Supporting Information

Microencapsulation improves chondrogenesis *in vitro* and cartilaginous matrix stability *in vivo* compared to bulk encapsulation

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Gelatin-norbornene synthesis

5-norbornene-2-carboxylic acid (1.00 g, 7.20 mmol) (Sigma) was first dissolved in 15 mL dichloromethane (DCM) (Merck Millipore). Then, *N*-hydroxysuccinimide (1.12 g, 9.72 mmol) (Sigma) was dissolved in DCM (5 mL) and mixed into the previous norbornene solution with gentle swirling. To this mixture, 1.79 g of *N*-(3-(dimethylamino)propyl)-*N*'-ethylcarbodiimide hydrochloride (9.36 mmol) was added, the final mixture was stirred and the reaction was allowed to continue over 15 h. The mixture was washed with 40 mL deionised water, dried with anhydrous magnesium sulphate (MgSO₄) and concentrated *in vacuo*. The resultant product was used directly in the next step with no further purification.

Next, type A gelatin (1.00 g) (Porcine skin, Sigma) was dissolved in 20 mL *N*,*N*-dimethylformamide (DMF)/water (1:1 v/v), stirred at 37 °C and recovered to room temperature when fully dissolved. Then, the above intermediate product (59 mg, 0.025 mmol) was dissolved in 5 mL DMF and further added into the gelatin solution with additional 50 μ L *N*,*N*-diisopropylethylamine (DIPEA) (Sigma) and stirred at ambient temperature for 10 h. The resultant product was transferred into a dialysis tube with 3.5 kDa cut-off and dialysed against deionised water in excess of water, changed over 5 days. The resulting purified solution was lyophilised to yield the final cotton-like GelNB product. Norbornene

functionalisation was confirmed via ¹H NMR. The percentage of substitution was characterised by fluoroaldehyde assay (ThermoFisher) by examining the content of amine groups before and post the funcrtionlisation process following the protocol provided by manufacturer. The norbornene substitution of the gelatin was measured to be 48% with the L-lysine as the reference.

Α

В



0 50 100 150 200 250 300 350 400 450 Lysine concentration (μg/mL)

Figure S1. (A) ¹H NMR of raw gelatin and Gelatin-norbornene (GelNB), the sample contains trace of D_2O , (B) The standard curve of fluorescence intensity versus amine concentration based on different L-lysine concentrations

PEGdiSH synthesis

4 g of PEG (2 mM, Mw = 2kDa) and 600 mg of triethylamine (6 mM) were dissolved in DCM (20 mL) and cooled on ice. Then, 458 mg of methanesulfonyl chloride (4 mM) were added dropwise with stirring over 30 min. The mixture was stirred for 4 h, and a white powder was precipitated in 100 mL diethyl ether.

The resultant product with additional potassium thioacetate (912 mg, 8 mM) was dissolved in 10 mL DMF. The solution was stirred and allowed to continue overnight at room temperature before being concentrated *in vacuo*. The mixture was dissolved in DCM (20 mL) and washed with 10 mL of brine and 10 mL of water, then dried over anhydrous MgSO₄. The dried solution was then evaporated *in vacuo*. The concentrated product was subsequently redissolved in HCl (5 N, 20 mL) and heated to reflux at 120 °C under Dean-Stark conditions overnight. The resultant product was extracted in DCM (20 mL x 3), dried over anhydrous MgSO₄ and concentrated to 5 mL. The final product was precipitated into diethyl ether as a white powder. ¹H NMR confirmed the thiol functionalisation of PEG. Ellman's test was conducted to determine the amount of free thiol from the synthesised PEG-diSH. an aqueous solution of PEG-SH (1.5 mL, 0.05 mM, Tris buffered pH 7.4) in a quartz cuvette equipped with a stirring bar was first corrected as background on the Cary60. To this solution was added a solution of 5,5'-dithiobis-(2-nitrobenzoic acid) (0.2 mM in Tris buffered pH 7.4) The solution was allowed to react with stirring in the dark for 10 min and the absorbance at 412 nm was recorded. L-cysteine solutions (0.025-0.2 mM) were used as standards for calculation of the amount of thiol release. Each measurement was done in triplicate.



Figure S2. (A) ¹H NMR of PEG-diSH, the sample contains trace of acetone and dichloromethane, (B) Cysteine calibration curve for Ellman's test.

Α

В

NHSA-microgels

Bulk

Bulk empty



Figure S3. H&E staining for hCC-laden bulk hydrogels, NHSA-microgels and bulk empty gel (without cells) after in vivo chondrogenesis at day 49.

mRNA		Primers	Ref.
Sox-9	Forward Reverse	5'-TCT GGA GAC TTC TGA ACG AGA GC-3' 5'-TGT AAT CCG GGT GGT CCT TC-3'	NM_000346.3
Aggrecan	Forward Reverse	5'-GAA TGG GAA CCA GCC TAT ACC-3' 5'-TCT GTA CTT TCC TCT GTT GCT G-3'	NM_01135_3
Col2A1	Forward Reverse	5'-GGA ATT CGG TGT GGA CAT AGG-3' 5'-ACT TGG GTC CTT TGG GTT TG-3'	NM_001844
Col1A1	Forward Reverse	5'-CAG CCG CTT CAC CTA CAG C-3' 5'-TTT TGT ATT CAA TCA CTG TCG CC-3'	NM_000088
RPL13a ¹	Forward Reverse	5'-AAG TAC CAG GCA GTG ACA G-3' 5'-CCT GTT TCC GTA GCC TCA TG-3'	NM_012423
GAPDH	Forward Reverse	5'-AGT CAG CCG CAT CTT CTT TT-3' 5'-CCA ATA CGA CCA AAT CCG TTG-3'	NM_002046

 Table S1.
 Primers used for RT-qPCR

1. D. Studer, S. Lischer, W. Jochum, M. Ehrbar, M. Zenobi-Wong and K. Maniura-Weber, *Tissue Engineering Part C: Methods*, 2012, **18**, 761-771.



Figure S4. Housekeeping genes quantification Ct results. (A) GAPDH; (B) RPL13a. RPL13a values were more stable across all the experimental conditions compared to GAPDH data. RPL13a was determined as the reference gene for other target genes.