Electronic Supporting Information

IBX-mediated oxidative addition of isocyanides to cyclic secondary amines:

total syntheses of alangiobussine and alangiobussinine

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1. General Information. All the reagents and solvents were used as received from commercial sources without further purification. All air and moisture sensitive reactions were conducted under inert atmosphere of nitrogen. Reactions were monitored by thinlayer chromatography carried out on silica plates (silica gel 60 F254, Merck) using uvlight, iodine, ninhydrin and *p*-anisaldehyde for visualization. Column chromatography was carried out using silica gel (100-200 mesh) packed in glass columns. Technical grade ethyl acetate and petroleum ether used for column chromatography were distilled prior to use. ¹H NMR and ¹³C NMR spectra were recorded in CDCl₃ as solvent on 300 MHz or 400 MHz spectrometer at ambient temperature. The coupling constant J is given in Hz. The chemical shifts (δ) are reported in ppm on scale downfield from TMS and using the residual solvent peak in CDCl₃ (H: δ = 7.26 ppm and C: δ = 77.00 ppm) or TMS (δ = (0.00) as internal standard and signal patterns are indicated as follows: s = singlet, d= doublet, dd = doublet of doublet, ddd = doublet of doublet of doublet, dt = doublet of triplet, t = triplet, q = quartet, m = multiplet. High-resolution mass spectra (HRMS) were recorded on a Thermo Scientific Exactive "ORBITRAP" spectrometer using H₂O/MeOH mixed with 0.1% formic acid as mobile phase. 2-Iodoxybenzoic acid (IBX),¹ tryptoline 1a,² 6-bromotryptoline 1b,³ 6-methoxy-1,2,3,4-tetrahydroisoquinoline 6b,⁴ 3,4-dihydro- β carboline 5,⁵ 3,4-dihydrohydroisoquinoline (DHIQ) 17,⁶ N(2)-benzyl-tryptoline 21,⁷ C(1)-**23**.⁸ benzylisocyanide **2d**,⁹ 2-bromobenzylisocyanide phenyl-tryptoline **2e**.¹⁰ **2f**,¹¹ 1-(2-isocyanoethyl)-4-methoxybenzene **2g**,¹² phenethylisocyanide 4-(2isocyanoethyl)-1,2-dimethoxybenzene **2h**,¹³ 1-bromo-2-isocyanobenzene **2k**,¹⁴ 3-(2isocyanoethyl)-1*H*-indole 2m,¹⁵ (3aR,4R,7S,7aS)-2,3,3a,4,7,7a-hexahydro-1H-4,7epoxyisoindole **10**,¹⁶ were prepared and characterized as previously reported.

Isocyanides used in the study:



Table S1. Optimization of the Reaction Conditions^a



Entry	Oxidant (equiv.)	Solvent	Temp (°C)	Time/(h)	3a (% yield) ^b
1.	IBX (2.5)	THF	rt	12	15
2.	IBX (2.5)	THF	60	12	20^{c}
3.	IBX (2.5)	DMSO	rt	2	68
4.	IBX (1.0)	DMSO	rt	12	d
5.	IBX (2.0)	DMSO	rt	12	40^{e}
6.	IBX (2.5)	DCM	rt	5	f
7.	IBX (2.5)	HFIP	rt	12	14^g
8.	PIDA (2.5)	DMSO	rt	5	f
9.	PIFA (2.5)	DMSO	rt	12	f
10.	DMP (2.5)	DMSO	rt	2	60

^{*a*}Reaction conditions: **1a** (0.11 mmol), **2a** (0.11 mmol), oxidant, solvent (1.0 mL). ^{*b*}Isolated yields. ^{*c*}along with **VII** (25%) and **4** (16%). ^{*d*}only oxidized product **5** (75%), ^{*e*}along with **5** (15%). ^{*f*}Complex mixture. ^{*g*}uncharacterized side products. IBX = 2-Iodoxybenzoic acid; DMP = Dess–Martin periodinane; PIDA = Phenyliododiacetate; PIFA = (Bis(trifluoroacetoxy)iodo)benzene; THF = Tetrahydrofuran; DCM = Dichloromethane; HFIP = Hexafluoroisopropanol; DMSO = Dimethyl sulfoxide; IBA = 2-iodosobenzoic acid

We began our investigation by reacting tryptoline 1a with *tert*-butyl isocyanide 2a in the presence of hypervalent iodine containing oxidizing agents (Table S1). Initially, the use of IBX in THF led to the formation of desired carboxamide 3a after 12 h albeit in a low yield of 15% with incomplete consumption of tryptoline 1a (Table S1, entry 1). After heating the reaction mixture at 60 °C for 12 h, product 3a was obtained in 20% yield along with two more byproducts; (i) a complex of 3a with 2-iodosobenzoic acid (IBA) (VII, 25%) and the over oxidized product (4, 16%) (Table S1, entry 2). Change of solvent from THF to DMSO gave encouraging results and tryptoline 1a got consumed at room temperature in 2 h affording 3a in 68% yield (Table S1, entry 3). Reducing the amount of IBX to 1.0 equivalent provided oxidized tryptoline 5 (75%), whereas with 2.0 equivalents of IBX, product 3a was formed in 40% yield with unreacted 5 (15%) (Table 1, entries 4 and 5). However, the reaction did not proceed in DCM and HFIP, and a mixture of uncharactizable products was obtained along with a low yield of 3a of 14% (Table S1, entries 6 and 7).

Screening of different hypervalent iodine reagents as oxidizing agents reflects that with PIDA and PIFA, a complex mixture was obtained whereas DMP favors the reaction to provide **3a** in 60% (Table S1, entries 8–10). It was inferred that IBX (2.5 equiv.) in DMSO provided the best results with complete conversion (Table S1, entry 3). Products **3a**, **4** and **VII** were fully characterized by spectroscopic analysis such as ¹H-, ¹³C-NMR, and HRMS analyses.

A probable complex of product **3a** with IBA was obtained as following structure based on NMR and Mass analysis:



2. Experimental Procedures and characterization data:





IBX (2.5 equiv) was taken in a round bottom flask, then DMSO (2.0 mL) was added and stirred at room temperature for 15 minutes. After complete solubilisation of IBX, tryptoline **1** (1.0 equiv) and isocyanide **2** (1.0 equiv) were added and stirred at room temperature for 2 h. After completion of reaction (based on TLC), the reaction mixture was diluted with ethyl acetate (10 mL) and then saturated solution of sodium thiosulfate (10 mL) was added and stirred for 10 minutes. The organic layer was separated; aqueous layer was again washed with ethyl acetate. The combined organic layers were washed with saturated solution of sodium bicarbonate (15 mL), dried over anhydrous Na₂SO₄, concentrated in *vacuo* and crude was purified by silica gel column chromatography to afford different tryptoline derived iminocarboxamides **3**.

N-(*tert*-butyl)-4,9-dihydro-3*H*-pyrido[3,4-*b*]indole-1-carboxamide (3a)



According to general procedure A by using tryptoline **1a** (50.0 mg, 0.29 mmol), *tert*-butyl isocyanide **2a** (0.04 mL, 0.29 mmol) and IBX (203.2 mg, 0.72 mmol), the titled compound was prepared to yield **3a** as a pale yellow solid (53.5 mg, 68%); **m.p**: 136-137 °C; $R_f = 0.5$ (20% EtOAc in hexane); ¹**H NMR (400 MHz, CDCl3)**: δ 7.57 (d, J = 8.1 Hz, 1H), 7.53 (s, 1H), 7.37 (d, J = 8.3 Hz, 1H), 7.30 – 7.25 (m, 2H), 7.14 – 7.09 (m, 1H), 4.00 (dd, J = 9.5, 8.2 Hz, 2H), 2.94 (dd, J = 9.5, 8.2 Hz, 2H), 1.47 (s, 9H); ¹³**C NMR (100 MHz, CDCl3)**: δ 163.6, 152.0, 136.9, 126.7, 124.9, 124.5, 120.0, 119.8, 117.6, 112.2, 51.0, 48.4, 28.6, 19.1; **IR**

(CHCl₃) v_{max} (cm⁻¹) = 3438, 3020, 2401, 1669, 1525, 1215, 1068, 760, 669; HRMS (ESI): calcd. for C₁₆H₂₀N₃O [M+H]⁺: 270.1606, found: 270.1602.

N-cyclohexyl-4,9-dihydro-3H-pyrido[3,4-b]indole-1-carboxamide (3b)



According to general procedure A by using tryptoline **1a** (50.0 mg, 0.29 mmol), cyclohexyl isocyanide **2b** (0.04 mL, 0.29 mmol) and IBX (203.2 mg, 0.72 mmol), the titled compound was prepared to yield **3b** as a yellow oil (51.5 mg, 60%); $R_f = 0.5$ (20% EtOAc in hexane); **¹H NMR (300 MHz, CDCl₃)**: δ 10.00 (s, 1H), 7.54 (t, J = 9.9 Hz, 2H), 7.38 (d, J = 8.3 Hz, 1H), 7.31 – 7.22 (m, 1H), 7.11 (dd, J = 11.0, 3.9 Hz, 1H), 4.07 – 3.95 (m, 2H), 3.94 – 3.79 (m, 1H), 3.03 – 2.87 (m, 2H), 2.04 – 1.93 (m, 2H), 1.82 – 1.72 (m, 2H), 1.69 – 1.60 (m, 1H), 1.46 – 1.39 (m, 1H), 1.33 (d, J = 12.3 Hz, 2H), 1.26 (dd, J = 7.2, 4.3 Hz, 2H); ¹³C NMR (125 MHz, CDCl₃): δ 163.3, 151.6, 136.9, 126.7, 124.8, 124.6, 119.9, 119.8, 117.4, 112.3, 48.4, 48.0, 32.8, 25.4, 24.7, 19.1; IR (CHCl₃) v_{max} (cm⁻¹) = 3360, 2931, 2854, 1663, 1589, 1523, 1447, 1371, 1319, 1249, 1188, 1066, 872, 756, 637; HRMS (ESI): calcd. for C₁₈H₂₂N₃O [M+H]⁺: 296.1763, found: 296.1754.

N-(2,4,4-trimethylpentan-2-yl)-4,9-dihydro-3*H*-pyrido[3,4-*b*]indole-1-carboxamide (3c)



According to general procedure A by using tryptoline **1a** (50.0 mg, 0.29 mmol), 2-isocyano-2,4,4-trimethylpentane **2c** (0.05 mL, 0.29 mmol) and IBX (203.2 mg, 0.72 mmol), the titled compound was prepared to yield **3c** as a yellow oil (61.4 mg, 65%); $R_f = 0.5$ (20% EtOAc in hexane); ¹H NMR (**400 MHz, CDCl₃**): δ 10.03 (s, 1H), 7.62 (s, 1H), 7.55 (dd, J = 8.0, 0.7 Hz, 1H), 7.36 (d, J = 8.3 Hz, 1H), 7.28 – 7.23 (m, 1H), 7.14 – 7.08 (m, 1H), 3.99 (dd, J = 9.5, 8.2 Hz, 2H), 2.92 (dd, J = 9.5, 8.2 Hz, 2H), 1.83 (s, 2H), 1.51 (s, 6H), 1.03 (s, 9H); ¹³C NMR (100 MHz, CDCl₃): δ 163.3, 151.9, 136.8, 126.7, 124.8, 124.5, 119.8, 117.4, 112.2, 54.7, 51.9, 48.4, 31.5, 28.9, 19.1; IR (CHCl₃) v_{max} (cm⁻¹) = 3743, 3430, 3357, 2952, 1673, 1590, 1523, 1447, 1367, 1320, 1231, 1188, 875, 743, 628, 510; HRMS (ESI): calcd. for C₂₀H₂₈N₃O [M+H]⁺: 326.2232, found: 326.2231.

N-benzyl-4,9-dihydro-3*H*-pyrido[3,4-*b*]indole-1-carboxamide (3d)



According to general procedure A by using tryptoline **1a** (50.0 mg, 0.29 mmol), benzylisocyanide **2d** (0.035 mL, 0.29 mmol) and IBX (203.2 mg, 0.72 mmol), the titled compound was prepared to yield **3d** as a yellow solid (54.0 mg, 61%); **m.p**: 129-130 °C; $R_f = 0.4$ (30% EtOAc in hexane); ¹H NMR (**300** MHz, CDCl₃) δ 9.94 (s, 1H), 7.95 (s, 1H), 7.58 (dd, J = 8.0, 0.9 Hz, 1H), 7.40 (dt, J = 8.3, 0.9 Hz, 1H), 7.37 – 7.32 (m, 4H), 7.32 – 7.26 (m, 2H), 7.12 (ddd, J = 8.0, 7.0, 1.0 Hz, 1H), 4.58 (d, J = 6.2 Hz, 2H), 4.07 – 3.97 (m, 2H), 2.99 – 2.90 (m, 2H); ¹³C NMR (75 MHz, CDCl₃): δ 164.2, 151.3, 137.7, 136.9, 128.8, 127.8, 127.6, 126.6, 125.0, 124.7, 120.1, 119.9, 117.6, 112.3, 48.5, 43.2, 19.1; IR (CHCl₃) v_{max} (cm⁻¹) = 3438, 3020, 1644, 1526, 1216, 761, 669; HRMS (ESI): calcd. for C₁₉H₁₈N₃O [M+H]⁺: 304.1450, found: 304.1445.

N-(2-bromobenzyl)-4,9-dihydro-3*H*-pyrido[3,4-*b*]indole-1-carboxamide (3e)



According to general procedure A by using tryptoline **1a** (50.0 mg, 0.29 mmol), 2bromobenzylisocyanide **2e** (0.035 mL, 0.29 mmol) and IBX (203.2 mg, 0.72 mmol), the titled compound was prepared to yield **3e** as a yellow oil (67.0 mg, 60%); $R_f = 0.4$ (30% EtOAc in hexane); **¹H NMR (300 MHz, CDCl3**): δ 9.90 (s, 1H), 8.13 – 7.98 (m, 1H), 7.58 (ddd, J =8.1, 2.2, 1.1 Hz, 2H), 7.47 – 7.36 (m, 2H), 7.34 – 7.26 (m, 2H), 7.21 – 7.09 (m, 2H), 4.67 (d, J = 6.4 Hz, 2H), 4.04 (dd, J = 9.6, 8.2 Hz, 2H), 2.95 (dd, J = 9.6, 8.3 Hz, 2H); ¹³C NMR (75 MHz, CDCl3): δ 164.2, 151.2, 136.9, 136.8, 132.9, 129.9, 129.3, 127.7, 126.6, 125.0, 124.6, 123.7, 120.1, 119.9, 117.6, 112.3, 48.6, 43.4, 19.1; IR (CHCl3) **v**_{max} (cm⁻¹) = 3368, 2924, 1668, 1522, 1067, 747; HRMS (ESI): calcd. for C₁₉H₁₇BrN₃O [M+H]⁺: 382.0555, found: 382.0555.

N-phenethyl-4,9-dihydro-3*H*-pyrido[3,4-*b*]indole-1-carboxamide (3f)¹⁷



According to general procedure A by using tryptoline **1a** (50.0 mg, 0.29 mmol), (2isocyanoethyl)benzene **2f** (0.035 mL, 0.29 mmol) and IBX (203.2 mg, 0.72 mmol), the titled compound was prepared to yield **3f** as a yellow oil (53.5 mg, 58%); $R_f = 0.5$ (30% EtOAc in hexane); ¹H NMR (400 MHz, CDCl₃): δ 9.97 (s, 1H), 7.71 (s, 1H), 7.58 – 7.54 (m, 1H), 7.38 (d, J = 8.3 Hz, 1H), 7.34 – 7.27 (m, 3H), 7.26 – 7.21 (m, 3H), 7.11 (ddd, J = 8.0, 7.0, 0.9 Hz, 1H), 3.98 (dd, J = 9.5, 8.3 Hz, 2H), 3.68 – 3.60 (m, 2H), 2.96 – 2.87 (m, 4H); ¹³C NMR (100 MHz, CDCl₃): δ 164.3, 151.3, 138.6, 136.9, 128.7, 128.6, 126.6, 126.5, 124.9, 124.6, 120.04, 119.8, 117.5, 112.3, 48.5, 40.4, 35.7, 19.1; IR (CHCl₃) v_{max} (cm⁻¹) = 3438, 3020, 1669, 1528, 1215, 1067, 760, 669; HRMS (ESI): calcd. for C₂₀H2₀N₃O [M+H]⁺: 318.1606, found: 318.1602.

N-(4-methoxyphenethyl)-4,9-dihydro-3*H*-pyrido[3,4-*b*]indole-1-carboxamide (3g)



According to general procedure A by using tryptoline **1a** (50.0 mg, 0.29 mmol), 1-(2isocyanoethyl)-4-methoxybenzene **2g** (46.2 mg, 0.29 mmol) and IBX (203.2 mg, 0.72 mmol), the titled compound was prepared to yield **3g** as a yellow oil (64.6 mg, 64%); $R_f = 0.4$ (30% EtOAc in hexane); ¹H NMR (400 MHz, CDCl₃): δ 9.97 (s, 1H), 7.69 (s, 1H), 7.56 (dd, J = 8.0, 0.6 Hz, 1H), 7.38 (d, J = 8.3 Hz, 1H), 7.31 – 7.24 (m, 1H), 7.18 – 7.09 (m, 3H), 6.89 – 6.83 (m, 2H), 3.99 (dd, J = 9.5, 8.3 Hz, 2H), 3.78 (s, 3H), 3.64 – 3.57 (m, 2H), 2.92 (dd, J = 9.5, 8.3 Hz, 2H), 2.84 (t, J = 7.3 Hz, 2H); ¹³C NMR (100 MHz, CDCl₃): δ 164.2, 158.3, 151.3, 136.9, 130.6, 129.6, 126.6, 124.9, 124.6, 120.0, 119.8, 117.5, 114.1, 112.3, 55.2, 48.5, 40.6, 34.9, 19.1; IR (CHCl₃) v_{max} (cm⁻¹) = 3441, 3019, 2401, 1671, 1591, 1519, 1440, 1215, 1037, 760, 669; HRMS (ESI): calcd. for C₂₁H₂₂N₃O₂ [M+H]⁺: 348.1712, found: 348.1708. *N*-(3,4-dimethoxyphenethyl)-4.9-dihydro-3*H*-pyrido[3,4-*b*]indole-1-carboxamide (3h)



According to general procedure A by using tryptoline **1a** (50.0 mg, 0.29 mmol), 4-(2isocyanoethyl)-1,2-dimethoxybenzene **2h** (55.5 mg, 0.29 mmol) and IBX (203.2 mg, 0.72 mmol), the titled compound was prepared to yield **3h** as a yellow oil (75.0 mg, 68%); $R_f =$ 0.4 (30% EtOAc in hexane); ¹H NMR (400 MHz, CDCl₃): δ 9.96 (s, 1H), 7.72 (s, 1H), 7.56 (dd, J = 8.0, 0.7 Hz, 1H), 7.39 (d, J = 8.3 Hz, 1H), 7.27 (ddd, J = 8.2, 7.0, 1.1 Hz, 1H), 7.12 (ddd, J = 8.0, 7.0, 0.9 Hz, 1H), 6.83 – 6.75 (m, 3H), 3.99 (dd, J = 9.6, 8.3 Hz, 2H), 3.86 (d, J =3.8 Hz, 6H), 3.62 (dd, J = 13.6, 7.0 Hz, 2H), 2.92 (dd, J = 9.6, 8.3 Hz, 2H), 2.85 (t, J = 7.2Hz, 2H); ¹³C NMR (100 MHz, CDCl₃): δ 164.2, 151.3, 148.9, 147.7, 136.9, 131.1, 126.6, 124.9, 124.6, 120.6, 120.1, 119.8, 117.5, 112.3, 111.9, 111.4, 55.9, 55.8, 48.4, 40.5, 35.3, 19.1; **IR (CHCl₃) v_{max} (cm⁻¹) = 3438, 3020, 2401, 1666, 1519, 1215, 1067, 758, 669; HRMS (ESI)**: calcd. for C₂₂H₂₄N₃O₃ [M+H]⁺: 378.1818, found: 378.1814.

N-(2-morpholinoethyl)-4,9-dihydro-3*H*-pyrido[3,4-*b*]indole-1-carboxamide (3i)



According to general procedure A by using tryptoline **1a** (50.0 mg, 0.29 mmol), 4-(2-isocyanoethyl)morpholine **2i** (0.035 mL, 0.29 mmol) and IBX (203.2 mg, 0.72 mmol), the titled compound was prepared to yield **3i** as a yellow oil (52.5 mg, 55%); $R_f = 0.4$ (5% MeOH in DCM); ¹H NMR (400 MHz, CDCl₃): δ 9.93 (s, 1H), 7.90 (s, 1H), 7.58 (d, J = 8.0 Hz, 1H), 7.40 (d, J = 8.3 Hz, 1H), 7.30 – 7.26 (m, 1H), 7.15 – 7.10 (m, 1H), 4.04 (dd, J = 9.5, 8.3 Hz, 2H), 3.77 – 3.72 (m, 4H), 3.51 (q, J = 6.1 Hz, 2H), 2.95 (dd, J = 9.5, 8.3 Hz, 2H), 2.59 (t, J = 6.3 Hz, 2H), 2.55 – 2.48 (m, 4H); ¹³C NMR (100 MHz, CDCl₃): δ 164.4, 151.4, 136.9, 126.7, 124.9, 124.7, 120.1, 119.9, 117.6, 112.3, 66.9, 57.2, 53.4, 48.6, 35.6, 19.1; IR (CHCl₃) v_{max} (cm⁻¹) = 3368, 2925, 1667, 1524, 1450, 1068, 748; HRMS (ESI): calcd. for C₁₈H₂₃N₄O₂ [M+H]⁺: 327.1821, found: 327.1821.

N-(4-methoxyphenyl)-4,9-dihydro-3*H*-pyrido[3,4-*b*]indole-1-carboxamide (3j)



According to general procedure A by using tryptoline **1a** (50.0 mg, 0.29 mmol), 1-isocyano-4-methoxybenzene **2j** (38.6 mg, 0.29 mmol) and IBX (203.2 mg, 0.72 mmol), the titled compound was prepared to yield **3j** as a yellow oil (37.2 mg, 40%); $R_f = 0.4$ (30% EtOAc in hexane); ¹H NMR (400 MHz, CDCl₃): δ 9.97 (s, 1H), 9.42 (s, 1H), 7.63 – 7.57 (m, 3H), 7.41 (d, J = 8.3 Hz, 1H), 7.29 (s, 1H), 7.13 (s, 1H), 6.95 – 6.88 (m, 2H), 4.09 (dd, J = 9.6, 8.3 Hz, 2H), 3.81 (s, 3H), 2.99 (dd, J = 9.6, 8.3 Hz, 2H); ¹³C NMR (125 MHz, CDCl₃): δ 161.7, 156.6, 151.6, 137.1, 130.3, 126.5, 125.2, 124.6, 121.3, 120.2, 119.9, 117.9, 114.3, 112.4, 55.5, 48.5, 19.2; IR (CHCl₃) v_{max} (cm⁻¹) = 3433, 2927, 1673, 1591, 1526, 1447, 1373, 1246, 1161, 1034, 828, 747, 666, 520; HRMS (ESI): calcd. for C₁₉H₁₈N₃O₂ [M+H]⁺: 320.1399, found: 320.1392.

6-bromo-N-(*tert*-butyl)-4,9-dihydro-3*H*-pyrido[3,4-*b*]indole-1-carboxamide (3m)



According to general procedure A by using 6-Bromo-1,2,3,4-tetrahydro-β-carboline **1b** (50.0 mg, 0.2 mmol), *tert*-butyl isocyanide **2a** (38.6 mg, 0.2 mmol) and IBX (139.38 mg, 0.49 mmol), the titled compound was prepared to yield **3m** as a yellow oil (43.0 mg, 62%); $\mathbf{R}_f = 0.6$ (30% EtOAc in hexane); ¹H NMR (**300** MHz, CDCl₃): δ 10.06 (s, 1H), 7.69 (d, J = 1.6 Hz, 1H), 7.51 (s, 1H), 7.33 (dd, J = 8.7, 1.8 Hz, 1H), 7.26 – 7.21 (m, 1H), 4.04 – 3.96 (m, 2H), 2.93 – 2.84 (m, 2H), 1.46 (s, 9H); ¹³C NMR (75 MHz, CDCl₃): δ 163.7, 151.9, 135.6, 127.9, 127.8, 126.5, 122.6, 116.9, 113.9, 113.4, 51.3, 48.5, 28.8, 19.2; **IR (CHCl₃) v_{max} (cm⁻¹)** = 3434, 3359, 3018, 2959, 2926, 2401, 1591, 1526, 1458, 1365, 1314, 1281, 1216, 932, 866, 760, 669, 586, 513; **HRMS (ESI)**: calcd. for C₁₆H₁₉BrN₃O [M+H]⁺: 348.0711, found: 348.0703 .

N-(*tert*-butyl)-2-(3-oxo-1 λ^3 -benzo[d][1,2]iodaoxol-1(3H)-yl)-2,3,4,9-tetrahydro-1Hpyrido[3,4-b]indole-1-carboxamide (VII)



Yellow solid (15.1 mg, 25%); **m.p**: 95-97 °C; $R_f = 0.2$ (20% EtOAc in hexane); ¹H NMR (400 MHz, CDCl₃): δ 11.40 (s, 1H), 7.81 (d, J = 8.1 Hz, 2H), 7.39 (d, J = 3.7 Hz, 3H), 7.31 – 7.26 (m, 1H), 7.20 – 7.14 (m, 2H), 7.06 – 7.01 (m, 1H), 6.14 (s, 1H), 3.84 – 3.78 (m, 2H), 3.54 (t, J = 6.7 Hz, 2H), 1.46 (s, 9H); ¹³C NMR (125 MHz, CDCl₃): δ 176.5, 169.38, 162.2, 142.3, 139.8, 137.4, 130.8, 129.4, 128.1, 128.1, 127.9, 127.3, 121.4, 121.0, 112.8, 92.4, 51.7, 40.6, 29.6, 28.3, 25.0; IR (CHCl₃) v_{max} (cm⁻¹) = 3431, 3020, 2401, 1641, 1514, 1216, 1047, 762, 669; HRMS (ESI): calcd. for C₂₃H₂₅IN₃O₃ [M+H]⁺: 518.0941, found: 518.0935. *N*-(*tert*-butyl)-9*H*-pyrido[3,4-*b*]indole-1-carboxamide (4)



Pale yellow oil (5.0 mg, 16%); $R_f = 0.5$ (20% EtOAc in hexane); ¹H NMR (400 MHz, CDCl₃): δ 10.37 (s, 1H), 8.34 (d, J = 5.1 Hz, 1H), 8.12 (dd, J = 5.4, 2.4 Hz, 2H), 8.06 (dd, J = 5.1, 0.5 Hz, 1H), 7.57 (ddd, J = 8.1, 7.0, 1.1 Hz, 1H), 7.54 – 7.48 (m, 1H), 7.29 (ddd, J = 8.0, 7.0, 1.1 Hz, 1H), 1.57 (s, 9H); ¹³C NMR (100 MHz, CDCl₃): δ 166.0, 141.0, 136.9, 135.4, 132.9, 131.3, 129.1, 121.8, 120.6, 120.1, 117.5, 111.8, 51.1, 28.9; IR (CHCl₃) v_{max} (cm⁻¹) = 3434, 3020,2401, 1657, 1527, 1215, 1048, 761,669; HRMS (ESI): calcd. for C₁₆H₁₈N₃O [M+H]⁺: 268.1450, found: 268.1450.

2.2 General procedure B for the synthesis of 7a-7m



IBX (2.5 equiv) was taken in a round bottom flask, then DMSO (2.0 mL) was added and stirred at room temperature for 15 minutes. After complete solubilisation of IBX, 1,2,3,4-tetrahydrosioquinoline (THIQ) **6** (1.0 equiv) and isocyanide **2** (1.0 equiv) were added and stirred at room temperature for 2 h. After completion of reaction (based on TLC), the reaction mixture was diluted with ethyl acetate (10 mL) and then saturated solution of sodium thiosulfate (10 mL) was added and stirred for 10 minutes. The organic layer was separated and aqueous layer was again washed with ethyl acetate. The combined organic layers were washed with saturated solution of sodium bicarbonate (15 mL), dried over anhydrous Na₂SO₄, concentrated *in vacuo* and the crude was purified by silica gel column chromatography to afford different THIQ derived imino-carboxamides **7**.

N-(*tert*-butyl)-3,4-dihydroisoquinoline-1-carboxamide (7a)¹⁸



According to general procedure B by using THIQ **6a** (50.0 mg, 0.37 mmol), *tert*-butyl isocyanide **2a** (0.035 mL, 0.37 mmol) and IBX (262.7 mg, 0.93 mmol), the titled compound was prepared to yield **7a** as a pale yellow oil (60.6 mg, 71%); $R_f = 0.5$ (20% EtOAc in hexane); ¹H NMR (**300 MHz, CDCl3**): δ 8.18 (dd, J = 7.6, 1.3 Hz, 1H), 7.41 – 7.26 (m, 3H), 7.20 – 7.14 (m, 1H), 3.79 – 3.70 (m, 2H), 2.74 – 2.66 (m, 2H), 1.46 (s, 9H); ¹³C NMR (**75 MHz, CDCl3**): δ 163.7, 160.5, 138.1, 131.1, 128.6, 127.0, 126.8, 126.3, 51.0, 47.1, 28.6,

25.9; **IR (CHCl₃)** v_{max} (cm⁻¹) = 3393, 2925, 1674, 1515, 1247, 1069, 750; HRMS (ESI): calcd. for C₁₄H₁₉N₂O [M+H]⁺: 231.1497, found: 231.1494.

N-cyclohexyl-3,4-dihydroisoquinoline-1-carboxamide (7b)



According to general procedure B by using THIQ **6a** (50.0 mg, 0.37 mmol), cyclohexyl isocyanide **2b** (0.035 mL, 0.37 mmol) and IBX (203.2 mg, 0.93 mmol), the titled compound was prepared to **7b** as a pale yellow solid (65.5 mg, 68%); m.p: 75-78 °C; $R_f = 0.5$ (20% EtOAc in hexane); ¹H NMR (**400 MHz, CDCl3**): δ 8.19 (dd, J = 7.7, 1.0 Hz, 1H), 7.37 (td, J = 7.4, 1.4 Hz, 1H), 7.31 (td, J = 7.6, 1.4 Hz, 2H), 7.17 (dd, J = 7.3, 0.7 Hz, 1H), 3.92 – 3.83 (m, 1H), 3.79 – 3.73 (m, 2H), 2.75 – 2.68 (m, 2H), 2.04 – 1.97 (m, 2H), 1.79 – 1.72 (m, 2H), 1.69 – 1.60 (m, 2H), 1.42 (ddd, J = 13.2, 10.1, 4.1 Hz, 2H), 1.30 – 1.23 (m, 2H); ¹³C NMR (**100 MHz, CDCl3**): δ 163.5, 160.1, 137.9, 131.2, 128.5, 127.1, 126.9, 126.3, 48.1, 47.2, 32.9, 25.8, 25.6, 24.9; **IR (CHCl3) vmax (cm⁻¹)** = 3392, 3016, 2933,2855, 1665, 1517, 1215, 757, 669; **HRMS (ESI)**: calcd. for C₁₆H₂₁N₂O [M+H]⁺: 257.1654, found: 257.1649.

N-(2,4,4-trimethylpentan-2-yl)-3,4-dihydroisoquinoline-1-carboxamide (7c)



According to general procedure B by using THIQ **6a** (50.0 mg, 0.37 mmol), 2-isocyano-2,4,4-trimethylpentane **2c** (52.2 mg, 0.37 mmol) and IBX (262.7 mg, 0.93 mmol), the titled compound was prepared to yield **7c** as a pale yellow oil (66.7 mg, 62%); $R_f = 0.5$ (20% EtOAc in hexane); ¹H NMR (400 MHz, CDCl₃): δ 8.20 (dd, J = 7.7, 1.1 Hz, 1H), 7.42 (s, 1H), 7.32 (dtd, J = 22.2, 7.5, 1.4 Hz, 2H), 7.16 (dd, J = 7.2, 0.7 Hz, 1H), 3.77 – 3.70 (m, 2H), 2.72 – 2.65 (m, 2H), 1.85 (s, 2H), 1.51 (s, 6H), 1.04 (s, 9H); ¹³C NMR (100 MHz, CDCl₃): δ 163.2, 160.2, 138.1, 130.9, 128.6, 126.9, 126.7, 126.3, 54.7, 51.6, 47.1, 31.6, 31.5, 28.9, 25.9; **IR (CHCl₃) v_{max} (cm⁻¹)** = 3377, 2952, 1676, 1611, 1517, 1240, 1066, 755, 605; HRMS (ESI): calcd. for C₁₈H₂₇N₂O [M+H]⁺: 287.2123, found: 287.2116.

N-benzyl-3,4-dihydroisoquinoline-1-carboxamide (7d)



According to general procedure B by using THIQ **6a** (50.0 mg, 0.37 mmol), benzyl isocyanide **2d** (0.027 mL, 0.37 mmol) and IBX (262.7 mg, 0.93 mmol), the titled compound was prepared to yield **7d** as a colourless oil 64.6 mg, 65%); $R_f = 0.4$ (30% EtOAc in hexane); ¹H NMR (400 MHz, CDCl₃): δ 8.24 (dd, J = 7.7, 0.9 Hz, 1H), 7.77 (s, 1H), 7.41 – 7.27 (m, 7H), 7.18 (d, J = 6.7 Hz, 1H), 4.58 (d, J = 6.1 Hz, 2H), 3.75 (dd, J = 8.4, 6.6 Hz, 2H), 2.74 – 2.69 (m, 2H); ¹³C NMR (100 MHz, CDCl₃): δ 164.2, 159.7, 138.1, 137.9, 131.3, 128.7, 128.5, 127.9, 127.5, 127.1, 126.9, 126.3, 47.2, 43.4, 25.8; IR (CHCl₃) v_{max} (cm⁻¹) = 3391, 3018, 1672, 1611, 1520, 1454, 1216, 1064, 765, 669; HRMS (ESI): calcd. for C₁₇H₁₇N₂O [M+H]⁺: 265.1341, found: 265.1337.





According to general procedure B by using THIQ **6a** (50.0 mg, 0.37 mmol), 2-bromo benzyl isocyanide **2e** (73.6 mg, 0.37 mmol) and IBX (262.7 mg, 0.93 mmol) the titled compound

was prepared to yield **7e** as a pale yellow oil (77.60 mg, 60%); $R_f = 0.4$ (30% EtOAc in hexane); ¹H NMR (300 MHz, CDCl₃): δ 8.22 (dd, J = 7.6, 1.1 Hz, 1H), 7.91 (s, 1H), 7.56 (dd, J = 7.9, 1.1 Hz, 1H), 7.45 (dd, J = 7.6, 1.5 Hz, 1H), 7.37 (td, J = 7.4, 1.5 Hz, 1H), 7.34 – 7.26 (m, 2H), 7.20 – 7.11 (m, 2H), 4.66 (d, J = 6.3 Hz, 2H), 3.81 – 3.73 (m, 2H), 2.75 – 2.67 (m, 2H); ¹³C NMR (75 MHz, CDCl₃): δ 164.2, 159.5, 137.9, 137.2, 132.8, 131.3, 130.1, 129.1, 128.5, 127.7, 127.1, 126.9, 126.2, 123.8, 47.2, 43.6, 25.8; IR (CHCl₃) v_{max} (cm⁻¹) = 3394, 3019, 2401, 1674, 1611, 1518, 1215, 1030, 931, 557, 669; HRMS (ESI): calcd. for C₁₇H₁₆BrN₂O [M+H]⁺: 343.0446, found: 343.0433.

N-phenethyl-3,4-dihydroisoquinoline-1-carboxamide (7f)¹⁹



According to general procedure B by using THIQ **6a** (50.0 mg, 0.37 mmol), (2isocyanoethyl)benzene **2f** (49.2 mg, 0.37 mmol) and IBX (262.7 mg, 0.93 mmol), the titled compound was prepared to yield **7f** as a pale yellow oil mixture (64.0 mg, 61%); $R_f = 0.5$ (40% EtOAc in hexane); ¹H NMR (400 MHz, CDCl₃): δ 8.16 (d, J = 7.5 Hz, 1H), 7.52 – 7.46 (m, 1H), 7.36 (dt, J = 7.4, 3.7 Hz, 1H), 7.33 – 7.29 (m, 2H), 7.24 (dd, J = 7.4, 4.3 Hz, 3H), 7.20 – 7.14 (m, 2H), 3.77 – 3.69 (m, 2H), 3.68 – 3.61 (m, 2H), 2.91 (t, J = 7.3 Hz, 2H), 2.73 – 2.66 (m, 2H); ¹³C NMR (100 MHz, CDCl₃): δ 164.3, 159.8, 138.9, 137.9, 131.2, 128.7, 128.5, 128.4, 127.1, 126.9, 126.4, 47.1, 40.6, 35.8, 25.7; IR (CHCl₃) v_{max} (cm⁻¹) = 3400, 3020, 1671, 1522, 1215, 1068, 760, 669; HRMS (ESI): calcd. for C₁₈H₁₉N₂O [M+H]⁺: 279.1497, found: 279.1495.

N-(4-methoxyphenethyl)-3,4-dihydroisoquinoline-1-carboxamide (7g)



According to general procedure B by using THIQ **6a** (50.0 mg, 0.37 mmol), 1-(2-isocyanoethyl)-4-methoxybenzene **2g** (60.5 mg, 0.37 mmol) and IBX (262.7 mg, 0.93 mmol), the titled compound was prepared to yield **7g** as a yellow oil (76.5, 66%); $R_f = 0.5$ (40% EtOAc in hexane); ¹H NMR (400 MHz, CDCl₃): δ 8.15 (dd, J = 7.7, 0.9 Hz, 1H), 7.47 (d, J = 4.4 Hz, 1H), 7.37 (td, J = 7.4, 1.4 Hz, 1H), 7.33 – 7.28 (m, 1H), 7.16 (dd, J = 6.9, 4.6 Hz, 3H), 6.89 – 6.83 (m, 2H), 3.79 (s, 3H), 3.74 (dd, J = 8.9, 6.1 Hz, 2H), 3.65 – 3.57 (m, 2H), 2.85 (t, J = 7.3 Hz, 2H), 2.73 – 2.67 (m, 2H); ¹³C NMR (100 MHz, CDCl₃): δ 164.3, 159.8, 158.2, 137.9, 131.2, 130.9, 129.7, 128.4, 127.1, 126.9, 126.2, 113.9, 55.2, 47.2, 40.8, 34.9, 25.8; IR (CHCl₃) v_{max} (cm⁻¹) =3408, 3020, 1616, 1514, 1216, 1068, 764, 669; HRMS (ESI): calcd. for C₁₉H₂₁N₂O₂ [M+H]⁺: 309.1603, found: 309.1592.

N-(3,4-dimethoxyphenethyl)-3,4-dihydroisoquinoline-1-carboxamide (7h)



According to general procedure B by using THIQ **6a** (50.0 mg, 0.37 mmol), 4-(2isocyanoethyl)-1,2-dimethoxybenzene **2h** (71.8 mg, 0.37 mmol) and IBX (262.7 mg, 0.93 mmol) the titled compound was prepared to yield **7h** as a yellow oil (85.1 mg, 67%); $R_f = 0.5$ (40% EtOAc in hexane); ¹H NMR (400 MHz, CDCl₃): δ 8.16 (dd, J = 7.7, 0.9 Hz, 1H), 7.51 (s, 1H), 7.37 (td, J = 7.4, 1.4 Hz, 1H), 7.31 (td, J = 7.6, 1.3 Hz, 1H), 7.20 – 7.15 (m, 1H), 6.84 – 6.77 (m, 3H), 3.87 (d, J = 5.3 Hz, 6H), 3.76 – 3.70 (m, 2H), 3.62 (dd, J = 13.5, 6.9 Hz, 2H), 2.86 (t, J = 7.1 Hz, 2H), 2.73 – 2.67 (m, 2H); ¹³C NMR (100 MHz, CDCl₃): δ 164.3, 159.8, 148.9, 147.6, 137.8, 131.4, 131.2, 128.3, 127.0, 126.9, 126.2, 120.6, 112.0, 111.4, 55.8, 55.8, 47.1, 40.7, 35.3, 25.7; IR (CHCl₃) v_{max} (cm⁻¹) = 3421, 3020, 1638, 1516,1069, 760, 669; HRMS (ESI): calcd. for C₂₀H₂₃N₂O₃ [M+H]⁺: 339.1709, found: 339.1706.

N-(2-(1*H*-indol-3-yl)ethyl)-3,4-dihydroisoquinoline-1-carboxamide (7i)



According to general procedure B by using THIQ **6a** (50.0 mg, 0.37 mmol), 3-(2-isocyanoethyl)-1*H*-indole **2m** (63.9 mg, 0.37 mmol) and IBX (262.7 mg, 0.93 mmol), the titled compound was prepared to yield **7i** as a pale brown solid (74.0 mg, 62%); m.p: 60-62 °C; $R_f = 0.4$ (40% EtOAc in hexane); ¹H NMR (400 MHz, CDCl₃): δ 8.26 (s, 1H), 8.17 (d, *J* = 7.6 Hz, 1H), 7.64 (d, *J* = 7.8 Hz, 1H), 7.53 (s, 1H), 7.39 – 7.28 (m, 3H), 7.21 – 7.15 (m, 2H), 7.13 – 7.09 (m, 1H), 7.04 (s, 1H), 3.79 – 3.67 (m, 4H), 3.07 (t, *J* = 7.0 Hz, 2H), 2.73 – 2.64 (m, 2H); ¹³C NMR (100 MHz, CDCl₃): δ 164.5, 159.9, 137.9, 136.4, 131.2, 128.4, 127.3, 127.1, 126.9, 126.2, 122.0, 119.3, 118.7, 112.9, 111.2, 47.1, 39.6, 25.7, 25.4; IR (CHCl₃) v_{max} (cm⁻¹) = 3401, 3019, 1669, 1523, 1426, 1216, 1068, 762, 669; HRMS (ESI): calcd. for C₂₀H₂₀N₃O [M+H]⁺: 318.1606, found: 318.1602.

N-(4-methoxyphenyl)-3,4-dihydroisoquinoline-1-carboxamide (7j)



According to general procedure by using THIQ **6a** (50.0 mg, 0.37 mmol), 1-isocyano-4methoxybenzene **2j** (50.0 mg, 0.37 mmol) and IBX (262.7 mg, 0.93 mmol), the titled compound was prepared to yield **7j** as a pale yellow oil (44.2 mg, 42%); $R_f = 0.4$ (30% EtOAc in hexane); ¹H NMR (400 MHz, CDCl₃): δ 9.97 (s, 1H), 9.42 (s, 1H), 7.63 – 7.57 (m, 3H), 7.41 (d, J = 8.3 Hz, 1H), 7.29 (s, 1H), 7.13 (s, 1H), 6.95 – 6.88 (m, 2H), 4.09 (dd, J = 9.6, 8.3 Hz, 2H), 3.81 (s, 3H), 2.99 (dd, J = 9.6, 8.3 Hz, 2H); ¹³C NMR (125 MHz, CDCl₃): δ 161.7, 156.6, 151.6, 137.1, 130.3, 126.5, 125.2, 124.6, 121.4, 120.2, 119.9, 117.9, 114.3, 112.4, 55.5, 48.5, 19.2; ; IR (CHCl₃) v_{max} (cm⁻¹) = 3396, 3020, 1613, 1522, 1215, 1045, 758, 669; HRMS (ESI): calcd. for C₁₇H₁₇N₂O₂ [M+H]⁺: 281.1290, found: 281.1282.

*N-(tert-*butyl)-7-methoxy-3,4-dihydroisoquinoline-1-carboxamide (7m)



According to general procedure B by using 6-methoxy-1,2,3,4-tetrahydroisoquinoline **6b** (50.0 mg, 0.306 mmol), *tert*-butyl isocyanide **2a** (0.03 mL, 0.306 mmol) and IBX (214.4 mg, 0.76 mmol), the titled compound was prepared to yield **7m** as a pale yellow oil (28.0 mg, 35%); $R_f = 0.5$ (30% EtOAc in hexane); ¹H NMR (300 MHz, CDCl₃): δ 7.87 (d, J = 2.7 Hz, 1H), 7.33 (s, 1H), 7.08 (d, J = 8.3 Hz, 1H), 6.93 (dd, J = 8.3, 2.7 Hz, 1H), 3.83 (s, 3H), 3.75 – 3.67 (m, 2H), 2.69 – 2.58 (m, 2H), 1.46 (s, 9H); ¹³C NMR (75 MHz, CDCl₃): δ 163.7, 160.1, 158.3, 130.2, 127.8, 126.9, 117.7, 113.5, 55.5, 50.9, 47.5, 28.6, 25.1; **IR (CHCl₃) v_{max} (cm⁻¹)** = 3401, 3020, 1672, 1518, 1216, 1045, 762, 669; **HRMS (ESI)**: calcd. for C₁₅H₂₁N₂O₂ [M+H]⁺: 261.1603, found: 261.1592.

N-benzyl-1*H*-pyrrole-2-carboxamide (9)



According to general procedure A by using pyrrolidine **8** (100.0 mg, 1.41 mmol), benzylisocyanide **2d** (164.7 mg, 1.41 mmol) and IBX (984.2 mg, 3.52 mmol), the titled compound was prepared to yield **9** as a pale brown sticky oil (40.0 mg, 14%); $R_f = 0.7$ (30% EtOAc in hexane); ¹H NMR (**300 MHz, CDCl3**): δ 9.90 (s, 1H), 7.37 – 7.27 (m, 5H), 6.94 – 6.86 (m, 1H), 6.58 – 6.53 (m, 1H), 6.25 (d, J = 14.4 Hz, 1H), 6.23 – 6.18 (m, 1H), 4.61 (d, J = 5.9 Hz, 2H); ¹³C NMR (**75 MHz, CDCl3**): δ 161.2, 138.4, 128.7, 127.7, 127.5, 125.7, 121.8, 109.7, 108.9, 43.4; **IR (CHCl3)** v_{max} (cm⁻¹) = 3355, 2924, 1638, 1448, 1323, 1217, 1088, 770; HRMS (ESI): calcd. for C₁₂H₁₃N₂O [M+H]⁺: 201.1028, found: 201.1021. (**3aS,4S,7R,7aR)**-*N*-benzyl-**3a,4,7,7a-tetrahydro**-1*H*-**4,7-epoxyisoindole-3-carboxamide**

(11)



According to general procedure A by using pyrrolidine **10** (100.0 mg, 1.41 mmol), benzyl isocyanide **2d** (85.4 mg, 1.41 mmol) and IBX (510.3 mg, 3.52 mmol), the titled compound was prepared to yield **11** as a white solid (60.0 mg, 31%); **m.p**: 124-125 °C; $R_f = 0.3$ (40% EtOAc in hexane); ¹**H NMR (300 MHz, CDCl₃)**: δ 7.43 (s, 1H), 7.37 – 7.26 (m, 5H), 6.44 (dd, J = 5.9, 1.7 Hz, 1H), 6.36 (dd, J = 5.9, 1.6 Hz, 1H), 5.22 (d, J = 1.2 Hz, 1H), 4.77 (d, J = 1.1 Hz, 1H), 4.51 (qd, J = 14.8, 6.0 Hz, 2H), 4.09 (ddd, J = 18.0, 8.7, 1.1 Hz, 1H), 3.79 (dt, J = 18.0, 3.2 Hz, 1H), 3.45 (dd, J = 7.4, 1.7 Hz, 1H), 2.62 (ddd, J = 8.6, 7.5, 3.6 Hz, 1H); ¹³**C NMR (75 MHz, CDCl₃)**: δ 167.6, 161.8, 137.6, 136.7, 136.6, 128.7, 127.8, 127.6, 84.3, 80.9, 63.5, 57.1, 43.9, 43.3; **IR (CHCl₃) v**_{max} (**cm**⁻¹) = 3396, 1624, 1531, 1071; **HRMS (ESI)**: calcd. for C₁₆H₁₇N₂O₂ [M+H]⁺: 269.1290, found: 269.1292.

1*H*-indole (14)



According to general procedure A by using indoline **12** (100.0 mg, 1.41 mmol), benzylisocyanide **2d** (98.3 mg, 1.41 mmol) and IBX (587.4 mg, 3.52 mmol), the titled compound was prepared to yield **14** as a white solid (40.0 mg, 14%); **m.p**: 50-54 °C; $R_f = 0.6$ (20% EtOAc in hexane); ¹H NMR (300 MHz, CDCl₃): δ 8.03 (s, 1H), 7.64 (dd, J = 7.8, 0.7 Hz, 1H), 7.39 – 7.33 (m, 1H), 7.23 – 7.08 (m, 3H), 6.54 (ddd, J = 3.0, 2.0, 0.9 Hz, 1H); ¹³C NMR (75 MHz, CDCl₃): δ 135.8, 127.8, 124.1, 121.9, 120.7, 119.8, 110.9, 102.6; IR (CHCl₃) v_{max} (cm⁻¹) =3414, 1632, 1409, 1217, 1087, 767; HRMS (ESI): calcd. for C₈H₈N [M+H]⁺: 118.0657, found: 118.065.

2.3 Total synthesis of Alangiobussine (I) and Alangiobussinine (II)

2.3.1. Total synthesis of Alangiobussine (I)¹⁷



According to general procedure A by using tryptoline **1a** (50.0 mg, 0.3 mmol), 3-(2isocyanoethyl)-1*H*-indole **2m** (49.4 mg, 0.3 mmol) and IBX (203.2 mg, 0.7 mmol), alangiobussine **I** was obtained as a yellow solid (66.0 mg, 63%); **m.p**: 152-153 °C; $R_f = 0.4$ (40% EtOAc in hexane); ¹**H NMR (400 MHz, CDCl3)**: δ 9.99 (s, 1H), 8.10 (s, 1H), 7.76 (t, *J* = 5.5 Hz, 1H), 7.62 (d, *J* = 7.8 Hz, 1H), 7.56 (d, *J* = 8.0 Hz, 1H), 7.35 (dd, *J* = 17.1, 8.2 Hz, 2H), 7.26 (dd, *J* = 11.3, 3.9 Hz, 1H), 7.23 – 7.16 (m, 1H), 7.11 (t, *J* = 7.5 Hz, 2H), 7.01 (d, *J* = 2.0 Hz, 1H), 4.01 – 3.91 (m, 2H), 3.71 (q, *J* = 6.9 Hz, 2H), 3.05 (t, *J* = 7.1 Hz, 2H), 2.94 – 2.85 (m, 2H); ¹³C **NMR (100 MHz, CDCl3)**: δ 164.3, 151.4, 136.9, 136.3, 127.2, 126.7, 124.9, 124.6, 122.1, 121.9, 120.0, 119.8, 119.4, 118.7, 117.5, 112.8, 112.3, 111.2, 48.4, 39.4, 25.3, 19.1; **IR (CHCl₃)** v_{max} (cm⁻¹) = 3419, 2924, 1663, 1526, 1451, 1074, 769; HRMS (ESI): calcd. for C₂₂H₂₁N₄O [M+H]⁺: 357.1715, found: 357.1706.

2.3.2 Total synthesis of Alangiobussinine (II)²⁰



To a stirred solution of alangiobussine I (50.0 mg, 0.37 mmol) in DMSO (1.5 mL), CuBr₂ (3.1 mg, 0.014 mmol) and DBU (0.02 mL, 0.37 mmol) were added and stirred at rt for 12 h under air. After completion of reaction (based on TLC), the reaction mixture was diluted with ammonia aqueous solution (5% w/w) and dichloromethane. Two phases were separated, and the aqueous phase was twice extracted with dichloromethane. The combined organic layers were separated and dried over Na₂SO₄, concentrated *in vacuo* and the crude was purified by silica gel column chromatography to afford alangiobussinine II as a pale yellow solid (35.0 mg, 70%); m.p: 190-192 °C; $R_f = 0.4$ (40% EtOAc in hexane); ¹H NMR (300 MHz, DMSO d_{δ} : δ 11.76 (s, 1H), 10.82 (s, 1H), 9.03 (t, J = 6.0 Hz, 1H), 8.39 (d, J = 5.0 Hz, 1H), 8.33 (d, *J* = 5.1 Hz, 1H), 8.27 (d, *J* = 7.9 Hz, 1H), 7.82 (d, *J* = 8.2 Hz, 1H), 7.67 (d, *J* = 7.8 Hz, 1H), 7.61 - 7.54 (m, 1H), 7.36 (d, J = 8.0 Hz, 1H), 7.30 - 7.22 (m, 2H), 7.12 - 7.05 (m, 1H), 7.03-6.96 (m, 1H), 3.73 (dd, J = 14.4, 6.7 Hz, 2H), 3.06 (t, J = 7.5 Hz, 2H); ¹³C NMR (75 MHz, **DMSO-** d_6): δ 165.4 141.6, 136.6, 136.3, 134.4, 132.6, 130.6, 128.7, 127.3, 122.6, 121.7, 120.9, 119.9, 119.7, 118.4, 118.2, 117.8, 112.9, 111.8, 111.4, 39.4, 25.4; IR (CHCl₃) v_{max} $(cm^{-1}) = 3431, 3019, 1654, 1528, 1453, 1215, 758, 669; HRMS (ESI): calcd. for C₂₂H₁₉N₄O$ [M+H]⁺: 355.1559, found: 355.1559.





IBX (8.31 g, 1.16 mmol) was taken in a round bottom flask, then DMSO (60.0 mL) was added and stirred at room temperature for 15 minutes. After complete solubilisation of IBX, tryptoline 1 (2.0 g, 11.61 mmol) and 3-(2-isocyanoethyl)-1*H*-indole 2m (1.98 g, 11.61 mmol) were added and stirred at room temperature for 2 h. After completion of reaction (based on TLC), the reaction mixture was diluted with ethyl acetate (15 mL) and saturated solution of sodium thiosulfate (15 mL) was added and stirred for 10 minutes. The organic layer was separated and the aqueous layer was again washed with ethyl acetate. The combined organic layers were washed with saturated solution of sodium bicarbonate (15 mL), dried over anhydrous Na₂SO₄, concentrated in *vacuo* and the crude obtained was used for next step without further purification. The crude was dissolved in DMSO, then CuBr₂ (259.37 mg, 1.16 mmol) and DBU (1.73 mL, 11.61 mmol) were added and stirred for 12 h under air. After completion of reaction (based on TLC), the reaction mixture was diluted with ammonia aqueous solution (5% w/w) and dichloromethane. Two phases were separated, and the aqueous phase was twice extracted with dichloromethane. The combined organic layers were separated and dried over Na₂SO₄, concentrated *in vacuo* and the crude was purified by silica gel column chromatography to afford alangiobussinine II as a pale yellow solid (1.85 g, 45%)

3. Control Experiments

3.1. Reaction of 3,4-dihydro- β -carboline 5 with *tert*-butyl isocyanide 2a:



IBX (205.6 mg, 0.73 mmol) was taken in a round bottom flask, then DMSO (2.0 mL) was added and stirred at room temperature for 15 minutes. After complete solubilisation of IBX, 3,4-dihydro- β -carboline **5** (50.0 mg, 0.29 mmol) and *tert*-butyl isocyanide **2a** (0.03 mL, 0.29 mmol) were added and stirred at room temperature for 2 h. After completion of reaction (based on TLC), the reaction mixture was diluted with ethyl acetate (10 mL) and then saturated solution of sodium thiosulfate (10 mL) was added and stirred for 10 minutes. The organic layer was separated and aqueous layer was again washed with ethyl acetate. The combined organic layers were washed with saturated solution of sodium bicarbonate (15 mL), dried over anhydrous Na₂SO₄, concentrated *in vacuo* and crude was purified by silica gel column chromatography to afford **3a** (54.5 mg, 69%).

3.2. Reaction of 3,4-dihydroisoquinoline 17 with tert-butyl isocyanide 2a:



IBX (266.8 mg, 0.95 mmol) was taken in a round bottom flask, then DMSO (2.0 mL) was added and stirred at room temperature for 15 minutes. After complete solubilisation of IBX, 3,4-Dihydroisoquinoline (DHIQ) **17** (50.0 mg, 0.38 mmol) and *tert*-butyl isocyanide **2a** (0.043 mL, 0.38 mmol) were added and stirred at room temperature for 2 h. After completion of reaction (based on TLC), the reaction mixture was diluted with ethyl acetate (10 mL) and then saturated solution of sodium thiosulfate (10 mL) was added and stirred for 10 minutes.

The organic layer was separated and aqueous layer was again washed with ethyl acetate. The combined organic layers were washed with saturated solution of sodium bicarbonate (15 mL), dried over anhydrous Na₂SO₄, concentrated *in vacuo* and crude was purified by silica gel column chromatography to afford **7a** (60.0 mg, 68%).

3.3. Effect of additives

Table S2.



Reaction Conditions:

1) IBX (81.3 mg, 0.29 mmol) was taken in a round bottom flask, then anhydrous DMSO (1.5 mL) was added and stirred at room temperature for 15 minutes under argon atmosphere. After complete solubilisation of IBX, $H_2^{18}O$ (0.2 mL, 35.0 equiv.) was added, simultaneously **1a** (20.0 mg, 0.116 mmol) and *tert*-butyl isocyanide **2a** (0.014 mL, 0.116mmol) were added and stirred at room temperature for 2 h. After completion of reaction (based on TLC), the reaction mixture was diluted with ethyl acetate (8 mL) and then saturated solution of sodium thiosulfate (8.0 mL) was added and stirred for 10 minutes. The organic layer was separated and aqueous layer was again washed with ethyl acetate. The combined organic layers were washed with saturated solution of sodium bicarbonate (10.0 mL), dried over anhydrous Na₂SO₄, concentrated *in vacuo* and crude was purified by silica gel column chromatography to afford **3a'** (19.5 mg, 62%). The percentage of ¹⁸O

enrichment was examined by mass spectrometry as shown in following Figure S1. The calculated data showed 80% ¹⁸O enrichment of **3a'**. HRMS (ESI): calcd. for $C_{16}H_{20}N_3^{18}O$ [M+H]⁺: 272.1649, found: 272.1642.



1) Figure S1: Observation of ¹⁸O incorporation by HRMS analysis

2) IBX (81.3 mg, 0.29 mmol) was taken in a round bottom flask, then anhydrous DMSO (1.8 mL) was added and stirred at room temperature for 15 minutes under argon atmosphere. After complete solubilisation of IBX, 4 Å MS (50 mg) was added, simultaneously **1a** (20.0 mg, 0.116 mmol) and *tert*-butyl isocyanide **2a** (0.014 mL, 0.116 mmol) were added stirred at room temperature for 2 h. After completion of reaction (based on TLC), the reaction mixture was diluted with ethyl acetate (8 mL) and then saturated solution of sodium thiosulfate (8 mL) was added and stirred for 10 minutes. The organic layer was separated and aqueous layer was again washed with ethyl acetate. The combined organic layers were washed with saturated solution of sodium bicarbonate (10 mL), dried

over anhydrous Na₂SO₄, concentrated *in vacuo* and crude was purified by silica gel column

chromatography to afford **3a** (15.8 mg, 50%).



Figure S2: HRMS analysis of compound 3a (entry 3, Table 1)

3) IBX (81.3 mg, 0.29 mmol) was taken in a round bottom flask, then anhydrous DMSO (2.2 mL) was added and stirred at room temperature for 15 minutes under argon atmosphere. After complete solubilisation of IBX, **1a** (20.0 mg, 0.116 mmol), acetic acid (0.02 ml, 0.116 mmol 1.0 equiv.) and *tert*-butyl isocyanide **2a** (0.013 mL, 0.16 mmol) were added and stirred at room temperature for 2 h. After completion of reaction (based on TLC), the reaction mixture was diluted with ethyl acetate (8 mL) and then saturated solution of sodium thiosulfate (8 mL) was added and stirred for 10 minutes. The organic layer was separated and aqueous layer was again washed with ethyl acetate. The combined organic layers were washed with saturated solution of sodium bicarbonate (10 mL), dried over anhydrous

Na₂SO₄, concentrated *in vacuo* and crude was purified by silica gel column chromatography

to afford **3a** (19.0 mg, 60%).



Figure S3: HRMS analysis of compound 3a (entry 4, Table 1)s

4) IBX (81.3 mg, 0.29 mmol) was taken in a round bottom flask, then anhydrous DMSO (2.2 mL) was added and stirred at room temperature for 15 minutes under argon atmosphere. After complete solubilisation of IBX, **1a** (20.0 mg, 0.116 mmol), benzoic acid (14.18 mg, 0.116 mmol 1.0 equiv.) and *tert*-butyl isocyanide **2a** (0.013 mL, 0.16 mmol) were added and stirred at room temperature for 2 h. After completion of reaction (based on TLC), the reaction mixture was diluted with ethyl acetate (8 mL) and then saturated solution of sodium thiosulfate (8 mL) was added and stirred for 10 minutes. The organic layer was separated and aqueous layer was again washed with ethyl acetate. The combined organic layers were washed with saturated solution of sodium bicarbonate (10 mL), dried over anhydrous

Na₂SO₄, concentrated *in vacuo* and crude was purified by silica gel column chromatography to afford **3a** (18.5 mg, 59%).

3.4. Reaction of N(2)-benzyl-tryptoline 23 with *tert*-butyl isocyanide 2a:



IBX (133.4 mg, 0.47 mmol) was taken in a round bottom flask, then DMSO (2.0 mL) was added and stirred at room temperature for 15 minutes. After complete solubilisation of IBX, N(2)-benzyl-tryptoline **21** (50.0 mg, 0.19 mmol) and *tert*-butyl isocyanide **2a** (0.021 mL, 0.19 mmol) were added and stirred at room temperature for 2 h. After completion of reaction (based on TLC), the reaction mixture was diluted with ethyl acetate (10 mL) and then saturated solution of sodium thiosulfate (10 mL) was added and stirred for 10 minutes. The organic layer was separated and aqueous layer was again washed with ethyl acetate. The combined organic layers were washed with saturated solution of sodium bicarbonate (15 mL), dried over anhydrous Na₂SO₄, concentrated *in vacuo* and crude was purified by silica gel column chromatography to afford compound **22**.

2-benzyl-1,2,3,4-tetrahydro-9*H***-pyrrolo[3,4-***b***]quinolin-9-one (22)²¹: Yellow solid (24.0 mg, 45%); m.p: 178-180 °C; R_f = 0.3 (60% EtOAc in hexane); ¹H NMR (300 MHz, DMSO***d***₆): δ 11.94 (s, 1H), 7.95 – 7.87 (m, 1H), 7.44 (dd,** *J* **= 6.1, 2.4 Hz, 1H), 7.36 (d,** *J* **= 4.0 Hz, 4H), 7.34 – 7.28 (m, 1H), 7.19 (pd,** *J* **= 7.2, 3.7 Hz, 2H), 3.87 (s, 2H), 3.78 (s, 2H), 3.24 (s, 2H); ¹³C NMR (75 MHz, DMSO-** *d***₆): δ 189.7, 150.5, 137.6, 136.1, 128.9, 128.4, 127.3, 123.7, 122.9, 121.9, 120.1, 112.0, 110.3, 61.3, 60.3, 48.5; IR (CHCl₃) v_{max} (cm⁻¹) = 3229, 3020, 2401, 1638, 1477, 1215, 1070, 758, 669; HRMS (ESI): calcd. for C₁₈H₁₇N₂O [M+H]⁺: 277.1341, found: 277.1347.**

3.5. Reaction of C(1)-phenyl-tryptoline 25 with tert-butyl isocyanide 2a:



IBX (140.9 mg 0.5 mmol) was taken in a round bottom flask, then DMSO (2.0 mL) was added and stirred at room temperature for 15 minutes. After complete solubilisation of IBX, C(1)-phenyl-tryptoline **23** (50.0 mg, 0.2 mmol) and *tert*-butyl isocyanide **2a** (0.02 mL, 0.2 mmol) were added and stirred at room temperature for 2 h. After completion of reaction (based on TLC), the reaction mixture was diluted with ethyl acetate (10 mL) and then saturated solution of sodium thiosulfate (10 mL) was added and stirred for 10 minutes. The organic layer was separated and aqueous layer was again washed with ethyl acetate. The combined organic layers were washed with saturated solution of sodium bicarbonate (15 mL), dried over anhydrous Na₂SO₄, concentrated *in vacuo* and crude was purified by silica gel column chromatography to afford **24a** and **24b** respectively.

1-phenyl-9*H***-pyrido[3,4-***b***]indole (24a)²²: White solid (5.0 mg, 10.0%); m.p: 240-241 °C; R_f = 0.6 (60% EtOAc in hexane); ¹H NMR (300 MHz, CDCl₃): \delta 8.58 (d, J = 5.2 Hz, 1H), 8.52 (s, 1H), 8.17 (d, J = 7.9 Hz, 1H), 7.98 (t, J = 1.7 Hz, 1H), 7.95 (t, J = 3.6 Hz, 2H), 7.63 – 7.46 (m, 5H), 7.31 (ddd, J = 8.0, 6.8, 1.3 Hz, 1H); ¹³C NMR (75 MHz, CDCl₃): \delta 143.0, 140.3, 139.7, 138.6, 133.5, 129.8, 129.2, 128.8, 128.5, 128.1, 121.9, 121.8, 120.3, 113.8, 111.5; IR (CHCl₃) v_{max} (cm⁻¹) = 3402, 3020, 1625, 1416, 1215, 1066, 759, 669; HRMS (ESI): calcd. for C₁₇H₁₃N₂ [M+H]⁺: 245.1079, found: 245.1070.**

1-phenyl-4,9-dihydro-3*H*-pyrido[3,4-*b*]indole (24b)²³: Pale yellow solid (20.0 mg, 40%);
m.p: 213-214 °C; R_f = 0.2 (60% EtOAc in hexane); ¹H NMR (300 MHz, CDCl₃): δ 8.15 (s, 1H), 7.78 – 7.69 (m, 2H), 7.65 (d, *J* = 7.9 Hz, 1H), 7.50 (dd, *J* = 6.7, 3.6 Hz, 3H), 7.36 (d, *J* = 8.2 Hz, 1H), 7.30 (dd, *J* = 6.9, 1.1 Hz, 1H), 7.22 – 7.13 (m, 1H), 4.05 (dd, *J* = 9.0, 7.7 Hz, 2H), 2.98 (dd, *J* = 9.1, 7.6 Hz, 2H); ¹³C NMR (75 MHz, CDCl₃): δ 159.3, 137.6, 136.5,

129.9, 128.8, 127.8, 127.7, 125.6, 124.6, 120.4, 120.0, 117.9, 111.9, 48.9, 19.2; **IR (CHCl3)** $\mathbf{v_{max}}$ (cm⁻¹) = 3459, 3020, 1638, 1542, 1215, 1069, 760, 669; **HRMS (ESI)**: calcd. for $C_{17}H_{15}N_2$ [M+H]⁺: 247.1235, found: 247.1226.

4. Real-time NMR Studies:





Spectrum A



3a ⁽⁾

Spectrum B





imine-IBX complex



Procedure: To a dry NMR tube were added tryptoline **1a** (10.0 mg, 0.1 mmol) and DMSO d_6 (0.5 mL) at room temperature. The NMR tube was shaken. At this point, ¹H NMR was recorded giving **spectrum A**.

To a dry NMR tube were added IBX (40.0 mg, mmol) and DMSO- d_6 at room temperature and shaken for 3 minutes. After complete solubilization, compound **1a** (10.0 mg, mmol) was added. The NMR tube was again shaken for 5 minutes, at this point ¹H NMR was recorded giving **spectrum B**.

After the ¹H NMR measurement, *tert*-butyl isocyanide (5.0 mg, mmol) was added to the mixture at room temperature. After addition, NMR tube was slightly shaken and ¹H NMR was recorded at this point giving **spectrum C1**.

Then reaction progress was monitored at 15 minutes interval for 1 h. After that, ¹H NMR was recorded after 2 h for respective time interval, giving **spectrum C2-C6**.

The **spectrum D** was taken for pure isolated compound **3a**.

Summary: Spectrum A represents the ¹H NMR of tryptoline 1a. Spectrum B gave the information about intermediate imine-IBX complex formation; as $C(1)-C\underline{H}_2$ - disappearance of the compound 1a and $C(1)-C\underline{H}$ - appeared more downfield (δ 8.99) region as compared to the 3,4-dihydro- β -carboline $C(1)-C\underline{H}$ - (δ 8.34) proton as shown in spectrum B. As *tert*-butyl isocyanide was added there is appearance of *tert*-butyl peak, $-C\underline{H}_2$ at C4 carbon shifted more shielded region and the -NH of amide peak appeared as shown in spectrum C1. Further progress of reaction was checked at 15 minutes interval of time. These results indicate that as the reaction proceeds, the $-C\underline{H}_2$ at C4 carbon (δ 2.83-2.89); $-N\underline{H}$ of amide, Ar-C<u>H</u> peak intensity goes on increasing while $-C\underline{H}_2$ at C4 carbon (δ 3.16-3.22), $C(1)-C\underline{H}$ -(δ 8.99) peak intensity goes on decreasing as shown in spectrum C2-C6.

5. References

- 1. M. Frigerio, M. Santagostino, S. Sputore, G. Palmisano, J. Org. Chem. 1995, 60, 7272.
- 2. J. Ye, J. Wu, T. Lv, G. Wu, Y. Gao, H. Chen, Angew. Chem., Int. Ed. 2017, 56, 14968.
- 3. N. Liu, C. Sheng, Y. Jiang, J. Tu, Z. Li, G. Dong, S. Wu, CN108623585A, 2018.
- A. Vasudevan, T. Penning, H. Chen, B. Liang, S. Wang, Z. Zhao, D. Chai, L. Yang, Y. Gao, WO2012097682A1, 2012.
- M. W. Smith, Z. Zhou, A. X.Gao, T. Shimbayashi, S. A. Snyder, Org. Lett. 2017, 19, 1004.
- J. Shi, G. Manolikakes, C. H. Yeh, C. A. Guerrero, R. A. Shenvi, H. Shigehisa, P.S. Baran, J. Am. Chem. Soc. 2011, 133, 8014.
- C. M. Adolph, J. Werth, R. Selvaraj, E. C. Wegener, C. Uyeda, J. Org. Chem. 2017, 82, 5959.
- 8. L. N. Wang, S. L. Shen, J. Qu, RSC Adv. 2014, 4, 30733.
- Z. Zhang, B. Huang, G. Qiao, L. Zhu, F. Xiao, F. Chen, B. Fu, Z. Zhang, Angew. Chem. Int. Ed. 2017, 56, 4320.
- 10. K. Kobayashi, Y. Yokoi, T. Nakahara, N. Matsumoto, Tetrahedron 2013, 69, 10304.
- N. Esmati, A. R. Maddirala, N. Hussein, H. Amawi, A. K. Tiwari, P. R. Andreana, Org. Biomol. Chem. 2018, 16, 5332.
- P. Mampuys, H. Neumann, S. Sergeyev, R. V. A. Orru, H. Jiao, A. Spannenberg, B. U. W. Maes, M. Beller, M. ACS Catal. 2017, 7, 5549.
- G. D. Ho, W. Michael Seganish, A. Bercovici, D. Tulshian, W. J. Greenlee, R. Van Rijn,
 A. Hruza, L. Xiao, D. Rindgen, D. Mullins, M. Guzzi, X. Zhang, C. Bleickardt, R.
 Hodgson, *Bioorg. Med. Chem. Lett.* 2012, 22, 2585.
- 14. H. Liu, Y. Fang, S. Y. Wang, S. J. Ji, Org. Lett. 2018, 20, 930.

- J. M. Saya, B. Oppelaar, R. C. Cioc, G. van der Heijden, C. M. L. Vande Velde, R. V. A. Orru, E. Ruijter, *Chem. Commun.*, 2016, **52**, 12482.
- C. de Graaff, L. Bensch, M. J. van Lint, E. Ruijter, R. V. A. Orru, Org. Biomol. Chem., 2015, 13, 10108.
- 17. A. O. Diallo, H. Mehri, L. Iouzalen, M. Plat, Phytochemistry, 1995, 40, 975.
- 18. T. Soeta, S. Fujinami, Y. Ukaji, Y. J. Org. Chem. 2012, 77, 9878.
- 19. G. Qi, Y. Q. Ji, Z. M. A. Judeh, Tetrahedron. 2010, 66, 4195.
- J. Baiget, S. Llona-Minguez, S. Lang, S. P. MacKay, C. J. Suckling, O. Sutcliffe, *Beilstein J. Org. Chem.* 2011, 7, 1407.
- 21. J. F. Carniaux, C. Kan-Fan, J. Royer, H. P. Husson, Tetrahedron Lett. 1997, 38, 2997.
- 22. J. Wu, D. Talwar, S. Johnston, M. Yan, J. Xiao, Angew. Chem. Int. Ed., 2013, 52, 6983.
- 23. A. E. Wendlandt, S. S. Stahl, J. Am. Chem. Soc., 2014, 136, 506.

6. Copies of ¹H and ¹³C NMR Spectra

Figure S5: ¹H NMR of compound 3a



Figure S7: ¹H NMR of compound 3b



Figure S9: ¹H NMR of compound 3c





Figure S13: ¹H NMR of compound 3e



Figure S15: ¹H NMR of compound 3f



Figure S17: ¹H NMR of compound 3g



Figure S19: ¹H NMR of compound 3h



Figure S21: ¹H NMR of compound 3i











f1 (ppm) . 150 . 120 . 110 . 100





Figure S29: ¹H NMR of compound 4







Figure S33: ¹H NMR of compound 7b



Figure S35: ¹H NMR of compound 7c



Figure S37: ¹H NMR of compound 7d



Figure S39: ¹H NMR of compound 7e



Figure S41: ¹H NMR of compound 7f







Figure S45: ¹H NMR of compound 7h



Figure S47: ¹H NMR of compound 7i







Figure S51: ¹H NMR of compound 7m



Figure S53: ¹H NMR of compound 9



Figure S55: ¹H NMR of compound 11

. . .


S61

f1 (ppm) . .

Figure S57: ¹H NMR of compound 14













Figure S61: ¹H NMR of compound II

Figure S63: ¹H NMR of compound 22

Figure S65: ¹H NMR of compound 24a

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Figure S67: ¹H NMR of compound 24b

