

SUPPORTING INFORMATION

¹H-Detected quadrupolar spin-lattice relaxation measurements under magic-angle spinning solid-state NMR

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1. Experimental parameters

Variable	²³ Na indirect Fig. 1c	²³ Na direct Fig. 1b	¹⁴ N indirect Fig. 2a	⁸¹ Br indirect Fig. 2b
Spectrometer details: Field / Console / Probe	14.1 T / Bruker AVIII / 4mm	14.1 T / Bruker AVIII / 4mm	9.4 T / Bruker AVIII / 4mm	14.1 T / Bruker AVIII / 4mm
Material and source	Na ₂ HPO ₄ ·2H ₂ O (Merck)	Na ₂ HPO ₄ ·2H ₂ O (Merck)	Glycine (Bio-Lab Chemicals)	Tetra-n-butylammonium bromide (Chem-Impex Int'l Inc.)
Spinning speed	14 kHz	14 kHz	14 kHz	14 kHz
PM pulse power / length (τ _{PM})	33.5 kHz / 10 T _R	33.5 kHz / 10 T _R	30 kHz / 50 T _R	40 kHz / 10 T _R
*Excitation pulse power / length (τ _p)	9.9 kHz / 12.6 μs	9.9 kHz / 12.6 μs	30 kHz / 41.5 μs	51.5 kHz / 2.4 μs
CP power ¹ H / CP power X / pulse length	39 kHz / 14.6 kHz / 1.5 ms		29 kHz / 15 kHz / 2.4 ms	45 kHz / 8.5 kHz / 3.0 ms
¹ H decoupling		100 kHz, swf-tppm		
Recycle delay	1 s	1 s	1.6 s	1 s
Scans	32	4	512	8192
Total experimental time	1.16 hrs	8.7 min	5.71 hrs	63.4 hrs
Temperature**	~45 °C	~45 °C	18.2 °C	~45 °C
Apodization	Exponential, 100 Hz	Exponential, 30 Hz	Exponential, 200 Hz	Exponential, 500 Hz

²³Na experiments: The ¹H-detected {²³Na}¹H CP experiment shown in the inset of Fig. 1c was conducted with 32 scans and all other experimental parameters were similar to Fig. 1c. The Bloch-decay ¹H spectrum was obtained using a 90° pulse of 100 kHz and a single scan. The ¹H T₁ value in this compound is ~7 min, hence a single scan was used for the Bloch-decay ¹H spectrum, and the measurement was performed after a long waiting period.

¹⁴N experiments: The ¹H-detected {¹⁴N}¹H CP experiment in the inset of Fig. 2a was conducted with 128 scans, a recycle delay of 5 s, and the same experimental parameters as the saturation recovery experiment (Fig. 2a). The Bloch-decay ¹H spectrum was obtained using 90° pulse of 100 kHz, a recycle delay of 3 s, and a single scan.

⁸¹Br experiments: The ¹H-detected {⁸¹Br}¹H CP experiment in the inset was conducted with the same experimental parameters used in the saturation recovery experiment. ¹H was saturated in the beginning of the sequence to avoid the presence of any residual signal from direct excitation. The Bloch-decay ¹H spectrum was obtained with a recycle delay of 35 s, and a single scan.

* ⁸¹Br power levels were calibrated using the signal of KBr. ¹⁴N power levels were calibrated using NH₄Cl.

** The temperature in the experiments on ²³Na and ⁸¹Br are the values of the temperature in the room (22 °C) corrected for frictional heating at a spinning speed of 14 kHz, as determined from experiments on PbNO₃ (approximately 23-25 °C). The temperatures in the experiments on ¹⁴N were determined by the second sensor in the probe. Temperature dependent experiments on ¹⁴N are detailed below (Figs. S2, S3) with their explicit experimental values.

> The lack of signal originating from direct excitation of the proton spins was verified by performing experiments without irradiation on the quadrupolar spins.

>>Phase cycle opt. 1: Quadrupolar excitation pulse: y, \bar{y} ; CP(¹H): $x, x, \bar{x}, \bar{x}, y, \bar{y}, y, \bar{y}$; CP(X): x ; receiver: $x, \bar{x}, \bar{x}, x, y, \bar{y}, \bar{y}, y$. (X=²³Na, ¹⁴N, ⁸¹Br)

>>Phase cycle opt. 2: Quadrupolar excitation pulse: y, \bar{y} ; CP(¹H): $(x)_4, (\bar{x})_4, (y)_4, (\bar{y})_4$; CP(X): $(x)_2, (\bar{x})_2$; receiver: $x, \bar{x}, \bar{x}, x, \bar{x}, x, \bar{x}, y, \bar{y}, \bar{y}, y, \bar{y}, y, \bar{y}$.

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2. Temperature dependent ^1H -detected ^{14}N PM-saturation recovery spin-lattice (T_1) relaxation measurements in natural-abundance glycine.

Experimental variables different from the Table above are indicated explicitly.

The fit values were used to produce Figure 3 in the article.

Figs. S2.1-S2.5 were acquired with a long ^{14}N excitation pulse τ_p ; Figs. S2.6-S2.9 were acquired with a short excitation pulse.

All curves were fit using the MATLAB tool 'cftools'. The errors report a confidence level of 95%.

Figure S2.1: Temperature (T)=18.2 °C.
The mono-exponential fit yields $m = 0.99 \pm 0.03$; $a = 1.07 \pm 0.03$; $T_1 = 590 \pm 77$ ms.
 $\tau_{\text{PM}} = 30$ TR; $\tau_p=41.5$ μs . Total experimental time (t_{exp}) = 5.71 hrs.

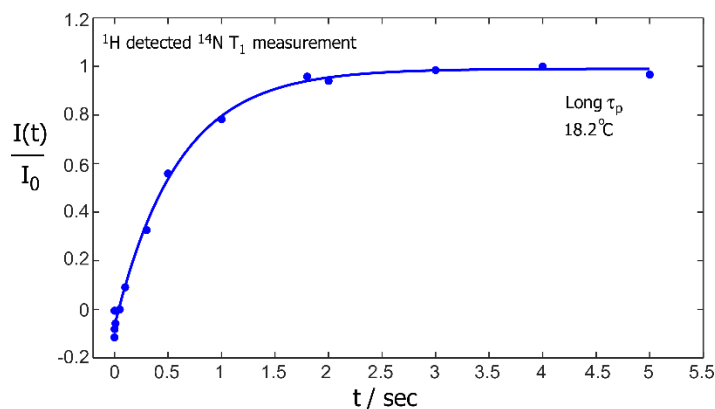
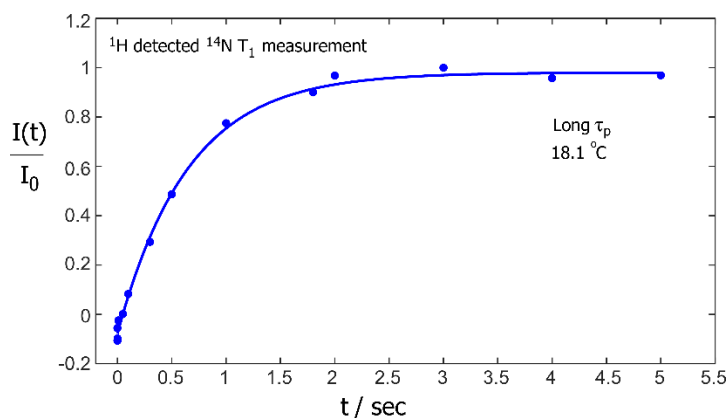


Figure S2.2: $T=18.1$ °C.
The mono-exponential fit yields $m = 0.98 \pm 0.03$; $a = 1.08 \pm 0.03$; $T_1 = 648 \pm 74$ ms.
 $\tau_p=48$ μs ; CP power ^1H : 29.8 kHz, power ^{14}N 15 kHz, length 3ms. $t_{\text{exp}} = 5.71$ hrs.



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Figure S2.3: T=-28.3 °C.

The mono-exponential fit yields $m = 0.98 \pm 0.02$; $a = 1.01 \pm 0.03$; $T_1 = 255 \pm 25$ ms.
 $\tau_p = 48 \mu\text{s}$; recycle delay = 9.6 s. $t_{\text{exp}} = 21.63$ hrs.

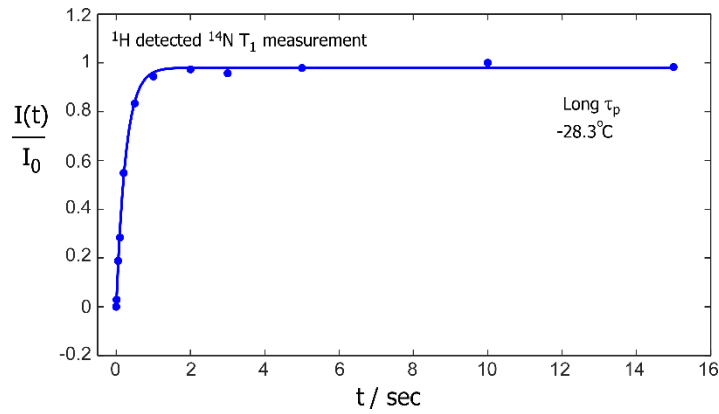


Figure S2.4: T = 47.2 °C.

Fit: $m = 0.97 \pm 0.04$; $a = 1.07 \pm 0.05$; $T_1 = 839 \pm 143$ ms.
 $\tau_p = 48 \mu\text{s}$; recycle delay = 1.3 s; 1024 scans. $t_{\text{exp}} = 11.85$ hrs.

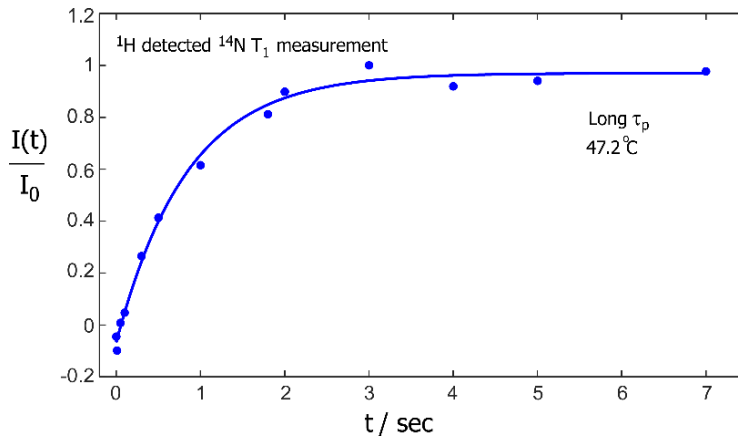
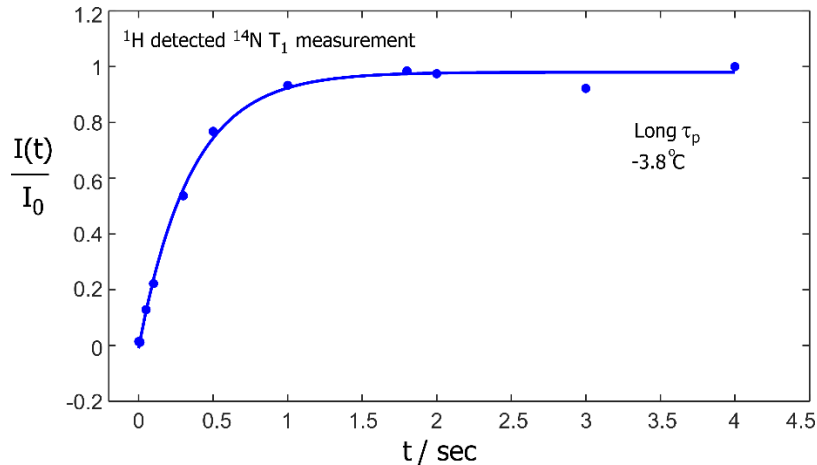


Figure S2.5: T = -3.8 °C.

Fit: $m = 0.98 \pm 0.03$; $a = 1.01 \pm 0.04$; $T_1 = 349 \pm 51$ ms.
 $\tau_p = 48 \mu\text{s}$; recycle delay = 3.2 s. $t_{\text{exp}} = 6.82$ hrs.



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Figure S2.6: $T = -3.9\text{ }^{\circ}\text{C}$.

Fit: $m = 0.99 \pm 0.05$; $a = 1.15 \pm 0.06$; $T_1 = 449 \pm 82\text{ ms}$.

$\tau_p = 8.3\text{ }\mu\text{s}$; recycle delay = 3.2 s; 1280 scans. $t_{\text{exp}} = 17.05\text{ hrs}$.

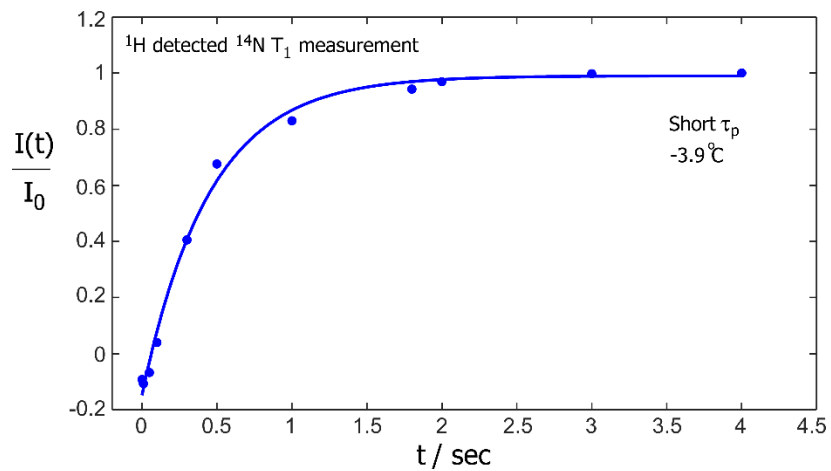
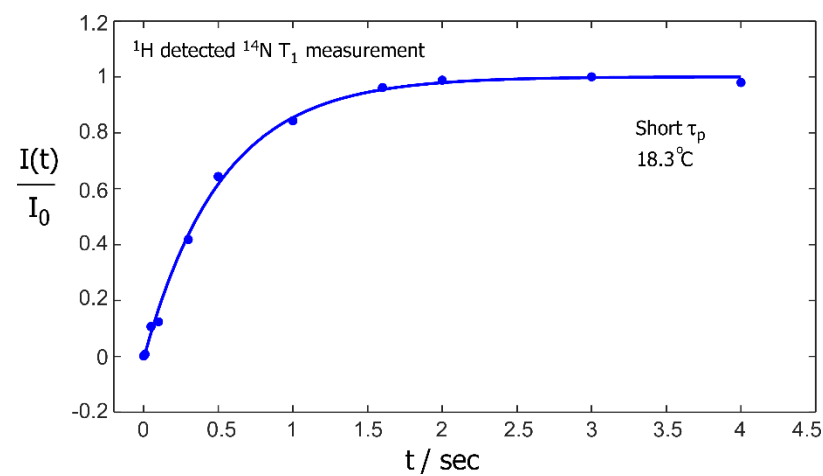


Figure S2.7: $T = 18.3\text{ }^{\circ}\text{C}$.

Fit: $m = 1.00 \pm 0.03$; $a = 1.01 \pm 0.03$; $T_1 = 517 \pm 64\text{ ms}$.

$\tau_p = 8.3\text{ }\mu\text{s}$; recycle delay = 1.6 s; 2048 scans. $t_{\text{exp}} = 17.16\text{ hrs}$.



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Figure S2.8: $T = 47.3\text{ }^{\circ}\text{C}$.
Fit: $m = 0.99 \pm 0.06$; $a = 1.27 \pm 0.06$; $T_1 = 862 \pm 160\text{ ms}$.
 $\tau_p = 8.3\text{ }\mu\text{s}$; recycle delay = 1.3 s; 2048 scans. $t_{\text{exp}} = 18.86\text{ hrs}$.

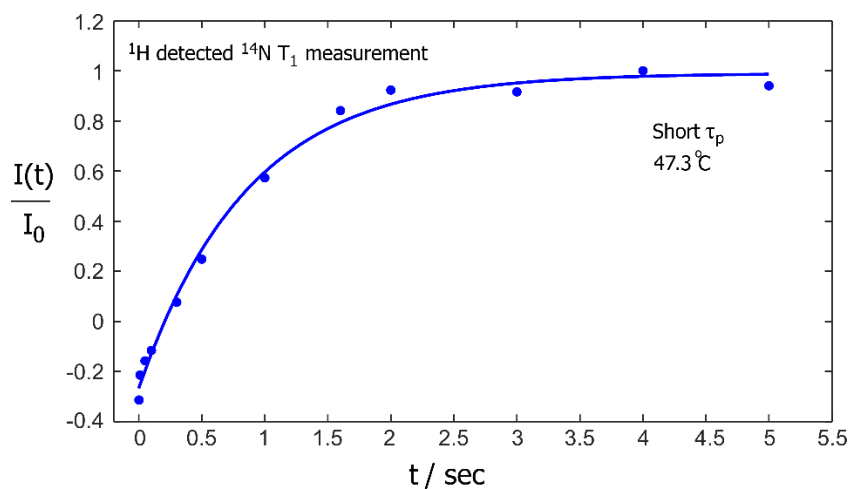
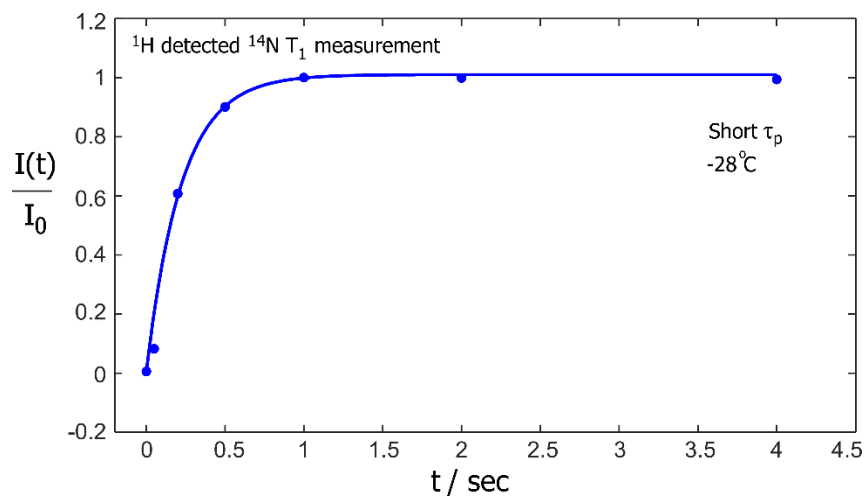


Figure S2.9: $T = -28\text{ }^{\circ}\text{C}$.
Fit: $m = 1.01 \pm 0.08$; $a = 1.0 \pm 0.1$; $T_1 = 223 \pm 85\text{ ms}$.
 $\tau_p = 8.3\text{ }\mu\text{s}$; recycle delay = 9.6 s; 768 scans. $t_{\text{exp}} = 15.99\text{ hrs}$.



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3. Direct-excitation ^{81}Br spectrum and phase-modulated T_1 saturation recovery curve.

Figure S3.1: Single excitation pulse spectrum of ^{81}Br in TBAB, Tetra-n-butylammonium bromide. The spectrum was acquired with an excitation pulse of 2.4 μs , rf power level of 51.5 kHz, a spinning speed of 14 kHz, 512 scans, and a recycle delay of 1 s. Proton decoupling of 100 kHz (swf-tppm) was applied. The spectrum was processed with an exponential apodization function with a value of 500 Hz. $t_{\text{exp}} = 9$ min.

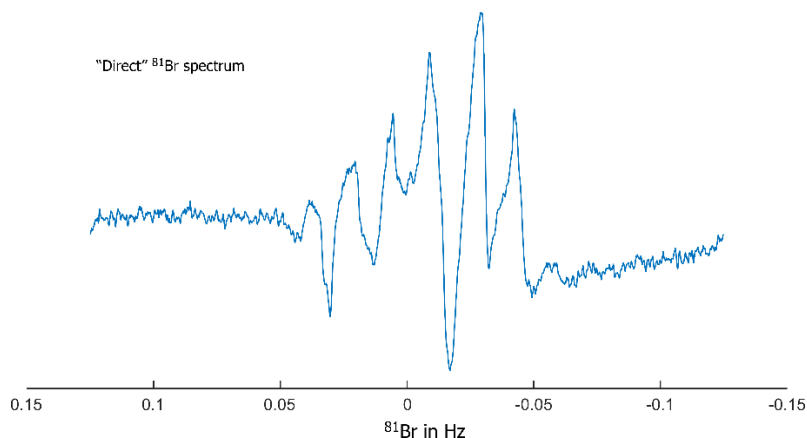
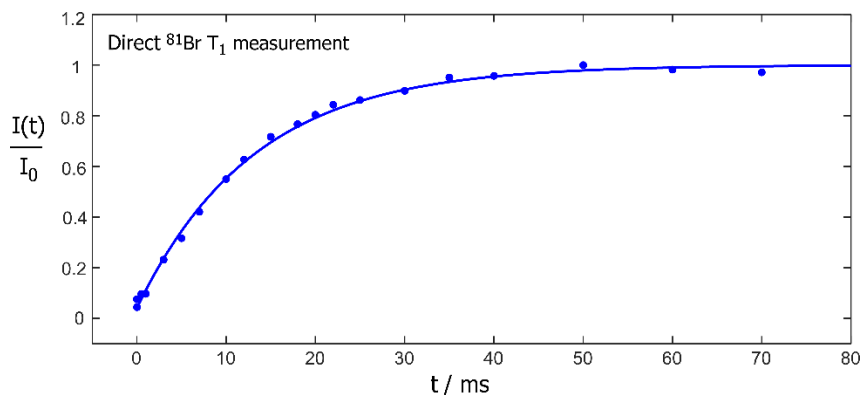


Figure S3.2: Direct ^{81}Br -detected phase-modulated saturation recovery experiment. Each data point was extracted by integrating the directly-detected spectrum. The saturation recovery sequence is given in [1] and the PM pulse lasted 10 rotor periods. The spinning speed was 14 kHz. The T_1 relaxation time is 13 ± 1 ms. $t_{\text{exp}} = 2.9$ hrs.



4. References

- [1] M. Makrinich, R. Gupta, T. Polenova, A. Goldbourt, Saturation capability of short phase modulated pulses facilitates the measurement of longitudinal relaxation times of quadrupolar nuclei, *Solid State Nucl. Magn. Reson.* 84 (2017) 196-203.