## Electronic Supplementary Information (ESI)

Photocontrolled morphological conversion and chiral transfer ofsnowflake-like supramolecular assembly based on azobenzene-bridgedbis(dibenzo-24-crown-8) and a cholesterol derivative
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Experimental section ..... S2
Scheme S1. Synthesis route of 1 and 2 ..... S3
Figure S1-S10. Characterization of 1, 3, 4 and 2 ..... S6
Figure S11. Absorption spectra of $\mathbf{1}$ ..... S11
Figure S12. ${ }^{1} \mathrm{H}$ NMR spectra of $\mathbf{1}$ upon UV/vis irradiation ..... S12
Figure S13. UV-vis titration spectra of $\mathbf{2} \subset$ trans- $\mathbf{1}$ ..... S12
Figure S14. SEM images and DLS data of $\mathbf{1}$ ..... S13
Scheme S2. Schematic representations of 2 ..... S13
Figure S15. SEM images of $\mathbf{2} \subset \mathbf{1}$ ..... S14

## Experimental Section

## Instrumentation and methods

All the reagents and solvents were commercially available and used as received without further purification. The precursor 4-chloromethylenedibenzo-24-crown-8 ether was prepared according to literature procedures ${ }^{1}$. Column chromatography was performed on silica gel (200-300 mesh). NMR spectra were recorded on Bruker AV400 instrument at $25{ }^{\circ} \mathrm{C}$ and chemical shifts were recorded in parts per million (ppm). Electrospray ionization (ESI) mass spectra were measured on a Q-TOF LC-MS. High-resolution matrix-assisted laser desorption/ionization spectra (HR-MALDI) were measured on a Varian 7.0T FT-MS. The UV light irradiation experiment was carried out using a ZF-7A lamp ( $365 \mathrm{~nm}, 8 \mathrm{~W}$ ), and the visible light irradiation experiment ( $\lambda>420 \mathrm{~nm}$ ) was carried out using a CEL HXF300 xenon lamp with cutoff filter. UV/vis spectra were recorded on a Shimadzu UV-3600 spectrophotometer equipped with a PTC-348WI temperature controller. The sample for TEM measurements was prepared by dropping the solution onto a copper grid. The grid was then air-dried. The samples were examined by a high-resolution TEM (Tecnai G2 F20 microscope, FEI) equipped with a CCD camera (Orius 832, Gatan) operating at an accelerating voltage with a CCD camera (Orius 832, Gatan) operating at an accelerating voltage of 200 Kv . The molecular packing model of supramolecular assemblies voltage of 200 kV . CD spectra were collected on a JASCO J-715 Circular Dichroism spectroimeter.

Scheme S1. Synthetic route of 1 and 2.





## Synthesis of azobenzene-bridged bis(dibenzo-24-crown-8) (1).

4,4'-Dihydroxyazobenzene ( $60.46 \mathrm{mg}, 0.28 \mathrm{mmol}$ ) was added to a stirred solution of 4-chloromethylenedibenzo-24-crown-8 ether ( $350 \mathrm{mg}, 0.7 \mathrm{mmol}$ ) in acetone ( 80 ml ), to which $\mathrm{K}_{2} \mathrm{CO}_{3}(681.16 \mathrm{mg}, 4.94 \mathrm{mmol})$ was added under argon at room temperature. The mixture was stirred under reflux for 7 days. After cooling to room temperature, the mixture was filtered and the filtrate was evaporated under reduced pressure to remove acetone. The residue was dissolved in chloroform ( 50 ml ) and washed with water ( $2 \times 50 \mathrm{ml}$ ), dried with anhydrous $\mathrm{MgSO}_{4}$ and evaporated to a yellow residue which was purified by silica gel column chromatography using chloroform-methanol solution ( $\mathrm{v} / \mathrm{v}=200: 1$ ) as elutent to give azobenzene-bridged bis(dibenzo-24-crown-8) ( 123.8 mg ) as a yellow powder (yield $38.97 \%$ ). ${ }^{1} \mathrm{H}$ NMR ( $400 \mathrm{MHz}, \mathrm{CDCl}_{3}$ ): $\delta 7.86(\mathrm{~d}, J=8.7 \mathrm{~Hz}, 4 \mathrm{H}), 7.05(\mathrm{~d}, J=8.8 \mathrm{~Hz}, 4 \mathrm{H}), 6.97(\mathrm{~d}, J=$ $5.1 \mathrm{~Hz}, 4 \mathrm{H}), 6.88(\mathrm{~s}, 10 \mathrm{H}), 5.03(\mathrm{~s}, 4 \mathrm{H}), 4.16(\mathrm{~s}, 16 \mathrm{H}), 3.92(\mathrm{~s}, 16 \mathrm{H}), 3.83(\mathrm{~s}, 16 \mathrm{H}) .{ }^{13} \mathrm{C}$ NMR ( $101 \mathrm{MHz}, \mathrm{CDCl}_{3}$ ): $\delta 159.70,148.01,147.86,147.82,146.12,128.44,123.30,120.37,119.79$, 114.03, 112.97, 112.70, 112.47, 70.26, 69.17, 68.43, 68.34. MALDI-MS for $\mathbf{1}\left(\mathrm{C}_{62} \mathrm{H}_{74} \mathrm{~N}_{2} \mathrm{O}_{18}\right)$ : calcd. $\left[\mathrm{M}+\mathrm{Na}^{+}\right]^{+}: 1157.4834$, found: 1157.4833.

## Synthesis of cholesteryl aldehyde benzoate (3).

4-Formylbenzoic acid ( $555 \mathrm{mg}, 3.7 \mathrm{mmol}$ ), cholesterol ( $1.43 \mathrm{~g}, 3.7 \mathrm{mmol}$ ), DMAP ( 0.23 g , $1.9 \mathrm{mmol})$ and $\operatorname{DCC}(0.84 \mathrm{~g}, 4.07 \mathrm{mmol})$ were dissolved in chloroform $(100 \mathrm{ml})$. The mixture was stirred 4 h under argon at room temperature. The mixture was evaporated under reduced pressure to remove chloroform. The residue was purified by silica gel column chromatography using petroleum ether-chloroform ( $\mathrm{v} / \mathrm{v}=5: 3$ ) as elutent to give product (1.13 $\mathrm{g})$ as a white solid (yield $58.9 \%) .{ }^{1} \mathrm{H}$ NMR ( $400 \mathrm{MHz}, \mathrm{CDCl}_{3}$ ): $\delta 10.09(\mathrm{~s}, 1 \mathrm{H}), 8.17(\mathrm{~d}, \mathrm{~J}=$ $7.6 \mathrm{~Hz}, 2 \mathrm{H}), 7.94(\mathrm{~d}, J=8.0 \mathrm{~Hz}, 2 \mathrm{H}), 5.42(\mathrm{~s}, 1 \mathrm{H}), 4.89(\mathrm{~s}, 1 \mathrm{H}), 2.46(\mathrm{~d}, J=7.6 \mathrm{~Hz}, 2 \mathrm{H}), 2.00$ $(\mathrm{s}, 3 \mathrm{H}), 1.85-1.71(\mathrm{~m}, 3 \mathrm{H}), 1.62-1.44(\mathrm{~m}, 7 \mathrm{H}), 1.38-1.29(\mathrm{~m}, 4 \mathrm{H}), 1.17-1.05(\mathrm{~m}, 9 \mathrm{H}), 0.99(\mathrm{~d}$, $J=7.0 \mathrm{~Hz}, 3 \mathrm{H}), 0.91(\mathrm{~d}, J=6.2 \mathrm{~Hz}, 3 \mathrm{H}), 0.86(\mathrm{~d}, J=6.1 \mathrm{~Hz}, 6 \mathrm{H}), 0.681(\mathrm{~s}, 3 \mathrm{H}) .{ }^{13} \mathrm{C}$ NMR (101 MHz, $\mathrm{CDCl}_{3}$ ): $\delta 190.54,163.86,138.38,138.04,134.83,129.11,128.39,122.00,74.35$, $55.69,55.18,49.06,38.52,36.00,34.79,30.93,30.88,27.22,27.00,26.84,22.85,21.80$, 21.56, 20.06, 18.35, 17.73, 10.86.

## Synthesis of methylenebenzylamine benzoic acid cholesterylamine (4).

Cholesteryl aldehyde benzoate ( $0.518 \mathrm{~g}, 1 \mathrm{mmol}$ ), benzylamine ( $0.13 \mathrm{~g}, 1.2 \mathrm{mmol}$ ) and triethylamine ( $0.24 \mathrm{~g}, 4 \mathrm{mmol}$ ) were dissolved in methanol and chloroform. The mixture was stirred under reflux for 1 day under argon. After cooling to room temperature, the sodium borohydride ( $0.61 \mathrm{~g}, 16 \mathrm{mmol}$ ) was added to the reaction system under ice bath conditions. Then the mixture was stirred 24 h under argon at room temperature. Then water was added to stop the reaction. After the solution was concentrated, the residue was dissolved with $\mathrm{CH}_{2} \mathrm{Cl}_{2}$ $(50 \mathrm{ml})$ and washed with $\mathrm{H}_{2} \mathrm{O}(2 \times 50 \mathrm{ml})$. The organic phase was dried by $\mathrm{MgSO}_{4}$ and evaporated off. Then the residue was purified by column chromatography using chloroform as elutent to give product $(0.41 \mathrm{~g})$ as a white solid (yield $68.8 \%) .{ }^{1} \mathrm{H}$ NMR $\left(400 \mathrm{MHz}, \mathrm{CDCl}_{3}\right): \delta$ $8.02(\mathrm{~d}, J=8.0 \mathrm{~Hz}, 2 \mathrm{H}), 7.42(\mathrm{~d}, J=8.0 \mathrm{~Hz}, 2 \mathrm{H}), 7.34(\mathrm{~d}, J=4.2 \mathrm{~Hz}, 5 \mathrm{H}), 5.44(\mathrm{~s}, 1 \mathrm{H}), 4.87$ ( $\mathrm{s}, 1 \mathrm{H}$ ), $3.83(\mathrm{~d}, ~ J=24.7 \mathrm{~Hz}, 4 \mathrm{H}), 2.48(\mathrm{~d}, ~ J=7.7 \mathrm{~Hz}, 2 \mathrm{H}), 2.10-1.95(\mathrm{~m}, 3 \mathrm{H}), 1.84-1.69(\mathrm{~m}$, $3 \mathrm{H}), 1.65-1.44(\mathrm{~m}, 7 \mathrm{H}), 1.43-1.32(\mathrm{~m}, 4 \mathrm{H}), 1.19-1.07(\mathrm{~m}, 9 \mathrm{H}), 1.05-0.99(\mathrm{~m}, 3 \mathrm{H}), 0.95(\mathrm{~d}, J=$ $6.3 \mathrm{~Hz}, 3 \mathrm{H}), 0.89(\mathrm{~d}, J=6.1 \mathrm{~Hz}, 6 \mathrm{H}), 0.71(\mathrm{~s}, 3 \mathrm{H}) .{ }^{13} \mathrm{C}$ NMR ( $101 \mathrm{MHz}, \mathrm{CDCl}_{3}$ ): $\delta 165.91$, $145.51,140.08,139.70,129.73,129.58,128.47,128.17,127.92,127.08,122.78,74.52,56.73$, $56.20,53.17,52.79,50.09,42.37,39.80,39.58,38.29,37.10,36.69,36.25,35.86,31.99$, 31.93, 28.30, 28.06, 27.96, 24.35, 23.91, 22.90, 22.64, 21.11, 19.44, 18.79, 11.93.

## Synthesis of 2.

Methylenebenzylamine benzoic acid cholesterylamine: ( $154.96 \mathrm{mg}, 0.25 \mathrm{mmol}$ ), trifluoroacetic acid ( $144.98 \mathrm{mg}, 1.25 \mathrm{mmol}$ ) were dissolved in chloroform. The mixture was stirred 2 h at room temperature. Then the solvent was evaporated to obtain a white solid. Then the solid was dissolved in chloroform and methonal. A saturated solution of $\mathrm{NH}_{4} \mathrm{PF}_{6}$ was added to the solution until no further precipitation was observed. The precipitate was filtered off, washed with water and dried to obtain aim product as a yellow solid ( 119.1 mg , yield $62 \%$ ). ${ }^{1} \mathrm{H}$ NMR ( $400 \mathrm{MHz}, \mathrm{CDCl}_{3}$ ): $\delta 8.01$ (d, $J=8.2 \mathrm{~Hz}, 2 \mathrm{H}$ ), 7.41 (d, $J=8.2 \mathrm{~Hz}, 2 \mathrm{H}$ ), 7.35 $(\mathrm{d}, J=4.4 \mathrm{~Hz}, 4 \mathrm{H}) 7.30-7.27(\mathrm{~m}, 1 \mathrm{H}), 5.42(\mathrm{~d}, J=3.9 \mathrm{~Hz}, 1 \mathrm{H}), 4.90-4.81(\mathrm{~m}, 1 \mathrm{H}), 3.868(\mathrm{~s}$, $2 \mathrm{H}), 3.808(\mathrm{~s}, 2 \mathrm{H}), 2.46(\mathrm{~d}, J=7.7 \mathrm{~Hz}, 2 \mathrm{H}), 2.07-1.96(\mathrm{~m}, 3 \mathrm{H}), 1.94-1.71(\mathrm{~m}, 3 \mathrm{H}), 1.63-1.42$ $(\mathrm{m}, 7 \mathrm{H}), 1.40-1.29(\mathrm{~m}, 4 \mathrm{H}), 1.19-1.07(\mathrm{~m}, 9 \mathrm{H}), 1.05-0.98(\mathrm{~m}, 3 \mathrm{H}), 0.93(\mathrm{~d}, J=6.5 \mathrm{~Hz}, 3 \mathrm{H})$,
$0.88(\mathrm{~d}, J=1.7 \mathrm{~Hz}, 3 \mathrm{H}), 0.87(\mathrm{~d}, J=1.6 \mathrm{~Hz}, 3 \mathrm{H}), 0.695(\mathrm{~s}, 3 \mathrm{H}) .{ }^{13} \mathrm{C}$ NMR ( $101 \mathrm{MHz}, \mathrm{CDCl}_{3}$ ): $\delta 164.85,143.74,138.65,138.36,128.71,127.48,127.23,127.00,126.20,121.73,73.51$, 55.68, 55.13, 51.92, 51.52, 49.04, 41.31, 38.73, 38.50, 37.21, 35.64, 35.17, 34.78, 30.92, 30.87, 27.22, 27.00, 26.88, 23.28, 22.82, 21.80, 21.55, 20.04, 18.37, 17.71, 10.85. ESI-MS calcd. $\left[\mathrm{M}-\mathrm{PF}_{6}{ }^{-}\right]^{-}: 610.4619$, found: 610.4620 .


Figure S1. ${ }^{1} \mathrm{H}$ NMR spectrum of $\mathbf{1}$ in $\mathrm{CDCl}_{3}, 400 \mathrm{MHz}, 298 \mathrm{~K}$.


Figure S2. ${ }^{13} \mathrm{C}$ NMR spectrum of $\mathbf{1}$ in $\mathrm{CDCl}_{3}, 100 \mathrm{MHz}, 298 \mathrm{~K}$.


Figure S3. MADILI-TOFMS spectrum of $1\left(\mathrm{C}_{62} \mathrm{H}_{74} \mathrm{~N}_{2} \mathrm{O}_{18}\right)$ : calcd. $\left[\mathrm{M}+\mathrm{Na}^{+}\right]^{+}$: 1157.4834, found: 1157.4833.


Figure S4. ${ }^{1} \mathrm{H}$ NMR spectrum of $\mathbf{3}$ in $\mathrm{CDCl}_{3}, 400 \mathrm{MHz}, 298 \mathrm{~K}$.


Figure S5. ${ }^{13} \mathrm{C}$ NMR spectrum of 3 in $\mathrm{CDCl}_{3}, 100 \mathrm{MHz}, 298 \mathrm{~K}$.


Figure S6. ${ }^{1} \mathrm{H}$ NMR spectrum of $\mathbf{4}$ in $\mathrm{CDCl}_{3}, 400 \mathrm{MHz}, 298 \mathrm{~K}$.


Figure S7. ${ }^{13} \mathrm{C}$ NMR spectrum of $\mathbf{4}$ in $\mathrm{CDCl}_{3}, 100 \mathrm{MHz}, 298 \mathrm{~K}$.


Figure S8. ${ }^{1} \mathrm{H}$ NMR spectrum of $\mathbf{2}$ in $\mathrm{CDCl}_{3}, 400 \mathrm{MHz}, 298 \mathrm{~K}$.



Figure S9. ${ }^{13} \mathrm{C}$ NMR spectrum of $\mathbf{2}$ in $\mathrm{CDCl}_{3}, 100 \mathrm{MHz}, 298 \mathrm{~K}$.

Figure S10. The ESI-MS spectrum of $2\left(\mathrm{C}_{42} \mathrm{H}_{60} \mathrm{NO}_{2} \mathrm{PF}_{6}\right)$ : calcd. $\left[\mathrm{M}-\mathrm{PF}_{6}{ }^{-}\right]^{-}: 610.4619$, found: 610.4620 .


Figure S11. Absorption spectra of $\mathbf{1}$ upon irradiation at 365 nm for different time in $\mathrm{CH}_{2} \mathrm{Cl}_{2}$ at $298 \mathrm{~K}\left([1]=3 \times 10^{-5} \mathrm{M}\right)$.


Figure S12. Partial ${ }^{1} \mathrm{H}$ NMR spectra ( $400 \mathrm{MHz}, \mathrm{CDCl}_{3}, 298 \mathrm{~K}$ ) of a) trans-1, b) sample (a) irradiated with 365 nm light for 50 min , c) sample (b) irradiated with > 420 nm light for 30 min.


Figure S13. UV-vis titration spectra of trans-1 $\left(2 \times 10^{-5} \mathrm{M}\right)$ upon addition of $\mathbf{2}\left((0-160) \times 10^{-6}\right.$ M) in $\mathrm{CH}_{2} \mathrm{Cl}_{2}$ at 298 K , Inset: Nonlinear least sqares fit of the absorption changes at 257 nm to determine the complex stability constant $\left(\mathrm{K}_{\mathrm{s}}\right)$ as $13400 \pm 700 \mathrm{M}^{-1}$.


Figure S14. SEM image of a) trans-1, b) cis-1, DLS data of c) trans-1, d) cis-1. All samples were prepared in $\mathrm{CH}_{2} \mathrm{Cl}_{2}$ at 298 K . ([trans-1 $]=[$ cis-1 $\left.]=3.3 \times 10^{-4} \mathrm{M}\right)$.

Scheme S2. Schematic representations of 2.



Figure S15. SEM images of $\mathbf{2} \subset$ trans $-\mathbf{1}$ in $\mathrm{CH}_{2} \mathrm{Cl}_{2}$ upon the irradiation of 365 nm light for a) 0 $\min ,\left(\mathbf{2} \subset\right.$ trans-1) b) $50 \mathrm{~min}(2 \subset c i s-1)\left(\mathbf{1}: \mathbf{2}=1: 2,[\mathbf{1}]=3.3 \times 10^{-4} \mathrm{M}\right)$.

## References

1. M. Han, H.-Y. Zhang, L.-X. Yang, Q. Jiang and Y. Liu, Org. Lett. 2008, 10, 5557-5560.
