Supporting Information

Nickel-Catalyzed Monofluoromethylation of (Hetero)aryl Bromides

via Reductive Cross-coupling

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General Information

NMR spectra were recorded on Bruker-400 (400 MHz for ¹H, 101 MHz for ¹³C and 376 MHz for ¹⁹F {¹H, ¹³C decoupled}) instruments internally referenced to SiMe₄ signal. High resolution mass spectra were recorded on P-SIMS-Gly of Bruker Daltonics Inc. using ESI-TOF (electrospray ionization-time of flight) or Micromass GCT using EI (electron impact). Anhydrous DMAc was obtained from Infsci. BrCH₂F was obtained from ShangFluoro. NiI₂ was obtained from Strem. Substrates **1a** and **1c** were obtained from Adamas-beta. DMAP and substrate **1e** were obtained from SCR. Substrates **1d**, **1h** and **1m** were obtained from Accela. Substrates **1b**, **1f**, **1g**, **1u** and **1ac** were obtained from Energy. Substrates **1i**, **1k**, **1l**, **1p**, **1q**, **1s**, **1t**, **1v**, **1x** and **1aa** were obtained from Bidepharm. Substrates **1r** and **1z** were obtained from Macklin. Mn powder and substrate **1ad** were obtained from Aladdin. Substrate **1y** was obtained from AikonChem. All reagents were used as received without further purification.

Tables of the Optimization of Reaction Conditions

	Ph + 1a	BrCH ₂ F 2	Nil ₂ Ligai Mi N ₂ , DM	2 (10 mol%) nds (x mol%) n (3 equiv) Ac, 40 °C, 24 h 3a	l₂F
Entry	Ligand (x mol%)	Yie l d(%) ^b	Entry	Ligands (x mol%)	Yield(%) ^b
1	None	0	11	dombpy(12)/DMAP(24)	76
2	dmbpy (12)	64	12	dtbpy(12)/DMAP(24)	85(86) ^c
3	dtbpy (12)	71	13	dmbpy(12)/DMAP(24)	72
4	dombpy (12)	79	14	phen(12)/DMAP(24)	69
5	phen (12)	52	15	bpy(12)/DMAP(24)	50
6	dppe (12)	12	16	dtbpy(12)/4-CN-Py(24)	74
7	DMAP (24)	15	17	dtbpy(12)/2,6-lutidine(24)	78
8	4-CN-Py (24)	0	18	dtbpy(12)/PPh ₃ (24)	50
9	2,6 -l utidine (24)	0	19	dmbpy(12)/4-CN-Py(24)	0
10	PPh ₃ (24)	0	20	dmbpy(12)/PPh ₃ (24)	64

Table S1. Ligands Screening^a

^a Reaction conditions (unless otherwise specified): **1a** (0.2 mmol, 1.0 equiv), **2** (2.5 equiv), Nil₂ (10 mol%), ligands, Mn (200 mesh, 3.0 equiv), DMAc (1 mL), 40 °C, under N₂ atmosphere, 24 h. ^b Yields determined by ¹⁹F NMR using PhCF₃ as an internal standard. ^c Isolated yield.



Table S2. Nickel Sources Screening^a

Ph	Br + 1a	BrCH ₂ F — N 2	[Ni] (10 mol%) dtbpy (12 mol%) DMAP (24 mol%) Mn (3 equiv) ₂ , DMAc, 40 °C, 24	h Ph 3a	H ₂ F
Entry	[Ni]	Yield(%) ^b	Entry	[Ni]	Yield(%) ^b
1	None	0	5	Ni(OAc) ₂	19
2	NiCl ₂	39	6	Ni(NO ₃) ₂ ·6H ₂ O	0
3	NiBr ₂	<5	7	Ni(acac) ₂	68
4	Nil ₂	86 ^c	8	Ni(OTf) ₂	0

^a Reaction conditions (unless otherwise specified): **1** (0.2 mmol, 1.0 equiv), **2** (2.5 equiv), [Ni] (10 mol%), dtbpy (12 mol%), DMAP (24 mol%), Mn (200 mesh, 3.0 equiv), DMAc (1 mL), 40 °C, under N₂ atmosphere, 24 h. ^b Yields determined by ¹⁹F NMR using PhCF₃ as an internal standard. ^c Isolated yield.

Nil₂ (10 mol%) dtbpy (x mol%) CH₂F Br DMAP (y mol%) BrCH₂F Mn (z equiv) Ph N₂, solvent, 40 °C, 24 h 1a 2 3a Entry z solvent Yield(%)^b Х у 1 10 20 3.0 DMAc 80 2 12 24 3.0 DMF 0 3 24 MeCN 12 3.0 41 4 12 24 3.0 DMSO 0 5 12 79 2.0 DMAc 24 DMAc 6^c 12 24 3.0 81 7^d 12 24 3.0 DMAc 76

Table S3. Optimization of Other Conditions^{*a*}

^{*a*} Standard reaction conditions (unless otherwise specified): **1a** (0.2 mmol, 1.0 equiv), **2** (2.5 equiv), Nil₂ (10 mol%), dtbpy (12 mol%), DMAP (24 mol%), Mn (200 mesh, 3.0 equiv), DMAc (1 mL), 40 °C, under N₂ atmosphere, 24 h. ^{*b*} Yields determined by ¹⁹F NMR using PhCF₃ as an internal standard. ^{*c*} H₂O (3 equiv) was added. ^{*d*} 60 °C.

Preparation of Substrates

Substrates $1j^1$, $1w^2$ and $1ab^3$ were prepared in accordance with methods described in the references.

Synthesis of Fenofibrate bromide (1ae):



Fenofibrate boronic acid⁴ (2 mmol, 0.74 g), 1,3-dibromo-5,5-dimethylhydantoin (2.2 mmol, 0.63 mg) and NaOMe (0.1 mmol, 5.4 mg) were added to a 50 mL Schlenk tube equipped with a magnetic stirring bar. The vessel was evacuated and backfilled with N₂ (repeated for 3 times). Acetonitrile (10 mL) were added to the tube at room temperature under a stream of nitrogen, and the tube was sealed and put into a preheated oil bath at 40 °C for 4 h under nitrogen atmosphere. After the resulting solution was cooled to room temperature, Na_2SO_3 (10% aq, ~20 mL) was added, and the aqueous layer was extracted with MTBE (3×15 mL). The combined organic phase was washed with NaOH (10% aq) and dried over anhydrous Na₂SO₄, filtrated and concentrated under vacuum. The residue was then purified by flash column chromatography (PE/EA = 5:1) to give the product as a white solid (> 80%). ¹H NMR (400 MHz, Chloroform-d) δ 7.76–7.70 (m, 2H), 7.61 (s, 4H), 6.89–6.82 (m, 2H), 5.08 (hept, J = 6.3 Hz, 1H), 1.66 (s, 6H), 1.20 (d, J = 6.3 Hz, 6H). ¹³C NMR (101 MHz, Chloroform-d) & 194.51, 173.21, 159.88, 136.98, 132.09, 131.63, 131.41, 130.25, 127.03, 117.33, 79.53, 69.48, 25.49, 21.66. HRMS ESI (m/z): [M+H]⁺ calcd. For C₂₀H₂₂BrO₄: 405.0701 found: 405.0685.

Preparation of BrCH₂F Stock Solution⁵

Dry DMAc (~23 mL) was added to a Schlenk graduated cylinder under nitrogen. The vessel and solvent were weighed. Next, BrCH₂F (~2mL) was added and the total volume of the solution reached approximately 25 mL. The vessel was sealed and weighed again. The concentration of the BrCH₂F stock solution was calculated based on the mass of BrCH₂F added and the total volume of the solution (~0.2 mol/L).

General Procedure for Nickel-Catalyzed Cross-Coupling between (Hetero)aryl Bromides and Bromofluoromethane

To a 5 mL of sealing tube were added NiI₂ (10 mol%), dtbpy (4,4'-Di-tert-butyl-2,2'-bipyridine, 12 mol%), DMAP (4-Dimethylaminopyridine, 24 mol%) and manganese powder (200 mesh, 3.0 equiv). The vessel was evacuated and backfilled with N₂ (repeated for 3 times), after that, aryl bromine **1** (0.2 mmol, 1.0 equiv) and dry DMAc (1 mL) were added. The solution then premix for 10 s before BrCH₂F **2** (solution in DMAc, 2.5 equiv) was added. The tube was sealed with a Teflon lined cap and heated in a preheated oil bath at 40°C for 24 h. The reaction mixture was then cooled to room temperature, diluted with EtOAc (~20 mL) and filtered through a pad of celite. The filtrate was added brine (30 mL) and extracted with EtOAc (2×15 mL), the combined organic layer was dried over Na₂SO₄, filtrated and concentrated under vacuum. The residue was then purified by flash column chromatography to give **3** as a colorless solid or oil.



4-(fluoromethyl)-1,1'-biphenyl (3a) was purified with silica gel chromatography (PE) as a white solid (86% yield). The compound is known⁵. ¹H NMR (400 MHz, Chloroform-*d*) δ 7.66 (t, *J* = 8.5 Hz, 4H), 7.53-7.47 (m, 4H), 7.41 (t, *J* = 7.3 Hz, 1H),

5.46 (d, J = 47.9 Hz, 2H). ¹³C NMR (101 MHz, Chloroform-*d*) δ 141.83 (d, J = 3.3 Hz), 140.69 (d, J = 1.1 Hz), 135.23 (d, J = 17.0 Hz), 128.95, 128.19 (d, J = 5.7 Hz), 127.66, 127.47 (d, J = 1.4 Hz), 127.27, 84.51 (d, J = 166.0 Hz). ¹⁹F NMR (376 MHz, Chloroform-*d*) δ -206.03 (t, J = 47.8 Hz).



3-(fluoromethyl)-1,1'-biphenyl (3b) was purified with silica gel chromatography (PE) as a colorless liquid (89% yield). The compound is known⁶. ¹H NMR (400 MHz, Chloroform-*d*) δ 7.67-7.60 (m, 4H), 7.53-7.45 (m, 3H), 7.43-7.36 (m, 2H), 5.48

(d, J = 47.8 Hz, 2H). ¹³C NMR (101 MHz, Chloroform-*d*) δ 141.79 (d, J = 1.0 Hz), 140.80, 136.85 (d, J = 17.1 Hz), 129.19, 128.95, 127.67, 127.65, 127.30, 126.43 (d, J = 2.0 Hz), 126.37 (d, J = 2.1 Hz), 84.72 (d, J = 166.6 Hz). ¹⁹F NMR (376 MHz, Chloroform-*d*) δ -207.15 (t, J = 47.8 Hz).



1-(fluoromethyl)-2-methylbenzene (3c) due to the low boiling point of the product, the yield (74%) was determined by ¹⁹F NMR using PhCF₃ as an internal standard. This compound is known⁷. The product was characterized by ¹⁹F NMR and GC-MS analysis.

2-(fluoromethyl)-1,3-dimethylbenzene (3d) due to the low

boiling point of the product, the yield (76%) was determined by 19 F NMR using PhCF₃ as an internal standard. This compound is

known⁷. The product was characterized by ¹⁹F NMR and GC-MS



analysis.



CH₂F OMe 3f

(Fluoromethyl)benzene (3e) due to the low boiling point of the product, the yield (83%) was determined by ¹⁹F NMR using PhCF₃ as an internal standard. This compound is known⁶. The product was characterized by ¹⁹F NMR and GC-MS analysis.



1-(fluoromethyl)-2-methoxybenzene (3f) due to the low boiling point of the product, the yield (77%) was determined by ¹⁹F NMR using PhCF₃ as an internal standard. This compound is known⁶. The product was characterized by ¹⁹F NMR and GC-MS analysis.



1-(fluoromethyl)-3-methoxybenzene (3g) due to the low boiling point of the product, the yield (82%) was determined by ¹⁹F NMR using PhCF₃ as an internal standard. This compound is known⁶. The product was characterized by ¹⁹F NMR and GC-

MS analysis.



1-(fluoromethyl)-3,5-dimethoxybenzene (3h) was purified with silica gel chromatography (PE/EA = 20:1) as a colorless liquid (84% yield). ¹H NMR (400 MHz, Chloroform-d) δ 6.52 (t, J = 1.7 Hz, 2H), 6.45 (q, J = 2.1 Hz, 1H), 5.32 (d, J = 47.6

Hz, 2H), 3.80 (s, 6H). ¹³C NMR (101 MHz, Chloroform-d) δ 161.13, 138.62 (d, J = 17.3 Hz), 105.03 (d, J = 6.5 Hz), 100.77 (d, J = 2.6 Hz), 84.60 (d, J = 167.5 Hz), 55.53. ¹⁹F NMR (376 MHz, Chloroform-d) δ -208.96 (t, J = 47.7 Hz). HRMS EI (m/z): [M]⁺ calcd. For C₉H₁₁FO₂: 170.0743 found: 170.0739.



1-(benzyloxy)-3-(fluoromethyl)benzene (3i) was purified with silica gel chromatography (PE/EA = 20:1) as a white solid (94%) yield). The compound is known⁸. ¹H NMR (400 MHz, Chloroform-d) δ 7.53–7.39 (m, 4H), 7.39–7.29 (m, 2H), 7.04 (s,

1H), 6.99 (d, J = 7.7 Hz, 2H), 5.38 (d, J = 47.7 Hz, 2H), 5.10 (s, 2H). ¹³C NMR (101 MHz, Chloroform-*d*) δ 159.11, 137.90 (d, J = 17.1 Hz), 136.91, 129.85, 128.73, 128.15, 127.61, 119.90 (d, J = 6.1 Hz), 115.30 (d, J = 2.8 Hz), 113.77 (d, J = 6.3 Hz), 84.54 (d, J = 166.9 Hz), 70.12. ¹⁹F NMR (376 MHz, Chloroform-*d*) δ -207.84 (t, J = 47.7 Hz).



4-(fluoromethyl)phenyl-4-methylbenzoate (3j) was purified with silica gel chromatography (PE/EA = 20:1) as a white solid (77% yield). ¹H NMR (400 MHz, Chloroform-*d*) δ 8.11 (d, *J* = 8.0 Hz, 2H), 7.45 (dd, *J* = 8.5, 1.9 Hz, 2H), 7.32 (d, *J* = 8.0 Hz, 2H), 7.26 (d, *J* =

8.2 Hz, 2H), 5.40 (d, J = 47.8 Hz, 2H), 2.46 (s, 3H). ¹³C NMR (101 MHz, Chloroformd) δ 165.23 (d, J = 0.4 Hz), 151.42 (d, J = 3.3 Hz), 144.68, 133.87 (d, J = 17.5 Hz), 130.35, 129.44, 128.92 (d, J = 5.8 Hz), 126.73, 122.13 (d, J = 1.1 Hz), 84.13 (d, J =166.6 Hz), 21.89. ¹⁹F NMR (376 MHz, Chloroform-*d*) δ -206.06 (t, J = 47.8 Hz). HRMS ESI (m/z): [M+Na]⁺ calcd. For C₁₅H₁₃FO₂Na: 267.0797 found: 267.0792.



(4-(fluoromethyl)phenyl)(phenyl)methanone (3k) was purified with silica gel chromatography (PE/EA = 15:1) as a pale yellow oil (95% yield). The compound is known⁵. ¹H NMR (400 MHz, Chloroform-*d*) δ 7.85-7.71 (m, 4H), 7.63–7.56 (m, 1H),

7.51-7.46 (m, 4H), 5.47 (d, J = 47.2 Hz, 2H). ¹³C NMR (101 MHz, Chloroform-*d*) δ 196.28, 140.72 (d, J = 17.2 Hz), 137.80 (d, J = 2.5 Hz), 137.51, 132.66, 130.41, 130.11, 128.44, 126.70 (d, J = 6.5 Hz), 83.83 (d, J = 168.4 Hz). ¹⁹F NMR (376 MHz, Chloroform-*d*) δ -212.32 (t, J = 47.2 Hz).



(3-(fluoromethyl)phenyl)(phenyl)methanone (3l) was purified with silica gel chromatography (PE/EA = 15:1) as a colorless liquid (83% yield). ¹H NMR (400 MHz, Chloroform-*d*) δ 7.95– 7.74 (m, 4H), 7.60 (t, *J* = 7.5 Hz, 2H), 7.57–7.44 (m, 3H), 5.44 (d,

J = 47.5 Hz, 2H). ¹³C NMR (101 MHz, Chloroform-*d*) δ 196.38, 138.08, 137.41, 136.67 (d, J = 17.5 Hz), 132.74, 131.27 (d, J = 5.8 Hz), 130.44 (d, J = 2.6 Hz), 130.14, 128.81 (d, J = 6.2 Hz), 128.74, 128.49, 84.06 (d, J = 167.7 Hz). ¹⁹F NMR (376 MHz, Chloroform-*d*) δ -208.92 (t, J = 47.7 Hz). HRMS ESI (m/z): [M+H]⁺ calcd. For C₁₄H₁₂FO: 215.0872 found: 215.0869.



(3-(fluoromethyl)phenyl)(thiophen-2-yl)methanone (3m) was purified with silica gel chromatography (PE/EA = 5:1) as a colorless liquid (68% yield). ¹H NMR (400 MHz,

Chloroform-*d*) δ 7.91–7.81 (m, 2H), 7.74 (dd, *J* = 5.0, 1.1 Hz, 1H), 7.64 (dd, *J* = 3.8, 1.1 Hz, 1H), 7.62-7.59 (m, 1H), 7.58–7.48 (m, 1H), 7.17 (dd, *J* = 4.9, 3.8 Hz, 1H), 5.46 (d, *J* = 47.5 Hz, 2H). ¹³C NMR (101 MHz, Chloroform-*d*) δ 187.85, 143.49, 138.62, 136.84 (d, *J* = 17.6 Hz), 135.08, 134.60, 131.04 (d, *J* = 5.9 Hz), 129.50 (d, *J* = 2.6 Hz), 128.88, 128.19, 127.95 (d, *J* = 6.3 Hz), 84.03 (d, *J* = 167.8 Hz). ¹⁹F NMR (376 MHz, Chloroform-*d*) δ -209.28 (t, *J* = 47.5 Hz). HRMS ESI (m/z): [M+H]⁺ calcd. For C₁₂H₁₀FOS: 221.0436 found: 221.0429.



1-chloro-3-(fluoromethyl)benzene (3n) due to the low boiling point of the product, the yield (78%) was determined by ¹⁹F NMR using PhCF₃ as an internal standard. This compound is known⁹. The product was characterized by ¹⁹F NMR and GC-MS analysis.



9-(fluoromethyl)phenanthrene (3o) was purified with silica gel chromatography (PE) as a white solid (95% yield). ¹H NMR (400 MHz, Chloroform-*d*) δ 8.74 (dd, *J* = 7.3, 2.2 Hz, 1H), 8.69 (d, *J* = 8.3 Hz, 1H), 8.11 (dt, *J* = 6.6, 1.8 Hz, 1H), 7.91 (d, *J* = 7.7 Hz,

1H), 7.81 (d, J = 3.1 Hz, 1H), 7.70 (ddt, J = 10.8, 7.0, 3.4 Hz, 3H), 7.65 – 7.59 (m, 1H), 5.89 (d, J = 47.8 Hz, 2H). ¹³C NMR (101 MHz, Chloroform-*d*) δ 131.13 (d, J = 1.3 Hz), 131.03 (d, J = 1.8 Hz), 130.74, 130.36 (d, J = 15.2 Hz), 130.04 (d, J = 1.1 Hz), 129.08 (d, J = 1.4 Hz), 128.03 (d, J = 9.5 Hz), 127.51 (d, J = 1.1 Hz), 127.16, 127.01 (d, J = 0.6 Hz), 126.92, 124.37 (d, J = 1.3 Hz), 123.29, 122.70 (d, J = 1.0 Hz), 83.87 (d, J = 166.4 Hz). ¹⁹F NMR (376 MHz, Chloroform-*d*) δ -207.89 (t, J = 47.8 Hz). HRMS EI (m/z): [M]⁺ calcd. For C₁₅H₁₁F: 210.0845 found: 210.0835.



9-(4-(fluoromethyl)phenyl)-9H-carbazole (**3p**) was purified with silica gel chromatography (PE/EA = 20:1) as a white solid (77% yield). ¹H NMR (400 MHz, Chloroform-*d*) δ 8.18 (dt, *J* = 7.7, 1.0 Hz, 2H), 7.63 (s, 4H), 7.47–7.40 (m, 4H), 7.36–7.28 (m, 2H), 5.52 (d, *J* = 47.7 Hz, 2H). ¹³C NMR

(101 MHz, Chloroform-*d*) δ 140.84, 138.25 (d, *J* = 3.1 Hz), 135.43 (d, *J* = 17.4 Hz), 129.17 (d, *J* = 5.8 Hz), 127.33 (d, *J* = 0.9 Hz), 126.14, 123.57, 120.48, 120.22, 109.81, 84.21 (d, *J* = 167.0 Hz). ¹⁹F NMR (376 MHz, Chloroform-*d*) δ -207.06 (t, *J* = 47.7 Hz). HRMS ESI (m/z): [M+H]⁺ calcd. For C₁₉H₁₅FN: 276.1189 found: 276.1180.



9-(3-(fluoromethyl)phenyl)-9H-carbazole (**3q**) was purified with silica gel chromatography (PE/EA = 20:1) as a colorless liquid (93% yield). ¹H NMR (400 MHz, Chloroform-*d*) δ 8.20 (dt, *J* = 7.8, 1.0 Hz, 2H), 7.70–7.58 (m, 3H), 7.54–7.42 (m, 5H), 7.39-7.31 (m, 2H), 5.52 (d, *J*

= 47.5 Hz, 2H). ¹³C NMR (101 MHz, Chloroform-*d*) δ 140.85, 138.42 (d, J = 17.5 Hz), 138.18, 130.29, 127.38 (d, J = 2.5 Hz), 126.25 (d, J = 6.0 Hz), 126.12, 125.89 (d, J = 6.3 Hz), 123.54, 120.46, 120.20, 109.78, 84.02 (d, J = 167.9 Hz). ¹⁹F NMR (376 MHz, Chloroform-*d*) δ -208.81 (t, J = 47.4 Hz). HRMS ESI (m/z): [M+H]⁺ calcd. For C₁₉H₁₅FN: 276.1189 found: 276.1177.



4-(3-(fluoromethyl)phenyl)morpholine (3r) was purified with silica gel chromatography (PE/EA = 5:1) as a colorless liquid (93% yield). ¹H NMR (400 MHz, Chloroform-*d*) δ 7.30 (t, *J* = 7.8 Hz, 1H), 6.97–6.84 (m, 3H), 5.35 (d, *J* = 47.8 Hz,

2H), 3.91–3.81 (m, 4H), 3.24–3.13 (m, 4H). ¹³C NMR (101 MHz, Chloroform-*d*) δ 151.66, 137.35 (d, *J* = 16.9 Hz), 129.56 (d, *J* = 0.8 Hz), 118.95 (d, *J* = 5.9 Hz), 116.02 (d, *J* = 2.9 Hz), 114.57 (d, *J* = 6.3 Hz), 84.97 (d, *J* = 166.5 Hz), 66.99, 49.31. ¹⁹F NMR (376 MHz, Chloroform-*d*) δ -207.25 (t, *J* = 47.8 Hz). HRMS EI (m/z): [M]⁺ calcd. For C₁₁H₁₄FNO: 195.1059 found: 195.1050.



2-(4-(fluoromethyl)phenyl)pyridine (3s) was purified with silica gel chromatography (PE/EA = 5:1) as a colorless liquid (80% yield). The compound is known¹⁰. ¹H NMR (400 MHz, Chloroform-*d*) δ 8.74 – 8.67 (m, 1H), 8.03 (dd, *J* = 8.4, 1.3 Hz,

2H), 7.81–7.70 (m, 2H), 7.56–7.44 (m, 2H), 7.23 (ddd, J = 6.0, 4.8, 2.5 Hz, 1H), 5.43 (d, J = 47.7 Hz, 2H). ¹³C NMR (101 MHz, Chloroform-*d*) δ 156.90 (d, J = 1.4 Hz), 149.83, 139.87 (d, J = 3.0 Hz), 136.96 (d, J = 17.3 Hz), 136.89, 127.86 (d, J = 5.9 Hz), 127.20 (d, J = 1.3 Hz), 122.43, 120.66, 84.36 (d, J = 166.6 Hz). ¹⁹F NMR (376 MHz, Chloroform-*d*) δ -208.00 (t, J = 47.7 Hz).



2-(4-(fluoromethyl)phenyl)thiophene (3t) was purified with silica gel chromatography (PE) as a white solid (75% yield). ¹H NMR (400 MHz, Chloroform-*d*) δ 7.71–7.60 (m, 2H), 7.40 (ddd, J = 6.5, 2.8, 1.1 Hz, 2H), 7.35 (dd, J = 3.6, 0.8 Hz, 1H), 7.31 (dd,

J = 5.1, 1.1 Hz, 1H), 7.10 (dd, J = 5.1, 3.6 Hz, 1H), 5.40 (d, J = 47.8 Hz, 2H). ¹³C NMR (101 MHz, Chloroform-*d*) δ 143.85 (d, J = 1.5 Hz), 135.38 (d, J = 17.0 Hz), 135.00 (d,

J = 3.3 Hz), 128.31 (d, J = 5.8 Hz), 128.22, 126.19 (d, J = 1.3 Hz), 125.31, 123.60 (d, J = 0.5 Hz), 84.40 (d, J = 166.2 Hz). ¹⁹F NMR (376 MHz, Chloroform-*d*) δ -206.37 (t, J = 47.8 Hz). HRMS EI (m/z): [M]⁺ calcd. For C₁₁H₉FS: 192.0409 found: 192.0398.



6-(fluoromethyl)-2-methylquinoline (**3u**) was purified with silica gel chromatography (PE/EA = 4:1) as a white solid (72% yield). The compound is known¹¹. ¹H NMR (400 MHz, Chloroform-*d*) δ 8.02 (d, *J* = 8.4 Hz, 2H), 7.75 (s, 1H), 7.65 (d,

J = 8.6 Hz, 1H), 7.29 (d, *J* = 8.4 Hz, 1H), 5.52 (d, *J* = 47.6 Hz, 2H), 2.74 (s, 3H). ¹³C NMR (101 MHz, Chloroform-*d*) δ 159.77 (d, *J* = 1.1 Hz), 147.93 (d, *J* = 2.1 Hz), 136.34 (d, *J* = 0.9 Hz), 133.64 (d, *J* = 17.1 Hz), 129.27, 128.52 (d, *J* = 4.8 Hz), 126.25 (d, *J* = 7.4 Hz), 126.24 (d, *J* = 1.0 Hz), 122.59, 84.38 (d, *J* = 167.2 Hz), 25.52. ¹⁹F NMR (376 MHz, Chloroform-*d*) δ -207.64 (t, *J* = 47.6 Hz).



7-(fluoromethyl)-2-methylquinoline (**3v**) was purified with silica gel chromatography (PE/EA = 4:1) as a white solid (79% yield). ¹H NMR (400 MHz, Chloroform-*d*) δ 8.01 (d, *J* = 8.4 Hz, 1H), 7.97 (s, 1H), 7.77 (d, *J* = 8.3 Hz, 1H), 7.47 (d, *J* = 8.3

Hz, 1H), 7.27 (d, J = 8.8 Hz, 1H), 5.55 (d, J = 47.4 Hz, 2H), 2.73 (s, 3H). ¹³C NMR (101 MHz, Chloroform-*d*) δ 159.68, 147.73, 137.58 (d, J = 17.1 Hz), 136.01, 128.11, 127.10 (d, J = 7.3 Hz), 126.42 (d, J = 1.9 Hz), 124.54 (d, J = 5.3 Hz), 122.55, 84.43 (d, J = 167.6 Hz), 25.47. ¹⁹F NMR (376 MHz, Chloroform-*d*) δ -209.44 (t, J = 47.5 Hz). HRMS ESI (m/z): [M+H]⁺ calcd. For C₁₁H₁₁FN: 176.0876 found: 176.0873.



5-(fluoromethyl)-1-tosyl-1H-indole (3w) was purified with silica gel chromatography (PE/EA = 5:1) as a colorless liquid (87% yield). The compound is known⁵. ¹H NMR (400 MHz, Chloroform-*d*) δ 8.02 (d, *J* = 8.5 Hz, 1H), 7.76 (d, *J* = 8.4 Hz,

2H), 7.60 (d, J = 3.7 Hz, 1H), 7.55 (t, J = 2.0 Hz, 1H), 7.34 (dt, J = 8.5, 1.7 Hz, 1H), 7.21 (d, J = 8.1 Hz, 2H), 6.66 (d, J = 3.6 Hz, 1H), 5.42 (d, J = 48.2 Hz, 2H), 2.33 (s, 3H). ¹³C NMR (101 MHz, Chloroform-*d*) δ 145.23, 135.22, 135.04 (d, J = 2.7 Hz), 131.36 (d, J = 17.2 Hz), 130.99 (d, J = 1.3 Hz), 130.03, 127.21, 126.89, 124.63 (d, J =4.9 Hz), 121.21 (d, J = 6.1 Hz), 113.80 (d, J = 1.1 Hz), 109.10, 84.94 (d, J = 165.8 Hz), 21.65. ¹⁹F NMR (376 MHz, Chloroform-*d*) δ -201.63 (t, J = 48.2 Hz).



3-(fluoromethyl)dibenzo[*b,d*]**furan** (**3x**) was purified with silica gel chromatography (PE) as a white solid (93% yield). ¹H NMR (400 MHz, Chloroform-*d*) δ 8.03–7.92 (m, 2H), 7.62–

7.56 (m, 2H), 7.49 (ddd, J = 8.4, 7.4, 1.3 Hz, 1H), 7.40–7.34 (m, 2H), 5.54 (d, J = 47.8 Hz, 2H). ¹³C NMR (101 MHz, Chloroform-*d*) δ 156.73 , 156.29 (d, J = 0.6 Hz), 135.59 (d, J = 17.1 Hz), 127.59, 124.78 (d, J = 2.7 Hz), 123.90 (d, J = 1.0 Hz), 122.99, 122.24 (d, J = 5.8 Hz), 120.91, 120.85 (d, J = 0.9 Hz), 111.89, 110.99 (d, J = 6.6 Hz), 84.76 (d, J = 167.3 Hz). ¹⁹F NMR (376 MHz, Chloroform-*d*) δ -204.77 (t, J = 47.8 Hz). HRMS EI (m/z): [M]⁺ calcd. For C₁₃H₉FO: 200.0637 found: 200.0630.



3-(fluoromethyl)dibenzo[b,d]thiophene (**3y**) was purified with silica gel chromatography (PE) as a white solid (77% yield). ¹H NMR (400 MHz, Chloroform-*d*) δ 8.21–8.09 (m, 2H), 7.87 (dtd, *J* = 6.6, 3.1, 1.7 Hz, 2H), 7.56–7.38 (m, 3H),

5.52 (d, J = 47.8 Hz, 2H). ¹³C NMR (101 MHz, Chloroform-*d*) δ 139.90 , 139.77 (d, J = 1.0 Hz), 135.96 (d, J = 2.7 Hz), 135.16 (d, J = 1.1 Hz), 134.90 (d, J = 17.1 Hz), 127.09, 124.60, 123.95 (d, J = 5.4 Hz), 122.97, 121.99 (d, J = 6.7 Hz), 121.86, 121.78 (d, J = 0.6 Hz), 84.67 (d, J = 167.2 Hz). ¹⁹F NMR (376 MHz, Chloroform-*d*) δ -205.39 (t, J = 47.8 Hz). HRMS EI (m/z): [M]⁺ calcd. For C₁₃H₉FS: 216.0409 found: 216.0400.



5-(fluoromethyl)-2-methylbenzo[d]oxazole (3z) was purified with silica gel chromatography (PE/EA = 5:1) as a white solid (89% yield). ¹H NMR (400 MHz, Chloroform-*d*) δ 7.66 (s, 1H), 7.47 (d, *J* = 8.3 Hz, 1H), 7.32 (d, *J* = 8.3 Hz, 1H), 5.45 (d, *J* =

48.1 Hz, 2H), 2.64 (s, 3H). ¹³C NMR (101 MHz, Chloroform-*d*) δ 164.79, 151.29 (d, *J* = 2.9 Hz), 141.90 (d, *J* = 1.0 Hz), 132.48 (d, *J* = 17.4 Hz), 124.62 (d, *J* = 5.4 Hz), 119.20 (d, *J* = 5.8 Hz), 110.41, 84.76 (d, *J* = 166.6 Hz), 14.69. ¹⁹F NMR (376 MHz, Chloroform-*d*) δ -201.70 (t, *J* = 48.1 Hz). HRMS ESI (m/z): [M+H]⁺ calcd. For C₉H₉FNO: 166.0668 found: 166.0664.



5-(fluoromethyl)-2-methylbenzo[d]thiazole (3aa) was purified with silica gel chromatography (PE/EA = 5:1) as a white solid (81% yield). ¹H NMR (400 MHz, Chloroform-*d*) δ 7.93 (s, 1H), 7.83 (d, *J* = 8.2 Hz, 1H), 7.37 (d, *J* = 8.2 Hz, 1H), 5.49 (d,

J = 47.9 Hz, 2H), 2.84 (s, 3H). ¹³C NMR (101 MHz, Chloroform-*d*) δ 168.02, 153.62, 136.20 (d, J = 2.8 Hz), 134.41 (d, J = 17.3 Hz), 124.28 (d, J = 5.3 Hz), 121.73 (d, J = 0.8 Hz), 121.62 (d, J = 6.4 Hz), 84.60 (d, J = 167.0 Hz), 20.32. ¹⁹F NMR (376 MHz, Chloroform-*d*) δ -204.70 (t, J = 47.8 Hz). HRMS ESI (m/z): [M+H]⁺ calcd. For C₉H₉FNS: 182.0440 found: 182.0435.



3-(fluoromethyl)-1-tosyl-1H-indole (3ab) was purified with silica gel chromatography (PE/EA = 5:1) as a white solid (65% yield). The compound is known¹². ¹H NMR (400 MHz, Chloroform-*d*) δ 8.00 (d,

 $\begin{array}{c} \textbf{J} = 8.3 \text{ Hz}, 1\text{H}, 7.79 \text{ (d, } J = 8.4 \text{ Hz}, 2\text{H}, 7.66 \text{ (d, } J = 4.7 \text{ Hz}, 1\text{H}), \\ 7.62 \text{ (d, } J = 7.8 \text{ Hz}, 1\text{H}), 7.40-7.33 \text{ (m, } 1\text{H}), 7.32-7.26 \text{ (m, } 1\text{H}), 7.23 \text{ (d, } J = 8.3 \text{ Hz}, \\ 2\text{H}), 5.52 \text{ (d, } J = 48.4 \text{ Hz}, 2\text{H}), 2.34 \text{ (s, } 3\text{H}). \ ^{13}\text{C} \text{ NMR} (101 \text{ MHz}, \text{Chloroform-}d) \delta \\ 145.38, 135.32, 135.18, 130.12, 129.36 \text{ (d, } J = 0.9 \text{ Hz}), 127.02, 125.97 \text{ (d, } J = 9.3 \text{ Hz}), \\ \end{array}$

125.34, 123.71, 119.87, 117.78 (d, J = 19.7 Hz), 113.77, 76.52 (d, J = 162.9 Hz), 21.69. ¹⁹F NMR (376 MHz, Chloroform-*d*) δ -207.22 (td, J = 48.4, 4.8 Hz).



3-(fluoromethyl)quinoline (3ac) was purified with silica gel chromatography (PE/EA = 4:1) as a white solid (81% yield). ¹H NMR (400 MHz, Chloroform-*d*) δ 8.93 (s, 1H), 8.22–8.07 (m, 2H), 7.83 (d, *J* = 8.2 Hz, 1H), 7.75 (t, *J* = 7.6 Hz, 1H), 7.57 (t, *J*

= 7.5 Hz, 1H), 5.58 (d, J = 47.6 Hz, 2H). ¹³C NMR (101 MHz, Chloroform-*d*) δ 149.94 (d, J = 4.7 Hz), 148.31 (d, J = 2.2 Hz), 135.15 (d, J = 6.5 Hz), 130.19 (d, J = 1.0 Hz), 129.51 (d, J = 1.0 Hz), 128.91 (d, J = 17.3 Hz), 128.08 (d, J = 1.2 Hz), 127.62 (d, J = 0.9 Hz), 127.25 (d, J = 0.4 Hz), 82.55 (d, J = 167.7 Hz). ¹⁹F NMR (376 MHz, Chloroform-*d*) δ -208.67 (t, J = 47.6 Hz). HRMS ESI (m/z): [M+H]⁺ calcd. For C₁₀H₁₉FN: 162.0719 found: 162.0716.



5-(fluoromethyl)-2-phenylpyridine (3ad) was purified with silica gel chromatography (PE/EA = 10:1) as a white solid (77% yield). ¹H NMR (400 MHz, Chloroform-*d*) δ 8.70 (s, 1H), 8.09–7.95 (m, 2H), 7.86–7.73 (m, 2H), 7.57–7.39 (m, 3H), 5.44 (d, *J* =

47.7 Hz, 2H). ¹³C NMR (101 MHz, Chloroform-*d*) δ 158.14 (d, *J* = 3.2 Hz), 149.09 (d, *J* = 6.0 Hz), 138.91, 136.49 (d, *J* = 4.8 Hz), 129.95 (d, *J* = 17.5 Hz), 129.39, 128.93, 127.10, 120.43, 82.25 (d, *J* = 167.0 Hz). ¹⁹F NMR (376 MHz, Chloroform-*d*) δ -208.42 (t, *J* = 47.8 Hz). HRMS ESI (m/z): [M+H]⁺ calcd. For C₁₂H₁₁FN: 188.0876 found: 188.0872.



Isopropyl 2-(4-(4-(fluoromethyl)benzoyl)phenoxy)-2-methylpropanoate (3ae) was purified with silica gel chromatography (PE/EA = 5:1) as a white solid (81% yield). The compound is known⁵. ¹H NMR (400 MHz, Chloroform-*d*) δ 7.80-7.71 (m, 4H), 7.50–

7.43 (m, 2H), 6.90–6.82 (m, 2H), 5.46 (d, J = 47.3 Hz, 2H), 5.08 (hept, J = 6.3 Hz, 1H),

1.66 (s, 6H), 1.20 (d, J = 6.3 Hz, 6H). ¹³C NMR (101 MHz, Chloroform-d) δ 195.06, 173.19, 159.76, 140.20 (d, J = 17.2 Hz), 138.37 (d, J = 2.4 Hz), 132.11, 130.48, 130.07, 126.69 (d, J = 6.4 Hz), 117.28, 83.86 (d, J = 168.2 Hz), 79.47, 69.40, 25.44, 21.59. ¹⁹F NMR (376 MHz, Chloroform-*d*) δ -211.76 (t, J = 47.6 Hz).

Mechanistic Studies

1. Radical Inhibition Experiment



To a 5 mL of sealing tube were added aryl bromine **1** (0.2 mmol, 1.0 equiv), NiI₂ (10 mol%), dtbpy (12 mol%), DMAP (24 mol%) and Mn powder (200 mesh, 3.0 equiv). The mixture was added DMAc (1 mL), TEMPO (1 equiv) and **2** (2.5 equiv) under N₂ atmosphere. The tube was sealed with a Teflon lined cap and heated up into a preheated oil bath (40 $^{\circ}$ C) for 48 h. Then the mixture was cooled to room temperature and diluted with ethyl acetate (2 mL) and filtered through a pad of silica gel. The mixture was detected by crude ¹⁹F NMR and GC-MS. No product **3a** was found.

2. Radical Clock Experiment



To a 5 mL of sealing tube were added aryl bromine **1** (0.2 mmol, 1.0 equiv), NiI₂ (10 mol%), dtbpy (12 mol%), DMAP (24 mol%) and Mn powder (200 mesh, 2.0 equiv). The mixture was added DMAc (1 mL), **4** (2 equiv) and **2** (2.5 equiv) under N₂ atmosphere. The tube was sealed with a Teflon lined cap and heated up into a preheated oil bath (40 $^{\circ}$ C) for 24 h. Then the mixture was cooled to room temperature and diluted with ethyl acetate (2 mL) and filtered through a pad of silica gel. The mixture was then concentrated under vacuum and purified with silica gel chromatography to give product

3a in 85% yield and product **5** was yield by crude ¹⁹F NMR and identified by GC-MS. HRMS EI (m/z): [M]+ calcd. For C₁₁H₁₇F: 168.1314 found: 168.1303.



3. Direct Insertion of Bromofluoromethane (2) with Mn⁰

$$\begin{array}{c} \mbox{1)Mn (2 equiv), DMAc,} \\ \mbox{N}_2, 40 \ {}^{\circ}\mbox{C}, 24 \ h \\ \mbox{2)2 M HCl, rt, 30 min} \end{array} \ CH_3 F \\ \mbox{2', N.D.} \end{array}$$

To a 5 mL of sealing tube was added Mn powder (3.0 equiv, 0.6 mmol) in air. The mixture was then added DMAc (1 mL) and **2** (2.5 equiv) under N₂ atmosphere. The tube was sealed with a Teflon lined cap and heated up into a preheated oil bath (40 $^{\circ}$ C) for 24 h. After the mixture was cooled to room temperature, 2 M HCl (1 mL) was added via syringe and the reaction was stirred at room temperature for 30 min. No product **2'** was detected by crude ¹⁹F NMR.

4. Direct Insertion of Bromofluoromethane (2) with Mn^0 at Standard

Condition



To a 5 mL of sealing tube was added dtbpy (12 mol%, 0.024 mmol), DMAP (24 mol%), Mn powder (3.0 equiv) and NiI₂ (10 mol%). The mixture was then added DMAc (1 mL) and **2** (2.5 equiv) under N₂ atmosphere. The tube was sealed with a Teflon lined cap and heated up into a preheated oil bath (40 $^{\circ}$ C) for 24 h. After the mixture was cooled to room temperature, 2 M HCl (1 mL) was added via syringe and the reaction was stirred at room temperature for 30 min. No product **2'** was detected by crude ¹⁹F NMR.

5. Proposed mechanism: a) Radical-cage-rebound process; b) A



radical chain process

References

- 1. E.-M. Kwon, C.-G. Kim, A.-R. Goh, J.-S. Park and J.-G. Jun, *Bull. Korean Chem. Soc.*, 2012, **33**, 1939-1944.
- X. Ji, J. Guo, Y. Liu, A. Lu, Z. Wang, Y. Li, S. Yang and Q. Wang, J. Agric. Food. Chem., 2018, 66, 4062-4072.
- 3. R. J. Tang, T. Milcent and B. Crousse, J. Org. Chem., 2018, 83, 930-938.
- 4. Q. Zhao, L. Lu and Q. Shen, Angew. Chem. Int. Ed., 2017, 56, 11575-11578.
- 5. J. Sheng, H.-Q. Ni, H.-R. Zhang, K.-F. Zhang, Y.-N. Wang and X.-S. Wang, *Angew. Chem. Int. Ed.*, 2018, **57**, 7634-7639.
- 6. J. Hu, B. Gao, L. Li, C. Ni and J. Hu, Org. Lett., 2015, 17, 3086-3089.
- 7. S. Stavber and M. Zupan, J. Org. Chem., 1991, 56, 7347-7350.
- K. G. Kulkarni, B. Miokovic, M. Sauder and G. K. Murphy, *Org. Biomol. Chem.*, 2016, 14, 9907-9911.
- J. Bernstein, J. S. Roth and W. T. Miller, J. Am. Chem. Soc., 1948, 70, 2310-2314.
- 10. A. M. Hua, D. N. Mai, R. Martinez and R. D. Baxter, *Org. Lett.*, 2017, **19**, 2949-2952.
- 11. J. Li, X. Zhang, H. Jin, J. Fan, H. Flores, J. S. Perlmutter and Z. Tu, *J. Med. Chem.*, 2015, **58**, 8584-8600.
- 12. M.-G. Braun, M. H. Katcher and A. G. Doyle, *Chem. Sci.*, 2013, 4, 1216.

¹H, ¹⁹F, and ¹³C NMR Spectra



















-205.93
 -206.06
 -206.19
 -206.19

















3о





















-208.69 -208.81 -208.94



























































-211.63
 -211.76
 -211.88



