
A Catalytic Asymmetric One-Pot [3+2] Cyclization/Semipinacol Rearrangement Sequence: An Efficient Construction of Multi-Substituted 3*H*-Spiro[benzofuran-2,1'-cyclopentane] Skeleton

Lin Liu,^a Lin-Sheng Lei,^a Zong-Song Zhan,^a Si-Zhan Liu,^a Yu-Xiao Wang,^a Yong-Qiang Tu,^{*a} Fu-Min Zhang,^a Xiao-Ming Zhang,^a Ai-Jun Ma,^b Shao-Hua Wang^{*ab}

^aState Key Laboratory of Applied Organic Chemistry & School of Pharmacy, Lanzhou University, Lanzhou 730000 (P. R. China)

^bSchool of Biotechnology and Health Science, Wuyi University, Jiangmen 529020 (P. R. China)

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1. General Information:

All reactions under standard conditions were monitored by thin-layer chromatography (TLC) on silica gel F254 plates. Column chromatography was performed on silica gel (200-300 meshes) or neutral alumina (200-300 meshes). Solvents for reaction were distilled prior to use, and all air- or moisture-sensitive reactions were conducted under an argon atmosphere. The melting points were measured using micro melting point apparatus. ^1H NMR and ^{13}C NMR spectra were recorded in CDCl_3 solution, C_6D_6 solution or $\text{Acetone-}d_6$ solution on instruments (400 MHz for ^1H NMR and 100 MHz for ^{13}C NMR) and spectra data were reported in ppm relative to tetramethylsilane (TMS) as internal standard. High-resolution mass analysis (HRMS) data were measured by means of the ESI technique on Fourier transform ion cyclotron resonance mass analyzer. Ee values were determined by high performance liquid chromatography (HPLC) analysis employing Darcel Chiracel IF-3 columns. Optical rotation was measured using a 0.1-mL cell with a 1-cm path length by the Perkin Elmer 341 polarimeter.

2. Optimization of Reaction Conditions

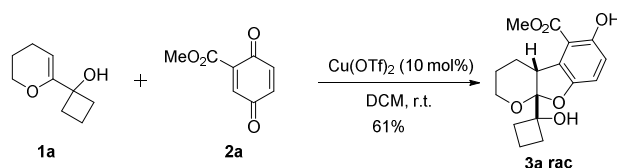
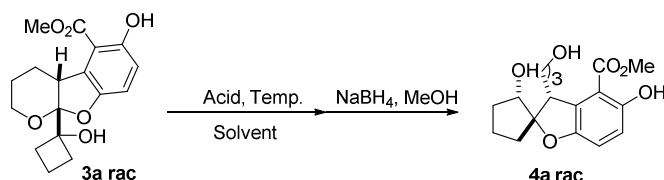


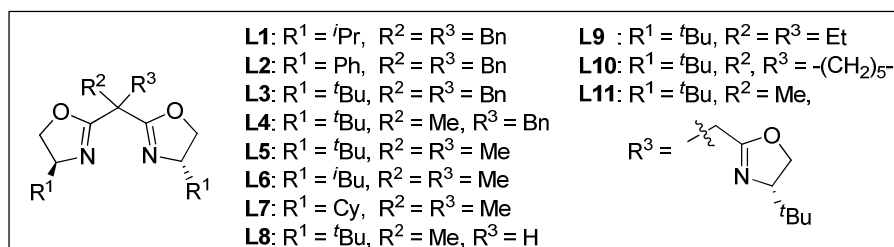
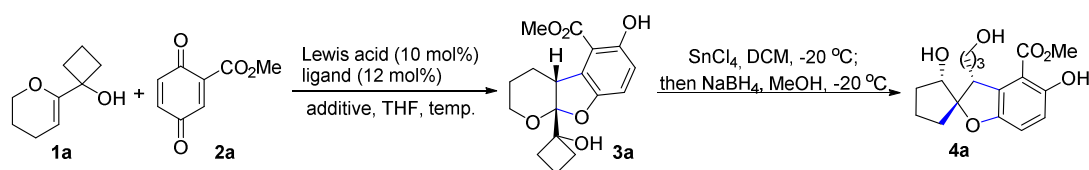
Table 1. Optimization of Semipinacol Rearrangement Reaction Conditions^a



Acid	Temp.	solvent	yield ^b	dr ^c
$\text{Cu}(\text{OTf})_2$	r.t.	DCM	NR	---
PTS	r.t.	DCM	NR	---
TfOH	0 °C	DCM	51%	4.0:1
SnCl_4	0 °C	DCM	66%	5.5:1
TMSOTf	0 °C	DCM	68%	4.5:1
BF_3OEt_2	0 °C	DCM	68%	2.2:1
SnCl_4	-20 °C	DCM	81%	7.2:1
SnCl_4	-40 °C	DCM	76%	7.2:1

^aUnless specified, reaction was conducted in DCM (4 mL) using **3a-rac** (0.1 mmol, 1.0 equiv), Lewis acid (2.0 equiv.) at the indicated temperature; ^bIsolated yields; ^cdr was determined by ^1H NMR.

Table 2. Optimization of the One-Pot Asymmetric Reaction Conditions^a



Entry	Lewis acid	Ligand	Additive	Product	Temp.	Yield ^b (%)	ee ^c (%)
1	Cu(OTf) ₂	-	-	3a	r.t.	61	-
2	Cu(OTf) ₂	L1	-	3a	-20 °C	57	3
3	Cu(OTf) ₂	L2	-	3a	-20 °C	67	38
4	Cu(OTf) ₂	L3	-	3a	-20 °C	47	41
5	Cu(OTf) ₂	L3	-	3a	-78 °C	41	57
6	Cu(OTf) ₂	L4	-	3a	-78 °C	45	61
7	Cu(OTf) ₂	L5	-	3a	-78 °C	37	81
8	Cu(BF ₄)·6H ₂ O	L5	-	3a	-78 °C	61	67
9	Cu(SbF ₆) ₂	L5	-	3a	-78 °C	78	7
10	Cu(PF ₆) ₂	L5	-	3a	-78 °C	51	12
11	Sc(OTf) ₃	L5	-	3a	-78 °C	91	2
12	Zn(OTf) ₂	L5	-	3a	-78 °C	60	20
13	Ni(ClO ₄) ₂ ·6H ₂ O	L5	-	3a	-78 °C	20	1
14	Cu(ClO ₄) ₂ ·6H ₂ O	L5	-	3a	-78 °C	41	87
15	Cu(ClO ₄) ₂ ·6H ₂ O	L6	-	3a	-78 °C	41	60
16	Cu(ClO ₄) ₂ ·6H ₂ O	L7	-	3a	-78 °C	67	77
17	Cu(ClO ₄) ₂ ·6H ₂ O	L8	-	3a	-78 °C	35	9
18	Cu(ClO ₄) ₂ ·6H ₂ O	L9	-	3a	-78 °C	51	54
19	Cu(ClO ₄) ₂ ·6H ₂ O	L10	-	3a	-78 °C	61	76
20	Cu(ClO ₄) ₂ ·6H ₂ O	L11	-	3a	-78 °C	31	11
21 ^d	Cu(ClO ₄) ₂ ·6H ₂ O	L5	3 Å MS	3a	-78 °C	60	86
22 ^e	Cu(ClO ₄) ₂ ·6H ₂ O	L5	4 Å MS	3a	-78 °C	55	87
23 ^f	Cu(ClO ₄) ₂ ·6H ₂ O	L5	5 Å MS	3a	-78 °C	87	91
24 ^{fg}	Cu(ClO ₄) ₂ ·6H ₂ O	L5	5 Å MS	3a	-78 °C	85	89
25 ^{fh}	Cu(ClO ₄) ₂ ·6H ₂ O	L5	5 Å MS	4a	-78 °C	61	91

^aUnless specified, reaction was conducted (also see supporting information) in THF (1.5 mL) using **1a** (0.1 mmol, 1.0 equiv), **2a** (0.15 mmol, 1.5 equiv), Lewis acid (10 mol%), ligand (12 mol%) at the indicated temperature;

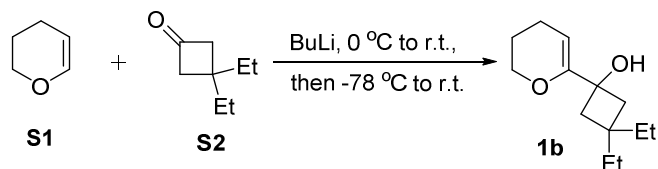
^bIsolated yields; ^cDetermined by chiral HPLC; ^d3 Å MS (50 mg) was added; ^e4 Å MS (50 mg) was added; ^f5 Å MS (50 mg) was added; ^gLewis acid (5 mol%), ligand (6 mol%) was used; ^hdr = 7.2:1 (determined by ¹H NMR).

3. Syntheses of the Substrates 1 and Spectroscopic Data of Substrates

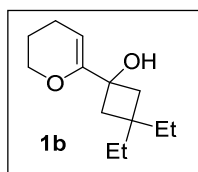
1a was synthesized according to a known procedure.^[1]

General procedure A: Allylic tertiary alcohol from enol-ethers (**1b** as an example):

3.1 Synthesis of the Substrate 1b



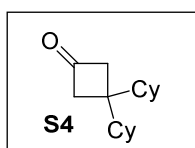
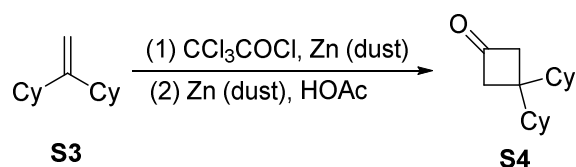
To a solution of 3,4-dihydro-2H-pyran **S1** (1 mL, 10.9 mmol, 1.5 equiv.) in anhydrous THF (15 mL) at 0 °C was added n-butyllithium (5.6 mL, 1.6 M in hexane, 9.1 mmol, 1.25 equiv.) over 10 minutes. The mixture was allowed to warm up to room temperature and stirred for a further 3 h, before it was cooled down to -78 °C. The substituted cyclobutanone **S2** (0.91 g, 7.2 mmol, 1 equiv.) was then added dropwise into the solution over 15 minutes. The reaction was then allowed to warm slowly to room temperature and then stirred for a further 2 h before quenched with saturated NaHCO₃ solution and extracted with EtOAc, the combined organic layer was washed with brine and dried over MgSO₄ before concentrated in vacuo. The product was purified by flash chromatography using neutral alumina to afford the desired product **1b** as a colorless oil (0.622 g, 2.95 mmol, 41%).



¹H NMR (400 MHz, Acetone-*d*₆) δ 4.81 (t, *J* = 3.6 Hz, 1H), 3.97 (t, *J* = 5.2 Hz, 2H), 3.75 (s, 1H), 2.82 (s, 2H), 2.13-2.10 (m, 2H), 1.78-1.60 (m, 6H), 1.41 (q, *J* = 7.2 Hz, 2H), 0.75 (t, *J* = 7.6 Hz, 3H), 0.70 (t, *J* = 7.6 Hz, 3H);

¹³C NMR (100 MHz, Acetone-*d*₆) δ 158.8, 93.7, 70.5, 66.6, 43.6, 34.8, 30.3, 23.1, 20.6, 8.2, 8.1;

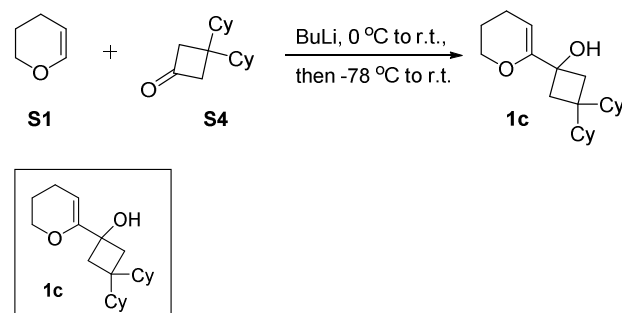
3.2 Synthesis of the Substrates S4 and 1c



To a 100 mL three-neck round bottomed flask were added 1,1-dicyclohexylethene **S3** (4.0 g, 20.8 mmol, 1 equiv.), Zn dust (1.76 g, 27.1 mmol, 1.3 equiv) and Et₂O (40 mL). Next, a solution of trichloroacetic chloride (3.04 mL, 27.1 mmol, 1.3 equiv) in Et₂O (10 mL) was added dropwise to the above suspension. This mixture was sonicated at room temperature for 3 h, and the solid was removed by filtration and the ether solution was washed with NH₄Cl, saturated NaHCO₃ and brine. Evaporation of the resulting solution dried over MgSO₄ gave a pale yellow liquid, which was directly used for the next step. A solution of the crude intermediate mentioned above in HOAc (25 mL) was added dropwise to a vigorously stirred suspension of zinc dust (5.41 g, 83.2 mmol, 4 equiv) in HOAc (25 mL) at 0 °C. After the addition, the reaction mixture was heated at 70 °C for 24 h, and cooled down to room temperature. The acetic acid was then removed by rotavapor. The residue was dissolved in Et₂O (50 mL) and then poured into a separation funnel containing H₂O (50 mL) and Et₂O (200 mL). The organic layer was washed with H₂O, saturated NaHCO₃ and brine, and dried over MgSO₄. Then the solution was concentrated followed by purification with flash chromatography giving the substituted cyclobutanone **S4** as a colorless oil (1.460 g, 6.24 mmol, 30%).

¹H NMR (400 MHz, CDCl₃) δ 2.72 (s, 4H), 1.72 (t, *J* = 12.8 Hz, 8H), 1.65-1.57 (m, 4H), 1.23-1.04 (m, 6H), 0.95-0.85 (m, 4H);

¹³C NMR (100 MHz, CDCl₃) δ 209.0, 49.2, 41.5, 38.2, 27.9, 26.7, 26.3.



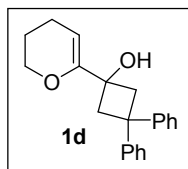
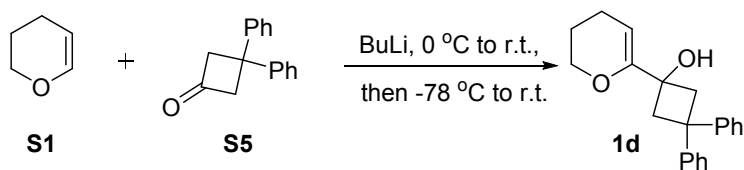
Preparation according to the general procedure A from **S4** (0.780 g, 3.33 mmol) afforded **1c** as an amorphous solid (0.413 g, 1.30 mmol, 39% yield).

¹H NMR (400 MHz, Acetone-*d*₆) δ 4.79 (t, *J* = 5.2 Hz, 1H), 3.97 (t, *J* = 5.2 Hz, 2H), 3.61 (s, 1H), 2.67-2.61 (m, 1H), 2.44-2.43 (m, 1H), 2.29-2.26 (dd, *J* = 2.8 Hz, 10.8 Hz, 2H), 2.00 (dt, *J* = 4.0 Hz, 6.4 Hz, 2H), 1.90-1.87 (m, 1H), 1.86-1.84 (m, 1H), 1.78-1.72 (m, 8H), 1.68-1.56 (m, 6H), 1.24-1.01 (m, 8H);

¹³C NMR (100 MHz, Acetone-*d*₆) δ 159.0, 94.3, 70.3, 66.6, 39.8, 39.1, 30.3, 29.7, 29.6, 29.1, 28.2, 28.1, 27.5, 27.4, 23.1, 20.7;

HRMS ESI Calcd for C₂₁H₃₄O₂Na [M+Na]⁺: 341.2451, Found: 341.2451.

3.3 Synthesis of the Substrate 1d



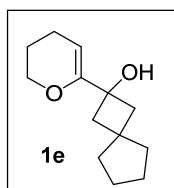
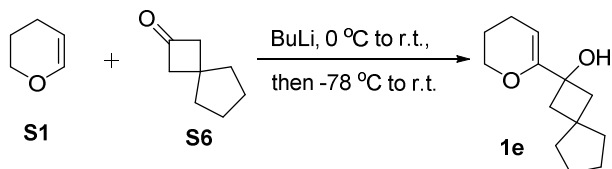
Preparation according to the general procedure A from **S5** (0.570 g, 2.57 mmol) afforded **1d** as an amorphous solid (0.401 g, 1.31 mmol, 51% yield).

^1H NMR (400 MHz, Acetone- d_6) δ 7.41 (dd, J = 0.8 Hz, 8.4 Hz, 2H), 7.35 (dd, J = 0.8 Hz, 8.4 Hz, 2H), 7.26-7.21 (m, 4H), 7.09-7.04 (m, 2H), 4.75 (t, J = 3.6 Hz, 1H), 3.96 (s, 1H), 3.84 (t, J = 5.2 Hz, 2H), 3.34 (d, J = 12.8 Hz, 2H), 2.88 (d, J = 12.8 Hz, 2H), 1.86 (dt, J = 3.6 Hz, 6.4 Hz, 2H), 1.66-1.60 (m, 2H);

^{13}C NMR (100 MHz, Acetone- d_6) δ 156.1, 150.8, 149.9, 127.7, 127.6, 126.0, 125.9, 124.7, 97.4, 70.2, 65.5, 46.2, 43.2, 21.8, 19.5;

HRMS ESI Calcd for $\text{C}_{21}\text{H}_{22}\text{O}_2\text{Na}$ $[\text{M}+\text{Na}]^+$: 329.1512, Found: 329.1502.

3.4 Synthesis of the Substrate **1e**



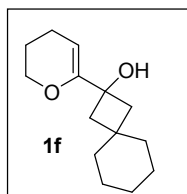
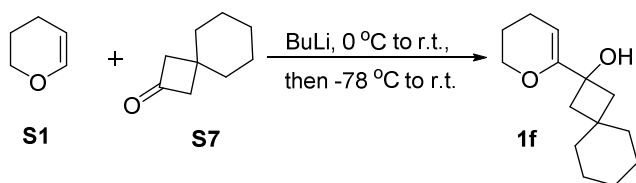
Preparation according to the general procedure A from **S6** (0.901 g, 7.25 mmol) afforded **1e** as an amorphous solid (0.628 g, 3.02 mmol, 42% yield).

^1H NMR (400 MHz, C_6D_6) δ 4.81 (t, J = 4.0 Hz, 1H), 3.70 (t, J = 5.2 Hz, 2H), 2.48 (dd, J = 2.8 Hz, 10.0 Hz, 2H), 2.10 (dd, J = 2.8 Hz, 10.0 Hz, 2H), 2.03 (s, 1H), 1.81-1.76 (m, 4H), 1.60 (t, J = 6.4 Hz, 2H), 1.54-1.49 (m, 4H), 1.43-1.37 (m, 2H);

^{13}C NMR (100 MHz, Acetone- d_6) δ 158.5, 94.1, 71.0, 66.7, 45.7, 41.6, 40.2, 38.8, 24.3, 23.1, 20.7;

HRMS ESI Calcd for $\text{C}_{13}\text{H}_{20}\text{O}_2\text{Na}$ $[\text{M}+\text{Na}]^+$: 231.1356, Found: 231.1345.

3.5 Synthesis of the Substrate **1f**



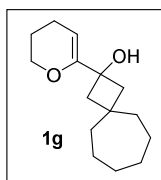
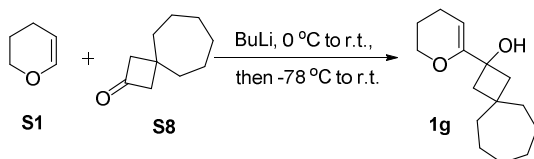
Preparation according to the general procedure A from **S7** (0.99 g, 7.2 mmol) afforded **1f** as an amorphous solid (0.590 g, 2.66 mmol, 37% yield).

¹H NMR (400 MHz, Actone-*d*₆) δ 4.83 (t, *J* = 3.6 Hz, 1H), 3.98 (t, *J* = 4.8 Hz, 2H), 3.79 (s, 1H), 2.84 (s, 1H), 2.16 (d, *J* = 12.8 Hz, 2H), 2.07 (s, 1H), 1.80-1.70 (m, 4H), 1.68-1.53 (m, 2H), 1.42-1.25 (m, 8H);

¹³C NMR (100 MHz, Actone-*d*₆) δ 158.7, 93.7, 70.7, 66.6, 44.9, 39.9, 38.9, 31.8, 26.5, 23.6, 23.3, 23.1, 20.6;

HRMS ESI Calcd for C₁₄H₂₂O₂Na [M+Na]⁺: 245.1512, Found: 245.1500.

3.6 Synthesis of the Substrate **1g**



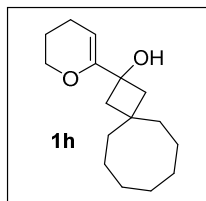
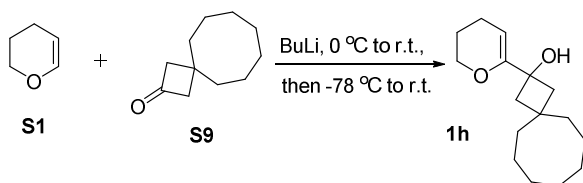
Preparation according to the general procedure A from **S8** (0.77 g, 5.1 mmol) afforded **1g** as an amorphous solid (0.550 g, 2.32 mmol, 46% yield).

¹H NMR (400 MHz, Actone-*d*₆) δ 4.81 (t, *J* = 3.6 Hz, 1H), 3.97 (t, *J* = 5.2 Hz, 2H), 3.77 (s, 1H), 2.18 (dd, *J* = 2.8 Hz, 10.0 Hz, 2H), 2.00 (dt, *J* = 3.6 Hz, 6.4 Hz, 2H), 1.82-1.77 (m, 2H), 1.76-1.74 (m, 2H), 1.73-1.71 (m, 2H), 1.60-1.56 (m, 2H), 1.52-1.47 (m, 6H), 1.47-1.41 (m, 2H);

¹³C NMR (100 MHz, Actone-*d*₆) δ 158.7, 93.7, 70.5, 66.6, 45.8, 43.3, 42.1, 34.6, 23.5, 23.4, 23.1, 20.6;

HRMS ESI Calcd for C₁₅H₂₄O₂Na [M+Na]⁺: 259.1669, Found: 259.1658.

3.7 Synthesis of the Substrate **1h**



Preparation according to the general procedure A from **S9** (1.12 g, 7.2 mmol) afforded **1h** as an amorphous solid (0.648 g, 2.58 mmol, 36% yield).

¹H NMR (400 MHz, Acetone-*d*₆) δ 4.82 (t, *J* = 3.6 Hz, 1H), 3.98 (t, *J* = 1.2 Hz, 2H), 3.77 (s, 1H), 2.83 (s, 2H), 2.15 (d, *J* = 13.2 Hz, 2H), 2.04-1.99 (m, 2H), 1.88-1.77 (m, 2H), 1.77-1.72 (m, 1H), 1.69 (d, *J* = 15.6 Hz, 1H), 1.64-1.62 (m, 2H), 1.60-1.30 (m, 10H);

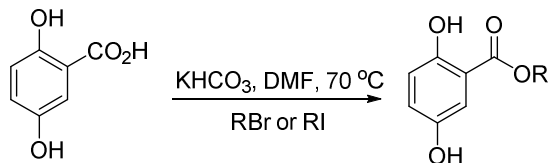
¹³C NMR (100 MHz, Acetone-*d*₆) δ 158.8, 93.7, 70.5, 66.6, 45.8, 38.1, 37.2, 34.2, 24.7, 23.2, 23.08, 22.99, 20.6;

HRMS ESI Calcd for C₁₆H₂₆O₂Na [M+Na]⁺: 273.1825, Found: 273.1816.

4. Syntheses of 1,4-dihydroxybenzene-2-carboxylates and the substrates 2 and spectroscopic data of substrates

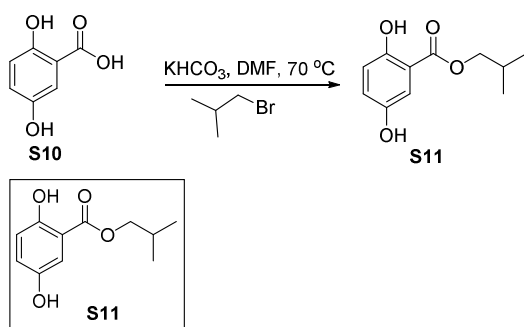
Substrates **2a**, **2k**, **2l** and **2q** were synthesized according to known literatures.^[2]

General procedure B: Synthesis of 1,4-dihydroxybenzene-2-carboxylate from 2,5-dihydroxybenzoic acid.



Solid KHCO₃ (2.619 g, 26.2 mmol, 2.7 equiv.) was added to a stirred mixture of 2,5-dihydroxybenzoic acid (1.50 g, 9.7 mmol, 1 equiv.) and RBr or RI (38.8 mmol, 4.0 equiv.) in DMF (30 mL). The mixture was heated to 70 °C and stirred for 5-24 h. The reaction mixture was cooled down to room temperature, diluted with 1 M HCl and extracted with EtOAc. The organic phases were washed with water and dried over MgSO₄. Then the solution was filtered, and concentrated followed by purification with flash chromatography giving the desired 1,4-dihydroxybenzene-2-carboxylate.

4.1 Synthesis of the Substrate S11



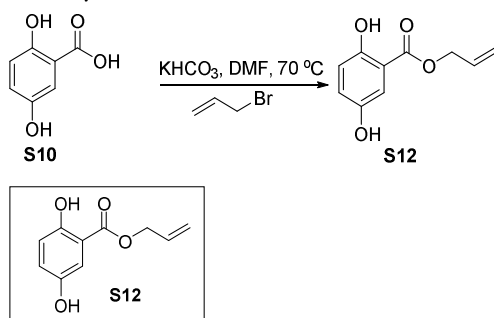
Preparation according to the general procedure B from **S10** (1.50 g, 9.7 mmol, 1 equiv.) and isobutyl bromide (4.2 mL, 38.8 mmol, 4 equiv.) afforded **S11** as a colorless oil (1.772 g, 8.43 mmol, 87% yield).

^1H NMR (400 MHz, CDCl_3) δ 10.38 (s, 1H), 7.32 (d, $J = 3.2$ Hz, 1H), 7.03 (dd, $J = 3.2$ Hz, 8.8 Hz, 1H), 6.84 (d, $J = 8.8$ Hz, 1H), 6.57 (s, 1H), 4.10 (d, $J = 6.8$ Hz, 2H), 2.93 (d, $J = 28$ Hz, 3H), 2.11-2.02 (m, 1H), 1.00 (d, $J = 6.8$ Hz, 6H);

^{13}C NMR (100 MHz, CDCl_3) δ 169.9, 155.4, 148.3, 124.0, 118.2, 114.7, 112.4, 71.3, 36.8, 27.7, 27.7, 19.0.

HRMS ESI Calcd for $\text{C}_{11}\text{H}_{14}\text{O}_4\text{Na}$ $[\text{M}+\text{Na}]^+$: 233.0784, Found: 233.0778.

4.2 Synthesis of the Substrate S12

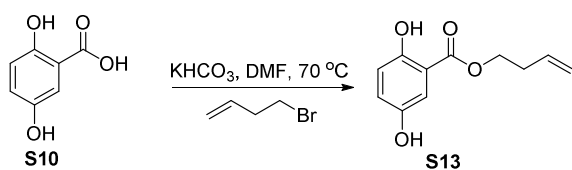


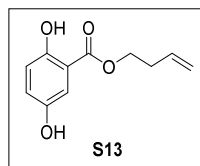
Preparation according to the general procedure B from **S10** (1.50 g, 9.7 mmol, 1 equiv.) and allyl bromide (2.85 mL, 38.8 mmol, 4 equiv.) afforded **S12** as an amorphous solid (1.787 g, 9.21 mmol, 95% yield).

^1H NMR (400 MHz, CDCl_3) δ 10.38 (s, 1H), 7.32 (d, $J = 3.2$ Hz, 1H), 7.02 (dd, $J = 3.2$ Hz, 8.8 Hz, 1H), 6.87 (d, $J = 8.8$ Hz, 1H), 6.02-5.95 (m, 1H), 5.41 (dd, $J = 1.2$ Hz, 17.2 Hz, 1H), 5.30 (dd, $J = 0.8$ Hz, 10.4 Hz, 1H), 4.81 (d, $J = 5.6$ Hz, 2H);

^{13}C NMR (100 MHz, CDCl_3) δ 169.4, 155.6, 147.8, 131.3, 124.2, 119.0, 118.5, 114.9, 112.2, 66.0.

4.3 Synthesis of the Substrate S13



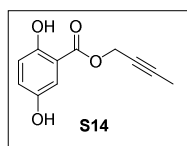
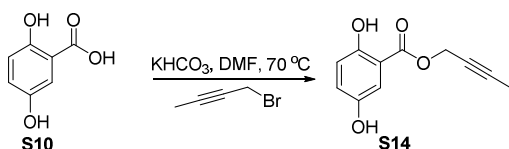


Preparation according to the general procedure B from **S10** (1.5 g, 9.7 mmol, 1 equiv.) and 4-bromo-1-butene (3.9 mL, 38.8 mmol, 4equiv.) afforded **S13** as an amorphous solid (1.836 g, 8.83 mmol, 91% yield).

¹H NMR (400 MHz, CDCl₃) δ 10.41 (s, 1H), 7.29 (d, *J* = 3.2Hz, 1H), 7.05 (s, 1H), 6.84 (d, *J* = 1.2 Hz, 2H), 5.82-5.89 (m, 1H), 5.17-5.11 (m, 2H), 4.38-4.34 (m, 2H), 2.52-2.49 (m, 2H);

¹³C NMR (100 MHz, CDCl₃) δ 169.7, 155.4, 147.9, 133.4, 124.1, 118.3, 117.7, 114.8, 112.3, 64.4, 32.8.

4.4 Synthesis of the Substrate S14



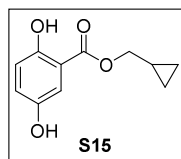
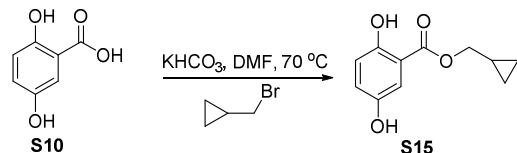
Preparation according to the general procedure B from **S10** (1.5 g, 9.7 mmol, 1 equiv.) and 1-bromo-2-butyne (3.34 mL, 38.8 mmol, 4 equiv.) afforded **S14** as an amorphous solid (1.758 g, 8.53 mmol, 88% yield).

¹H NMR (400 MHz, CDCl₃) δ 10.18 (s, 1H), 7.32 (d, *J* = 3.2 Hz, 1H), 7.03 (dd, *J* = 3.2 Hz, 5.2 Hz, 1H), 6.87 (d, *J* = 8.8 Hz, 1H), 4.91-4.86 (m, 3H), 1.88 (t, *J* = 2.4 Hz, 3H);

¹³C NMR (100 MHz, CDCl₃) δ 169.1, 155.9, 147.8, 124.4, 118.6, 114.9, 111.9, 84.1, 72.6, 53.7, 3.6;

HRMS ESI Calcd for C₁₁H₁₀O₄Na [M+Na]⁺: 229.0471, Found: 229.0463.

4.5 Synthesis of the Substrate S15



Preparation according to the general procedure B from **S10** (1.5 g, 9.7 mmol, 1 equiv.) and

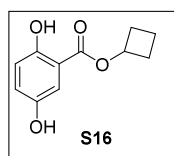
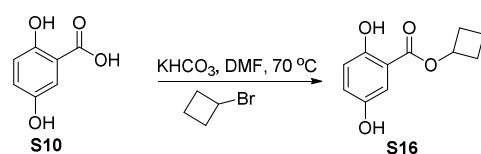
(bromomethyl) cyclopropane (3.76 mL, 38.8 mmol, 4 equiv.) afforded **S15** as a colorless oil (1.896 g, 9.11 mmol, 94% yield).

¹H NMR (400 MHz, CDCl₃) δ 10.40 (d, *J* = 2.8 Hz, 1H), 7.35 (d, *J* = 3.2 Hz, 1H), 7.02 (dd, *J* = 3.2 Hz, 8.8 Hz, 1H), 6.86 (d, *J* = 8.8 Hz, 1H), 5.90-5.20 (m, 1H), 4.16 (d, *J* = 7.2 Hz, 2H), 1.28-1.22 (m, 1H), 0.65-0.61 (m, 2H), 0.38-0.35 (m, 2H);

¹³C NMR (100 MHz, CDCl₃) δ 169.9, 155.7, 147.9, 124.0, 118.4, 114.9, 112.5, 70.2, 9.7, 3.4;

HRMS ESI Calcd for C₁₁H₁₂O₄Na [M+Na]⁺: 231.0628, Found: 231.0626.

4.6 Synthesis of the Substrate S16



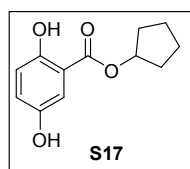
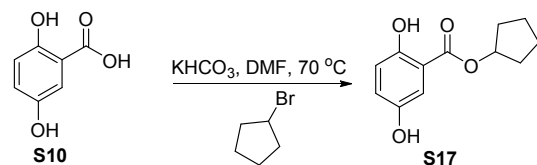
Preparation according to the general procedure B from **S10** (1.5 g, 9.7 mmol, 1 equiv.) and cyclobutyl bromide (3.65 mL, 38.8 mmol, 4 equiv.) afforded **S16** as a colorless oil (1.532 g, 7.37 mmol, 76% yield).

¹H NMR (400 MHz, CDCl₃) δ 10.40 (s, 1H), 7.31 (d, *J* = 3.2 Hz, 1H), 7.01 (dd, *J* = 3.2 Hz, 4.8 Hz, 1H), 6.86 (d, *J* = 8.8 Hz, 1H), 5.20 (t, *J* = 7.6 Hz, 1H), 2.48-2.41 (m, 2H), 2.38-2.18 (m, 2H), 1.92-1.84 (m, 1H), 1.74-1.67 (m, 1H);

¹³C NMR (100 MHz, CDCl₃) δ 169.2, 155.7, 147.8, 124.0, 118.4, 114.8, 112.3, 69.9, 30.2, 13.5.

HRMS ESI Calcd for C₁₁H₁₂O₄Na [M+Na]⁺: 231.0628, Found: 231.0630.

4.7 Synthesis of the Substrate S17



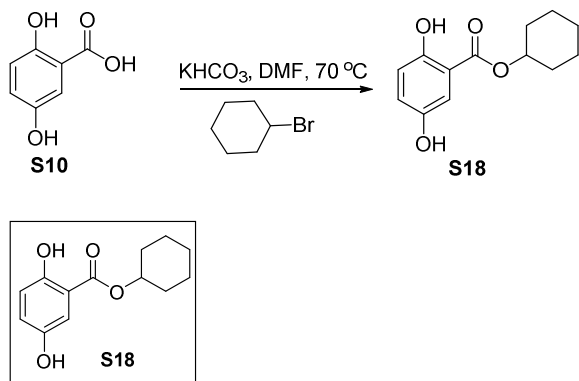
Preparation according to the general procedure B from **S10** (1.5 g, 9.7 mmol, 1 equiv.) and cyclopentyl bromide (4.20 mL, 38.8 mmol, 4 equiv.) afforded **S17** as a colorless oil (1.486 g, 6.69 mmol, 69% yield).

¹H NMR (400 MHz, CDCl₃) δ 10.50 (s, 1H), 7.27 (s, 1H), 7.01-6.98 (m, 1H), 6.88-6.86 (m, 1H), 5.42 (t, *J* = 2.8 Hz, 1H), 1.98-1.95 (m, 2H), 1.86-1.79 (m, 4H), 1.73-1.10 (m, 2H);

¹³C NMR (100 MHz, CDCl₃) δ 169.6, 155.6, 147.7, 123.8, 118.3, 114.80, 114.78, 112.8, 78.6, 32.7, 23.7;

HRMS ESI Calcd for C₁₂H₁₄O₄Na [M+Na]⁺: 245.0784, Found: 245.0788.

4.8 Synthesis of the Substrate S18



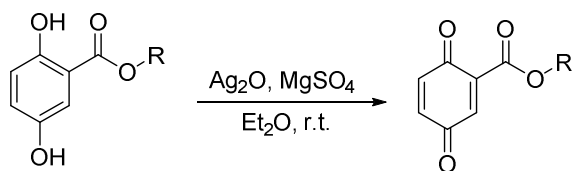
Preparation according to the general procedure B from **S10** (1.5 g, 9.7 mmol, 1 equiv.) and cyclohexyl bromide (4.77 mL, 38.8 mmol, 4 equiv.) afforded **S18** as a colorless oil (1.488 g, 6.30 mmol, 65% yield).

¹H NMR (400 MHz, CDCl₃) δ 10.49 (s, 1H), 7.31 (d, *J* = 3.2 Hz, 1H), 7.00 (dd, *J* = 3.2 Hz, 9.2 Hz, 1H), 6.88 (d, *J* = 8.8 Hz, 1H), 5.08-5.02 (m, 1H), 1.92-1.78 (m, 2H), 1.77-1.64 (m, 2H), 1.61-1.49 (m, 2H), 1.46-1.24 (m, 3H);

¹³C NMR (100 MHz, CDCl₃) δ 169.2, 155.9, 147.6, 123.7, 118.4, 114.8, 112.9, 73.9, 31.5, 25.3, 23.5;

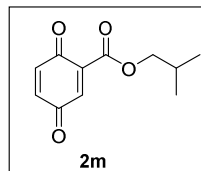
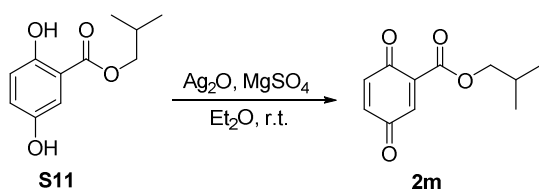
HRMS ESI Calcd for C₁₃H₁₆O₄Na [M+Na]⁺: 259.0941, Found: 259.0941.

General procedure C: Synthesis of 2-alkoxycarbonyl-1,4-benzoquinone from 1,4-dihydroxybenzene-2-carboxylate.



Silver oxide (1.392 g, 6.0 mmol, 3.0 equiv.) and magnesium sulfate (0.72 g, 6.0 mmol, 3.0 equiv.) were added to a solution of 1,4-dihydroxybenzene-2-carboxylate (2.0 mmol, 1 equiv.) in diethyl ether (20 mL). The reaction mixture was stirred at 25 °C for 1.5 h. After filtration, the filtrate was evaporated in vacuo to furnish the desired quinone **2**.

4.9 Synthesis of the Substrate 2m



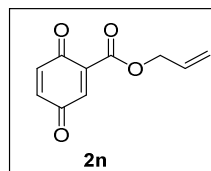
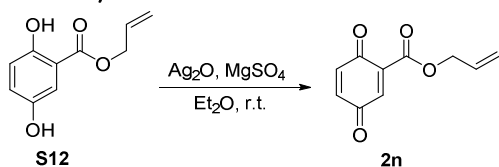
Preparation according to the general procedure C from **S11** (0.42 g, 2 mmol) afforded **2m** as a red oil (0.391 g, 1.88 mmol, 92% yield).

¹H NMR (400 MHz, CDCl₃) δ 7.05 (s, 1H), 6.79 (s, 2H), 4.05 (d, *J* = 6.8 Hz, 2H), 2.03-1.97 (m, 1H), 0.95 (d, *J* = 6.8 Hz, 6H);

¹³C NMR (100 MHz, CDCl₃) δ 186.9, 182.9, 162.6, 137.4, 136.8, 136.00, 135.96, 72.2, 27.6, 18.9;

HRMS ESI Calcd for C₁₁H₁₂O₄Na [M+Na]⁺: 231.0628, Found: 231.0626.

4.10 Synthesis of the Substrate 2n



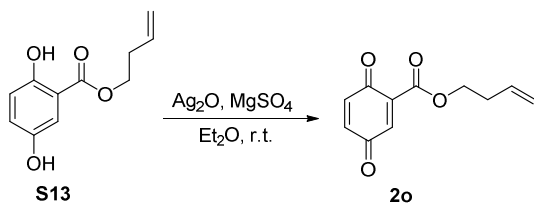
Preparation according to the general procedure C from **S12** (0.388 g, 2 mmol) afforded **2n** as a red oil (0.365 g, 1.9 mmol, 95% yield).

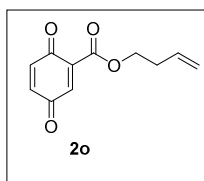
¹H NMR (400 MHz, CDCl₃) δ 7.10 (dd, *J* = 0.8 Hz, 1.6 Hz, 1H), 6.81 (d, *J* = 2.4 Hz, 2H), 5.99-5.91 (m, 1H), 5.43 (d, *J* = 1.6 Hz, 1H), 5.31 (dd, *J* = 0.8 Hz, 10.4 Hz, 1H), 4.79 (td, *J* = 1.2 Hz, 6.0 Hz, 2H);

¹³C NMR (100 MHz, CDCl₃) δ 186.8, 182.9, 162.3, 137.1, 136.9, 136.4, 136.1, 130.9, 119.5, 66.7;

HRMS ESI Calcd for C₁₀H₈O₄Na [M+Na]⁺: 215.0315, Found: 215.0312.

4.11 Synthesis of the Substrate 2o





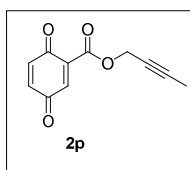
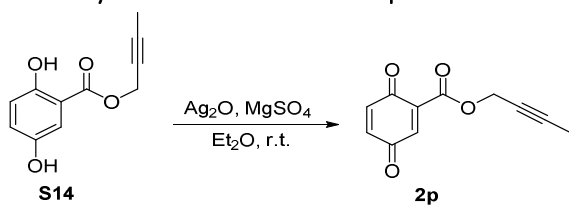
Preparation according to the general procedure C from **S13** (0.42 g, 2 mmol) afforded **2o** as a red oil (0.383 g, 1.86 mmol, 93% yield).

¹H NMR (400 MHz, CDCl₃) δ 7.05 (s, 1H), 6.85 (dd, *J* = 1.2 Hz, 8.8 Hz, 1H), 5.85-5.78 (m, 2H), 5.18-5.08 (m, 2H), 4.37 (dt, *J* = 2.0 Hz, 6.8 Hz, 2H), 2.52-2.47 (m, 2H);

¹³C NMR (100 MHz, CDCl₃) δ 186.9, 182.9, 162.5, 137.2, 136.86, 136.85, 136.1, 133.3, 117.8, 65.2, 32.8;

HRMS ESI Calcd for C₁₁H₁₀O₄Na [M+Na]⁺: 229.0471, Found: 229.0471.

4.12 Synthesis of the Substrate 2p



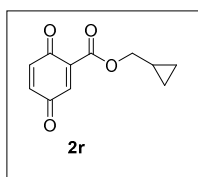
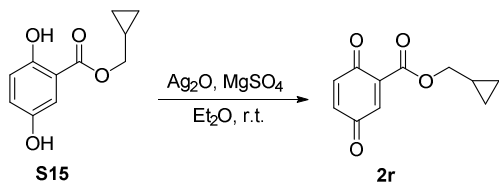
Preparation according to the general procedure C from **S14** (0.412 g, 2 mmol) afforded **2p** as a red oil (0.347 g, 1.70 mmol, 85% yield).

¹H NMR (400 MHz, CDCl₃) δ 7.12 (d, *J* = 2.0 Hz, 1H), 6.81 (d, *J* = 2.0 Hz, 2H), 4.82 (q, *J* = 2.4 Hz, 2H), 1.83 (t, *J* = 2.4 Hz, 3H);

¹³C NMR (100 MHz, CDCl₃) δ 186.7, 182.7, 161.8, 136.8, 136.7, 136.4, 136.1, 84.5, 71.9, 54.4, 3.6;

HRMS ESI Calcd for C₁₁H₈O₄Na [M+Na]⁺: 227.0315, Found: 227.0304.

4.13 Synthesis of the Substrate 2r



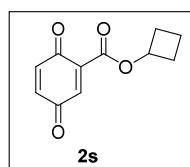
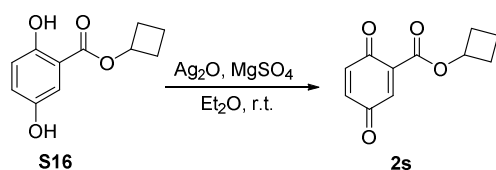
Preparation according to the general procedure C from **S15** (0.415 g, 2 mmol) afforded **2r** as a red oil (0.376 g, 1.83 mmol, 92% yield).

¹H NMR (400 MHz, CDCl₃) δ 7.07 (s, 1H), 6.80 (s, 2H), 4.11 (d, *J* = 7.2 Hz, 2H), 1.22-1.15 (m, 1H), 0.61 (dd, *J* = 5.6 Hz, 13.2 Hz, 2H), 0.32 (dd, *J* = 5.6 Hz, 10.0 Hz, 2H);

¹³C NMR (100 MHz, CDCl₃) δ 186.9, 183.0, 162.6, 137.3, 136.8, 136.02, 135.99, 71.1, 9.5, 3.4;

HRMS ESI Calcd for C₁₁H₁₀O₄Na [M+Na]⁺: 229.0471, Found: 229.0461.

4.14 Synthesis of the Substrate 2s



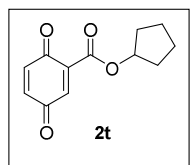
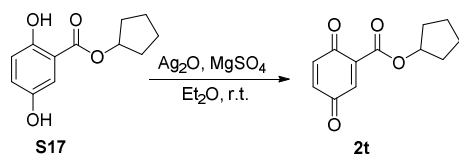
Preparation according to the general procedure C from **S16** (0.417 g, 2 mmol) afforded **2s** as a red oil (0.391 g, 1.88 mmol, 94% yield).

¹H NMR (400 MHz, CDCl₃) δ 7.06 (d, *J* = 1.6 Hz, 1H), 6.80 (d, *J* = 2.4 Hz, 2H), 5.20-5.13 (m, 1H), 2.46-2.38 (m, 2H), 2.22-2.11 (m, 2H), 1.90-1.81 (m, 1H), 1.73-1.61 (m, 1H);

¹³C NMR (100 MHz, CDCl₃) δ 186.9, 183.0, 161.9, 137.2, 136.9, 136.04, 136.02, 70.7, 30.2, 13.5;

HRMS ESI Calcd for C₁₁H₁₀O₄Na [M+Na]⁺: 229.0471, Found: 229.0463.

4.15 Synthesis of the Substrate 2t



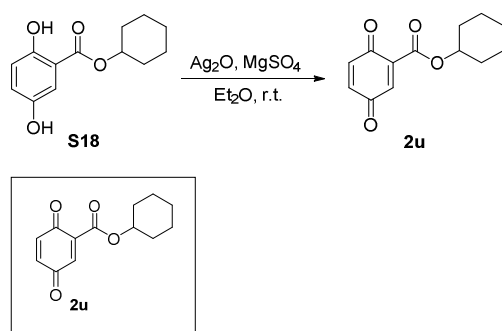
Preparation according to the general procedure C from **S17** (0.444 g, 2 mmol) afforded **2t** as a red oil (0.42 g, 1.90 mmol, 95% yield).

¹H NMR (400 MHz, CDCl₃) δ 7.01 (d, *J* = 2.0 Hz, 1H), 6.79 (d, *J* = 2.4 Hz, 2H), 5.38 (dt, *J* = 2.8 Hz, 5.6 Hz, 1H), 1.95-1.90 (m, 2H), 1.81-1.73 (m, 4H), 1.64-1.61 (m, 2H);

¹³C NMR (100 MHz, CDCl₃) δ 187.0, 183.1, 162.4, 137.7, 136.8, 136.0, 135.6, 79.6, 32.6, 23.6;

HRMS ESI Calcd for C₁₂H₁₂O₄Na [M+Na]⁺: 243.0628, Found: 243.0624.

4.16 Synthesis of the Substrate 2u



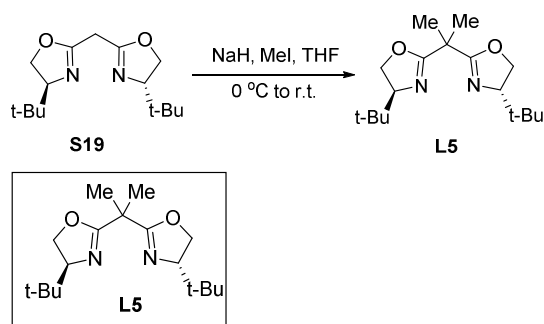
Preparation according to the general procedure C from **S18** (0.471 g, 2 mmol) afforded **2u** as a red oil (0.441 g, 1.88 mmol, 94% yield).

¹H NMR (400 MHz, CDCl₃) δ 7.04 (dd, *J* = 0.4 Hz, 2.0 Hz, 1H), 6.81 (d, *J* = 2.0 Hz, 2H), 5.04-4.98 (m, 1H), 1.94-1.89 (m, 2H), 1.80-1.74 (m, 2H), 1.60-1.51 (m, 3H), 1.47-1.25 (m, 3H);

¹³C NMR (100 MHz, CDCl₃) δ 187.0, 183.1, 162.1, 137.8, 136.9, 136.0, 135.6, 75.1, 31.3, 25.2, 23.4;

HRMS ESI Calcd for C₁₃H₁₁O₄Na [M+Na]⁺: 257.0784, Found: 257.0784.

5. Synthesis of the Ligand L5

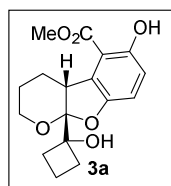


To a solution of **S19** (750 mg 2.82 mmol) in THF (50 mL) in a flame-dried round-bottom flask was added NaH (338 mg, 60 % dispersion in mineral oil, 8.46 mmol, 3 equiv.) at 0 °C. The mixture was stirred at the same temperature for 30 min, and iodomethane (526 μL, 8.46 mmol, 3.0 equiv) was added via syringe. After the addition, the cold bath was removed and the mixture was allowed to stir at room temperature for an additional 6 h. Next, the reaction mixture was quenched with saturated aqueous NH₄Cl solution and extracted with EtOAc. The combined organic layers were washed with brine, dried over MgSO₄, and concentrated. The residue was purified by column chromatography to give **L5** as a colorless solid (581 mg, 1.97 mmol, 71% yield).

¹H NMR (400 MHz, CDCl₃) δ 4.14-4.03 (m, 4H), 3.83 (dt, *J* = 2.8 Hz, 10.0 Hz, 2H), 1.484 (s, 3H), 1.478 (s, 3H), 0.843 (s, 9H), 0.838 (s, 9H);

¹³C NMR (100 MHz, CDCl₃) δ 168.5, 75.2, 68.9, 38.5, 33.9, 25.6, 24.4.

6. Experimental Procedures of the [3+2] Cyclization and Spectroscopic Data of Products (3a as an example)



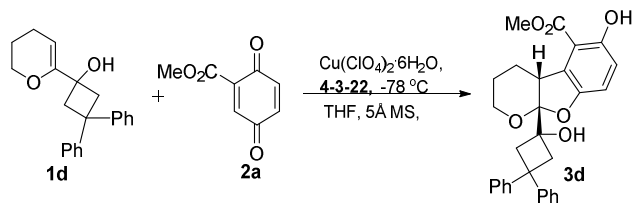
A mixture of $\text{Cu}(\text{ClO}_4)_2 \cdot 6\text{H}_2\text{O}$ (3.7 mg, 0.01 mmol, 0.1 equiv.) and ligand **L5** (3.5 mg, 0.012 mmol, 0.12 equiv.) in THF (1.0 mL) with activated 5Å MS (50 mg) was stirred at room temperature for 2 h under an argon atmosphere. Then the mixture was cooled down to $-78\text{ }^\circ\text{C}$ for 30 minutes, and substituted benzoquinone **2a** (24.9 mg, 0.15 mmol, 1.5 equiv.) was added followed by allylic alcohol **1a** (15.4 mg, 0.1 mmol, 1 equiv.) in THF (0.5 mL). The resulting solution was stirred until **1a** was completely consumed (monitored by TLC, PE/EtOAc = 2/1). Then the mixture was passed through a short silica gel column and eluted with EtOAc (20 mL). The combined elution was concentrated under reduced pressure, which was further purified by flash chromatography on silica gel (PE/EtOAc = 15/1) to give compound **3a** as a colorless oil (28.2 mg, 0.088 mmol, 88%).

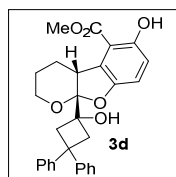
^1H NMR (400 MHz, CDCl_3) δ 10.51 (s, 1H), 6.96 (d, $J = 8.8$ Hz, 1H), 6.79 (d, $J = 8.8$ Hz, 1H), 3.94 (s, 3H), 3.90-3.77 (m, 3H), 2.55-2.50 (m, 1H), 2.37 (d, $J = 8.8$ Hz, 1H), 2.08-2.00 (m, 1H), 1.98-1.87 (m, 3H), 1.86-1.76 (m, 2H), 1.71-1.53 (m, 2H), 1.38-1.23 (m, 1H);

^{13}C NMR (100 MHz, CDCl_3) δ 170.2, 156.5, 151.4, 130.5, 117.1, 116.2, 112.4, 109.2, 78.6, 60.2, 52.2, 40.1, 30.4, 29.2, 23.4, 18.3, 12.9;

HRMS ESI Calcd for $\text{C}_{17}\text{H}_{20}\text{O}_6\text{Na}$ $[\text{M}+\text{Na}]^+$: 343.1152, Found: 343.1144;

HPLC: Chiralpak IF-3 column, n-hexane/isopropanol = 95/5, flow rate = 1.0 mL/min, $\lambda = 254$ nm, retention time: $t_{\text{major}} = 8.409$ min, $t_{\text{minor}} = 10.195$ min, 91% ee; $[\alpha]_{\text{D}}^{26.1} = 28.0$ ($c = 5.0$, CHCl_3).





Preparation according to the general procedure of **3a** from **1d** (30.6 mg, 0.1 mmol) and **2a** (24.9 mg, 0.15 mmol) afforded **3d** as a colorless solid (42.2 mg, 0.090 mmol, 90%).

Mp 69.3–72.1 °C;

¹H NMR (400 MHz, CDCl₃) δ 10.52 (s, 1H), 7.31–7.21 (m, 5H), 7.19–7.06 (m, 5H), 6.66 (d, *J* = 8.8 Hz, 1H), 6.28 (d, *J* = 8.8 Hz, 1H), 3.91 (s, 3H), 3.86–3.71 (m, 2H), 3.53–3.50 (m, 1H), 1.94–1.90 (m, 1H), 1.75–1.70 (m, 1H), 1.56–1.51 (m, 2H), 1.33–1.26 (m, 3H), 0.9–0.83 (m, 2H).

¹³C NMR (100 MHz, CDCl₃) δ 170.3, 156.6, 151.0, 150.6, 149.0, 130.2, 128.2, 128.1, 126.4, 126.3, 125.4, 125.2, 117.0, 116.8, 111.7, 108.9, 74.4, 60.5, 52.2, 44.3, 42.9, 42.8, 40.1, 26.9, 23.6, 18.4;

HRMS ESI Calcd for C₂₉H₂₈O₆Na [M+Na]⁺: 495.1778, Found: 495.1757;

HPLC: Chiralpak IF-3 column, n-hexane/isopropanol = 98/2, flow rate = 1.0 mL/min, λ = 254 nm, retention time: *t*_{major} = 27.313 min, *t*_{minor} = 30.269 min, 87% ee; [α]_D^{26.0} = 30.0 (*c* = 2.0, CHCl₃).

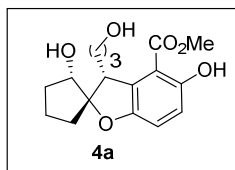
7. Experimental Procedures of the One-Pot [3+2] Cyclization/Semipinacol Rearrangement Cascade and Spectroscopic Data of Products

General procedure D: Experimental Procedures of the One-Pot [3+2] Cyclization/Semipinacol Rearrangement Cascade (using **4a** as an example)

A mixture of Cu(ClO₄)₂·6H₂O (3.7 mg, 0.01 mmol, 0.1 equiv.) and ligand **L5** (3.5 mg, 0.012 mmol, 0.12 equiv.) in THF (1.0 mL) with activated 5 Å MS (50 mg) was stirred at room temperature for 2 h under an argon atmosphere. Then the mixture was cooled down to -78 °C for 30 minutes, and substituted benzoquinone **2a** (24.9 mg, 0.15 mmol, 1.5 equiv.) was added followed by allylic alcohol **1a** (15.4 mg, 0.1 mmol, 1 equiv.) in THF (0.5 mL). The resulting solution was stirred until **1a** was completely consumed (monitored by TLC, PE/EtOAc = 2/1). Then the mixture was passed through a short silica gel column and eluted with EtOAc (20 mL). The combined elution was concentrated under reduced pressure, and then was added dry DCM (4.0 mL) and cooled down to -20 °C for 10 minutes, SnCl₄ (0.20 mL, 0.2 mmol, 1 mol/L in DCM, 2.0 equiv.) was added and stirred at this temperature until **3a** was completely consumed (monitored by TLC, PE/EtOAc = 2/1). And then MeOH (2.0 mL) and NaBH₄ (15.4 mg, 0.4 mmol, 4.0 equiv.) was added and stirred at this temperature for 2 h before it was quenched with the saturated aqueous NH₄Cl. The organic layer was separated and the aqueous layer was extracted with DCM. The combined organic layer was washed with brine, dried over MgSO₄ and concentrated under vacuum, which was further purified by flash chromatography on silica gel (PE/EtOAc = 2/1) to give compound **4a** as a colorless oil (19.5

mg, 0.0609 mmol, 61%, dr = 7.2:1).

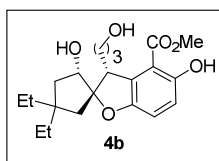
Preparation according to the general procedure D from **1a** (154.1 mg, 1 mmol) and **2a** (249.0 mg, 1.5 mmol) afforded **4a** as a colorless oil (174.1 mg, 0.54 mmol, 54% yield, dr = 7.2:1).



¹H NMR (400 MHz, CDCl₃) δ 10.50 (s, 1H), 6.95 (d, *J* = 8.8 Hz, 1H), 6.79 (d, *J* = 8.8 Hz, 1H), 4.36 (t, *J* = 7.2 Hz, 1H), 3.96 (s, 3H), 3.54 (t, *J* = 5.6 Hz, 2H), 3.45 (dd, *J* = 2.4 Hz, 7.6 Hz, 1H), 2.65 (br, 1H), 2.18-2.16 (m, 1H), 2.04-1.98 (m, 2H), 1.97-1.80 (m, 3H), 1.63-1.54 (m, 4H), 1.46-1.32 (m, 1H);
¹³C NMR (100 MHz, CDCl₃) δ 170.4, 156.6, 150.1, 132.4, 118.0, 117.0, 109.3, 97.0, 73.4, 62.5, 52.2, 50.2, 37.6, 33.0, 30.5, 27.2, 18.6;

HRMS ESI Calcd for C₁₇H₂₂O₆Na [M+Na]⁺: 345.1309, Found: 345.1324;

HPLC: Chiralpak IF-3 column, n-hexane/isopropanol = 90/10, flow rate = 1.0 mL/min, λ = 254 nm, retention time: t_{major} = 15.416 min, t_{minor} = 23.265 min, 91% ee; [α]_D^{17.4} = 4.8 (c = 6.2, CHCl₃).



Preparation according to the general procedure D from **1b** (20.9 mg, 0.1 mmol) and **2a** (24.9 mg, 0.15 mmol) afforded **4b** as a colorless oil (15.3 mg, 0.041 mmol, 41% yield, dr = 10:1).

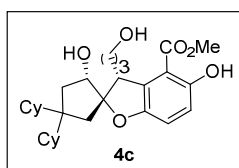
Preparation according to the general procedure D from **1b** (208.8 mg, 1 mmol) and **2a** (249.0 mg, 1.5 mmol) afforded **4b** as a colorless oil (124.2 mg, 0.33 mmol, 33% yield, dr = 10:1).

¹H NMR (400 MHz, CDCl₃) δ 10.50 (s, 1H), 6.92 (d, *J* = 8.8 Hz, 1H), 6.75 (d, *J* = 8.4 Hz, 1H), 4.38 (br, 1H), 3.93 (s, 3H), 3.52-3.46 (m, 2H), 3.37 (d, *J* = 1.6 Hz, 1H), 2.75-2.25 (m, 1H), 2.10-1.96 (m, 3H), 1.84 (d, *J* = 14.8 Hz, 1H), 1.72 (dd, *J* = 10.8 Hz, 12.4 Hz, 1H), 1.58-1.49 (m, 2H), 1.47-1.40 (m, 2H), 1.39-1.26 (m, 3H), 1.22 (t, *J* = 11.2 Hz, 1H), 0.76 (t, *J* = 7.2 Hz, 3H), 0.72 (t, *J* = 7.6 Hz, 3H);

¹³C NMR (100 MHz, CDCl₃) δ 170.4, 156.5, 149.7, 132.8, 118.0, 116.9, 109.1, 97.6, 72.7, 62.3, 52.2, 50.7, 49.4, 45.0, 39.3, 32.4, 32.0, 30.8, 27.2, 8.7, 8.6;

HRMS ESI Calcd for C₂₁H₃₀O₆Na [M+Na]⁺: 401.1935, Found: 401.1942;

HPLC: Chiralpak IF-3 column, n-hexane/isopropanol = 90/10, flow rate = 1.0 mL/min, λ = 254 nm, retention time: t_{major} = 13.494 min, t_{minor} = 20.138 min, 91% ee; [α]_D^{19.8} = -10.0 (c = 1.0, CHCl₃).



Preparation according to the general procedure D from **1c** (31.8 mg, 0.1 mmol) and **2a** (24.9 mg, 0.15 mmol) afforded **4c** as a colorless oil (19.4 mg, 0.040 mmol, 40% yield, dr > 20:1).

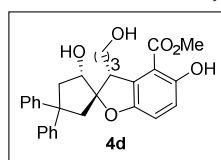
Preparation according to the general procedure D from **1c** (319.4 mg, 1 mmol) and **2a** (249.0 mg, 1.5 mmol) afforded **4c** as a colorless oil (180.1 mg, 0.37 mmol, 37% yield, dr > 20:1).

¹H NMR (400 MHz, CDCl₃) δ 10.52 (s, 1H), 6.93 (d, *J* = 8.8 Hz, 1H), 6.80 (d, *J* = 8.8 Hz, 1H), 4.37 (t, *J* = 9.2 Hz, 1H), 3.98 (s, 3H), 3.97-3.91 (m, 1H), 3.56 (t, *J* = 8 Hz, 2H), 3.41 (dd, *J* = 2.0 Hz, 8.4 Hz, 1H), 2.47 (br, 1H), 3.13-2.01 (m, 1H), 1.85-1.76 (m, 10H), 1.69-1.57 (m, 10H), 1.55-1.45 (m, 2H), 1.89-1.06 (m, 4H), 0.91-0.81 (m, 2H);

¹³C NMR (100 MHz, CDCl₃) δ 170.5, 156.6, 149.4, 133.4, 118.1, 117.0, 109.2, 94.4, 77.2, 73.4, 62.5, 52.3, 51.1, 45.1, 44.0, 43.3, 43.1, 39.9, 30.9, 29.7, 28.1, 27.9, 27.8, 27.4, 27.3, 27.1, 27.0, 26.8, 26.7;

HRMS ESI Calcd for C₂₉H₄₂O₆Na [M+Na]⁺: 509.2874, Found: 509.2862;

HPLC: Chiralpak IF-3 column, n-hexane/isopropanol = 90/10, flow rate = 1.0 mL/min, λ = 254 nm, retention time: *t*_{major} = 13.962 min, *t*_{minor} = 25.945 min, 89% ee; [α]_D^{20.2} = -16.7 (*c* = 1.2, CHCl₃).



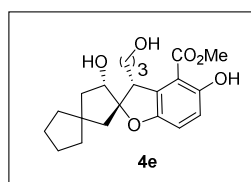
Preparation according to the general procedure D from **1d** (30.0 mg, 0.1 mmol) and **2a** (24.9 mg, 0.15 mmol) afforded **4d** as a colorless oil (30.9 mg, 0.067 mmol, 67% yield, dr = 15.5:1).

¹H NMR (400 MHz, CDCl₃) δ 10.52 (s, 1H), 7.32-7.24 (m, 5H), 7.22-7.16 (m, 2H), 7.14-7.12 (m, 3H), 6.93 (d, *J* = 8.8 Hz, 1H), 6.81 (d, *J* = 8.8 Hz, 1H), 4.52-4.48 (m, 1H), 3.94 (s, 3H), 3.51-3.46 (m, 2H), 3.16 (dd, *J* = 6.0 Hz, 12.4 Hz, 1H), 2.90 (d, *J* = 14.8 Hz, 1H), 2.89 (br, 1H), 2.75 (d, *J* = 14.8 Hz, 1H), 2.55 (dd, *J* = 2.8 Hz, 8.4 Hz, 1H), 2.32-2.26 (m, 1H), 2.04 (s, 1H), 1.97-1.87 (m, 1H), 1.68 (br, 1H), 1.57-1.42 (m, 1H), 1.32-1.23 (m, 1H);

¹³C NMR (100 MHz, CDCl₃) δ 170.3, 156.8, 149.8, 149.6, 148.3, 132.3, 128.5, 128.1, 126.7, 126.5, 126.0, 125.7, 118.1, 117.2, 109.3, 96.4, 72.1, 62.4, 60.4, 52.9, 52.2, 51.6, 49.3, 46.7, 30.3, 27.2, 14.1;

HRMS ESI Calcd for C₂₉H₃₀O₆Na [M+Na]⁺: 497.1935, Found: 497.1938;

HPLC: Chiralpak IF-3 column, n-hexane/isopropanol = 90/10, flow rate = 1.0 mL/min, λ = 254 nm, retention time: *t*_{major} = 23.816 min, *t*_{minor} = 30.598 min, 92% ee; [α]_D^{20.3} = -72.7 (*c* = 1.1, CHCl₃).



Preparation according to the general procedure D from **1e** (20.9 mg, 0.1 mmol) and **2a** (24.9 mg, 0.15 mmol) afforded **4e** as a colorless oil (17.3 mg, 0.047 mmol, 47% yield, dr > 20:1).

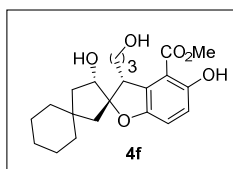
Preparation according to the general procedure D from **1e** (208.8 mg, 1 mmol) and **2a** (249.0 mg, 1.5 mmol) afforded **4e** as a colorless oil (165.0 mg, 0.44 mmol, 44% yield, dr > 20:1).

¹H NMR (400 MHz, CDCl₃) δ 10.49 (s, 1H), 6.96 (d, *J* = 8.8 Hz, 1H), 6.80 (d, *J* = 8.8 Hz, 1H), 4.46-4.41 (m, 1H), 3.97 (s, 3H), 3.57-3.52 (m, 2H), 3.45 (dd, *J* = 2.4 Hz, 8.0 Hz, 1H), 2.46 (d, *J* = 9.6 Hz, 1H), 2.10-1.98 (m, 3H), 1.89-1.78 (m, 2H), 1.67-1.44 (m, 10H), 1.28-1.24 (m, 2H);

¹³C NMR (100 MHz, CDCl₃) δ 170.4, 156.7, 150.0, 132.6, 118.1, 117.1, 109.3, 97.5, 73.2, 62.6, 52.2, 51.5, 50.9, 46.8, 44.3, 41.8, 41.7, 30.8, 27.2, 23.8, 23.6;

HRMS ESI Calcd for C₂₁H₂₈O₆Na [M+Na]⁺: 399.1778, Found: 399.1771;

HPLC: Chiralpak IF-3 column, n-hexane/isopropanol = 90/10, flow rate = 1.0 mL/min, λ = 254 nm, retention time: *t*_{major} = 16.083 min, *t*_{minor} = 23.715 min, 86% ee; [α]_D^{20.0} = -22.2 (c = 0.9, CHCl₃).



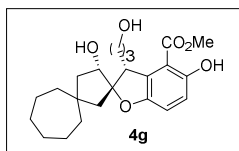
Preparation according to the general procedure D from **1f** (22.1 mg, 0.1 mmol) and **2a** (24.9 mg, 0.15 mmol) afforded **4f** as a colorless oil (19.6 mg, 0.051 mmol, 51% yield, dr > 20:1).

¹H NMR (400 MHz, CDCl₃) δ 10.50 (s, 1H), 6.95 (d, *J* = 8.4 Hz, 1H), 6.79 (d, *J* = 8.8 Hz, 1H), 4.45-4.43 (m, 1H), 3.97 (s, 3H), 3.57 (t, *J* = 5.6 Hz, 2H), 3.45 (dd, *J* = 2.4 Hz, 5.6 Hz, 1H), 2.40 (br, 1H), 2.18-2.09 (m, 1H), 2.05 (d, *J* = 14.0 Hz, 1H), 1.96 (d, *J* = 14.8 Hz, 1H), 1.88-1.78 (m, 1H), 1.75-1.65 (m, 1H), 1.63-1.47 (m, 6H), 1.47-1.25 (m, 8H);

¹³C NMR (100 MHz, CDCl₃) δ 170.4, 156.6, 149.8, 132.7, 118.1, 117.0, 109.2, 97.4, 72.6, 62.5, 52.2, 50.8, 40.5, 40.4, 36.9, 30.9, 27.2, 25.7, 23.3, 23.2;

HRMS ESI Calcd for C₂₂H₃₀O₆Na [M+Na]⁺: 413.1935, Found: 413.1917;

HPLC: Chiralpak IF-3 column, n-hexane/isopropanol = 90/10, flow rate = 1.0 mL/min, λ = 254 nm, retention time: *t*_{major} = 15.517 min, *t*_{minor} = 22.968 min, 88% ee; [α]_D^{20.1} = -12.5 (c = 1.6, CHCl₃).



Preparation according to the general procedure D from **1g** (23.7 mg, 0.1 mmol) and **2a** (24.9 mg, 0.15 mmol) afforded **4g** as a colorless oil (22.4 mg, 0.056 mmol, 56% yield, dr > 20:1).

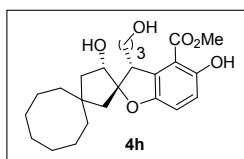
Preparation according to the general procedure D from **1g** (235.7 mg, 1 mmol) and **2a** (249.0 mg, 1.5 mmol) afforded **4g** as a colorless oil (190.0 mg, 0.47 mmol, 47% yield, dr > 20:1).

¹H NMR (400 MHz, CDCl₃) δ 10.49 (s, 1H), 6.95 (d, *J* = 8.8 Hz, 1H), 6.79 (d, *J* = 8.8 Hz, 1H), 4.41 (dd, *J* = 7.6 Hz, 10.4 Hz, 1H), 3.97-3.94 (m, 1H), 3.96 (s, 3H), 3.57 (t, *J* = 5.6 Hz, 2H), 3.43 (dd, *J* = 2.4 Hz, 8.0 Hz, 1H), 2.17-2.05 (m, 2H), 2.00 (d, *J* = 14.4 Hz, 1H), 1.74 (dd, *J* = 10.8 Hz, 12.4 Hz, 2H), 1.66-1.52 (m, 8H), 1.51-1.37 (m, 8H);

¹³C NMR (100 MHz, CDCl₃) δ 170.4, 156.6, 149.8, 132.7, 118.1, 117.0, 109.2, 97.6, 72.6, 62.5, 52.6, 52.2, 50.9, 47.9, 44.0, 43.5, 40.0, 30.9, 29.0, 28.9, 27.2, 23.6, 23.4;

HRMS ESI Calcd for C₂₃H₃₂O₆Na [M+Na]⁺: 427.2091, Found: 427.2072;

HPLC: Chiralpak IF-3 column, n-hexane/isopropanol = 90/10, flow rate = 1.0 mL/min, λ = 254 nm, retention time: *t*_{major} = 16.898 min, *t*_{minor} = 25.782 min, 92% ee; [α]_D^{20.2} = -18.2 (c = 1.1, CHCl₃).



Preparation according to the general procedure D from **1h** (24.9 mg, 0.1 mmol) and **2a** (24.9 mg, 0.15 mmol) afforded **4h** as a colorless oil (22.8 mg, 0.055 mmol, 55% yield, dr > 20:1).

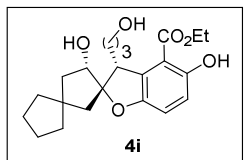
Preparation according to the general procedure D from **1h** (249.4 mg, 1 mmol) and **2a** (249.0 mg, 1.5 mmol) afforded **4h** as a colorless oil (199.7 mg, 0.48 mmol, 48% yield, dr > 20:1).

¹H NMR (400 MHz, CDCl₃) δ 10.49 (s, 1H), 6.95 (d, *J* = 8.8 Hz, 1H), 6.79 (d, *J* = 8.4 Hz, 1H), 4.43 (br, 1H), 3.96 (s, 3H), 3.56 (t, *J* = 5.6 Hz, 2H), 3.42 (dd, *J* = 2.4 Hz, 8.4 Hz, 1H), 2.48 (br, 1H), 2.18-2.01 (m, 3H), 1.97 (d, *J* = 10.8 Hz, 1H), 1.70 (dd, *J* = 10.8 Hz, 12.8 Hz, 3H), 1.61-1.60 (m, 3H), 1.60-1.56 (m, 1H), 1.56-1.45 (m, 9H), 1.44-1.35 (m, 3H);

¹³C NMR (100 MHz, CDCl₃) δ 170.4, 156.6, 149.8, 132.8, 118.0, 117.0, 109.2, 97.7, 72.6, 62.4, 52.2, 52.1, 50.9, 47.3, 39.9, 38.5, 38.1, 30.9, 28.6, 28.3, 27.3, 24.7, 23.2, 23.1;

HRMS ESI Calcd for C₂₄H₃₄O₆Na [M+Na]⁺: 441.2248, Found: 441.2235;

HPLC: Chiralpak IF-3 column, n-hexane/isopropanol = 90/10, flow rate = 1.0 mL/min, λ = 254 nm, retention time: *t*_{major} = 19.329 min, *t*_{minor} = 25.994 min, 92% ee; [α]_D^{20.3} = -14.3 (c = 1.4, CHCl₃).



Preparation according to the general procedure D from **1e** (20.7 mg, 0.1 mmol) and **2b** (27.0 mg, 0.15 mmol) afforded **4i** as a colorless oil (19.7 mg, 0.051 mmol, 51% yield, dr > 20:1).

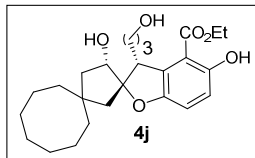
¹H NMR (400 MHz, CDCl₃) δ 10.60 (s, 1H), 6.96 (d, *J* = 8.8 Hz, 1H), 6.69 (d, *J* = 8.8 Hz, 1H), 4.48-4.41 (m, 3H), 3.56 (t, *J* = 5.6 Hz, 2H), 3.46 (dd, *J* = 2.0 Hz, 8.4 Hz, 1H), 2.09-2.02 (m, 2H), 2.00 (d, *J* = 14.4 Hz, 1H), 1.91-1.88 (m, 1H), 1.67-1.46 (m, 12H), 1.46 (t, *J* = 7.2 Hz, 5H);

¹³C NMR (100 MHz, CDCl₃) δ 170.1, 156.8, 149.9, 132.5, 117.9, 117.1, 109.5, 97.4, 73.2, 62.6, 61.6,

51.3, 50.8, 46.7, 41.8, 30.5, 27.1, 23.8, 23.6, 14.3;

HRMS ESI Calcd for $C_{22}H_{30}O_6Na$ $[M+Na]^+$: 413.1935, Found: 413.1944;

HPLC: Chiralpak IF-3 column, n-hexane/isopropanol = 90/10, flow rate = 1.0 mL/min, λ = 254 nm, retention time: t_{major} = 13.789 min, t_{minor} = 23.829 min, 75% ee; $[\alpha]_D^{20.1}$ = -15.4 (c = 1.3, $CHCl_3$).



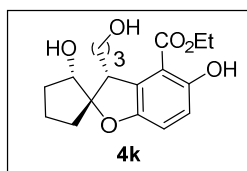
Preparation according to the general procedure D from **1h** (25.0 mg, 0.1 mmol) and **2b** (27.0 mg, 0.15 mmol) afforded **4j** as a colorless oil (22.5 mg, 0.051 mmol, 53% yield, dr > 20:1).

1H NMR (400 MHz, $CDCl_3$) δ 10.60 (s, 1H), 6.95 (d, J = 8.4 Hz, 1H), 6.79 (d, J = 8.8 Hz, 1H), 4.48-4.40 (m, 3H), 3.56 (t, J = 5.6 Hz, 2H), 3.44 (dd, J = 2.0 Hz, 8.8 Hz, 1H), 2.53-2.32 (m, 1H), 2.16-2.047 (m, 1H), 2.02-1.96 (m, 2H), 1.92-1.79 (m, 1H), 1.73-1.56 (m, 8H), 1.56-1.52 (m, 2H), 1.46-1.41 (m, 8H), 1.37-1.33 (m, 2H);

^{13}C NMR (100 MHz, $CDCl_3$) δ 170.1, 156.7, 149.7, 132.7, 118.0, 117.0, 109.4, 97.7, 72.7, 62.6, 61.6, 51.9, 50.9, 47.3, 40.0, 38.5, 38.2, 30.7, 28.6, 28.3, 27.2, 24.7, 23.24, 23.15, 14.3;

HRMS ESI Calcd for $C_{25}H_{36}O_6Na$ $[M+Na]^+$: 455.2404, Found: 455.2403;

HPLC: Chiralpak IF-3 column, n-hexane/isopropanol = 90/10, flow rate = 1.0 mL/min, λ = 254 nm, retention time: t_{major} = 16.813 min, t_{minor} = 27.576 min, 86% ee; $[\alpha]_D^{20.2}$ = -13.3 (c = 1.5, $CHCl_3$).



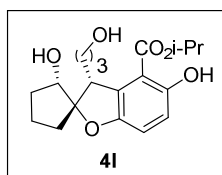
Preparation according to the general procedure D from **1a** (15.4 mg, 0.1 mmol) and **2b** (27.0 mg, 0.15 mmol) afforded **4k** as a colorless oil (20.2 mg, 0.06 mmol, 60% yield, dr = 12.6:1).

1H NMR (400 MHz, $CDCl_3$) δ 10.62 (s, 1H), 6.95 (d, J = 8.8 Hz, 1H), 6.79 (d, J = 8.8 Hz, 1H), 4.49-4.37 (m, 3H), 3.54 (t, J = 6.0 Hz, 2H), 3.47 (dd, J = 2.4 Hz, 8.0 Hz, 1H), 2.57 (br, 1H), 2.20-2.12 (m, 1H), 2.04-1.86 (m, 3H), 1.84-1.64 (m, 2H), 1.63-1.50 (m, 4H), 1.43 (t, J = 7.2 Hz, 3H);

^{13}C NMR (100 MHz, $CDCl_3$) δ 170.1, 156.8, 150.1, 132.4, 117.9, 117.1, 109.5, 97.0, 73.4, 62.6, 61.7, 50.2, 37.6, 33.0, 30.3, 27.1, 18.5, 14.3;

HRMS ESI Calcd for $C_{18}H_{24}O_6Na$ $[M+Na]^+$: 359.1465, Found: 359.1454;

HPLC: Chiralpak IF-3 column, n-hexane/isopropanol = 90/10, flow rate = 1.0 mL/min, λ = 254 nm, retention time: t_{major} = 14.317 min, t_{minor} = 23.956 min, 88% ee; $[\alpha]_D^{17.9}$ = -8.3 (c = 1.0, $CHCl_3$).



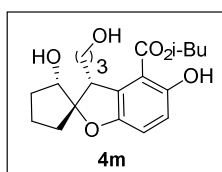
Preparation according to the general procedure D from **1a** (15.4 mg, 0.1 mmol) and **2c** (29.2 mg, 0.15 mmol) afforded **4l** as a colorless oil (19.7 mg, 0.057 mmol, 57% yield, dr = 9.4:1).

¹H NMR (400 MHz, CDCl₃) δ 10.73 (s, 1H), 6.95 (d, *J* = 8.8 Hz, 1H), 6.79 (d, *J* = 8.8 Hz, 1H), 5.37-5.31 (m, 1H), 4.40 (t, *J* = 7.2 Hz, 1H), 3.54 (t, *J* = 6.0 Hz, 2H), 3.47 (dd, *J* = 2.0 Hz, 8.0 Hz, 1H), 2.47 (br, 1H), 2.20-2.16 (m, 1H), 2.05-2.00 (m, 2H), 1.85-1.77 (m, 2H), 1.66-1.56 (m, 5H), 1.42 (d, *J* = 4.8 Hz, 3H), 1.41 (d, *J* = 5.2 Hz, 3H);

¹³C NMR (100 MHz, CDCl₃) δ 169.3, 156.9, 150.0, 132.3, 117.8, 117.1, 109.9, 96.9, 73.5, 67.9, 62.6, 50.1, 37.5, 33.0, 30.2, 27.1, 22.0, 21.8, 18.5;

HRMS ESI Calcd for C₁₉H₂₆O₆Na [M+Na]⁺: 373.1622, Found: 373.1617;

HPLC: Chiralpak IF-3 column, n-hexane/isopropanol = 90/10, flow rate = 1.0 mL/min, λ = 254 nm, retention time: *t*_{major} = 13.468 min, *t*_{minor} = 20.366 min, 80% ee; [α]_D^{18.2} = -14.3 (c = 0.7, CHCl₃).



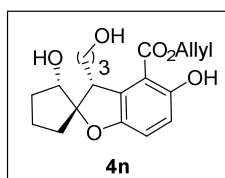
Preparation according to the general procedure D from **1a** (15.4 mg, 0.1 mmol) and **2m** (31.4 mg, 0.15 mmol) afforded **4m** as a colorless oil (19.4 mg, 0.054 mmol, 54% yield, dr = 4.7:1).

¹H NMR (400 MHz, CDCl₃) δ 10.58 (s, 1H), 6.95 (d, *J* = 8.8 Hz, 1H), 6.78 (d, *J* = 8.8 Hz, 1H), 4.37 (t, *J* = 7.2 Hz, 1H), 4.78 (dd, *J* = 3.2 Hz, 10.8 Hz, 1H), 4.08 (d, *J* = 6.4 Hz, 1H), 3.51 (t, *J* = 6.4 Hz, 3H), 2.75 (br, 1H), 2.17-2.08 (m, 2H), 2.03-1.84 (m, 2H), 1.83-1.62 (m, 3H), 1.61-1.53 (m, 4H), 1.30-1.25 (m, 1H), 1.03 (d, *J* = 2.8 Hz, 3H), 1.01 (d, *J* = 3.2 Hz, 3H);

¹³C NMR (100 MHz, CDCl₃) δ 170.1, 156.7, 150.2, 132.2, 117.8, 117.1, 109.6, 97.0, 73.3, 71.8, 62.5, 50.1, 37.4, 32.9, 29.8, 27.8, 27.1, 19.4, 19.2, 18.5;

HRMS ESI Calcd for C₂₀H₂₈O₆Na [M+Na]⁺: 387.1778, Found: 387.1769;

HPLC: Chiralpak IF-3 column, n-hexane/isopropanol = 90/10, flow rate = 1.0 mL/min, λ = 254 nm, retention time: *t*_{major} = 11.103 min, *t*_{minor} = 18.840 min, 80% ee; [α]_D^{19.1} = 7.1 (c = 1.4, CHCl₃).



Preparation according to the general procedure D from **1a** (15.4 mg, 0.1 mmol) and **2n** (28.7 mg,

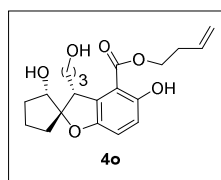
0.15 mmol) afforded **4n** as a colorless oil (17.7 mg, 0.051 mmol, 51% yield, dr = 4.7:1).

¹H NMR (400 MHz, CDCl₃) δ 10.52 (s, 1H), 6.96 (d, *J* = 8.8 Hz, 1H), 6.80 (d, *J* = 8.8 Hz, 1H), 6.11-6.04 (m, 1H), 5.37 (dt, *J* = 1.2 Hz, 7.6 Hz, 2H), 4.92-4.89 (m, 1H), 4.39 (d, *J* = 7.2 Hz, 2H), 3.55-3.51 (m, 2H), 3.47 (dd, *J* = 2.0 Hz, 8.0 Hz, 1H), 2.20-2.14 (m, 1H), 2.05-1.90 (m, 2H), 1.86-1.77 (m, 4H), 1.66-1.54 (m, 4H), 1.43-1.26 (m, 1H);

¹³C NMR (100 MHz, CDCl₃) δ 169.7, 156.9, 150.1, 132.5, 131.4, 120.1, 118.1, 117.1, 109.3, 97.0, 66.4, 62.6, 50.2, 37.6, 33.0, 30.4, 27.1, 18.6;

HRMS ESI Calcd for C₁₉H₂₄O₆Na [M+Na]⁺: 371.1465, Found: 371.1459;

HPLC: Chiralpak IF-3 column, n-hexane/isopropanol = 90/10, flow rate = 1.0 mL/min, λ = 254 nm, retention time: t_{major} = 13.772 min, t_{minor} = 22.585 min, 84% ee; [α]_D^{19.4} -14.3 (c = 1.4, CHCl₃).



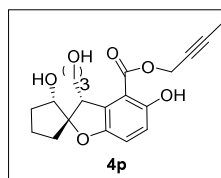
Preparation according to the general procedure D from **1a** (15.4 mg, 0.1 mmol) and **2o** (31.2 mg, 0.15 mmol) afforded **4o** as a colorless oil (19.8 mg, 0.055 mmol, 55% yield, dr = 4.6:1).

¹H NMR (400 MHz, CDCl₃) δ 10.52 (s, 1H), 6.95 (d, *J* = 8.8 Hz, 1H), 6.78 (d, *J* = 8.8 Hz, 1H), 5.87-5.79 (m, 1H), 5.20 (m, 2H), 4.52-4.42 (m, 1H), 4.41-4.35 (m, 2H), 3.55-3.48 (m, 2H), 2.57-2.52 (m, 2H), 2.23-2.12 (m, 1H), 2.04-1.88 (m, 2H), 1.87-1.63 (m, 4H), 1.63-1.53 (m, 4H), 1.42-1.40 (m, 1H), 1.25 (t, *J* = 7.2 Hz, 1H);

¹³C NMR (100 MHz, CDCl₃) δ 170.0, 156.7, 150.2, 132.3, 117.94, 117.87, 117.1, 109.5, 97.1, 77.3, 73.4, 64.6, 62.5, 50.0, 37.3, 33.0, 32.9, 29.9, 27.0, 18.5;

HRMS ESI Calcd for C₂₀H₂₆O₆K [M+K]⁺: 401.1361, Found: 401.1382;

HPLC: Chiralpak IF-3 column, n-hexane/isopropanol = 90/10, flow rate = 1.0 mL/min, λ = 254 nm, retention time: t_{major} = 13.119 min, t_{minor} = 21.234 min, 83% ee; [α]_D^{20.6} = -10.0 (c = 2.0, CHCl₃).



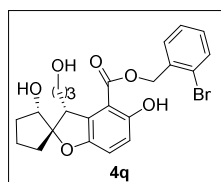
Preparation according to the general procedure D from **1a** (15.4 mg, 0.1 mmol) and **2p** (30.5 mg, 0.15 mmol) afforded **4p** as a colorless oil (18.8 mg, 0.057 mmol, 52% yield, dr = 3.9:1).

¹H NMR (400 MHz, CDCl₃) δ 10.41 (s, 1H), 6.96 (d, *J* = 8.8 Hz, 1H), 6.78 (d, *J* = 8.4 Hz, 1H), 5.05-4.95 (m, 1H), 4.88-4.80 (m, 1H), 4.39 (t, *J* = 7.2 Hz, 1H), 3.58-3.45 (m, 3H), 2.76 (br, 1H), 2.21-2.05 (m, 2H), 2.03-1.93 (m, 1H), 1.88-1.80 (m, 3H), 1.87 (s, 3H), 1.74-1.44 (m, 5H);

¹³C NMR (100 MHz, CDCl₃) δ 169.7, 169.4, 156.8, 156.4, 150.5, 150.1, 133.2, 132.7, 118.2, 118.1, 117.0, 116.8, 109.2, 109.0, 101.1, 97.0, 84.3, 84.1, 73.4, 72.5, 63.0, 62.6, 53.6, 53.4, 50.3, 47.8, 37.6, 35.2, 33.7, 32.9, 30.7, 29.2, 27.7, 27.3, 19.5, 18.5, 3.61, 3.58;

HRMS ESI Calcd for C₂₀H₂₅O₆ [M+H]⁺: 361.1646, Found: 361.1633;

HPLC: Chiralpak IF-3 column, n-hexane/isopropanol = 90/10, flow rate = 1.0 mL/min, λ = 254 nm, retention time: t_{major} = 14.961 min, t_{minor} = 25.074 min, 86% ee; [α]_D^{19.2} = -9.1 (c = 2.2, CHCl₃).



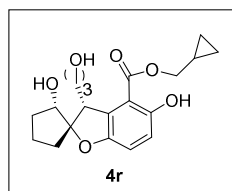
Preparation according to the general procedure D from **1a** (15.4 mg, 0.1 mmol) and **2q** (48.1 mg, 0.15 mmol) afforded **4q** as a colorless oil (22.2 mg, 0.047 mmol, 47% yield, dr = 11.4:1).

¹H NMR (400 MHz, CDCl₃) δ 10.54 (s, 1H), 7.65-7.60 (m, 1H), 7.52-7.46 (m, 1H), 7.37 (dt, *J* = 0.8 Hz, 7.6 Hz, 1H), 7.28 (dd, *J* = 0.8 Hz, 2 Hz, 1H), 6.95 (d, *J* = 9.6 Hz, 1H), 6.79 (d, *J* = 8.8 Hz, 1H), 5.48 (d, *J* = 3.2 Hz, 2H), 4.30 (t, *J* = 7.2 Hz, 1H), 3.41 (dd, *J* = 2.0 Hz, 8.4 Hz, 1H), 3.33 (t, *J* = 6.0 Hz, 2H), 2.17-2.08 (m, 1H), 2.02-1.90 (m, 2H), 1.86-1.68 (m, 4H), 1.60-1.52 (m, 2H), 1.45-1.36 (m, 1H);

¹³C NMR (100 MHz, CDCl₃) δ 169.7, 156.9, 150.1, 134.3, 133.3, 132.8, 131.7, 130.8, 127.8, 124.8, 118.2, 117.0, 109.0, 97.0, 73.2, 67.0, 62.5, 50.0, 37.1, 32.8, 30.0, 27.0, 18.4;

HRMS ESI Calcd for C₂₃H₂₅BrO₆Na [M+Na]⁺: 499.0727, Found: 499.0717;

HPLC: Chiralpak IF-3 column, n-hexane/isopropanol = 90/10, flow rate = 1.0 mL/min, λ = 254 nm, retention time: t_{major} = 18.563 min, t_{minor} = 37.175 min, 74% ee; [α]_D^{20.8} = -3.8 (c = 2.6, CHCl₃).



Preparation according to the general procedure D from **1a** (15.4 mg, 0.1 mmol) and **2r** (30.6 mg, 0.15 mmol) afforded **4r** as a colorless oil (18.6 mg, 0.052 mmol, 52% yield, dr = 4.7:1).

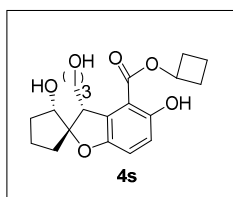
¹H NMR (400 MHz, CDCl₃) δ 10.62 (s, 1H), 6.93 (d, *J* = 8.8 Hz, 1H), 6.77 (d, *J* = 8.8 Hz, 1H), 4.37 (t, *J* = 6.8 Hz, 1H), 4.25-4.11 (m, 2H), 3.50 (m, 3H), 2.88 (br, 1H), 2.61 (br, 1H), 2.20-1.82 (m, 3H), 1.81-1.75 (m, 3H), 1.70-1.42 (m, 3H), 1.28-1.25 (m, 1H), 1.24-1.22 (m, 1H), 0.67-0.61 (m, 2H), 0.36 (dd, *J* = 4.8 Hz, 9.6 Hz, 2H);

¹³C NMR (100 MHz, CDCl₃) δ 170.1, 156.7, 150.1, 132.4, 117.8, 116.9, 109.5, 100.8, 97.0, 73.3, 70.6, 62.5, 50.2, 37.5, 32.8, 30.1, 27.1, 18.5, 9.7, 3.7;

HRMS ESI Calcd for C₂₀H₂₆O₆Na [M+Na]⁺: 385.1622, Found: 385.1627;

HPLC: Chiralpak IF-3 column, n-hexane/isopropanol = 90/10, flow rate = 1.0 mL/min, λ = 254 nm,

retention time: $t_{\text{major}} = 13.792$ min, $t_{\text{minor}} = 23.171$ min, 78% ee; $[\alpha]_{\text{D}}^{19.5} = -5.0$ ($c = 2.0$, CHCl_3).



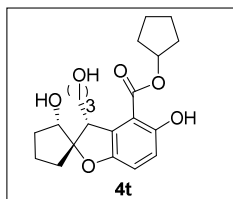
Preparation according to the general procedure D from **1a** (15.4 mg, 0.1 mmol) and **2s** (30.6 mg, 0.15 mmol) afforded **4s** as a colorless oil (20.9 mg, 0.057 mmol, 57% yield, dr = 7.8:1).

^1H NMR (400 MHz, CDCl_3) δ 10.61 (s, 1H), 6.94 (d, $J = 8.8$ Hz, 1H), 6.77 (d, $J = 8.8$ Hz, 1H), 5.24 (t, $J = 7.6$ Hz, 1H), 4.39 (t, $J = 7.6$ Hz, 1H), 3.54 (t, $J = 5.6$ Hz, 2H), 3.50 (dd, $J = 2.4$ Hz, 8.0 Hz, 1H), 2.52-2.44 (m, 2H), 2.27-2.12 (m, 3H), 2.11-1.98 (m, 2H), 1.95-1.79 (m, 3H), 1.78-1.69 (m, 1H), 1.68-1.54 (m, 4H), 1.48-1.37 (m, 1H), 1.29-1.23 (m, 1H), 0.93-0.84 (m, 1H);

^{13}C NMR (100 MHz, CDCl_3) δ 169.5, 156.8, 150.0, 132.3, 117.9, 117.0, 109.4, 96.9, 73.4, 70.1, 62.6, 50.2, 37.6, 33.0, 30.5, 30.31, 30.29, 27.2, 18.5, 13.7;

HRMS ESI Calcd for $\text{C}_{20}\text{H}_{26}\text{O}_6\text{Na}$ $[\text{M}+\text{Na}]^+$: 385.1622, Found: 385.1634;

HPLC: Chiralpak IF-3 column, n-hexane/isopropanol = 90/10, flow rate = 1.0 mL/min, $\lambda = 254$ nm, retention time: $t_{\text{major}} = 14.345$ min, $t_{\text{minor}} = 24.116$ min, 77% ee; $[\alpha]_{\text{D}}^{20.4} = -9.5$ ($c = 2.1$, CHCl_3).



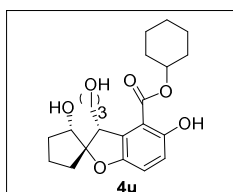
Preparation according to the general procedure D from **1a** (15.4 mg, 0.1 mmol) and **2t** (32.8 mg, 0.15 mmol) afforded **4t** as a colorless oil (20.2 mg, 0.054 mmol, 54% yield, dr = 6.7:1).

^1H NMR (400 MHz, CDCl_3) δ 10.69 (s, 1H), 6.94 (d, $J = 8.8$ Hz, 1H), 6.78 (d, $J = 8.4$ Hz, 1H), 5.49-5.43 (m, 1H), 4.38 (t, $J = 7.2$ Hz, 1H), 3.54-3.48 (m, 2H), 3.46 (dd, $J = 2.8$ Hz, 8 Hz, 1H), 2.21-2.14 (m, 1H), 2.03-1.93 (m, 4H), 1.89-1.78 (m, 6H), 1.76-1.61 (m, 6H), 1.59-1.50 (m, 2H);

^{13}C NMR (100 MHz, CDCl_3) δ 170.0, 156.7, 150.1, 132.0, 117.7, 117.1, 109.8, 96.9, 79.0, 73.3, 62.5, 50.1, 37.4, 32.9, 32.8, 32.5, 29.6, 27.0, 23.9, 23.7, 18.5;

HRMS ESI Calcd for $\text{C}_{21}\text{H}_{28}\text{O}_6\text{Na}$ $[\text{M}+\text{Na}]^+$: 399.1778, Found: 399.1791;

HPLC: Chiralpak IF-3 column, n-hexane/isopropanol = 90/10, flow rate = 1.0 mL/min, $\lambda = 254$ nm, retention time: $t_{\text{major}} = 13.156$ min, $t_{\text{minor}} = 21.162$ min, 81% ee; $[\alpha]_{\text{D}}^{20.5} = -21.4$ ($c = 1.4$, CHCl_3).



Preparation according to the general procedure D from **1a** (15.4 mg, 0.1 mmol) and **2u** (35.2 mg,

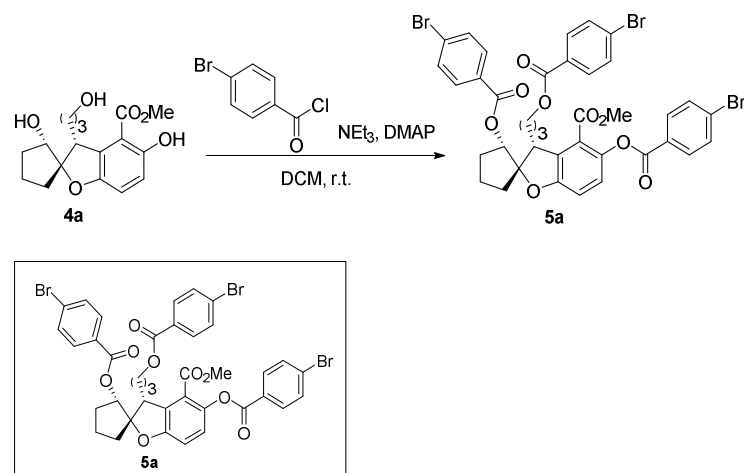
0.15 mmol) afforded **4u** as a colorless oil (22.3 mg, 0.057 mmol, 57% yield, dr = 5.4:1).

¹H NMR (400 MHz, CDCl₃) δ 10.70 (s, 1H), 6.92 (d, *J* = 8.8 Hz, 1H), 6.76 (d, *J* = 8.8 Hz, 1H), 5.09-5.02 (m, 1H), 4.37 (t, *J* = 7.2 Hz, 1H), 3.52-3.47 (m, 3H), 2.85 (br, 1H), 2.12-2.06 (m, 1H), 2.11-1.98 (m, 5H), 1.83-1.75 (m, 5H), 1.64-1.54 (m, 6H), 1.54-1.24 (m, 4H);

¹³C NMR (100 MHz, CDCl₃) δ 169.5, 156.7, 150.1, 132.2, 117.7, 117.0, 109.8, 96.9, 74.9, 73.3, 62.5, 50.1, 37.4, 32.9, 31.8, 31.7, 29.8, 27.1, 25.2, 24.1, 24.0, 18.5;

HRMS ESI Calcd for C₂₂H₃₀O₆Na [M+Na]⁺: 413.1935, Found: 413.1942;

HPLC: Chiralpak IF-3 column, n-hexane/isopropanol = 90/10, flow rate = 1.0 mL/min, λ = 254 nm, retention time: *t*_{major} = 11.736 min, *t*_{minor} = 20.430 min, 78% ee; [α]_D^{19.7} = -3.7 (c = 2.7, CHCl₃).



To a stirred solution of **4a** (30.1 mg, 0.093 mmol, 1 equiv.) in dry DCM (2 mL) was added DMAP (11.4 mg, 1.0 equiv.), Et₃N (52.0 μL, 4.0 equiv.) and bromobenzoyl chloride (82.1 mg, 4.0 equiv.) successively at r.t. and stirred overnight before it was quenched with the saturated aqueous NaHCO₃. The organic layer was separated and the aqueous layer was extracted with DCM. The combined organic layer was washed with brine, dried over MgSO₄ and concentrated under vacuum. The residue was purified via column chromatography on silica gel to give product compound **5a** as a colorless solid (58.3 mg, 0.067 mmol, 72% yield).

Mp 159.9-162.3 °C;

¹H NMR (400 MHz, CDCl₃) δ 8.05-8.02 (m, 2H), 7.76 (dd, *J* = 0.2 Hz, 8.0 Hz, 4H), 7.67-7.65 (m, 2H), 7.56-7.50 (m, 4H), 7.03 (d, *J* = 8.4 Hz, 11.6Hz, 2H), 5.50 (t, *J* = 6.8Hz, 1H), 4.13-4.08 (m, 2H), 3.74 (q, *J* = 3.6 Hz, 1H), 3.62 (s, 3H), 2.51-2.27 (m, 2H), 2.14-1.90 (m, 4H), 1.86-1.64 (m, 4H);

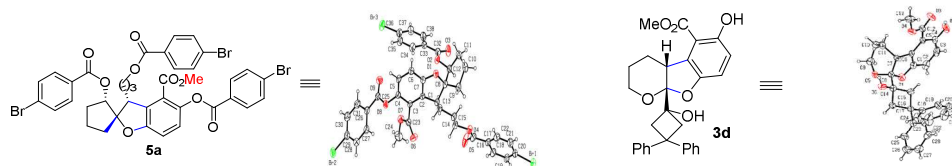
¹³C NMR (100 MHz, CDCl₃) δ 165.6, 165.2, 165.0, 156.4, 143.2, 133.1, 132.0, 131.8, 131.6, 131.1, 131.0, 128.9, 128.8, 128.7, 128.2, 123.5, 113.6, 97.6, 75.3, 64.9, 52.1, 47.9, 36.5, 29.7, 29.1, 27.2, 26.2, 18.6;

HRMS ESI Calcd for C₃₈H₃₁Br₃O₉Na [M+Na]⁺: 890.9410, Found: 890.9399;

HPLC: Chiralpak IF-3 column, n-hexane/isopropanol = 95/5, flow rate = 1.0 mL/min, λ = 254 nm,

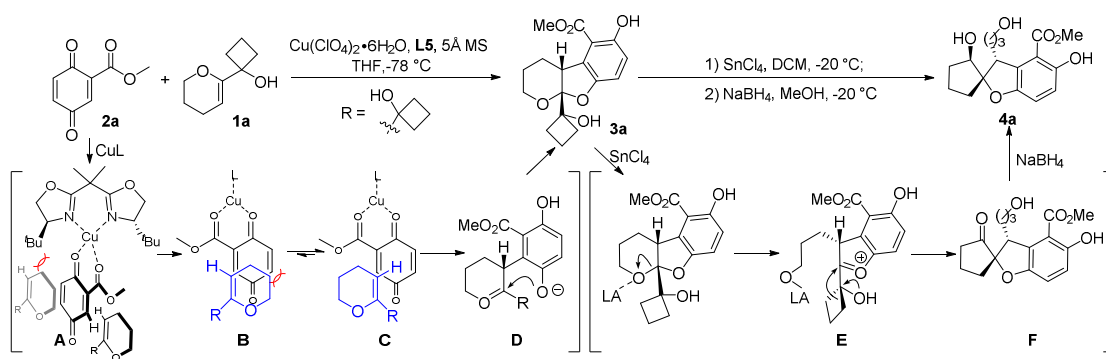
retention time: $t_{\text{major}} = 30.810$ min, $t_{\text{minor}} = 36.724$ min, 94% ee; $[\alpha]_{\text{D}}^{20.7} = 42.9$ ($c = 1.4$, CHCl_3).

8. X-Ray Ellipsoid Plots of 3d and 5a



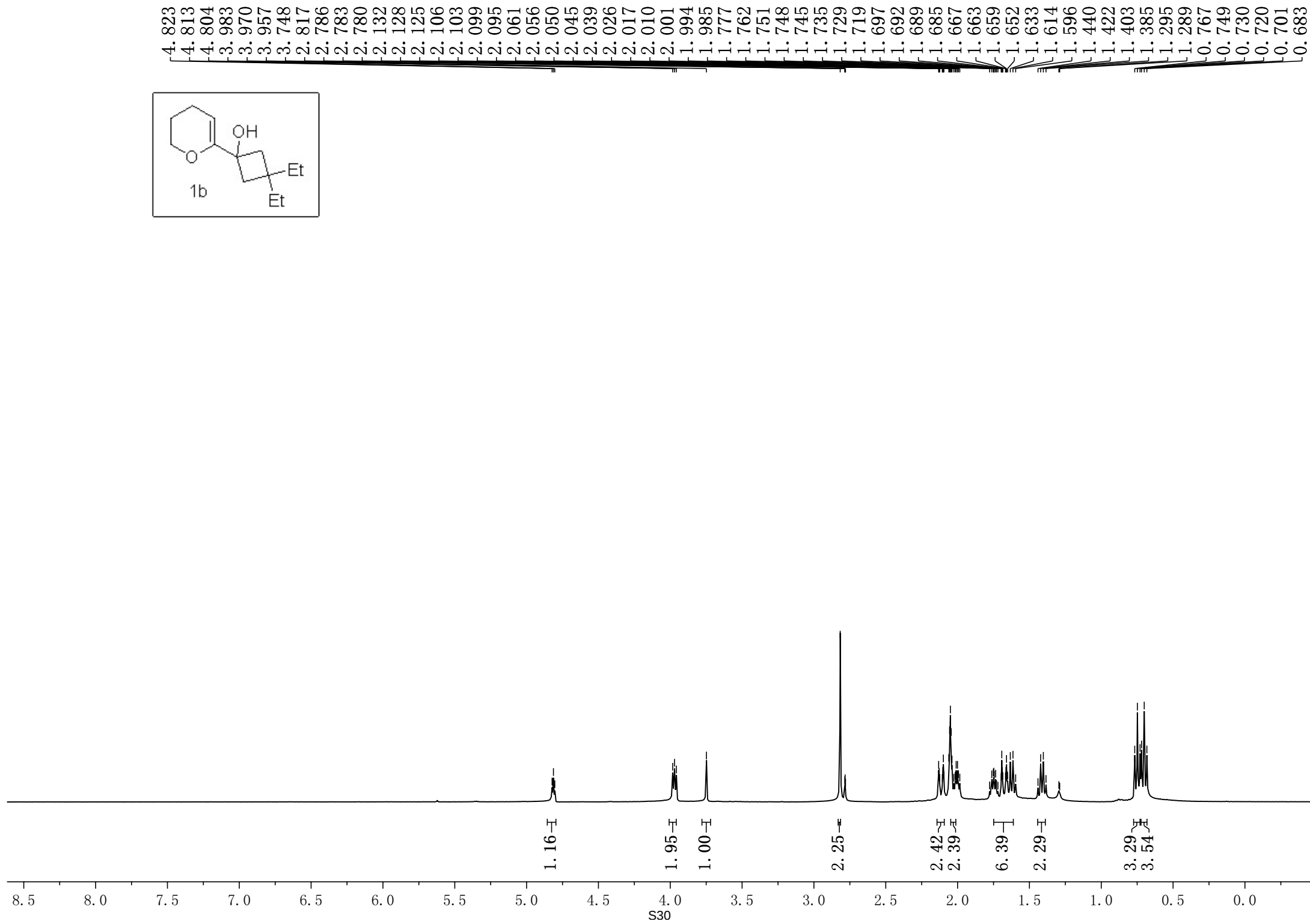
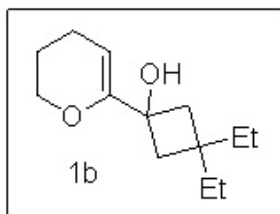
The crystal structure of **3d** and **5a** have been deposited at the Cambridge Crystallographic Data Centre and allocated the deposition numbers (CCDC number): 1883807 and 1883805.

9. Proposed Reaction Mechanism

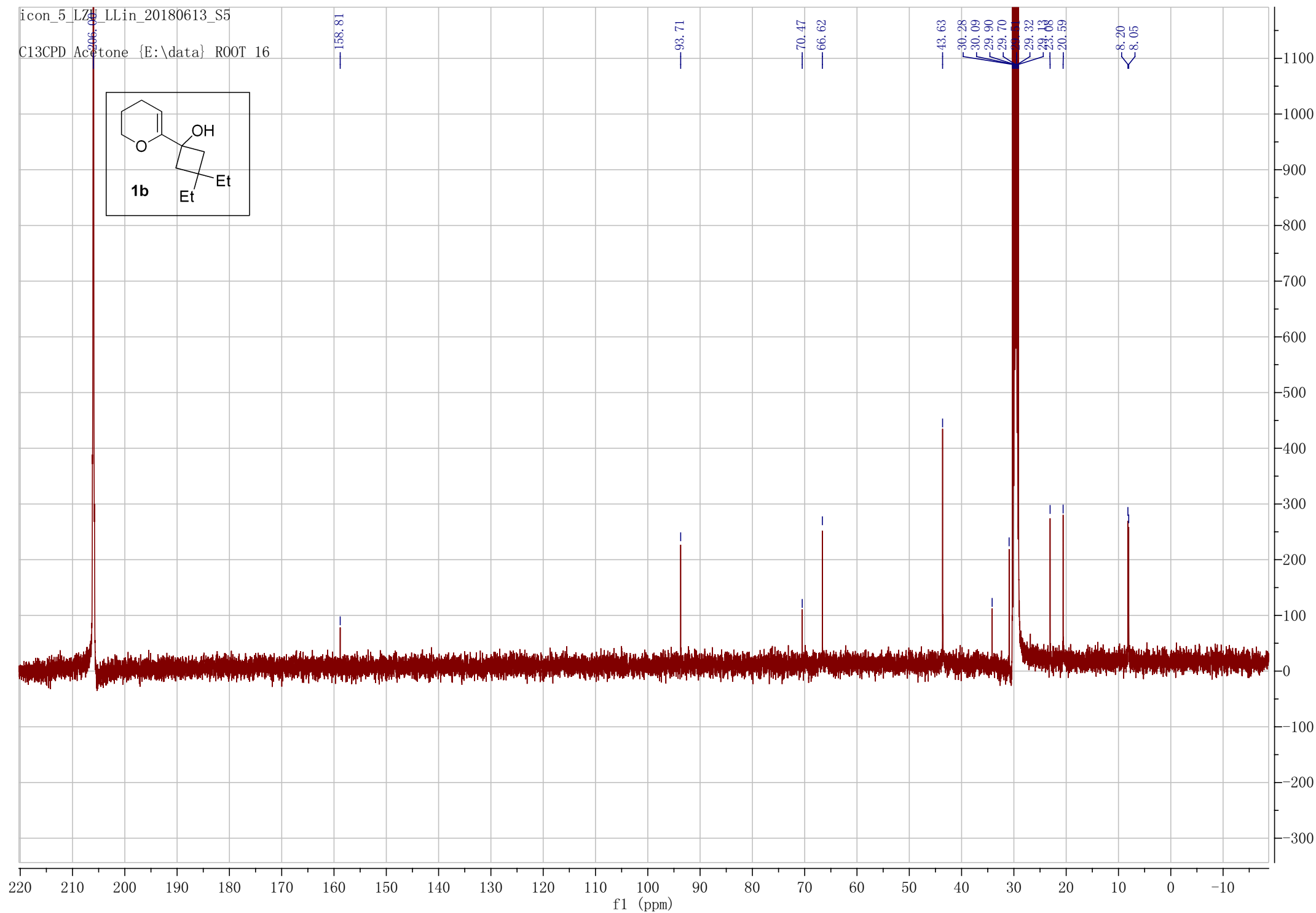
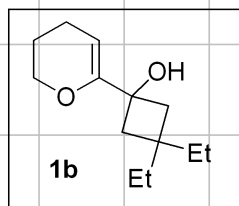


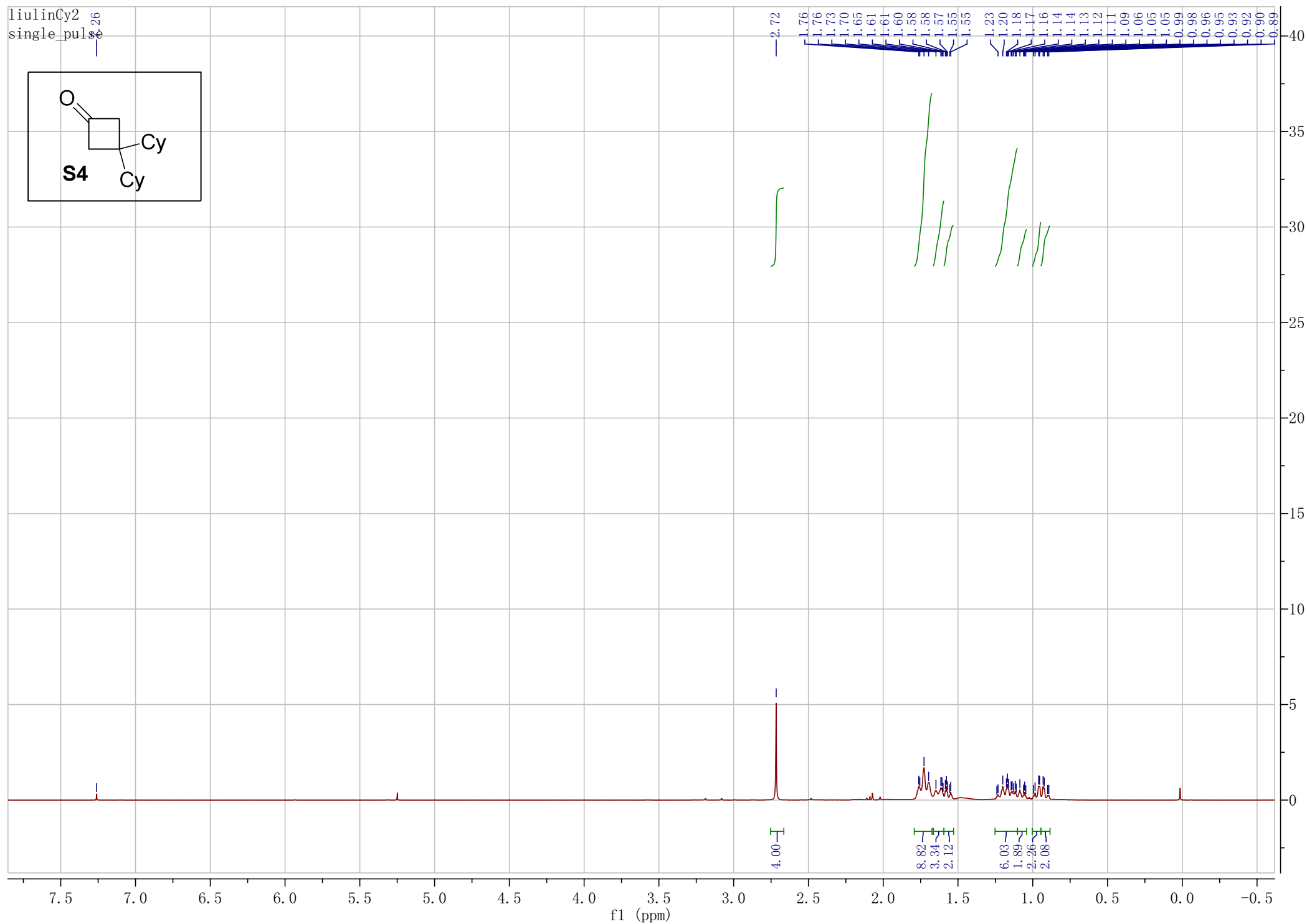
10. References

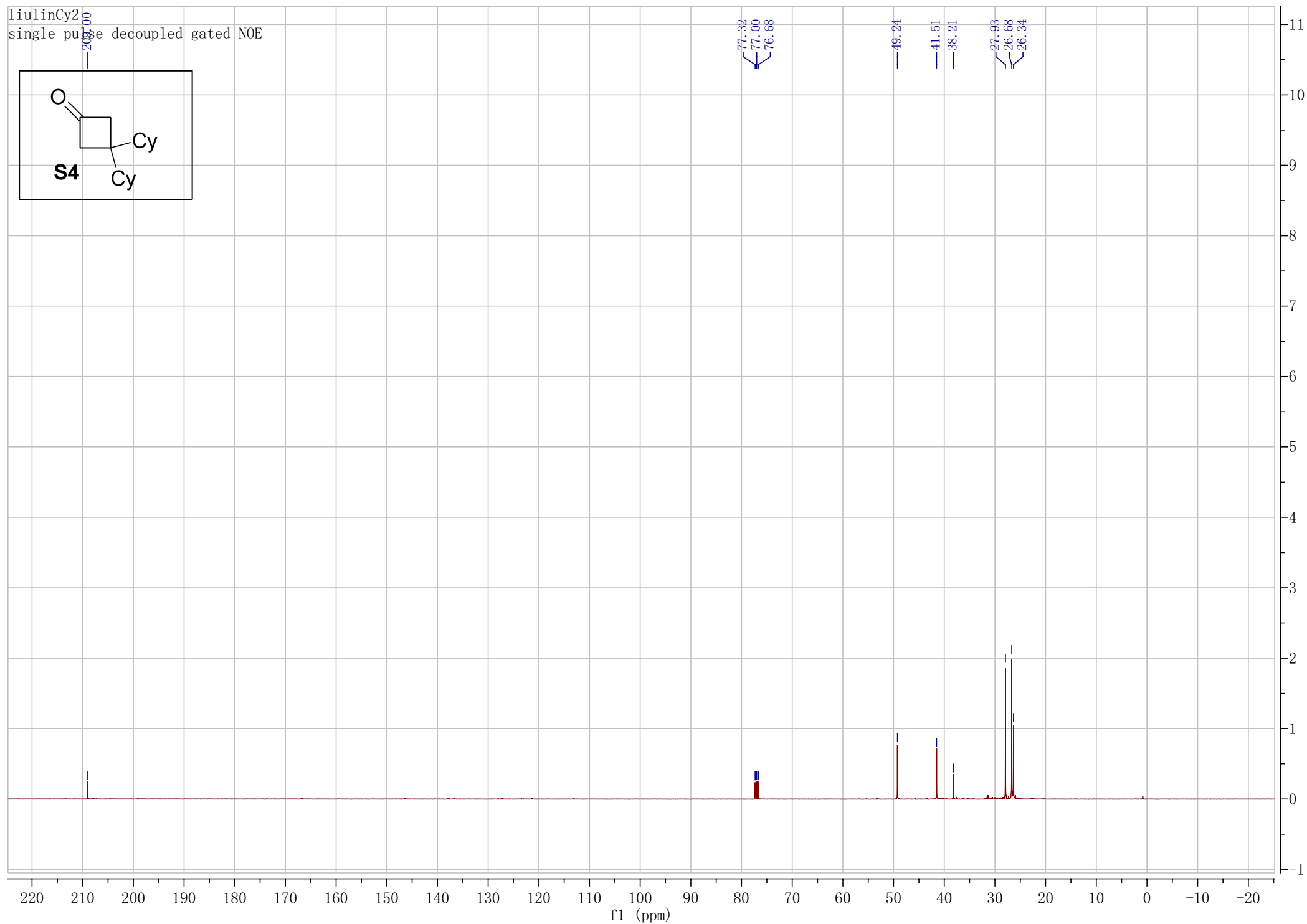
- [1] Q.-W. Zhang, C.-A. Fan, H.-J. Zhang, Y.-Q. Tu, Y.-M. Zhao, P. Gu, and Z.-M. Chen, *Angew. Chem. Int. Ed.* 2009, **48**, 8572-8574.
- [2] Y.-H. Chen, D.-J. Cheng, J. Zhang, Y. Wang, X.-Y. Liu, B. Tan, *J. Am. Chem. Soc.*, 2015, **137**, 15062–15065.



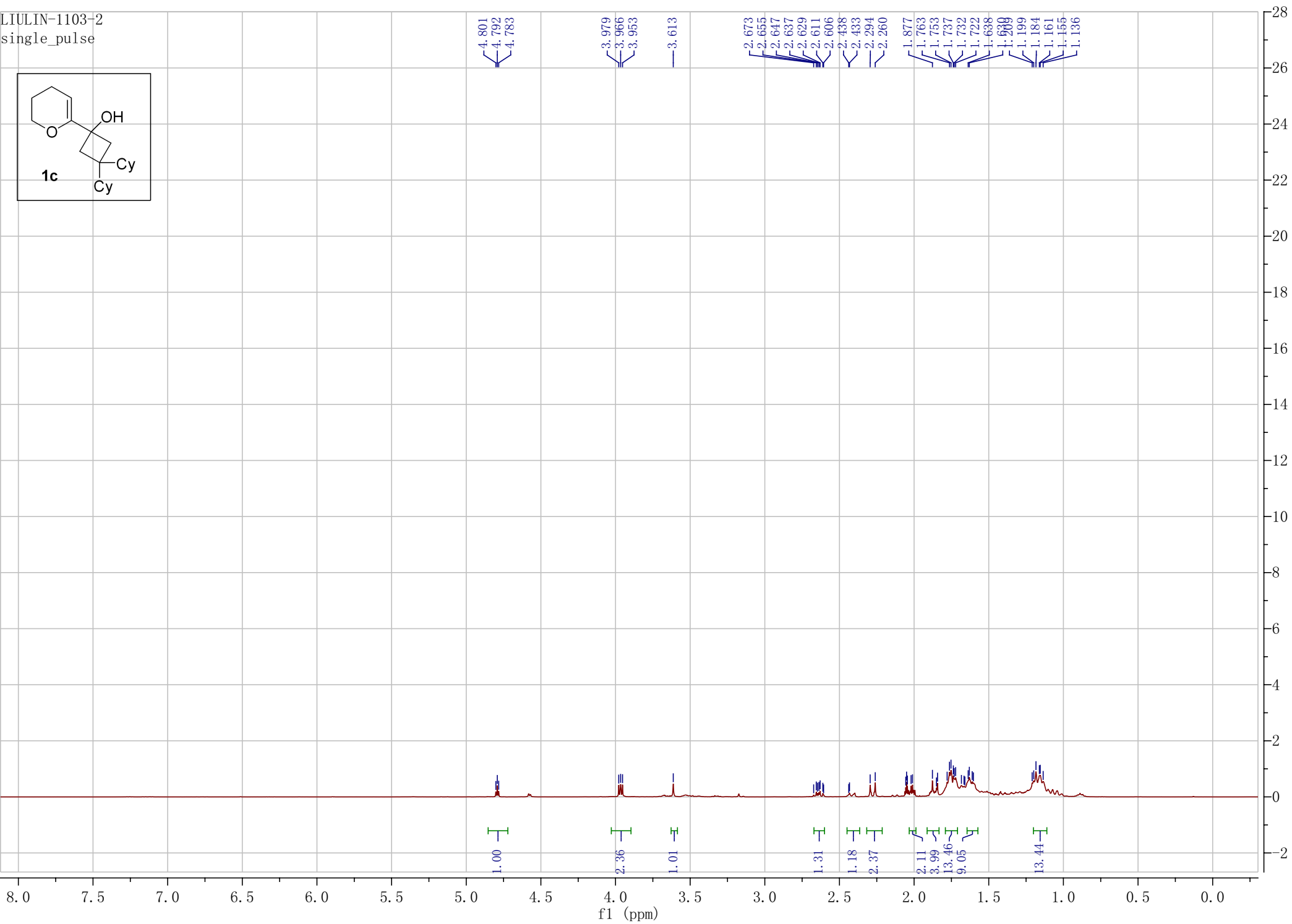
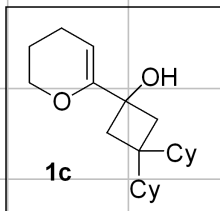
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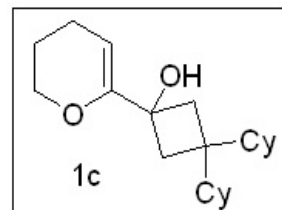






LIULIN-1103-2
single_pulse





—205.986

—158.949

—94.281

—70.318

—66.569

39.792

39.123

30.301

30.110

29.915

29.724

29.584

29.531

29.338

29.146

28.179

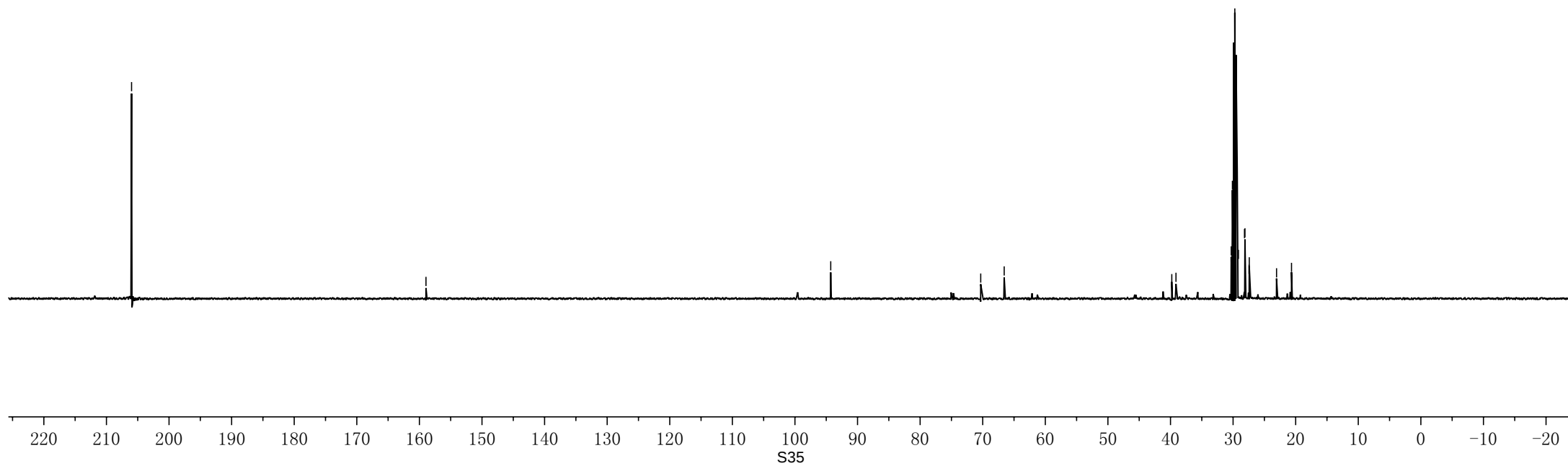
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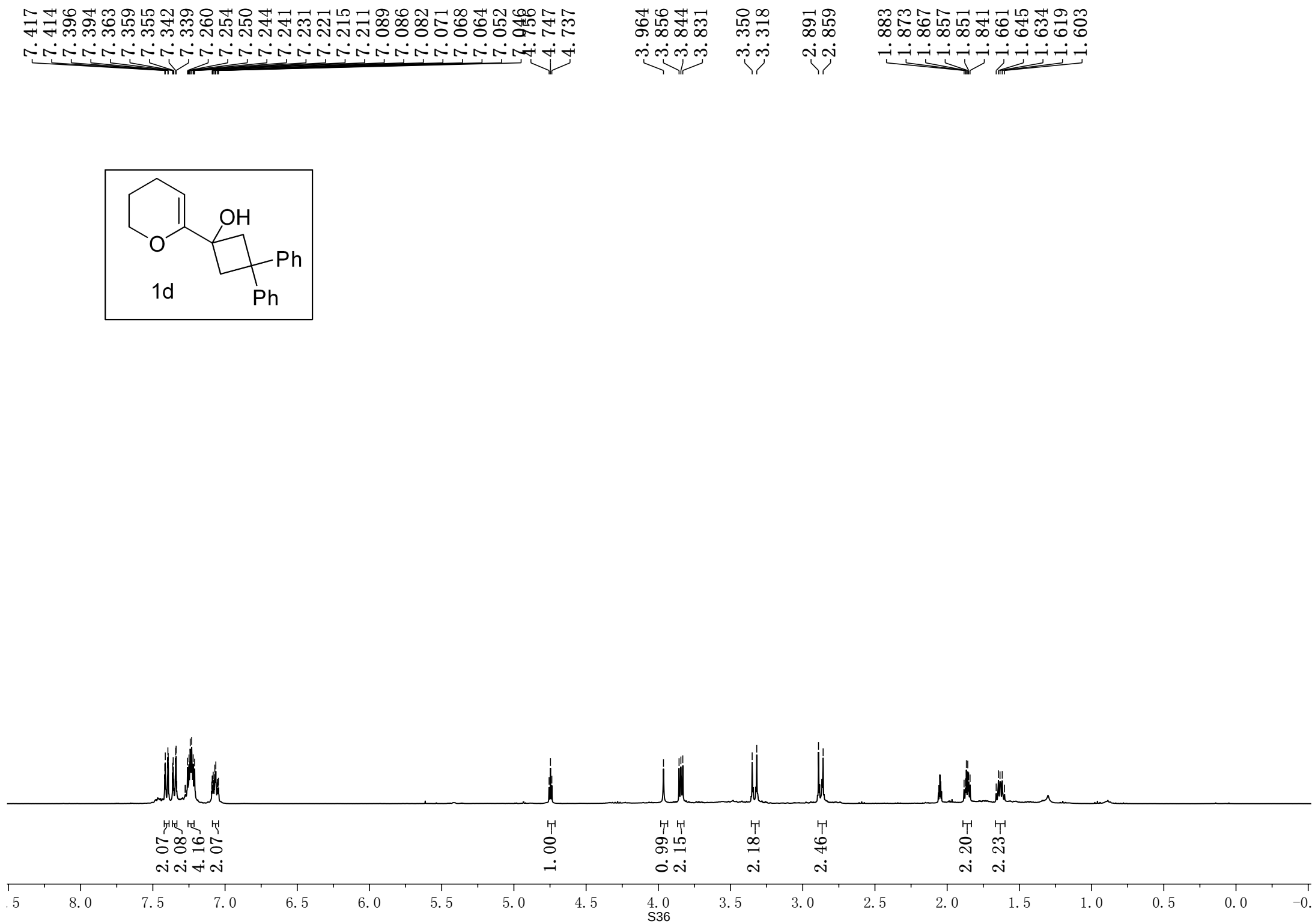
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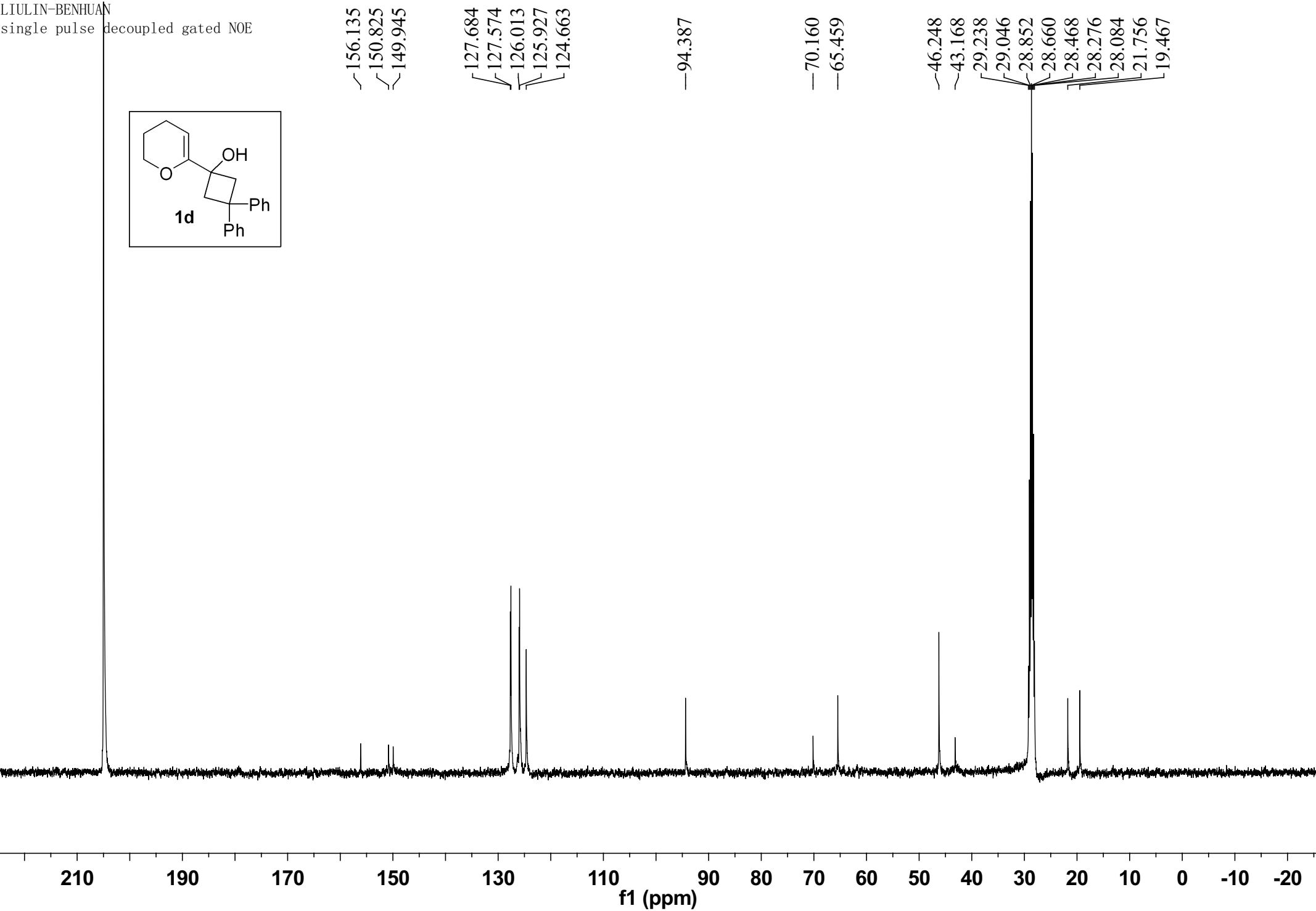
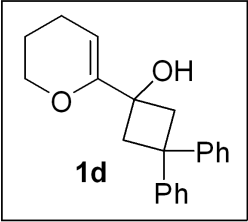
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20.665



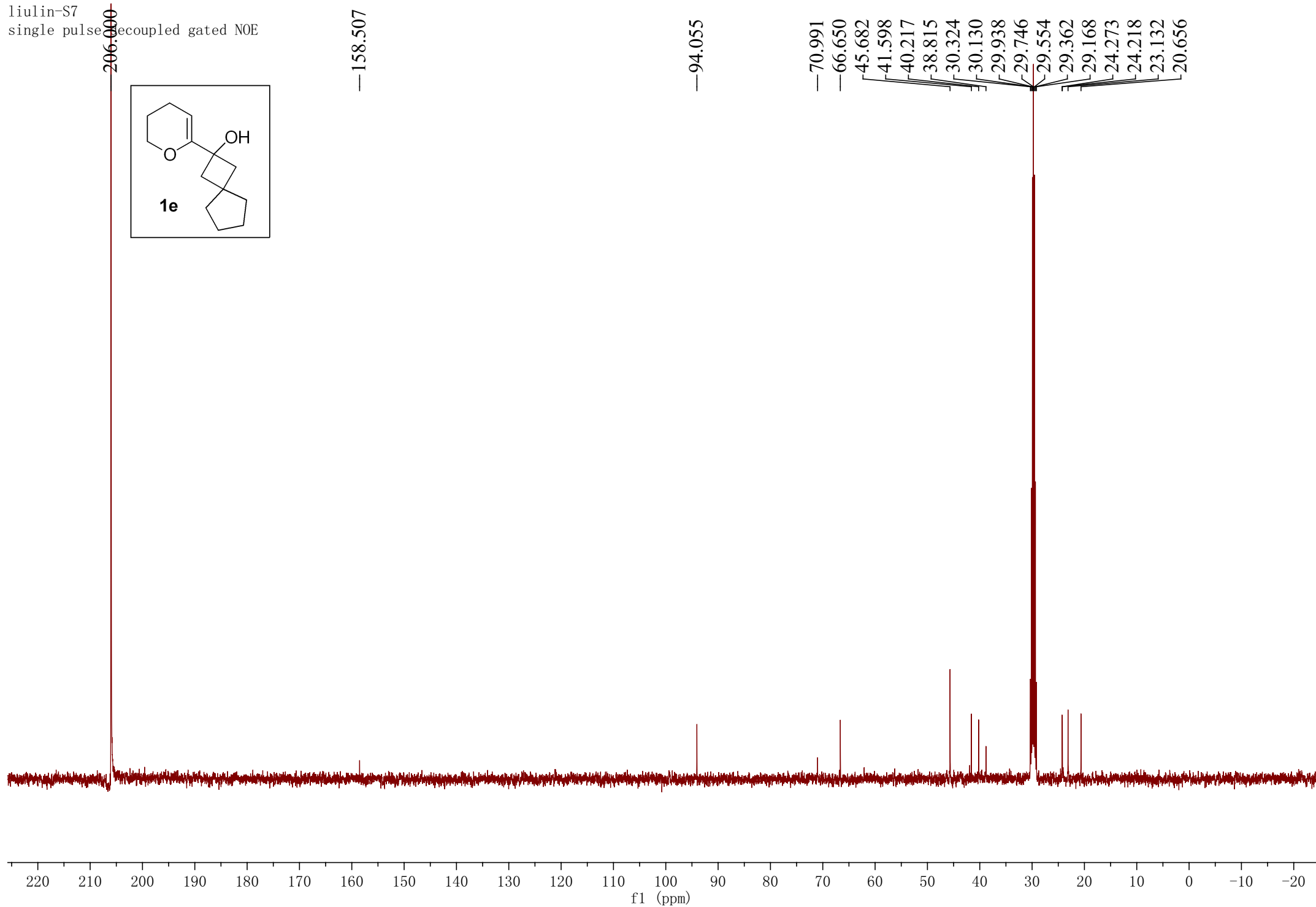
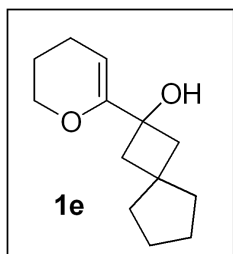


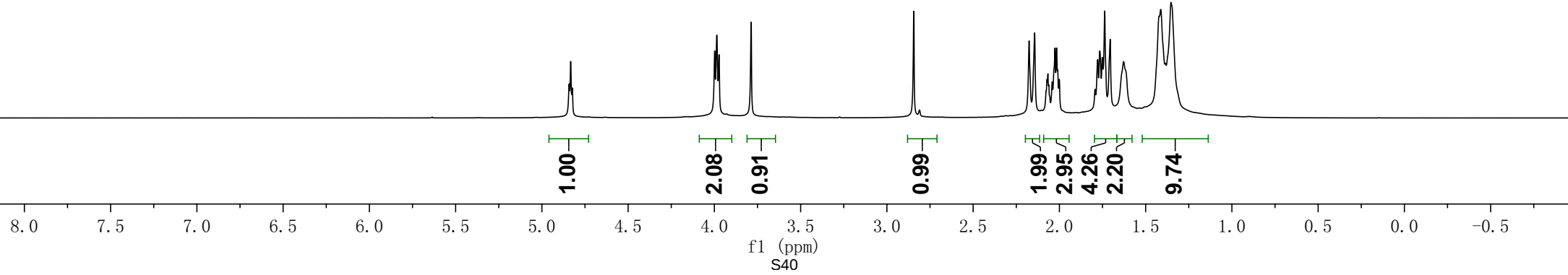
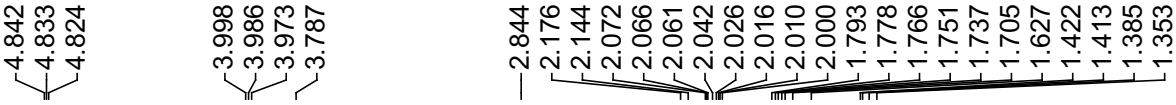
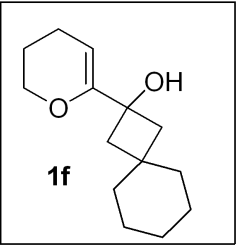
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single pulse decoupled gated NOE

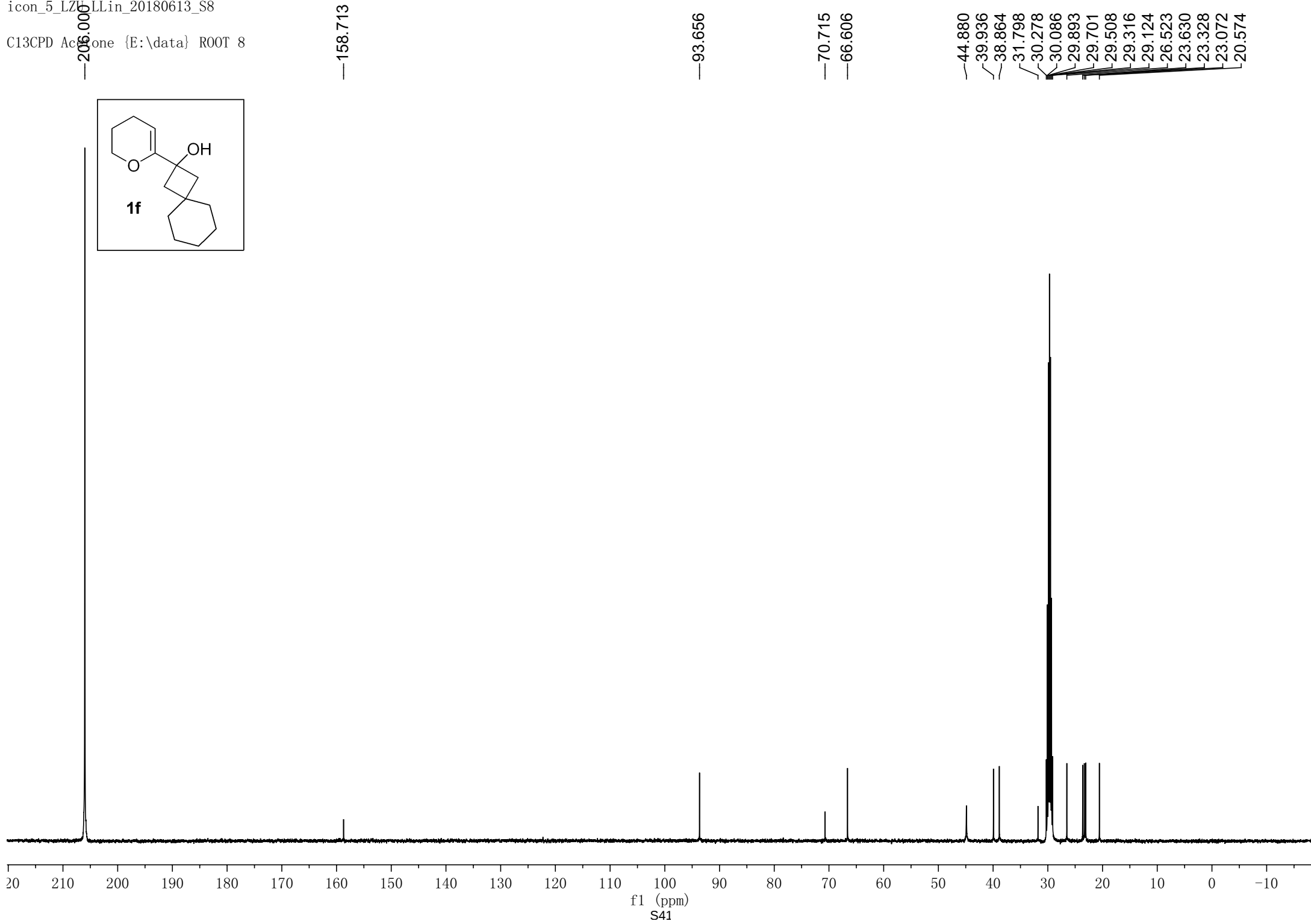




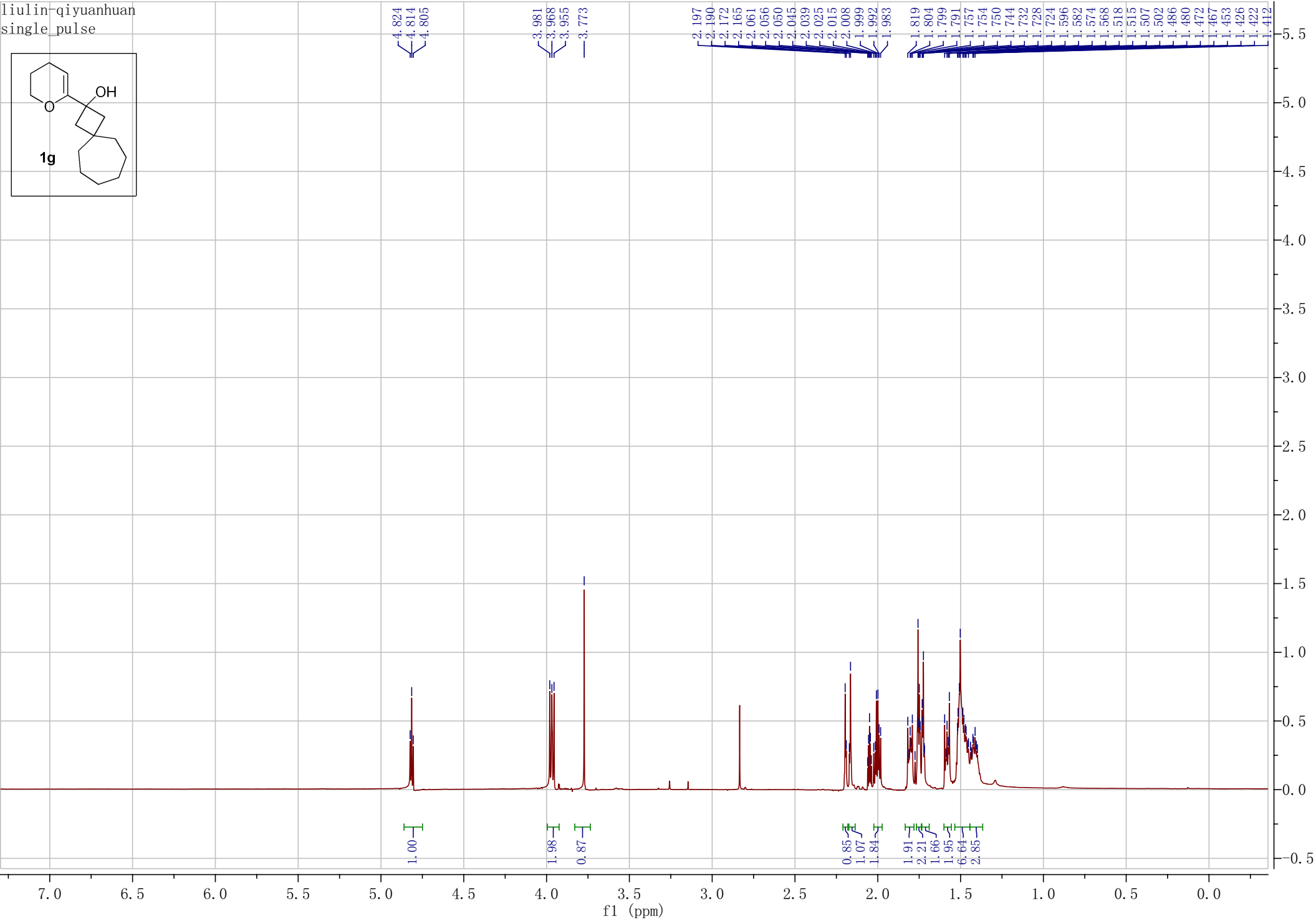
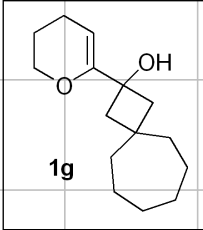
liulin-S7
single pulse decoupled gated NOE

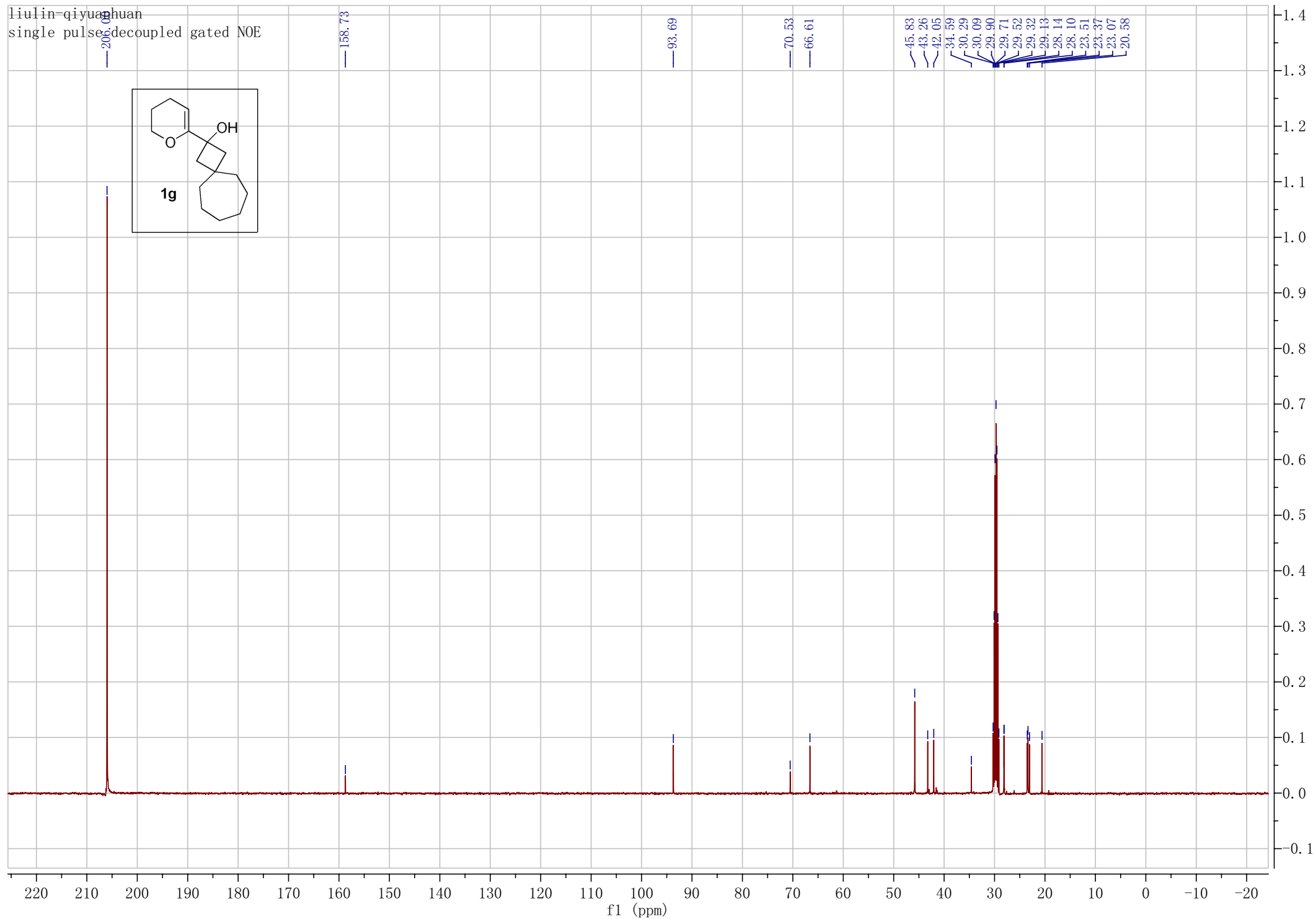


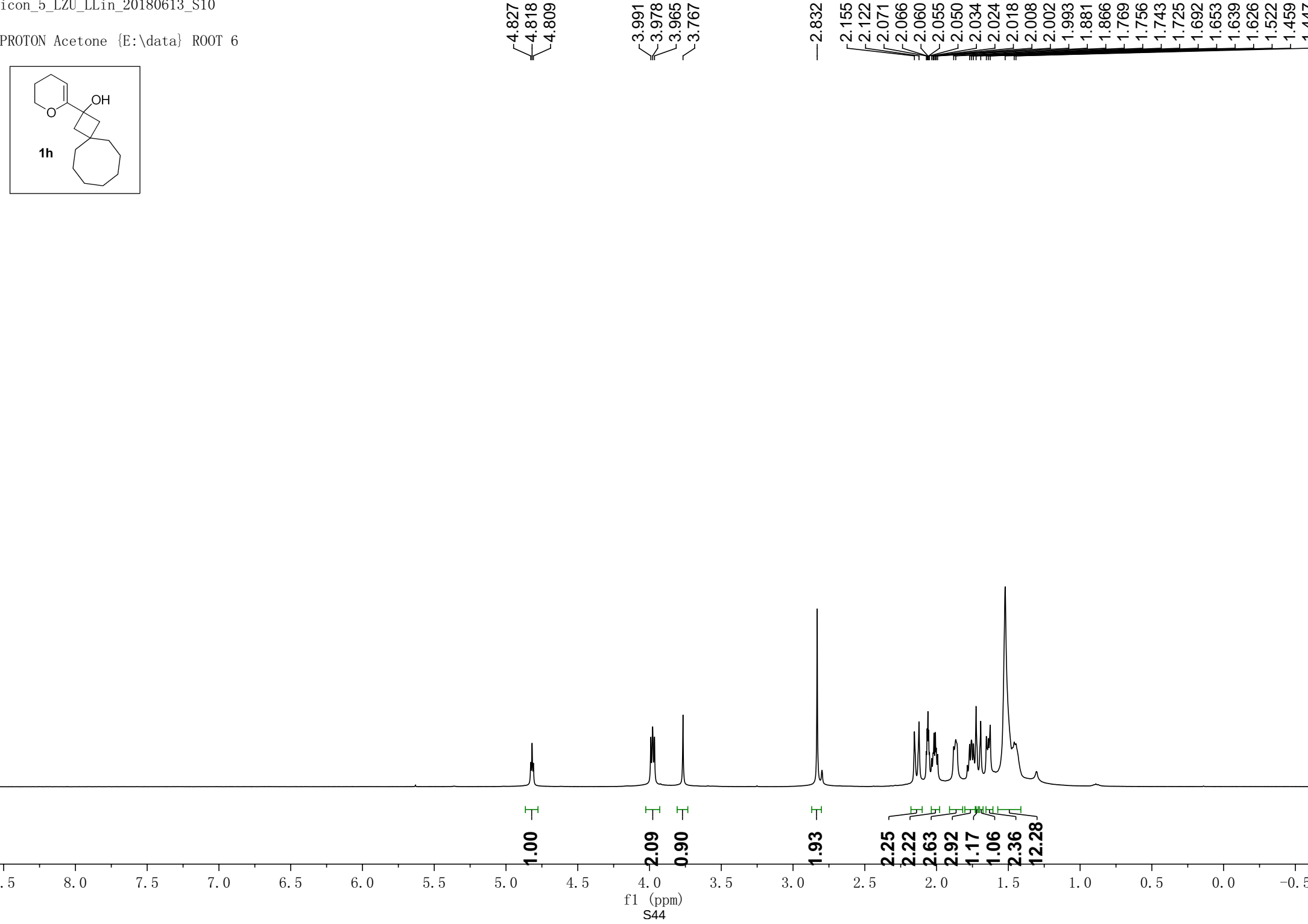
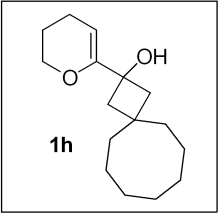




liulin-qiyuanhuan
single_pulse

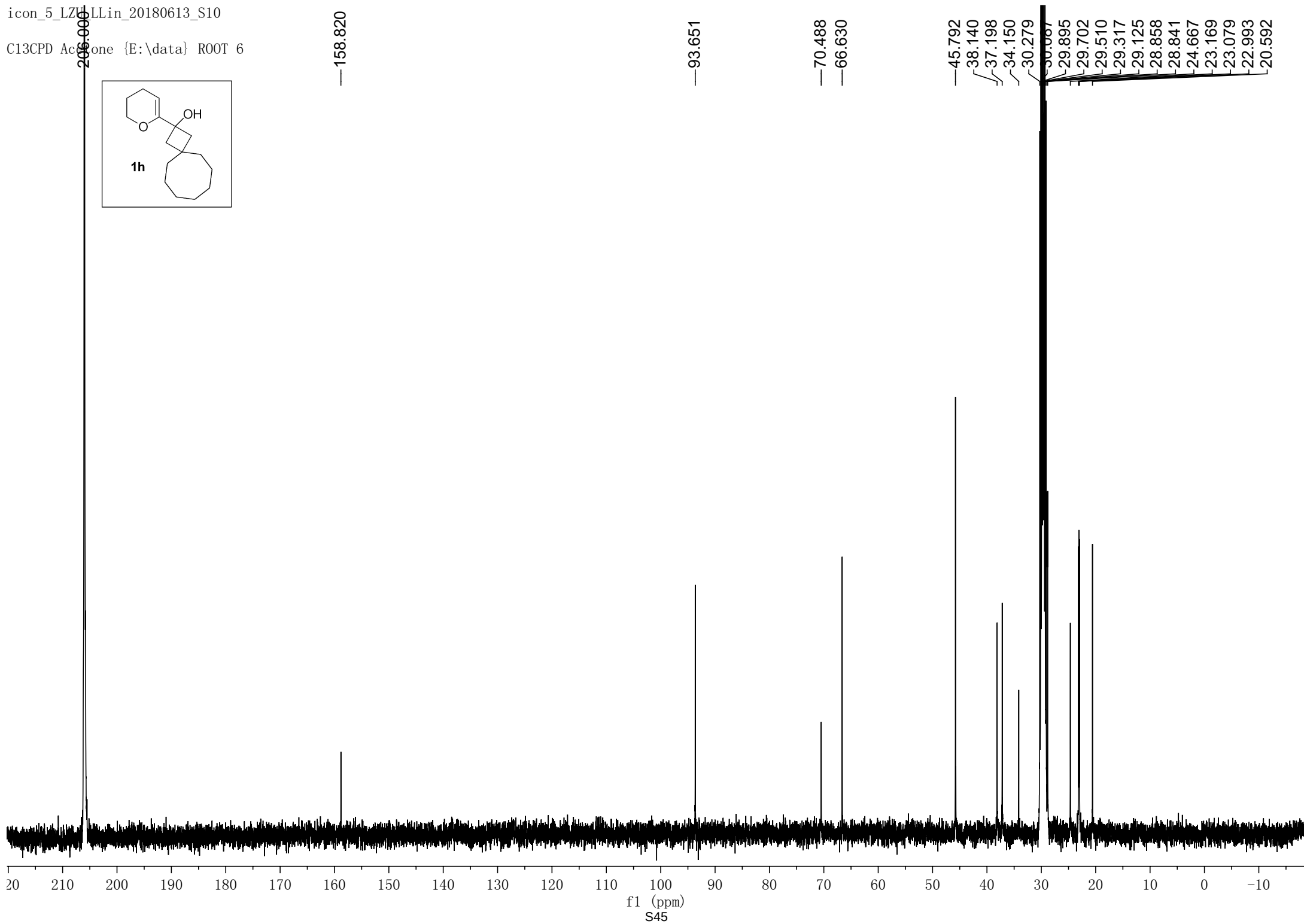
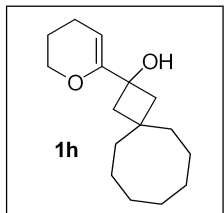


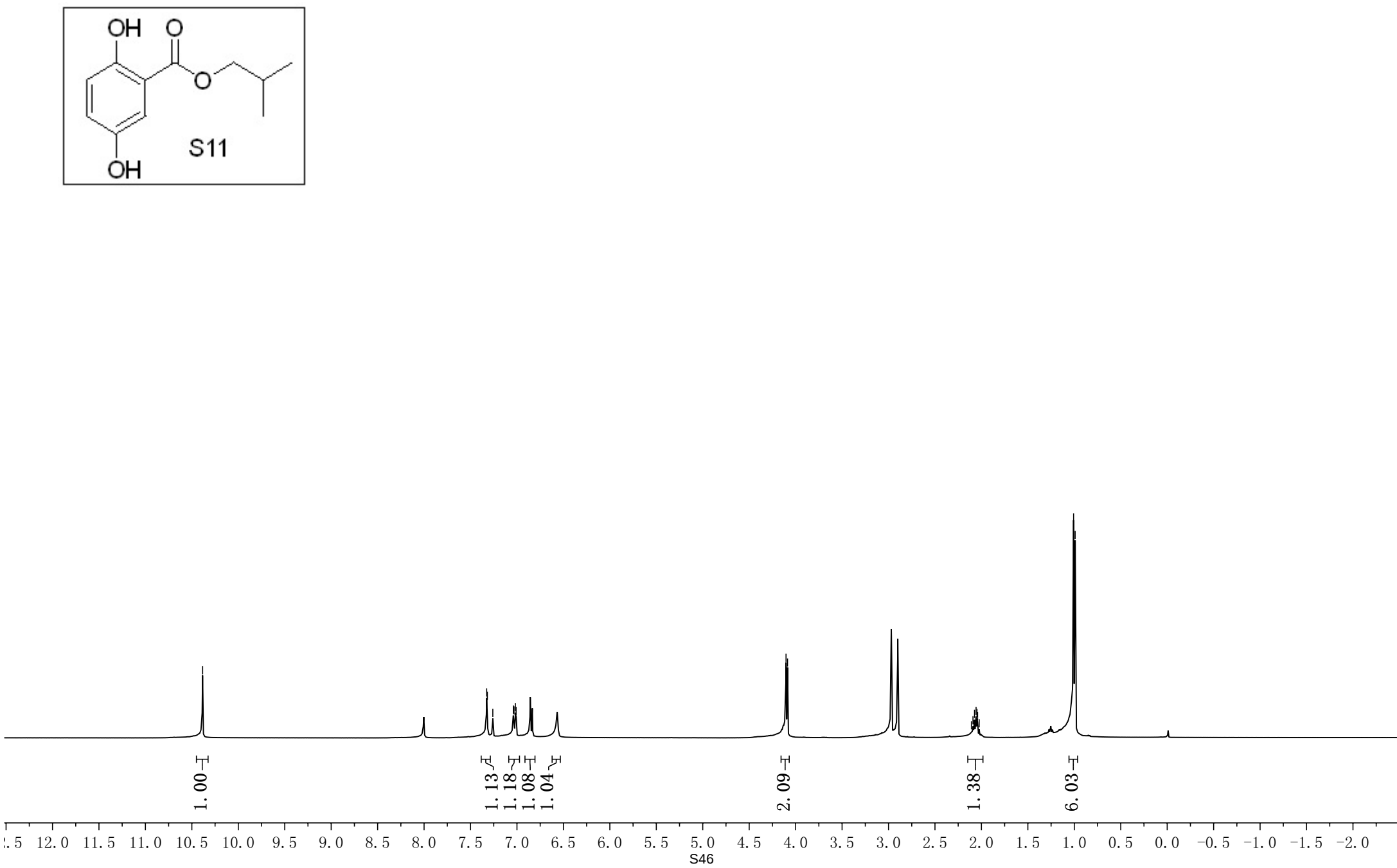


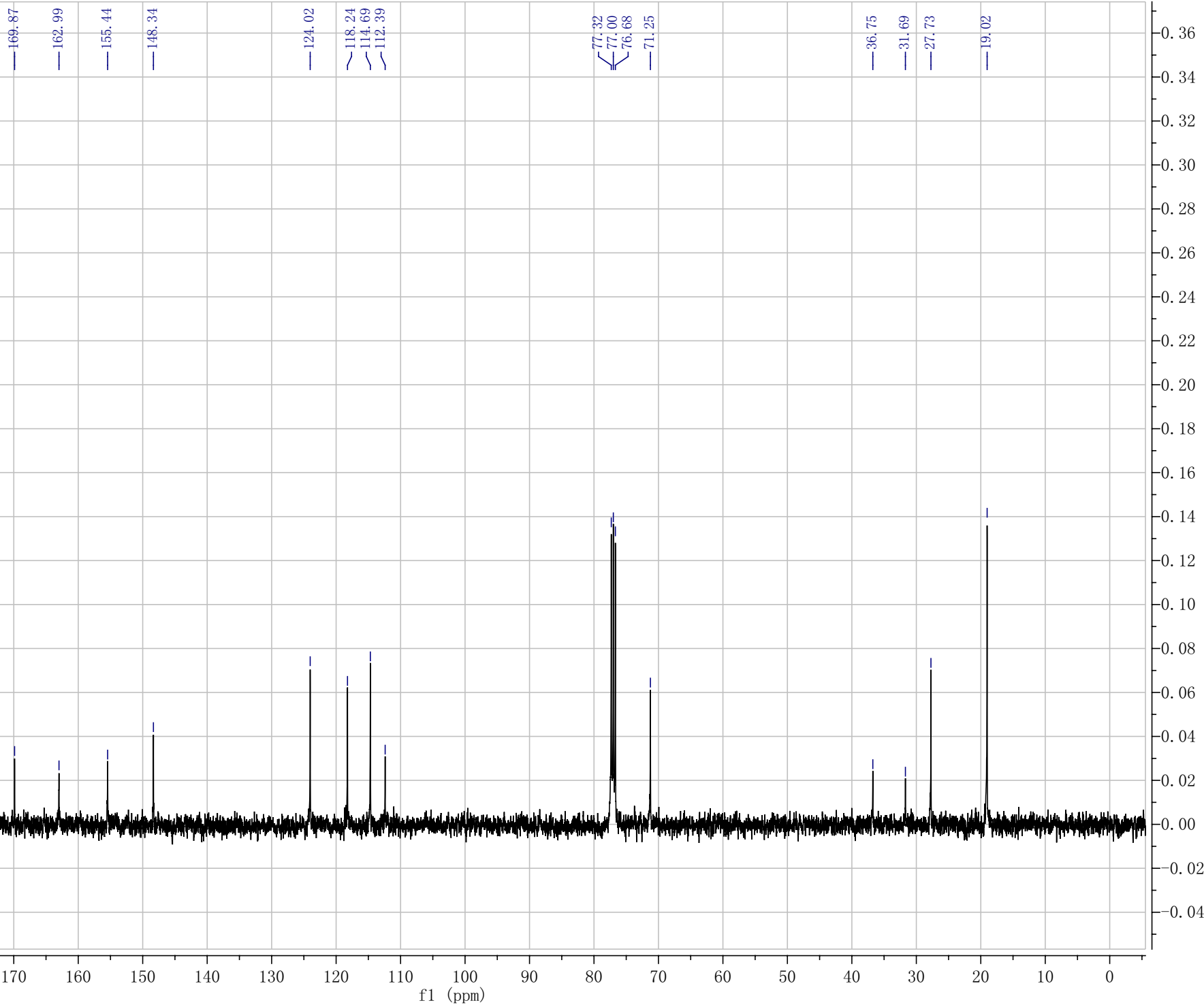
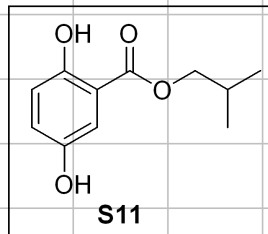


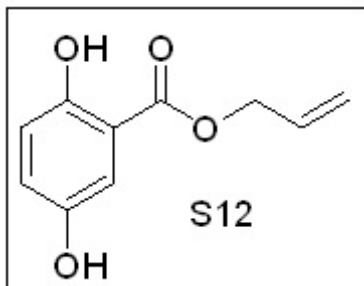
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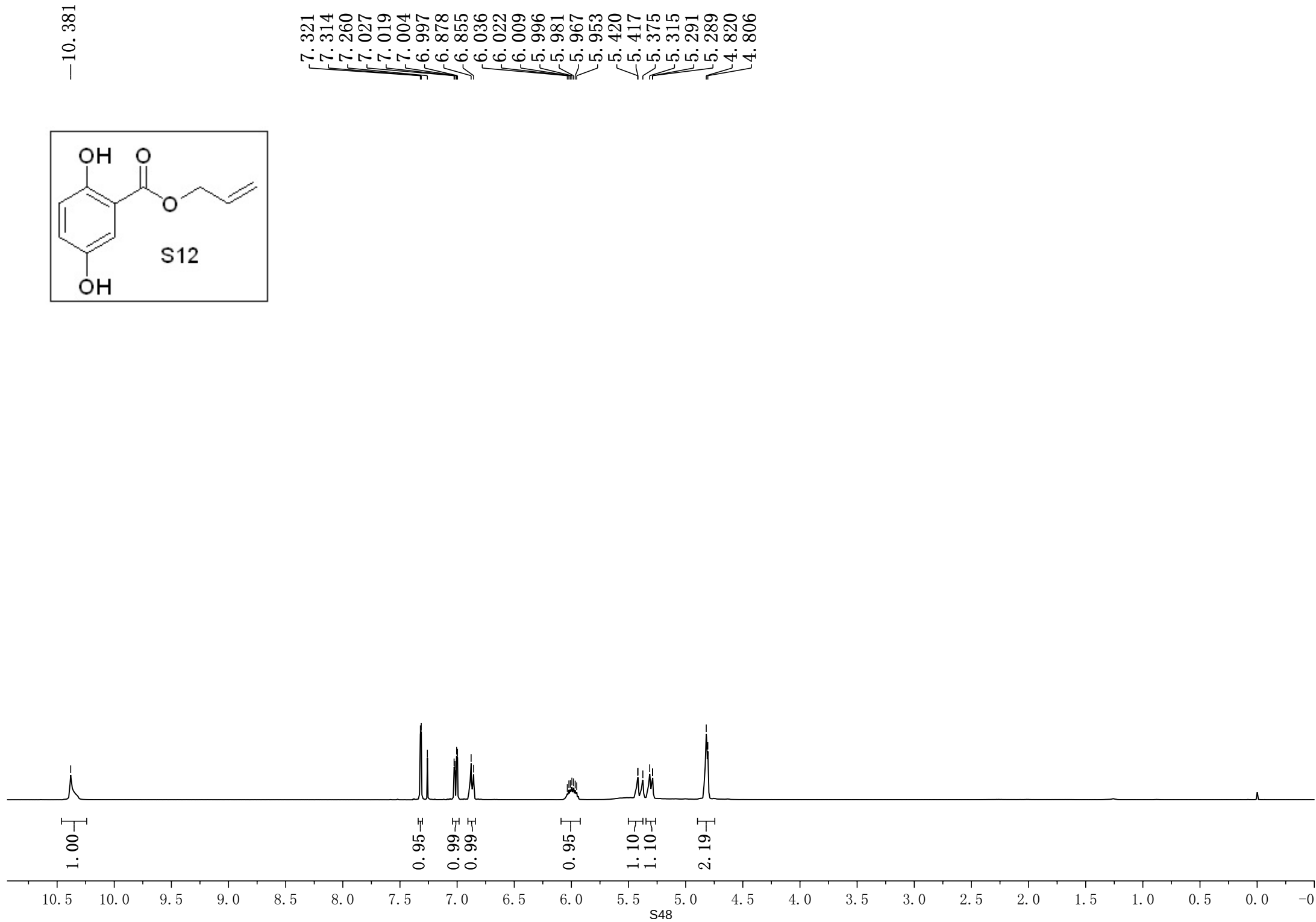


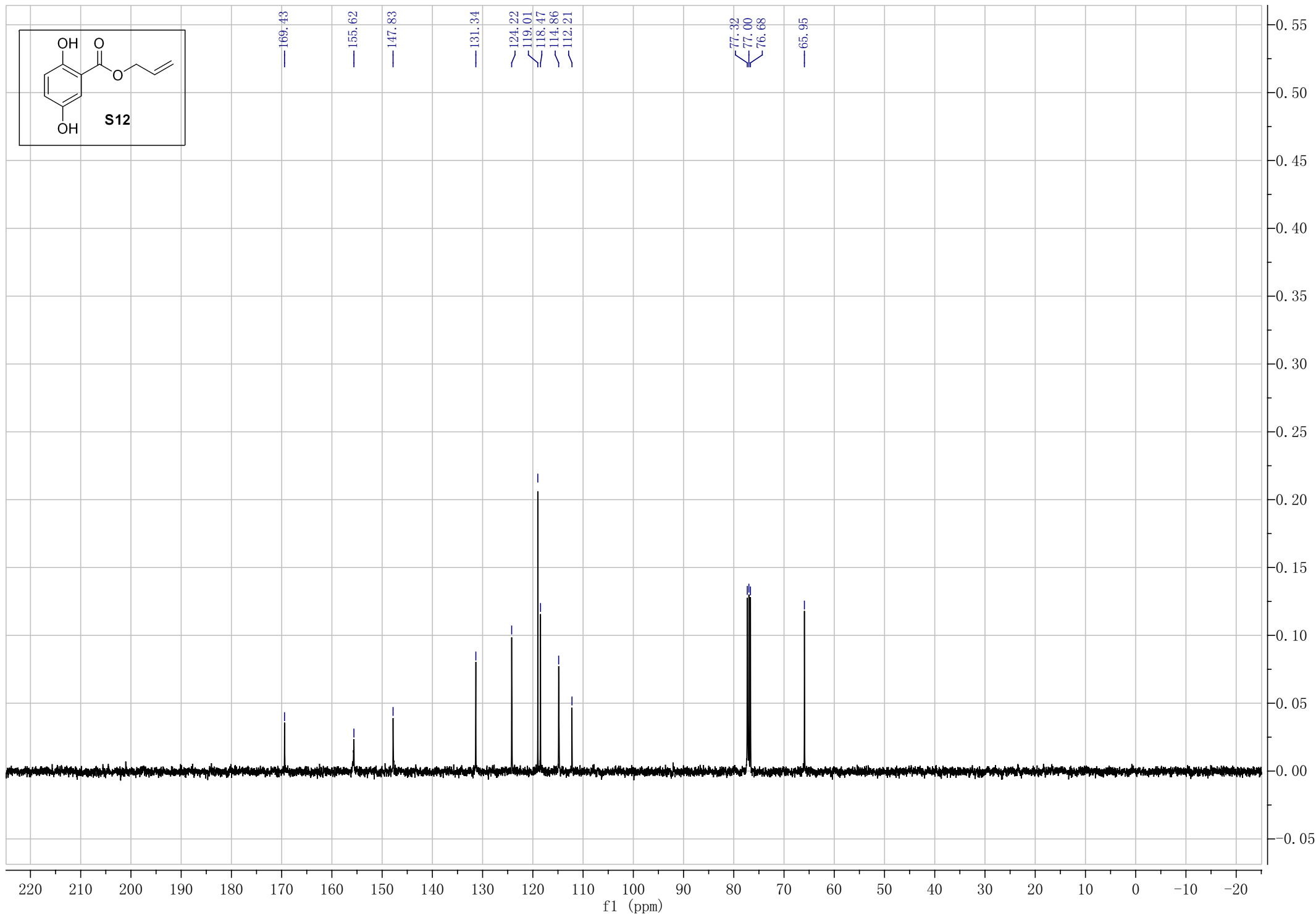
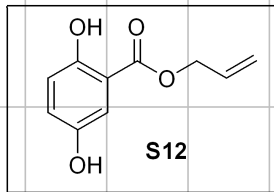


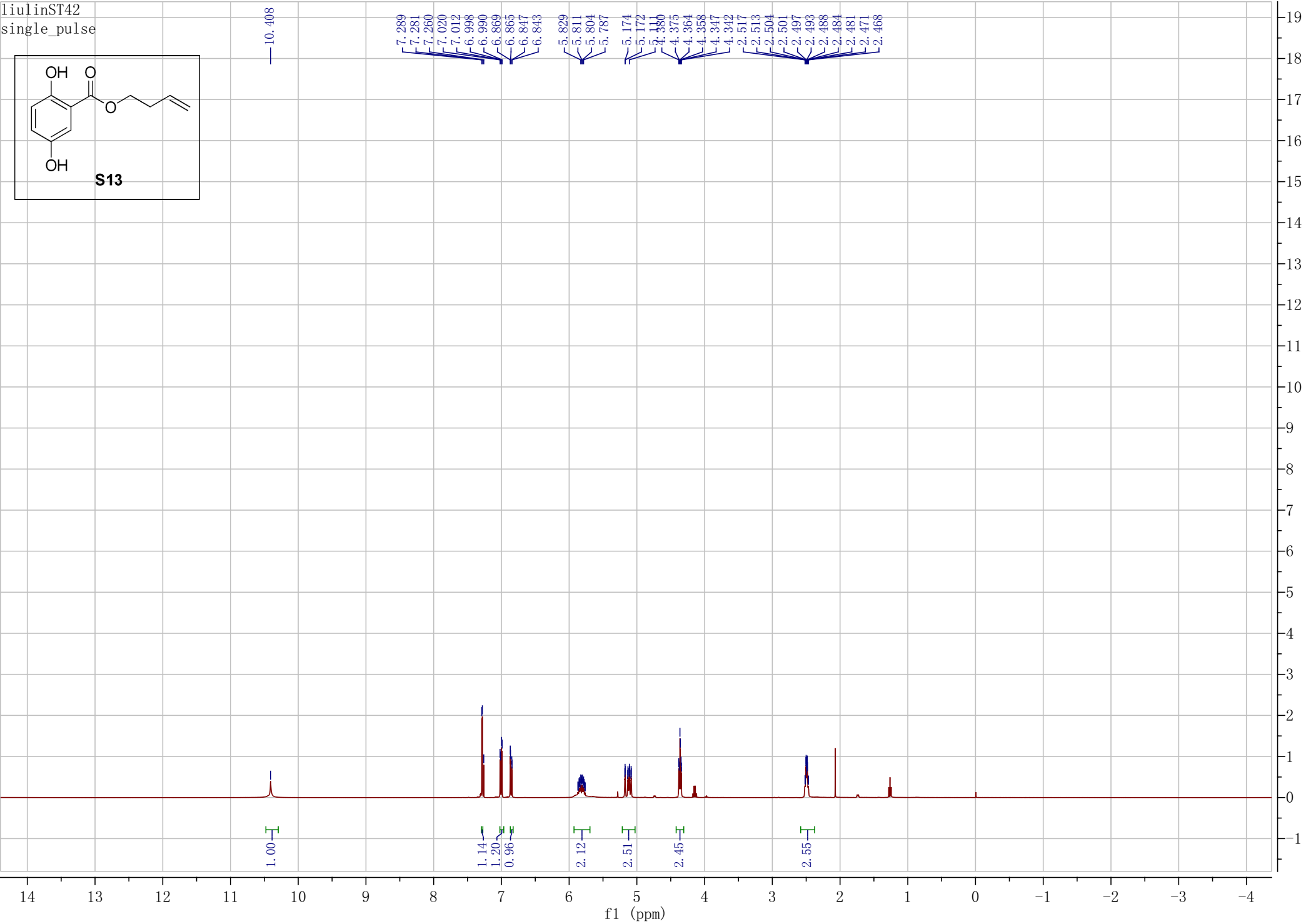


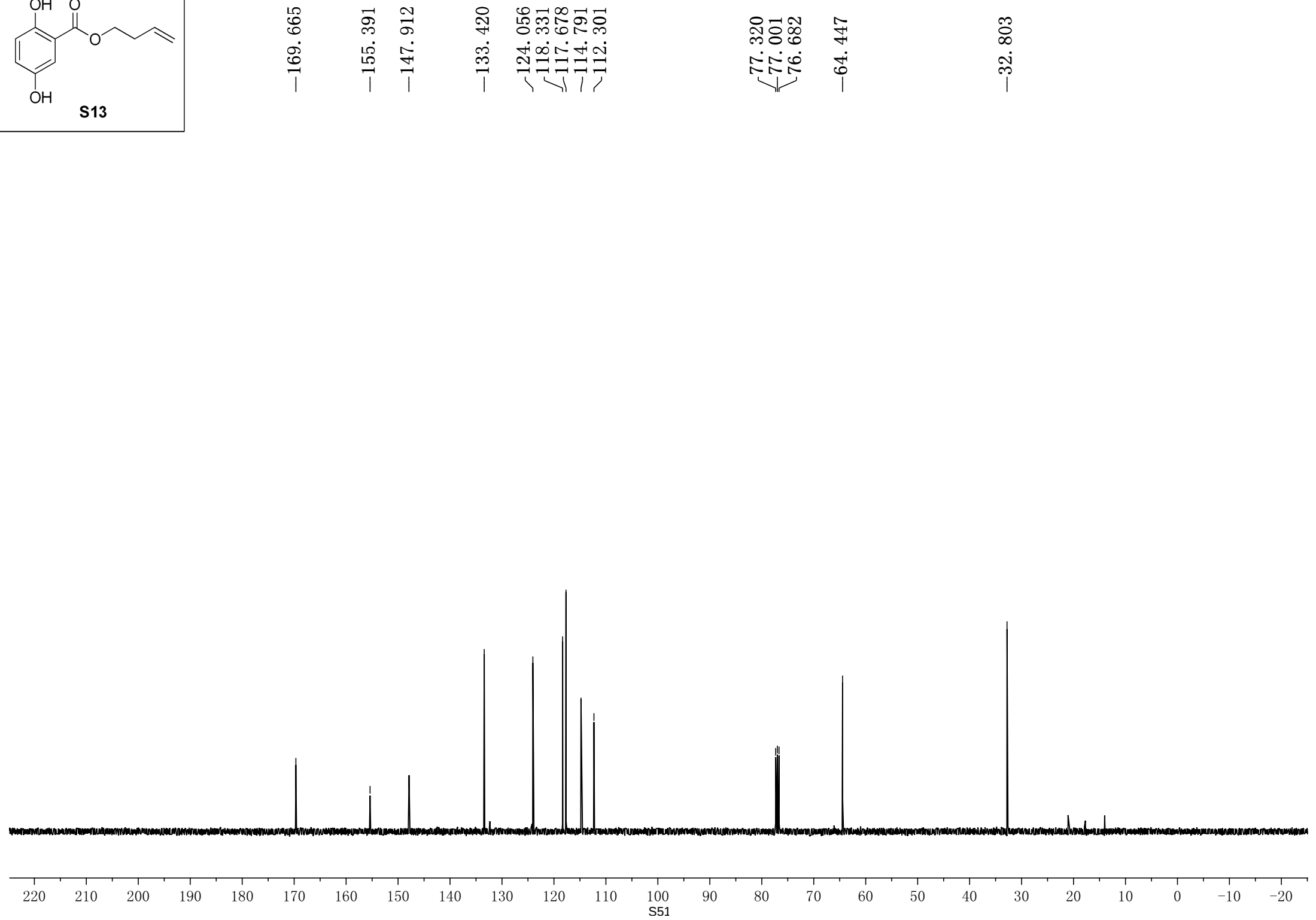
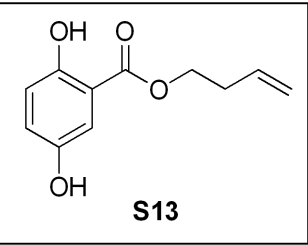


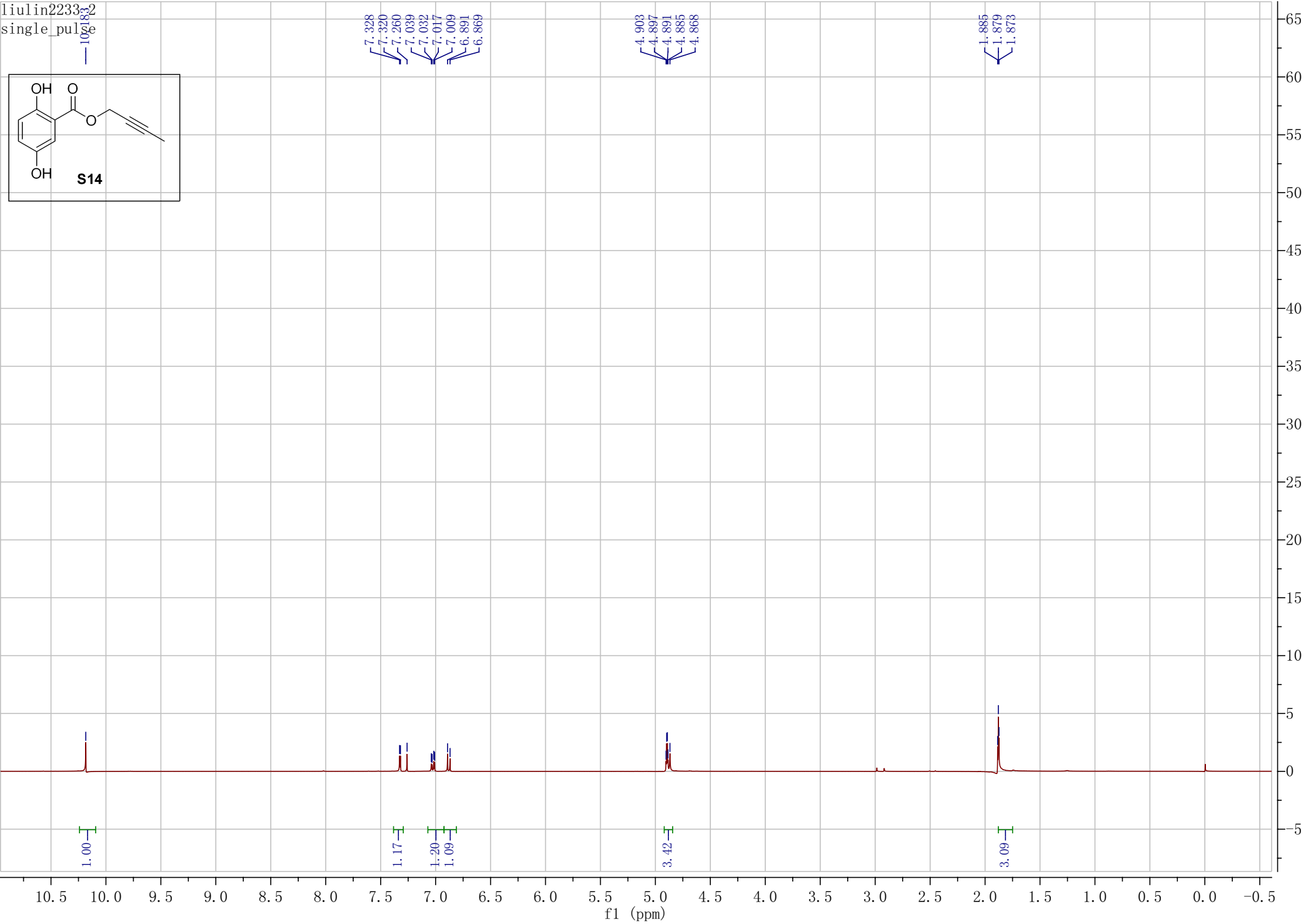
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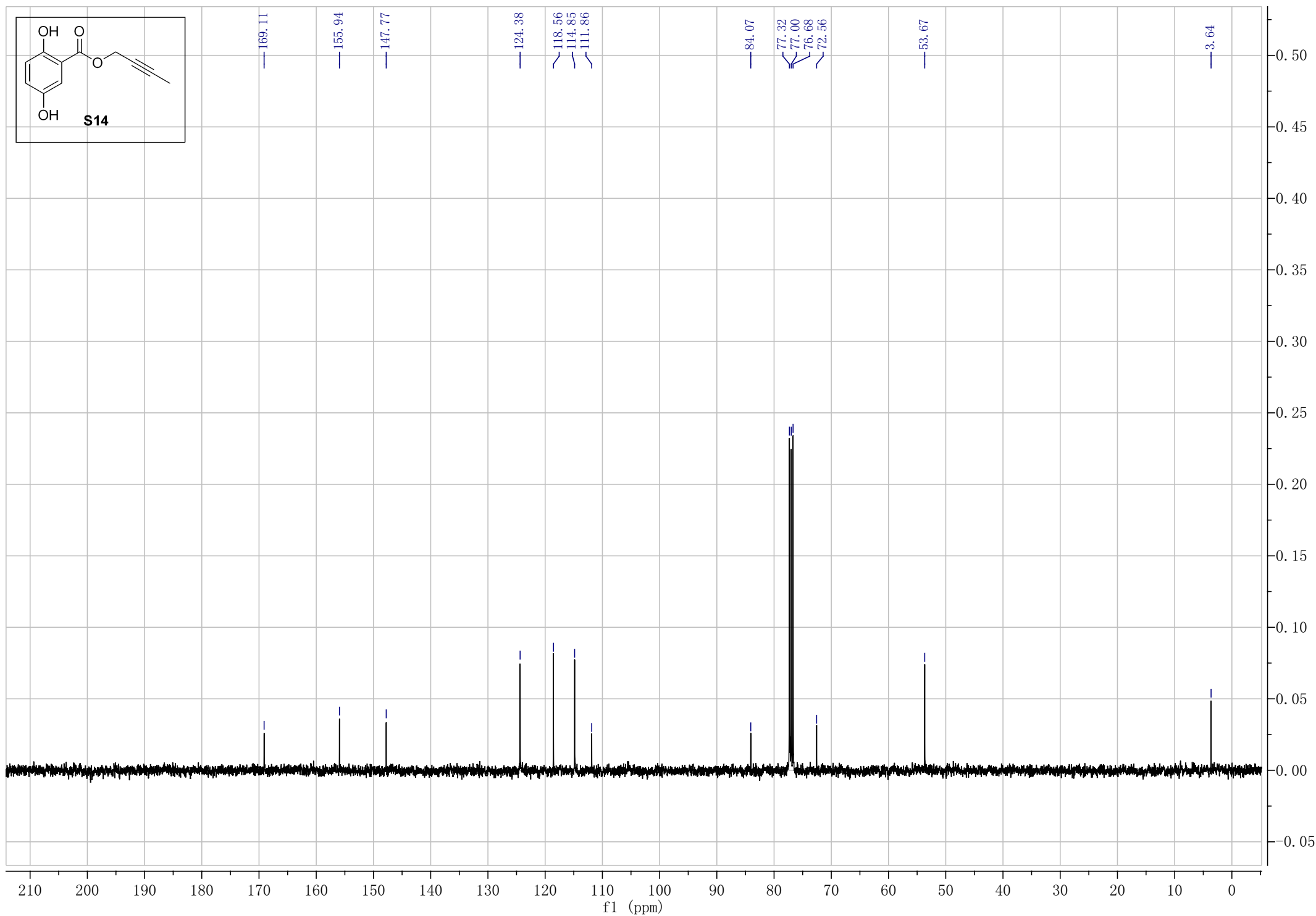


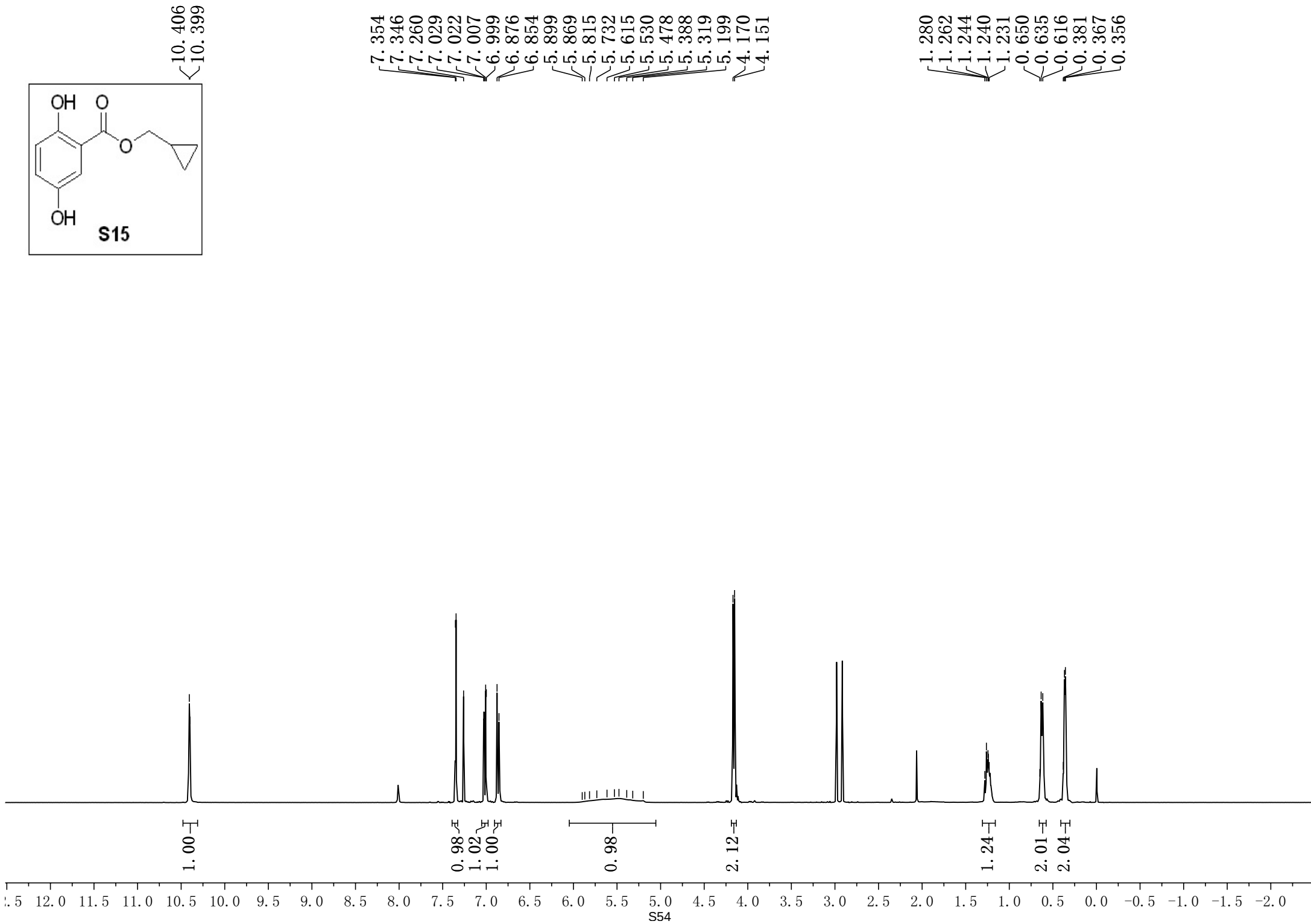
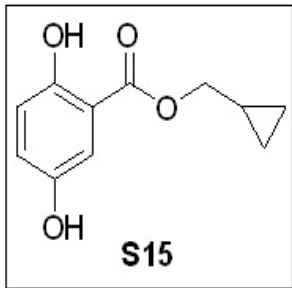


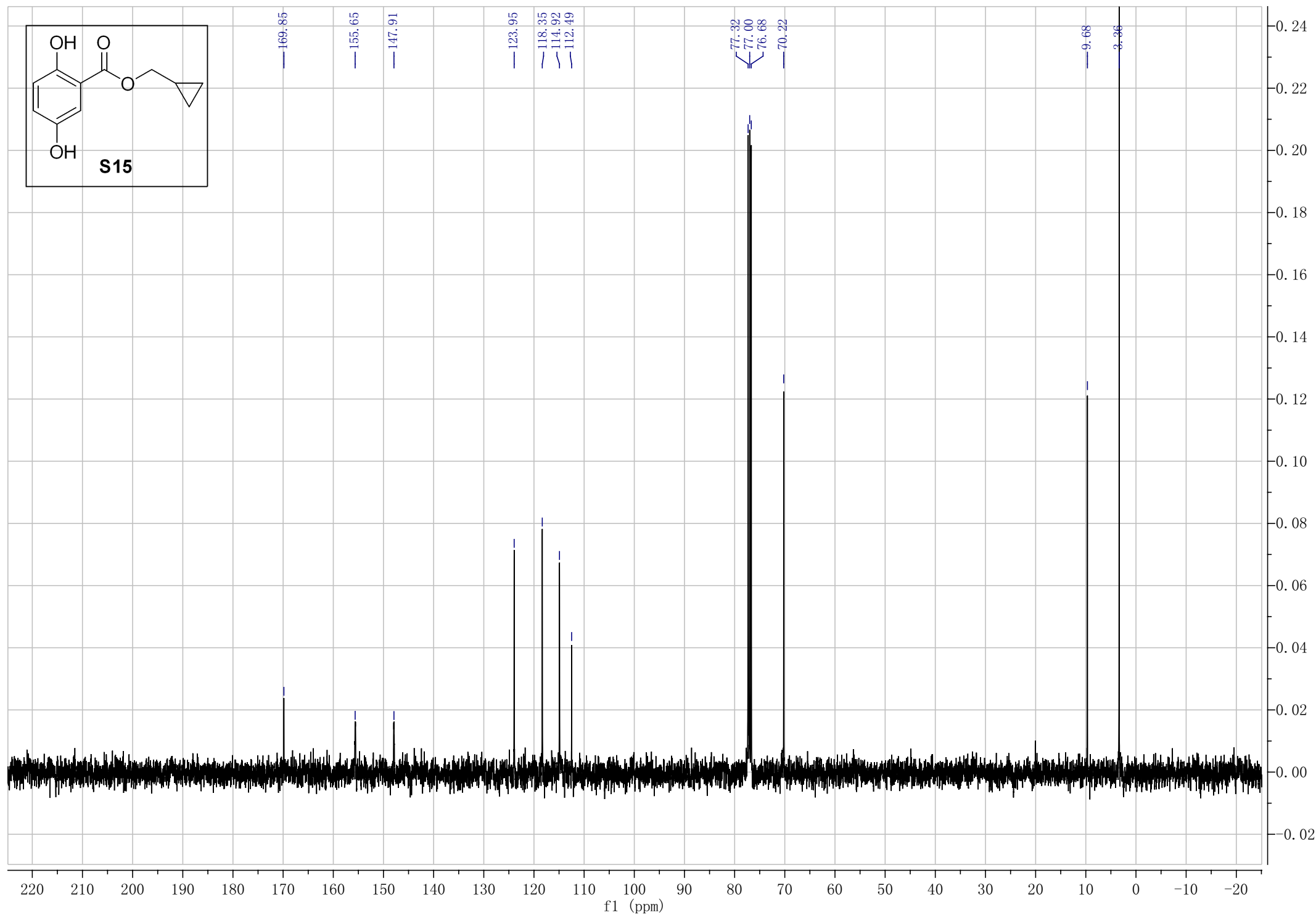
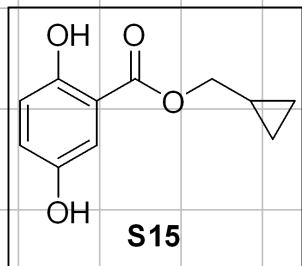


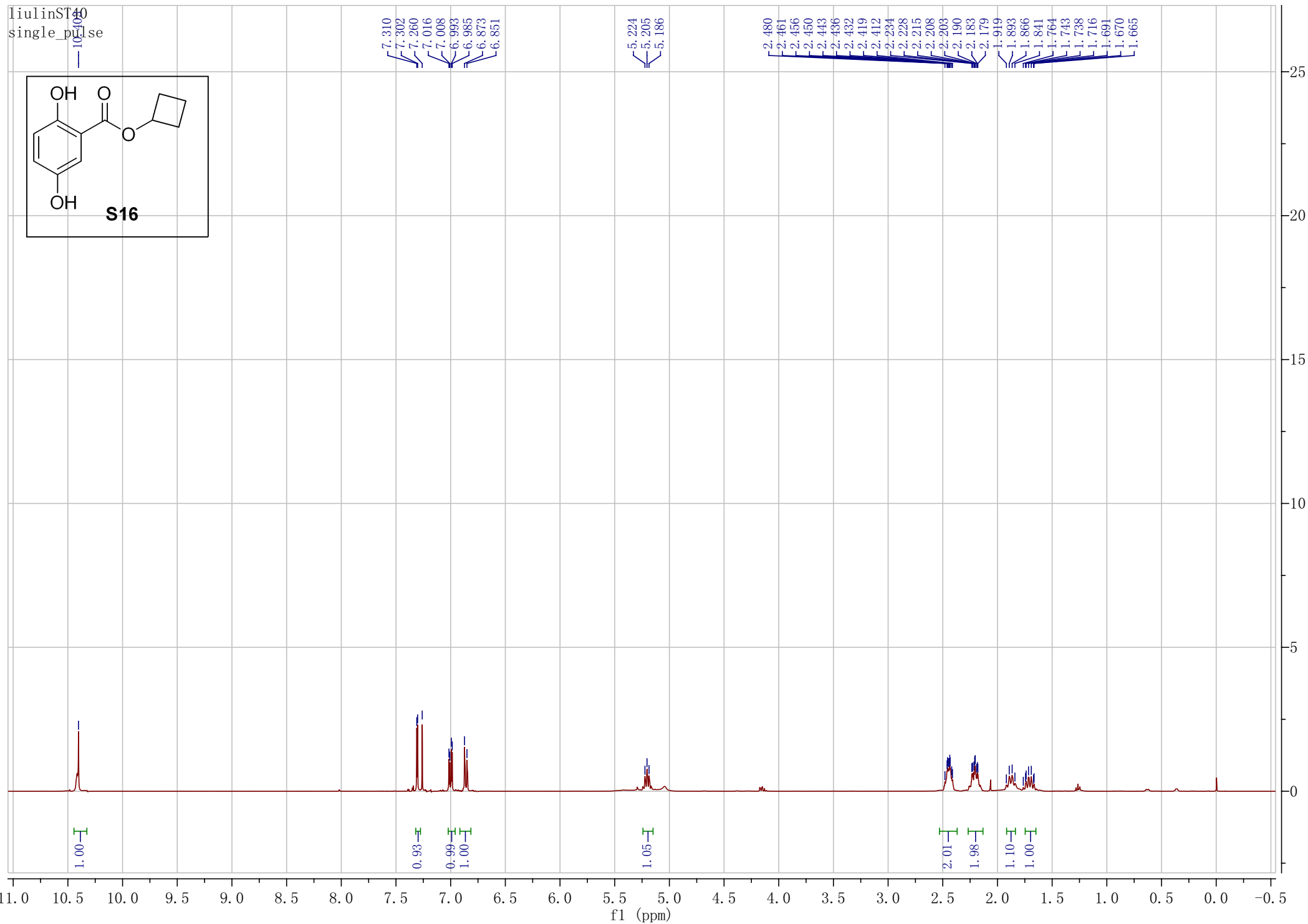


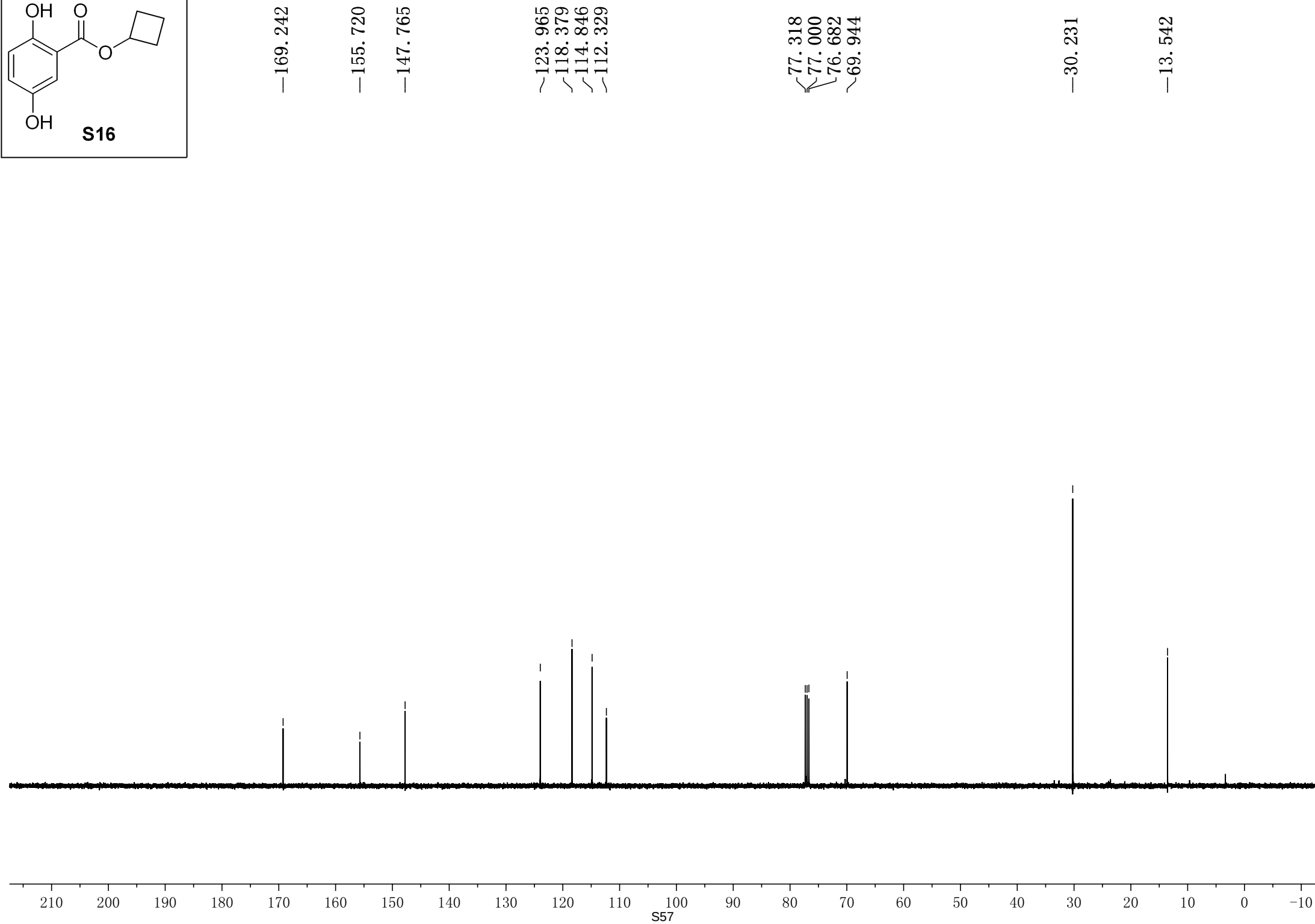
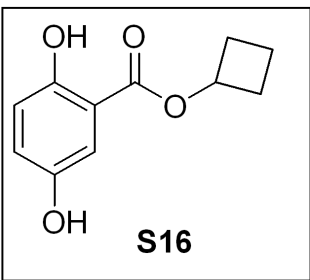




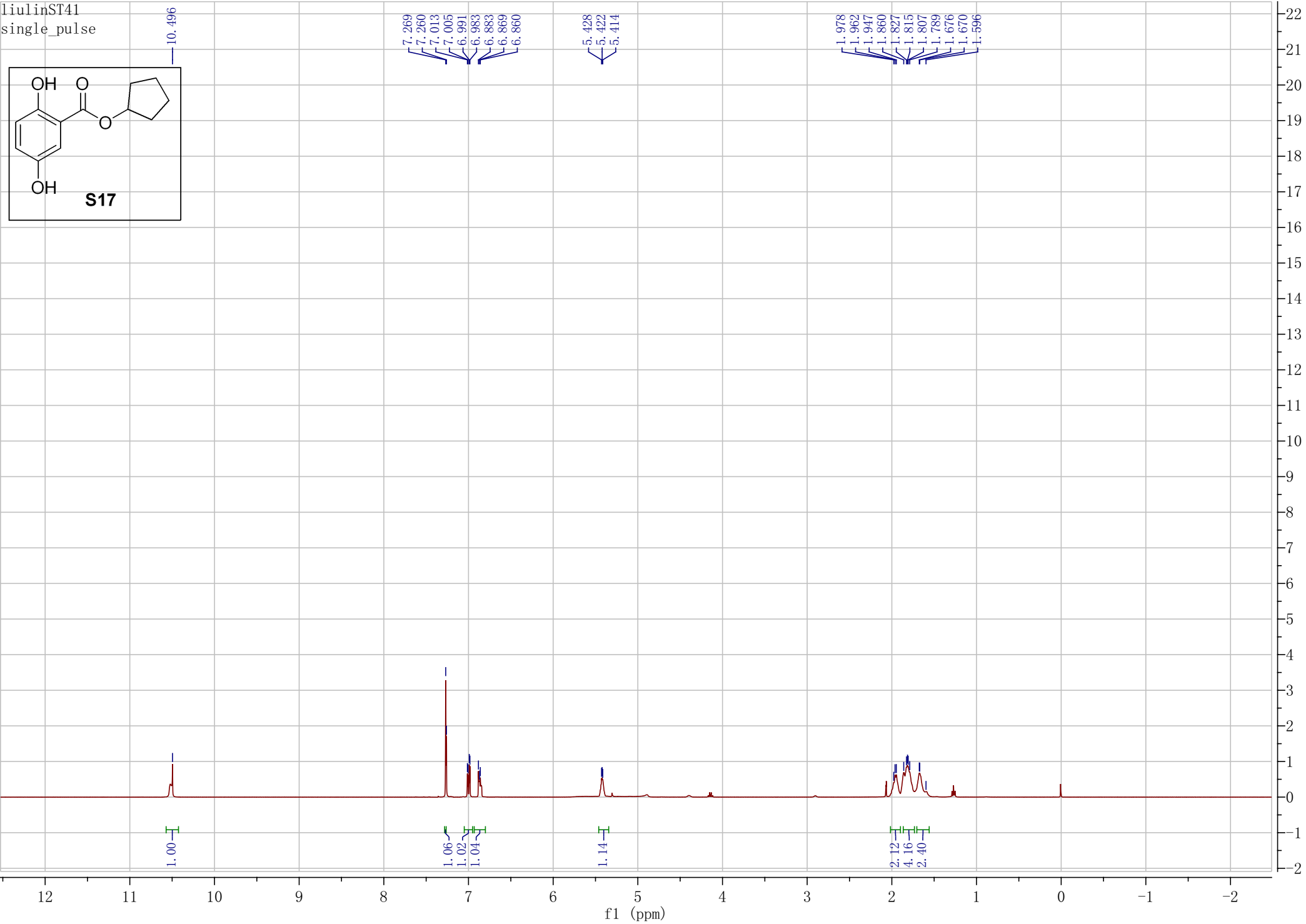
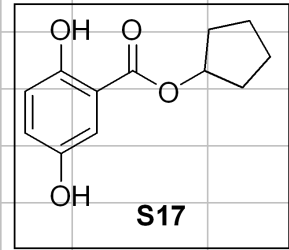


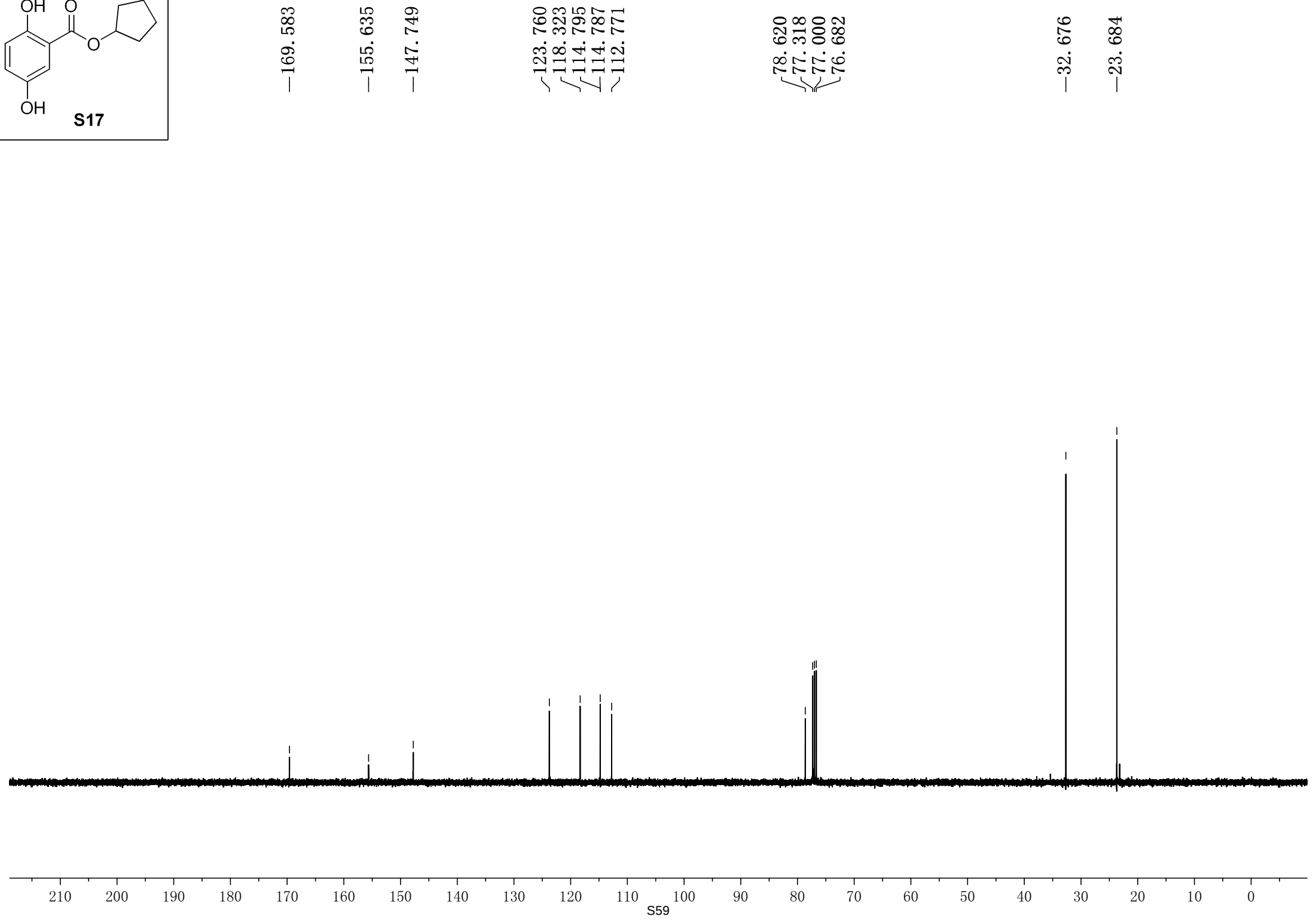
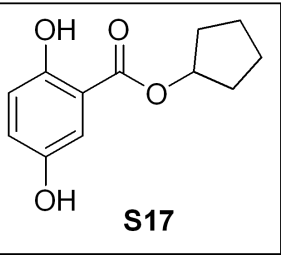


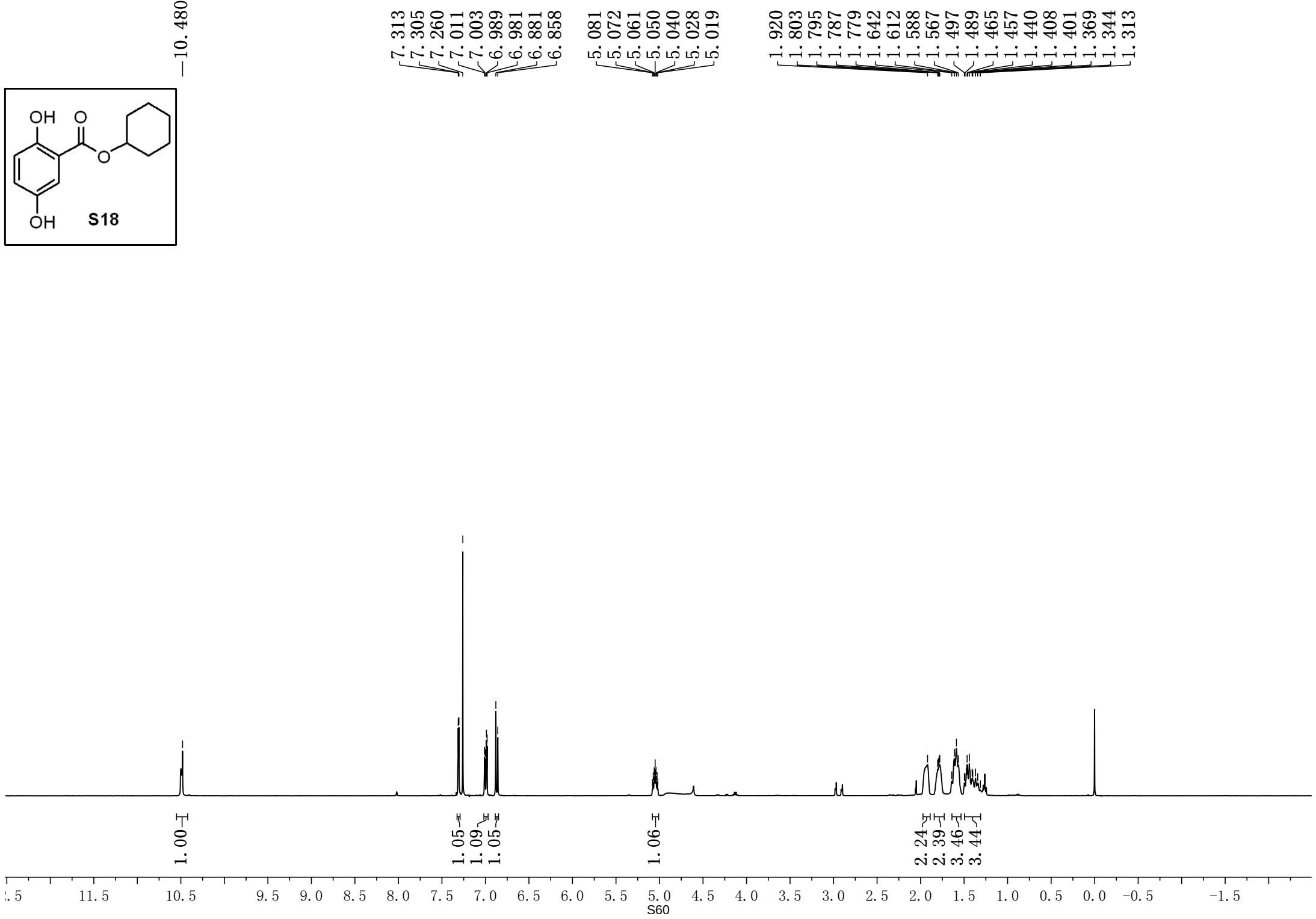
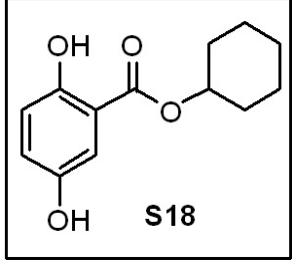


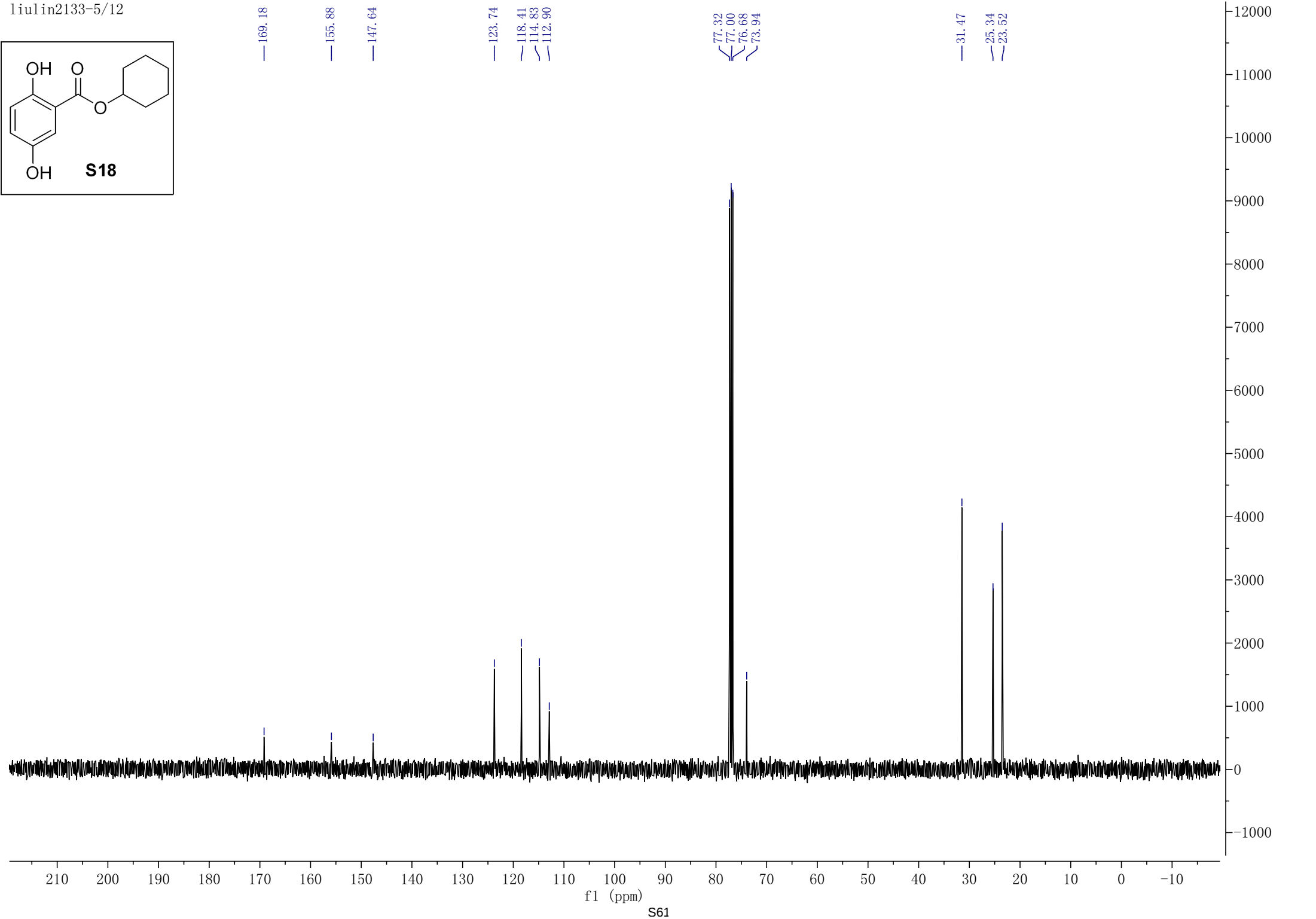
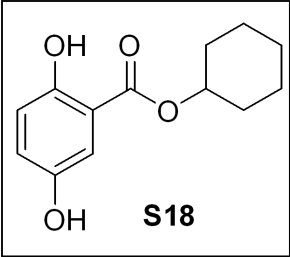


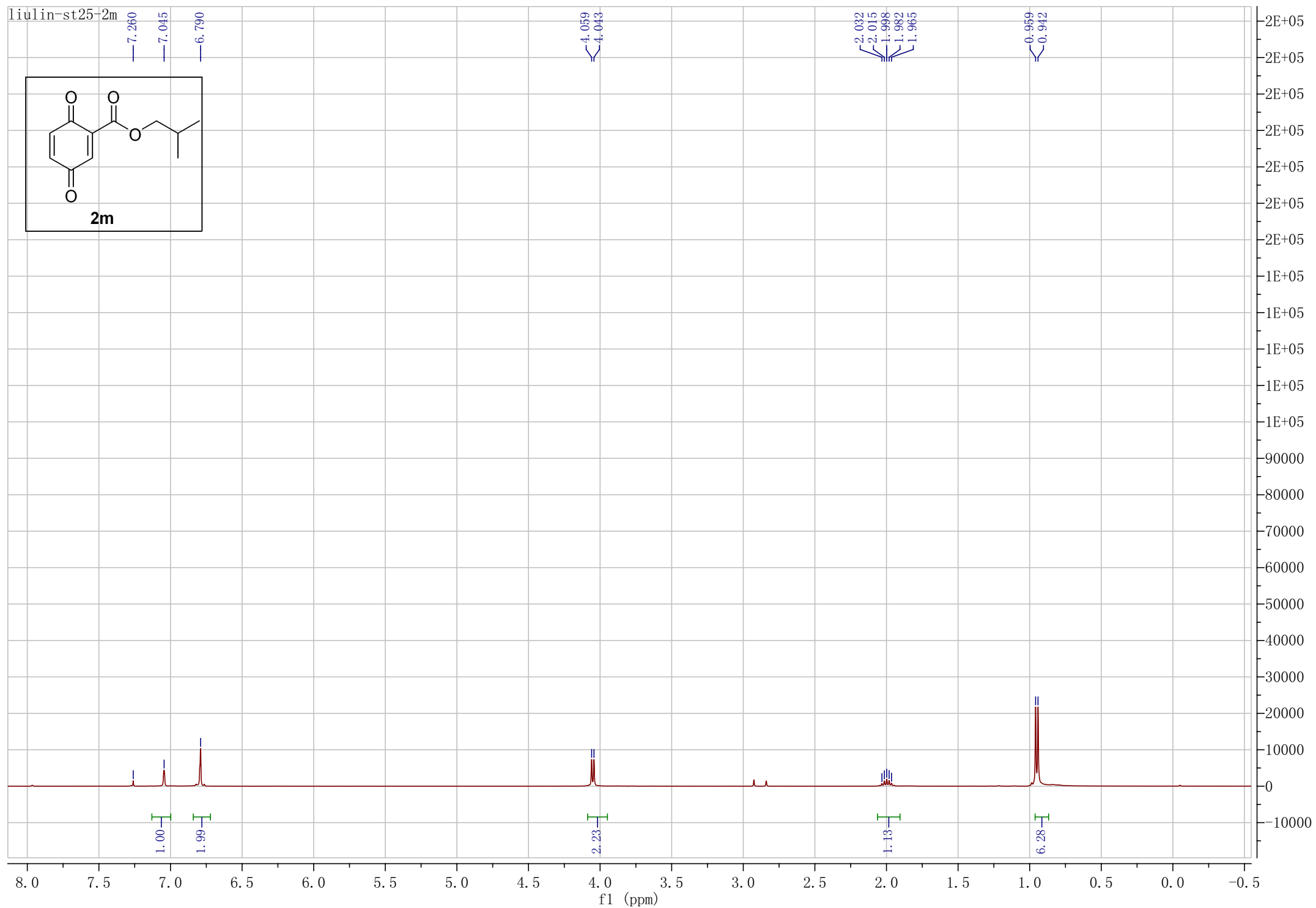
liulinST41
single_pulse

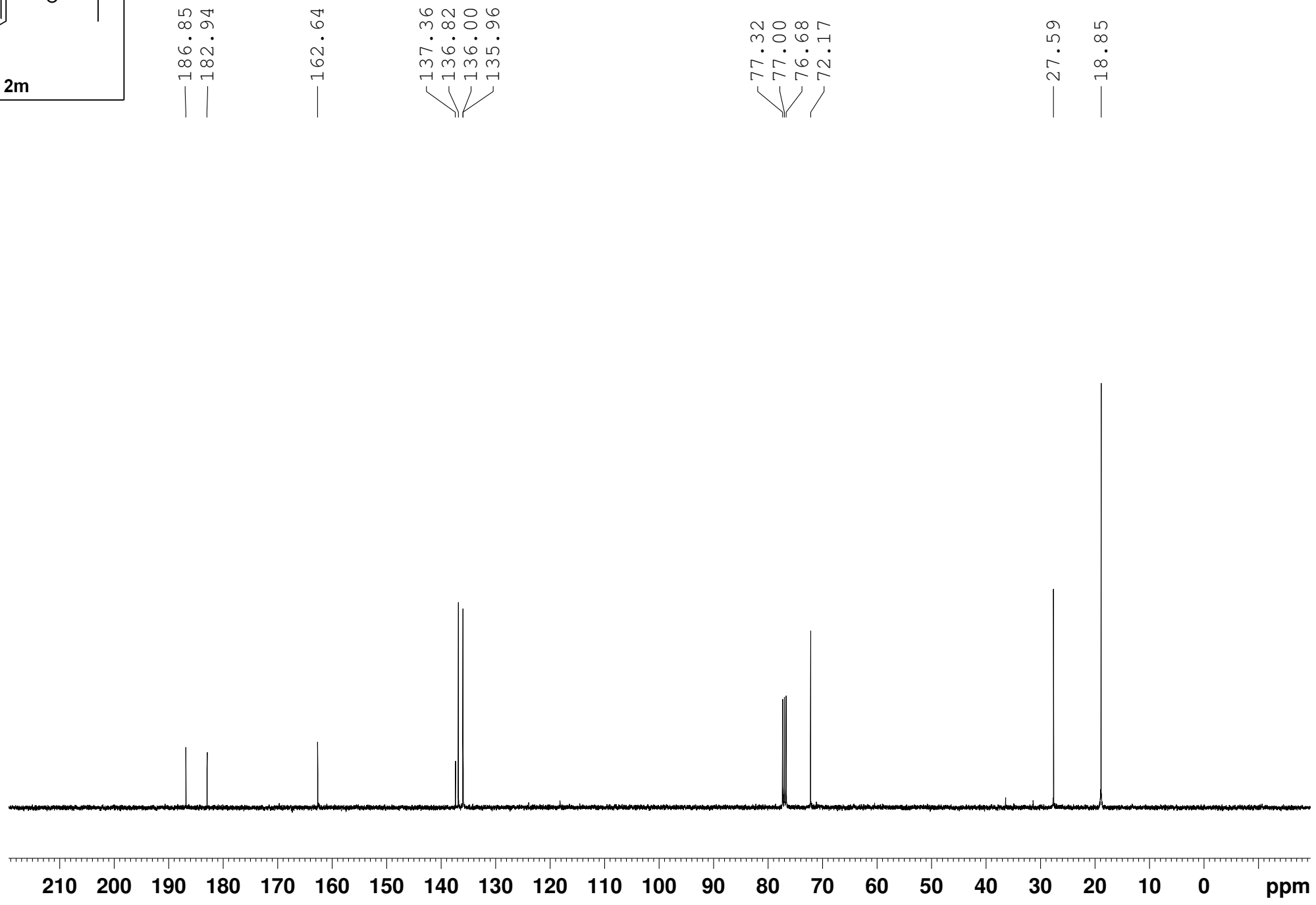
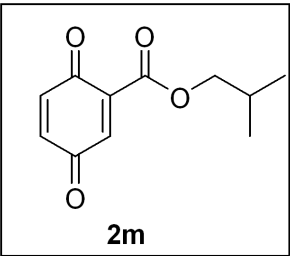


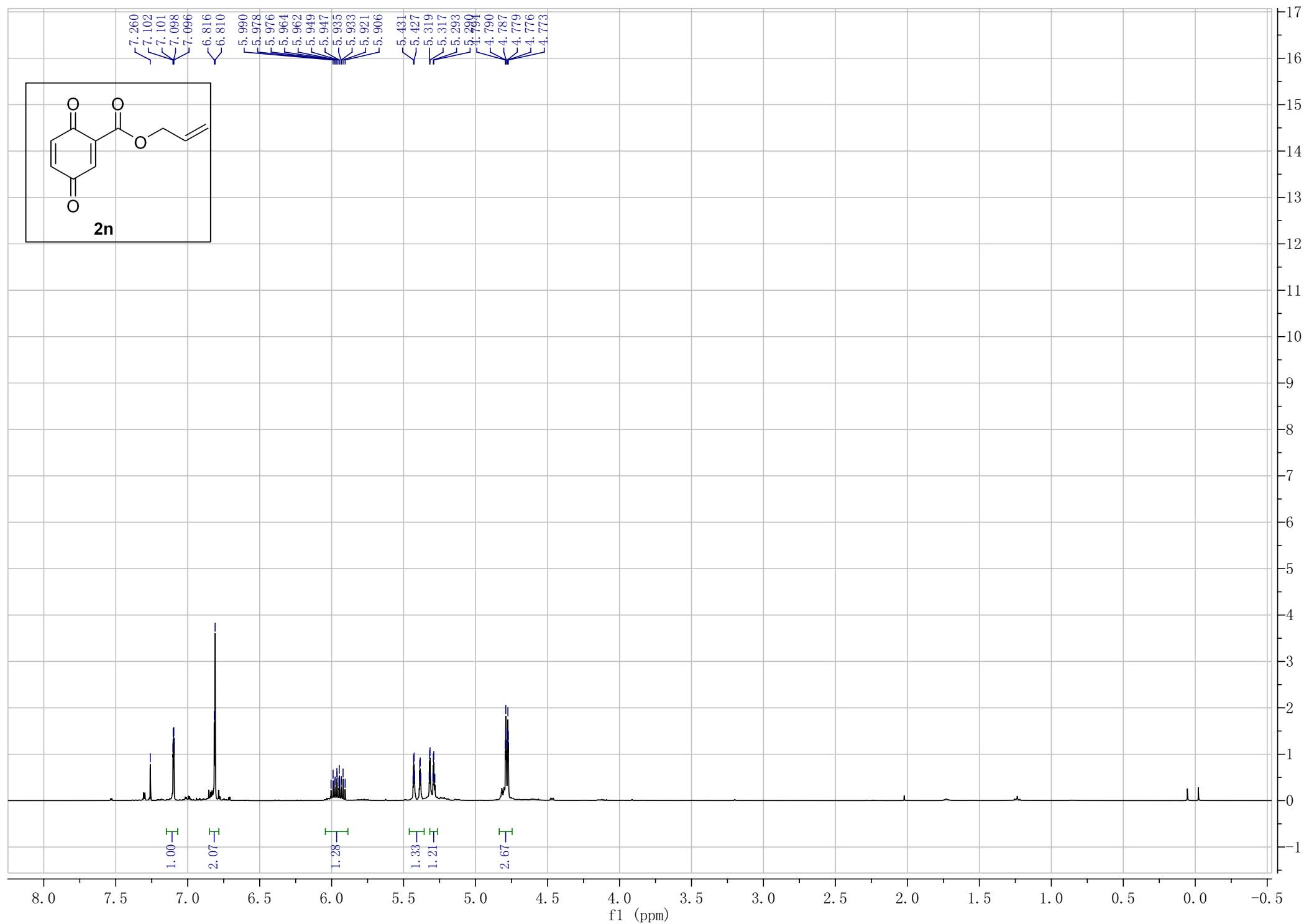




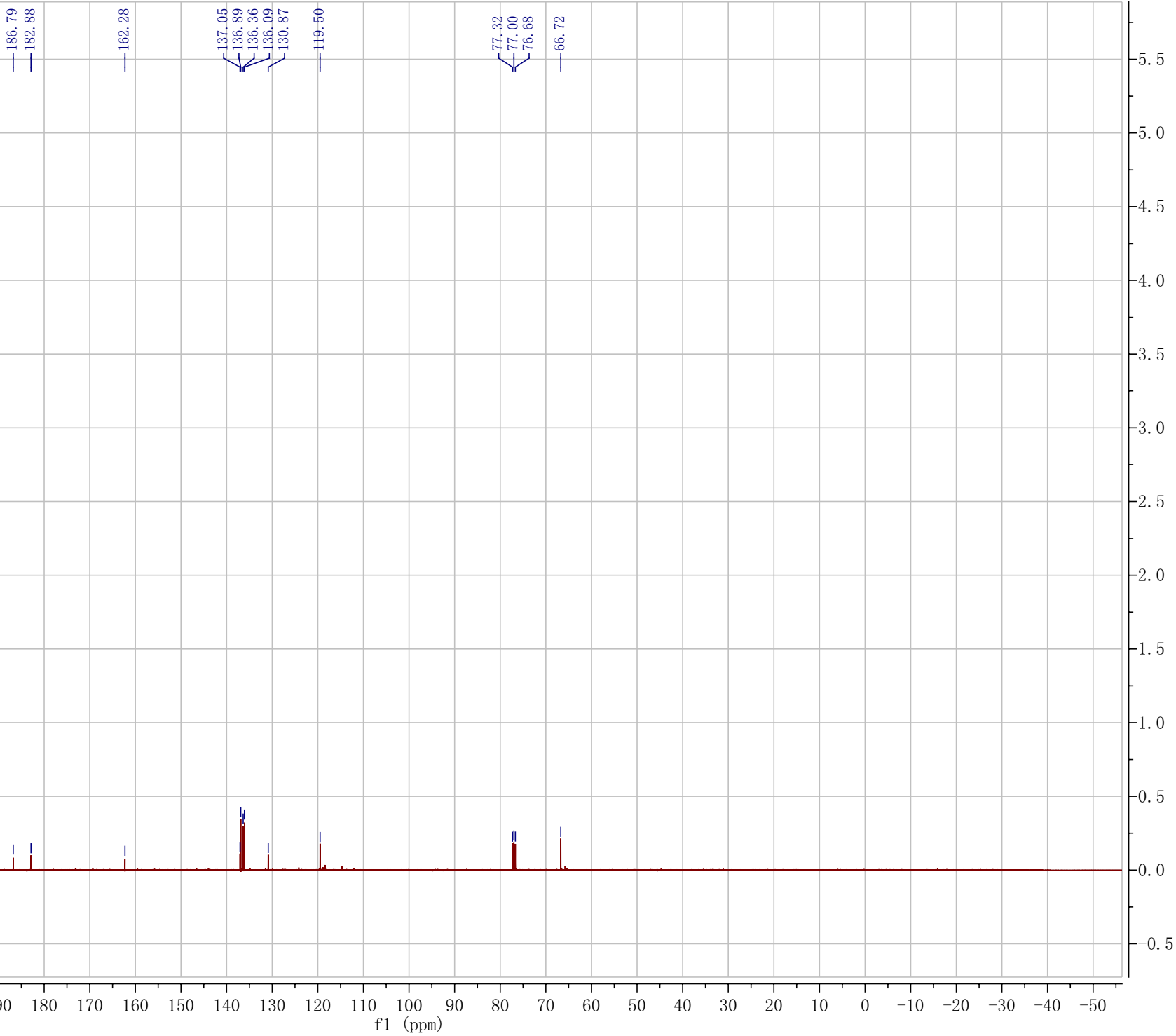
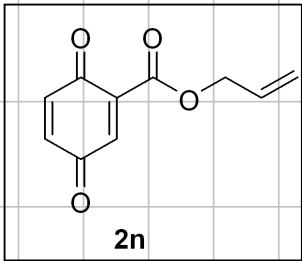


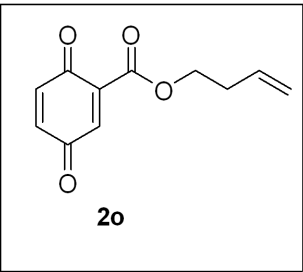






liulininst23
single pulse decoupled gated NOE



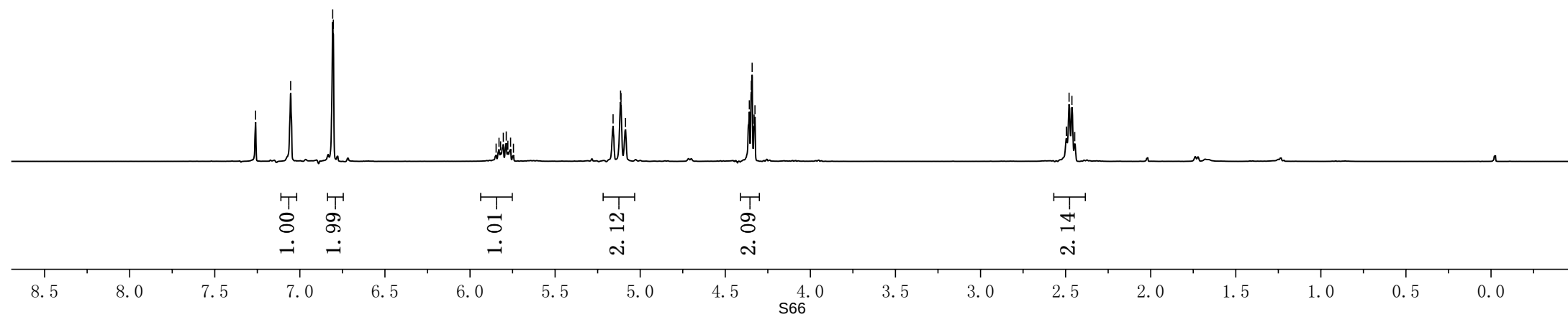


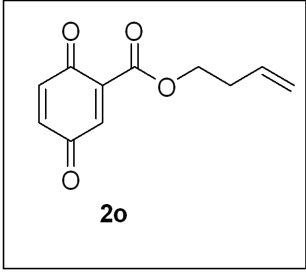
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7.054
6.807
6.803

5.847
5.830
5.820
5.804
5.787
5.778
5.762
5.745
5.159
5.115
5.113
5.086

4.359
4.347
4.342
4.331
4.326

2.496
2.494
2.480
2.463
2.446





—186.856
—182.922

—162.494

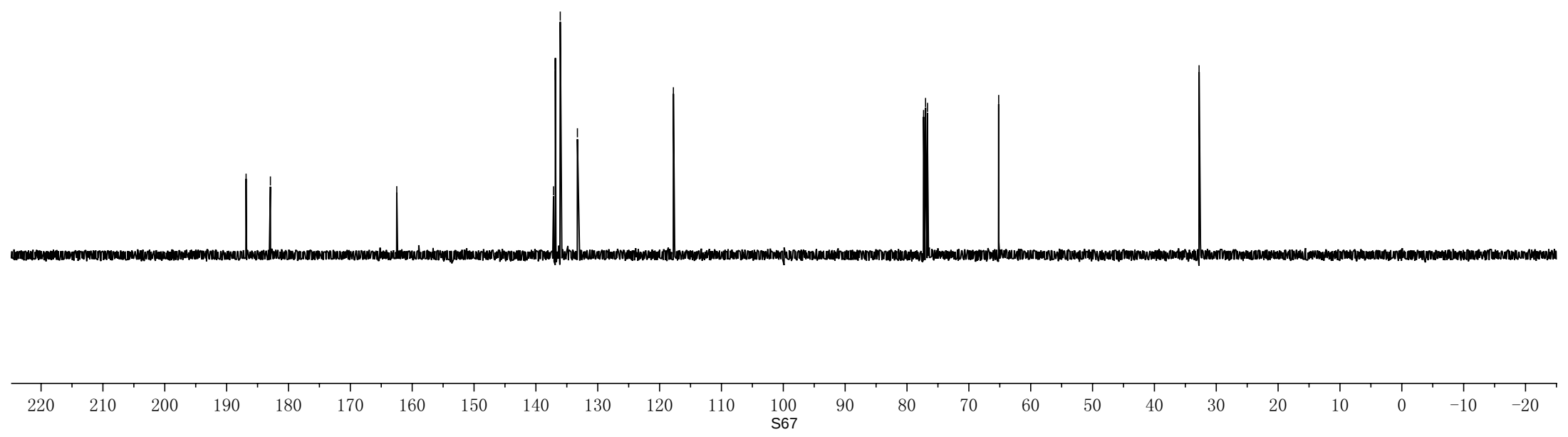
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136.855
136.853
136.060
133.286

—117.769

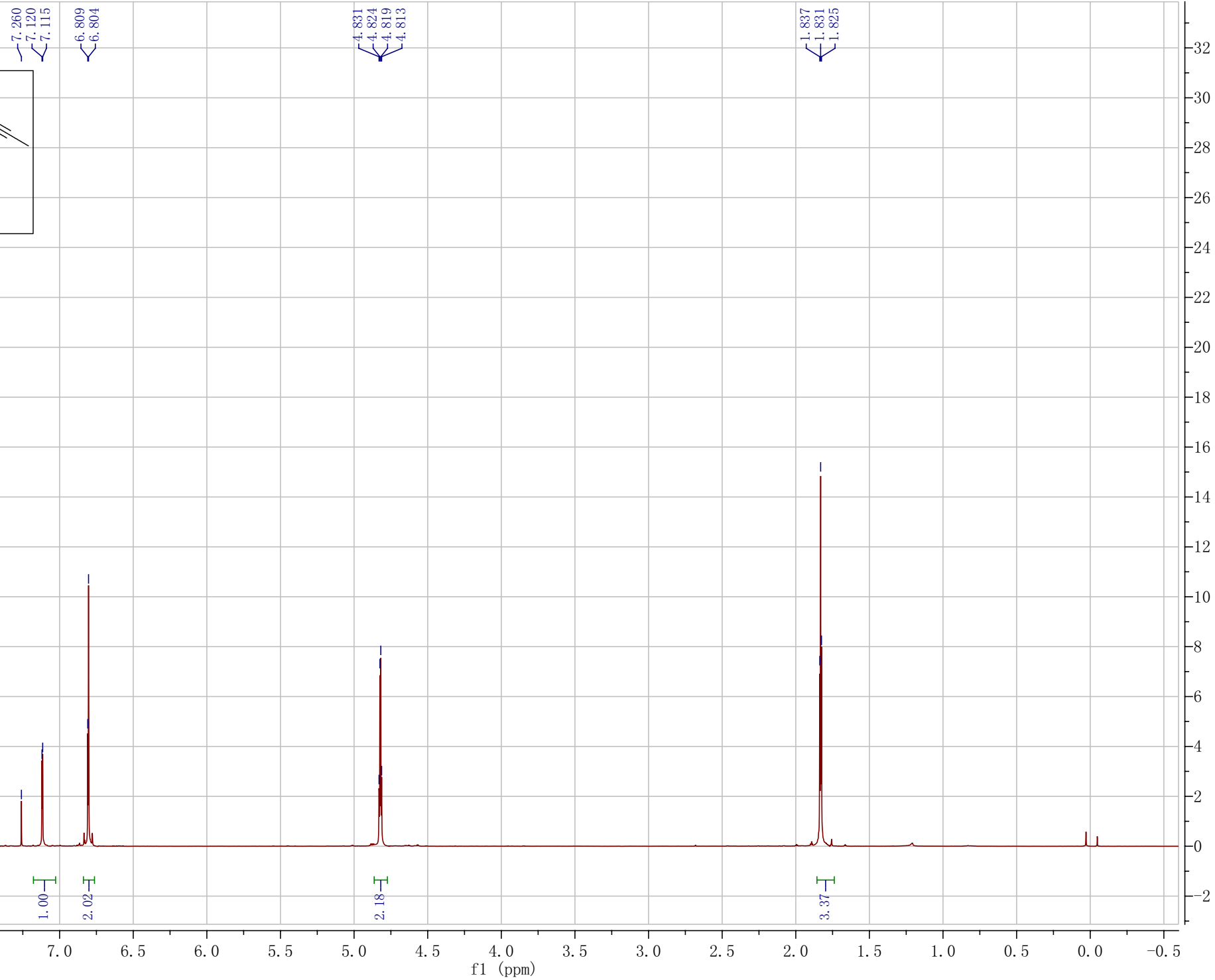
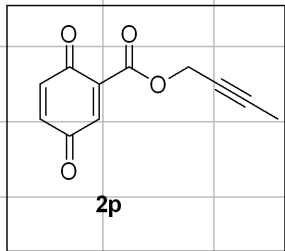
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77.000
76.681

—65.177

—32.780



liulin-181106
single_pulse

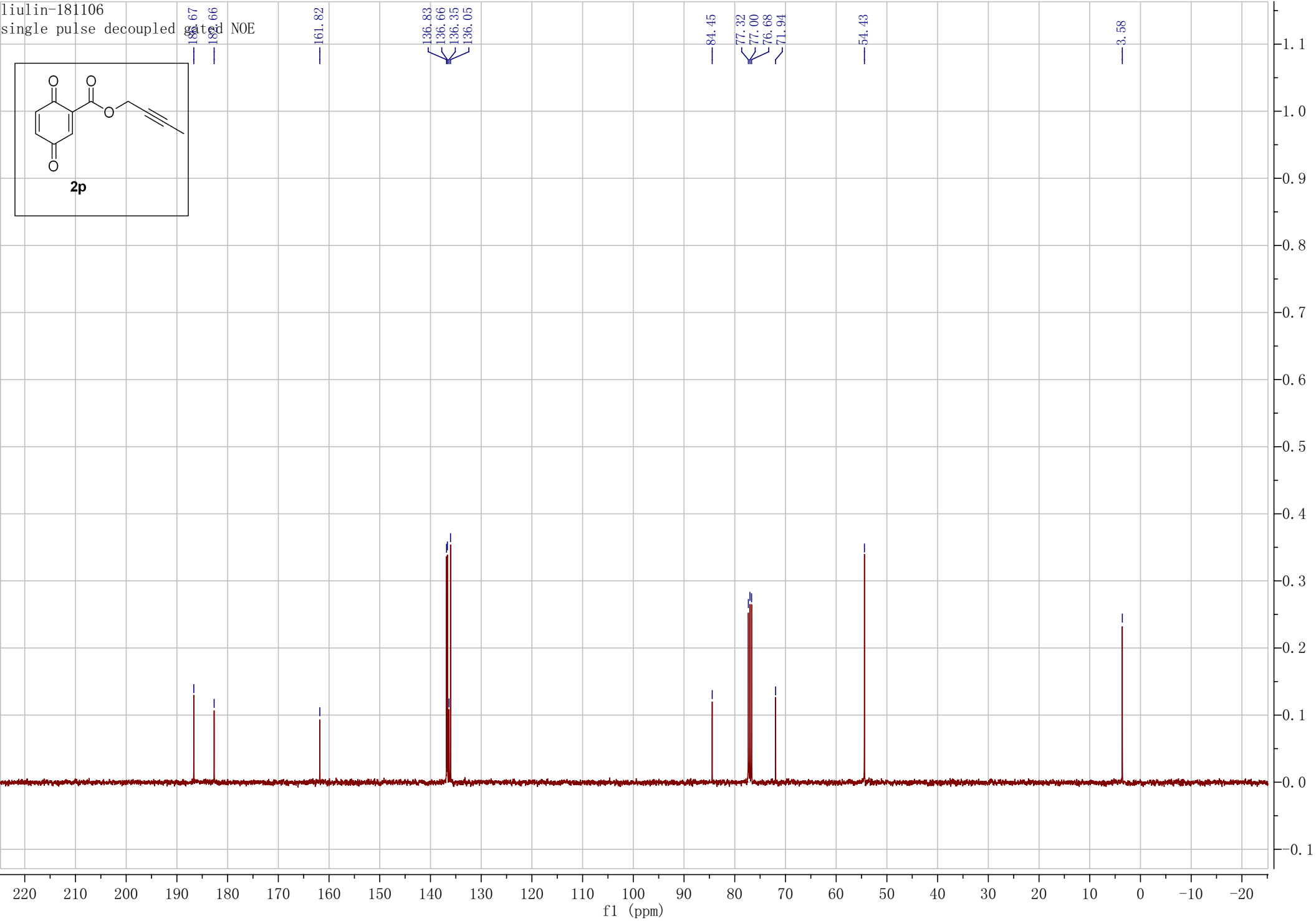
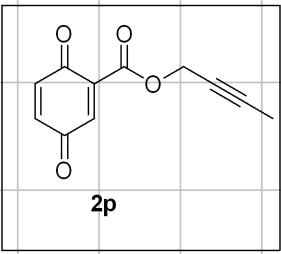


liulin-181106

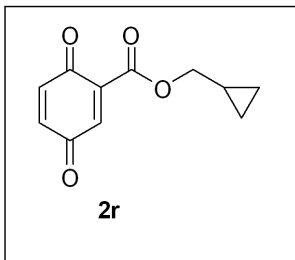
single pulse decoupled

gated

NOE



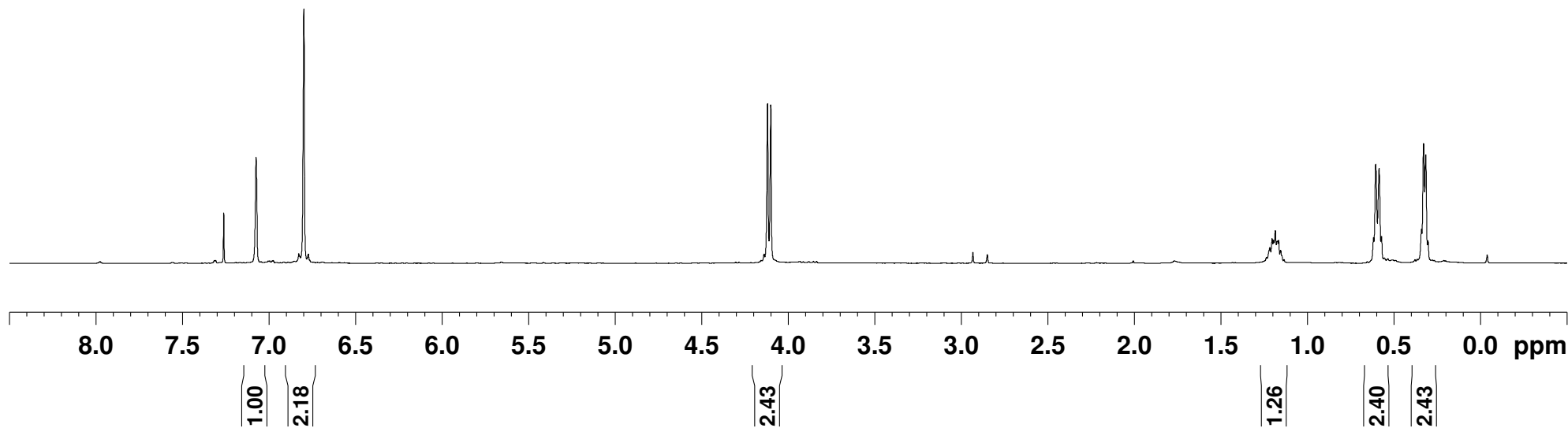
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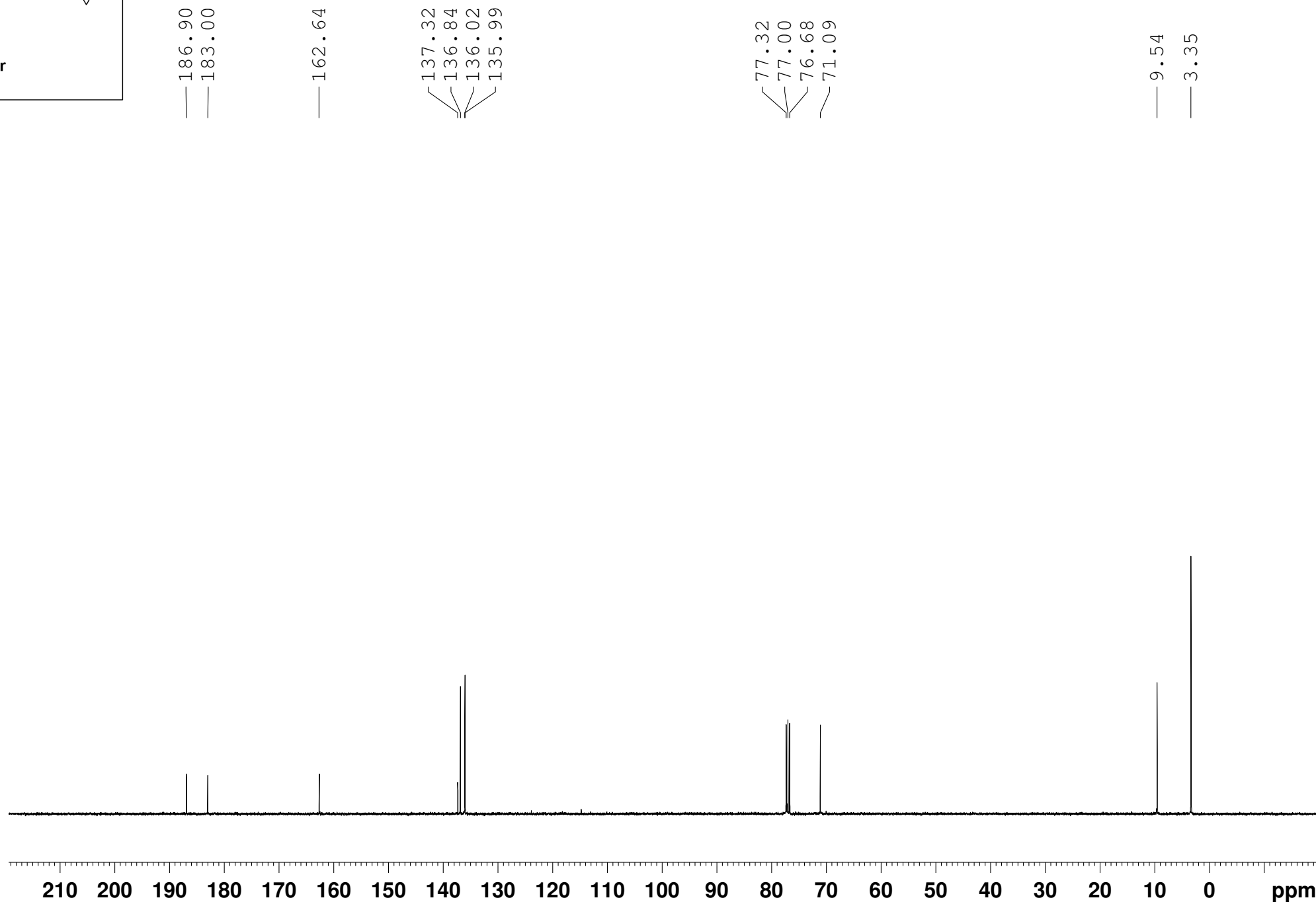
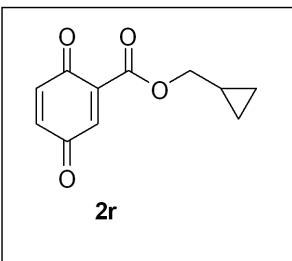


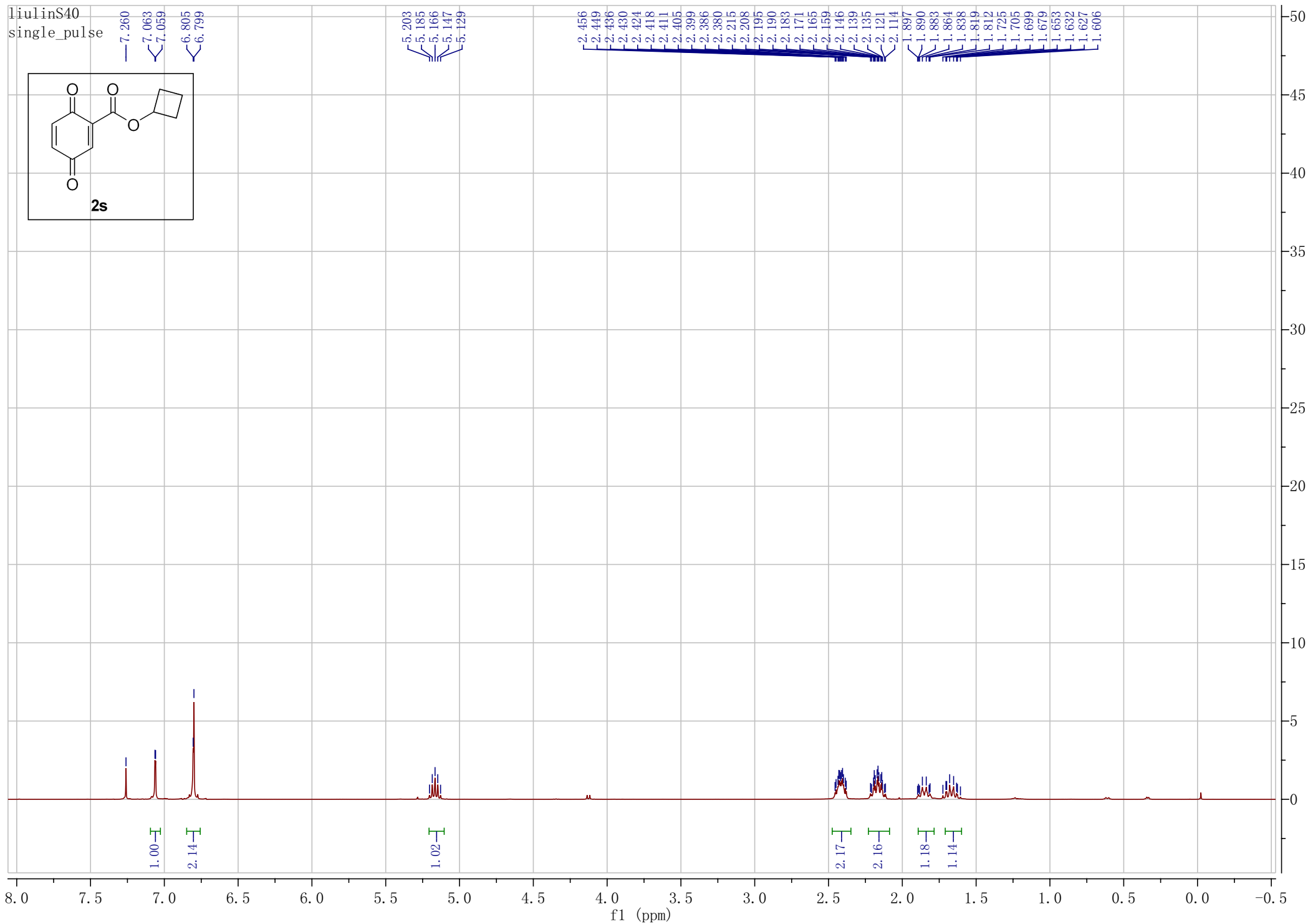
— 7.260
 — 7.074
 — 6.800

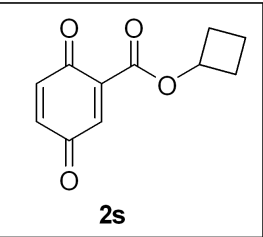
< 4.120
 < 4.102

1.217
 1.205
 1.197
 1.186
 1.175
 1.167
 1.155
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 0.586
 0.572
 0.341
 0.329
 0.317
 0.303









—186.913
—183.031

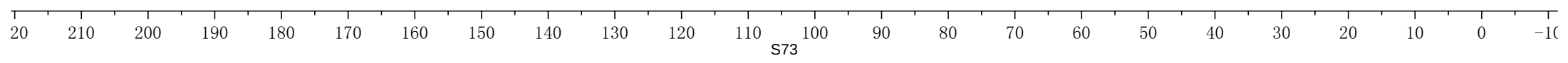
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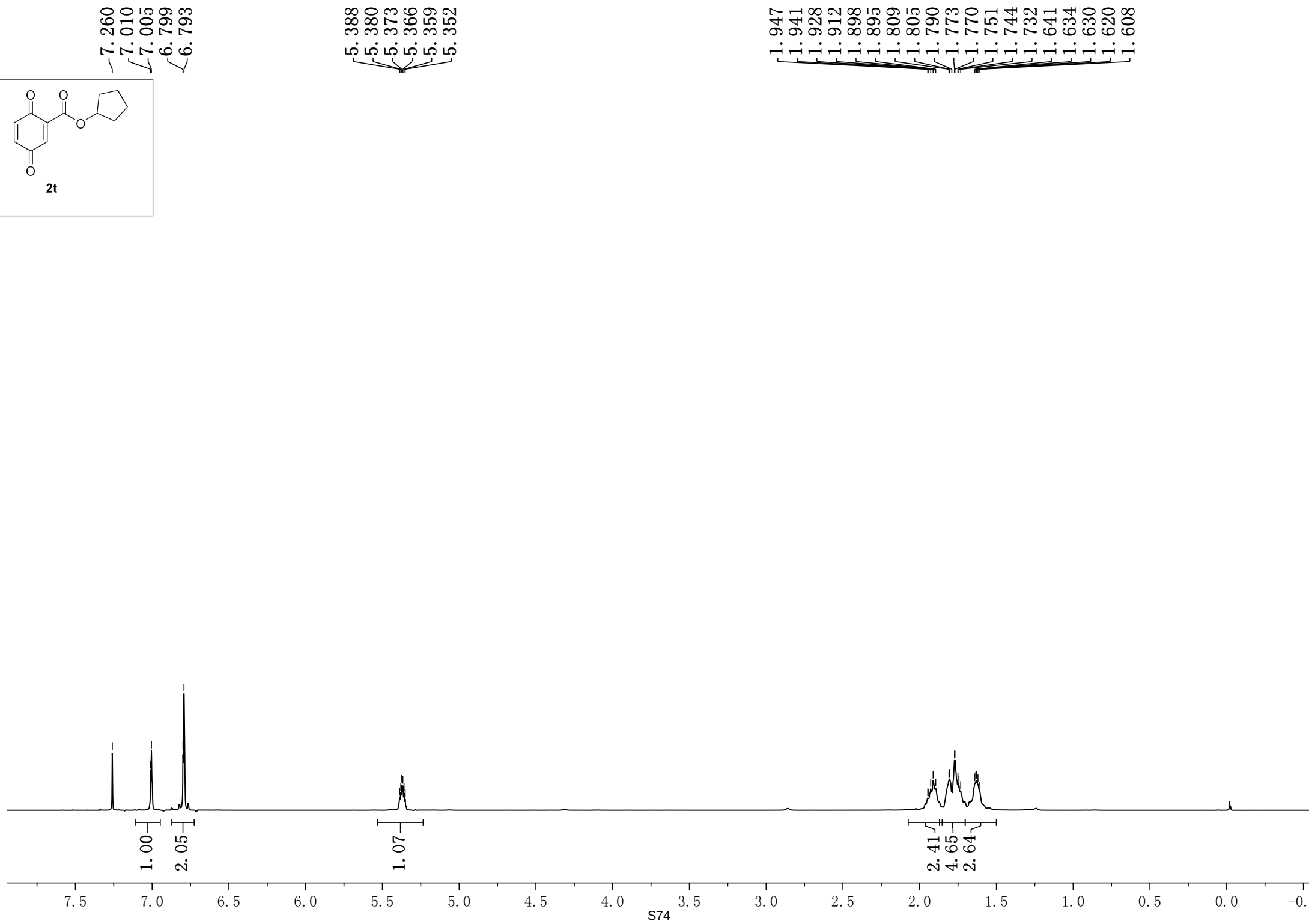
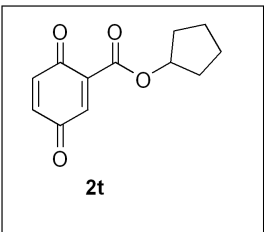
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136.022

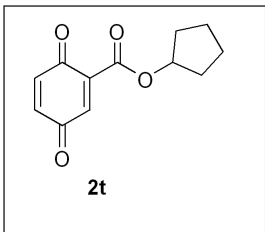
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77.000
76.682
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—30.160

—13.477







—186.992

—183.129

—162.407

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135.597

79.626

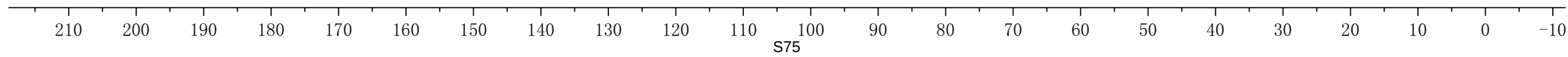
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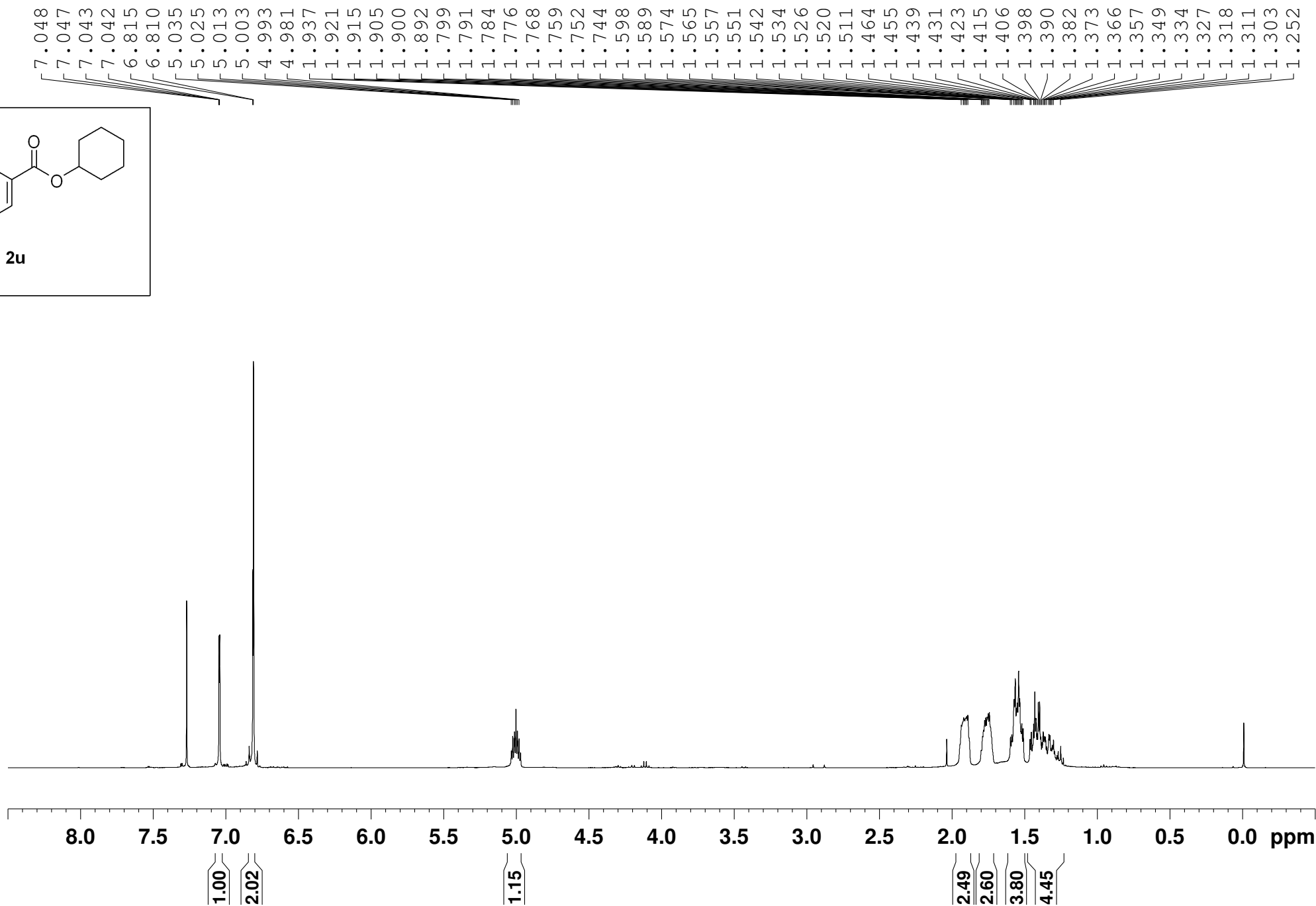
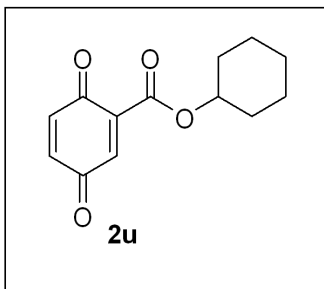
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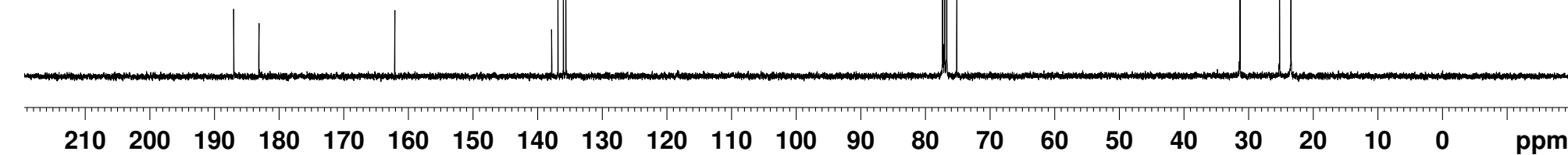
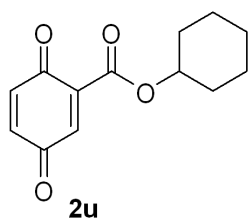
76.681

—32.617

—23.611







— 187.03
— 183.12

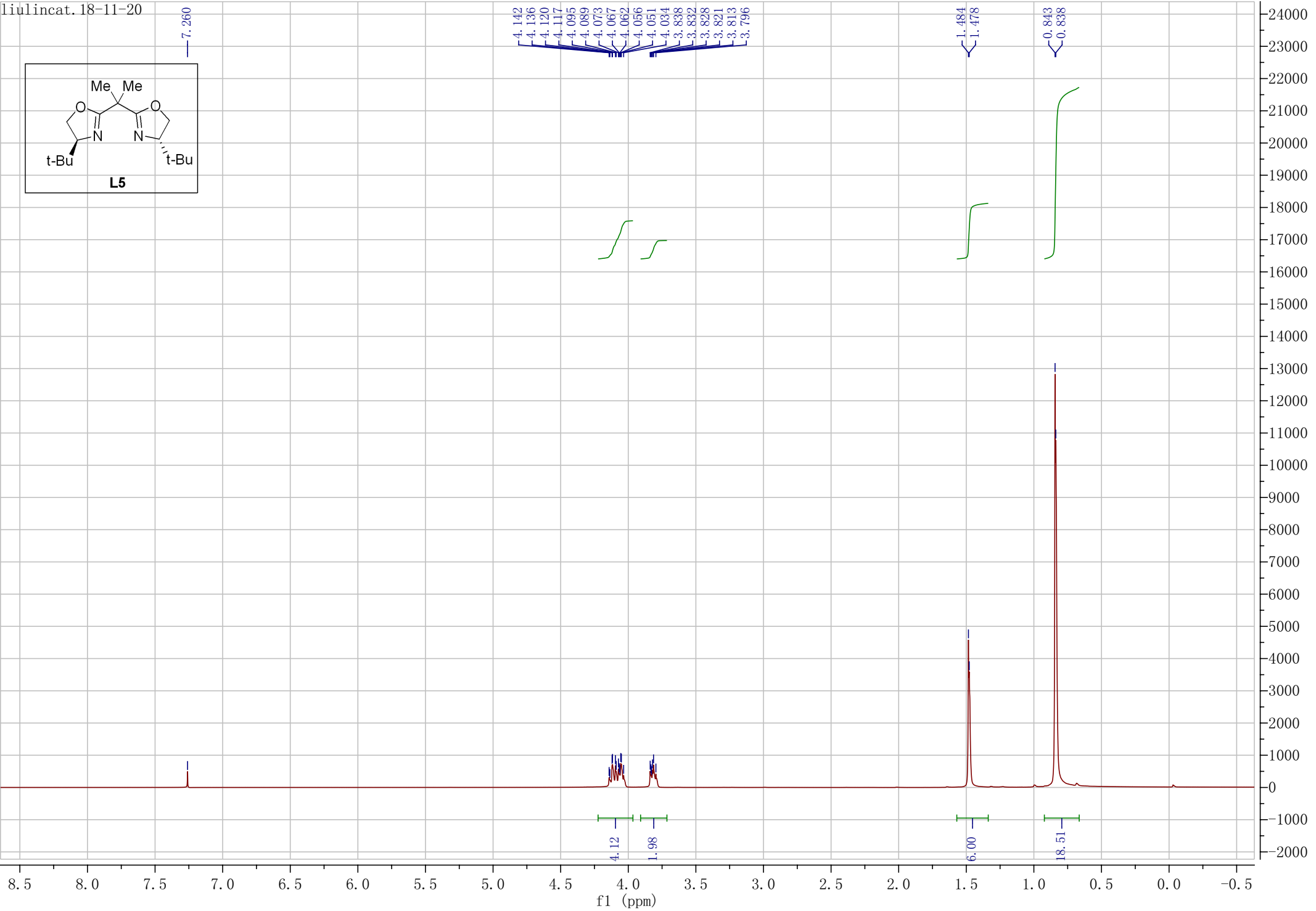
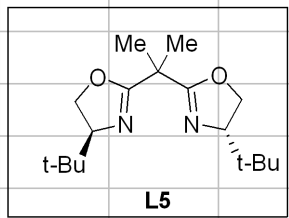
— 162.10

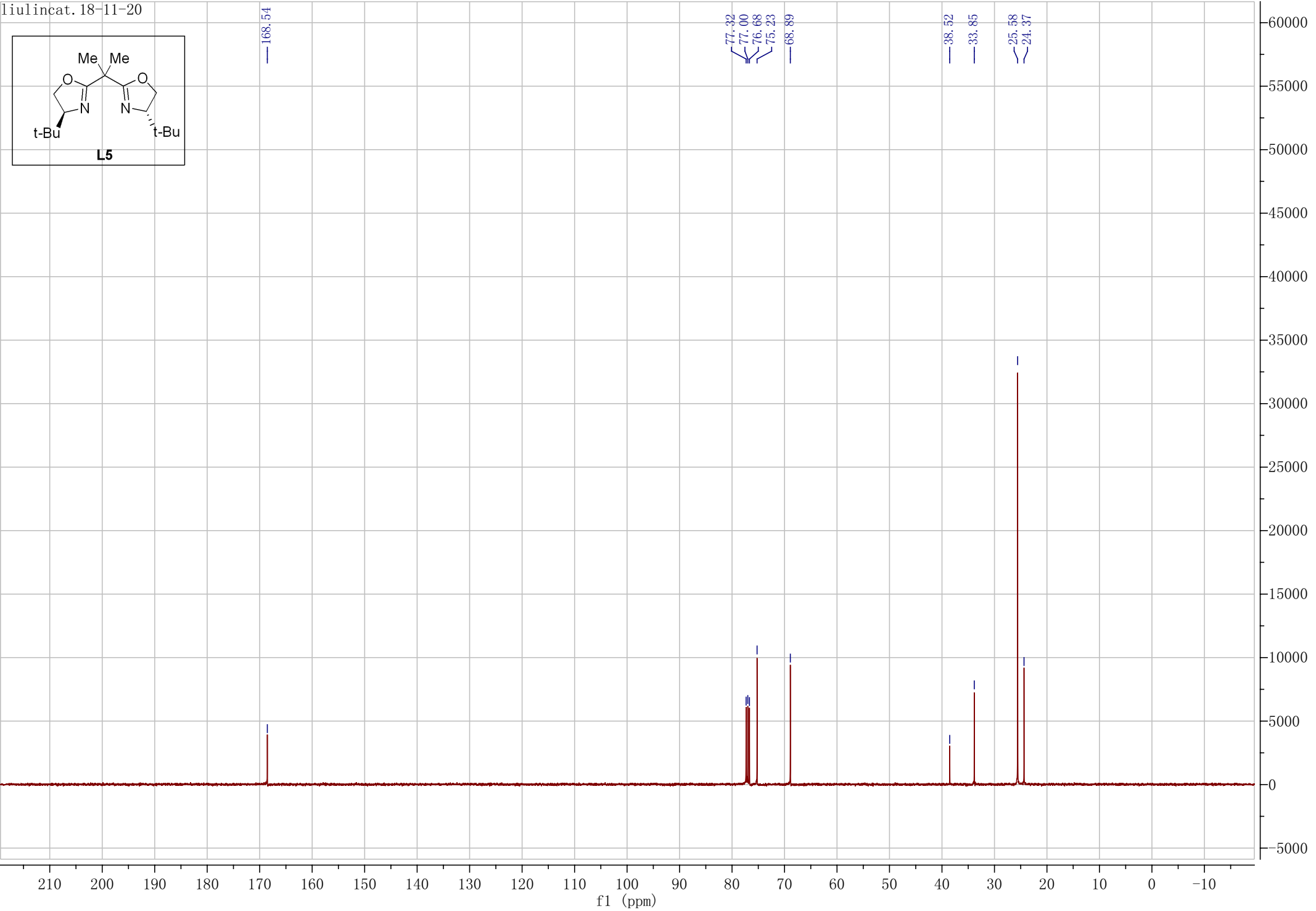
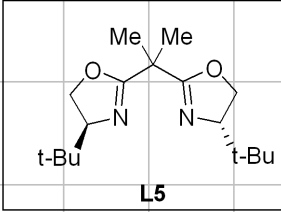
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136.03
135.63

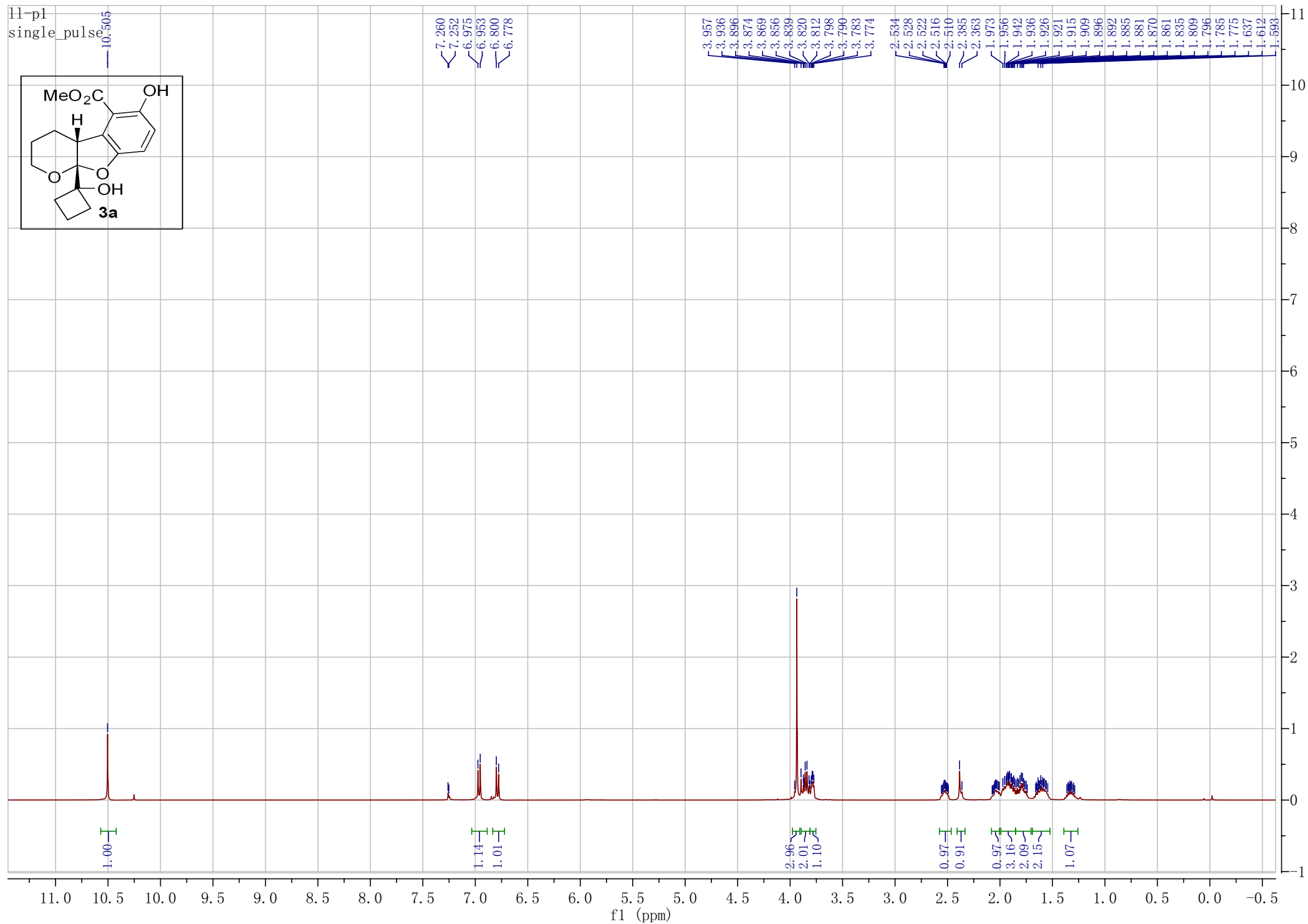
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77.00
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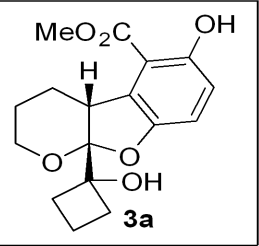
— 31.28
— 25.19
— 23.40

liulincat.18-11-20

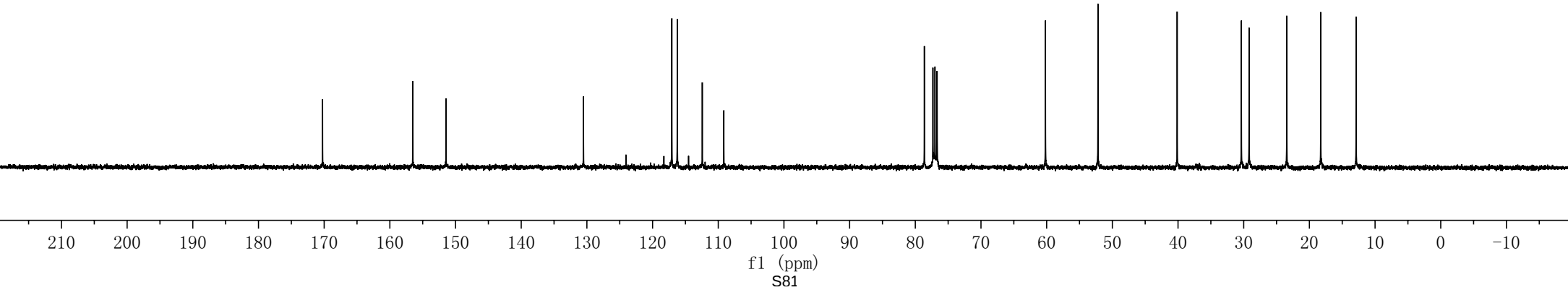


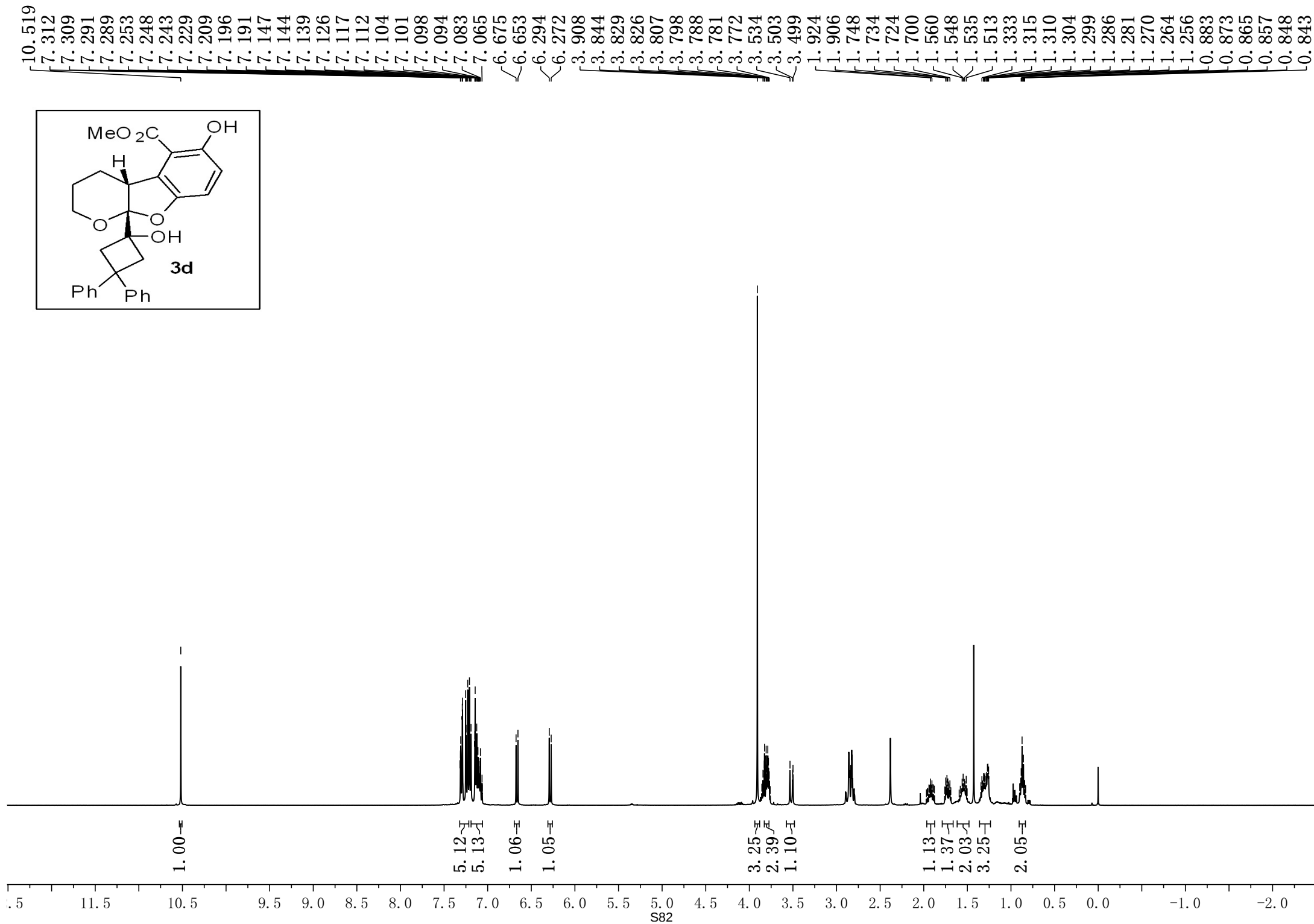


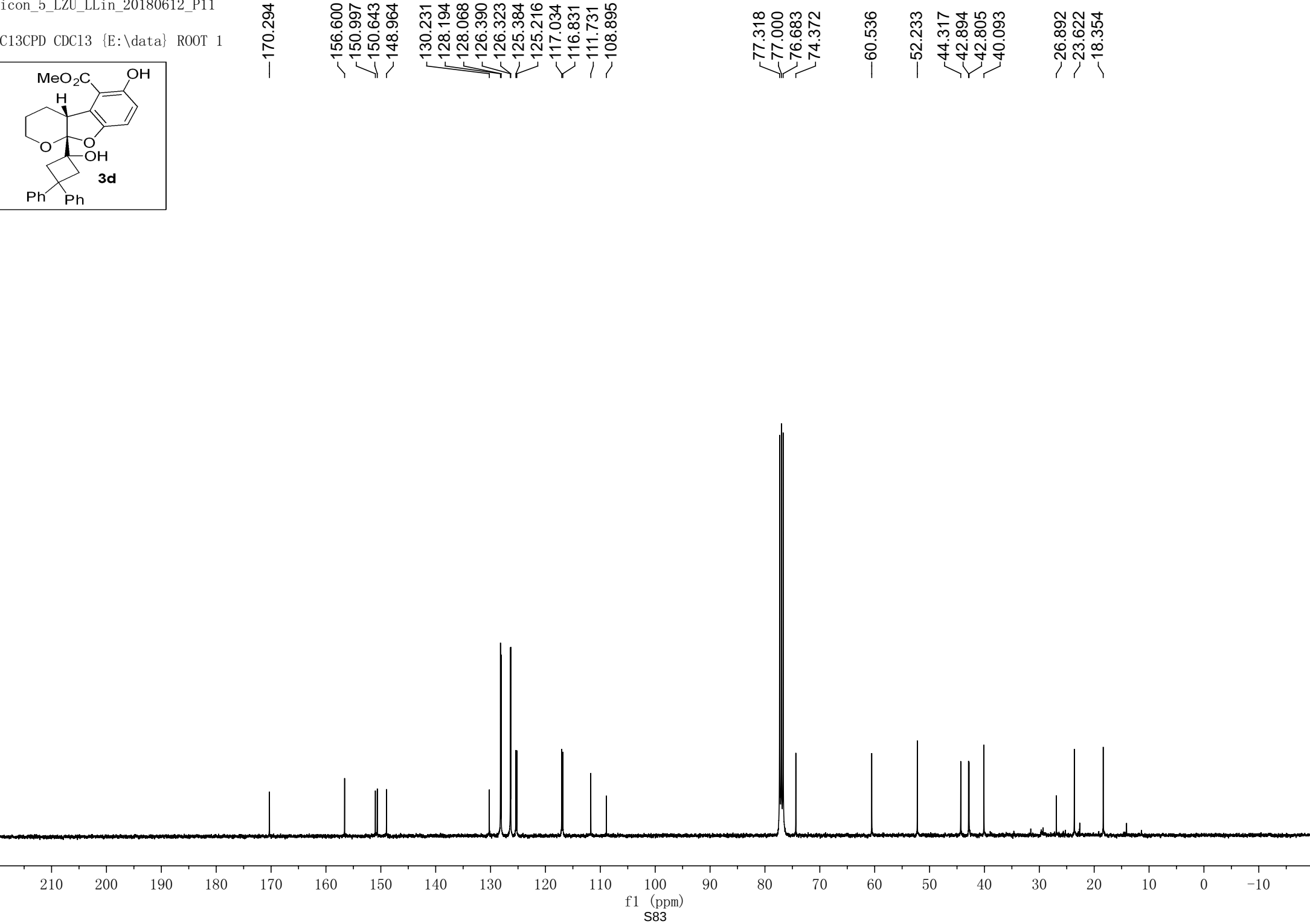
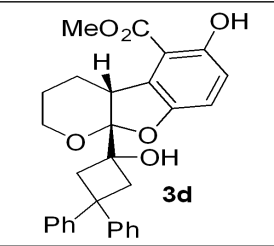




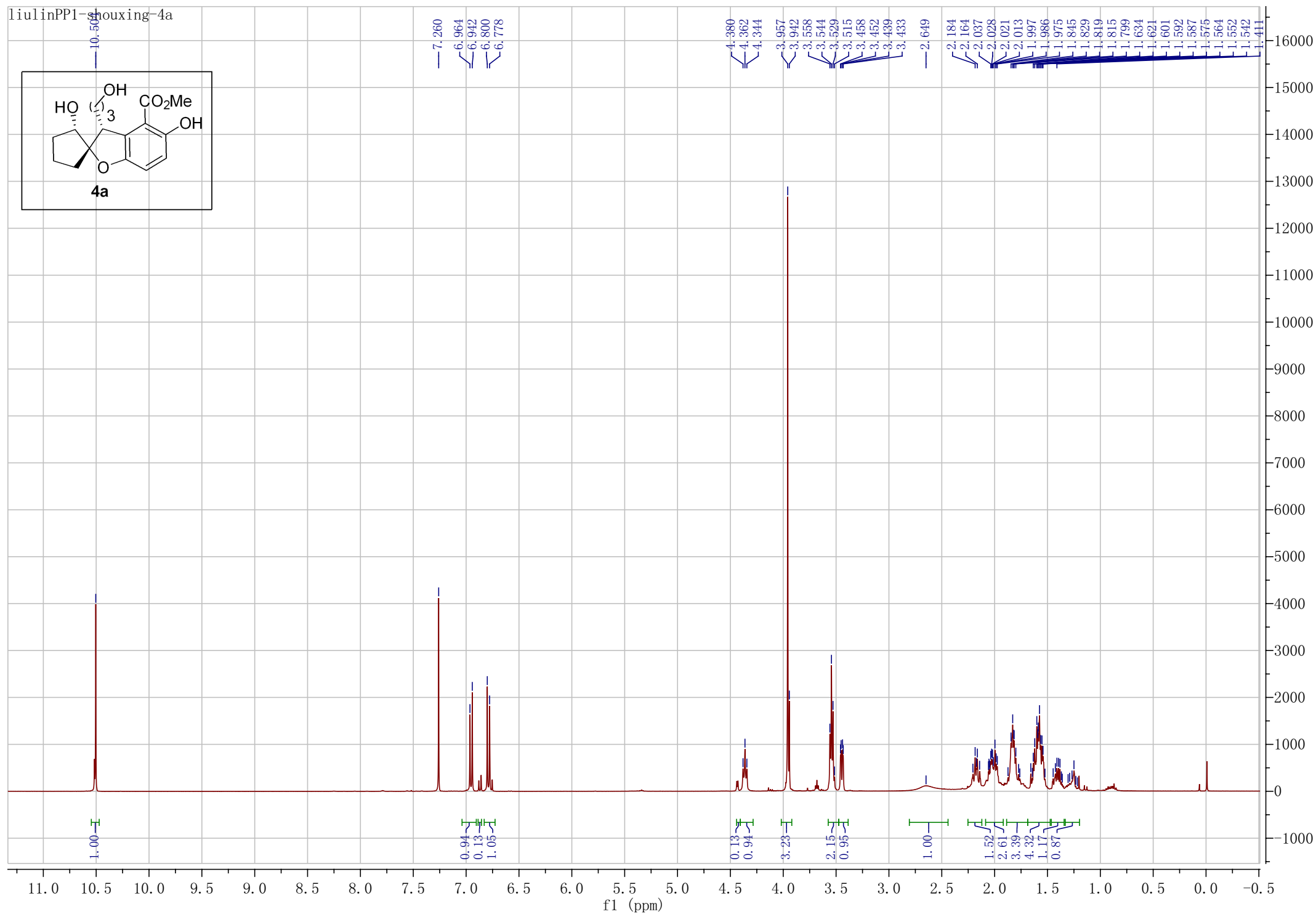
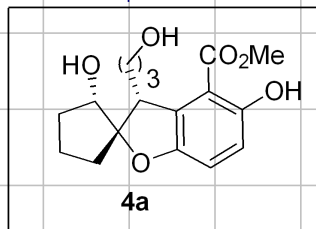
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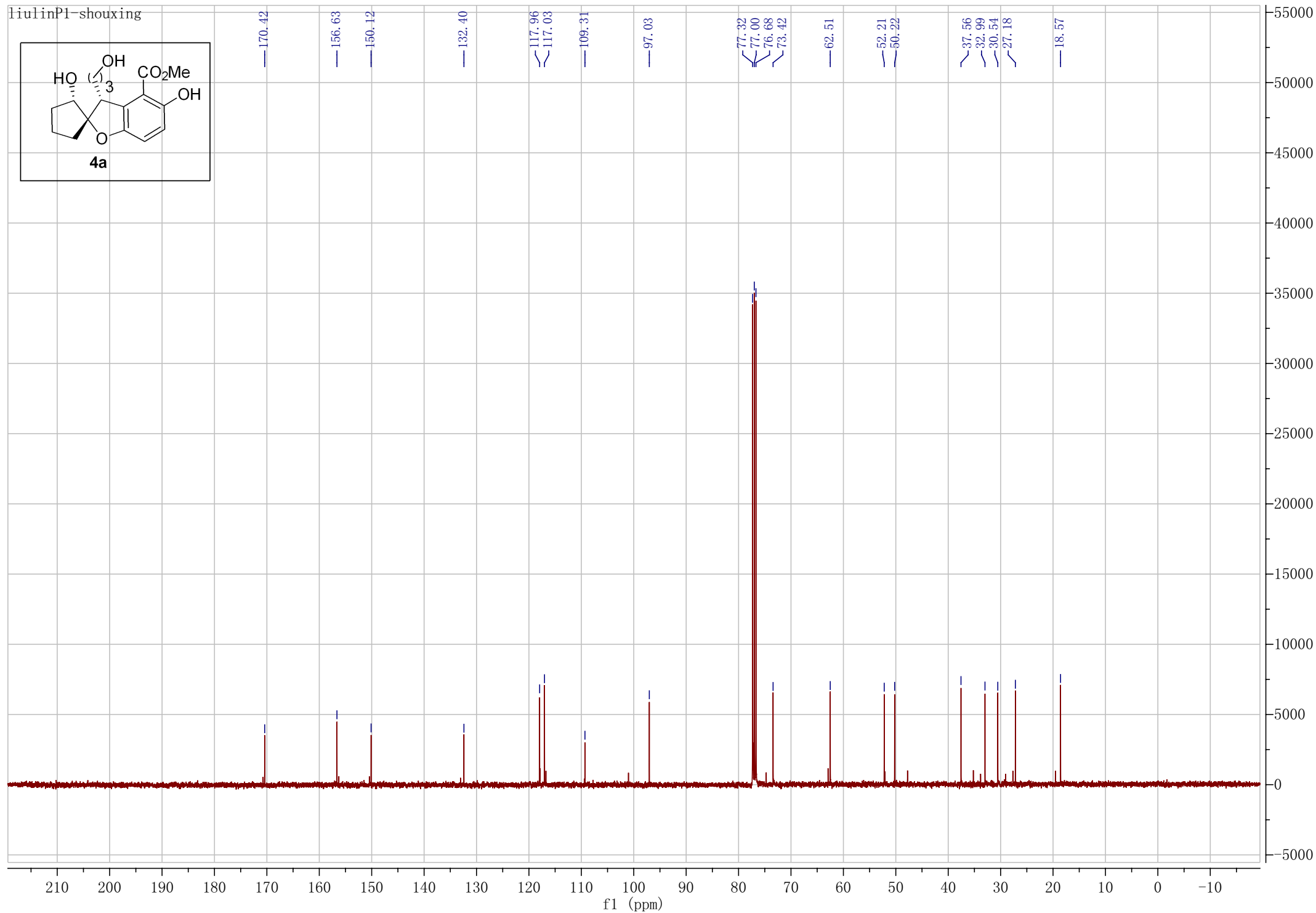
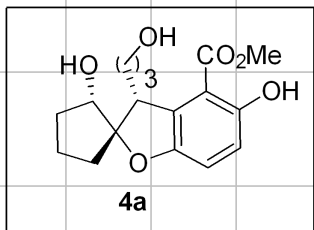


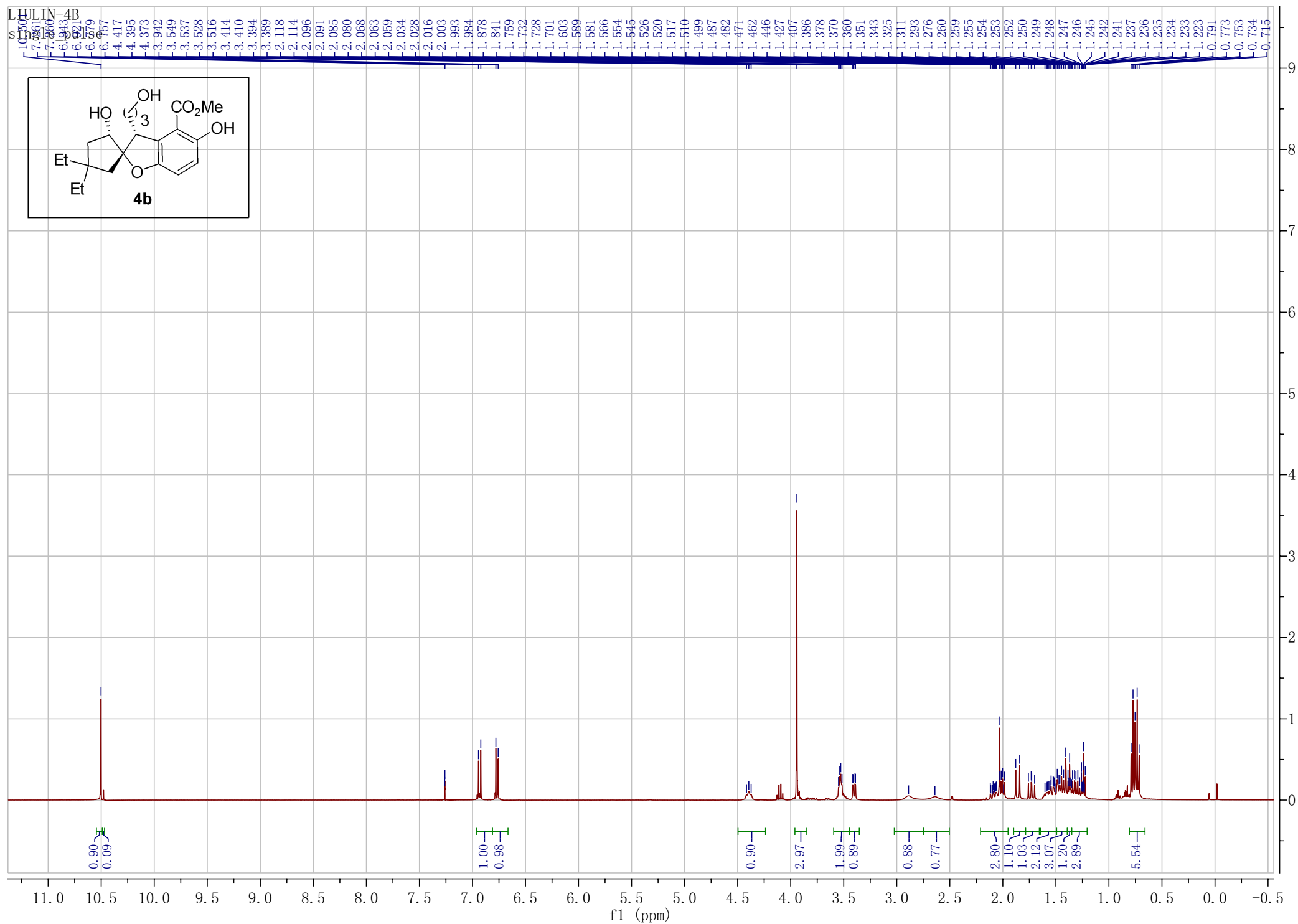


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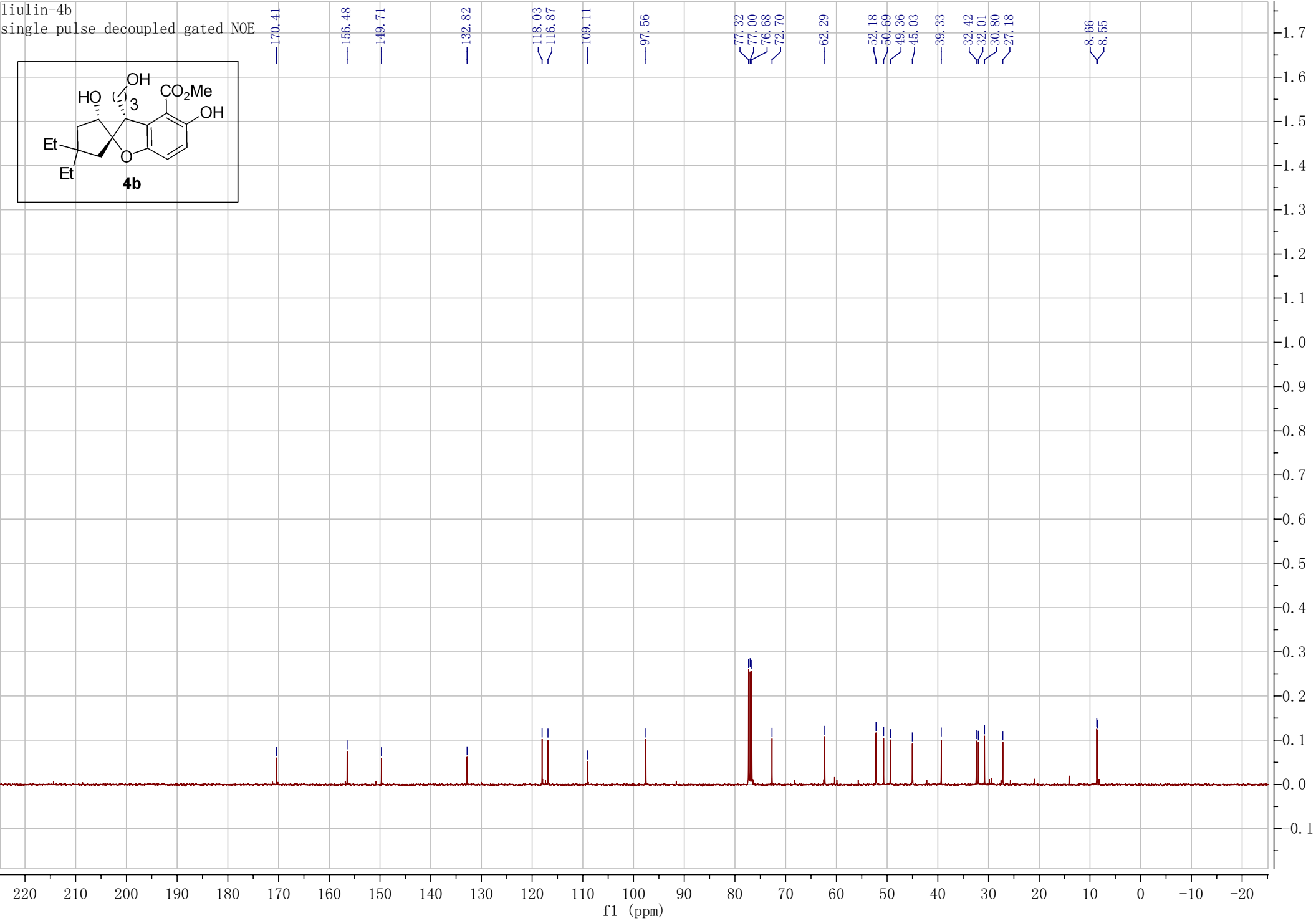
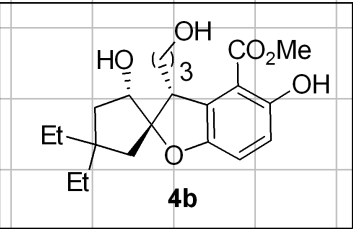


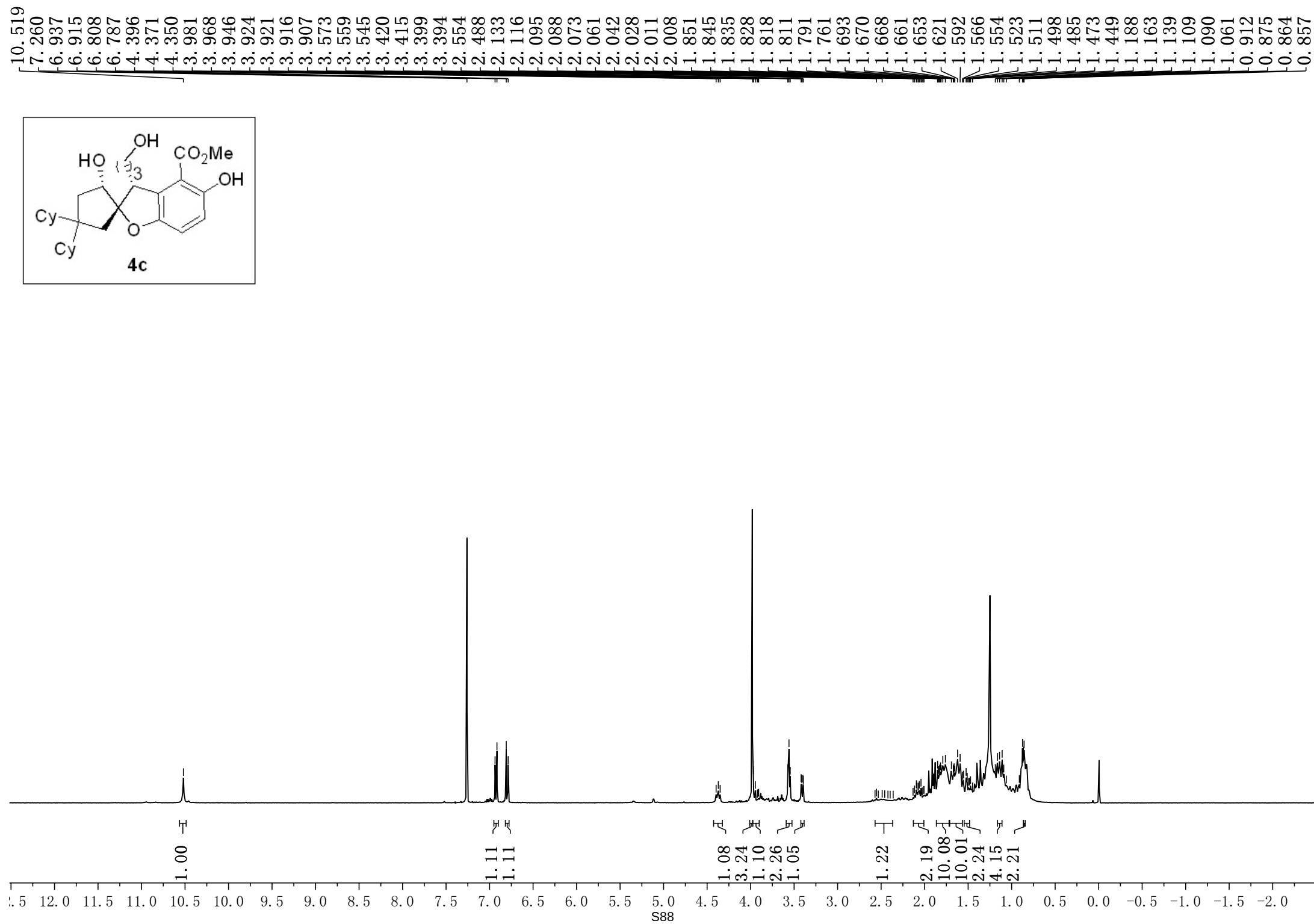
tiulinP1-shouxing





liulin-4b
single pulse decoupled gated NOE



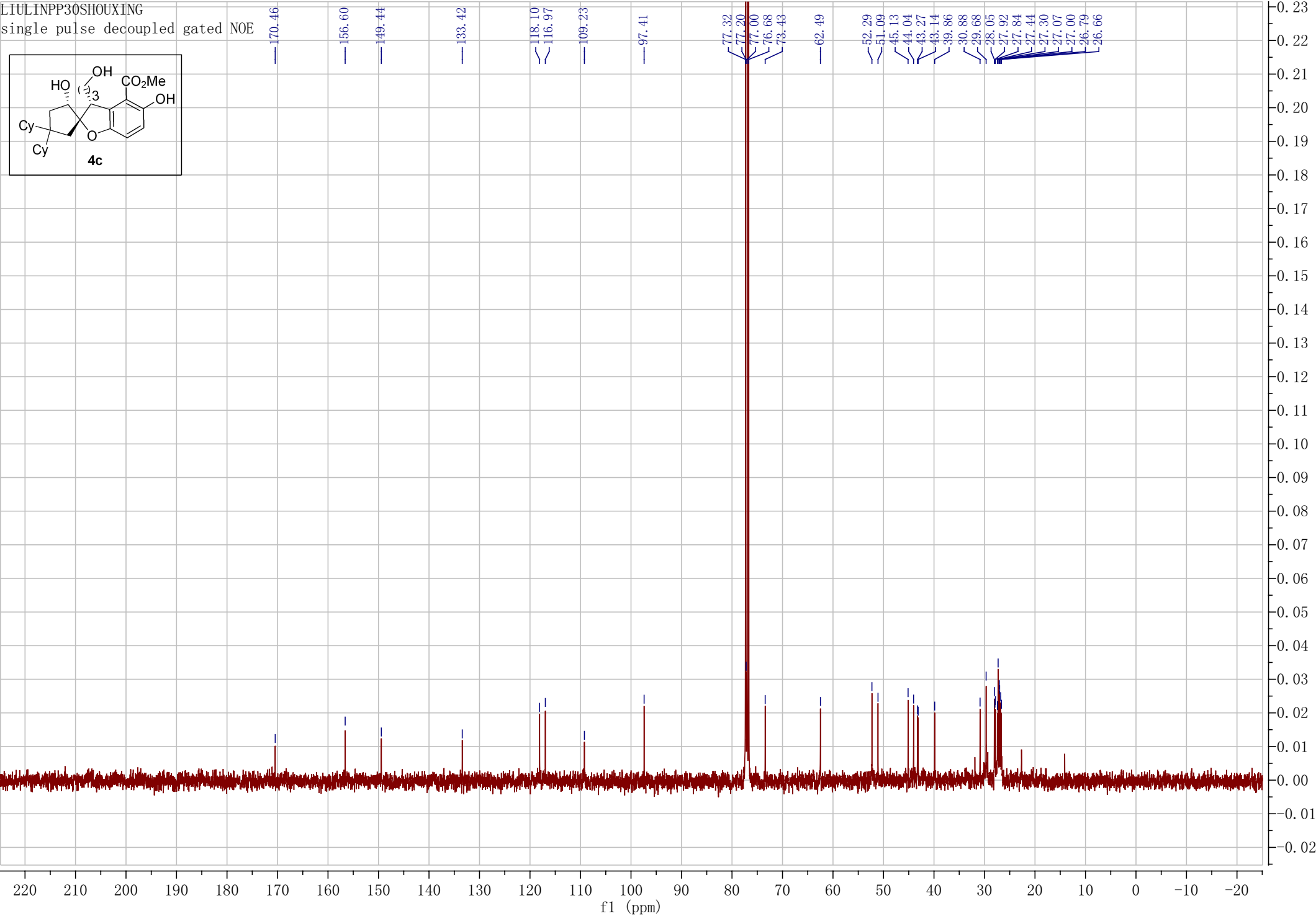
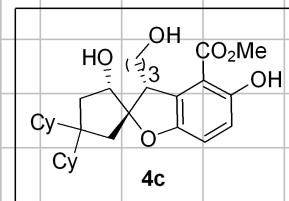


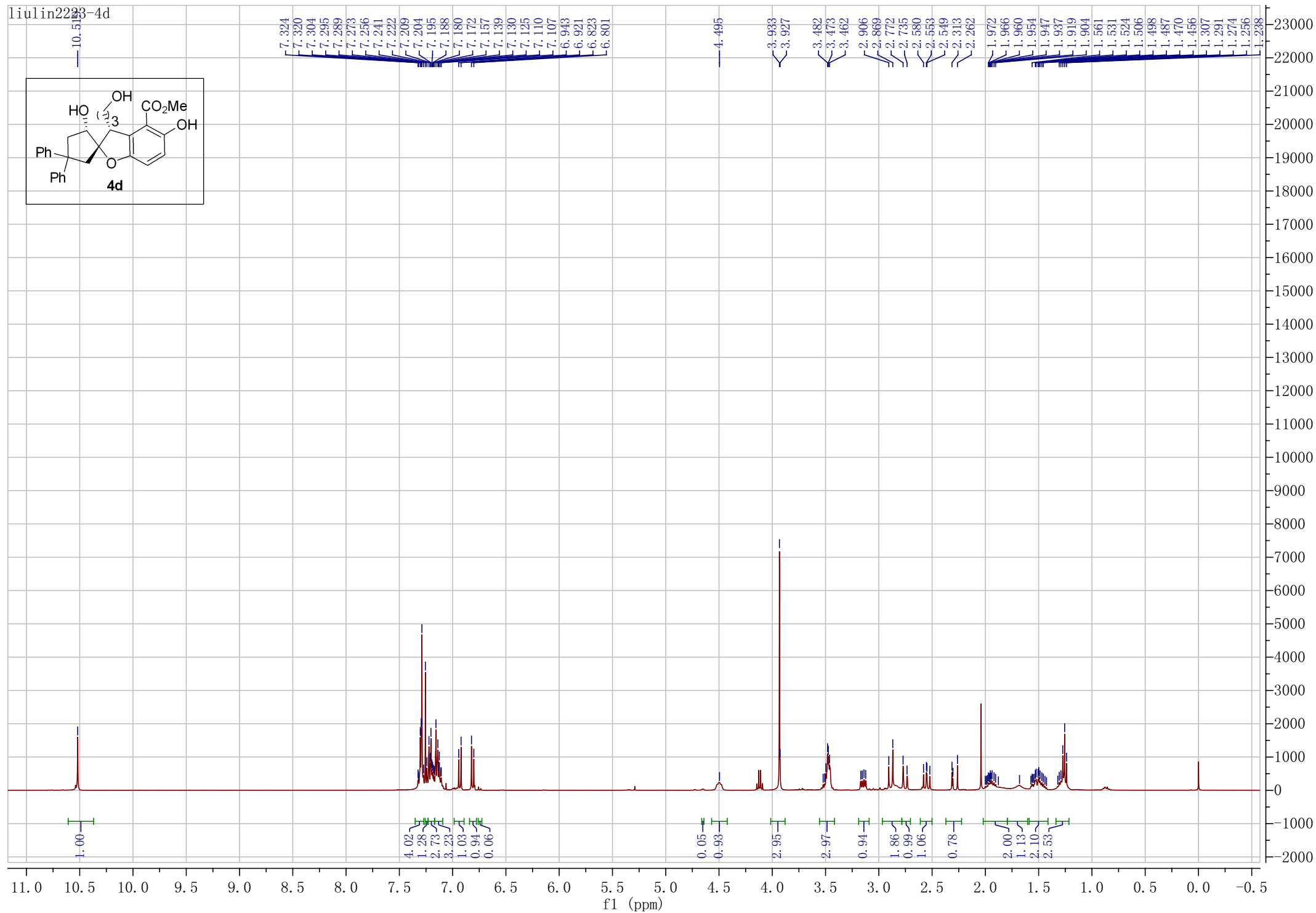
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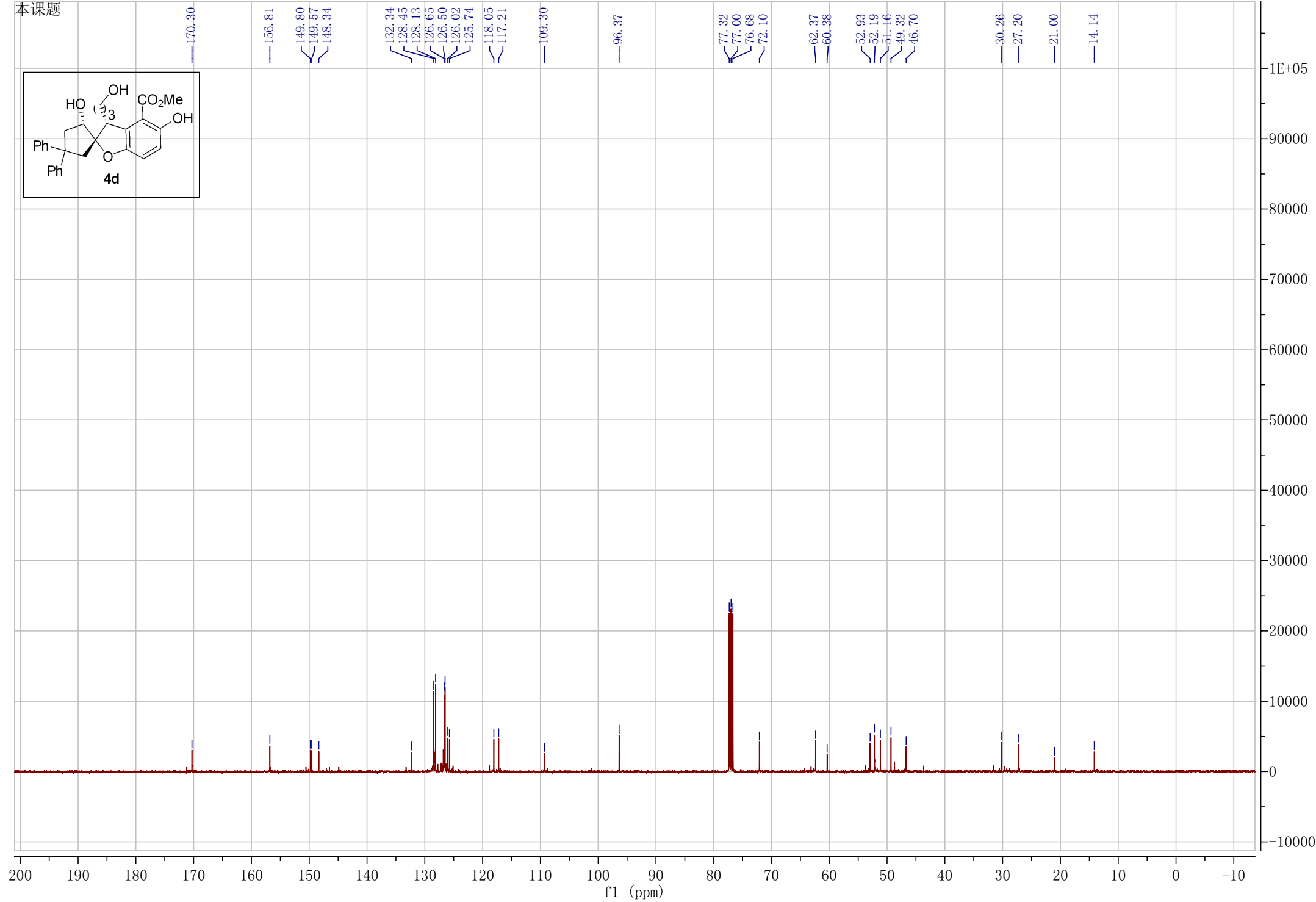
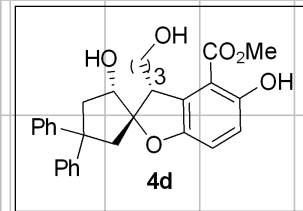
single pulse decoupled

gated

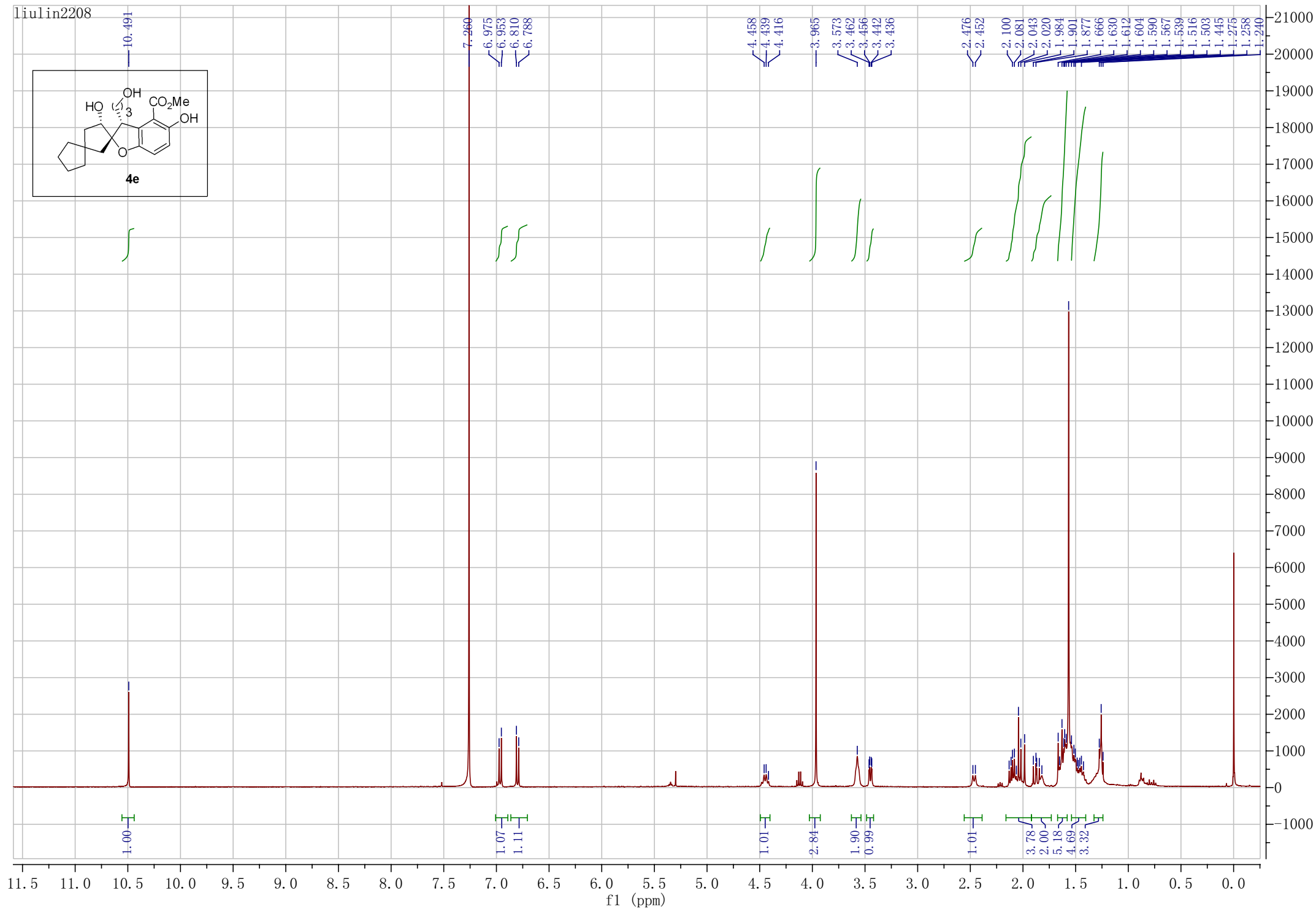
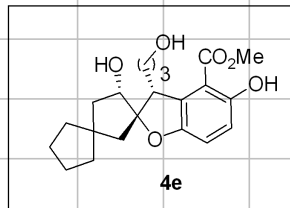
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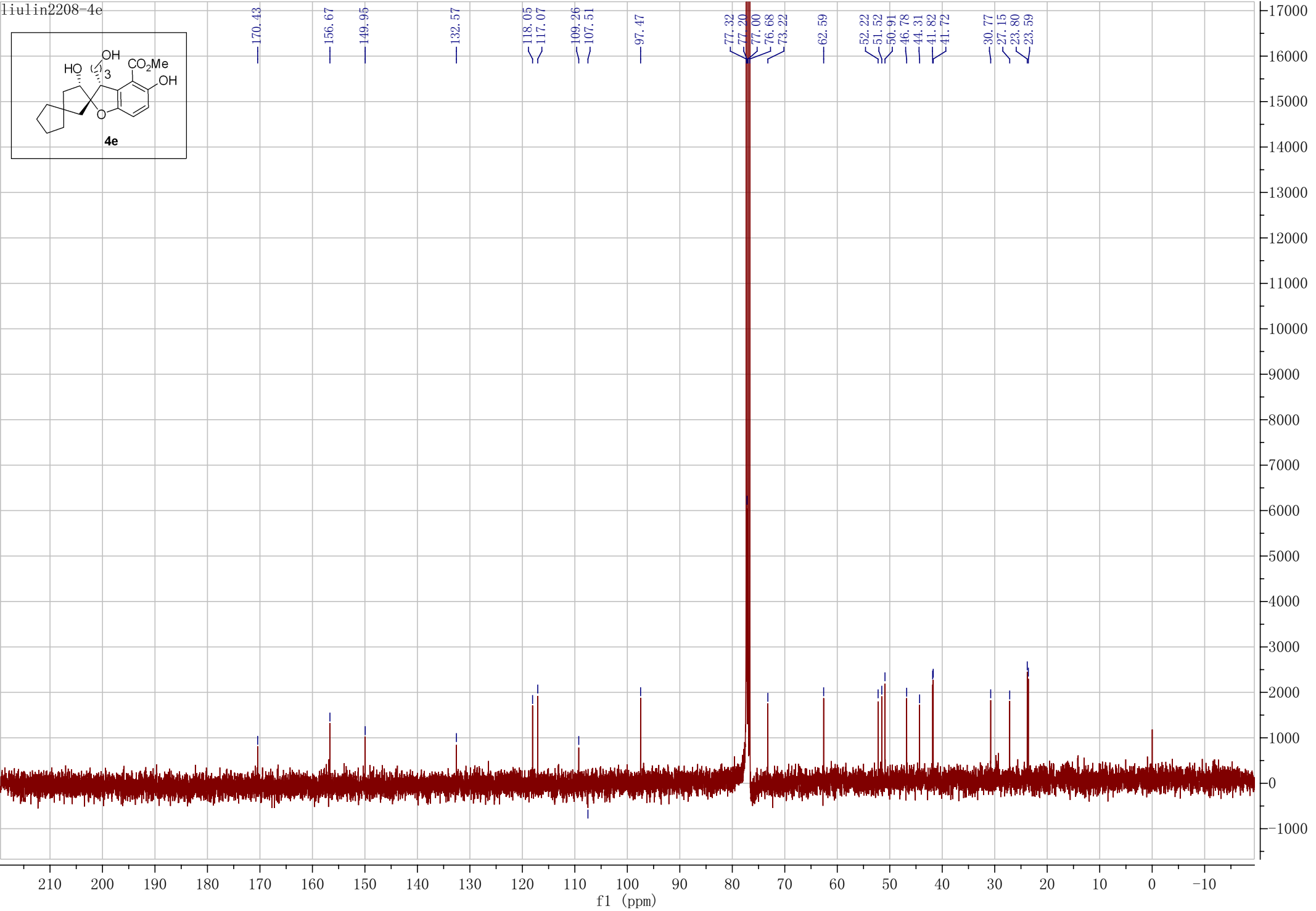
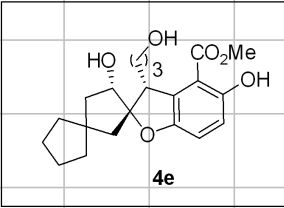




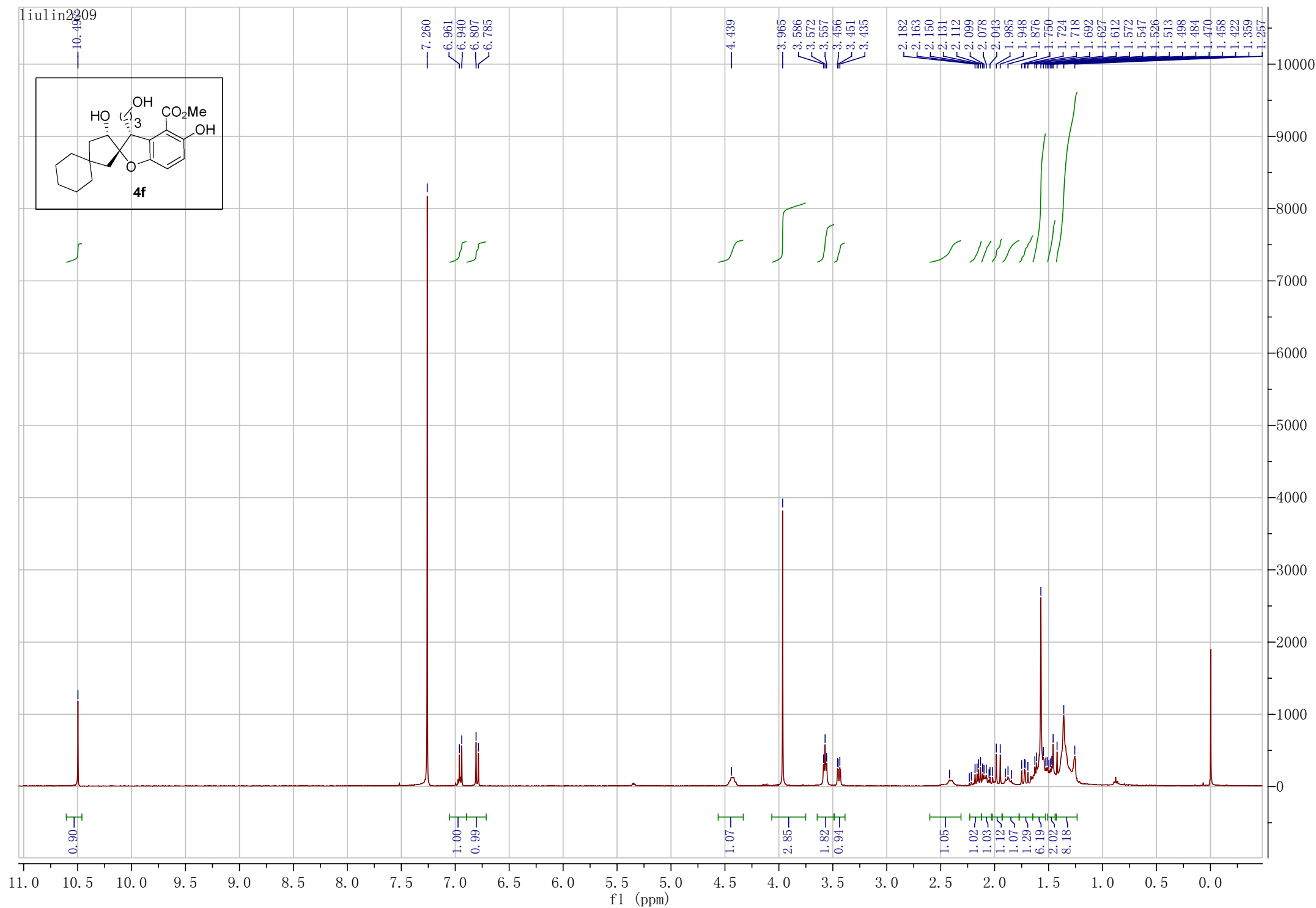
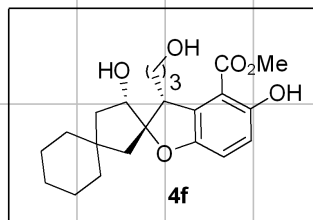
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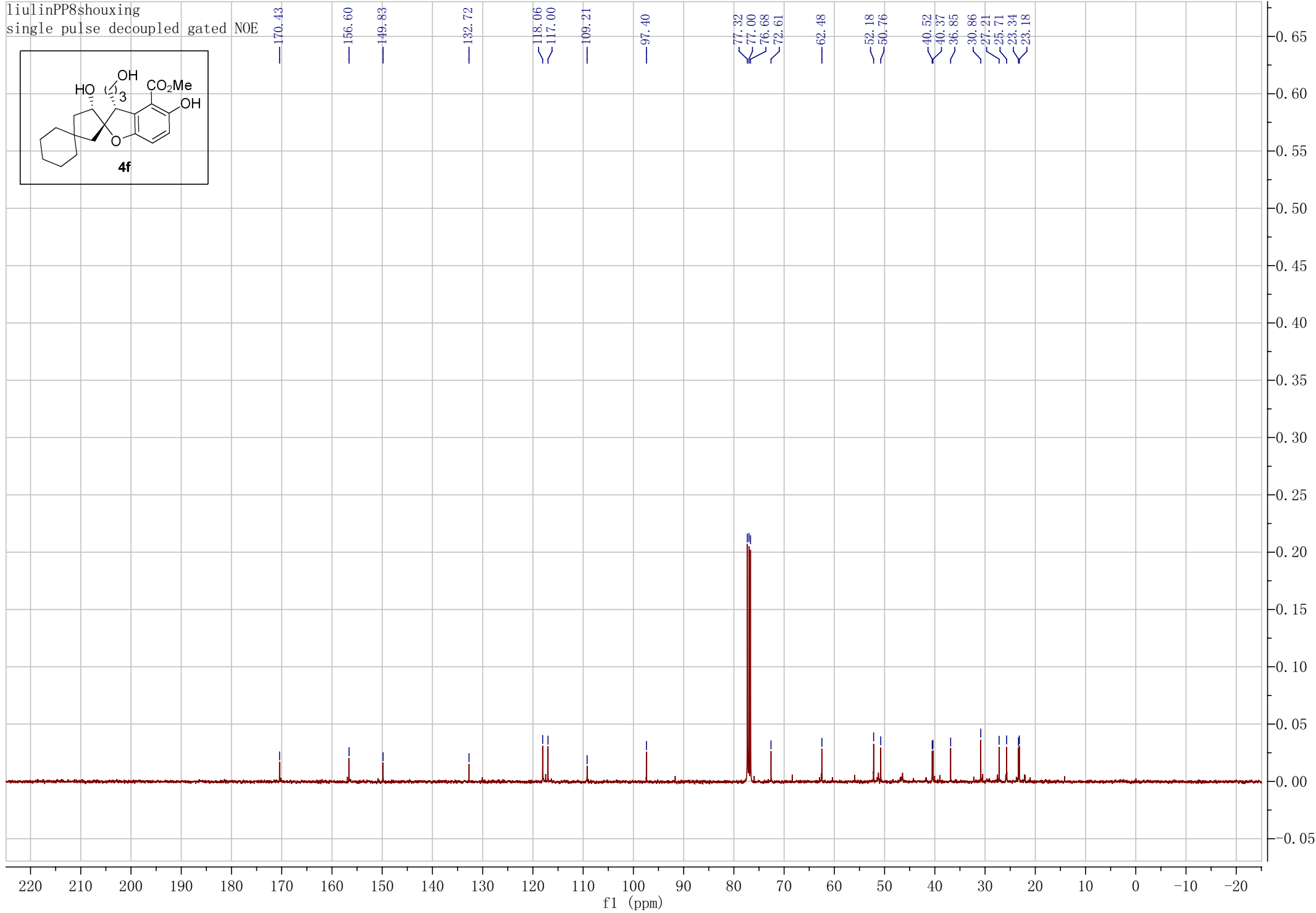
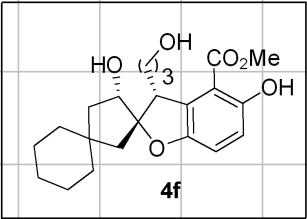
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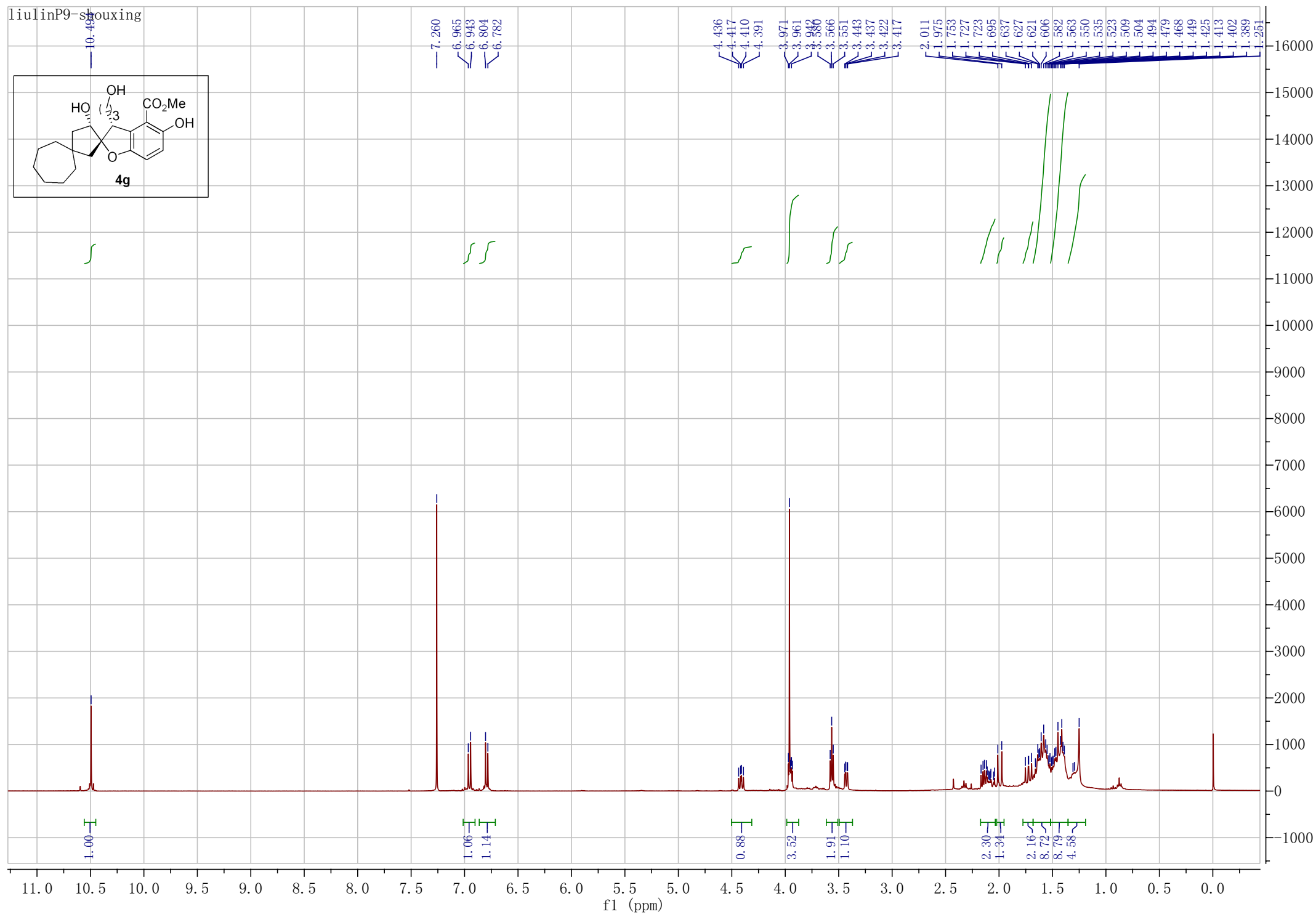


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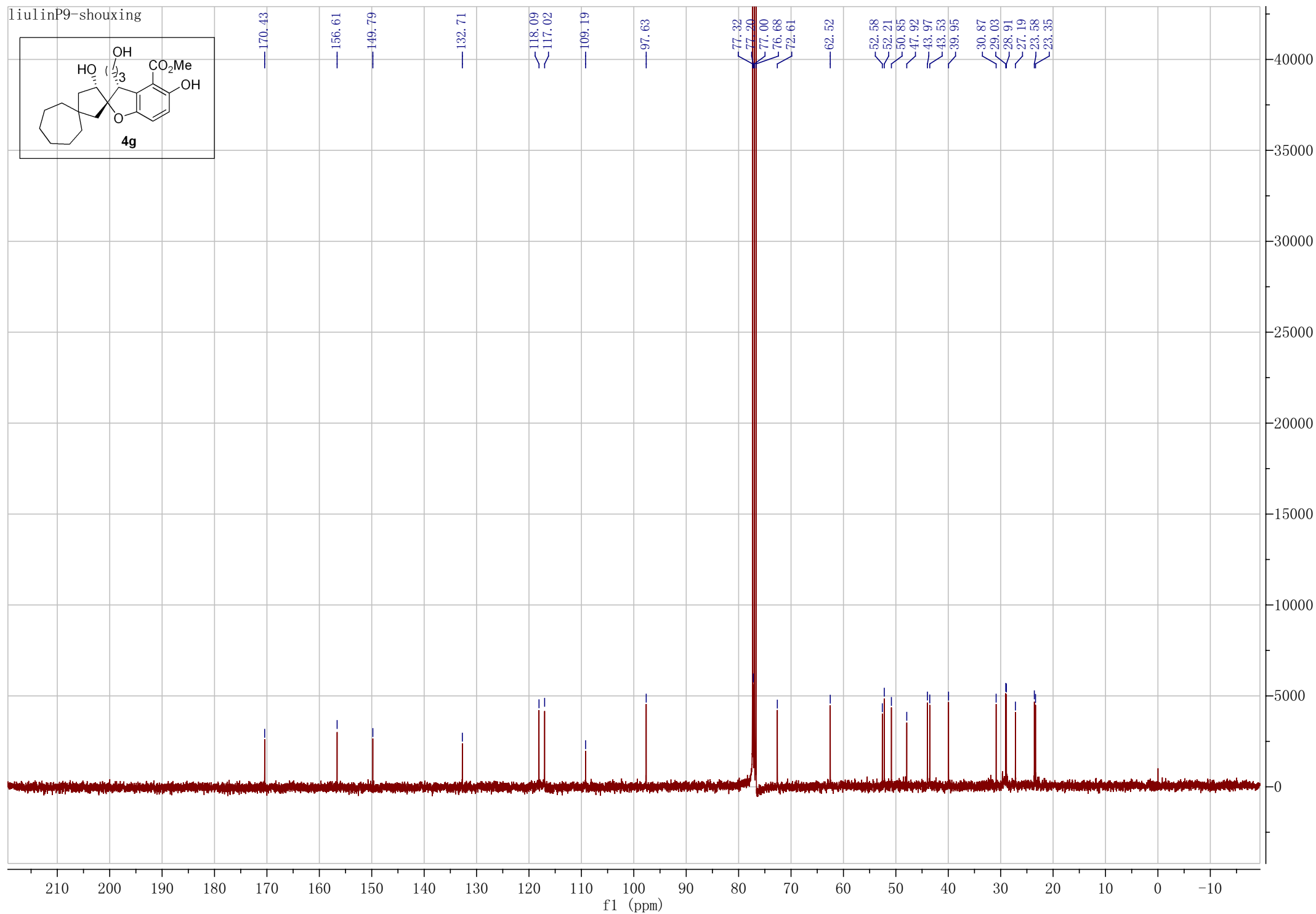
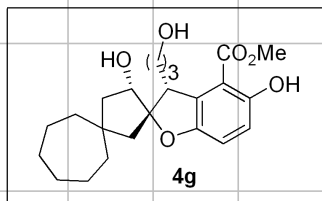


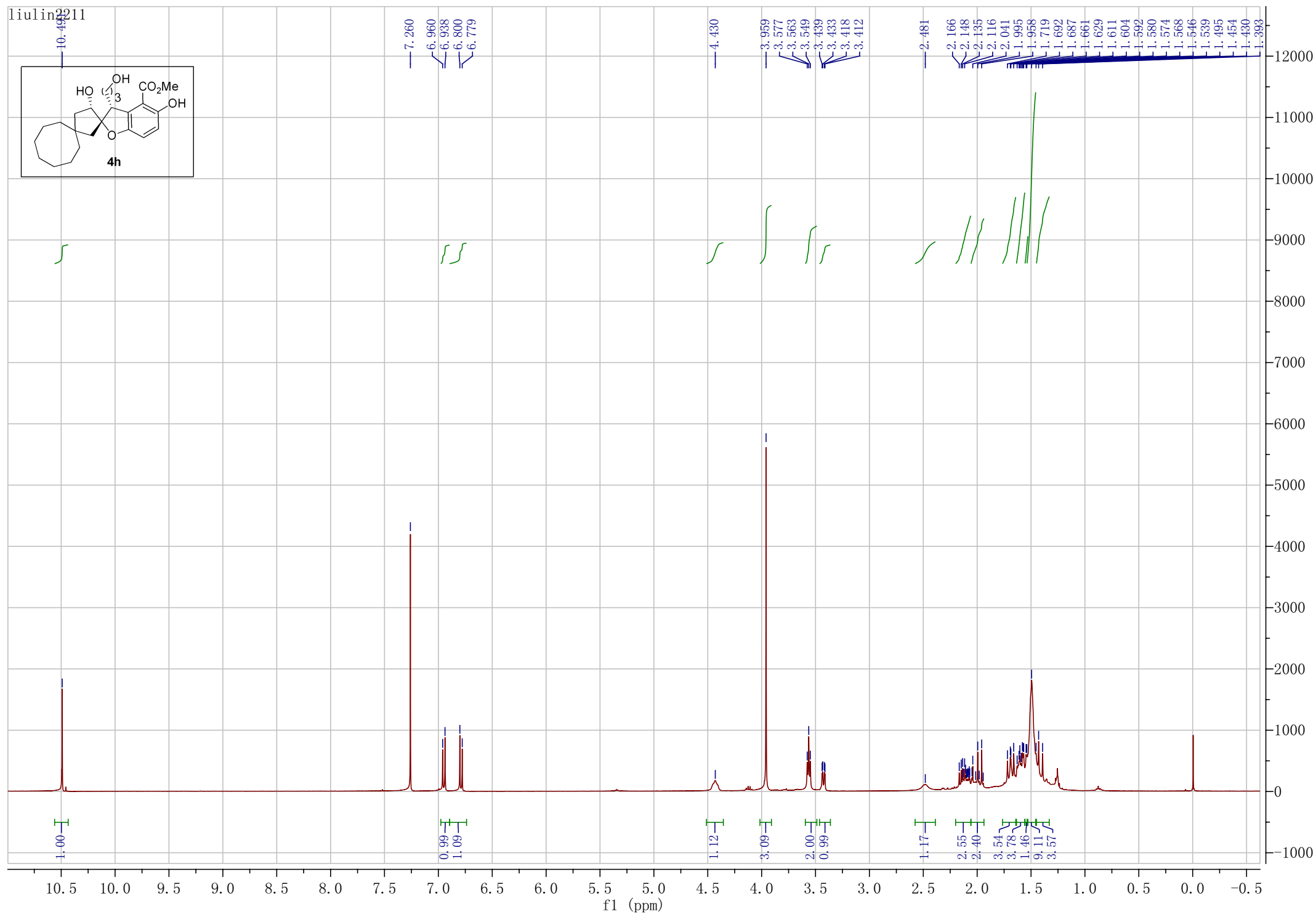
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single pulse decoupled gated NOE





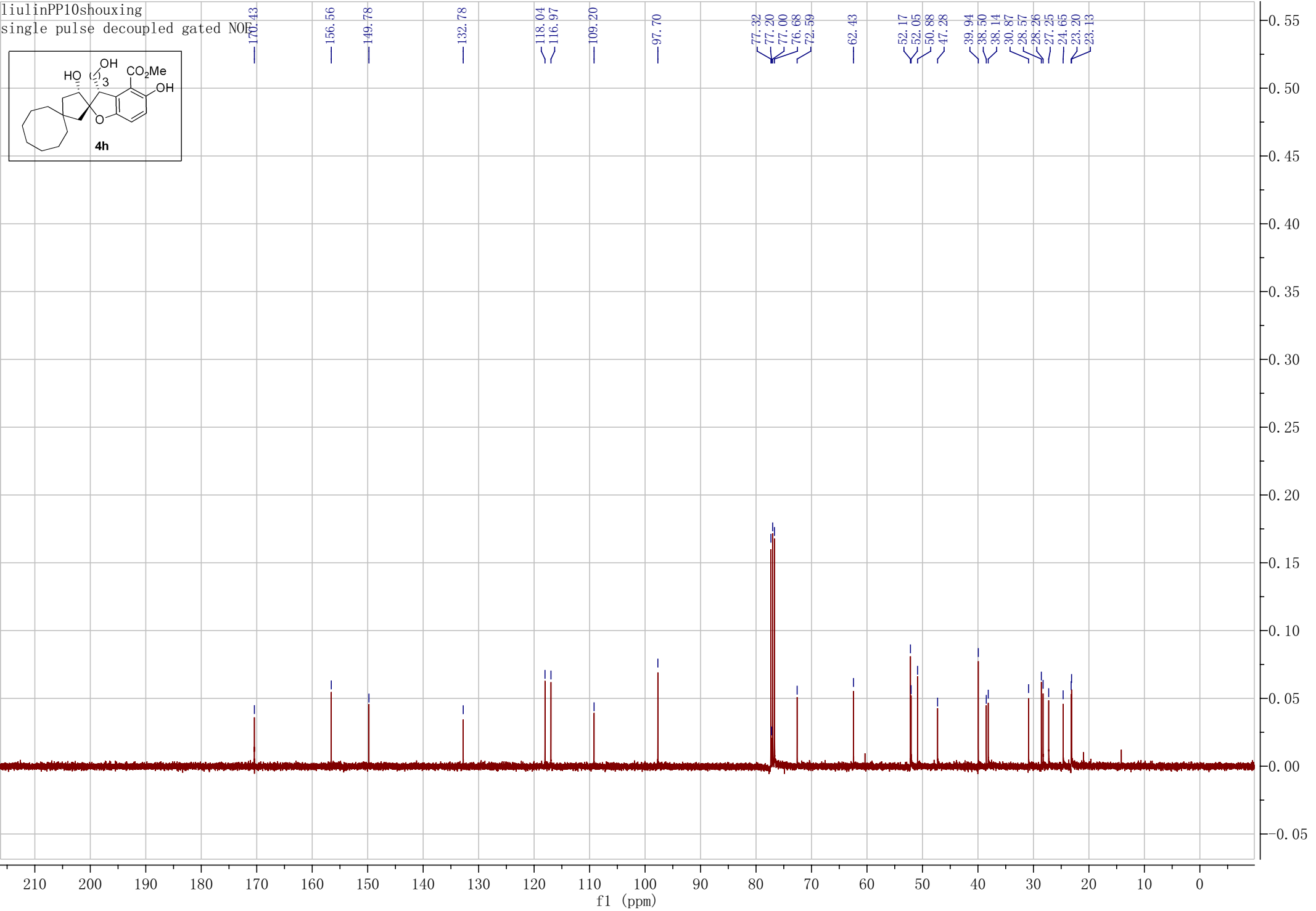
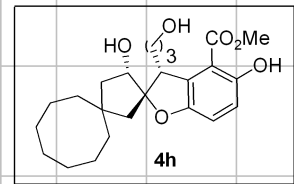
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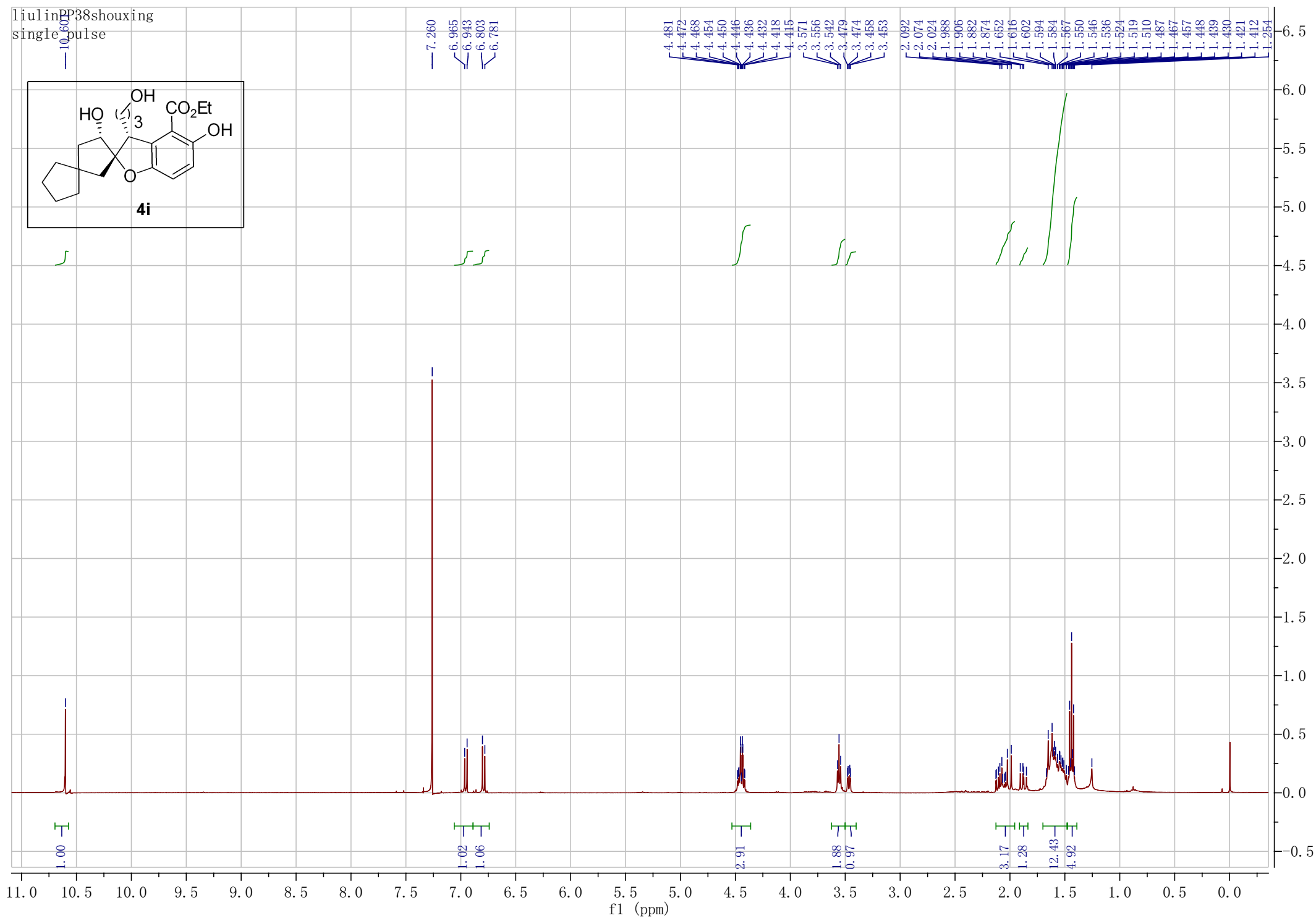
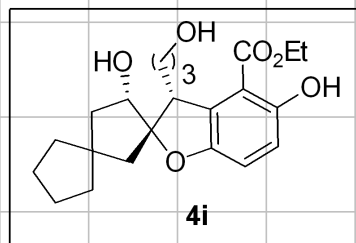


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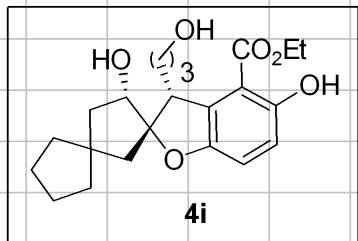
single pulse decoupled gated NOE



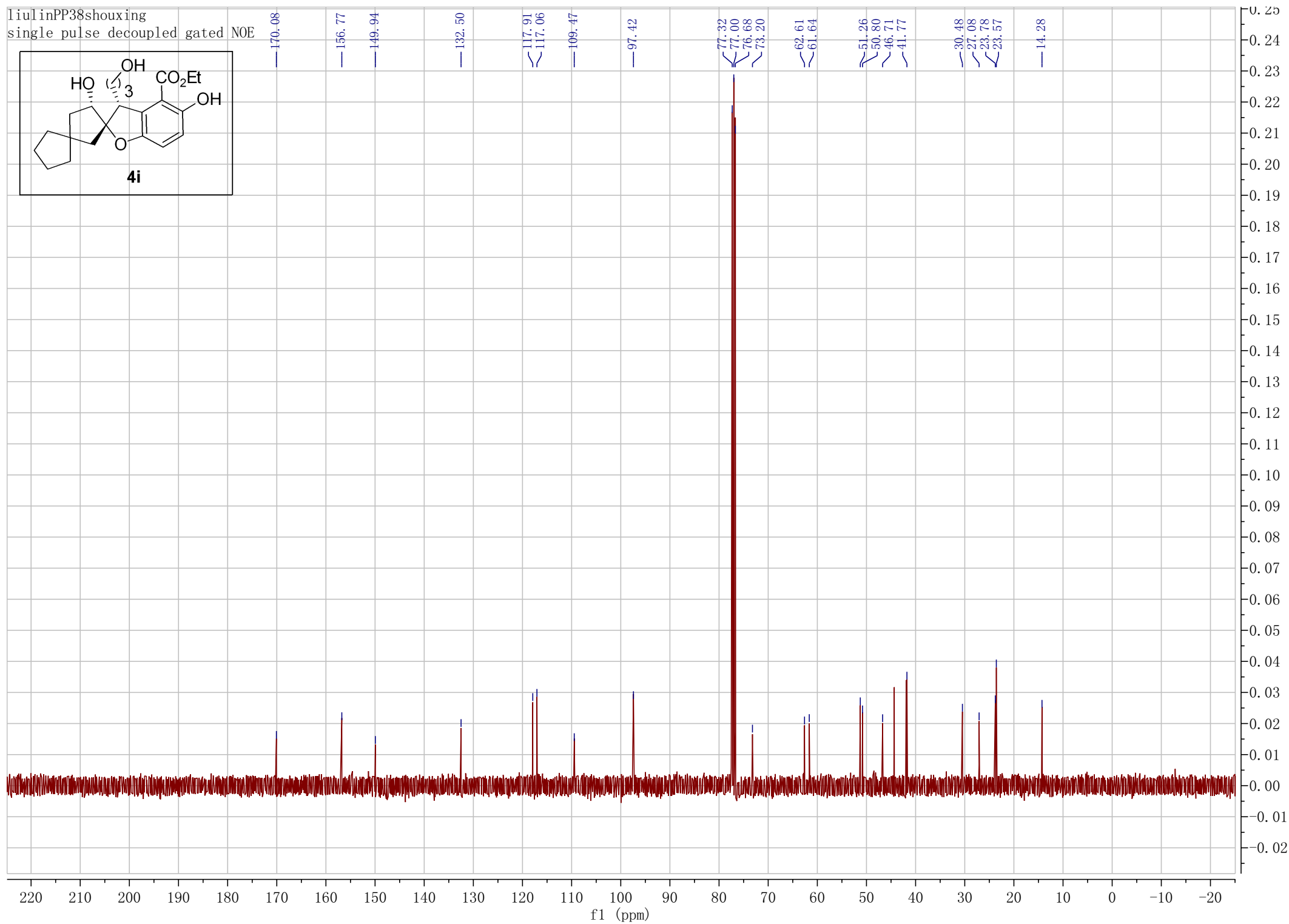
liulinPP38shouxiang
single pulse



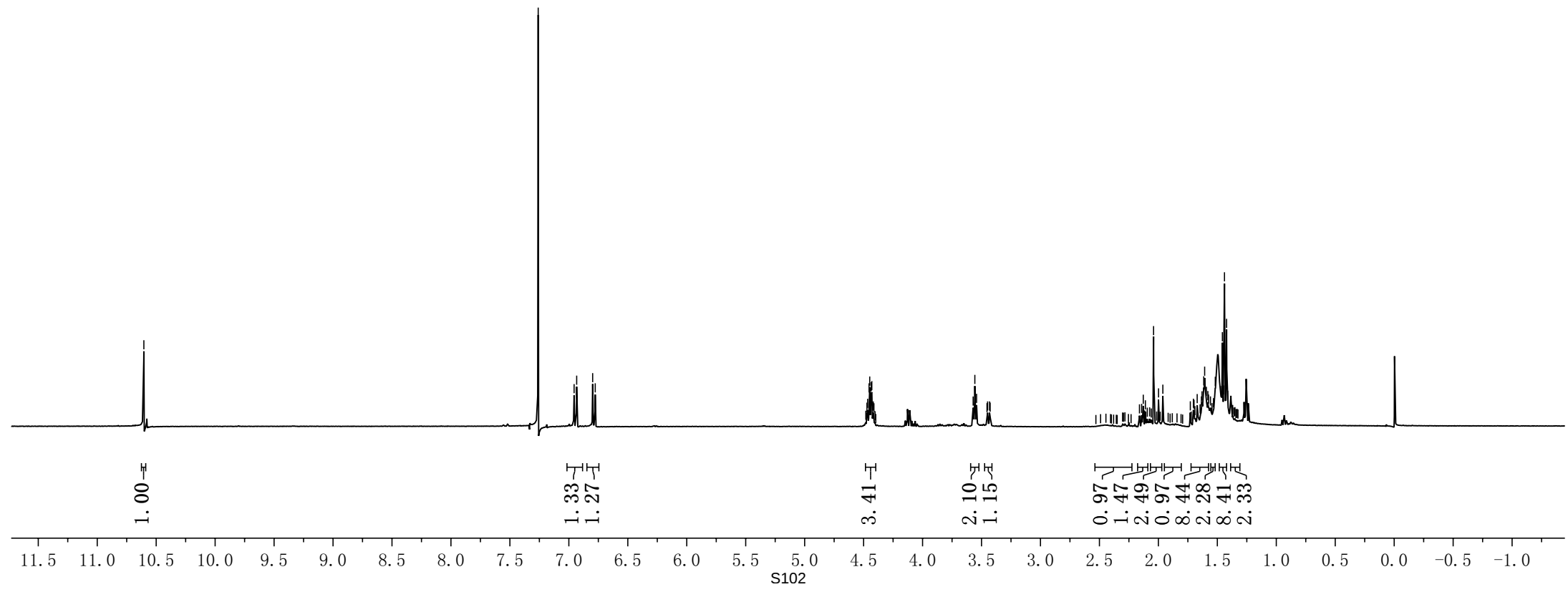
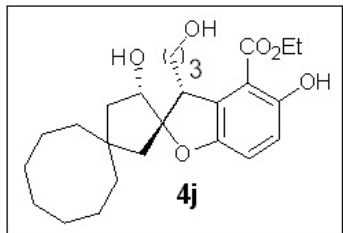
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single pulse decoupled gated NOE



NOE

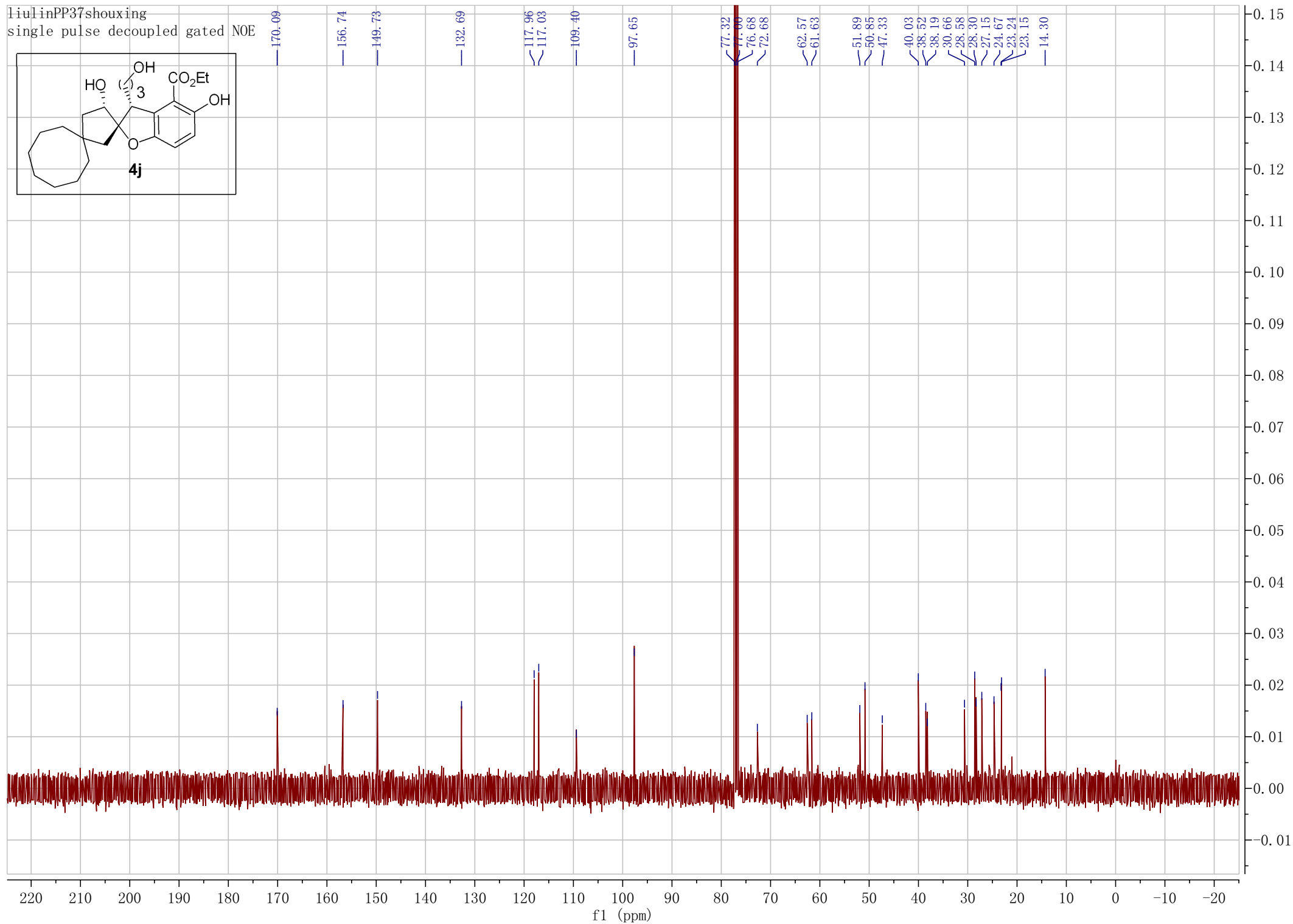
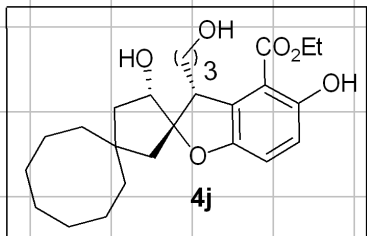


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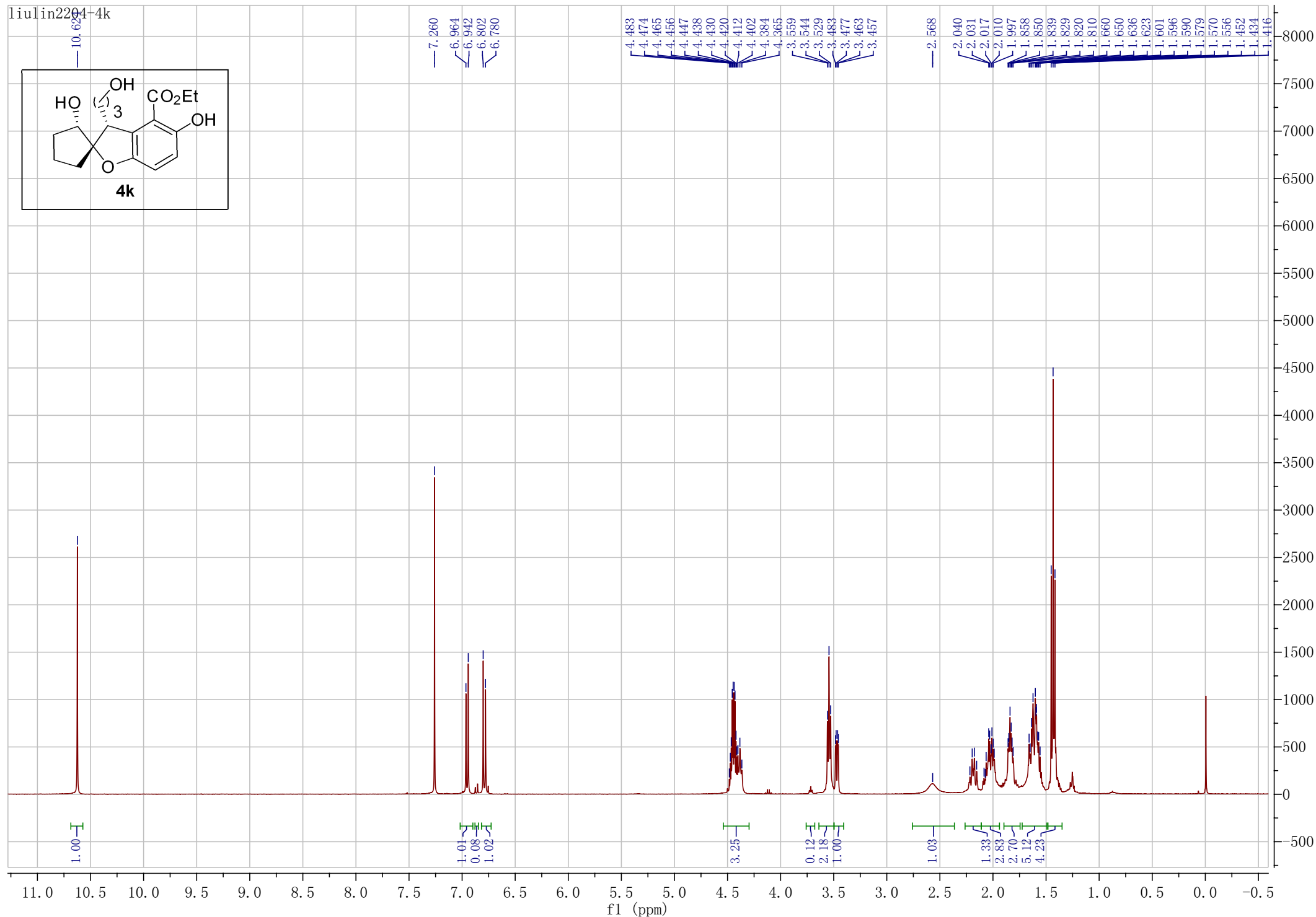
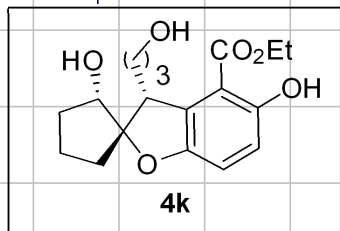
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single pulse decoupled gated NOE

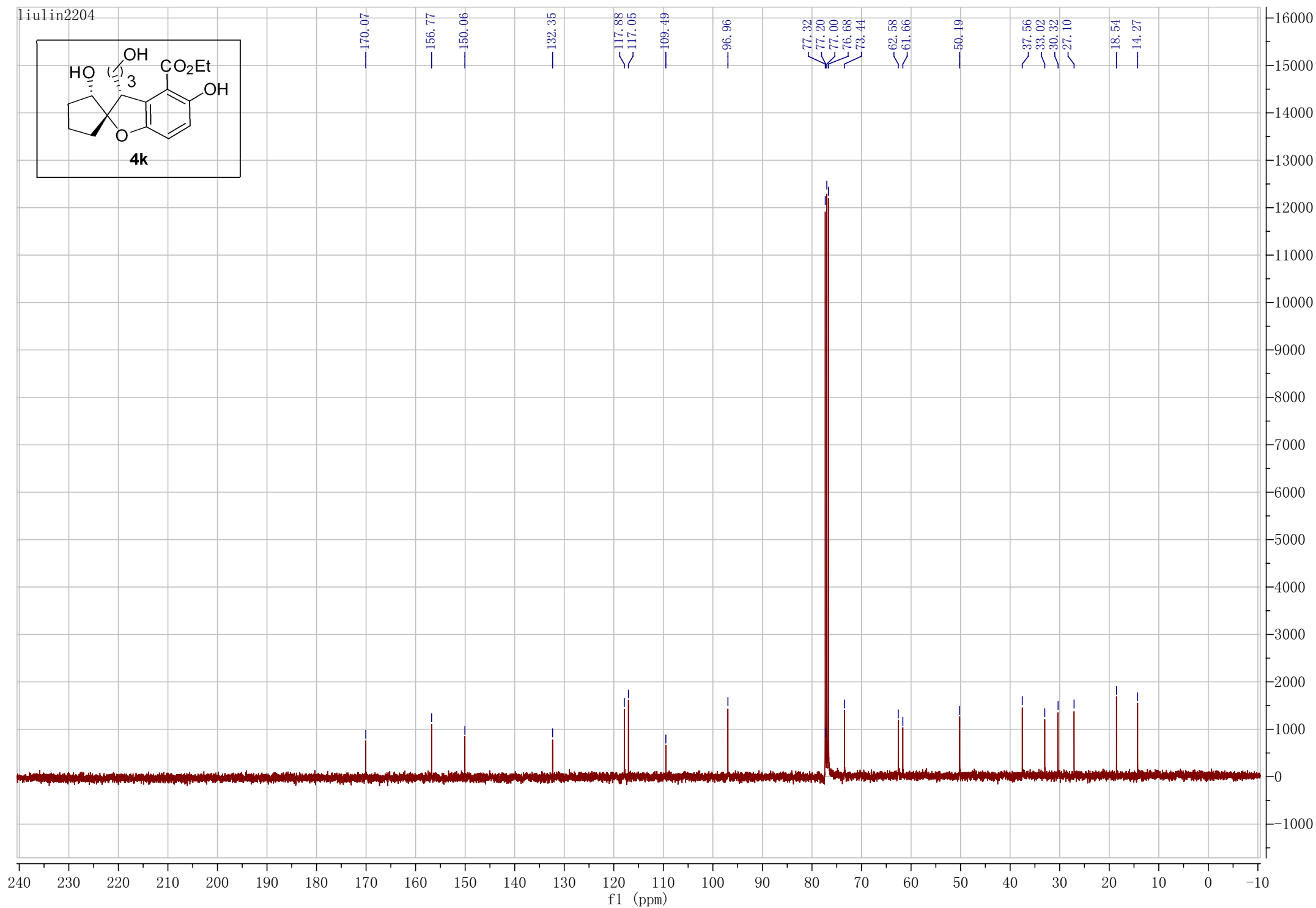
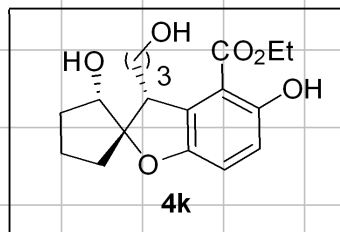


S103

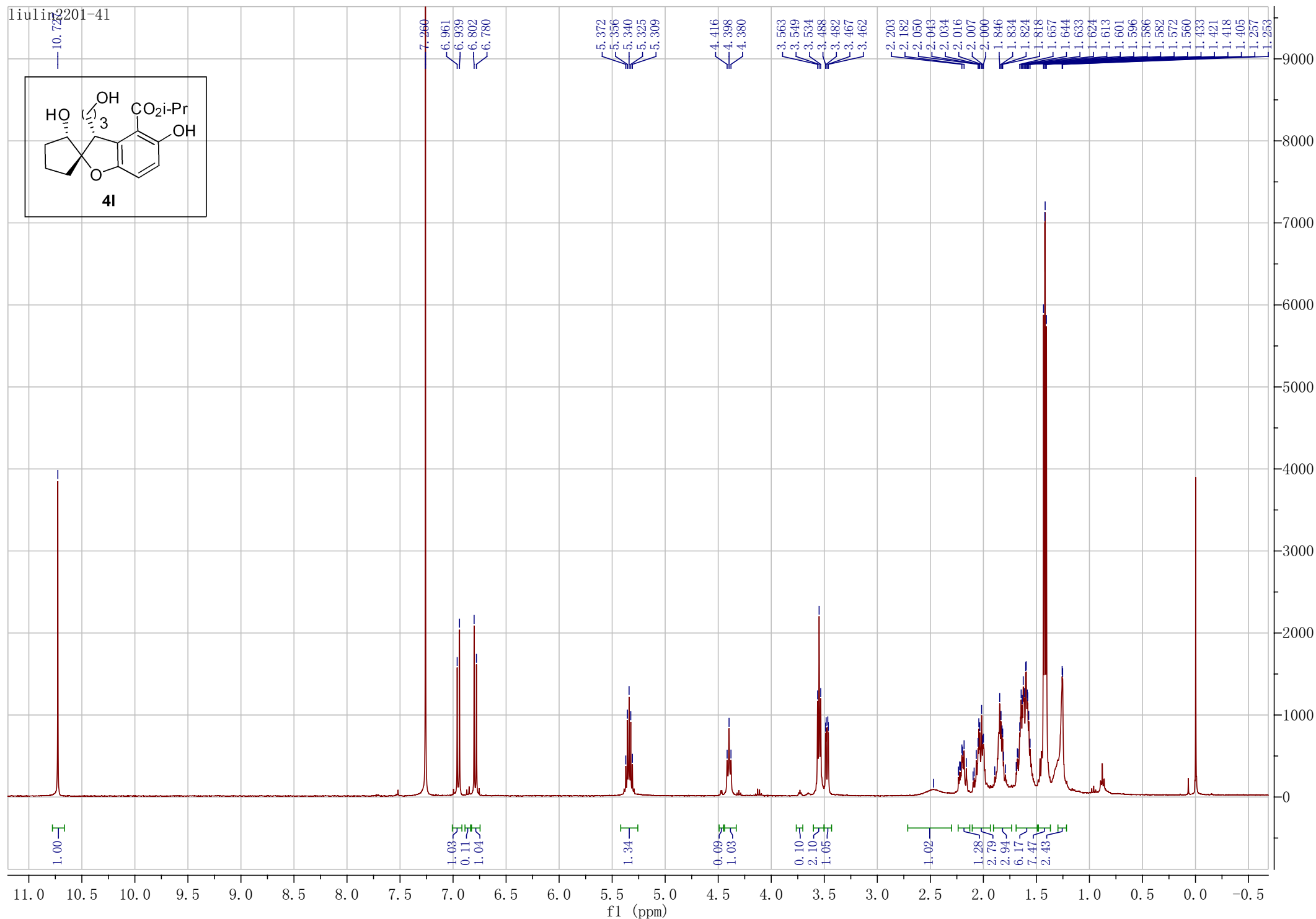
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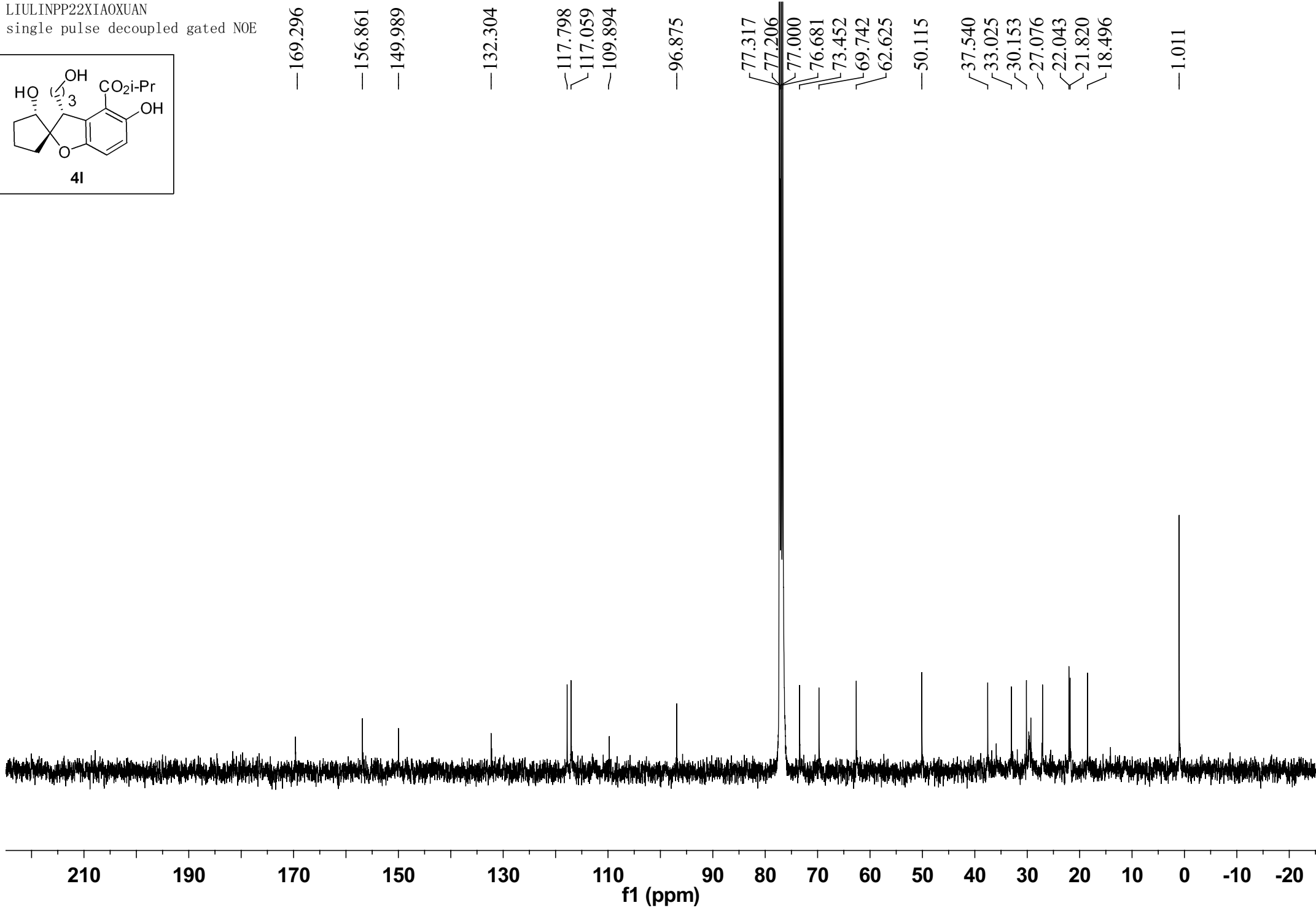
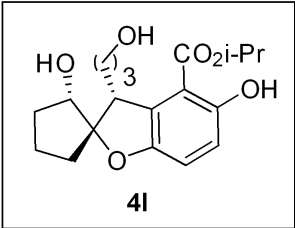
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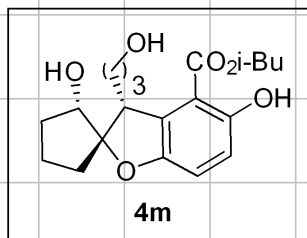
S105



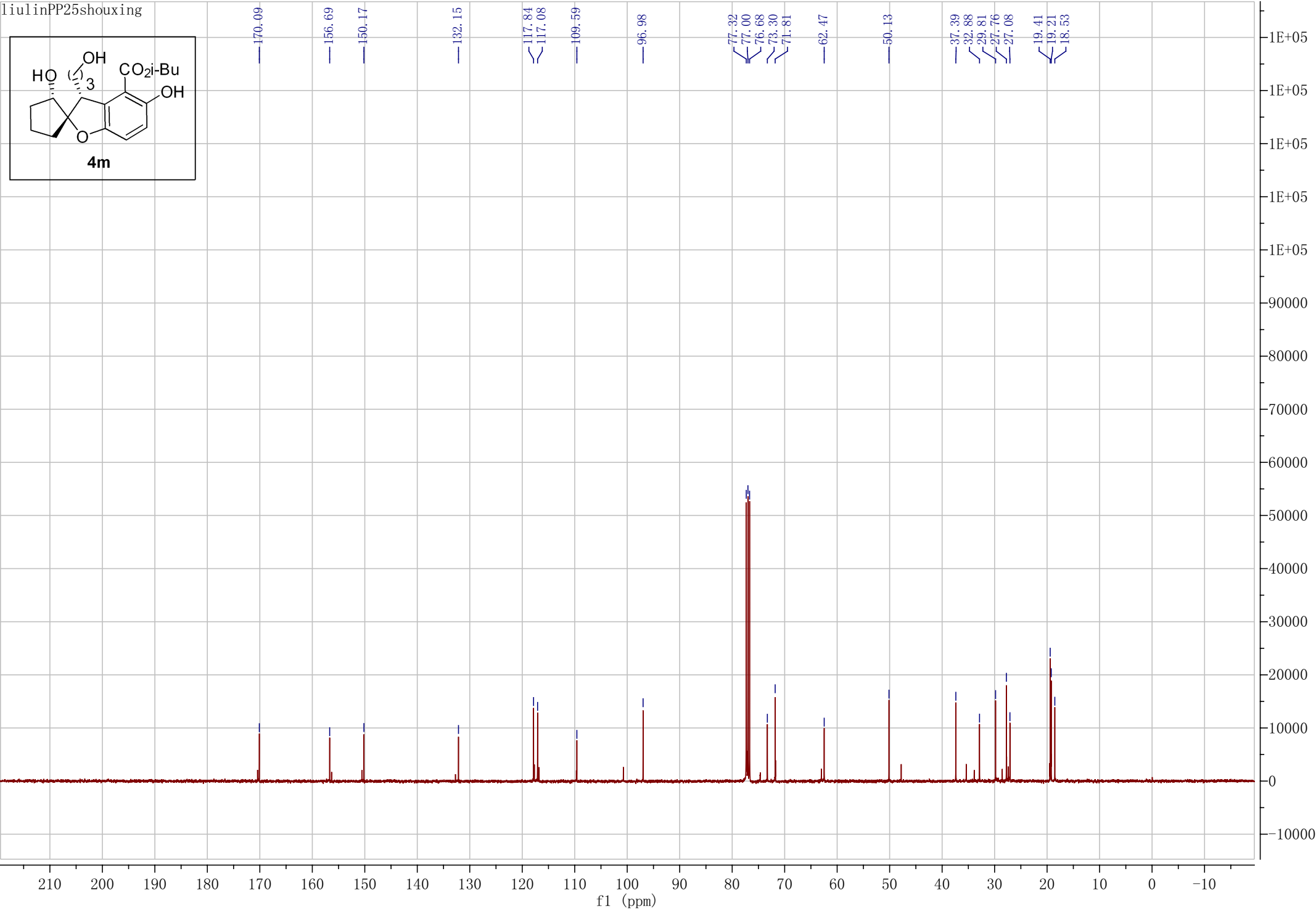
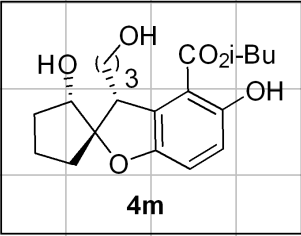
LIULINPP22XIAOXUAN
single pulse decoupled gated NOE

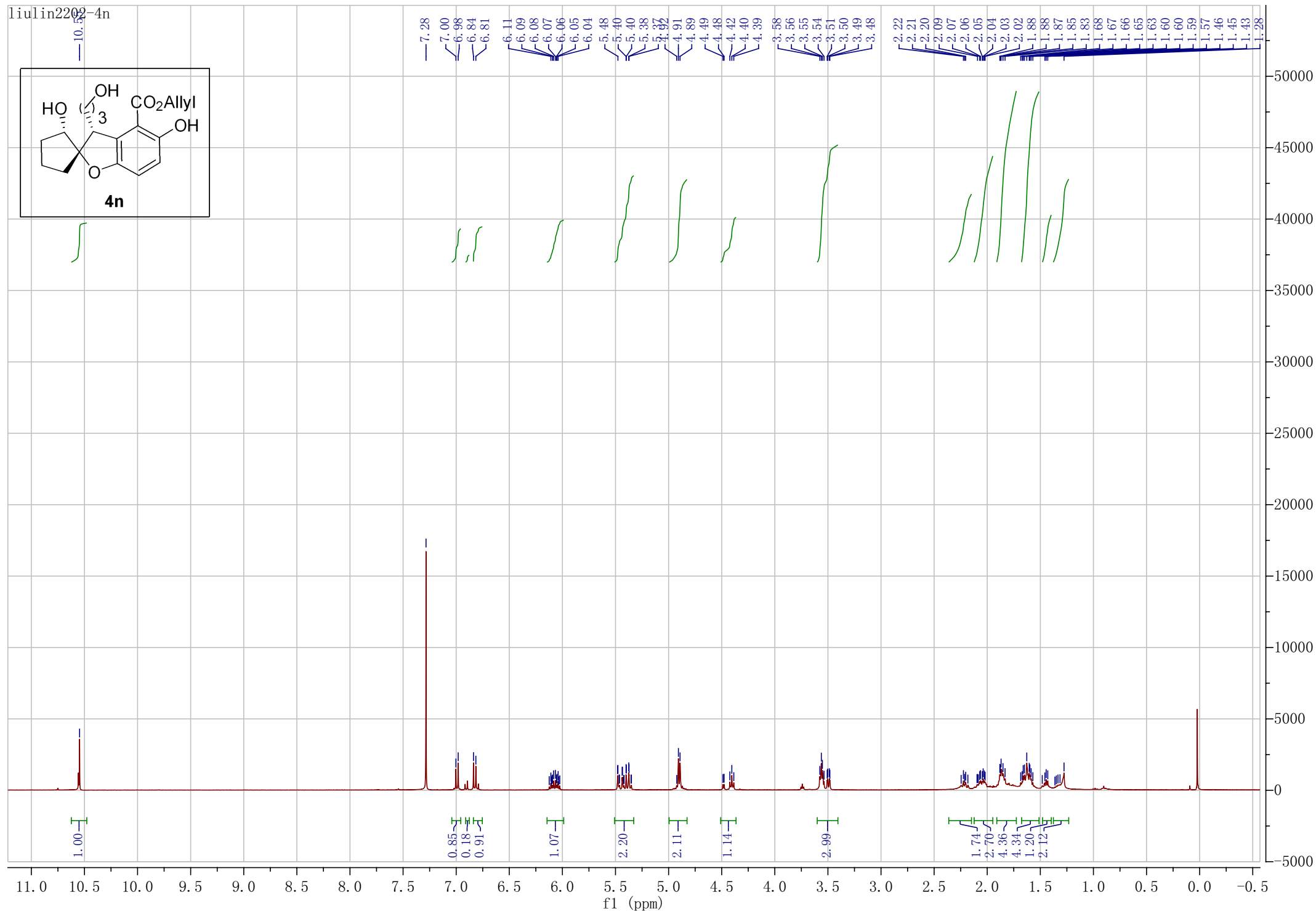


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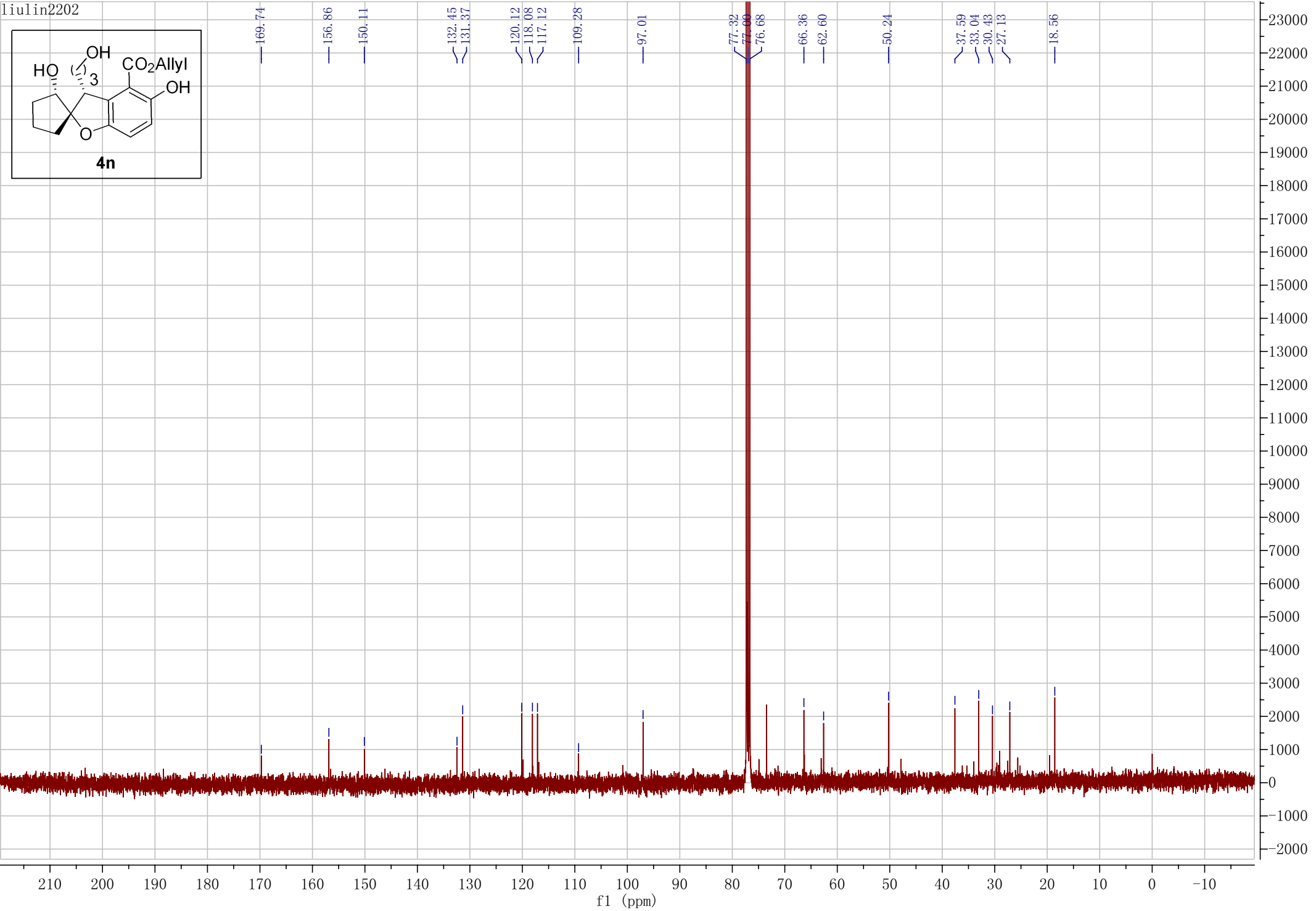
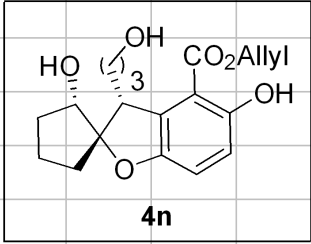


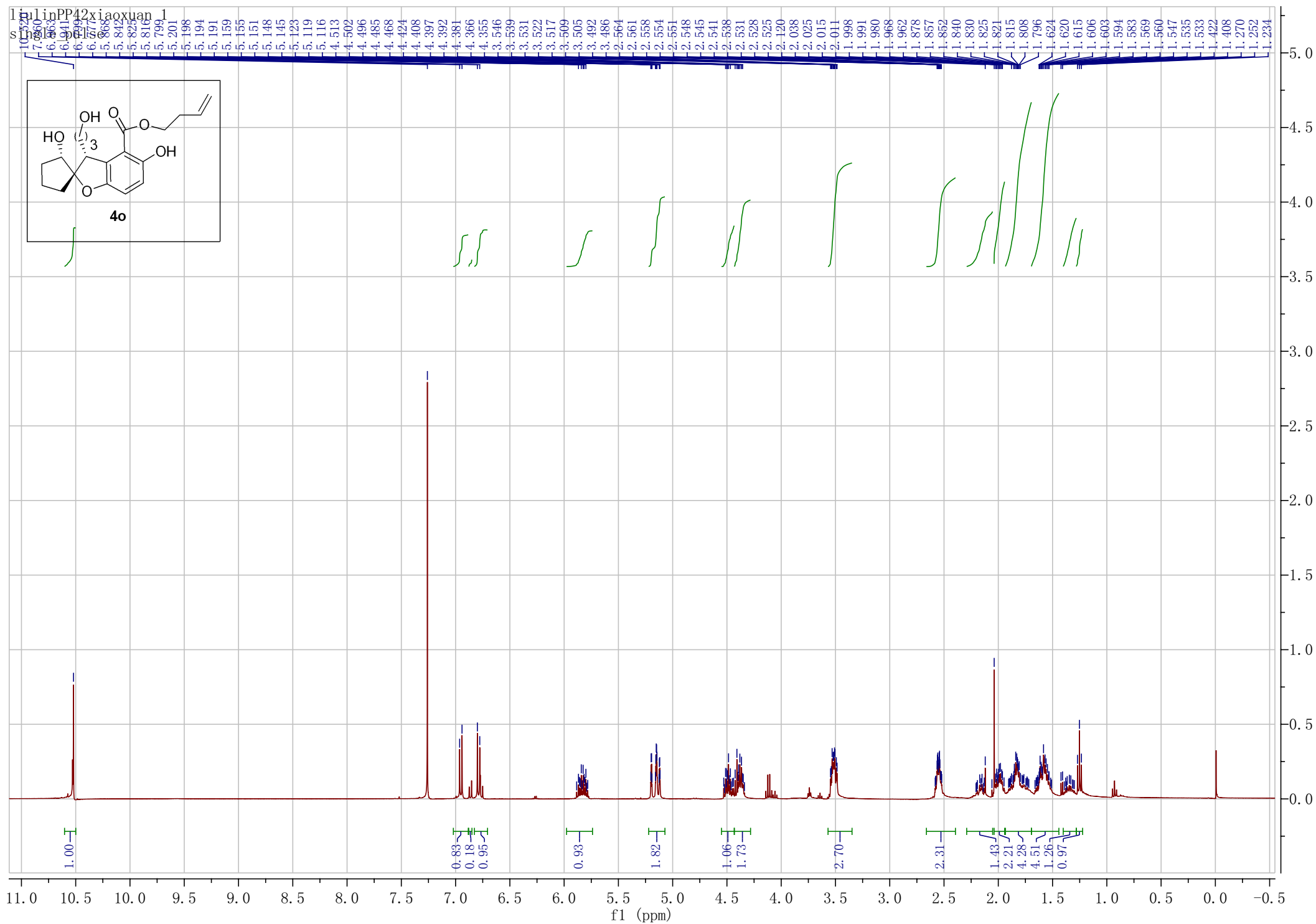
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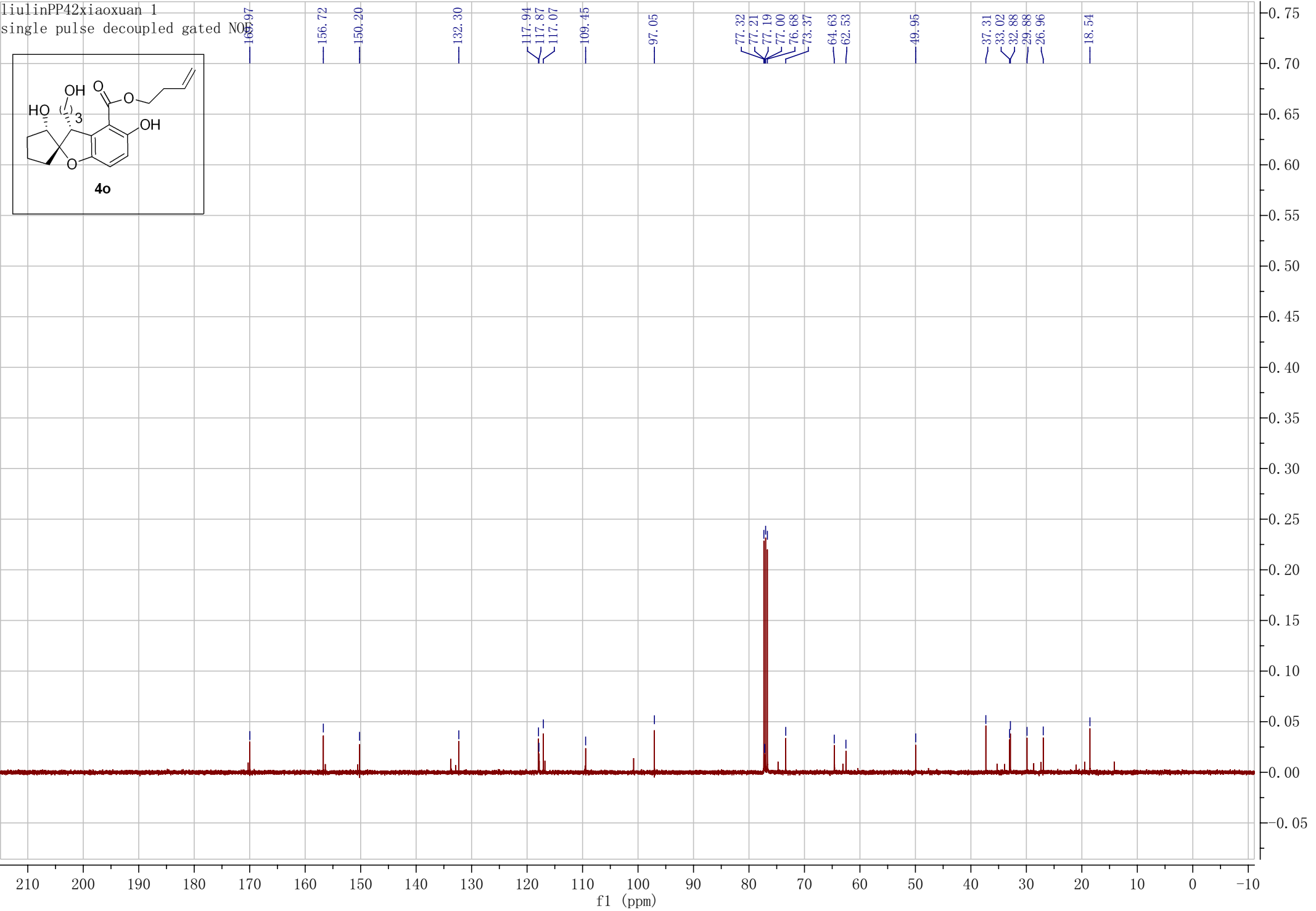
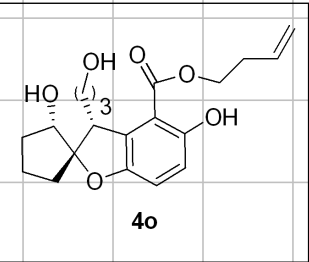
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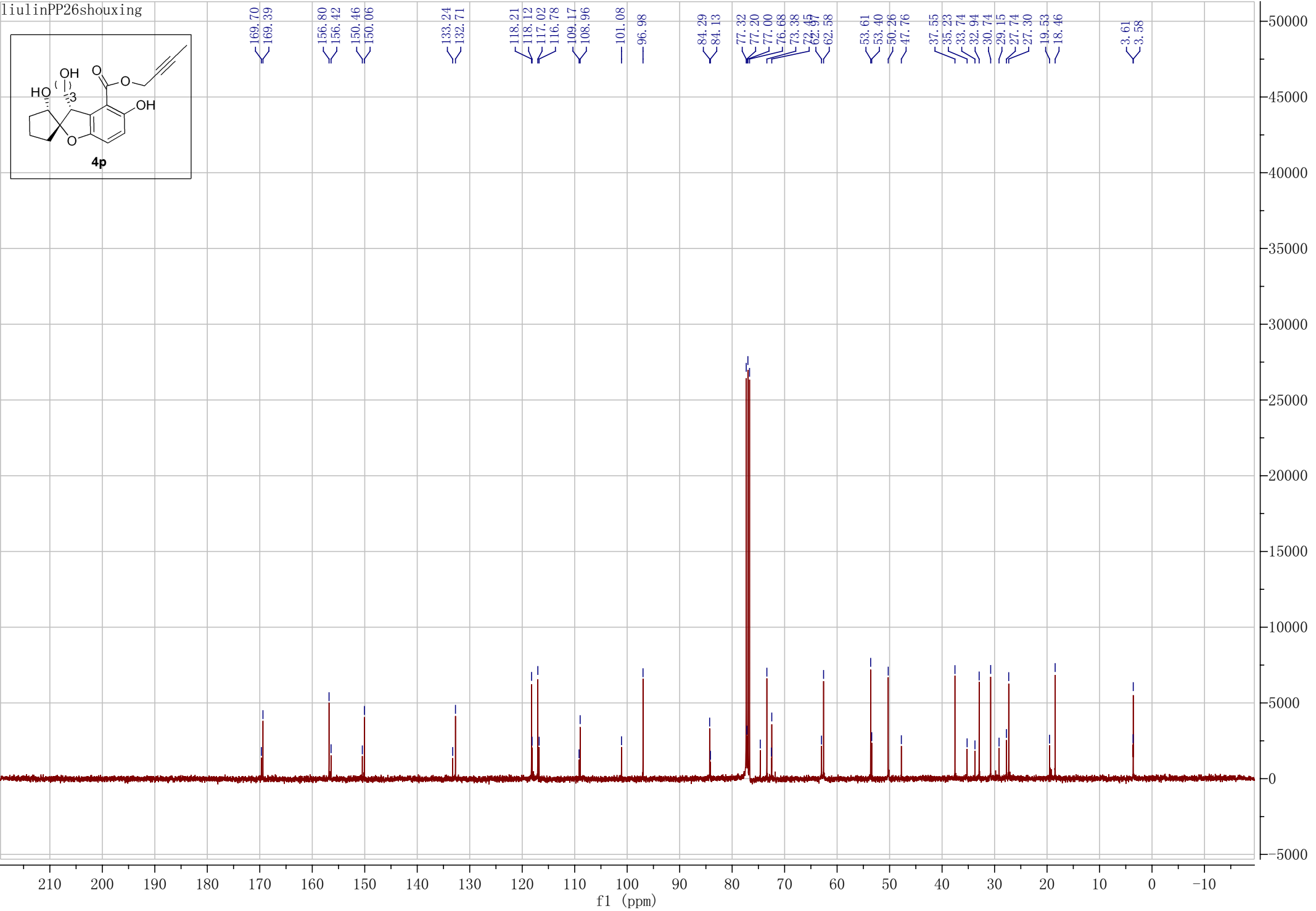
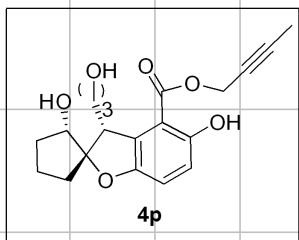


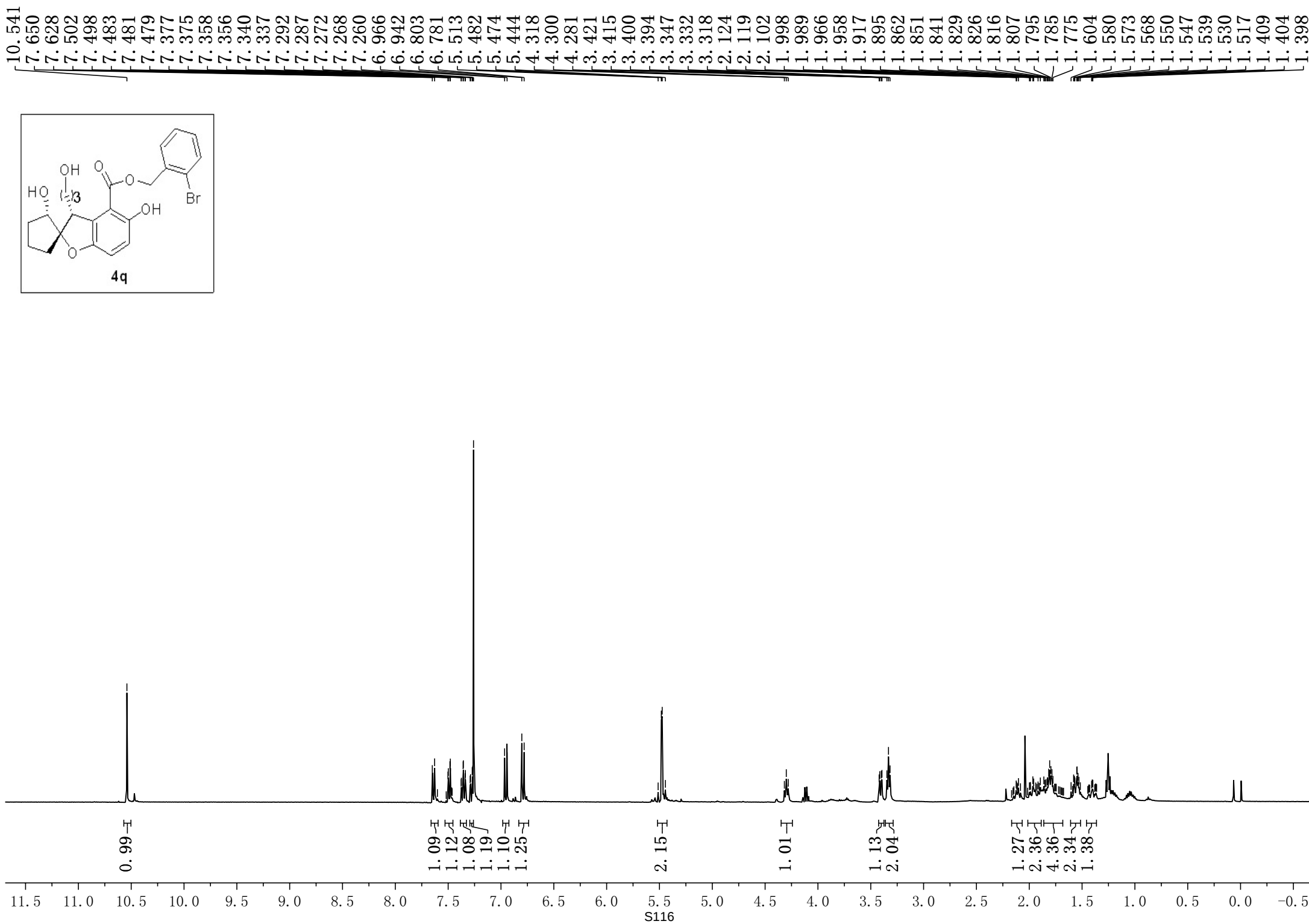
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single pulse decoupled gated NOESY



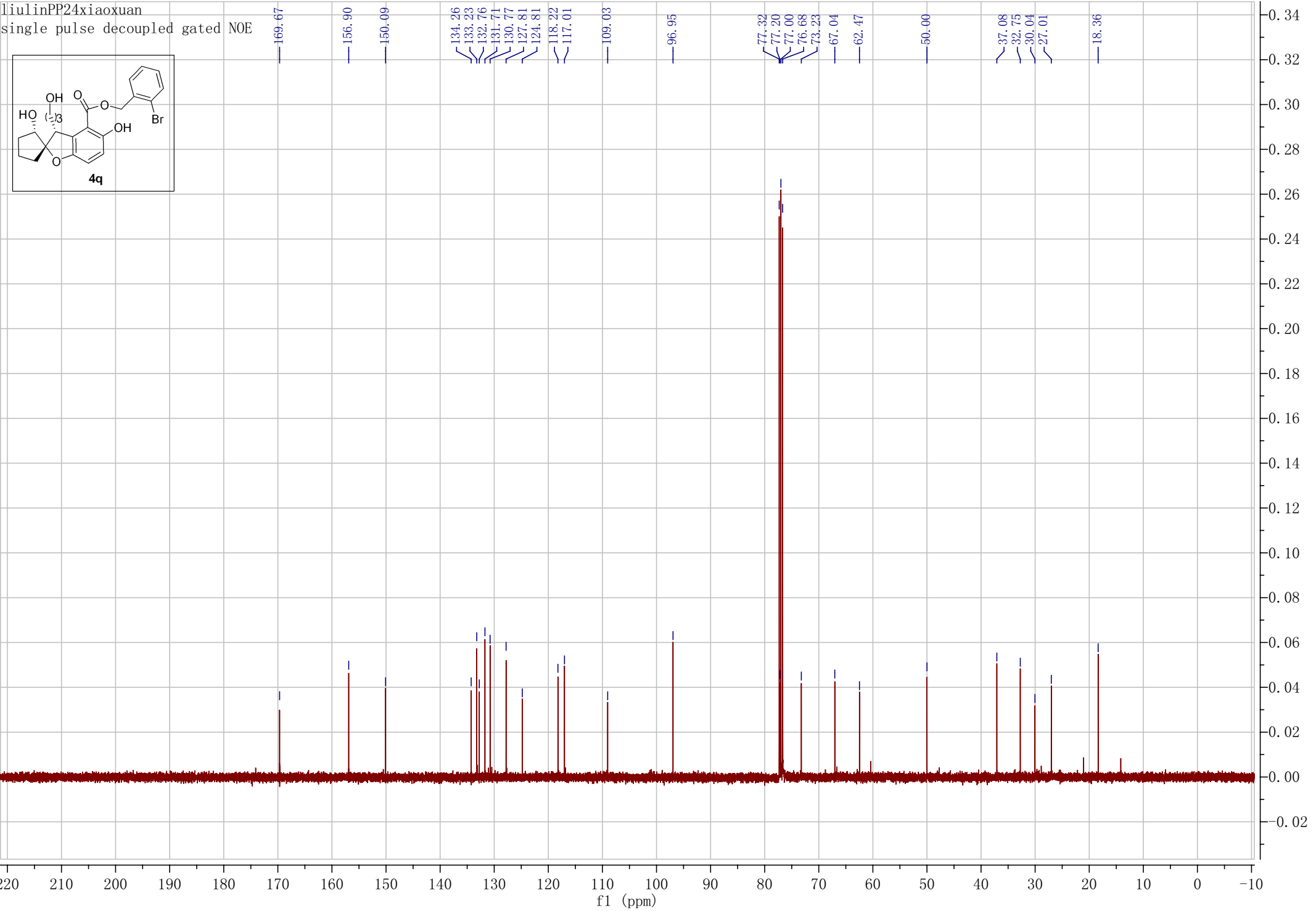
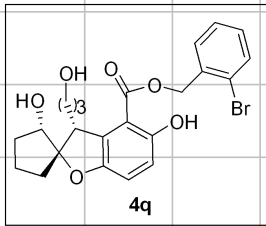
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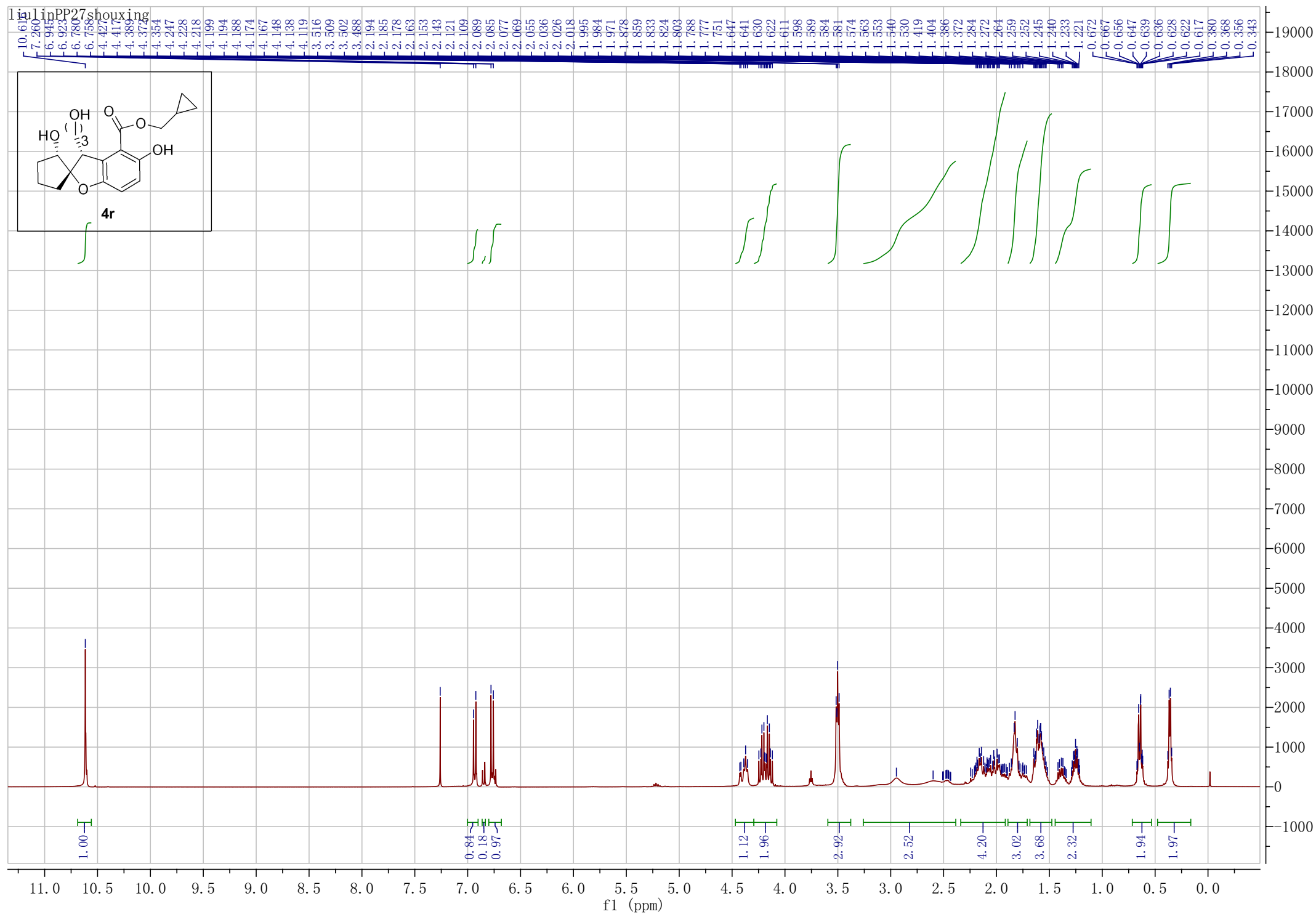




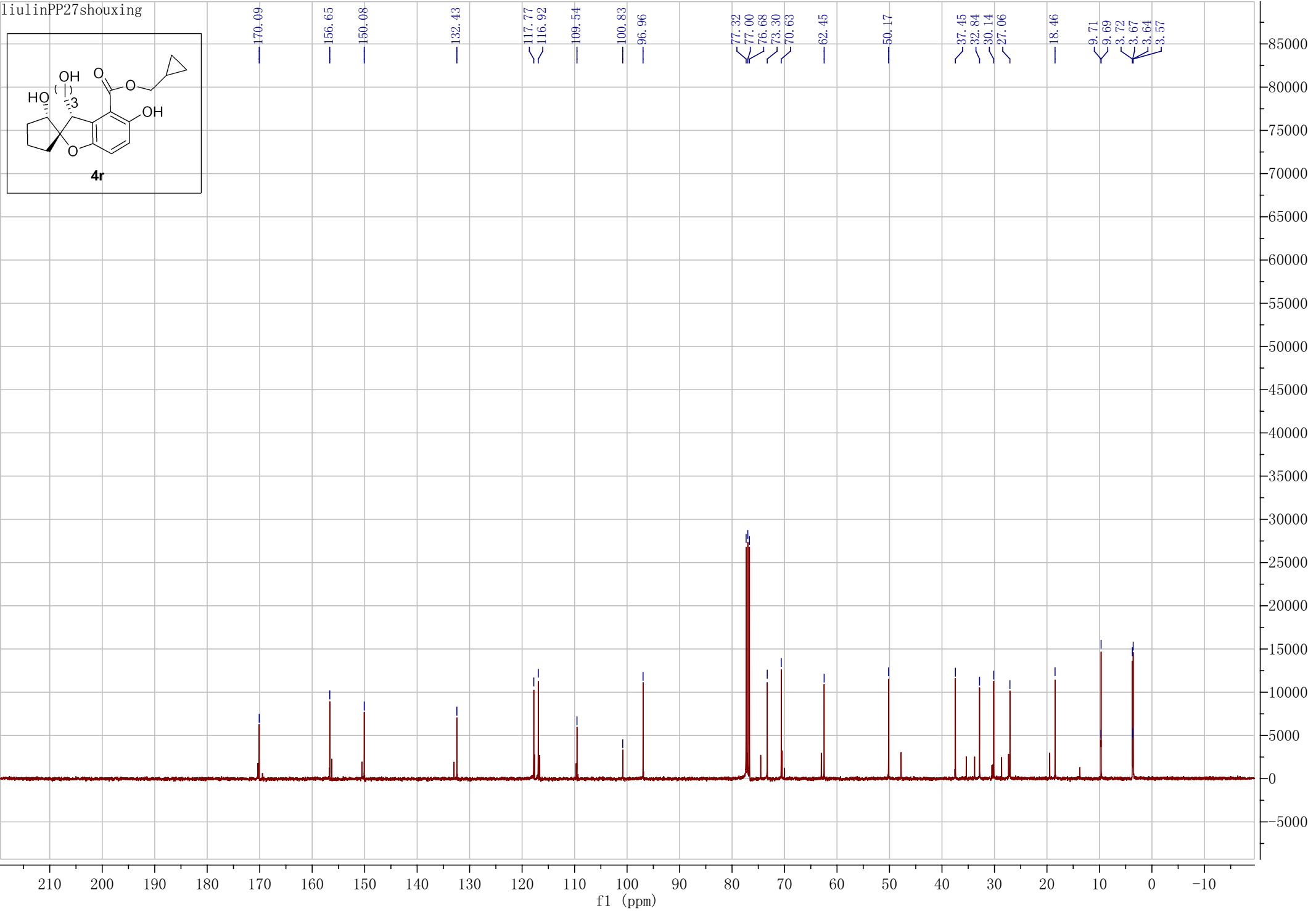
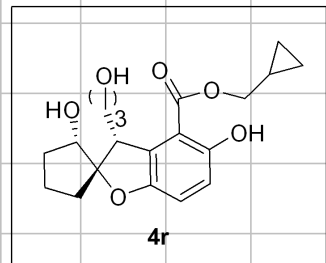
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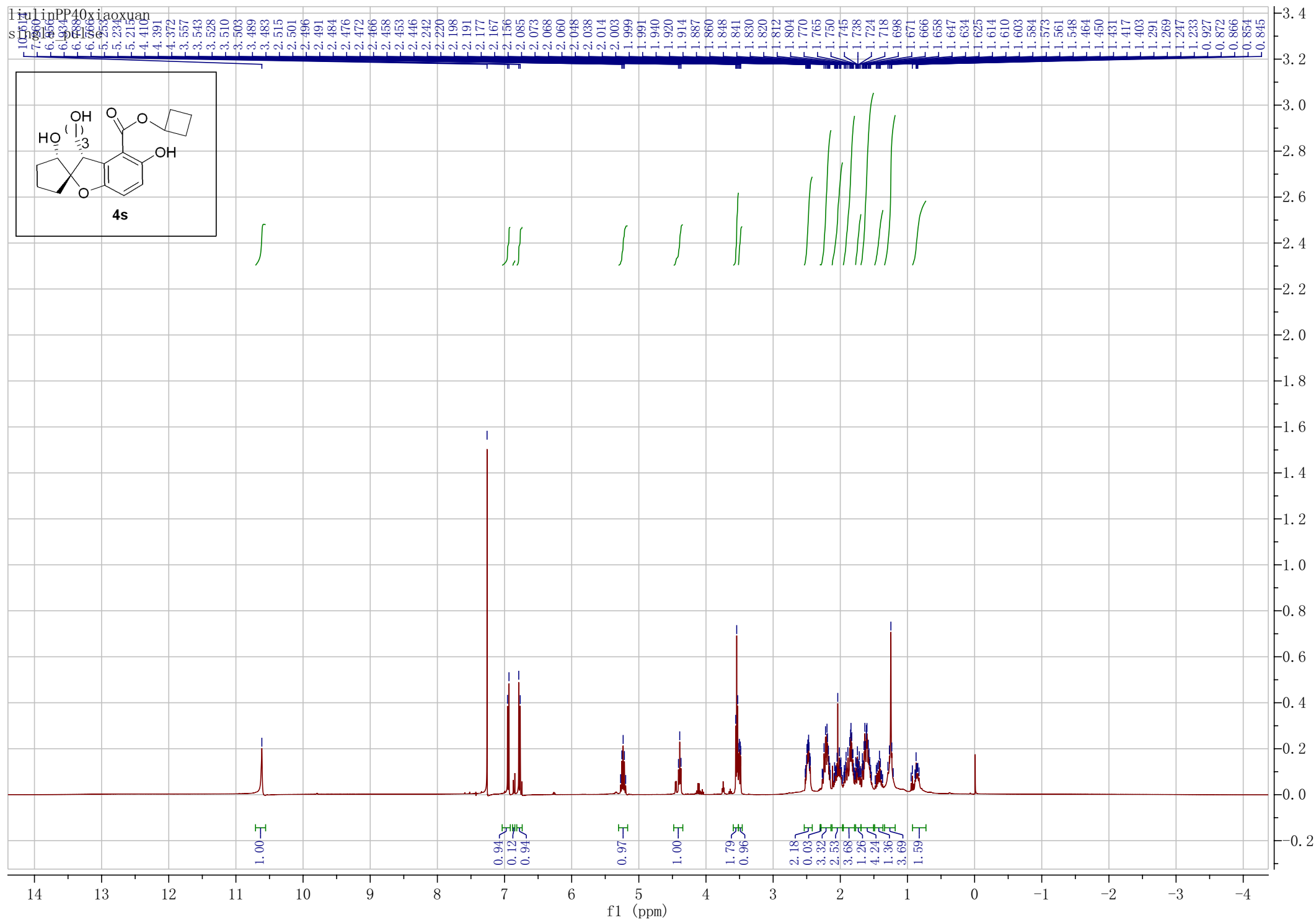
single pulse decoupled gated NOE





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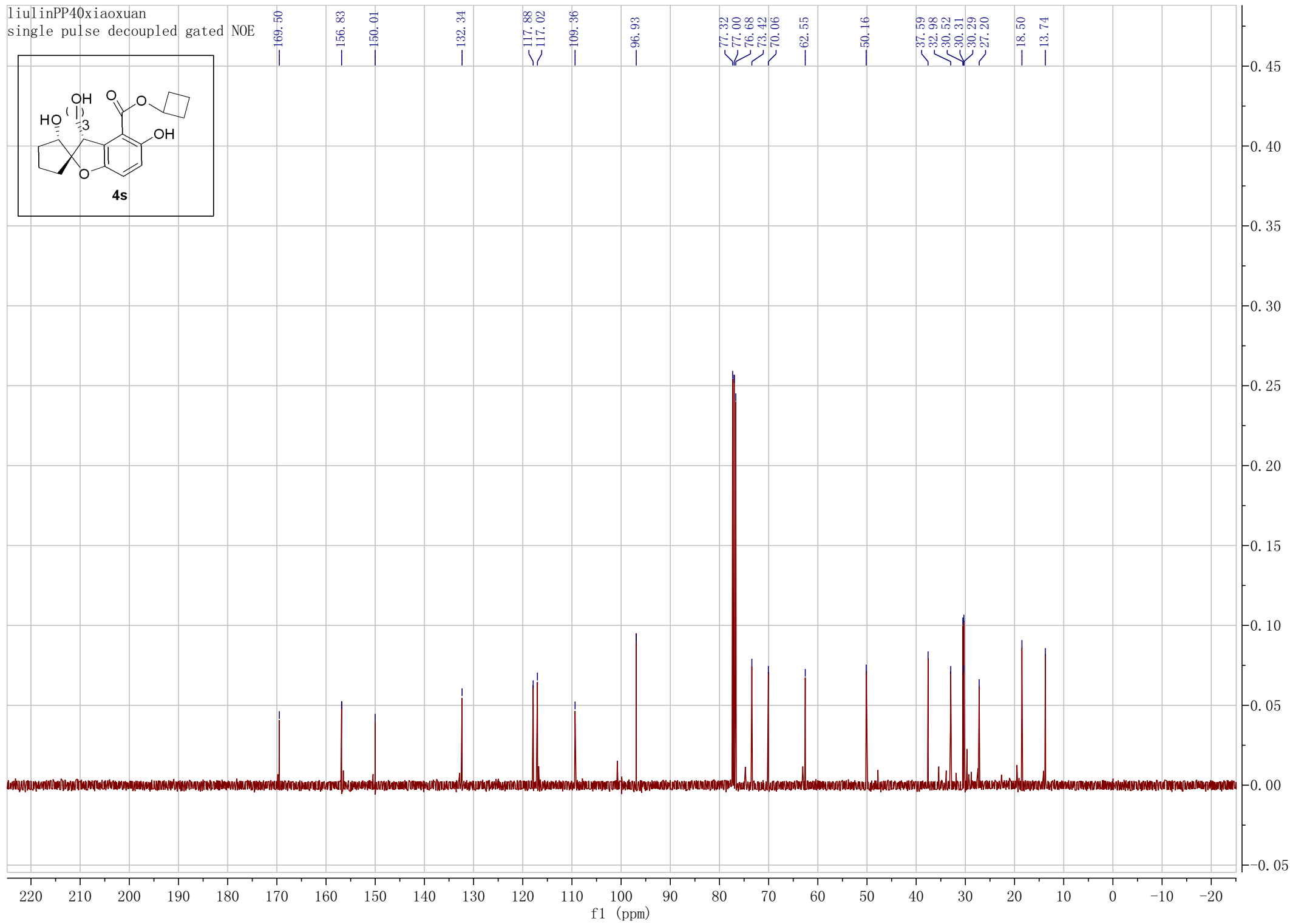
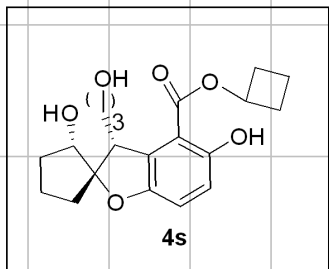




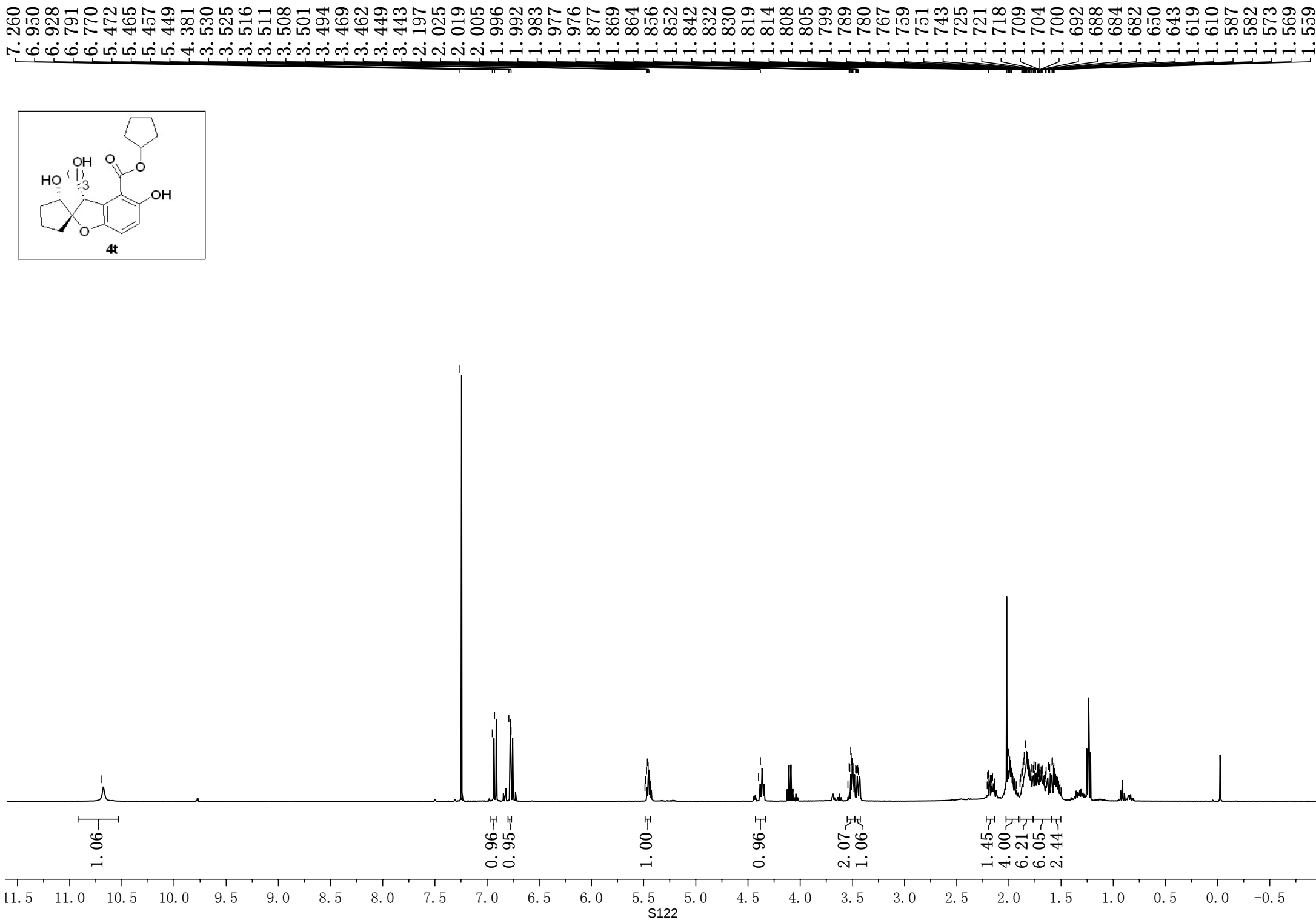
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single pulse decoupled gated NOE

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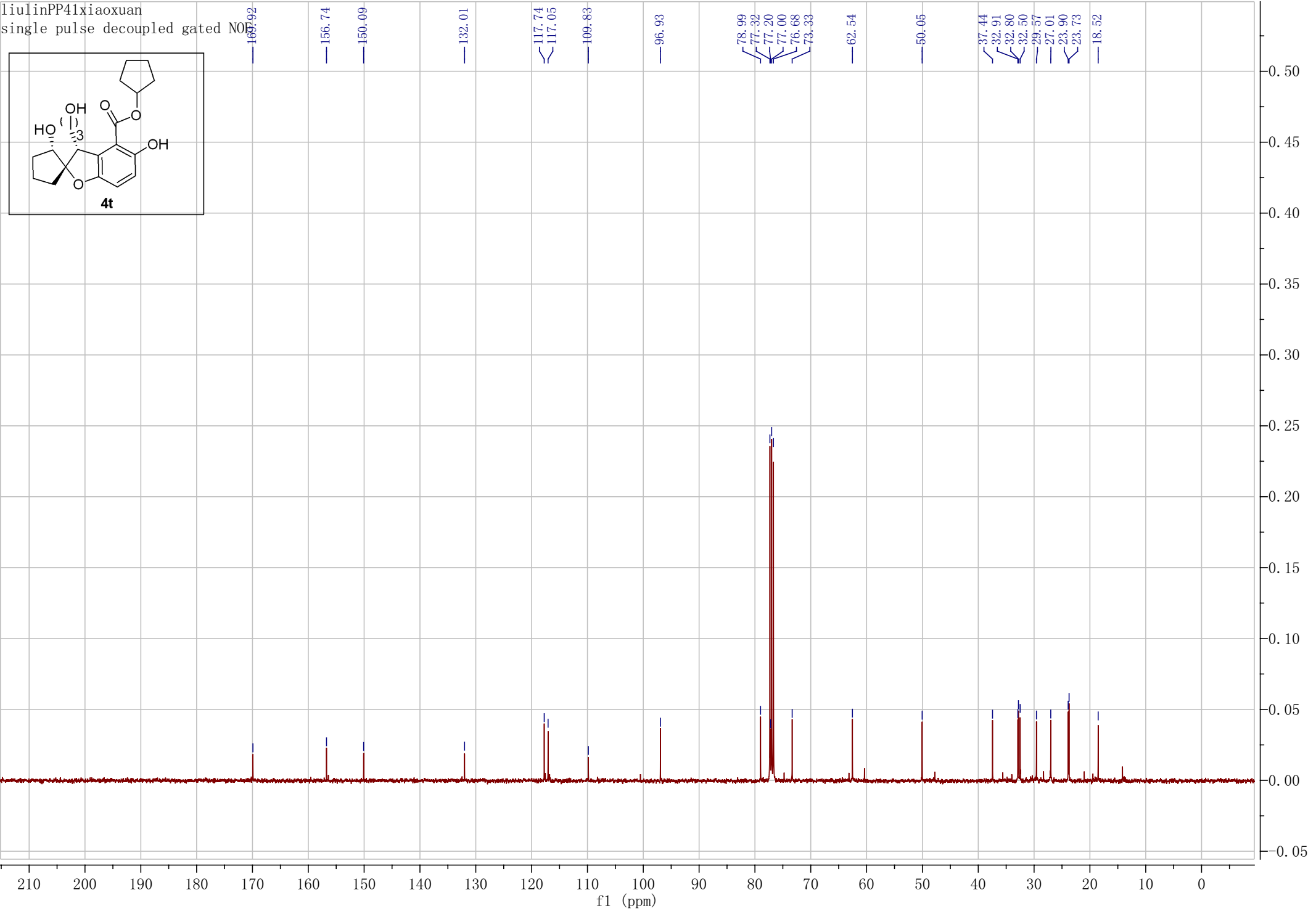
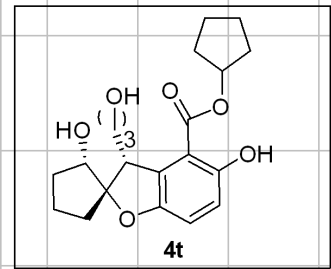


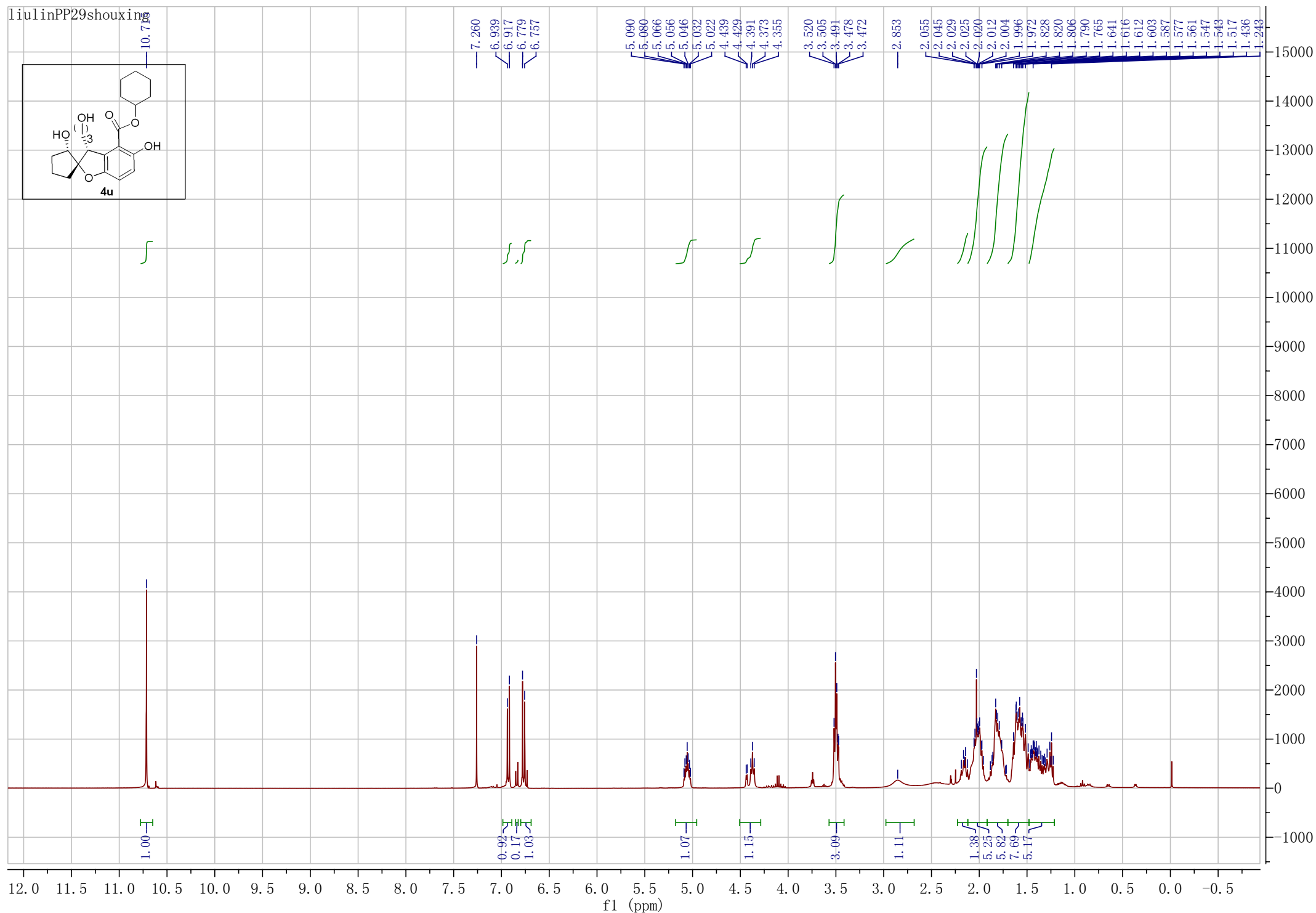
S121



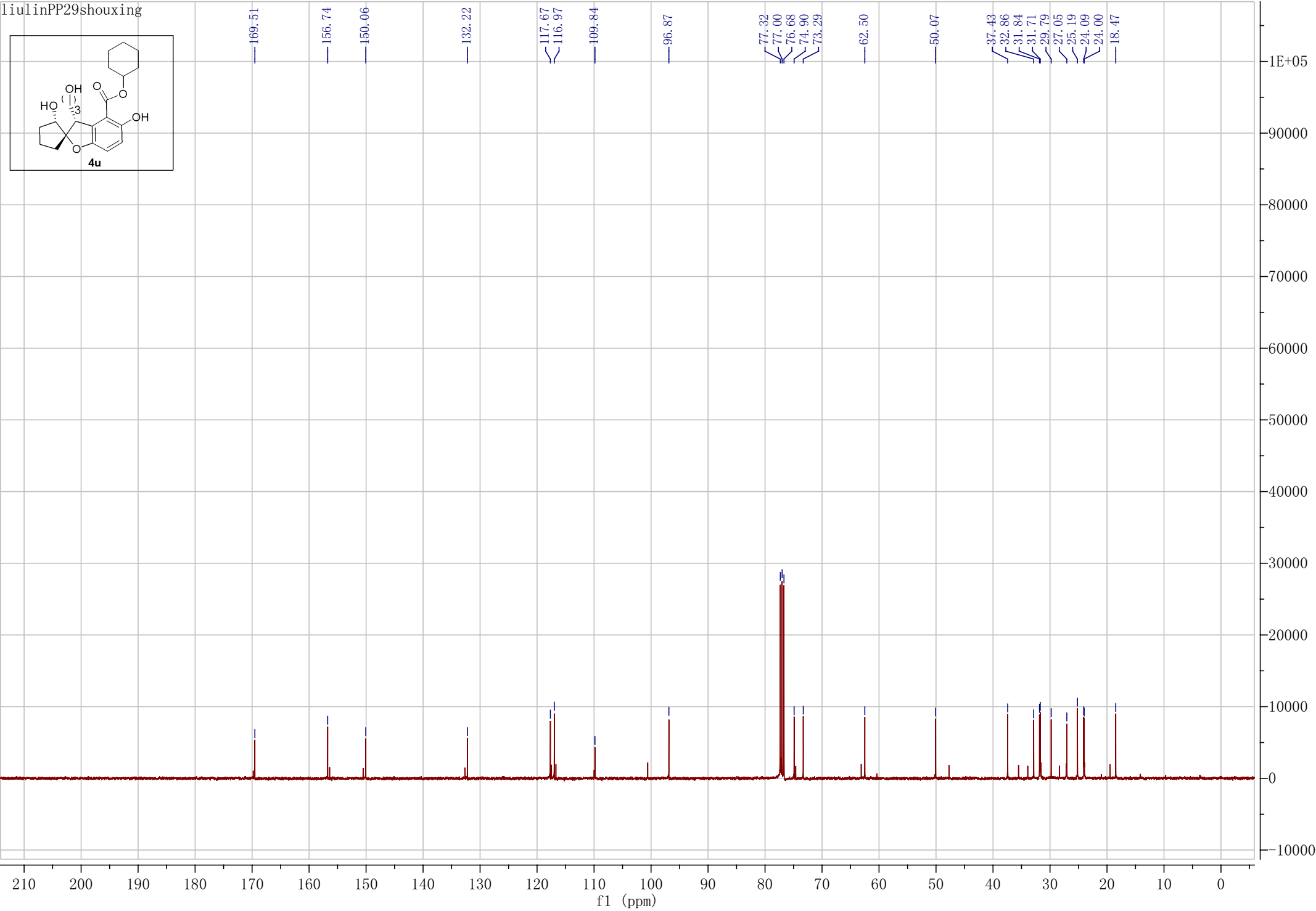
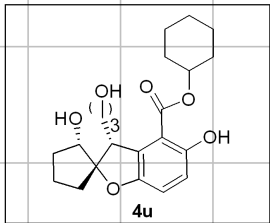
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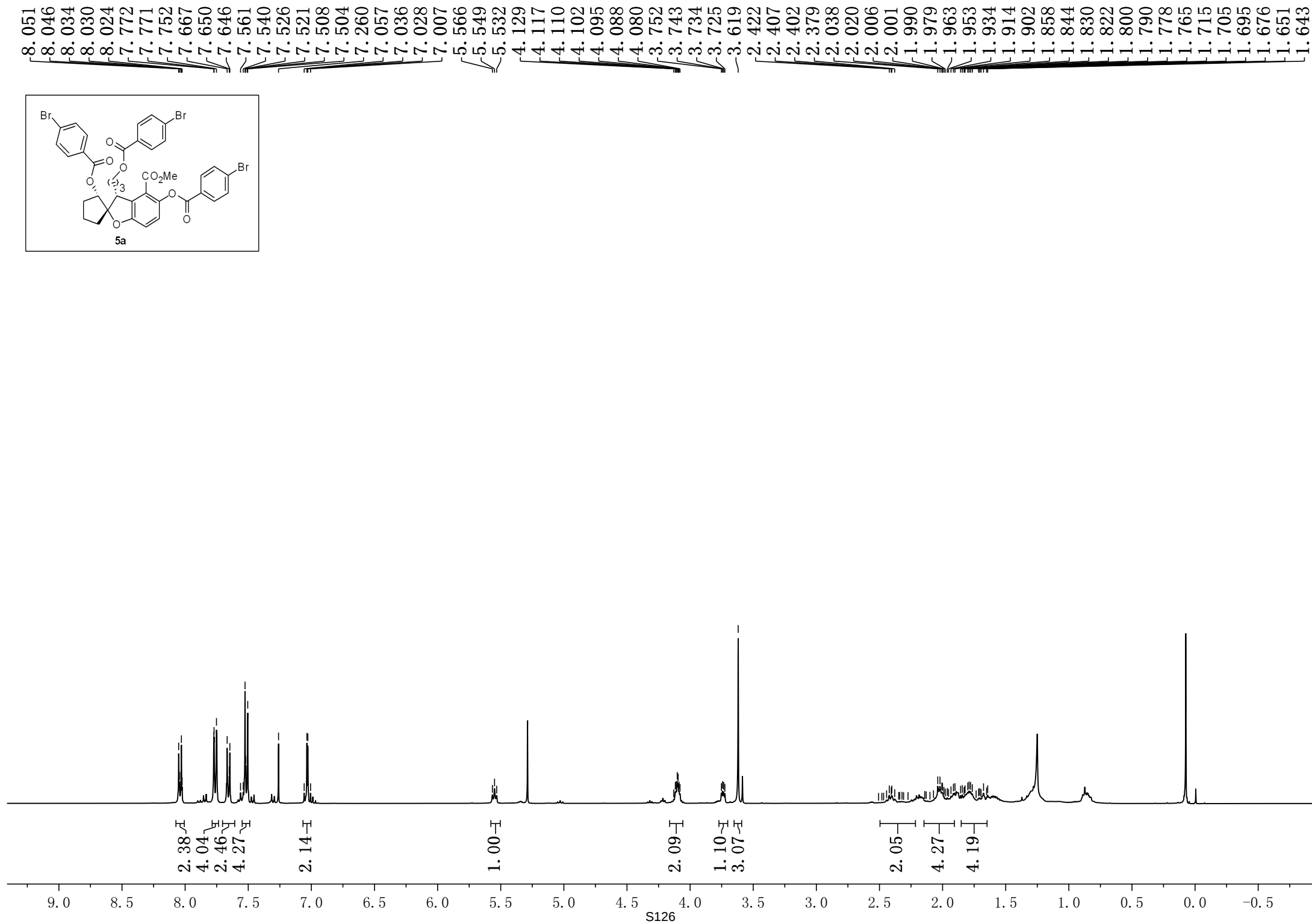
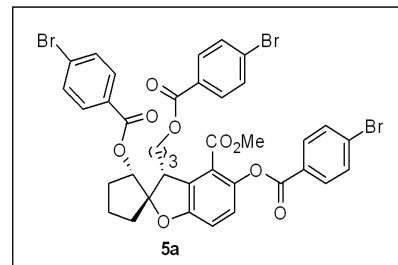
single pulse decoupled gated NOE



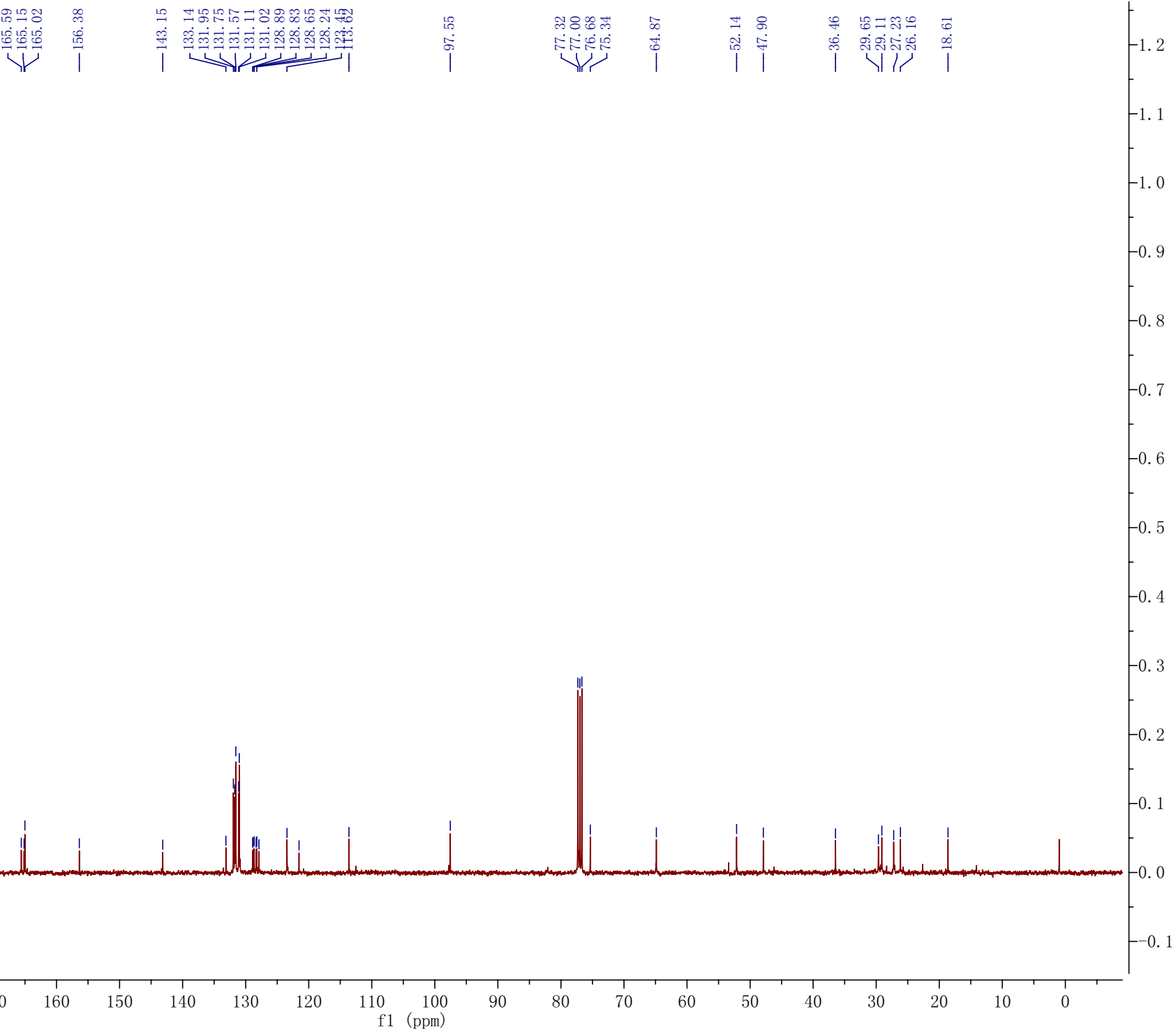
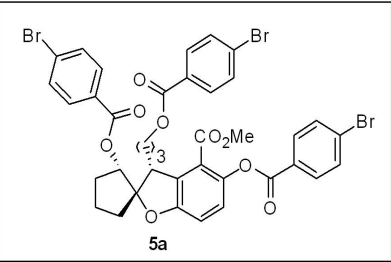


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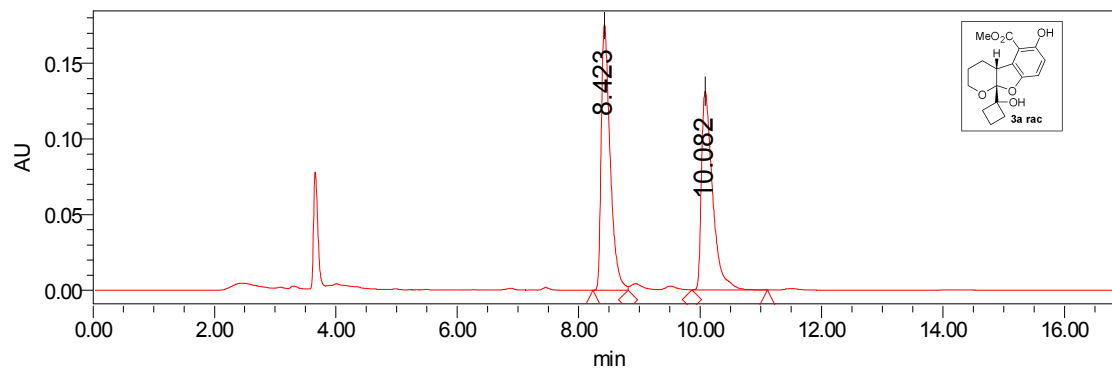


LIULINSHANGXIU
single pulse decoupled gated NOE



Sample Name: 3a-rac
Column: 10.00 μ l

Wave Length: PDA 254 nm



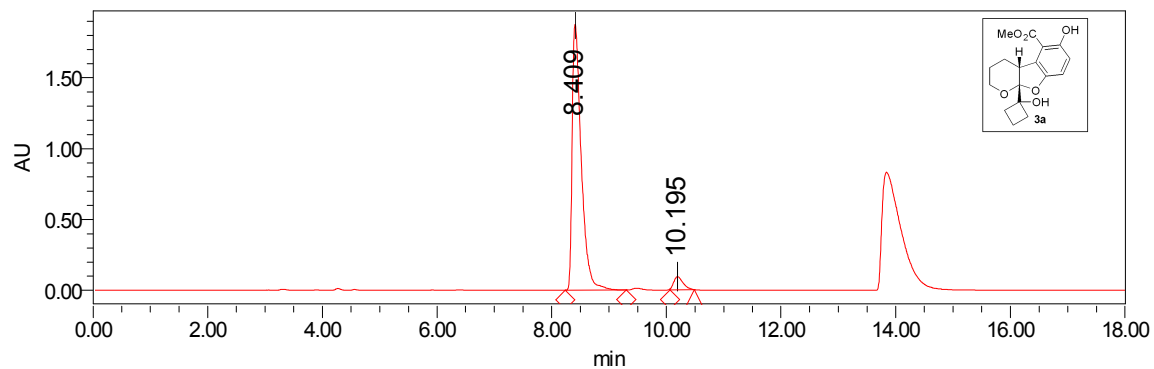
Peak Information:

	RetTime (min)	Area (μ V*S)	Area (%)	Height (μ V)
1	8.423	1792840.003	51.738	175776
2	10.082	1672395.158	48.262	131378

Sample Information

Sample Name: 3a
Column: 10.00 μ l

Wave Length: PDA 254 nm

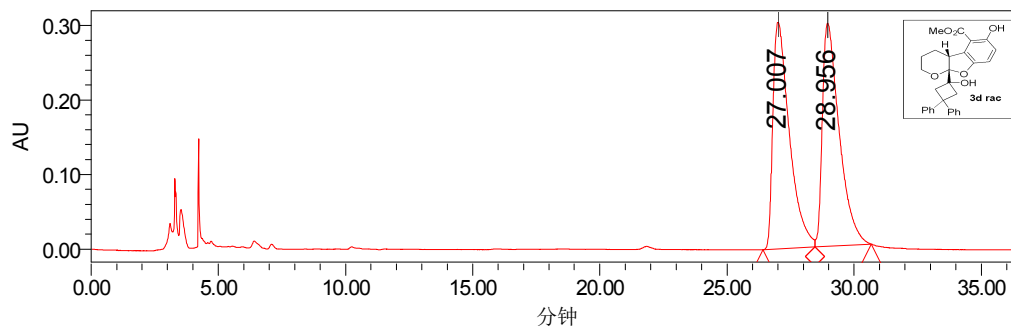


Peak Information:

	RetTime (min)	Area (μ V*S)	Area (%)	Height (μ V)
1	8.409	20647135.691	95.253	1877227
2	10.195	1029046.778	4.747	92423

Sample Name: 3d-rac
Column: 10.00 μ l

Wave Length: PDA 254 nm



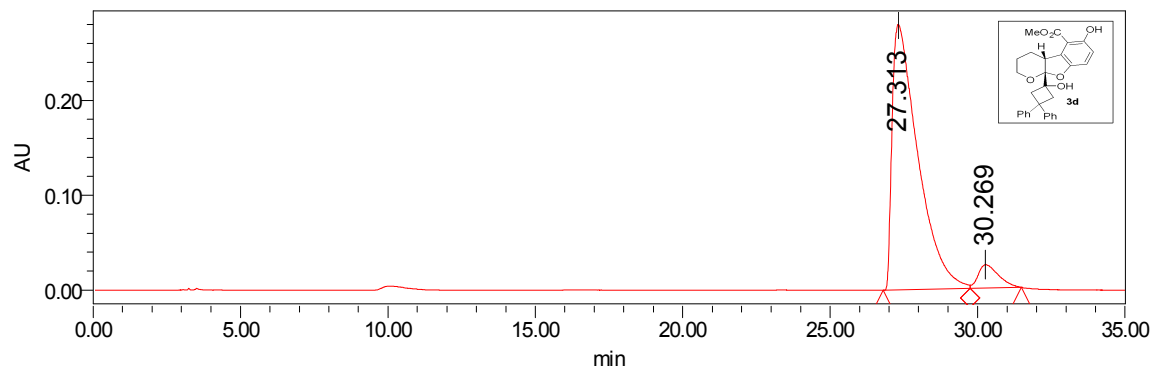
Peak Information:

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1	27.007	13780336.166	49.849	304233
2	28.956	13863744.802	50.151	299443

Sample Information

Sample Name: 3d
Column: 10.00 μ l

Wave Length: PDA 254 nm



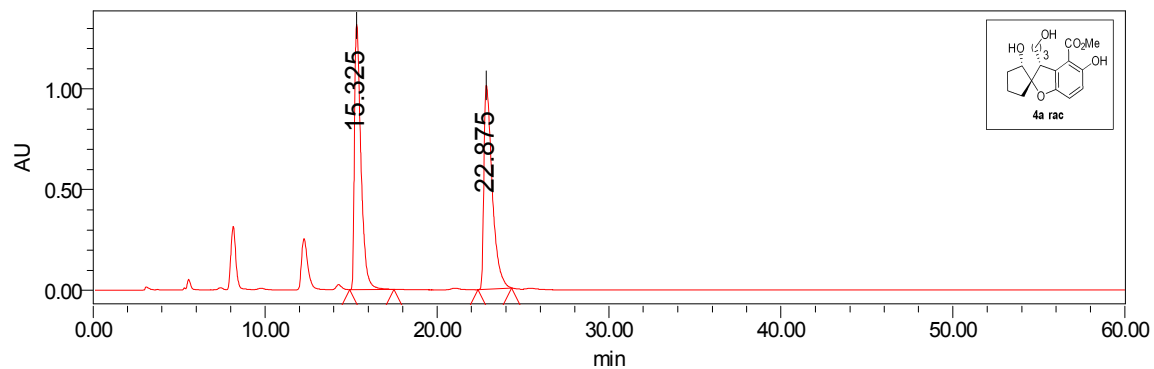
Peak Information:

	RetTime (min)	Area (μ V*S)	Area (%)	Height (μ V)
1	27.313	17021892.191	93.486	280036
2	30.269	1186013.144	6.514	24603

Sample Information

Sample Name: 4a rac
Column: 10.00 μ l

Wave Length: PDA 254 nm



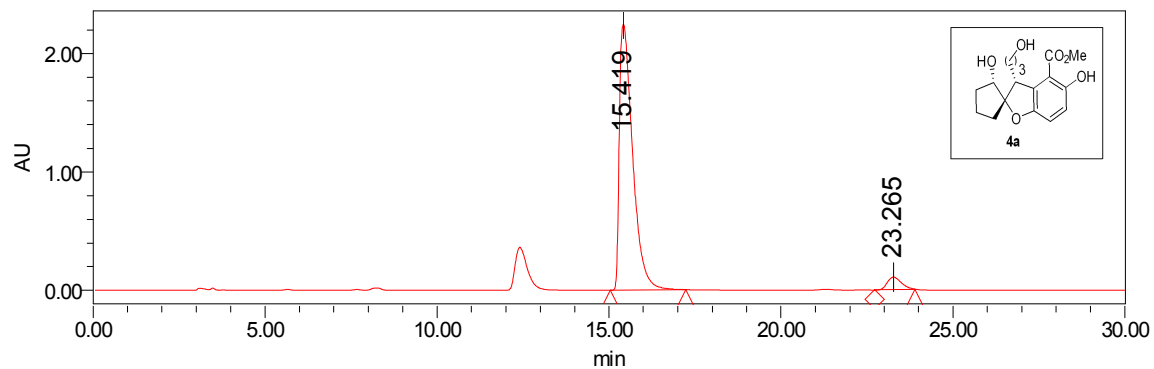
Peak Information:

	RetTime (min)	Area (μ V*S)	Area (%)	Height (μ V)
1	15.325	34076912.804	49.955	1316017
2	22.875	34138291.791	50.045	1011606

Sample Information

Sample Name: 4a
Column: 10.00 μ l

Wave Length: PDA 254 nm



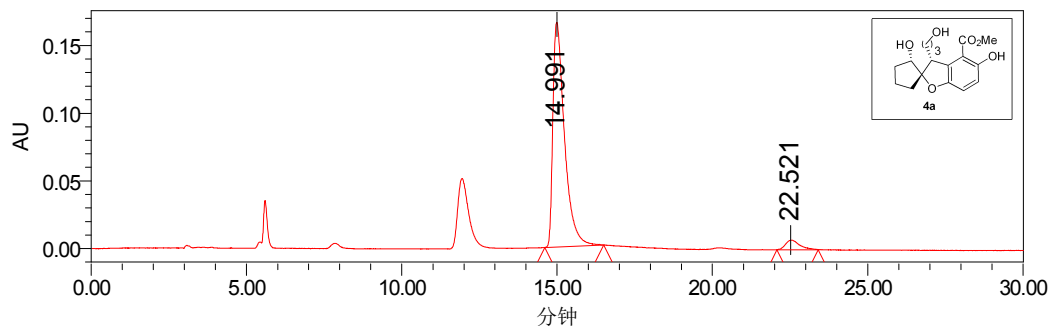
Peak Information:

	RetTime (min)	Area (μ V*S)	Area (%)	Height (μ V)
1	15.419	60225242.981	95.404	2245084
2	23.265	2901615.567	4.596	103391

2019/2/21 16:03:54 CST

Sample Name: 4a (1 mmol scale)
Column: 10.00 μ l

Wave Length: PDA 254 nm



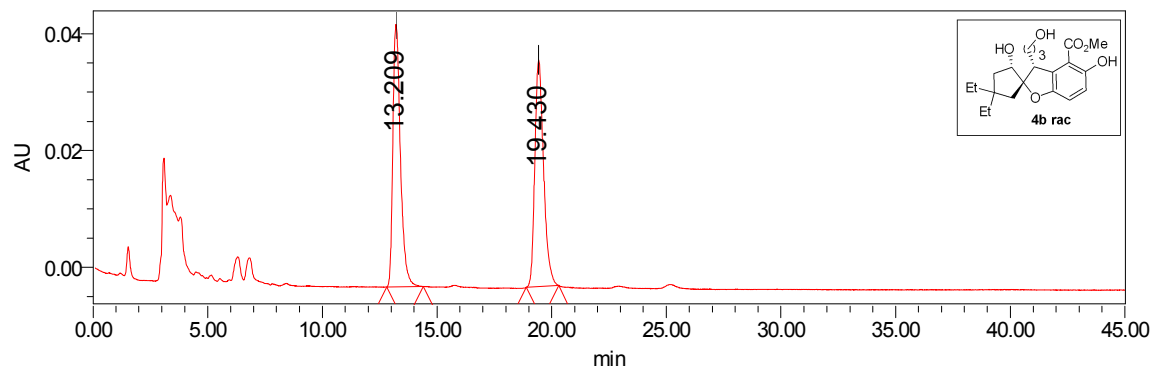
Peak Information:

	RetTime (min)	Area (μ V*S)	Area (%)	Height (μ V)
1	14.991	4312807.627	95.107	166151
2	22.521	221896.209	4.893	7162

Sample Information

Sample Name: 4b rac
Column: 10.00 μ l

Wave Length: PDA 254 nm

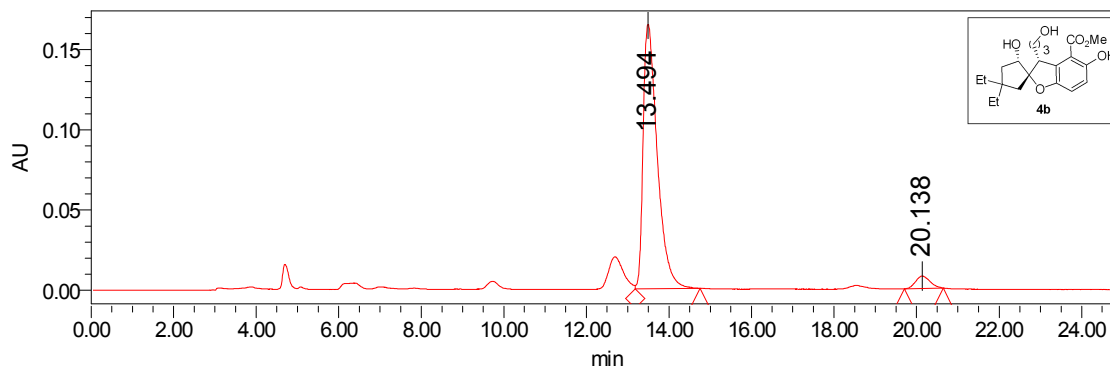


Peak Information:

	RetTime (min)	Area (μ V*S)	Area (%)	Height (μ V)
1	13.209	1029518.176	49.954	44988
2	19.430	1031422.731	50.046	38880

Sample Name: 4b
Column: 10.00 μ l

Wave Length: PDA 254 nm



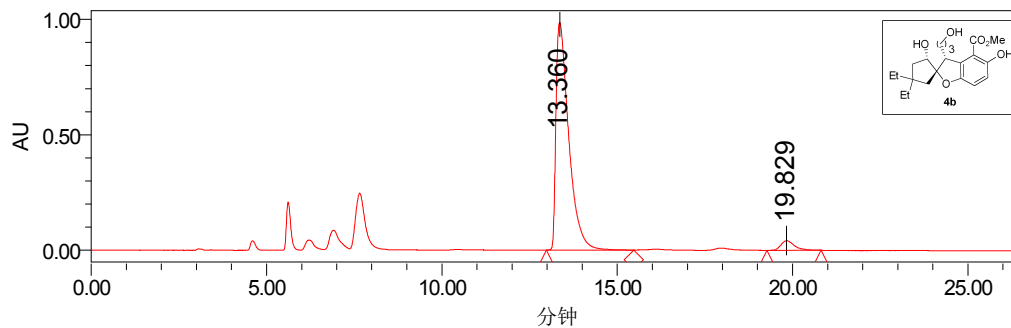
Peak Information:

	RetTime (min)	Area (μ V*S)	Area (%)	Height (μ V)
1	13.494	3906930.236	95.421	164704
2	20.138	187475.789	4.579	7689

2019/2/21 16:34:49 CST

Sample Name: 4b (1 mmol scale)
Column: 10.00 μ l

Wave Length: PDA 254 nm



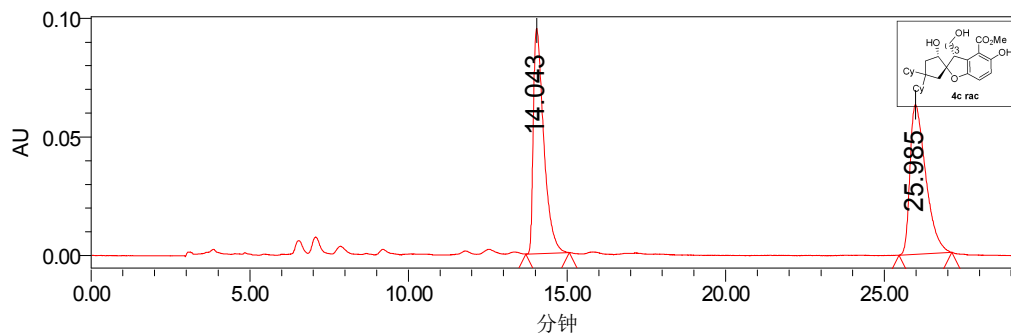
Peak Information:

	RetTime (min)	Area (μ V*S)	Area (%)	Height (μ V)
1	13.360	24877492.542	95.610	987389
2	19.829	1142344.137	4.390	42121

Sample Information

Sample Name: 4c rac
Column: 10.00 μ l

Wave Length: PDA 254 nm

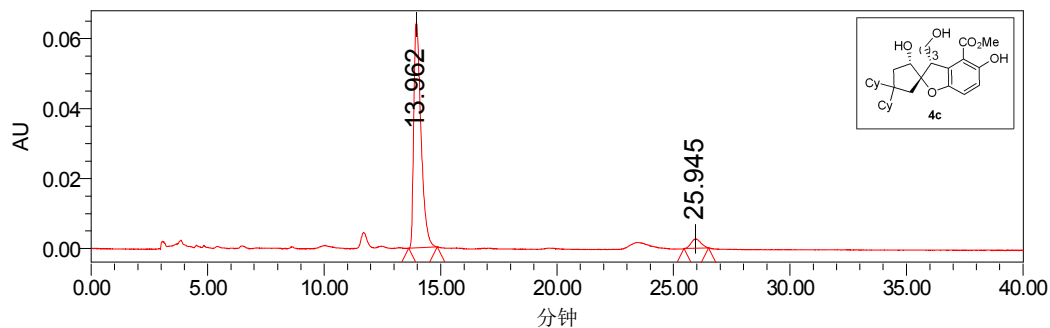


Peak Information:

	RetTime (min)	Area (μ V*S)	Area (%)	Height (μ V)
1	14.043	2179845.825	49.931	95136
2	25.985	2185858.346	50.069	63084

Sample Name: 4c
Column: 10.00 μ l

Wave Length: PDA 254 nm



Peak Information:

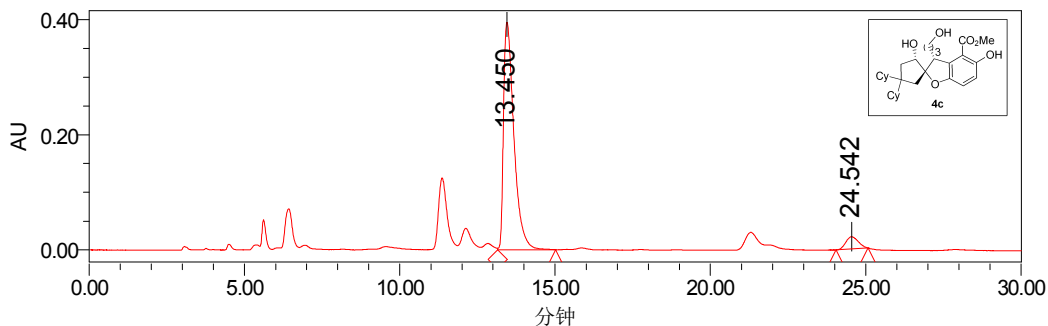
	RetTime (min)	Area (μ V*S)	Area (%)	Height (μ V)
1	13.962	1402808.198	94.650	64603
2	25.945	79292.152	5.350	2716

2019/2/21 17:02:06 CST

Sample Name: 4c (1 mmol scale)
Column: 10.00 ul

Wave Length:

PDA 254 nm



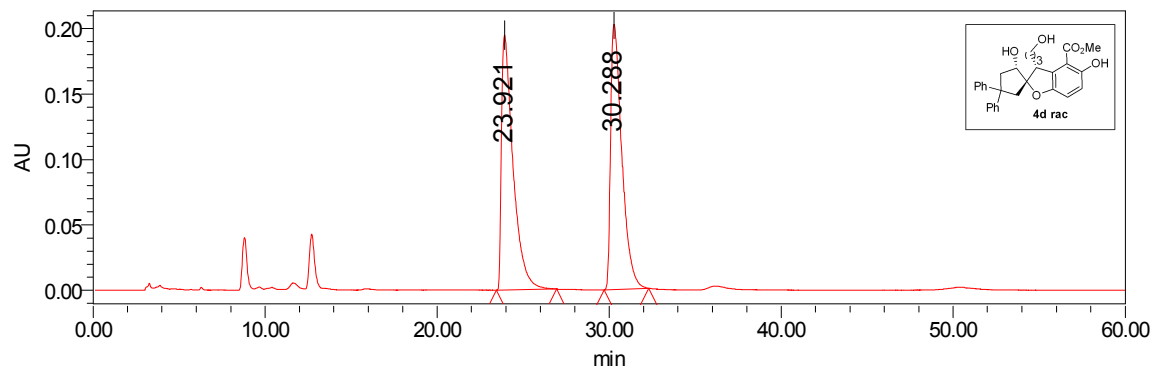
Peak Information:

	RetTime (min)	Area ($\mu\text{V}\cdot\text{S}$)	Area (%)	Height (μV)
1	13.450	9061835.833	93.851	395104
2	24.542	593763.960	6.149	21304

Sample Information

Sample Name: 4d rac
Column: 10.00 μ l

Wave Length: PDA 254 nm



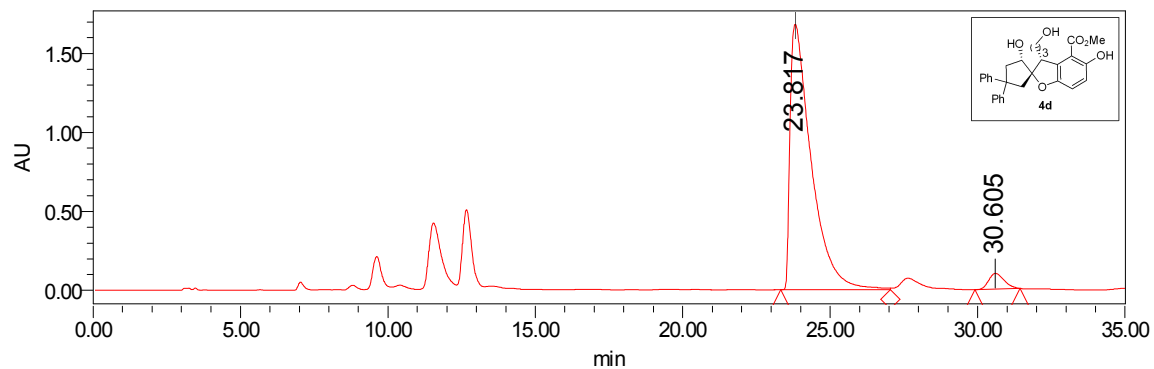
Peak Information:

	RetTime (min)	Area (μ V*S)	Area (%)	Height (μ V)
1	23.921	9125828.977	49.876	194818
2	30.288	9171253.028	50.124	202643

Sample Information

Sample Name: 4d
Column: 10.00 μ l

Wave Length: PDA 254 nm



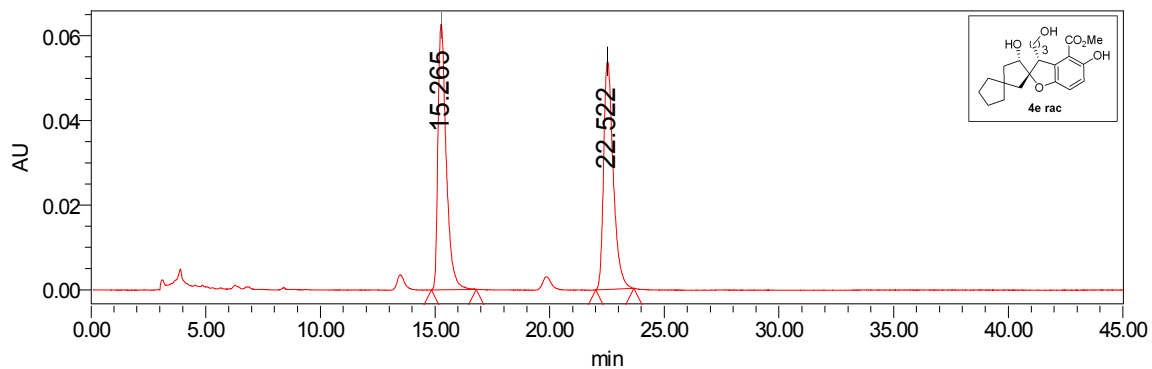
Peak Information:

	RetTime (min)	Area (μ V*S)	Area (%)	Height (μ V)
1	23.817	84027973.697	95.863	1681745
2	30.605	3626505.197	4.137	99298

Sample Information

Sample Name: 4e rac
Column: 10.00 μ l

Wave Length: PDA 254 nm



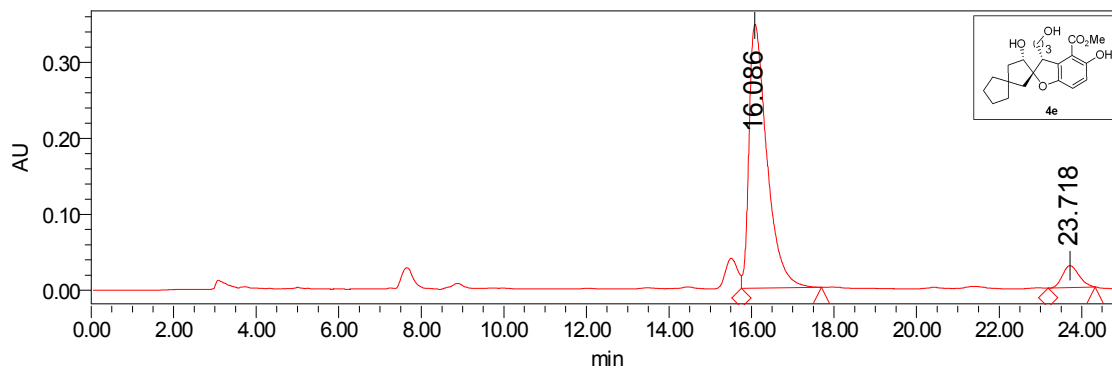
Peak Information:

	RetTime (min)	Area (μ V*S)	Area (%)	Height (μ V)
1	15.265	1571150.017	50.015	62681
2	22.522	1570222.247	49.985	53852

Sample Information

Sample Name: 4e
Column: 10.00 μ l

Wave Length: PDA 254 nm



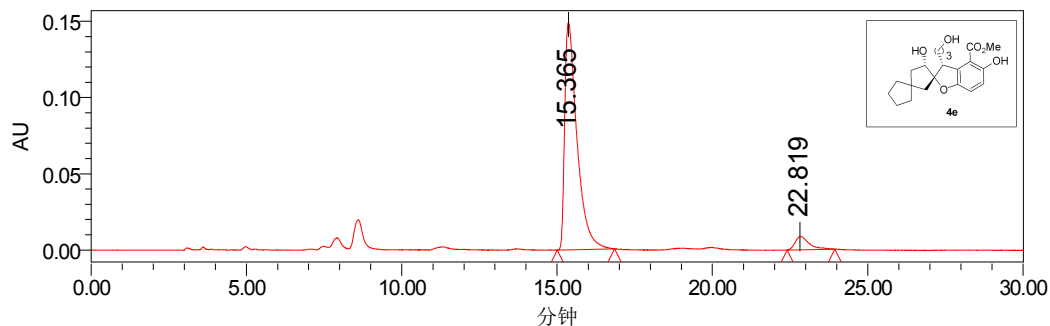
Peak Information:

	RetTime (min)	Area (μ V*S)	Area (%)	Height (μ V)
1	16.086	10432703.879	92.865	347459
2	23.718	801613.460	7.135	28908

2019/2/21 18:13:29 CST

Sample Name: 4e (1 mmol scale)
Column: 10.00 ul

Wave Length: PDA 254 nm

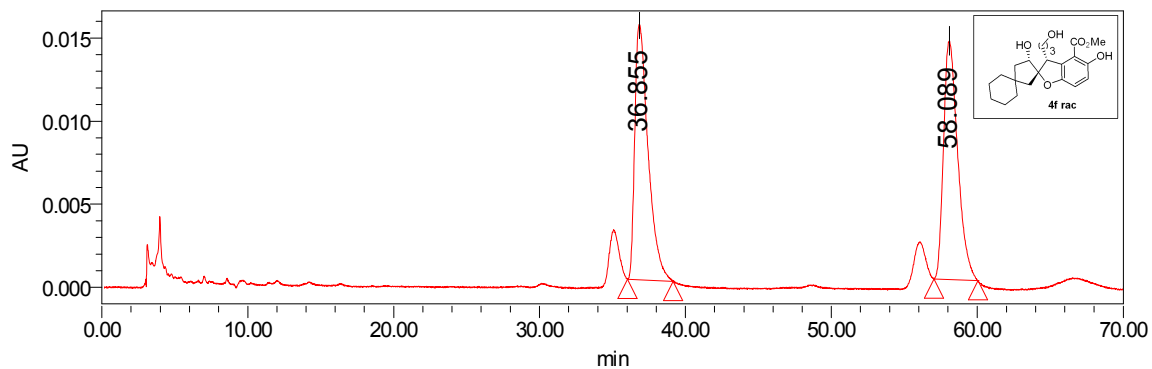


Peak Information:

	RetTime (min)	Area ($\mu\text{V}\cdot\text{S}$)	Area (%)	Height (μV)
1	15.365	4109921.339	93.635	149381
2	22.819	279360.275	6.365	8731

Sample Name: 4f rac
Column: 10.00 ul

Wave Length: PDA 254 nm

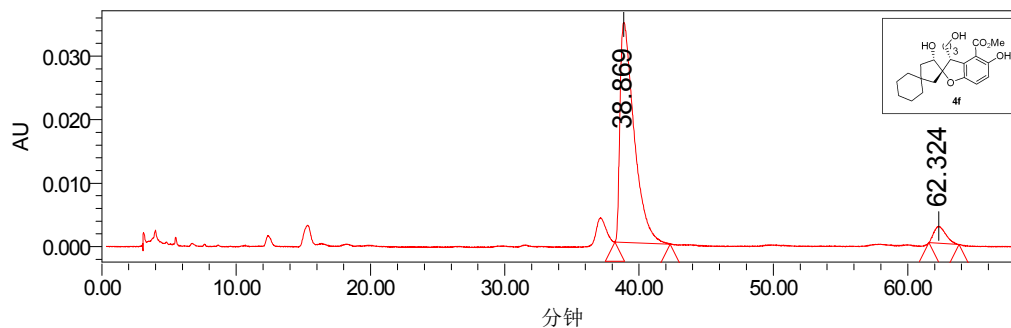


Peak Information:

	RetTime (min)	Area ($\mu\text{V}\cdot\text{S}$)	Area (%)	Height (μV)
1	36.855	977696.305	50.192	15381
2	58.089	970228.898	49.808	14366

Sample Name: 4f
Column: 10.00 ul

Wave Length: PDA 254 nm



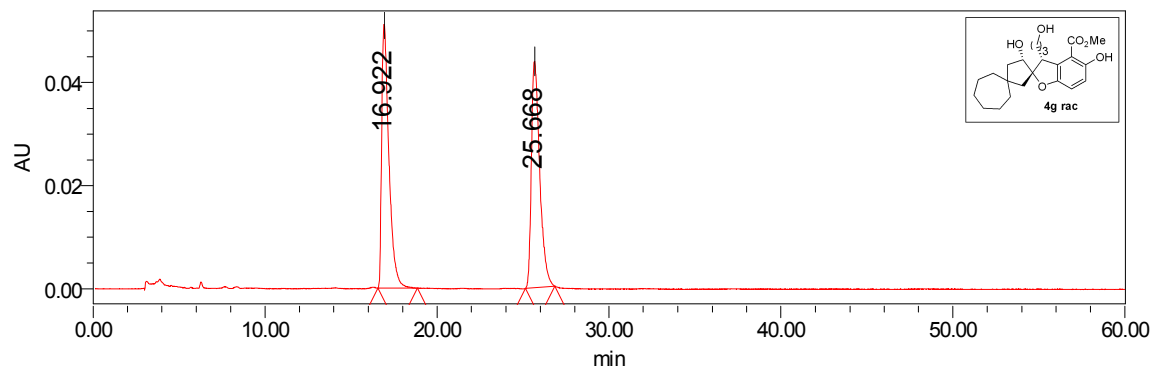
Peak Information:

	RetTime (min)	Area ($\mu\text{V}\cdot\text{S}$)	Area (%)	Height (μV)
1	38.869	2540269.039	93.894	34727
2	62.324	165194.234	6.106	2681

Sample Information

Sample Name: 4g rac
Column: 10.00 μ l

Wave Length: PDA 254 nm



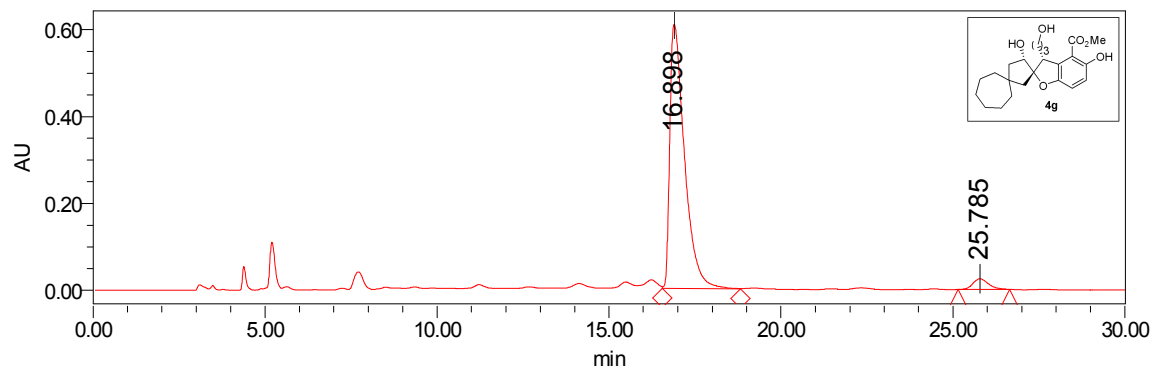
Peak Information:

	RetTime (min)	Area (μ V*S)	Area (%)	Height (μ V)
1	16.922	1479067.415	50.071	51131
2	25.668	1474864.437	49.929	43925

Sample Information

Sample Name: 4g
Column: 10.00 μ l

Wave Length: PDA 254 nm



Peak Information:

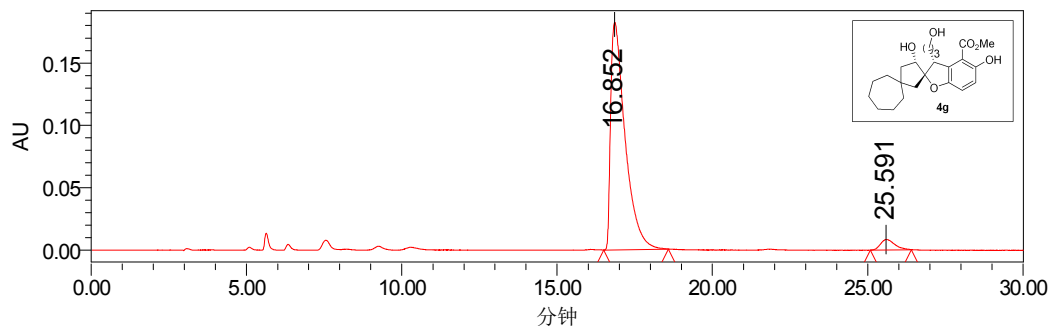
	RetTime (min)	Area (μ V*S)	Area (%)	Height (μ V)
1	16.898	18014229.865	95.917	607962
2	25.785	766763.260	4.083	24253

2019/2/21 18:44:11 CST

Sample Name: 4g (1 mmol scale)
Column: 10.00 ul

Wave Length:

PDA 254 nm

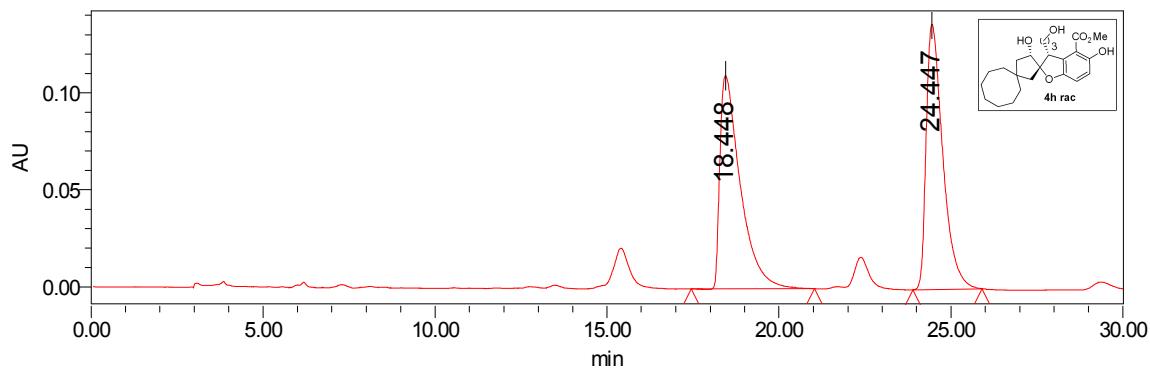


Peak Information:

	RetTime (min)	Area ($\mu\text{V}\cdot\text{S}$)	Area (%)	Height (μV)
1	16.852	5889494.928	95.615	182672
2	25.591	270120.267	4.385	8463

Sample Name: 4h rac
Column: 10.00 ul

Wave Length: PDA 254 nm



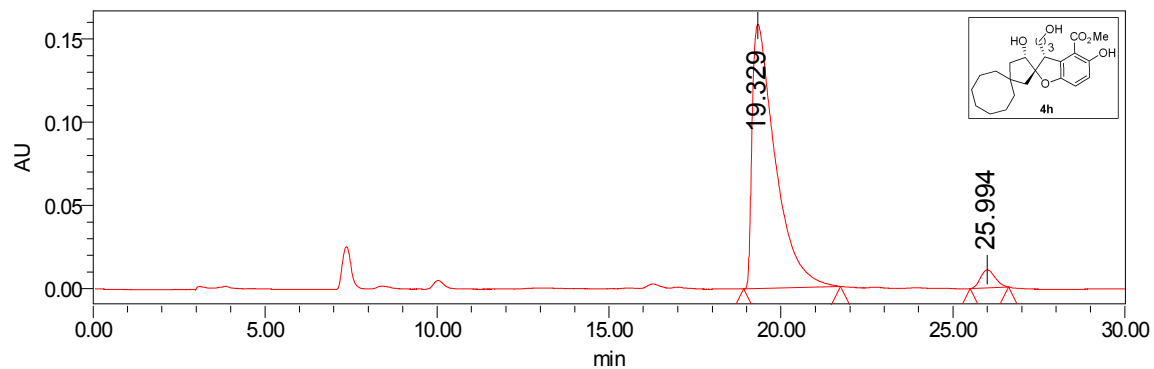
Peak Information:

	RetTime (min)	Area ($\mu\text{V}\cdot\text{S}$)	Area (%)	Height (μV)
1	18.448	4539503.975	49.772	109924
2	24.447	4581045.852	50.228	136754

Sample Information

Sample Name: 4h
Column: 10.00 μ l

Wave Length: PDA 254 nm



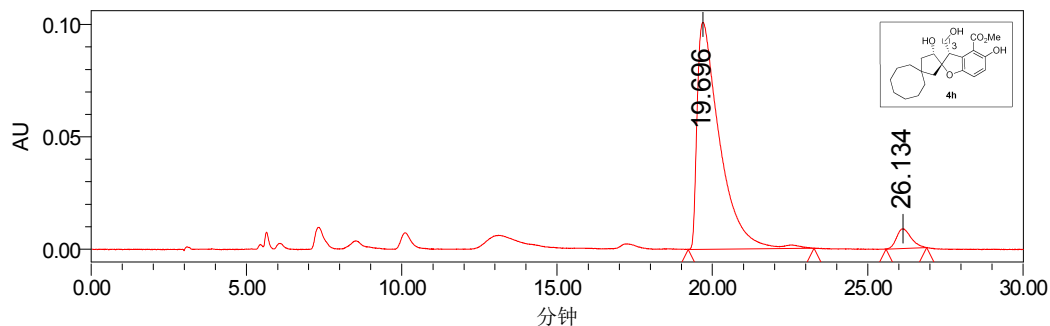
Peak Information:

	RetTime (min)	Area (μ V*S)	Area (%)	Height (μ V)
1	19.329	7395252.364	95.848	158741
2	25.994	320372.181	4.152	10845

2019/2/21 19:14:52 CST

Sample Name: 4h (1 mmol scale)
Column: 10.00 ul

Wave Length: PDA 254 nm



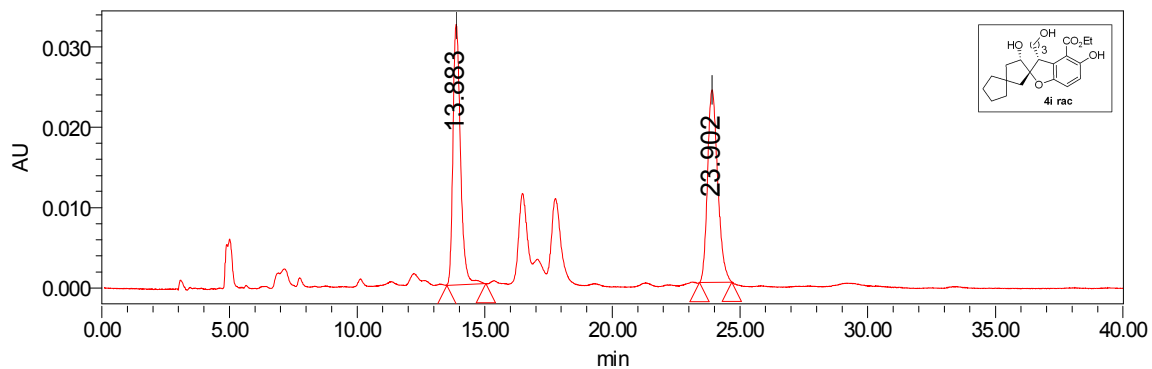
Peak Information:

	RetTime (min)	Area ($\mu\text{V}\cdot\text{S}$)	Area (%)	Height (μV)
1	19.696	5102531.106	94.761	101134
2	26.134	282101.096	5.239	8738

Sample Information

Sample Name: 4i rac
Column: 10.00 μ l

Wave Length: PDA 254 nm

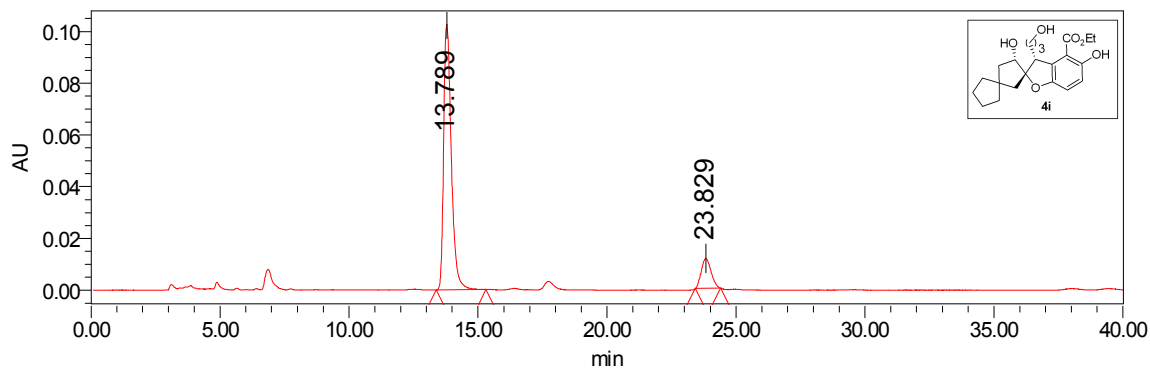


Peak Information:

	RetTime (min)	Area (μ V*S)	Area (%)	Height (μ V)
1	13.883	663309.241	49.830	32451
2	23.902	667828.803	50.170	23966

Sample Name: 4i
Column: 10.00 μ l

Wave Length: PDA 254 nm



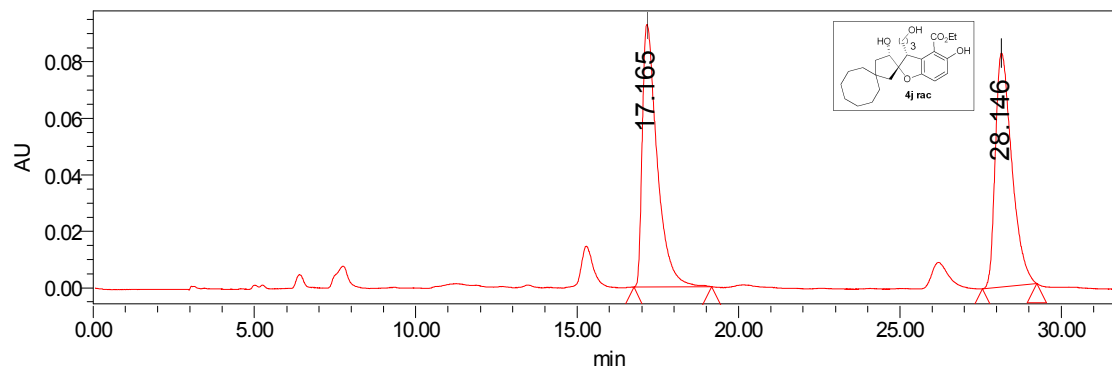
Peak Information:

	RetTime (min)	Area (μ V*S)	Area (%)	Height (μ V)
1	13.789	2114825.123	87.431	102605
2	23.829	304019.061	12.569	11486

Sample Information

Sample Name: 4j rac
Column: 10.00 μ l

Wave Length: PDA 254 nm



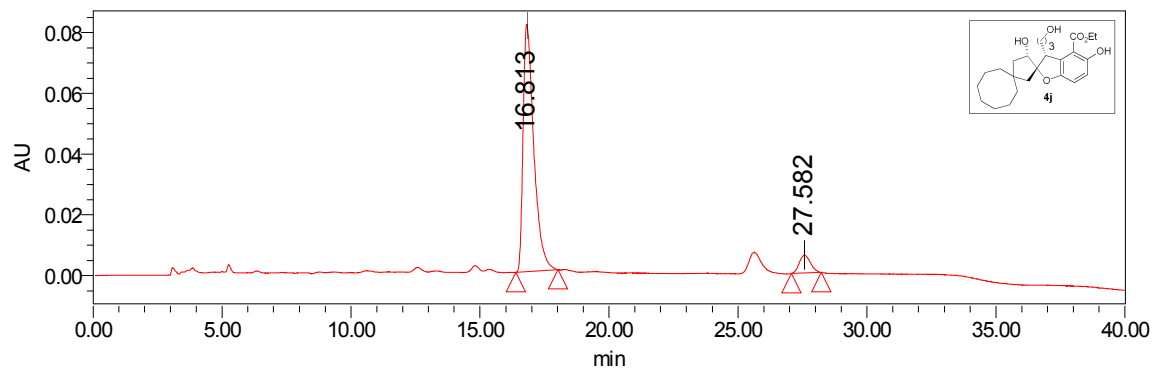
Peak Information:

	RetTime (min)	Area (μ V*S)	Area (%)	Height (μ V)
1	17.165	2967333.124	50.089	92985
2	28.146	2956799.620	49.911	82836

Sample Information

Sample Name: 4j
Column: 10.00 μ l

Wave Length: PDA 254 nm



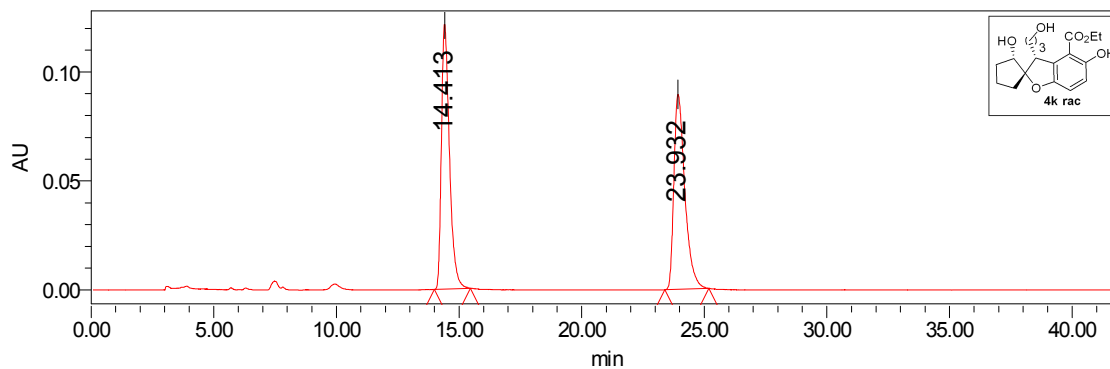
Peak Information:

	RetTime (min)	Area (μ V*S)	Area (%)	Height (μ V)
1	16.813	2349475.329	92.948	81506
2	27.582	178246.132	7.052	5926

Sample Information

Sample Name: 4k rac
Column: 10.00 ul

Wave Length: PDA 254 nm

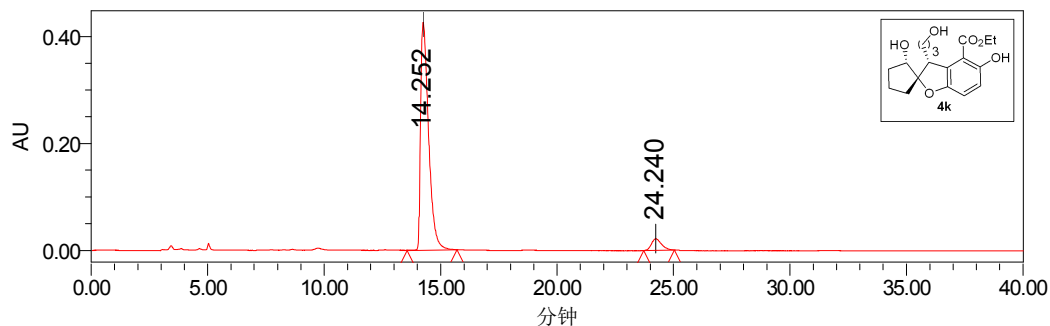


Peak Information:

	RetTime (min)	Area ($\mu\text{V}\cdot\text{S}$)	Area (%)	Height (μV)
1	14.413	2776633.743	50.021	121418
2	23.932	2774288.533	49.979	89423

Sample Name: 4k
Column: 10.00 ul

Wave Length: PDA 254 nm



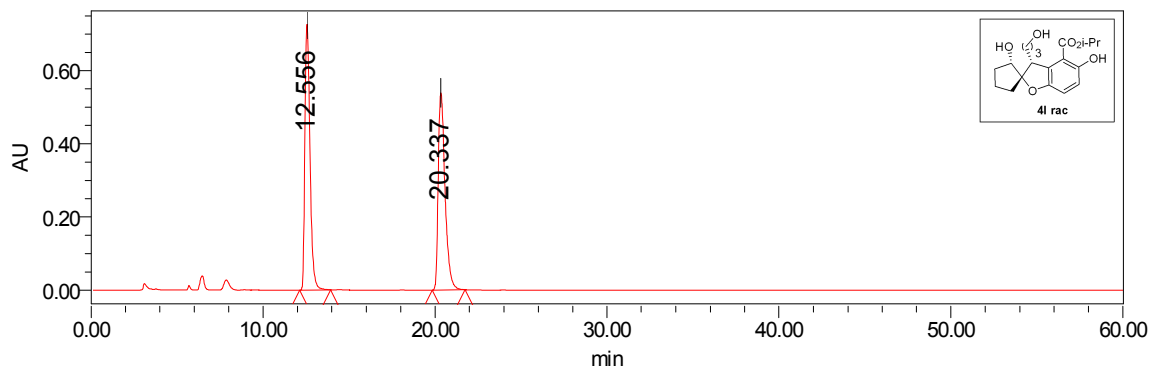
Peak Information:

	RetTime (min)	Area ($\mu\text{V}\cdot\text{S}$)	Area (%)	Height (μV)
1	14.252	9984056.972	93.808	426511
2	24.240	659068.739	6.192	21783

Sample Information

Sample Name: 4l rac
Column: 10.00 ul

Wave Length: PDA 254 nm



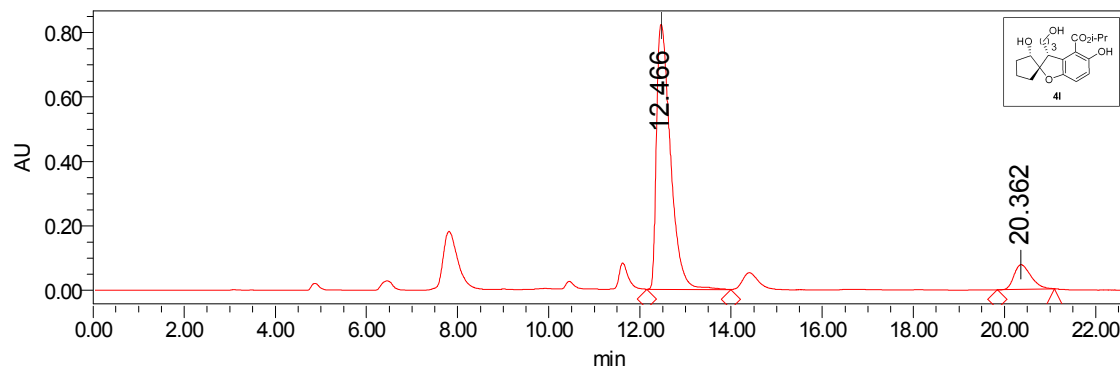
Peak Information:

	RetTime (min)	Area ($\mu\text{V}\cdot\text{S}$)	Area (%)	Height (μV)
1	12.556	14834610.284	50.083	726490
2	20.337	14785611.804	49.917	538905

Sample Information

Sample Name: 4l
Column: 10.00 μ l

Wave Length: PDA 254 nm



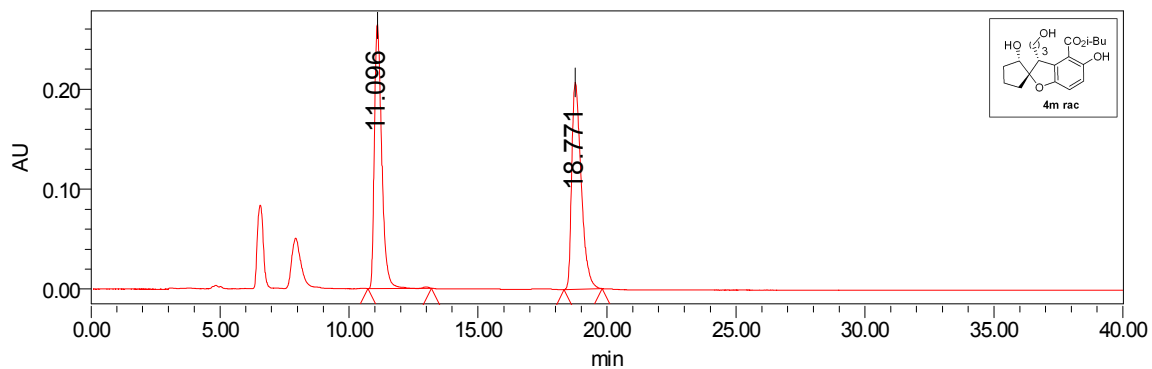
Peak Information:

	RetTime (min)	Area (μ V*S)	Area (%)	Height (μ V)
1	12.466	17455492.174	89.833	822711
2	20.362	1975451.642	10.167	76637

Sample Information

Sample Name: 4m rac
Column: 10.00 μ l

Wave Length: PDA 254 nm



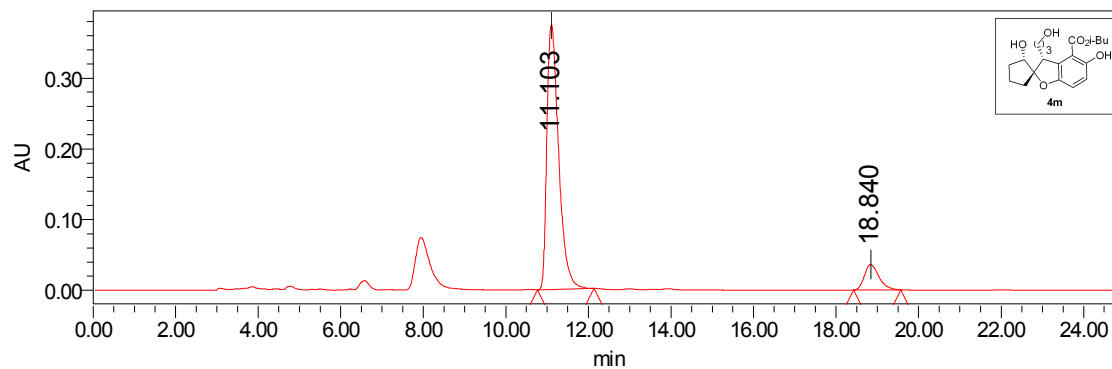
Peak Information:

	RetTime (min)	Area (μ V*S)	Area (%)	Height (μ V)
1	11.096	5227057.791	49.899	264259
2	18.771	5248209.155	50.101	206546

Sample Information

Sample Name: 4m
Column: 10.00 μ l

Wave Length: PDA 254 nm



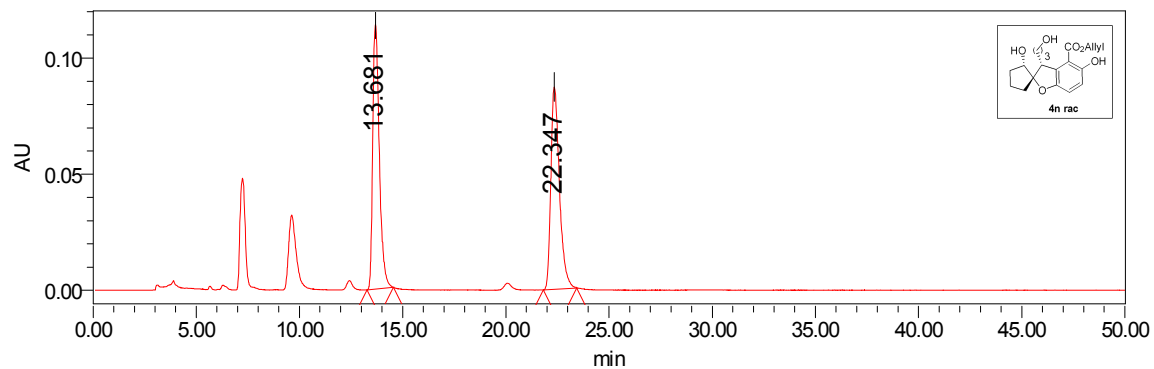
Peak Information:

	RetTime (min)	Area (μ V*S)	Area (%)	Height (μ V)
1	11.103	7403126.473	89.349	374850
2	18.840	882510.661	10.651	36001

Sample Information

Sample Name: 4n rac
Column: 10.00 μ l

Wave Length: PDA 254 nm



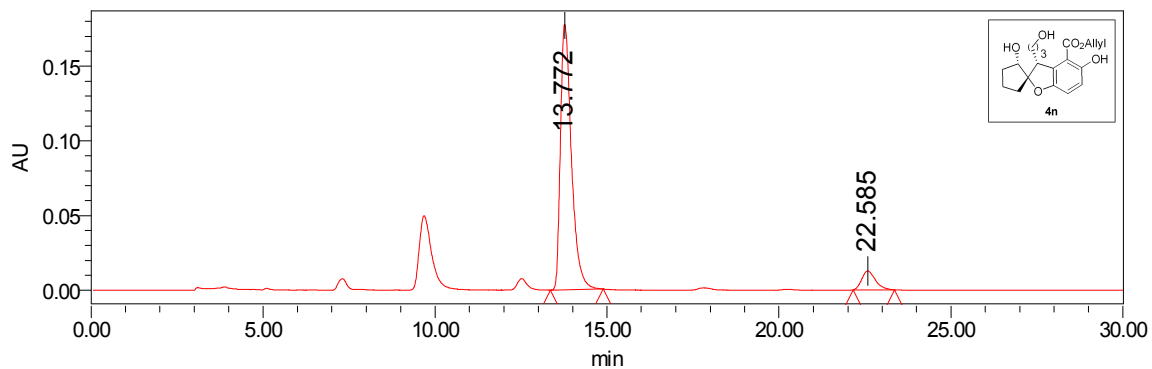
Peak Information:

	RetTime (min)	Area (μ V*S)	Area (%)	Height (μ V)
1	13.681	2519354.239	49.995	114075
2	22.347	2519892.671	50.005	87156

Sample Information

Sample Name: 4n
Column: 10.00 μ l

Wave Length: PDA 254 nm



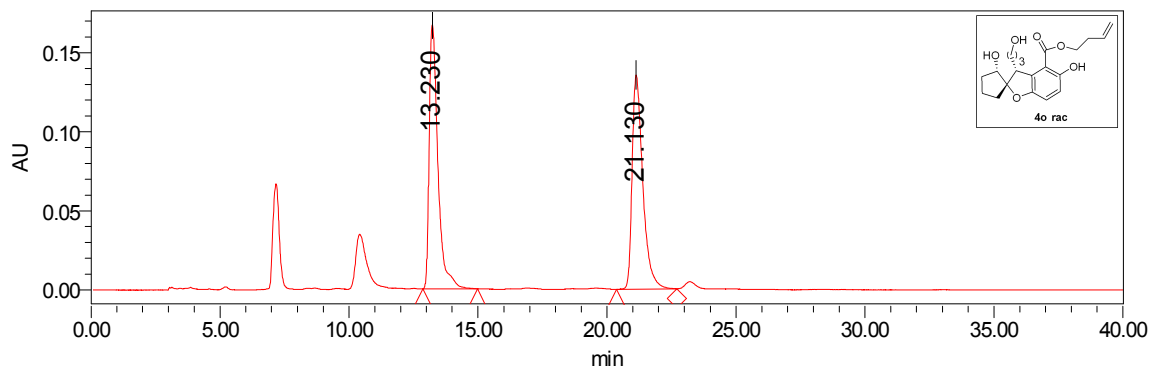
Peak Information:

	RetTime (min)	Area (μ V*S)	Area (%)	Height (μ V)
1	13.772	3949836.133	91.914	177759
2	22.585	347476.858	8.086	12574

Sample Information

Sample Name: 4o rac
Column: 10.00 μ l

Wave Length: PDA 254 nm



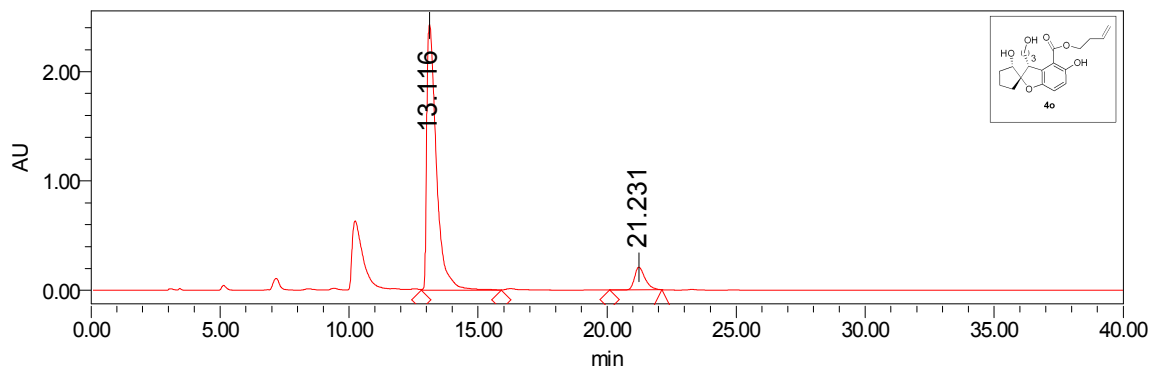
Peak Information:

	RetTime (min)	Area (μ V*S)	Area (%)	Height (μ V)
1	13.230	3923853.165	50.031	167386
2	21.130	3918926.863	49.969	135511

Sample Information

Sample Name: 4o
Column: 10.00 μ l

Wave Length: PDA 254 nm



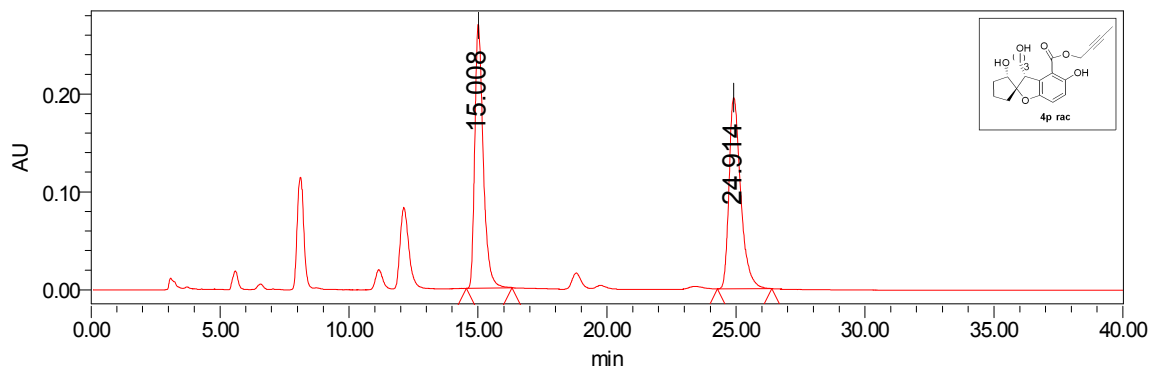
Peak Information:

	RetTime (min)	Area (μ V*S)	Area (%)	Height (μ V)
1	13.116	63101618.153	91.516	2430432
2	21.231	5849544.457	8.484	205636

Sample Information

Sample Name: 4p rac
Column: 10.00 μ l

Wave Length: PDA 254 nm



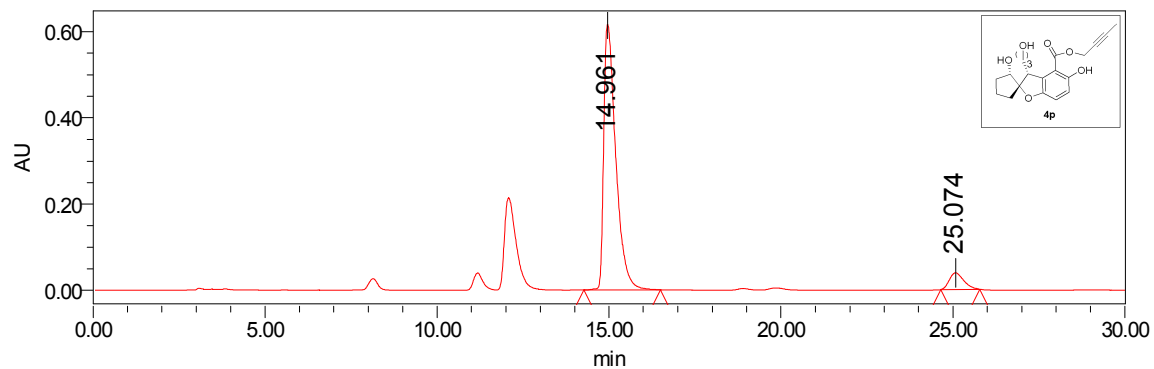
Peak Information:

	RetTime (min)	Area (μ V*S)	Area (%)	Height (μ V)
1	15.008	6179372.165	50.081	269603
2	24.914	6159451.777	49.919	195302

Sample Information

Sample Name: 4p
Column: 10.00 μ l

Wave Length: PDA 254 nm

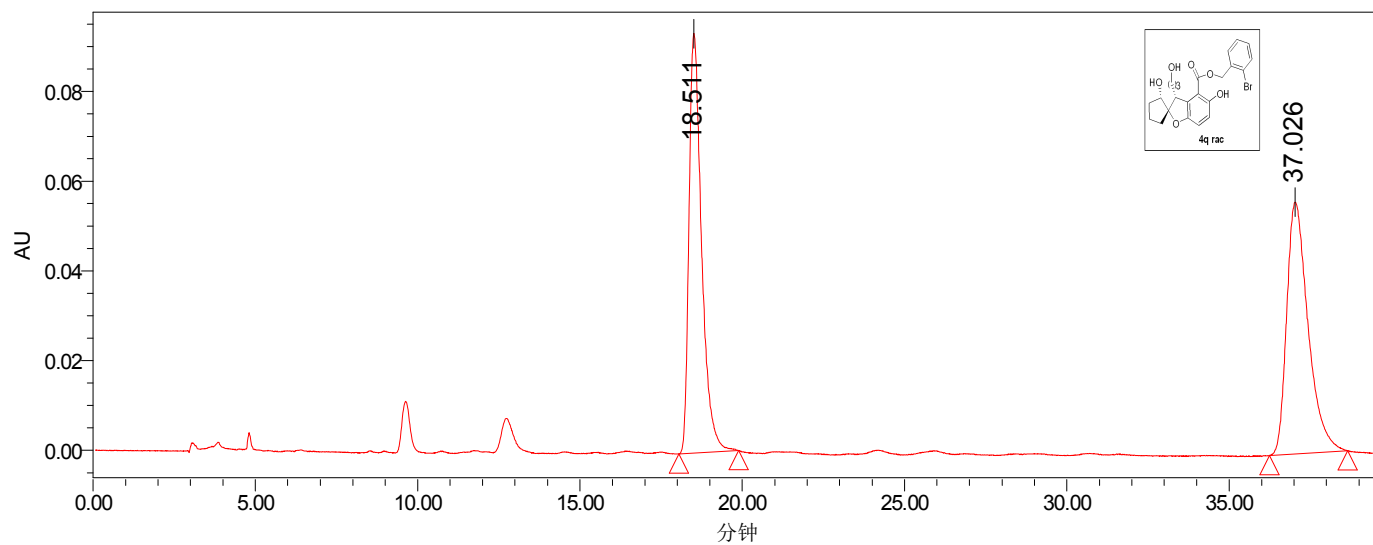


Peak Information:

	RetTime (min)	Area (μ V*S)	Area (%)	Height (μ V)
1	14.961	14945157.820	93.084	615974
2	25.074	1110454.297	6.916	38201

Sample Name: 4q rac
Column: 10.00 ul

Wave Length: PDA 254 nm

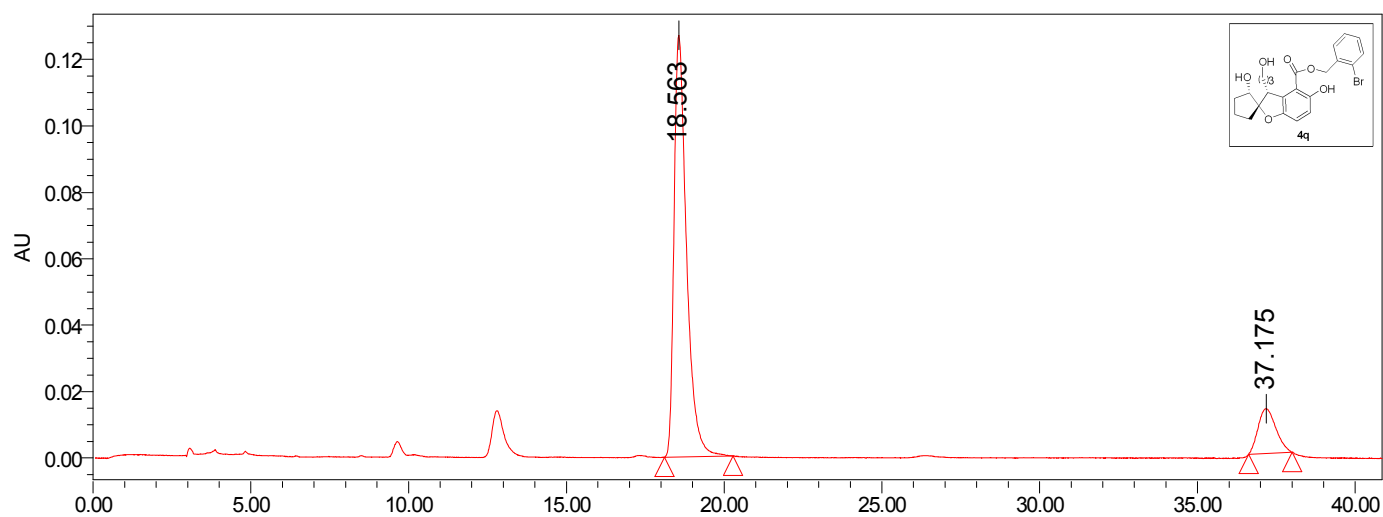


Peak Information:

	RetTime (min)	Area ($\mu\text{V}\cdot\text{S}$)	Area (%)	Height (μV)
1	18.511	2510695	49.41	93528
2	37.026	2570163	50.59	56176

Sample Name: 4q
Column: 10.00 ul

Wave Length: PDA 254 nm



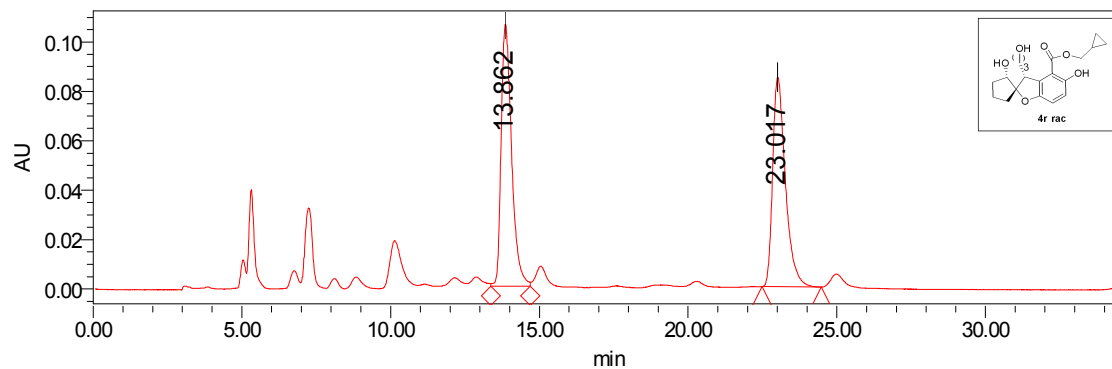
Peak Information:

	RetTime (min)	Area ($\mu\text{V}\cdot\text{S}$)	Area (%)	Height (μV)
1	18.563	3502751	86.80	126891
2	37.175	532642	13.20	13529

Sample Information

Sample Name: 4r rac
Column: 10.00 μ l

Wave Length: PDA 254 nm

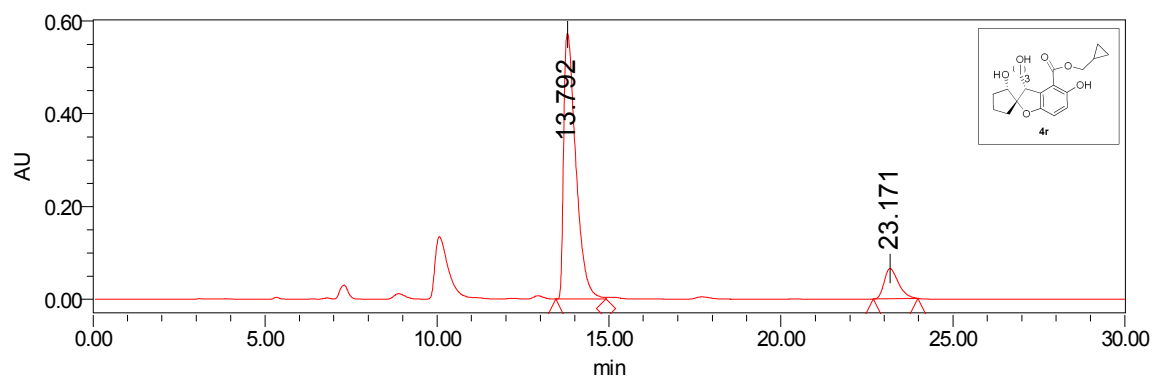


Peak Information:

	RetTime (min)	Area (μ V*S)	Area (%)	Height (μ V)
1	13.862	2578133.577	50.457	105991
2	23.017	2531408.469	49.543	84867

Sample Name: 4r
Column: 10.00 ul

Wave Length: PDA 254 nm



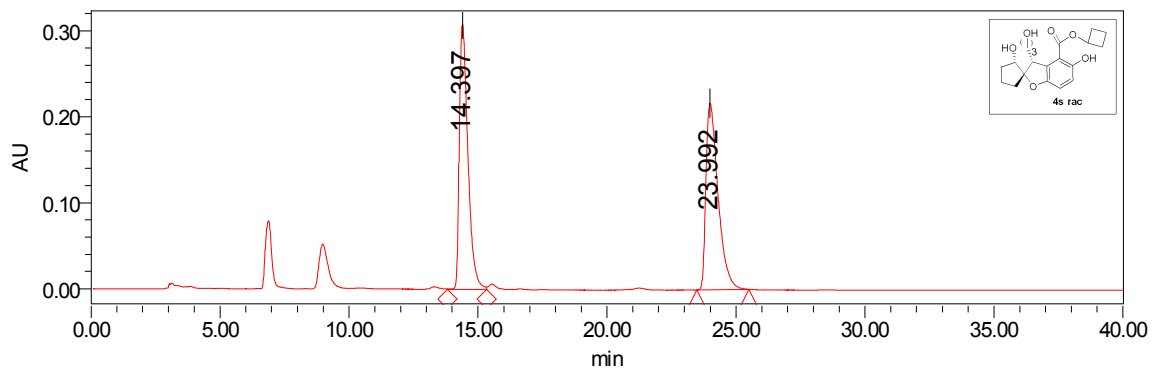
Peak Information:

	RetTime (min)	Area ($\mu\text{V}\cdot\text{S}$)	Area (%)	Height (μV)
1	13.792	14089218.211	88.253	572798
2	23.171	1875434.940	11.747	65144

Sample Information

Sample Name: 4s rac
Column: 10.00 μ l

Wave Length: PDA 254 nm



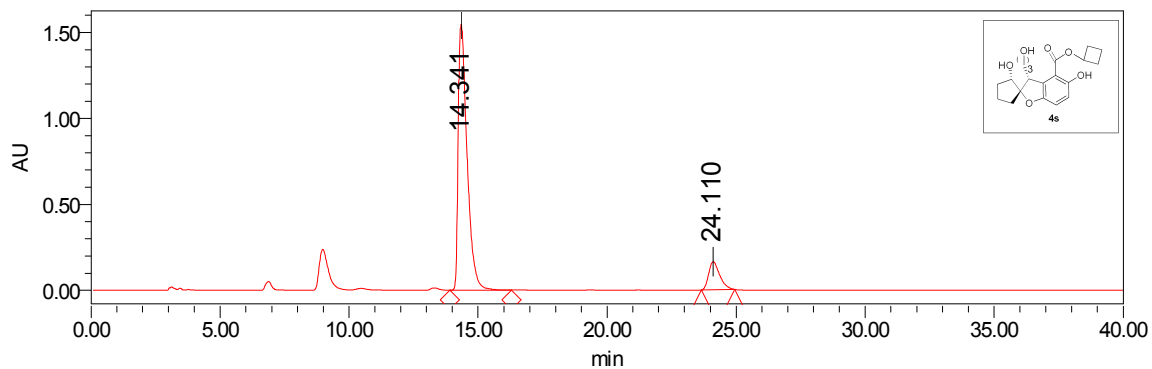
Peak Information:

	RetTime (min)	Area (μ V*S)	Area (%)	Height (μ V)
1	14.397	7080663.682	49.933	308136
2	23.992	7099623.474	50.067	217190

Sample Information

Sample Name: 4s
Column: 10.00 μ l

Wave Length: PDA 254 nm



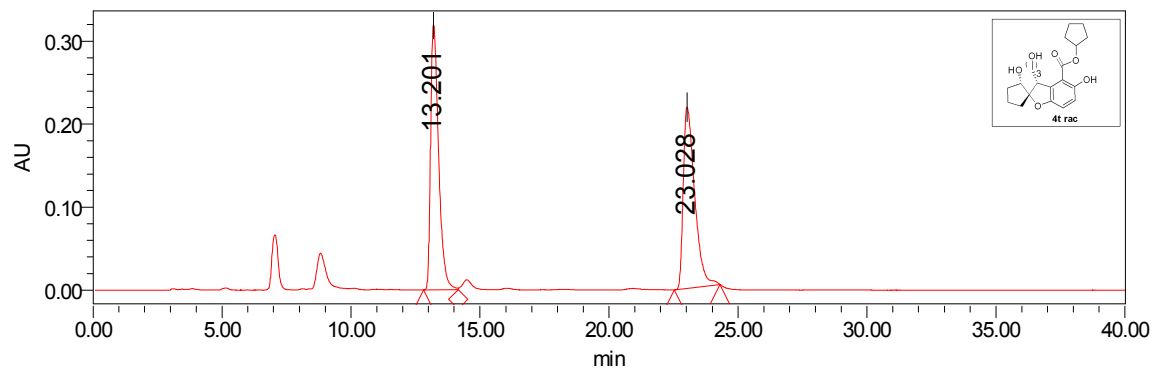
Peak Information:

	RetTime (min)	Area (μ V*S)	Area (%)	Height (μ V)
1	14.341	37110036.273	88.353	1547286
2	24.110	4891996.030	11.647	163167

Sample Information

Sample Name: 4t rac
Column: 10.00 μ l

Wave Length: PDA 254 nm



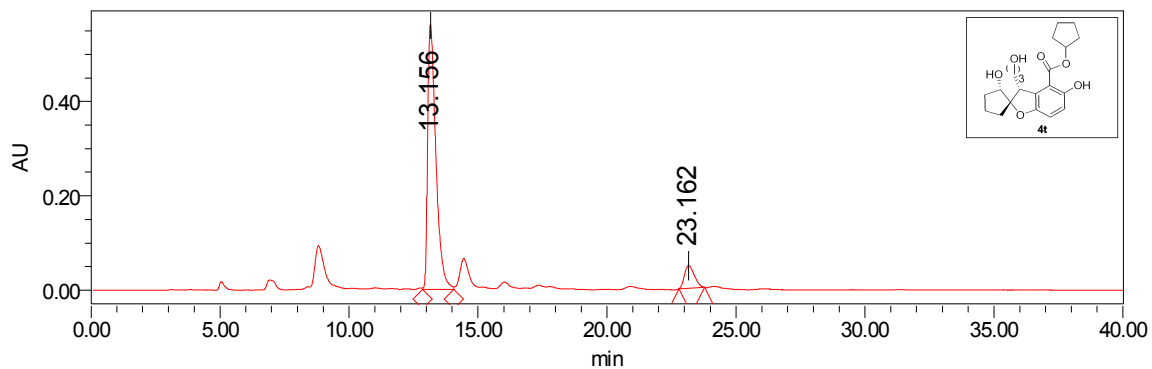
Peak Information:

	RetTime (min)	Area (μ V*S)	Area (%)	Height (μ V)
1	13.201	6866809.017	50.459	319985
2	23.028	6741962.430	49.541	218346

Sample Information

Sample Name: 4t
Column: 10.00 μ l

Wave Length: PDA 254 nm

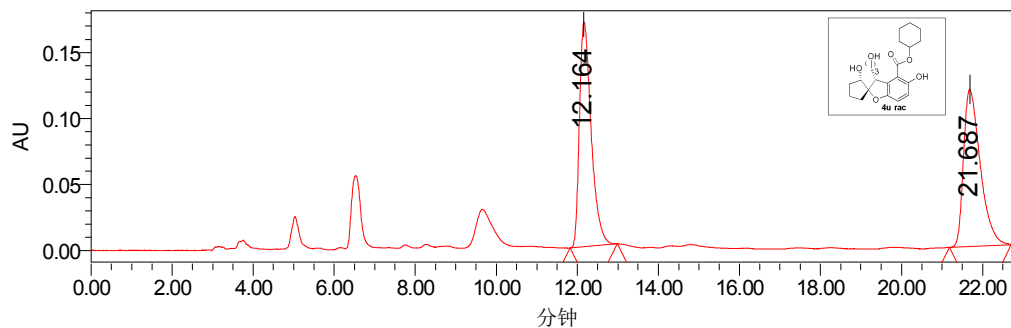


Peak Information:

	RetTime (min)	Area (μ V*S)	Area (%)	Height (μ V)
1	13.156	12347927.869	90.565	561633
2	23.162	1286419.999	9.435	48300

Sample Name: 4u rac
Column: 10.00 ul

Wave Length: PDA 254 nm



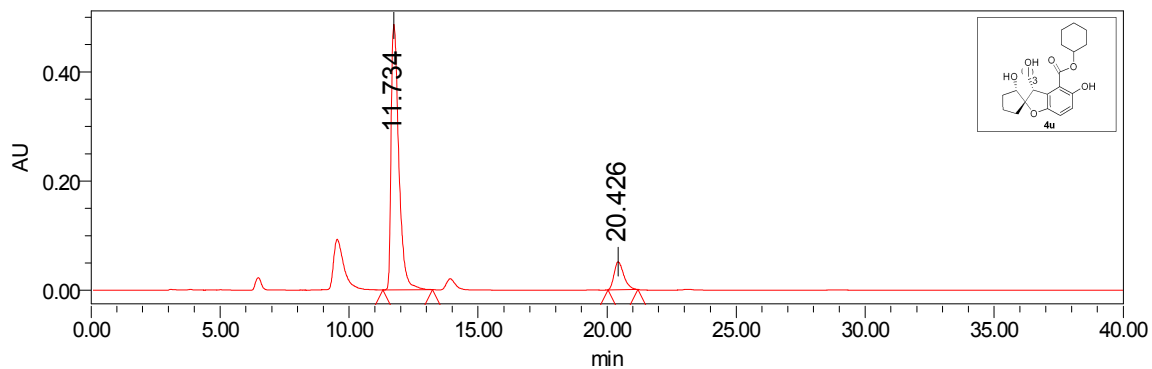
Peak Information:

	RetTime (min)	Area ($\mu\text{V}\cdot\text{S}$)	Area (%)	Height (μV)
1	12.164	3426936.987	50.159	170376
2	21.687	3405262.470	49.841	119052

Sample Information

Sample Name: 4u
Column: 10.00 μ l

Wave Length: PDA 254 nm

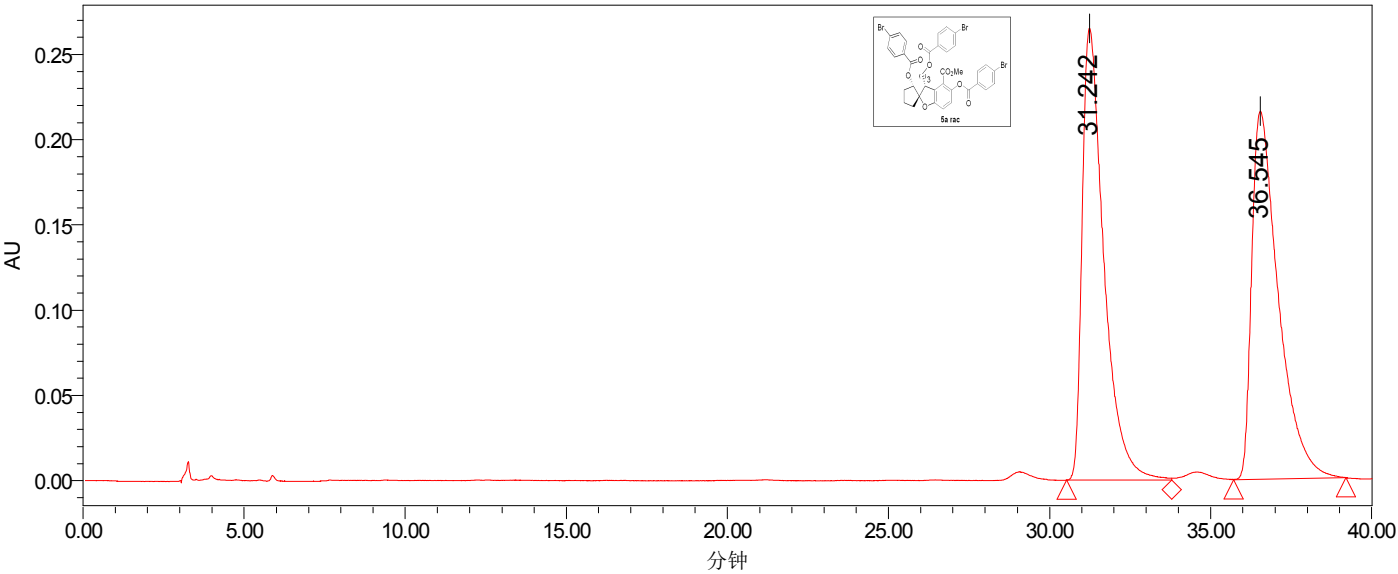


Peak Information:

	RetTime (min)	Area (μ V*S)	Area (%)	Height (μ V)
1	11.734	10081317.903	88.362	485878
2	20.426	1327789.476	11.638	51017

Sample Name: 5a rac
Column: 10.00 ul

Wave Length: PDA 254 nm

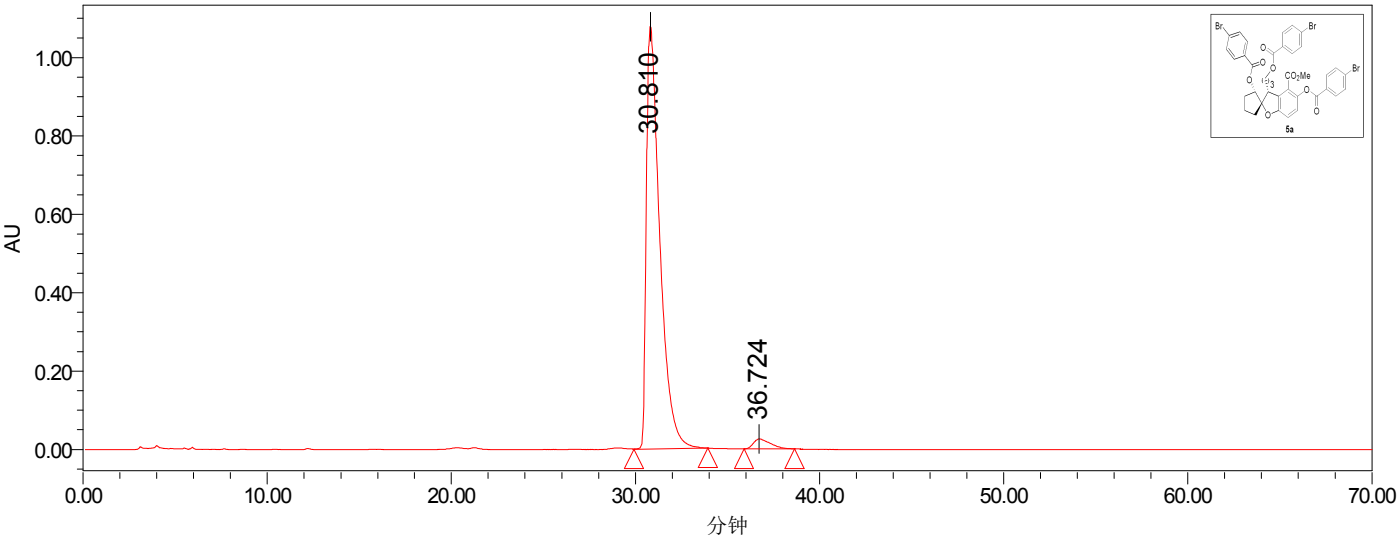


Peak Information:

	RetTime (min)	Area (μV*S)	Area (%)	Height (μV)
1	31.242	12379456	49.95	265110
2	36.545	12403999	50.05	215831

Sample Name: 5a
Column: 10.00 ul

Wave Length: PDA 254 nm



Peak Information:

	RetTime (min)	Area ($\mu\text{V}\cdot\text{S}$)	Area (%)	Height (μV)
1	30.810	54542601	97.04	1077964
2	36.724	1665753	2.96	25449