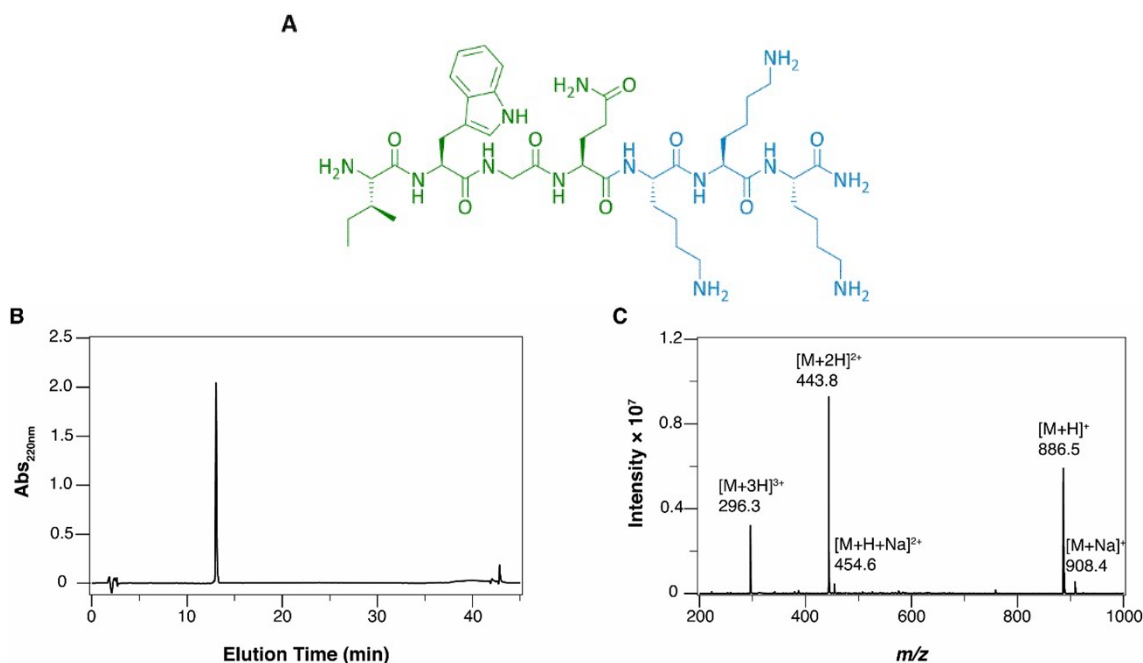


Fig. S7 Characterization of PF ($NH_2-IWGQKKK-CONH_2$): (A) chemical structure; (B) analytical RP-HPLC chromatogram under the gradient of 98% to 0% H_2O (2% to 100% ACN) with 0.1% TFA from 5 to 35 min showing high purity; (C) ESI-MS spectrum showing the expected molecular mass ($C_{42}H_{71}N_{13}O_8$, Mw: 886.10 g/mol).

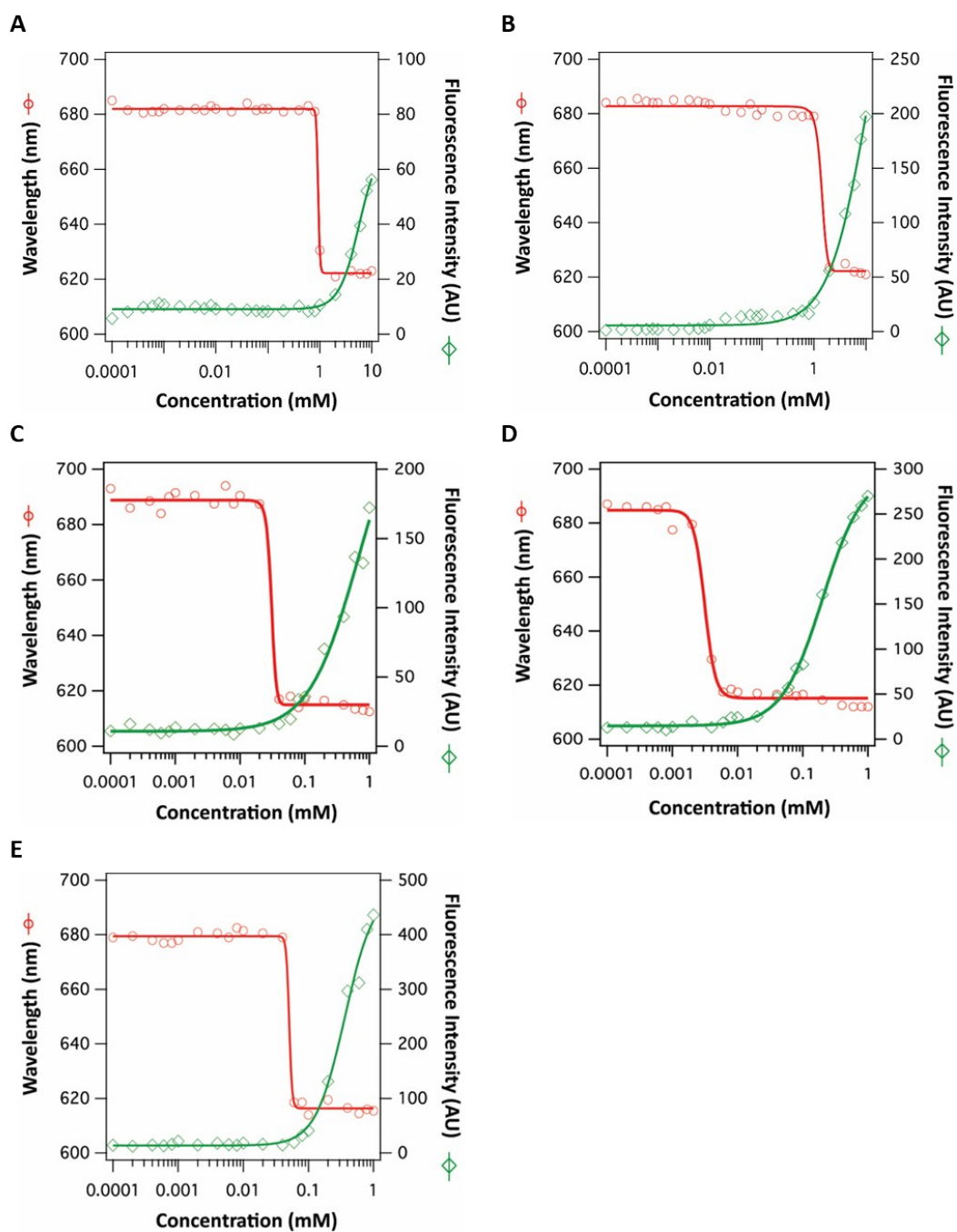


Fig. S8 CAC determination of PS and PAs: maximum fluorescence emission wavelength and intensity of Nile red as a function of PS and PA concentration to determine the CAC of PS at 2 mM (A), PA1 at 2 mM (B), PA2 at 0.04 mM (C), PA3 at 0.004 mM (D), and PA4 at 0.06 mM (E).

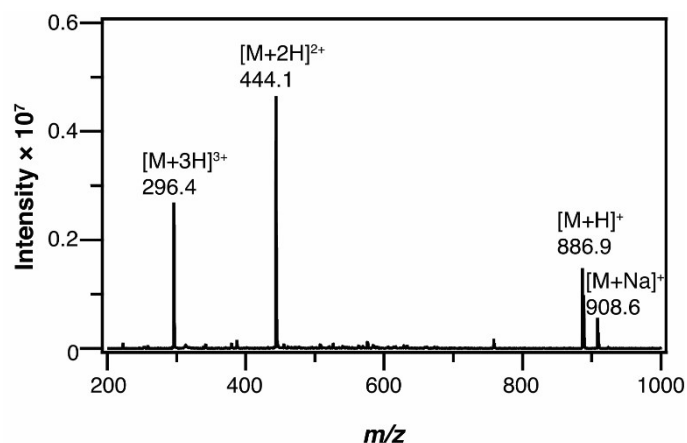


Fig. S9 Identification of the peptide fragment from MMP-1 degradation: representative ESI-MS spectrum of the fractions collected from the RP-HPLC (peak at 13 min in **Figure 4**) showing the molecular mass corresponding to the expected *N*-terminal PF ($\text{NH}_2\text{-IWGQKKK-CONH}_2$, $\text{C}_{42}\text{H}_{71}\text{N}_{13}\text{O}_8$, Mw: 886.10 g/mol).

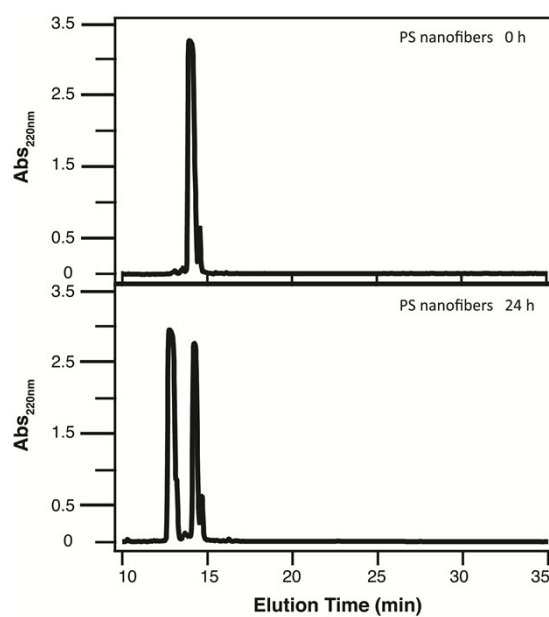


Fig. S10 MMP-1 degradation of PS nanofibers: analytical RP-HPLC traces of 5 mM PS before and after incubation with 20 nM active MMP-1 in TCNB buffer (pH 7) at 37 °C for 24 h.

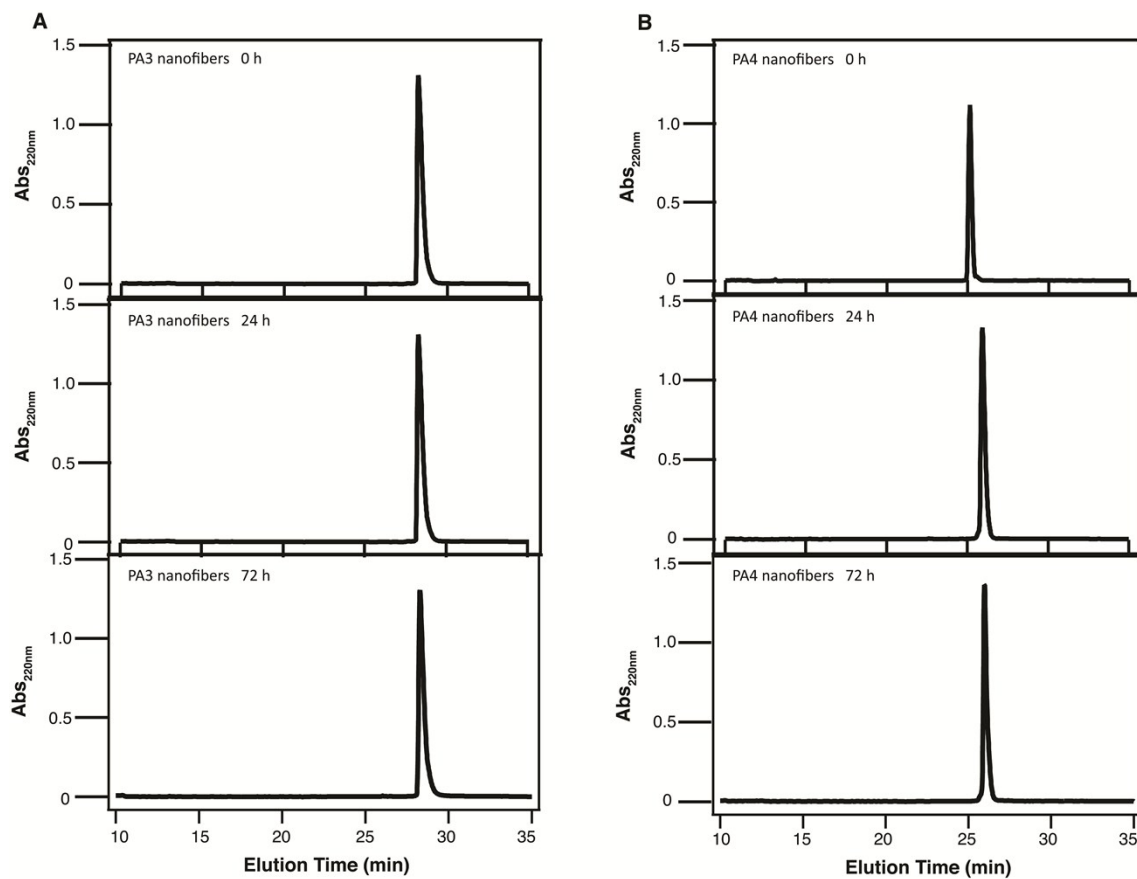


Fig. S11 MMP-1 degradation of PA3 and PA4 nanofibers: analytical RP-HPLC traces of 0.5 mM PA3 (A) and PA4 (B) before and after incubation with 20 nM active MMP-1 in TCNB buffer (pH 7) at 37 °C for 72 h.

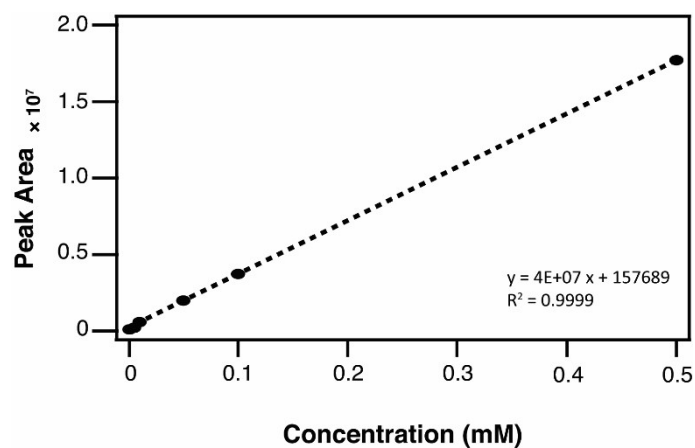


Fig. S12 Standard curve of PF: peak area of PF measured from the RP-HPLC chromatogram as function of its concentration.