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Orchestrated Catalytic Double Rollover Annulation:Rapid Access to N-Enriched Cationic and Neutral PAHs

Pirudhan Karak,^a Champak Dutta,^a Tanoy Dutta,^b Apurba Lal Koner^{*b} and Joyanta Choudhury^{*a}

^aOrganometallics & Smart Materials Laboratory, Department of Chemistry, Indian Institute of Science Education and Research Bhopal, Bhopal 462 066, India

^bDepartment of Chemistry, Indian Institute of Science Education and Research Bhopal, Bhopal 462 066, India

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1. General Methods and Materials

¹H, ¹³C{¹H}, ¹⁹F and ³¹P{¹H} NMR spectra were recorded on Bruker AVANCE III 400 and 500 MHz NMR spectrometers at room temperature unless mentioned otherwise. Chemical shifts (δ) are expressed in ppm using the residual proton resonance of the solvent as an internal standard (CHCl₃: $\delta = 7.26$ ppm for ¹H spectra, 77.2 ppm for ¹³C{¹H} spectra; CH₃CN: $\delta = 1.94$ ppm for ¹H spectra, 1.3 ppm for ¹³C{¹H} spectra; DMSO: 2.50 ppm for ¹H spectra, 39.5 ppm for ${}^{13}C{}^{1}H$ spectra). All coupling constants (J) are expressed in hertz (Hz) and only given for ¹H–¹H couplings unless mentioned otherwise. The following abbreviations were used to indicate multiplicity: s (singlet), d (doublet), t (triplet), q (quartet), dd (doublet of doublets), dt (doublet of triplets), ddd (doublet of doublet of doublets), m (multiplet). ESI mass spectroscopy was performed on a Bruker microTOF QII spectrometer. Single-crystal Xray diffraction data were collected using a Bruker SMART APEX II CCD diffractometer with graphite monochromated Mo K α ($\lambda = 0.71073$ Å) radiation at low temperatures. Dry solvents and reagents were obtained from commercial suppliers and used without further purification. Deuterated solvents, RhCl₃.xH₂O and other chemicals were purchased from Aldrich. [Cp*RhCl₂]2^{S1} and [Cp*Co(CO)I₂]^{S2} were synthesized according to reported procedure.

2. General Procedure for the Synthesis of Imidazolium Substituted Pyridines/Pyrazines^{S3}



A mixture of 2,6-dibromopyridine or 2,6-dichloropyrazine (5 mmol) and N-alkylimidazole (15 mmol) was stirred in a sealed tube at 120 °C for 30 h (for pyridines) or 5 h (for pyrazines). After completion, THF (10 mL) was added to the mixture and the resulting solid residue was filtered and washed with THF (10 mL x 5) and diethyl ether (20 mL). The solid product was dried under vacuum to give an off-white powder. Next, this halide salt was dissolved in minimum amount of distilled water and an aqueous solution of KPF₆ (20 mmol) was added dropwise with vigorous stirring. After 3 h, the resulting white precipitate was filtered, washed with distilled water (20 mL x 3) and diethyl ether (20 mL x 2) to yield the desired product.

2,6-*Bis*(**1-methylimidazolium**) pyridine hexafluorophosphate (1a): Yield: 81%. ¹H NMR (500 MHz, DMSO-d₆): δ 10.25 (s, 2H), 8.73 (s, 2H), 8.61 (d, *J* = 16.2 Hz, 1H), 8.18 (t, *J* = 8.9 Hz, 2H), 8.09 (s, 2H), 4.01 (s, 6H). HRMS (ESI, positive ion): M²⁺ = 120.5664 (calculated = 120.5658 for [C₁₃H₁₅N₅]²⁺).

2,6-*Bis*(1-methylimidazolium) pyrazine hexafluorophosphate (1b): A mixture of 2,6dichloropyrazine (300 mg, 2.0 mmol) and N-methylimidazole (0.5 mL, 6.1 mmol) was stirred in a sealed tube at 120 °C for 3 h. The work up procedure was similar as mentioned above for synthesis of **1a**. Yield: 82%. ¹H NMR (500 MHz, DMSO-d₆): δ 10.30 (s, 2H), 9.49 (s, 2H), 8.75 (d, *J* = 1.8 Hz, 2H), 8.09 (s, 2H), 4.04 (s, 6H). HRMS (ESI, positive ion): M²⁺ = 121.0625 (calculated = 121.0634 for [C₁₂H₁₄N₆]²⁺).

1,1'-(pyridine-2,6-diyl)bis(3-butyl-1H-imidazol-3-ium) hexafluorophosphate (1c): Yield: 50%. ¹H NMR (400 MHz, CD₃CN): δ 9.60 (s, 2H), 8.41 (t, *J* = 8.1 Hz, 1H), 8.22 (t, *J* = 1.8 Hz, 2H), 7.91 (d, *J* = 8.1 Hz, 2H), 7.69 – 7.63 (m, 2H), 4.30 (t, *J* = 7.3 Hz, 4H), 1.93 (d, *J* = 2.5 Hz, 4H), 1.41 (dd, *J* = 15.1, 7.5 Hz, 4H), 0.98 (t, *J* = 7.4 Hz, 6H). HRMS (ESI, positive ion): M²⁺ = 162.6125 (calculated = 162.6128 for [C₁₉H₂₇N₅]²⁺).

3,3'-(pyridine-2,6-diyl)bis(1-methyl-1H-benzo[*d*]imidazol-3-ium) hexafluorophosphate (1d): Yield: 83%. ¹H NMR (500 MHz, DMSO-d₆): δ 10.63 (s, 2H), 8.78 (t, *J* = 8.1 Hz, 1H), 8.48 (d, *J* = 8.4 Hz, 2H), 8.32 (d, *J* = 8.1 Hz, 2H), 8.24 (d, *J* = 8.3 Hz, 2H), 7.87 (t, *J* = 7.7

Hz, 2H), 7.80 (t, J = 7.8 Hz, 2H), 4.31 (s, 6H). HRMS (ESI, positive ion): $M^{2+} = 170.5804$ (calculated = 170.5815 for $[C_{21}H_{19}N_5]^{2+}$).

1,1'-(pyrazine-2,6-diyl)bis(3-methyl-1H-benzo[d]imidazol-3-ium) hexafluorophosphate (1e): Yield: 70%. ¹H NMR (400 MHz, DMSO-d₆): δ 10.72 (s, 2H), 9.57 (s, 2H), 8.44 (d, J = 8.4 Hz, 2H), 8.24 (d, J = 8.2 Hz, 2H), 7.86 (t, J = 7.7 Hz, 2H), 7.80 (t, J = 7.8 Hz, 2H), 4.30 (s, 6H). HRMS (ESI, positive ion): M²⁺ = 171.0780 (calculated = 171.0791 for [C₂₀H₁₈N₆]²⁺). **1,1'-(pyrazine-2,5-diyl)bis(3-methyl-1H-imidazol-3-ium)** hexafluorophosphate (1f): Yield: 70%. ¹H NMR (500 MHz, DMSO-d₆): δ 10.18 (s, 2H), 9.37 (s, 2H), 8.62 (s, 2H), 8.05 (s, 2H), 4.03 (s, 6H). ¹³C NMR (126 MHz, DMSO-d₆): δ 36.58, 119.42, 125.18, 134.97, 136.34, 143.05. HRMS (ESI, positive ion): M²⁺ = 121.0606 (calculated = 121.0634 for [C₁₂H₁₄N₆]²⁺).

3. Synthesis of 2,6-bis-(benz)imidazolyl pyridines/pyrazines^{S4}



A mixture of CuI (1.04 mmol), Cs_2CO_3 (16.6 mmol), imidazole/benzimidazole (12.6 mmol), and 2,6-dibromopyridine or 2,6-dichloropyrazine (4.2 mmol) in dry DMF (9 mL) was stirred at room temperature for 30 min. in a sealed tube and then allowed to be stirred at 130 °C for 24 h. After completion, it was cooled to room temperature. To it CH_2Cl_2 (40 mL) was added and it was washed with H_2O (20 mL x 2) by solvent extraction procedure. The CH_2Cl_2 layer was collected and concentrated under reduced pressure. The resulting residue was subjected to column chromatography (silica gel, 1-2% MeOH in CHCl₃) to isolate the desired product.

2,6-di(1H-imidazol-1-yl)pyridine (1g): Yield: 85%. ¹H NMR (400 MHz, CDCl₃) δ 8.38 (s, 2H), 7.97 (t, J = 8.0 Hz, 1H), 7.67 (s, 2H), 7.29 (d, J = 8.0 Hz, 2H), 7.23 (s, 2H). HRMS (ESI, positive ion): $[M+H]^+ = 212.0936$ (calculated = 212.0931 for $[C_{11}H_{10}N_5]^+$).

2,6-di(**1H-imidazol-1-yl**)**pyrazine** (**1h**): Yield: 62%. ¹H NMR (500 MHz, CDCl₃) δ 8.72 (s, 2H), 8.42 (s, 2H), 7.71 (t, *J* = 1.4 Hz, 2H), 7.30 (dd, *J* = 1.3, 0.8 Hz, 2H). HRMS (ESI, positive ion): [M+H]⁺ = 213.0863 (calculated = 213.0883 for [C₁₀H₉N₆]⁺).

2,6-bis(1H-benzo[d]imidazol-1-yl)pyridine (**1i):**Yield: 80%. ¹H NMR (500 MHz, DMSOd₆) δ 9.17 (s, 2H), 8.41 – 8.31 (m, 1H), 8.28 (s, 2H), 7.98 (d, *J* = 7.0 Hz, 2H), 7.85 (s, 2H), 7.38 (s, 4H). HRMS (ESI, positive ion): [M+H]⁺ = 312.1249 (calculated = 312.1244 for [C₁₉H₁₄N₅]⁺).

2,5-di(1H-imidazol-1-yl)pyrazine (1j): Yield: 72%. ¹H NMR (500 MHz, CDCl₃) δ 8.63 (s, 2H), 8.38 (bs, 2H), 7.67 (t, *J* = 1.4 Hz, 2H), 7.29 (bs, 2H), 7.26 (s, 2H). HRMS (ESI, positive ion): [M+H]⁺ = 213.0867 (calculated = 213.0883 for [C₁₀H₉N₆]⁺).

4. Optimization of the Reaction Conditions

To an oven dried sealed tube, **1a** (0.05 mmol), catalyst (3 mol%), **2a** (0.11 mmol) and additives were loaded. To this mixture, dry and degassed DCE (2.0 mL) was added and the reaction mixture was left with stirring at 120 °C in dark. After a certain time, the whole reaction mixture was passed through a short celite pad which was thereafter washed with dichloromethane (10 mL x 3). The combined filtrate was concentrated under reduced pressure and the whole crude mixture was subjected to ¹H NMR spectroscopy in DMSO-d₆ for checking the yield by using 1,3,5-trimethoxybenzene as internal standard.

Table S1: Optimization Studies ^a



No	Catalyst (mol%)	Base (equiv.)	Solvent	Oxidant (equiv.)	Temp. (°C)	Time (h)	Yield (%) ^b
1.	[RhCp*Cl ₂] ₂ (3)	NaOAc (10)	DCE	AgOTf (5)	120	24	94
2.	[RhCp*Cl ₂] ₂ (3)	NaOAc (10)	DCE	AgOTf (5)	120	12	93
3.	[RhCp*Cl ₂] ₂ (3)	NaOAc (5)	DCE	AgOTf (2.5)	120	12	52
4.	[RhCp*Cl ₂] ₂ (3)	NaOAc (5)	DCE	AgOTf (5)	120	6	68
5.	[RhCp*Cl ₂] ₂ (3)	NaOAc (5)	DCE	AgOTf (5)	120	12	93
6.	[RhCp*Cl ₂] ₂ (3)	NaOAc (5)	DCE	AgOAc (5)	120	12	Trace
7.	[RhCp*Cl ₂] ₂ (3)	NaOAc (5)	DCE	Cu(OAc) ₂ . H ₂ O (5)	120	12	Mix. (~50)
8.	[RhCp*Cl ₂] ₂ (3)	NaOAc (5)	MeOH	Cu(OAc) ₂ . H ₂ O (5)	120	12	Trace
9.	[RhCp*Cl ₂] ₂ (3)	NaOAc (5)	MeOH	Cu(BF ₄) ₂ (5)	120	12	Trace
10.	[RhCp*Cl ₂] ₂ (3)	NaOAc (5)	DCE	Cu(BF ₄) ₂ (5)	120	12	~30

11.	[RhCp*Cl ₂] ₂ (3)	NaOAc (5)	DCE	AgOTf (5)	30	24	Nd
12.	[RhCp*Cl ₂] ₂ (2)	NaOAc (5)	DCE	AgOTf (5)	120	12	65
13.	[RhCp*Cl ₂] ₂ (3)	KOAc (5)	DCE	AgOTf (5)	120	12	88
14.	[RhCp*Cl ₂] ₂ (3)	NaHCO ₃ (5)	DCE	AgOTf (5)	120	12	40
15.	[RhCp*Cl ₂] ₂ (3)	K ₂ CO ₃ (5)	DCE	AgOTf (5)	120	12	22
16.		NaOAc (5)	DCE	AgOTf (5)	120	12	Nd
17.	[RhCp*Cl ₂] ₂ (3)		DCE	AgOTf (5)	120	12	Nd
18.	[RhCp*Cl ₂] ₂ (3)	NaOAc (5)	DCE		120	12	Nd
19.	[CoCp*COI ₂] (6)	NaOAc (5)	DCE		120	12	Nd

^a Reaction conditions: **1a** (0.05 mmol), **2a** (0.11 mmol), catalyst (0.0015 mmol), additives (as mentioned), DCE (2.0 mL). ^b Crude product was detected by ¹H NMR spectroscopy by using 1,3,5-tri-methoxybenzene as internal standard. Nd = Not detected.

5. General Catalytic Procedure of Annulation Reactions for Scheme 2A, Main Article

To an oven dried sealed tube, **1** (0.1 mmol), NaOAc (0.5 mmol), [RhCp*Cl₂]₂ (0.003 mmol), AgOTf (0.5 mmol) and **2** (0.21 mmol) were loaded. To this mixture, dry and degassed DCE (2.0 mL) was added and the reaction mixture was left with stirring at 120 °C in dark. After 12 h stirring, the whole reaction mixture was passed through a short celite pad and which was washed with dichloromethane (20 mL x 2). The combined filtrate was concentrated under reduced pressure. Final product was separated and purified by alumina column chromatography, using CHCl₃/MeOH (95:5) as eluent.

6. Experimental Characterization Data of the Products (3a-3r) (*vide* Scheme 2B in main article)

3,9-Dimethyl-4,5,7,8-tetraphenyl-3,9-dihydroimidazo[1',2':1,6]pyrido[3,2-g]imidazo[1,2a][1,8]naphthyridium Salt (3a): Yield = 90%. ¹H NMR (500 MHz, DMSO-d₆, 298K): δ



9.80 (d, J = 2.2 Hz, 2H), 8.52 (d, J = 2.2 Hz, 2H), 7.94 (s, 1H), 7.42 (m, J = 5.1, 3.0 Hz, 10H), 7.20 (s, 6H), 7.09 (m, J = 6.5, 3.0 Hz, 4H), 3.39 (s, 6H). ¹³C NMR (126 MHz, DMSO-d₆, 298K): δ 143.9, 141.0, 140.3, 139.3, 132.5, 131.7, 131.3, 130.2, 130.0, 129.2, 129.0, 128.9, 128.6, 125.5, 119.6, 114.8, 38.3. ¹⁹F NMR (471 MHz, DMSO-d₆) δ -70.25 (d, J = 711.6 Hz), -77.76. ³¹P

NMR (202 MHz, DMSO-d₆, 298K): δ -144.3 (hept, J = 711.37 Hz). HRMS (ESI, positive ion): $M^{2+} = 296.6292$ (calculated =296.6284 for $[C_{41}H_{31}N_5]^{2+}$).

3,9-Dimethyl-4,5,7,8-tetra-p-tolyl-3H-imidazo[1',2':1,6]pyrido[3,2-g]imidazo[1,2-



a][1,8]naphthyridium Salt (3b): Yield = 80%. ¹H NMR (500 MHz, DMSO-d₆, 298K): δ 9.75 (d, J = 2.2 Hz, 2H), 8.50 (d, J = 2.2 Hz, 2H), 7.81 (s, 1H), 7.32 (d, J = 8.0 Hz, 4H), 7.24 (d, J = 7.9 Hz, 4H), 7.07 (d, J = 7.9 Hz, 4H), 6.95 (d, J = 8.0 Hz, 4H), 3.39 (s, 6H), 2.31 (s, 6H), 2.24 (s, 6H). ¹³C NMR (126 MHz, DMSO-d₆, 298K): δ 143.6, 140.9, 140.9, 139.7, 139.1, 139.0,

138.0, 130.7, 129.7, 129.7, 129.1, 128.7, 128.4, 124.8, 119.5, 114.2, 37.7, 20.9, 20.6. ¹⁹F NMR (471 MHz, DMSO-d₆, 298K): δ -70.20 (d, J = 711.3 Hz). ³¹P NMR (202 MHz, DMSO-d₆, 298K): δ -144.23 (hept, J = 711.4 Hz). HRMS (ESI, positive ion): M²⁺ = 324.6613 (calculated = 324.6597 for [C₄₅H₃₉N₅]²⁺).

4,5,7,8-Tetrakis(4-methoxyphenyl)-3,9-dimethyl-3,9 dihydroimidazo[1',2':1,6]pyrido[3,2-g]imidazo[1,2-a][1,8]naphthyridine-12,14-diium



Salt (3c): Yield = 63% ¹H NMR (500 MHz, CD₃CN, 298K) δ 9.24 (d, J = 2.3 Hz, 2H), 8.12 (s, 1H), 7.97 (d, J= 2.3 Hz, 2H), 7.38 – 7.27 (m, 4H), 7.10 – 7.03 (m, 4H), 7.02 – 6.98 (m, 4H), 6.90 – 6.80 (m, 4H), 3.83 (s, 6H), 3.78 (s, 6H), 3.45 (s, 6H). ¹³C NMR (126 MHz, CD₃CN, 298K) δ 161.6, 160.9, 146.2, 141.4, 141.0, 140.7, 133.2, 132.5, 129.5 126.1, 125.8, 124.2, 121.3, 115.0, 114.8,

114.7, 56.2, 56.1, 39.2. ¹⁹F NMR (471 MHz, CD₃CN, 298K) δ -72.99 (d, *J* = 706.8 Hz). ³¹P NMR (202 MHz, CD₃CN, 298K) δ -139.44 (d, *J* = 706.7 Hz). HRMS (ESI, positive ion): M²⁺ = 356.6493 (calculated = 356.6496 for [C₄₅H₃₉N₅O₄]²⁺).

3,9-Dimethyl-4,5,7,8-tetrapropyl-3,9-dihydroimidazo[1',2':1,6]pyrido[3,2-g]imidazo[1,2a][1,8]naphthyridium Salt (3d): Yield = 71%. ¹H NMR (500 MHz, DMSO-d₆, 298K): δ



9.57 (d, J = 2.0 Hz, 2H), 9.38 (s, 1H), 8.43 (d, J = 2.1 Hz, 2H), 4.39 (s, 6H), 3.49 – 3.44 (m, 4H), 3.26 – 3.18 (m, 4H), 1.72 (tt, J = 14.7, 7.4 Hz, 8H), 1.18 (dd, J = 16.4, 7.4 Hz, 12H). ¹³C NMR (126 MHz, DMSO-d₆, 298K): δ 143.4, 140.4, 139.6, 137.1, 129.0, 124.9, 118.4, 114.3, 38.8, 29.3, 28.8, 24.7, 24.5, 14.5, 14.1. ¹⁹F NMR (471 MHz, DMSO-d₆,

298K): δ -70.20 (dd, J = 711.3, 9.6 Hz). ³¹P NMR (202 MHz, DMSO-d₆, 298K): δ -144.22 (hept, J = 711.5 Hz). HRMS (ESI, positive ion): M²⁺ = 228.6585 (calculated = 228.6597 for $[C_{29}H_{39}N_5]^{2+}$).

4,5,7,8-Tetraethyl-3,9-dimethylimidazo[1',2':1,6]pyrido[3,2-g]imidazo[1,2-

a][1,8]naphthyridine-3,9-diium Salt (3e): Yield = 50%. ¹H NMR (500 MHz, CD₃CN,



298K) δ 9.33 (s, 1H), 9.14 (d, J = 2.3 Hz, 2H), 7.92 (d, J = 2.3 Hz, 2H), 4.37 (s, 6H), 3.45 (q, J = 7.6 Hz, 4H), 3.31 (q, J = 7.6 Hz, 4H), 1.50 – 1.31 (m, 12H). ¹³C NMR (126 MHz, CD₃CN) δ 146.5, 141.6, 140.6, 137.4, 129.3, 126.9, 119.5, 114.6, 39.6, 21.9, 21.1, 15.6, 15.5. ¹⁹F NMR (376 MHz, CD₃CN) δ -79.31.

HRMS (ESI, positive ion): $M^{2+} = 200.6292$ (calculated = 200.6284 for $[C_{25}H_{31}N_5]^{2+}$).

3,9-Dimethyl-4,8-diphenyl-5,7-di-p-tolyl-3,9-dihydroimidazo[1',2':1,6]pyrido[3,2g]imidazo[1,2-a][1,8]naphthyridine-12,14-diium Salts (3f): Yield = 50% (a mixture of



three regioisomers in **3f/3f'/3f**" = 2/1/1 ratio determined by ¹H NMR spectroscopy). ¹H NMR (400 MHz, DMSO, 298K) δ 9.79 (s, 2H), 8.53 (s, 2H), 7.92 – 7.83 (m, 1H), 7.45 (bs, 6H), 7.35 – 7.30 (m, 2H), 7.23 (d, *J* = 6.0 Hz, 4H), 7.05 (m, 4H), 6.99 – 6.94 (m, 2H), 3.41 (d, *J* = 4.0 Hz, 3H), 3.39 (s, 3H), 2.31 (s, 3H), 2.22 (d, *J* = 8.2 Hz, 3H). ¹³C NMR (126 MHz, DMSO, 298K) δ 144.0, 143.9, 143.9, 141.2, 140.2, 140.2, 139.5, 139.5, 139.4, 138.6, 138.5, 133.1, 133.1, 131.9, 131.3,

131.2, 130.1, 130.1, 130.1, 129.5, 129.2, 129.1, 129.0, 128.9, 128.8, 128.8, 128.7, 128.6, 125.5, 125.4, 125.2, 125.2, 122.4, 119.9, 119.9, 119.8, 114.8, 114.7, 38.4, 38.3, 21.4, 21.1. ³¹P NMR (202 MHz, DMSO, 298K) δ -138.99 (d, J = 711.3 Hz). ¹⁹F NMR (471 MHz, DMSO, 298K) δ -70.29 (d, J = 711.4 Hz), -77.75 (s). HRMS (ESI, positive ion): M²⁺ = 310.6459 (calculated = 310.6441 for [C₄₃H₃₅N₅]²⁺).

3,9-Dimethyl-4,8-diphenyl-5,7-bis(4-(trifluoromethyl)phenyl)-3,9-



 $R^{1} = R^{4} = CF_{3}, R^{2} = R^{3} = H (3g)$ $R^{1} = R^{3} = CF_{3}, R^{2} = R^{4} = H (3g')$ $R^{1} = R^{4} = H, R^{2} = R^{3} = CF_{3} (3g'')$ dihydroimidazo[1',2':1,6]pyrido[3,2-

g]imidazo[1,2-a][1,8]naphthyridine-12,14-

diium Salt (3g): Yield = 53% (a mixture of three regioisomers in **3g/3g'/3g"** = 4/1.5/1 ratio determined by ¹H NMR spectroscopy). ¹H NMR (500 MHz, CD₃CN) δ 9.46 (m, 3H), 8.04 (s, 1H), 8.01 (m, 3H), 7.81 (s, 1H), 7.79 (s, 1H), 7.70 (m, 5H), 7.50 (m, 7H), 7.46 – 7.40 (m, 5H), 7.36 (m, 4H), 7.24 – 7.19 (m, 5H), 7.12 (m, 2H), 3.38 (m, 9H). ¹³C NMR (126 MHz, CD₃CN) δ 146.1, 146.0, 144.0, 143.9, 142.6, 141.9, 141.5, 141.3,

141.3, 140.2, 140.2, 140.0, 138.1, 138.0, 136.8, 133.4, 133.4, 133.0, 132.9, 132.0, 132.0, 131.9, 131.9, 131.7, 131.0, 130.9, 130.8, 129.9, 129.89, 129.7, 129.7, 129.6, 129.6, 129.5, 129.2, 129.2, 129.0, 126.8, 126.6, 126.5, 126.4, 126.4, 126.0, 126.0, 125.9, 125.0, 124.9,

123.9, 123.8, 123.7, 123.3, 121.0, 120.8, 120.7, 120.5, 115.5, 115.4, 39.3, 39.1. HRMS (ESI, positive ion): $M^{2+} = 364.6146$ (calculated = 364.6158 for $[C_{43}H_{29}F_6N_5]^{2+}$).

5,7-Diethyl-3,9-dimethyl-4,8-diphenyl-3H-imidazo[1',2':1,6]pyrido[3,2-g]imidazo[1,2a][1,8]naphthyridine-9,14-diium Salt (3h): Yield = 78%. ¹H NMR (400 MHz, DMSO-d₆, 298K): δ 9.69 (s, 2H), 9.61 (s, 1H), 8.41 (s, 2H), 7.70-7.71 (m, 6H), 7.62 (d, J = 3.8 Hz, 4H),



3.32 (s, 6H), 3.15 (q, J = 6.7 Hz, 4H), 1.21 (t, J = 7.3 Hz, 6H). ¹³C NMR (126 MHz, DMSO-d₆, 298K): δ 145.7, 140.1, 138.9, 137.8, 131.8, 130.2, 130.1, 129.3, 128.1, 124.3, 121.9, 119.4, 117.6, 113.8, 37.4, 21.9, 15.2. ¹⁹F NMR (471 MHz, DMSO-d₆, 300K): δ -77.80 (s). HRMS (ESI, positive ion): M²⁺ = 248.6272

 $(calculated = 248.6284 \text{ for } [C_{33}H_{31}N_5]^{2+}).$

3,9-Dibutyl-4,5,7,8-tetraphenyl-3,9-dihydroimidazo[1',2':1,6]pyrido[3,2-*g*]imidazo[1,2*a*][**1,8]naphthyridine-12,14-diium Salt (3i):** Yield=50%. ¹H NMR (400 MHz, CD₃CN,



298K): δ 9.34 (s, 2H), 8.08 (s, 2H), 8.01 (s, 1H), 7.44 (bs, 10H), 7.29 – 7.18 (m, 6H), 7.10 (d, J = 4.8 Hz, 4H), 3.84 – 3.59 (m, 4H), 1.61 (m, 4H), 0.98 (m, 4H), 0.77 (t, J = 7.3 Hz, 6H). ¹³C NMR (126 MHz, CD₃CN, 298K): δ 146.2, 142.9, 141.3, 140.0, 133.6, 132.4, 131.6, 130.8, 129.8, 129.2, 128.1, 126.1, 120.9, 115.6, 50.9, 33.5, 20.3, 13.7. ¹⁹F

NMR (471 MHz, CD₃CN, 298K): δ -72.98 (d, J = 706.7 Hz). HRMS (ESI, positive ion): M²⁺ = 338.6751 (calculated = 338.6754 for [C₄₇H₄₃N₅]²⁺).

3,9-Dimethyl-4,5,7,8-tetraphenyl-3H-imidazo[1',2':1,6]pyrido[2,3-

b]imidazo[1',2':1,6]pyrido[3,2-*e*]pyrazine-9,14-diium Salt (3j): Yield = 68%. ¹H NMR



(500 MHz, CD₃CN, 298K): δ 9.28 (d, J = 2.3 Hz, 2H), 8.02 (d, J = 2.3 Hz, 2H), 7.49 – 7.44 (m, 5H), 7.45 – 7.39 (m, 5H), 7.17 – 7.13 (m, 2H), 7.12 – 7.08 (m, 4H), 7.07 – 7.03 (m, 4H), 3.40 (s, 6H). ¹³C NMR (126 MHz, CD₃CN, 298K): δ 146.3, 140.0, 137.9, 136.2, 132.7, 132.1, 131.8, 131.7, 130.9, 130.4, 130.2, 129.7, 129.2, 128.4, 115.9, 39.4. ¹⁹F

NMR (471 MHz, CD₃CN, 298K): δ -73.01 (d, J = 706.6 Hz). ³¹P NMR (202 MHz, CD₃CN, 298K): δ -144.68 (hept, J = 706.6 Hz). HRMS (ESI, positive ion): M²⁺ = 297.1269 (calculated = 297.1260 for [C₄₀H₃₀N₆]²⁺).

3,9-Dimethyl-4,5,7,8-tetra-p-tolyl-3H-imidazo[1',2':1,6]pyrido[2,3

b]imidazo[1',2':1,6]pyrido[3,2-*e*]pyrazine-9,14-diium Salt (3k): Yield = 51%. ¹H NMR (400 MHz, CD₃CN, 298K): δ 9.21 (s, 2H), 7.98 (s, 2H), 7.29 (s, 8H), 6.95 (s, 8H), 3.38 (s,



6H), 2.37 (s, 6H), 2.27 (s, 6H). ¹³C NMR (126 MHz, DMSO-d₆, 298K): δ 146.4, 141.3, 140.2, 139.1, 138.0, 136.0, 131.7, 131.6, 130.3, 130.1, 130.0, 129.1, 129.0, 118.6, 115.6, 39.3, 21.3, 21.3. ¹⁹F NMR (471 MHz, DMSO-d₆, 298K): δ -72.44 (d, J = 706.6 Hz). ³¹P NMR (202 MHz, DMSO-d₆, 298K): δ -144.11 (hept, J = 706.6 Hz). HRMS

(ESI, positive ion): $M^{2+} = 325.1586$ (calculated = 325.1573 for $[C_{44}H_{38}N_6]^{2+}$).

3,9-Dimethyl-4,5,7,8-tetrapropyl-3H-imidazo[1',2':1,6]pyrido[2,3-

b]imidazo[1',2':1,6]pyrido[3,2-*e*]pyrazine-9,14-diium Salt (31): Yield = 67%. ¹H NMR



(500 MHz, CD₃CN, 298K): δ 9.01 (d, J = 2.3 Hz, 2H), 7.92 (d, J = 2.3 Hz, 2H), 4.36 (s, 6H), 3.51 – 3.43 (m, 4H), 3.32 – 3.25 (m, 4H), 1.83 – 1.74 (m, 8H), 1.20 (td, J = 7.3, 1.5 Hz, 12H). ¹³C NMR (126 MHz, CD₃CN, 298K): δ 146.6, 141.3, 137.0, 135.4, 130.2, 130.1, 115.1, 39.8, 30.2, 30.1, 25.1, 24.9, 14.9,

13.9. ¹⁹F NMR (471 MHz, CD₃CN, 298K): δ -72.97 (d, J = 706.6 Hz). ³¹P NMR (202 MHz, CD₃CN, 298K): δ -144.66 (hept, J = 706.6 Hz). HRMS (ESI, positive ion): M²⁺ = 229.1579 (calculated = 229.1573 for [C₂₈H₃₈N₆]²⁺).

5,11-Dimethyl-6,7,9,10-tetraphenyl-5H-benzo[4',5']imidazo[1',2':1,6]pyrido[3,2g]benzo[4,5]imidazo[1,2-a][1,8]naphthyridine-11,18-diium Salt (3m): Yield = 75% . ¹H NMR (500 MHz, DMSO-d₆, 298K): δ 9.67 (d, J = 8.3 Hz, 2H), 8.42 (d, J = 8.4 Hz, 2H), 8.24



(t, J = 7.6 Hz, 2H), 8.17 (t, J = 7.8 Hz, 2H), 8.10 (s, 1H), 7.48 (s, 10H), 7.29 – 7.22 (m, 6H), 7.17 – 7.10 (m, 4H), 3.60 (s, 6H). ¹³C NMR (126 MHz, DMSO-d₆, 298K): δ 148.7, 144.3, 143.2, 141.0, 134.3, 132.8, 132.0, 131.4, 130.2, 129.9, 129.1, 128.6, 128.5, 127.2, 125.6, 122.3, 119.8, 119.5, 117.2, 114.7, 34.9. ¹⁹F NMR (471 MHz, DMSO-d₆, 298K): δ -77.75 (s). HRMS (ESI, positive

ion): $M^{2+} = 346.6468$ (calculated = 346.6441 for $[C_{49}H_{35}N_5]^{2+}$).

5,11-Dimethyl-6,7,9,10-tetrapropyl-5H-benzo[4',5']imidazo[1',2':1,6]pyrido[3,2g]benzo[4,5]imidazo[1,2-a][1,8]naphthyridine-11,18-diium Salt (3n): Yield = 76%. ¹H



NMR (500 MHz, DMSO-d₆, 298K): δ 9.60 (s, 1H), 9.47 (d, J = 8.1 Hz, 2H), 8.49 (d, J = 8.1 Hz, 2H), 8.12 – 7.99 (m, 4H), 4.59 (s, 6H), 3.66 – 3.58 (m, 4H), 3.44 – 3.38 (m, 4H), 1.87 – 1.75 (m, 8H), 1.22 (q, J = 6.9 Hz, 12H). ¹³C NMR (126 MHz, DMSO-d₆, 298K): δ 148.9, 145.0, 142.1, 136.5, 134.0, 129.0, 127.1, 126.7, 124.5, 121.9, 119.4, 117.6, 117.0, 114.0,

34.6, 29.6, 28.8, 24.4, 24.0, 14.1, 13.6. ¹⁹F NMR (471 MHz, DMSO-d₆, 298K): δ -77.75 (s). HRMS (ESI, positive ion): M²⁺ = 278.6776 (calculated = 278.6754 for [C₃₇H₄₃N₅]²⁺).

5,11-Dimethyl-6,7,9,10-tetraphenyl-5,11

dihydrobenzo[4',5']imidazo[1',2':1,6]pyrido[2,3-



b]benzo[4',5']imidazo[1',2':1,6]pyrido[3,2-e/pyrazine-

16,18-diium Salt (30): Yield = 69% ¹H NMR (500 MHz, CD₃CN, 298K) δ 9.65 (d, J = 8.5 Hz, 2H), 8.26 (ddd, J = 8.4, 7.0, 1.3 Hz, 2H), 8.22 – 8.13 (m, 4H), 7.62 – 7.48 (m, 10H), 7.26 – 7.21 (m, 2H), 7.19 – 7.11 (m, 8H), 3.62 (s, 6H). ¹³C NMR (126 MHz, CD₃CN, 298K) δ 150.9, 144.4, 139.0,

136.9, 135.4, 132.8, 132.4, 132.0, 131.5, 131.2, 131.0, 130.1, 130.1, 130.0, 129.5, 128.5, 128.4, 115.2, 35.9. ³¹P NMR (202 MHz, CD₃CN, 298K) δ -146.47 (d, *J* = 706.6 Hz). ¹⁹F NMR (471 MHz, CD₃CN, 298K) δ -72.33 (d, *J* = 2.3 Hz), -73.08 (dd, *J* = 706.6, 2.3 Hz), -73.83 (d, *J* = 2.3 Hz). HRMS (ESI, positive ion): M²⁺ = 347.1410 (calculated = 347.1417 for [C₄₈H₃₄N₆]²⁺).

5,11-Dimethyl-6,7,9,10-tetrapropyl-5,11

dihydrobenzo[4',5']imidazo[1',2':1,6]pyrido[2,3-



b]benzo[4',5']imidazo[1',2':1,6]pyrido[3,2-*e*]pyrazine-16,18diium Salt (3p): Yield = 70%. ¹H NMR (500 MHz, CD₃CN, 298K) δ 9.46 (dd, *J* = 7.1, 1.2 Hz, 2H), 8.26 (dd, *J* = 7.4, 1.6 Hz, 2H), 8.11 (pd, *J* = 7.4, 1.3 Hz, 4H), 4.59 (s, 6H), 3.67 – 3.60 (m, 4H), 3.49 (dd, *J* = 9.7, 7.3 Hz, 4H), 1.91 (m, 8H), 1.29 (td, *J* = 7.3, 1.7 Hz, 12H). ¹³C NMR (126 MHz, CD₃CN, 298K) δ 151.9, 145.7, 138.2, 135.9, 135.2, 130.5, 130.0, 129.4, 128.4, 117.8, 115.0, 36.0, 30.9, 30.6, 25.1, 25.0, 15.0, 13.9. ¹⁹F NMR (376 MHz, CD₃CN, 298K) δ -73.07 (d, *J* = 706.7 Hz), -79.37 (s). HRMS (ESI, positive ion): M²⁺ = 279.1727 (calculated = 279.1730 for [C₃₆H₄₂N₆]²⁺).

3,10-Dimethyl-4,5,11,12-tetraphenylimidazo[1',2':1,6]pyrido[2,3

b]imidazo[1',2':1,6]pyrido[2,3-e]pyrazine-3,10-diium salt (3q): Yield = 67% ¹H NMR



(400 MHz, DMSO-d₆) δ 8.41 (s, 2H), 8.36 (s, 2H), 7.48 (s, 8H), 7.43 (s, 8H), 7.33 (bs, 4H), 3.40 (s, 6H). ¹³C NMR (126 MHz, DMSO-d₆) δ 143.5, 138.5, 136.9, 135.2, 132.5, 131.6, 131.1, 130.3, 130.2, 129.9, 129.1, 129.0, 128.8, 128.3, 122.4, 121.9, 119.8, 114.2, 38.5. ¹⁹F NMR (376 MHz, DMSO-d₆) δ -70.29 (d, J = 711.3 Hz), -77.77 (s). HRMS (ESI, positive ion): M²⁺ =

297.1271 (calculated = 297.1260 for $[C_{40}H_{30}N_6]^{2+}$).

3,10-Dimethyl-4,5,11,12-tetrapropylimidazo[1',2':1,6]pyrido[2,3-

b]imidazo[1',2':1,6]pyrido[2,3-*e*]pyrazine-3,10-diium (3r): Yield = $65\%^{-1}$ H NMR (500)



MHz, CD₃CN) δ 8.93 (d, J = 2.3 Hz, 2H), 7.94 (d, J = 2.3 Hz, 2H), 4.37 (s, 6H), 3.54 – 3.43 (m, 4H), 3.34 – 3.24 (m, 4H), 1.87 – 1.71 (m, 8H), 1.23 – 1.13 (m, 12H). ¹³C NMR (126 MHz, CD₃CN) δ 145.6, 140.7, 137.1, 135.6, 131.9, 129.9, 114.9, 39.8, 30.3, 29.6, 25.1, 24.7, 14.6, 14.0. ¹⁹F NMR (376 MHz, CD₃CN)

 δ -79.30. HRMS (ESI, positive ion): M^{2+} = 229.1580 (calculated = 229.1573 for $[C_{28}H_{38}N_6]^{2+}).$

7. General Catalytic Procedure of Annulation Reactions for Scheme 2B, Main Article

In an oven dried sealed tube **1g-1i** (0.1 mmol), [RhCp*Cl₂]₂ (3 mol%), **2a** (0.21 mmol) and Cu(OAc)₂.H₂O (0.3 mmol) were taken .To this 2 mL of toluene was added and stirred at 140 °C for 12 h. After the reaction time the mixture was cooled to room temperature and CH₂Cl₂ (15 mL) was added to this mixture followed by washing with saturated K₂CO₃ solution The organic layer was collected and the extraction was repeated by CH₂Cl₂ (10 mL x 2) and the organic layers were combined and evaporated to dryness. Final product was separated and purified by alumina column chromatography, using CHCl₃ as eluent.

8. Experimental Characterization Data of the Products (3s-3af) (*vide* Scheme 2B in main article)

4,5,7,8-Detraphenylimidazo[1',2':1,6]pyrido[3,2-g]imidazo[1,2-*a*][1,8]naphthyridine (3s): Yield = 78%. ¹H NMR (500 MHz, CDCl₃, 298K) δ 8.73 (d, *J* = 1.4 Hz, 2H), 8.13 (s,



1H), 7.83 (d, J = 1.3 Hz, 2H), 7.36 – 7.30 (m, 4H), 7.30 – 7.27 (m, 6H), 7.18 – 7.15 (m, 6H), 7.05 – 7.03 (m, 4H). ¹³C NMR (126 MHz, CDCl₃, 298K) δ 145.3, 140.8, 138.4, 135.2, 134.8, 134.2, 133.4, 130.8, 130.7, 129.3, 128.0, 127.9, 127.8, 127.5,

117.6, 112.7. HRMS (ESI, positive ion): $[M+H]^+ = 564.2163$ (calculated = 564.2183 for $[C_{39}H_{26}N_5]^+$).



Molecular structure of product 3s (50% probability level).

4,5,7,8-Tetrapropylimidazo[1',2':1,6]pyrido[3,2-g]imidazo[1,2-*a***][1,8]naphthyridine (3t):** Yield = 72%. ¹H NMR (500 MHz, CDCl₃): δ 8.60 (s, 1H), 8.39 (bs, 2H), 7.61 (d, *J* = 1.1 Hz,



2H), 3.12 - 3.07 (m, 4H), 3.05 - 2.99 (m, 4H), 1.80 (dq, J = 15.1, 7.4 Hz, 4H), 1.72 (dq, J = 15.0, 7.4 Hz, 4H), 1.16 (t, J = 7.3 Hz, 6H), 1.12 (t, J = 7.3 Hz, 6H). ¹³C NMR (126 MHz, CDCl₃): δ 14.7, 14.7, 23.4, 24.1, 30.2, 30.4, 112.2, 116.5, 128.9, 131.3,

131.8, 132.2, 140.2, 145.7. HRMS (ESI, positive ion): $[M+H]^+ = 428.2821$ (calculated = 428.2809 for $[C_{27}H_{34}N_5]^+$).

4,5,7,8-Tetraphenylimidazo[1',2':1,6]pyrido[2,3-b]imidazo[1',2':1,6]pyrido[3,2-



e]pyrazine (3u): Yield = 78%. ¹H NMR (500 MHz, CDCl₃, 298K) δ 8.68 (d, J = 1.4 Hz, 2H), 7.80 (d, J = 1.4 Hz, 2H), 7.39 (dt, J = 3.9, 2.1 Hz, 4H), 7.37 – 7.22 (m, 6H), 7.16 – 7.07 (m, 2H), 7.03 (bs, 4H), 7.02 (bs, 4H). ¹³C NMR (126 MHz, CDCl₃,

298K) δ 145.0, 135.5, 135.5, 134.9, 134.7, 134.1, 134.1, 133.1, 131.1, 130.9, 128.1, 128.0, 127.3, 126.9, 113.5. HRMS (ESI, positive ion): $[M+H]^+ = 565.2110$ (calculated = 565.2135 for $[C_{38}H_{25}N_6]^+$).

6,7,9,10-Tetraphenylbenzo[4',5']imidazo[1',2':1,6]pyrido[3,2-g]benzo[4,5]imidazo[1,2a][1,8]naphthyridine (3v): Yield = 80%. ¹H NMR (500 MHz, CDCl₃, 298K) δ 9.60 (d, J =



8.0 Hz, 2H), 8.11 (d, J = 7.7 Hz, 2H), 8.05 (s, 1H), 7.74 – 7.69 (m, 2H), 7.68 – 7.63 (m, 2H), 7.44 – 7.37 (m, 4H), 7.34 – 7.26 (m, 6H), 7.21 – 7.14 (m, 6H), 7.06 (dd, J = 6.4, 3.2 Hz, 4H). 13C NMR (126 MHz, CDCl₃) δ 149.5, 144.8, 144.2,138.8, 137.5, 135.2, 134.8, 130.9, 130.8, 130.4, 129.3, 128.0, 128.0, 127.9,

127.7, 125.5, 123.6, 120.9, 116.7, 116.1. HRMS (ESI, positive ion): $[M+H]^+ = 664.2486$ (calculated = 664.2496 for $[C_{47}H_{30}N_5]^+$).



Molecular structure of product 3v (50% probability level).

6,7,9,10-Tetrakis(4-bromophenyl)benzo[4',5']imidazo[1',2':1,6]pyrido[3,2g]benzo[4,5]imidazo[1,2-a][1,8]naphthyridine(3w): Yield = 61%. ¹H NMR (500 MHz,



CDCl₃) δ 9.54 (d, J = 8.0 Hz, 2H), 8.15 (d, J = 8.0 Hz, 2H), 7.87 (s, 1H), 7.78 – 7.70 (m, 2H), 7.72 – 7.64 (m, 2H), 7.49 (d, J = 8.4 Hz, 4H), 7.43 (d, J = 8.3 Hz, 4H), 7.30 (d, J = 8.4 Hz, 4H), 6.93 (d, J = 8.3 Hz, 4H).¹³C NMR (176 MHz, CDCl₃) δ 147.6, 144.1, 137.6, 135.1, 133.2, 133.0, 132.5, 132.0, 132.0, 131.9, 131.3, 130.9,

130.2, 126.9, 125.0, 123.5, 123.2, 120.7, 117.2, 116.1. HRMS (ESI, positive ion): $[M+H]^+ =$ 979.8891 (calculated = 979.8875 for $[C_{47}H_{26}Br_4N_5]^+$).

6,7,9,10-Tetrakis(4-iodophenyl)benzo[4',5']imidazo[1',2':1,6]pyrido[3,2-

g]benzo[4,5]imidazo[1,2-a][1,8]naphthyridine (3x): Yield = 63%. ¹H NMR (500 MHz,



CDCl₃) δ 9.52 (d, J = 8.1 Hz, 2H), 8.15 (t, J = 6.3 Hz, 2H), 7.91 (s, 1H), 7.73 – 7.61 (m, 6H), 7.16 (d, J = 8.4 Hz, 2H), 6.80 (d, J = 8.3 Hz, 2H). ¹³C NMR (176 MHz, CDCl₃) δ 148.2, 144.2, 137.8, 137.7, 137.2, 134.0, 133.3, 132.7, 132.0, 130.8, 130.4, 128.9, 126.4, 124.5, 120.9, 116.7, 116.5, 116.0, 95.2, 94.6.HRMS (ESI, positive ion): [M+H]⁺ = 1167.8340 (calculated = 1167.8361 for [C₄₇H₂₆I₄N₅]⁺).

4,4',4'',4'''-(Benzo[4',5']imidazo[1',2':1,6]pyrido[3,2-g]benzo[4,5]imidazo[1,2*a*][**1,8]naphthyridine-6,7,9,10-tetrayl)tetrabenzonitrile (3y):** Yield = 44%. ¹H NMR (500



MHz, CDCl₃) δ 9.81 (s, 1H), 9.54 (d, J = 8.2 Hz, 1H), 9.08 (d, J = 7.9 Hz, 1H), 8.30 (d, J = 7.1 Hz, 1H), 8.21 – 8.01 (m, 2H), 7.81 – 7.77 (m, 1H), 7.74 (m, 3H), 7.65 (dd, J = 8.3, 6.1 Hz, 3H), 7.62 – 7.57 (m, 3H), 7.51 (dd, J = 8.2, 5.9 Hz, 2H), 7.40 (d, J = 8.3 Hz, 2H), 7.22 (d, J = 8.3 Hz, 2H). ¹³C NMR (126 MHz, CDCl₃) δ 148.11, 147.97, 147.44, 145.70, 145.02, 144.81, 144.46, 140.30,

139.78, 139.53, 138.96, 138.92, 137.24, 137.06, 136.33, 133.04, 132.61, 132.59, 131.99, 131.77, 131.55, 130.88, 130.77, 129.63, 129.01, 126.84, 126.65, 126.21, 125.30, 125.17, 121.58, 121.16, 118.68, 118.63, 118.22, 117.66, 116.93, 116.40, 116.04, 113.57, 113.49,

113.40, 113.06, 112.92.HRMS (ESI, positive ion): $[M+H]^+ = 764.2345$ (calculated = 764.2306 for $[C_{51}H_{26}N_9]^+$).

6,7,9,10-Tetra(pyridin-3-yl)benzo[4',5']imidazo[1',2':1,6]pyrido[3,2-

g]benzo[4,5]imidazo[1,2-a][1,8]naphthyridine (3z): Yield = 72%. ¹H NMR (500 MHz,



CDCl₃) δ 9.55 (d, J = 8.1 Hz, 2H), 8.64 – 8.48 (m, 6H), 8.35 (d, 2H), 8.11 (d, J = 8.0 Hz, 2H), 7.90 (d, J = 6.2Hz, 2H), 7.84 (s, 1H), 7.76 (dd, J = 11.3, 4.1 Hz, 2H), 7.74 – 7.68 (m, 2H), 7.49 (m, 2H), 7.38 (dd, J = 7.7, 5.1 Hz, 2H) 7.24 (m, 2H). ¹³C NMR (126 MHz, CDCl₃) δ 151.1, 151.0, 150.8, 149.9, 149.8, 149.1, 148.5, 144.8,

139.0, 138.8, 138.0, 137.7, 136.2, 135.6, 130.8, 130.6, 130.5, 127.5, 126.3, 124.6, 123.5, 123.4, 122.9, 121.3, 116.3, 116.2.HRMS (ESI, positive ion): $[M+H]^+ = 668.2300$ (calculated = 668.2306 for $[C_{43}H_{26}N_9]^+$).

6,7,9,10-Tetrapropylbenzo[4',5']imidazo[1',2':1,6]pyrido[3,2-g]benzo[4,5]imidazo[1,2*a*][1,8]naphthyridine (3aa): Yield = 74%. ¹H NMR (500 MHz, CDCl₃): δ 9.49 – 9.35 (m,



2H), 8.76 (s, 1H), 8.13 – 8.05 (m, 2H), 7.66 – 7.56 (m, 4H), 3.31 – 3.21 (m, 4H), 3.16 (dd, J = 9.4, 6.9 Hz, 4H), 1.95 – 1.87 (m, 4H), 1.82 (dt, J = 14.9, 7.6 Hz, 4H), 1.23 (t, J = 7.3 Hz, 6H), 1.17 (t, J = 7.3 Hz, 6H). ¹³C NMR (126 MHz, CDCl₃): δ 149.3, 142.9, 135.5, 130.3, 128.3, 124.6, 122.4, 119.5, 115.48, 115.0, 77.2, 30.0, 29.9, 23.4, 22.6, 14.2, 14.0. HRMS (ESI, positive

ion): $[M+H]^+ = 528.3122$ (calculated = 528.3122 for $[C_{35}H_{38}N_5]^+$).

6,10-Diphenyl-7,9-di-p-tolylbenzo[4',5']imidazo[1',2':1,6]pyrido[3,2-

g]benzo[4,5]imidazo[1,2-*a*][1,8]naphthyridine(3ab): Yield = 63% (a mixture of three regioisomers in 3ab/3ab'/3ab'' = 2/1/1 ratio determined by ¹H NMR spectroscopy). ¹H NMR



(500 MHz, CDCl₃) δ 9.55 (d, J = 8.0 Hz, 2H), 8.12 (d, J = 7.8 Hz, 2H), 7.99 (s, 1H), 7.71 – 7.55 (m, 4H), 7.47 – 7.38 (m, 2H), 7.31 (m, 5H), 7.24 – 7.16 (m, 3H), 7.11 (dd, J = 8.0, 2.3 Hz, 2H), 7.06 (m, 2H), 6.98 (t, J = 8.5 Hz, 2H), 6.95 – 6.89 (m, 2H), 2.32 (s, 6H). ¹³C NMR (126 MHz, CDCl₃) δ 149.4, 149.2, 144.1, 144.0, 139.6, 138.9, 138.2, 137.9, 137.9, 137.8, 137.6, 137.6, 137.5, 135.4, 135.3, 134.7, 132.1, 132.1, 131.6, 131.0, 130.9, 130.7, 130.6, 130.5, 130.4, 130.4,

129.2, 128.9, 128.9, 128.6, 128.1, 127.7, 127.6, 125.7, 125.7, 123.8, 123.7, 120.8, 120.7, 117.2, 117.1, 117.0, 116.9, 116.1, 21.5, 21.3. HRMS (ESI, positive ion): $[M+H]^+ = 692.2805$ (calculated = 692.2809 for $[C_{49}H_{34}N_5]^+$).



Molecular structure of product **3ab** (50% probability level).

7,9-Diphenyl-6,10-bis(4

(trifluoromethyl)phenyl)benzo[4',5']imidazo[1',2':1,6]pyrido[3,2-

g]benzo[4,5]imidazo[1,2-a][1,8]naphthyridine (3ac): Yield = 65% (a mixture of three



 $\begin{aligned} & \mathsf{R}^1 = \mathsf{R}^4 = \mathsf{CF}_3, \, \mathsf{R}^2 = \mathsf{R}^3 = \mathsf{H} \; \textbf{(3ac)} \\ & \mathsf{R}^1 = \mathsf{R}^3 = \mathsf{CF}_3, \, \mathsf{R}^2 = \mathsf{R}^4 = \mathsf{H} \; \textbf{(3ac')} \\ & \mathsf{R}^1 = \mathsf{R}^4 = \mathsf{H}, \, \mathsf{R}^2 = \mathsf{R}^3 = \mathsf{CF}_3 \; \textbf{(3ac'')} \end{aligned}$

regioisomers in **3ac/3ac'/3ac"** = 4/1/1.2 ratio determined by ¹H NMR spectroscopy). ¹H NMR (500 MHz, CDCl₃) δ 9.54 (d, *J* = 8.0 Hz, 4H), 8.11 (dd, *J* = 12.3, 8.1 Hz, 4H), 8.04 (s, 1H), 7.82 (s, 1H), 7.78 (s, 1H), 7.72 – 7.60 (m, 8H), 7.58 – 7.51 (m, 10H), 7.46 (d, *J* = 8.1 Hz, 2H), 7.43 – 7.31 (m, 8H), 7.25 – 7.17 (m, 10H), 7.08 – 6.97 (m, 6H). ¹³C NMR (126 MHz, CDCl₃) δ 148.7, 144.5, 144.5, 144.4, 144.35, 144.2, 144.2, 139.6, 139.5, 139.1, 138.9, 138.5, 138.3, 138.1, 137.4, 136.3,

134.5, 134.5, 133.9, 131.5, 131.5, 131.0, 130.9, 130.9, 130.7, 130.6, 130.6, 130.4, 130.3, 130.2, 130.1, 130.0, 130.0, 129.9, 129.7, 128.7, 128.5, 128.5, 128.4, 128.0, 127.9, 126.0, 125.9, 125.3, 125.3, 125.2, 125.2, 125.1, 124.2, 124.1, 124.1, 123.1, 122.8, 121.0, 121.0, 120.9, 116.9, 116.7, 116.5, 116.1, 117.0. HRMS (ESI, positive ion): $[M+H]^+ = 800.2256$ (calculated = 800.2243 for $[C_{49}H_{28}F_6N_5]^+$).

7,9-Diethyl-6,10-diphenylbenzo[4',5']imidazo[1',2':1,6]pyrido[3,2-



g]benzo[4,5]imidazo[1,2-*a*][1,8]naphthyridine (3ad): Yield = 70% (a mixture of three regioisomers in 1.2/1 ratio determined by ¹H NMR spectroscopy). ¹H NMR (500 MHz, CDCl₃) δ 9.58 – 9.30 (m, 4H), 8.95 (s, 1H), 8.21 (s, 1H), 8.18 – 8.13 (m, 1H), 8.08 – 8.01 (m, 2H), 7.68 – 7.62 (m, 10H), 7.59 (m, 10H), 7.51 – 7.43 (m, 4H), 7.42 – 7.37 (m, 1H), 7.22 – 7.17 (m, 1H), 3.04 (m, 6H), 2.66 (m, 2H), 1.39 (m, 6H), 1.05 (t, *J* = 7.5 Hz, 3H). ¹³C NMR (126 MHz, CDCl₃) δ 149.8, 149.4, 144.6, 144.6, 144.2,

143.4, 139.4, 139.2, 135.8, 135.6, 135.6, 134.4, 132.3, 130.7, 130.7, 129.9, 129.7, 129.8, 129.6, 129.5, 129.3, 129.1, 129.0, 128.9, 128.8, 128.6, 128.6, 128.5, 128.2, 125.6, 125.3, 125.3, 123.5, 123.4, 123.3, 120.8, 120.7, 120.2, 117.2, 116.2, 116.1, 116.0, 115.9, 115.2, 77.4, 77.2, 76.9, 22.9, 22.7, 22.5, 15.4, 14.8, 14.5. HRMS (ESI, positive ion): $[M+H]^+ = 568.2502$ (calculated = 568.2496 for $[C_{39}H_{30}N_5]^+$).

4,5,11,12-Tetraphenylimidazo[1',2':1,6]pyrido[2,3-b]imidazo[1',2':1,6]pyrido[2,3-

e]pyrazine (3ae): Yield = 84%.¹H NMR (500 MHz, CDCl₃) δ 8.24 (d, J = 1.4 Hz, 2H), 7.67



(d, J = 1.4 Hz, 2H), 7.45 – 7.41 (m, 4H), 7.38 – 7.31 (m, 16H). ¹³C NMR (126 MHz, CDCl₃) δ 144.3, 136.9, 135.3, 134.7, 134.7, 134.4, 133.9, 133.8, 132.0, 131.0, 128.4, 128.1, 127.8, 127.7, 113.7. HRMS (ESI, positive ion): [M+H]⁺ = 565.2150 (calculated = 565.2135 for [C₃₈H₂₅N₆]⁺).

4,5,11,12-Tetrapropylimidazo[1',2':1,6]pyrido[2,3-*b*]imidazo[1',2':1,6]pyrido[2,3-

e]pyrazine (3af): Yield = 80%. ¹H NMR (500 MHz, CDCl₃) δ 8.46 (d, J = 1.2 Hz, 2H), 7.67



(d, J = 1.2 Hz, 2H), 3.28 - 3.21 (m, 4H), 3.17 (dd, J = 9.2, 6.9 Hz, 4H), 1.85 (dd, J = 15.6, 7.6 Hz, 4H), 1.79 (dd, J = 15.3, 7.5 Hz, 4H), 1.13 (tt, J = 12.3, 6.2 Hz, 12H).¹³C NMR (126 MHz, CDCl₃) δ 145.2, 136.6, 135.1, 134.1, 133.7, 132.8, 112.9, 31.2, 29.2, 24.1, 23.6, 15.1, 15.0. HRMS (ESI, positive ion): [M+H]⁺

= 429.2778 (calculated = 429.2761 for $[C_{26}H_{33}N_6]^+$).

9. Mechanistic Studies

A. Synthesis of the 1st Rollover Intermediate 4:



(a): [Cp*RhCl₂]₂ (x equiv.), NaOAc (10 equiv), DCE, reflux, 24 h isolated <u>yield of</u> 4: 63% (x = 1.0 equiv.); 30% (x = 0.5 equiv.)

To an oven dried Schlenk tube, a mixture of **1a** (27 mg, 0.05 mmol), NaOAc (41 mg, 0.5 mmol) and [Cp*RhCl₂]₂ (1.0 equiv. or 0.5 equiv.) were loaded and then the tube was kept under vacuum for 15 minutes. After that the tube was filled with N₂ gas. To this mixture, dry and degassed DCE (5 mL) was added under Schlenk technique. After 24 h of reflux, the reaction mixture was allowed to cool down to room temperature and the solution was passed through a short celite pad followed by washing with dichloromethane (3×10 mL). The combined filtrate was concentrated under reduced pressure. Next complex was purified by flash alumina column using CH₂Cl₂/MeOH (98:2) as eluent. Finally, reprecipitation using CH₂Cl₂/n-pentane gives the desired complex **4** (Yield = 63% or 30%) as orange solid after drying under reduced pressure.

¹H NMR (500 MHz, CD₃CN, 298K): δ 9.36 (d, J = 2.0 Hz, 1H), 8.37 (d, J = 7.8 Hz, 1H), 7.70 (d, J = 2.0 Hz, 1H), 7.40 (m, 2H), 7.22 (d, J = 2.0 Hz, 1H), 4.00 (s, 3H), 3.91 (s, 3H), 1.78 (s, 15H), 1.41 (s, 15H). ³¹P NMR (202 MHz, CD₃CN, 298K): δ -139.40 (d, J = 706.4Hz). ¹⁹F NMR (471 MHz, CD₃CN, 298K): δ -72.94 (d, J = 706.5 Hz). HRMS (ESI, positive ion): M⁺ = 784.0916 (calculated = 784.0927 for [C₃₃H₄₂Cl₂N₅Rh₂]⁺).



Molecular structure of intermediate 4 (50% probability level).

B. Synthesis of the 2nd Rollover Intermediate 5:



To an oven dried Schlenk tube, a mixture of **4** (47 mg, 0.05 mmol), NaOAc(41 mg, 0.50 mmol) and AgPF₆ (64 mg, 0.25 mmol) were loaded and then the tube was kept under vacuum for 15 minutes. After that the tube was filled with N₂ gas. To this mixture, dry and degassed CH₃CN (2 mL) was added under Schlenk technique. After 12 h of heating, the reaction mixture was allowed to cool down to room temperature and the solution was passed through a short celite pad followed by washing with acetonitrile (3×5 mL). The combined filtrate was concentrated under reduced pressure and the solid formed was re-dissolved in minimum quantity of CH₃CN. To this solution diethyl ether (~5 times) was added and again all volatiles were evaporated. The resulting yellow solid was washed with cold pentane to afford the desired complex **5** (Yield= 71%) after drying under reduced pressure.

¹H NMR (500 MHz, CD₃CN, 298K) δ 8.39 (s, 1H), 7.87 (d, J = 2.0 Hz, 2H), 7.38 (d, J = 2.0 Hz, 2H), 4.05 (s, 6H), 1.79 (s, 30H). ¹³C NMR (126 MHz, CD₃CN, 298K): δ 177.9 (d, J = 53.4 Hz),156.5, 155.3, 147.3 (d, J = 34.1 Hz),126.1, 117.8, 101.26 (d, J = 5.4 Hz), 39.1, 11.0 ³¹P NMR (202 MHz, CD₃CN, 298K) δ -139.39 (d, J = 706.4 Hz). ¹⁹F NMR (471 MHz, CD₃CN, 298K) δ -72.89 (d, J = 706.3 Hz). HRMS (ESI, positive ion): M⁺² = 356.5720 (calculated = 356.5731 for [C₃₃H₄₁N₅Rh₂]²⁺).



Molecular structure of intermediate 5 (50% probability level).

C. Catalytic Reaction with Intermediate 4:



To an oven dried sealed tube, **1a** (26.5 mg, 0.05 mmol), NaOAc (21 mg, 0.25 mmol), **4** (0.0015 mmol), AgOTf (65 mg, 0.25 mmol) and **2d** (19 mg, 0.11 mmol) were loaded. To this mixture, dry and degassed DCE (2.0 mL) was added and the reaction mixture was left with stirring at 120 °C in dark. After 12 h, the whole reaction mixture was passed through a short celite pad which was washed with dichloromethane (3×5 mL). The combined filtrate was concentrated under reduced pressure. Final product was separated and purified by alumina column chromatography, using CHCl₃/MeOH (95:5) as eluent. Yield of **3d**= 80%.

D. Stoichiometric Reaction with Intermediate 4:



To an oven dried sealed tube, **4** (47 mg, 0.025 mmol), NaOAc (21 mg, 0.25 mmol), AgOTf (32 mg, 0.125 mmol) and **2d** (10 μ L, 0.06 mmol) were loaded. To this mixture, dry and degassed DCE (1.5 mL) was added and the reaction mixture was left with stirring at 120 °C in dark. After 12 h, the whole reaction mixture was passed through a short celite pad which was washed with dichloromethane (3×5 mL). The combined filtrate was concentrated under reduced pressure. Final product was separated and purified by alumina column chromatography, using CHCl₃/MeOH (95:5) as eluent. Yield of **3d**= 64%.

E. Catalytic Reaction with Intermediate 5:



To an oven dried sealed tube, **1a** (27 mg, 0.05 mmol), NaOAc (21 mg, 0.25 mmol), **5** (0.0015 mmol), AgOTf (64 mg, 0.25 mmol) and **2d** (19 μ L, 0.12 mmol) were loaded. To this mixture, dry and degassed DCE (2.0 mL) was added and the reaction mixture was left with stirring at 120 °C in dark. After 12 h, the whole reaction mixture was passed through a short celite pad which was washed with dichloromethane (3×5 mL). The combined filtrate was concentrated under reduced pressure. Final product was separated and purified by alumina column chromatography, using CHCl₃/MeOH (95:5) as eluent. Yield of **3d**=62%.

F. Stoichiometric Reaction with Intermediate 5:



To an oven dried sealed tube, **5** (27 mg, 0.025 mmol), NaOAc (10 mg, 0.125 mmol), AgOTf (32 mg, 0.125 mmol) and **2d** (10.0 mg, 0.055 mmol) were loaded. To this mixture, dry and degassed DCE (2.0 mL) was added and the reaction mixture was left with stirring at 120 °C in dark. After 12 h, the whole reaction mixture was passed through a short celite pad which was washed with dichloromethane (3×5 mL). The combined filtrate was concentrated under reduced pressure. Final product was separated and purified by alumina column chromatography, using CHCl₃/MeOH (95:5) as eluent. Yield of **3d**= 43%.

10. Spectroscopic Measurements

Steady-state absorption spectra were recorded on SHIMADZU UV-1800 spectrophotometer. All steady-state fluorescence measurements were done on HORIBA Jobin Yvon Fluorolog spectrofluorimeter. The fluorescence and UV-Vis. spectra were recorded by using a 1 cm path length quartz cuvette. Both excitation and emission slit were kept at 1 nm while recording the fluorescence spectra. All the experiments were carried out at ambient temperature (298K).

Time-resolved fluorescence measurements were performed using a Hamamatsu MCP photomultiplier (R-3809U-50). The time-correlated single photon counting (TCSPC) setup consists of an Ortec 9327 pico-timing amplifier and using pulse Diode laser (λ_{ex} 405 nm) with fwhm ~143 ps with a setup target 10,000 counts. The instrument response function (IRF) was measured before and after fluorescence lifetime measurement using a dilute suspension of Ludox (colloidal silica, purchased from Sigma-Aldrich). The emission polarizer was positioned at the magic angle (54.7°) polarization with respect to excitation polarizer. The single exponential fitting function was employed by iterative deconvolution method using supplied software EzTime (HORIBA Scientific, v3.2.9.6). The general form of the fitting function was

$$I(t) = I(0) \sum \alpha_i \exp(-t/\tau_i)$$

where I(t) and I(0) are the fluorescence intensity at time t and 0 respectively, t is the time α_i and τ_i are the contributing amplitude and its corresponding lifetime. The quality of the fitted data was judged from the reduced chi-squared value (χ^2), calculated using the IBH software provided with the instrument. All the measurements were carried out at ambient temperature (298K).

11. Cell Culture and Imaging Studies

A. Materials and Instruments: Dulbecco's Modified Eagle Medium (DMEM), Trypsin, Antibiotic cocktail and Fetal Bovine Serum (FBS) were purchased from HiMedia (USA). LysoTracker Red and MitoTracker Red were purchased from Thermo Fisher SCIENTIFIC (USA). The 35 mm glass bottom imaging dishes were obtained from Ibidi (Germany, Cat#

81158). All the confocal microscopy imaging were performed with an Olympus FV3000 Confocal Laser Scanning Microscope (LSM). The image processing was done with the help of cellSens software (Olympus).

B. Culture Method: BHK-21 and CHO cells were obtained from NCCS, Pune, India and were grown in a 25 cm² cell culture flask (Corning, USA) using DMEM (phenol red free) containing 10% (v/v) FBS and 1% (v/v) antibiotic cocktail in 5% CO₂ at 37 °C in a CO₂ incubator. For imaging purpose, cells were grown to 75% - 80% confluency in the 35 mm glass bottom imaging dishes $(170 \pm 5 \,\mu\text{m})$ in DMEM with 10% FBS. The cells were washed twice with PBS (pH 7.4) containing 5 mM MgCl₂. Then the cells were co-incubated with 5 μ M of the dye of interest and 300 nM of LysoTracker Red/MitoTracker Red (whichever applicable) for 15 minutes and washed with PBS (pH 7.4) twice before imaging.

C. MTT Assay: Around 10,000 of cell per well were seeded in a 96 well-plate and grown for 24 h in the aforementioned condition. After that, the dye of interest was added according to the concentration from a concentrated stock in DMSO. The added amount of DMSO was not more than 2 μ L. After 24 h of incubation, 20 μ L of MTT dye solution (5 mg/ mL in PBS buffer) was added to each well and incubated for 4 h. The media was removed gently from each well and 200 μ L DMSO was added to each well and allowed to dissolve the purple color crystal for 30 min. The absorption at 570 nm was recorded in a microplate reader from BioTek (Synergy H1 Hybrid Multimode Reader).

D. Confocal Microscopy: For fluorescence imaging, 488 and 561 nm excitation lasers were used. For 488 nm and 561 nm excitation, the emission windows were kept at 500-560 nm and 565-650 nm, respectively. The confocal aperture was kept at 0.81 Airy Disk (AU) while the dwell time was 4 μ s/pixel.

Entry	Solvent	λ_{abs}^{max} (nm)	λ_{em}^{max} (nm)	Ф (%)	au (ns)
1	ACN	405	558	21.2	5.8
2	Acetone	406	560	11.3	4.9
3	CHCl ₃	404	560	12.6	3.5
4	DMSO	413	488	0.8	0.6
5	Ethylene glycol	409	560	10.4	4.7
6	EtOH	406	490	1.6	1.3
7	MeOH	406	550	6.3	2.4
8	THF	405	548	19.7	4.6
9	Water	406	565	9.8	4.1

Table S2: Solvent-dependent photophysical properties of 3j

Table S3: Solvent-dependent photophysical properties of 3s

Entry	Solvent	λ_{abs}^{max} (nm)	λ_{em}^{max} (nm)	Ф (%)	au (ns)
1	ACN	385	455	14.2	6.7
2	Acetone	388	455	15.6	6.4
3	CHCl ₃	380	455	10.3	3.9
4	DMSO	390	462	17.2	7.3
5	EtOH	385	452	22.8	6.4
6	MeOH	381	452	14.3	6.9
7	THF	390	455	18.1	6.6
8	Water	400	510	0.8	1.8



Absorption spectra of 3j (2 μ M) in different solvents



Emission spectra of 3j (2 μ M) in different solvents



Fluorescence lifetime of 3j (2 μ M) in different solvents



Scatter plot (**3j** and MitoTracker Red) obtained by fluorescence colocalization microscopy with Pearson's co-efficient 0.92±0.04



Absorption spectra of 3s (5 μ M) in different solvents



Emission spectra of 3s (5 μ M) in different solvents



Fluorescence lifetime of 3s (5 μ M) in different solvents



Scatter plot (**3s** and LysoTracker Red) obtained by fluorescence colocalization microscopy with Pearson's co-efficient 0.94±0.02

12. Computational Studies

Gaussian 09 software was used to carry out all the computational studies.^{S5} DFT calculations were performed with B3LYP exchange-correlation functional by using 6-31G (d) basis set for H, C, and N atoms.^{S6} Only gas phase calculations were performed to get optimized geometry of the structures. Frequency calculations were carried out to check the true minima of the optimized structures. For ground state optimized geometries, it showed 0 imaginary frequency. Later, images of HOMO and LUMO were obtained from optimized geometry to check electronic distributions and energy gap in between.

The x, y and z coordinates for the optimized structures of 3a, 3b and 3j are as below

_____ 3a _____ Symbolic Z-matrix: Charge = 2 Multiplicity = 1С -1.46555 -5.72544 -1.35355 С -3.35376 -4.95254 -2.50499 С -3.98998 -6.19177 -2.32676 С -3.33221 -7.21623 -1.6405 Η -3.85872 -4.16921 -3.03063 Ν -0.13411 -5.41527 -0.8645 С 0.49913 -4.25168 -1.01553 С 0.86686 -6.06048 -0.16519 С 2.00896 -5.64812 0.19907 Η 2.81722 -6.09798 0.73689 Ν 1.79969 -4.28513 -0.41081 С 2.76558 -3.1772 -0.3894 Η 3.7602 -3.57139 -0.37378

Н	2.63496 -2.57322 -1.26292
Н	2.60463 -2.58055 0.4841
С	-1.47269 -3.39907 -2.23513
С	-0.21212 -3.13059 -1.76261
С	0.07804 -1.88363 -0.90674
С	1.37713 -1.64034 -0.44083
С	-0.95702 -0.9922 -0.59381
С	1.64118 -0.50561 0.33801
Н	2.16742 -2.32097 -0.67976
С	-0.69298 0.14253 0.18503
Н	-1.94891 -1.17796 -0.94955
С	0.60612 0.38582 0.65095
Н	2.63307 -0.31986 0.69375
Н	-1.48326 0.82316 0.42397
Н	0.80772 1.25221 1.24561
С	-2.03933 -2.55804 -3.39409
С	-2.75908 -3.17894 -4.42383
С	-1.83522 -1.17181 -3.419
С	-3.27473 -2.4136 -5.47848
Н	-2.91492 -4.23736 -4.40481
С	-2.35087 -0.40647 -4.47366
Н	-1.28568 -0.69774 -2.63278
С	-3.07062 -1.02737 -5.5034
Н	-3.82427 -2.88767 -6.26471

Н	-2.19503 0.65195 -4.49268
Н	-3.46433 -0.44302 -6.30865
С	-2.11707 -4.73971 -2.02233
Ν	-2.04985 -6.97923 -1.14618
Ν	-3.97508 -8.52403 -1.44743
С	-5.24553 -8.876 -1.89372
С	-3.41327 -9.61382 -0.78877
С	-4.33436 -10.67026 -0.81537
Н	-2.41554 -9.56343 -0.3547
Н	-4.19559 -11.65711 -0.38679
Ν	-5.49592 -10.20246 -1.51595
С	-6.7193 -10.97129 -1.78641
Н	-7.56185 -10.45844 -1.37169
Н	-6.84939 -11.07438 -2.84345
Н	-6.63714 -11.94068 -1.34094
С	-6.1317 -7.88012 -2.66478
С	-5.40673 -6.42088 -2.88525
С	-7.51093 -8.3709 -3.14276
С	-5.99317 -5.19692 -3.61301
С	-7.41349 -4.70913 -3.27191
С	-5.26364 -4.54763 -4.55257
Н	-4.27679 -4.88655 -4.78956
С	-5.85008 -3.32367 -5.28032
С	-7.99993 -3.48517 -3.99967
Н	-8.98677 -3.14625 -3.76267
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С	-8.58027 -8.31554 -2.3121
Н	-8.46589 -7.92952 -1.32073
С	-7.67556 -8.92648 -4.56959
С	-9.05479 -9.41726 -5.04757
Н	-9.16917 -9.80328 -6.03894
С	-9.9595 -8.80632 -2.79008
Н	-6.83494 -8.97268 -5.22998
Н	-7.99428 -5.2177 -2.53101
Н	-10.8038 -8.76262 -2.13423
С	-10.26995 -9.35435 -4.10363
С	-7.14257 -2.87978 -4.96993
Н	-5.27408 -2.81102 -6.02215
Н	-6.95813 -2.12529 -5.64641
Н	-11.22225 -9.88639 -4.24528
Н	0.96609 -7.1415 -0.34279

-----3b

Symbolic Z-matrix:

Charge = 2 Multiplicity = 1		
С	-1.46555 -5.72544 -1.35355	
С	-3.35376 -4.95254 -2.50499	
С	-3.98998 -6.19177 -2.32676	
С	-3.33221 -7.21623 -1.6405	

Н	-3.85872 -4.16921 -3.03063
N	-0.13411 -5.41527 -0.8645
С	0.49913 -4.25168 -1.01553
С	0.86686 -6.06048 -0.16519
С	2.00896 -5.64812 0.19907
Н	2.81722 -6.09798 0.73689
Ν	1.79969 -4.28513 -0.41081
С	2.76558 -3.1772 -0.3894
Н	3.7602 -3.57139 -0.37378
Н	2.63496 -2.57322 -1.26292
Н	2.60463 -2.58055 0.4841
С	-1.47269 -3.39907 -2.23513
С	-0.21212 -3.13059 -1.76261
С	0.07804 -1.88363 -0.90674
С	1.37713 -1.64034 -0.44083
С	-0.95702 -0.9922 -0.59381
С	1.64118 -0.50561 0.33801
Н	2.16742 -2.32097 -0.67976
С	-0.69298 0.14253 0.18503
Н	-1.94891 -1.17796 -0.94955
С	0.60612 0.38582 0.65095
Н	2.63307 -0.31986 0.69375
Н	-1.48326 0.82316 0.42397
С	-2.03933 -2.55804 -3.39409

С	-2.75908 -3.17894 -4.42383
С	-1.83522 -1.17181 -3.419
С	-3.27473 -2.4136 -5.47848
Н	-2.91492 -4.23736 -4.40481
С	-2.35087 -0.40647 -4.47366
Н	-1.28568 -0.69774 -2.63278
С	-3.07062 -1.02737 -5.5034
Н	-3.82427 -2.88767 -6.26471
Н	-2.19503 0.65195 -4.49268
С	-2.11707 -4.73971 -2.02233
N	-2.04985 -6.97923 -1.14618
Ν	-3.97508 -8.52403 -1.44743
С	-5.24553 -8.876 -1.89372
С	-3.41327 -9.61382 -0.78877
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Н	-4.19559 -11.65711 -0.38679
N	-5.49592 -10.20246 -1.51595
С	-6.7193 -10.97129 -1.78641
Н	-7.56185 -10.45844 -1.37169
Н	-6.84939 -11.07438 -2.84345
Н	-6.63714 -11.94068 -1.34094
С	-6.1317 -7.88012 -2.66478
С	-5.40673 -6.42088 -2.88525

С	-7.51093 -8.3709 -3.14276
С	-5.99317 -5.19692 -3.61301
С	-7.41349 -4.70913 -3.27191
С	-5.26364 -4.54763 -4.55257
Н	-4.27679 -4.88655 -4.78956
С	-5.85008 -3.32367 -5.28032
С	-7.99993 -3.48517 -3.99967
Н	-8.98677 -3.14625 -3.76267
С	-8.58027 -8.31554 -2.3121
Н	-8.46589 -7.92952 -1.32073
С	-7.67556 -8.92648 -4.56959
С	-9.05479 -9.41726 -5.04757
Н	-9.16917 -9.80328 -6.03894
С	-9.9595 -8.80632 -2.79008
Н	-6.83494 -8.97268 -5.22998
Н	-7.99428 -5.2177 -2.53101
Н	-10.8038 -8.76262 -2.13423
С	-10.26995 -9.35435 -4.10363
С	-7.14257 -2.87978 -4.96993
Н	-5.27408 -2.81102 -6.02215
Н	0.96609 -7.1415 -0.34279
С	-11.60317 -10.0992 -4.30194
Н	-12.40642 -9.49778 -3.93048
Н	-11.75324 -10.29059 -5.34394

Н	-11.57619 -11.02676 -3.7692
С	-6.86681 -1.7517 -5.98136
Н	-6.23951 -1.01239 -5.52881
Н	-6.37706 -2.15645 -6.84231
Н	-7.79226 -1.30249 -6.27572
С	-3.63726 -0.18634 -6.66236
Н	-3.96058 -0.83537 -7.4492
Н	-4.46761 0.39034 -6.31183
Н	-2.87731 0.47036 -7.03129
С	0.89628 1.63277 1.50682
Н	1.27356 1.33032 2.46133
Н	1.62334 2.24303 1.01299
Н	-0.00646 2.19135 1.64081

3j

Symbolic Z-matrix:

Charge $= 2 N$	fultiplicity = 1
С	-1.46555 -5.72544 -1.35355
С	-3.98998 -6.19177 -2.32676
С	-3.33221 -7.21623 -1.6405
Ν	-0.13411 -5.41527 -0.8645
С	0.49913 -4.25168 -1.01553
С	0.86686 -6.06048 -0.16519
С	2.00896 -5.64812 0.19907
Н	2.81722 -6.09798 0.73689
Ν	1.79969 -4.28513 -0.41081
С	2.76558 -3.1772 -0.3894
Н	3.7602 -3.57139 -0.37378

Н	2.63496 -2.57322 -1.26292
Н	2.60463 -2.58055 0.4841
С	-1.47269 -3.39907 -2.23513
С	-0.21212 -3.13059 -1.76261
С	0.07804 -1.88363 -0.90674
С	1.37713 -1.64034 -0.44083
С	-0.95702 -0.9922 -0.59381
С	1.64118 -0.50561 0.33801
Н	2.16742 -2.32097 -0.67976
С	-0.69298 0.14253 0.18503
Н	-1.94891 -1.17796 -0.94955
С	0.60612 0.38582 0.65095
Н	2.63307 -0.31986 0.69375
Н	-1.48326 0.82316 0.42397
Н	0.80772 1.25221 1.24561
С	-2.03933 -2.55804 -3.39409
С	-2.75908 -3.17894 -4.42383
С	-1.83522 -1.17181 -3.419
С	-3.27473 -2.4136 -5.47848
Н	-2.91492 -4.23736 -4.40481
С	-2.35087 -0.40647 -4.47366
Н	-1.28568 -0.69774 -2.63278
С	-3.07062 -1.02737 -5.5034
Н	-3.82427 -2.88767 -6.26471
Н	-2.19503 0.65195 -4.49268
Н	-3.46433 -0.44302 -6.30865
С	-2.11707 -4.73971 -2.02233
Ν	-2.04985 -6.97923 -1.14618
Ν	-3.97508 -8.52403 -1.44743
С	-5.24553 -8.876 -1.89372
С	-3.41327 -9.61382 -0.78877
С	-4.33436 -10.67026 -0.81537
Н	-2.41554 -9.56343 -0.3547
Н	-4.19559 -11.65711 -0.38679

N	-5.49592 -10.20246 -1.51595
С	-6.7193 -10.97129 -1.78641
Н	-7.56185 -10.45844 -1.37169
Н	-6.84939 -11.07438 -2.84345
Н	-6.63714 -11.94068 -1.34094
С	-6.1317 -7.88012 -2.66478
С	-5.40673 -6.42088 -2.88525
С	-7.51093 -8.3709 -3.14276
С	-5.99317 -5.19692 -3.61301
С	-7.41349 -4.70913 -3.27191
С	-5.26364 -4.54763 -4.55257
Н	-4.27679 -4.88655 -4.78956
С	-5.85008 -3.32367 -5.28032
С	-7.99993 -3.48517 -3.99967
Н	-8.98677 -3.14625 -3.76267
С	-8.58027 -8.31554 -2.3121
Н	-8.46589 -7.92952 -1.32073
С	-7.67556 -8.92648 -4.56959
С	-9.05479 -9.41726 -5.04757
Н	-9.16917 -9.80328 -6.03894
С	-9.9595 -8.80632 -2.79008
Н	-6.83494 -8.97268 -5.22998
Н	-7.99428 -5.2177 -2.53101
Н	-10.8038 -8.76262 -2.13423
С	-10.26995 -9.35435 -4.10363
С	-7.14257 -2.87978 -4.96993
Н	-5.27408 -2.81102 -6.02215
Н	-6.95813 -2.12529 -5.64641
Н	-11.22225 -9.88639 -4.24528
Н	0.96609 -7.1415 -0.34279
Ν	-3.35376 -4.95254 -2.50499

13. ¹H, ¹³C{¹H}, ¹⁹F & ³¹P{¹H} NMR and ESI Mass Spectra



¹H NMR spectrum of **1a** (500 MHz, DMSO-d₆, 298 K).



ESI-HRMS (positive ion mode) spectrum of 1a.







ESI-HRMS (positive ion mode) spectrum of 1b.



¹H NMR spectrum of 1c (500 MHz, DMSO-d₆, 298 K).



ESI-HRMS (positive ion mode) spectrum of 1c



¹H NMR spectrum of **1d** (500 MHz, DMSO-d₆, 298 K).



ESI-HRMS (positive ion mode) spectrum of 1d



¹H NMR spectrum of **1e** (500 MHz, DMSO-d₆, 298 K)



ESI-HRMS (positive ion mode) spectrum of 1e.



 $^{^{13}}C\{^1H\}$ NMR spectrum of 1f (126 MHz, DMSO-d_6, 298 K).



ESI-HRMS (positive ion mode) spectrum of 1f



¹H NMR spectrum of **1g** (500 MHz, CDCl₃, 298 K).



ESI-HRMS (positive ion mode) spectrum of 1g



¹H NMR spectrum of **1h** (500 MHz, CDCl₃, 298K).



ESI-HRMS (positive ion mode) spectrum of 1h.



¹H NMR spectrum of **1i** (500 MHz, DMSO-d₆, 298K).







¹H NMR spectrum of **1j** (500 MHz, CDCl₃, 298 K).



ESI-HRMS (positive ion mode) spectrum of 1j



¹H NMR spectrum of **3a** (500 MHz, DMSO-d₆, 298 K).



¹³C{¹H} NMR spectrum of **3a** (126 MHz, DMSO-d₆, 298 K).



¹⁹F NMR spectrum of **3a** (471 MHz, DMSO-d₆, 298 K).



 ^{31}P NMR spectrum of **3a** (202 MHz, DMSO-d_6, 298 K).



ESI-HRMS (positive ion mode) spectrum of 3a



¹H NMR spectrum of **3b** (500 MHz, DMSO-d₆, 298 K).



³¹P NMR spectrum of **3b** (202 MHz, DMSO-d₆, 298 K).







ESI-HRMS (positive ion mode) spectrum of 3b.



¹H NMR spectrum of **3c** (400 MHz, CD₃CN, 298 K).



 $^{13}C\{^{1}H\}$ NMR spectrum of **3c** (126 MHz, CD₃CN, 298 K).



¹⁹F NMR spectrum of **3c** (471 MHz, CD₃CN, 298 K).



ESI-HRMS (positive ion mode) spectrum of 3c.



 1 H NMR spectrum of **3d** (500 MHz, DMSO-d₆, 298K).



 ^{31}P NMR spectrum of 3d (202 MHz, DMSO-d₆, 298K).

JC-PK-02-23-E(500MHz)









ESI-HRMS (positive ion mode) spectrum of 3d





¹³C{¹H} NMR spectrum of **3e** (126 MHz, CD₃CN, 298K).



--79.31

¹⁹F NMR spectrum of **3e** (471 MHz, DMSO-d₆, 298K).



ESI-HRMS (positive ion mode) spectrum of 3e.



¹H NMR spectrum of **3f** (mixture of 3 isomers) (500 MHz, DMSO-d₆, 298 K).



¹H NMR spectrum of **3f** (mixture of 3 isomers) (500 MHz, DMSO-d₆, 298 K).



¹³C{¹H} NMR spectrum of **3f** (mixture of 3 isomers) (126 MHz, DMSO-d₆, 298 K).



³¹P NMR spectrum of **3f** (mixture of 3 isomers) (202 MHz, DMSO-d₆, 298 K).



¹⁹F NMR spectrum of **3f** (mixture of 3 isomers) (471 MHz, DMSO-d₆, 298 K).



ESI-HRMS (positive ion mode) spectrum of 3f.



¹H NMR spectrum of **3g** (mixture of 3 isomers) (500 MHz, CD₃CN, 298 K).



¹H NMR spectrum of **3g** (mixture of 3 isomers) (500 MHz, CD₃CN, 298 K).



¹³C{¹H} NMR spectrum of **3g** (mixture of 3 isomers) (126 MHz, CD₃CN, 298 K).



ESI-HRMS (positive ion mode) spectrum of 3g.



¹H NMR spectrum of **3h** (500 MHz, DMSO-d₆, 298 K).



 $^{13}\text{C}\{^1\text{H}\}$ NMR spectrum of **3h** (126 MHz, DMSO-d_6, 298 K).






 ^{19}F NMR spectrum of **3h** (471 MHz, DMSO-d₆, 298K).



ESI-HRMS (positive ion mode) spectrum of 3h.





¹H NMR spectrum of **3i** (400 MHz, CD₃CN, 298K).



 $^{13}C\{^1H\}$ NMR spectrum of **3i** (126 MHz, CD₃CN, 298 K).



¹⁹F NMR spectrum of **3i** (471 MHz, CD₃CN, 298 K).



ESI-HRMS (positive ion mode) spectrum of 3i.



¹³C{¹H} NMR spectrum of **3j** (126 MHz, CD₃CN, 298 K).



¹⁹F NMR spectrum of **3j** (471 MHz, CD₃CN, 298K).



ESI-HRMS (positive ion mode) spectrum of 3j.



¹H NMR spectrum of **3k** (400 MHz, CD₃CN, 298 K).



 ^{31}P NMR spectrum of 3k (202 MHz, DMSO-d_6, 298K).

--73.19

-71.68





¹⁹F NMR spectrum of **3k** (471 MHz, DMSO-d₆, 298 K).



ESI-HRMS (positive ion mode) spectrum of 3k.





¹H NMR spectrum of **3l** (500 MHz, CD₃CN, 298 K).



 $^{13}C\{^{1}H\}$ NMR spectrum of **3l** (126 MHz, CD₃CN, 298K).





 ^{19}F NMR spectrum of **3l** (471 MHz, CD₃CN, 298 K).



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¹H NMR spectrum of **3m** (500 MHz, DMSO-d₆, 298 K).



¹⁹F NMR spectrum of **3m** (471 MHz, DMSO-d₆, 298 K).







¹H NMR spectrum of **3n** (500 MHz, DMSO-d₆, 298K).



¹⁹F NMR spectrum of **3n** (471 MHz, DMSO-d₆, 298 K).



ESI-HRMS (positive ion mode) spectrum of 3n.



¹H NMR spectrum of **30** (500 MHz, CD₃CN, 298 K).







JC-PK-02-111(500MHz)









¹⁹F NMR spectrum of **30** (471 MHz, CD₃CN, 298 K).



ESI-HRMS (positive ion mode) spectrum of 30.



¹H NMR spectrum of **3p** (500 MHz, CD₃CN, 298 K).



ppm

¹³C{¹H} NMR spectrum of **3p** (126 MHz, CD₃CN, 298 K).





¹⁹F NMR spectrum of **3p** (471 MHz, CD₃CN, 298 K).



ESI-HRMS (positive ion mode) spectrum of 3p.



¹H NMR spectrum of **3q** (500 MHz, DMSO-d₆, 298 K).



¹⁹F NMR spectrum of **3q** (376 MHz, DMSO-d₆, 298 K).



ESI-HRMS (positive ion mode) spectrum of 3q.

4.370 3.507⊬M (50 137) 3.48 3.48 3.48 3.29 3.29 3.29 1.77 1.22 1.22 1.22 1.19 1.19 1.19 1.19 1.18 1.18 1.16 46 48 3.31 3.28 2.16 .95 .82 .80 ..79 3.47 L.97 .95 82 .79 3.31 -94 9 6 .78 8 5



¹H NMR spectrum of **3r** (500 MHz, CD₃CN, 298 K).



 ^{19}F NMR spectrum of 3r (376 MHz, DMSO-d₆, 298 K).



ESI-HRMS (positive ion mode) spectrum of **3r**.



¹H NMR spectrum of **3s** (500 MHz, CDCl₃, 298 K).



 $^{13}C\{^1H\}$ NMR spectrum of **3s** (126 MHz, CDCl₃, 298 K).



ESI-HRMS (positive ion mode) spectrum of 3s.



¹³C{¹H} NMR spectrum of **3t** (126 MHz, CDCl₃, 298 K).







¹H NMR spectrum of **3u** (500 MHz, CDCl₃, 298 K).



180 170 160 150 140 130 120 110 100 90 80 70 60 50 40 30 20 10 0 -1 ppm

¹³C{¹H} NMR spectrum of **3u** (126 MHz, CDCl₃, 298 K).



ESI-HRMS (positive ion mode) spectrum of 3u.



¹H NMR spectrum of **3v** (500 MHz, CDCl₃, 298 K).



 $^{13}C\{^{1}H\}$ NMR spectrum of **3v** (126 MHz, CDCl₃, 298 K).



ESI-HRMS (positive ion mode) spectrum of 3v.





¹H NMR spectrum of **3w** (500 MHz, CDCl₃, 298 K).



 $^{13}C\{^{1}H\}$ NMR spectrum of **3w** (176 MHz, CDCl₃, 298 K).



ESI-HRMS (positive ion mode) spectrum of 3w.



¹H NMR spectrum of **3x** (500 MHz, CDCl₃, 298 K).



 $^{13}C\{^1H\}$ NMR spectrum of **3x** (176 MHz, CDCl₃, 298 K).



ESI-HRMS (positive ion mode) spectrum of 3x.



¹H NMR spectrum of **3y** (500 MHz, CDCl₃, 298 K).





 $^{13}\mathrm{C}\{^{1}\mathrm{H}\}$ NMR spectrum of **3y** (126 MHz, CDCl3, 298 K).







¹H NMR spectrum of **3z** (500 MHz, CDCl₃, 298 K).



¹³C{¹H} NMR spectrum of **3z** (126 MHz, CDCl3, 298 K).



ESI-HRMS (positive ion mode) spectrum of 3z.



¹H NMR spectrum of **3aa** (500 MHz, CDCl₃, 298 K).






ESI-HRMS (positive ion mode) spectrum of 3aa.



¹H NMR spectrum of **3ab** (mixture of 3 isomers) (500 MHz, CDCl₃, 298 K).



¹H NMR spectrum of **3ab** (mixture of 3 isomers) (500 MHz, CDCl₃, 298 K).



¹³C{¹H} NMR spectrum of **3ab** (mixture of 3 isomers) (126 MHz, CDCl₃, 298 K).



ESI-HRMS (positive ion mode) spectrum of 3ab.



¹H NMR spectrum of **3ac** (mixture of 3 isomers) (500 MHz, CDCl₃, 298 K).



¹H NMR spectrum of **3ac** (mixture of 3 isomers) (500 MHz, CDCl₃, 298 K).



¹³C{¹H} NMR spectrum of **3ac** (mixture of 3 isomers) (126 MHz, CDCl₃, 298 K).



ESI-HRMS (positive ion mode) spectrum of 3ac.



¹H NMR spectrum of **3ad** (mixture of two isomers) (500 MHz, CDCl₃, 298 K).



¹³C{¹H} NMR spectrum of **3ad** (mixture of two isomers) (126 MHz, CDCl₃, 298 K).



ESI-HRMS (positive ion mode) spectrum of 3ad.



¹H NMR spectrum of **3ae** (500 MHz, CDCl₃, 298 K).



 $^{13}C\{^{1}H\}$ NMR spectrum of **3ae** (126 MHz, CDCl₃, 298 K).



ESI-HRMS (positive ion mode) spectrum of 3ae.



¹³C{¹H} NMR spectrum of **3af** (126 MHz, CDCl₃, 298 K).



ESI-HRMS (positive ion mode) spectrum of 3af.



¹H NMR spectrum of **4** (500 MHz, CD₃CN, 298 K).

JC-PK-02-126-3(500MHz)







¹⁹F NMR spectrum of **4** (471 MHz, CD₃CN, 298 K).



ESI-HRMS (positive ion mode) spectrum of 4.



¹H NMR spectrum of **5** (500 MHz, CD₃CN, 298 K).



 $^{13}C\{^1H\}$ NMR spectrum of **5** (126 MHz, CD₃CN, 298 K).



³¹P NMR spectrum of **5** (202 MHz, CD₃CN, 298 K).





¹⁹F NMR spectrum of **5** (471 MHz, CD₃CN, 298 K).



ESI-HRMS (positive ion mode) spectrum of 5.

14. References

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