Supplementary Material

A new lysosome-targetable fluorescent probe for detection of endogenous hydrogen polysulfides in living cells and inflamed mouse model

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Table S1. Comparison of fluorescent probes for H_2S_n .

Probes	Fluorescence recovery	Stokes shift (nm)	Limit of detection	Response time	Analytical applications	Reference
	About 4-fold	25	3 nM	10 min	Living cells	Angew. Chem. Int. Ed. 2015 ¹
	About 4.7-fold	27	2 nM	5 min	Living cells and Zebrafish	Chem. Sci., 2018 ²
	7.3-fold	270	30 nM	180 min	Living cells and mouse	Chem. Commun., 2019 . ³
	20.3-fold	178	1 nM	1min	Living cells and organism	This work

1. Materials and instruments

2-Benzoylpyridine, salicylaldehyde, ammonium acetate, glacial acetic acid, thiosalicylic acid, benzoyl chloride, NaHCO_{3.} Na_2CO_3 N,N'-Dicyclohexylcarbodiimide, 4-Dimethylaminopyridine, and hexadecyl trimethyl ammonium bromide (CTAB) are purchased from Bidepharm (Shanghai, China) and utilized without further purification. The commercial lysosomal dye (LysoRed, KGMP006) and the commercial mitochondria dye (MitoGreen, KGMP0072) were obtained from KeyGen Biotech (Nanjing, China). LPS are purchased from Shanghai yuanye Bio-Technology Co., Ltd (Shanghai, China). Thin-layer chromatography (TLC) on 0.25 mm silica gel plates with a fluorescent indicator (GF254) visualized under UV light was used to monitor the reactions. Distilled water was prepared through a water purification system. ¹H NMR and ¹³C NMR spectra were recorded using a Bruker AV-500 instrument (Bruker, USA) with tetramethylsilane (TMS) as an internal standard. Highresolution mass spectra (HRMS) were performed on an LCMS-2010A instrument (Shimadzu, Japan). High-performance liquid chromatography (HPLC) analysis was used the Agilent 1260. UV-vis and fluorescence spectra were recorded on a Shimadzu UV-Vis spectrophotometer, UV-2550 and Shimadzu Instruments Spectrofluorometer, RF-6000, respectively.

2. General procedure for fluorometric analysis

All commercial chemicals and solvents are reagent grade and were used without further treatment unless otherwise noted. In *vitro* spectroscopy of the probe **PP-PS** (10 μ M) was performed in phosphate buffered saline (PBS, 10 mM, pH 7.4, containing 1 mM CTAB). In this work, CTAB was employed in the detection buffer to accelerate the response. The acceleration effect of CTAB is similar with that of previous reports which may be resulted from two reasons: first, CTAB can increase the solubility of PP-PS; second, the cationic CTAB may absorb the polysulfide anion to facilitate the reaction ^{4,5}. After incubating the test solution for 30 minutes at room temperature, all fluorescence was measured at an excitation wavelength of 300 nm (slit width: 5 nm / 5 nm) unless otherwise specified. Besides, Na₂S₂ and various other analytes were dissolved in ultrapure water to prepare a standard solution.

3 Cell cytotoxicity evaluated by MTT assay

A549 cells (adenocarcinomic human alveolar basal epithelial cells), MCF-7 cells (human breast adenocarcinoma cells), and U87 cell (human glioma cells) were purchased from American Type Culture Collection (ATCC, USA). The cell cytotoxicity was evaluated in three cell lines (A549, MCF-7, and U87 cells) by an MTT assay. The cells were seeded into 96-well cell culture plates (1×10^{5} /well) and subsequently incubated in CO₂ culture box for 24 h. The cells were retained at 37 °C under 5 % CO₂ for 24 h after incubation with **PP-PS** at different concentrations (5 µM, 10 µM, 20 µM, 50 µM, 100 µM, 250 µM, 500 µM, 750 µM and 1000 µM). And then the cells were incubated with MTT solution (20μ L, 5.0 mg mL^{-1}) for another 4 h. DMSO (150μ L/well) was further incubated with cells to dissolve the

precipitated formazan violet crystals at 37 °C for 20 min. The absorbance was measured at 490 nm by a multi-detection microplate reader. The following formula was used to calculate the viability of cell growth: viability (%) = (mean absorbance of test wells - mean absorbance of medium control wells) / (mean absorbance of untreated wells - mean absorbance of medium control well) \times 100 %.

4. Fluorescence quantum yield measurement

The quantum yield of the compound was calculated according to the equation:

$$\Phi_{\mathbf{u}} = \Phi_s \frac{I_u}{I_s} \times \frac{A_s}{A_u} \times \left(\frac{n_u}{n_s}\right)^2$$

 Φ denotes the quantum yield; *I* denotes the area under the fluorescence curve; *A* denotes the absorbance at the excitation wavelength; *n* denotes the refractive index of the solvent. Quinine sulfate ($\Phi = 0.542$ in 0.05 M sulfuric acid solution) was used as the reference standard.

5. The limit of detection measurement

The detection limit of **PP-PS** towards Na_2S_2 was determined through fluorescence titration, which was measured 5 times. The standard deviation of the blank solution was also measured for 20 times. After the linear slope of fluorescence intensity vs. concentrations of Na_2S_2 was obtained, the detection of limit (LOD) was calculated by following equation:

$LOD = 3\sigma / \kappa$

Where σ denotes the standard deviation of blank measurement, κ denotes the slope of thefluorescenceintensityvs.Na2S2concentrations.

6. ¹H NMR spectrum of PP-OH



Fig. S1 ¹H NMR spectrum of PP-OH.

7. ¹³C NMR spectrum of PP-OH



Fig. S2 ¹³C NMR spectrum of PP-OH.





Fig. S3 HRMS spectrum of PP-OH.



9. ¹H NMR spectrum of 2-Benzoylsulfanyl-benzoic acid

Fig. S4 ¹H NMR spectrum of 2-Benzoylsulfanyl-benzoic acid.

10. ¹H NMR spectrum of PP-PS



11. ¹³C NMR spectrum of PP-PS



Fig. S6 ¹³C NMR spectrum of **PP-PS**.





Fig. S7 HRMS spectrum of PP-PS.

13. The HPLC purity of PP-PS.



Peak	Ret. Time	Тур	Peak With	Peak Area	Peak Height	Peak Area
#	[min]	e	[min]	[mAU*s]	[mAU]	%
1	7.021	BB	0.1636	17.31207	1.71734	0.2497
2	8.967	VV	0.1919	6914.70264	544.37622	99.7503
Total				6932.01471	546.09356	

Signal 1: VWD1 A, Wavelength = 254 nm

Fig. S8 The HPLC purity of **PP-PS**.

14. Linear regression of the fluorescence intensity of PP-PS against the concentration of

 Na_2S_2



Fig. S9 Fluorescence intensity changes of PP-PS (10 μ M) at 478 nm against concentration of Na₂S₂ from 0 to 20 μ M.

15. The effects on pH research for detection of H_2S_n .



Fig. S10 Fluorescence intensity changes of the probe PP-PS (10 $\mu M)$ at different pH values in the absence

(black) or presence (blue) of Na_2S_2 (50 μ M).

16. Cell images of H_2S_n in living MCF-7 cells



Fig. S11 Cell images of H_2S_n in living MCF-7 cells. (a1-a3) cells incubated with **PP-PS** (10 μ M) for 20 min at 37 °C; (b1-b3) cells pretreated with 1mM NEM for 30 min and then incubated with **PP-PS** (10 μ M) for 20 min at 37 °C; (c1-c3) cells pretreated with 15 μ M Na₂S₂ for 20 min and then incubated with **PP-PS** (10 μ M) for 20 min at 37 °C; (d1-d3) cells preincubated with 1 μ g/mL LPS for 14 h and incubated with **PP-PS** (10 μ M) for 20 min at 37 °C.



17. Cell images of H_2S_n in living U87 cells

Fig. S12 Cell images of H_2S_n in living U87 cells. (a1-a3) cells incubated with **PP-PS** (10 µM) for 20 min at 37 °C; (b1-b3) cells pretreated with 1mM NEM for 30 min and then incubated with **PP-PS** (10 µM) for 20 min at 37 °C; (c1-c3) cells pretreated with 15 µM Na₂S₂ for 20 min and then incubated with **PP-PS** (10 µM) for 20 min at 37 °C; (d1-d3) cells preincubated with 1 µg/mL LPS for 14 h and incubated with **PP-PS** (10 µM) for 20 min at 37 °C.

18. Detection of H_2S_n in mitochondria



Fig. S13 Cell images of A549 cells treated with LPS (1 μ g/mL), **PP-PS** (10 μ M) and MitoGreen (1 μ M). The cells were pretreated with 1 μ g/mL LPS for 14 h, and then incubated with MitoGreen (1 μ M) for 30 min, and **PP-PS** (10 μ M) for another 20 min. (a) cells imaging merged green and blue channels; (b) cells imaging of **PP-PS** with blue channel; (c) cells imaging of MitoGreen with green channel; (d) cells imaging of bright field; (e) cells imaging merged green, blue channels and bright field; (f) the colocalization areas of the images of b and c.

Notes and references

1 W. Chen, E. W. Rosser, T. Matsunaga, A. Pacheco, T. Akaike and M. Xian, *Angew. Chem. Int. Ed. Engl.*, 2015, **54**, 13961–13965.

2 F. Yang, H. Gao, S.-S. Li, R.-B. An, X.-Y. Sun, B. Kang, J.-J. Xu and H.-Y. Chen, *Chemical Science*, 2018, **9**, 5556–5563.

3 J.-B. Li, Q. Wang, H.-W. Liu, L. Yuan and X.-B. Zhang, *Chem. Commun. (Camb.)*, 2019, **55**, 4487–4490.

4 W. Li, S. Zhou, L. Zhang, Z. Yang, H. Chen, W. Chen, J. Qin, X. Shen and S. Zhao, *Sensors and Actuators B: Chemical*, 2019, **284**, 30–35.

5 W. Chen, E. W. Rosser, D. Zhang, W. Shi, Y. Li, W.-J. Dong, H. Ma, D. Hu and M. Xian, *Org. Lett.*, 2015, **17**, 2776–2779.