Supporting Information

Intercepted dehomologation of aldoses by *N*-heterocyclic carbene catalysis – a novel transformation in carbohydrate chemistry

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Contents

1	Ge	neral scheme					
2	2 General information						
3	An	alytical dehomologation experiments8					
	3.1	Calibration method for reducing sugars8					
	3.2	General procedure for the dehomologation reaction and analysis thereof					
	3.3	Time resolved dehomologation of 3-O-Bn-glucose 1a with catalyst 211					
	3.4	Time resolved dehomologation of 3-O-Bn-mannose 1b with catalyst 212					
	3.5	Screening of reaction conditions with 3-O-Bn-Glucose 1a as starting material13					
	3.6	Time resolved dehomologation of 3-O-Bn-mannose 1b with catalyst 1614					
	3.7	Time resolved dehomologation of 3-O-Bn-glucose 1a with catalyst 1715					
3.8 Time resolved dehomologation of 3-O-Bn-mannose 1b with catalyst 1 catalyst loading							
3.9 Time resolved dehomologation of 3-O-Bn-glucose 1a with catalyst 17 – c							
	3.10	Time resolved dehomologation of 2-O-Bn-arabinose 3 with catalyst 16					
4	Sy	nthesis and characterisation via preparative dehomologation experiments19					
	4.1	1-O-Acetyl-2,3-O-isopropylidene-L-erythrose 7b via preparative dehomologation19					
	4.2	(3S)-β-Acetoxy-γ-butyrolactone 9b via preparative dehomologation20					

	4.3	2-O-Benzyl-D-arabinose 3 via preparative dehomologation21				
	4.4	3,5-O-Diacetyl-2-deoxy-D-ribono-1,4-lactone 4b via preparative dehomologation22				
	4.5 3- <i>0</i> -(4	3,5-O-Diacetyl-2-deoxy-D-ribono-1,4-lactone 4a <i>via</i> preparative dehomologation from 4-nitrophenyl)-D-glucose 1023				
5	Syı	nthesis and characterization of substrates and reference materials				
	5.1	3-O-Benzyl-1,2:5,6-di-O-isopropylidene-α-D-glucofuranose 2125				
	5.2	3-O-Benzyl-D-glucose 1a26				
	5.3	Methyl 6-O-(<i>tert</i> -butyldimethylsilyl)-α-D-mannopyranoside 2327				
	5.4	Methyl 6-O-(<i>tert</i> -butyldimethylsilyl)-3-O-benzyl-α-D-mannopyranoside 2428				
	5.5	Methyl 3-O-benzyl-α-D-mannopyranoside 2529				
	5.6	1,2,4,6-Tetra-O-acetyl-3-O-benzyl-α-D-mannopyranose 27				
	5.7	3-O-Benzyl-α-D-mannose 1b				
	5.8	2-O-Benzyl-D-arabinose 3 ¹⁹				
	5.9	2-Deoxy-D-ribono-1,4-lactone 4a ²⁰				
	5.10	3,4-O-Isopropylidene L-arabinose 6 ²²				
	5.11	3-O-(4-Nitrophenyl)-1,2:5,6-di-O-isopropylidene-α-D-glucofuranose 31				
	5.12	3-O-(4-Nitrophenyl)-D-glucose 10				
6	Syı	nthesis and characterization of precatalysts				
	6.1 and 3	General procedure for the synthesis of cyclohepta[<i>d</i>]thiazole-2-thiones 34, 35, 36, 37 8 (procedure A) ²⁷				
	6.2 15 an	General procedure for the synthesis of cyclohepta[<i>d</i>]thiazole precatalysts 12, 13, 14 d 16 (procedure B) ²⁷				
	6.3	3,4,5-Trimethylthiazol-3-ium iodide 2 ²⁸ 40				
	6.4	3-Ethyl-4,5-dimethylthiazol-3-ium bromide 40 ²⁸ 40				
	6.5	2-Bromocycloheptanone 33 ³¹ 41				
	6.6	3-Phenyl-3,4,5,6,7,8-hexahydro-2 <i>H</i> -cyclohepta[<i>d</i>]thiazole-2-thione 3442				
	6.7	3-Phenyl-5,6,7,8-tetrahydro-4 <i>H</i> -cyclohepta[<i>d</i>]thiazol-3-ium perchlorate 1243				
	6.8	3-(4-Methoxyphenyl)-3,4,5,6,7,8-hexahydro-2H-cyclohepta[d]thiazole-2-thione 35.43				
	6.9 13	3-(4-Methoxyphenyl)-5,6,7,8-tetrahydro-4H-cyclohepta[d]thiazol-3-ium perchlorate				

	6.10	3-(4-Nitrophenyl)-5,6,7,8-tetrahydro-4H-cyclohepta[d]thiazol-3-ium perchlorate 14 45						
	6.11	3-Mesityl-3,4,5,6,7,8-hexahydro-2 <i>H</i> -cyclohepta[<i>d</i>]thiazole-2-thione 37 ²⁷ 46						
	6.12	3-Mesityl-5,6,7,8-tetrahydro-4 <i>H</i> -cyclohepta[<i>d</i>]thiazol-3-ium perchlorate 15 ²⁷ 47						
	6.13 38 ²⁷	3-(2,6-Diisopropylphenyl)-3,4,5,6,7,8-hexahydro-2 <i>H</i> -cyclohepta[<i>d</i>]thiazole-2-thione 47						
	6.14 perch	5.14 3-(2,6-Diisopropylphenyl)-5,6,7,8-tetrahydro-4 <i>H</i> -cyclohepta[<i>d</i>]thiazol-3-ium berchlorate 16 ²⁷ 4						
	6.15	1,3,4-Triphenyl-1,2,4-triazol-1-ium perchlorate 17 ³³ 49						
7	NM	IR spectra of dehomologation products51						
	7.1	1-O-Acetyl-2,3-O-isopropylidene-L-erythrose 7b51						
	7.2	(3 <i>S</i>)-β-acetoxy-γ-butyrolactone 9b52						
	7.3	3,5-O-Diacetyl-2-deoxy-D-ribono-1,4-lactone 4b53						
8	NM	VR spectra of sugar substrates54						
	8.1 21	NMR-spectra of isolated 3-O-benzyl-1,2:5,6-di-O-isopropylidene-α-D-glucofuranose 54						
	8.2	NMR-spectra of isolated 3-O-benzyl-D-glucose 1a55						
	8.3	NMR-spectra of isolated methyl 6- <i>O</i> -(<i>tert</i> -butyldimethylsilyl)- α -D-mannopyranoside 23 56						
	8.4 mann	NMR-spectra of isolated methyl 6- <i>O</i> -(<i>tert</i> -butyldimethylsilyl)-3- <i>O</i> -benzyl-α-D- opyranoside 24						
	8.5	NMR-spectra of isolated methyl 3-O-benzyl-α-D-mannopyranoside 2558						
	8.6	NMR-spectra of isolated 1,2,4,6-tetra-O-acetyl-3-O-benzyl-α-D-mannopyranose 27 59						
	8.7	NMR-spectra of isolated 3-O-benzyl-α-D-mannose 1b60						
	8.8	NMR-spectra of isolated 2-O-benzyl-D-arabinose 361						
	8.9	NMR-spectra of isolated 2-deoxy-D-ribono-1,4-lactone 4a62						
	8.10	NMR-spectra of isolated 3,4-O-isopropylidene L-arabinose 663						
	8.11 glucot	NMR-spectra of isolated 3-O-(4-nitrophenyl)-1,2:5,6-di-O-isopropylidene-α-D- furanose 3164						
	8.12	NMR-spectra of isolated 3-O-(4-nitrophenyl)-D-glucose 1065						

9 NMR spectra of precatalysts66
9.1 NMR-spectra of isolated 3,4,5-trimethylthiazol-3-ium iodide 2
9.2 NMR-spectra of isolated 3-Ethyl-4,5-dimethylthiazol-3-ium iodide 4067
9.3 NMR-spectra of isolated 2-bromocycloheptanone 3368
9.4 NMR-spectra of isolated 3-phenyl-3,4,5,6,7,8-hexahydro-2 <i>H</i> -cyclohepta[<i>d</i>]thiazole-2- thione 34
9.5 NMR-spectra of isolated 3-phenyl-5,6,7,8-tetrahydro-4 <i>H</i> -cyclohepta[<i>d</i>]thiazol-3-ium perchlorate 1270
9.6 NMR-spectra of isolated 3-(4-methoxyphenyl)-3,4,5,6,7,8-hexahydro-2H- cyclohepta[d]thiazole-2-thione 3571
9.7 NMR-spectra of isolated 3-(4-methoxyphenyl)-5,6,7,8-tetrahydro-4H- cyclohepta[d]thiazol-3-ium perchlorate 1372
9.8 NMR-spectra of crude 3-(4-nitroxyphenyl)-3,4,5,6,7,8-hexahydro-2H cyclohepta[d]thiazole-2-thione 36
9.9 NMR-spectra of isolated 3-(4-nitrophenyl)-5,6,7,8-tetrahydro-4 <i>H</i> cyclohepta[<i>d</i>]thiazol-3-ium perchlorate 1474
9.10 NMR-spectra of isolated 3-mesityl-3,4,5,6,7,8-hexahydro-2H-cyclohepta[d]thiazole-2- thione 37
9.11 NMR-spectra of isolated 3-mesityl-5,6,7,8-tetrahydro-4 <i>H</i> -cyclohepta[<i>d</i>]thiazol-3-ium perchlorate 15
9.12 NMR-spectra of isolated 3-(2,6-diisopropylphenyl)-3,4,5,6,7,8-hexahydro-2H cyclohepta[d]thiazole-2-thione 3877
9.13 NMR-spectra of isolated 3-(2,6-diisopropylphenyl)-5,6,7,8-tetrahydro-4 <i>H</i> cyclohepta[<i>d</i>]thiazol-3-ium perchlorate 1678
9.14 NMR-spectra of crude <i>N</i> , <i>N</i> -diphenylbenzohydrazonamide 42
9.15 NMR-spectra of isolated 1,3,4-triphenyl-1,2,4-triazol-1-ium perchlorate 1780
10 References

General scheme 1







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2 General information

All chemicals were used directly from commercial sources and used without further purification. Water-free solvents were available at the institute from a PureSolv EN 1-4 Enclosed solvent drying plant. Light petrol for column chromatography was distilled prior to use. MeCN was dried by refluxing it over CaH₂ for several hours, before collecting it by distillation. Amberlyst 15 was washed with the respective solvent prior to use. TLC analysis for reaction monitoring and analyzing fraction from column chromatography was performed on silica gel 60 F254-plates or HPTLC-plates (silica gel 60 F₂₅₄ with concentration zone 20 \times 2.5 cm). The reverse phase columns LiChrolut® RP18 (100 mg) used for the removal of aliphatic residues in the derivatisation step of the dehomologation experiments were purchased from VWR. The spots were visualized using UV light (254 nm) followed by staining the plates with anisaldehyde solution (180 ml EtOH, 10 ml anisaldehyde, 10 ml H₂SO₄ conc., 2 ml AcOH), potassium permanganate solution (3.0 g KMnO₄, 20.0 g K₂CO₃, 250 mg KOH, 300 ml H₂O) or cerium molybdate solution ("Mostain", 21 g (NH₄)₆Mo₇O₂₄·4 H₂O, 1 g Ce(SO₄)₂ 31 ml H₂SO₄ conc., 500 ml H₂O). NMR spectra were recorded at 297 K in the solvent indicated with an Avance UltraShield 400 and an Avance III HD 600 spectrometer. All spectra were calibrated to the solvent residual peak. Chemical shifts (δ) and coupling constants (J) were expressed in ppm and Hz, respectively. GC analysis was carried out on a Thermo Finnigan Focus GC/DSQ II equipped with a standard capillary column (BGB5, 30 m x 0.25 mm ID, 0.50 µm film) with FID detector. Carrier gas: helium, injector: 230 °C; column flow: 2.0 mL/min; method for quantification: $50-180 \degree C (60 \degree C/min) \rightarrow 180-310 \degree C (20 \degree C/min) \rightarrow 310 \degree C (2 min)$. Accurate mass analysis was obtained on an Agilent 6230 AJS ESI-TOF mass spectrometer with ESI ionization method or (for the cases denoted with a *) Q Exactive Focus, ESI, FIA injection, mobile phase 18% MeCN with 0.1% formic acid. GC-MS spectra were measured on a Thermo Trace 1300 / ISQ LT (single quadrupole MS (EI)) using a standard capillary column BGB 5 (30 m x 0.25 mm ID). Optical rotation was measured on an Anton Paar MCP 500 at the specified conditions, $[\alpha]_D$ values are given in $10^{-1} \text{ deg cm}^2 \text{ g}^{-1}$. Melting points were recorded on a Koflertype Leica Galen III or a BÜCHI Melting Point B-545 with a 40%/90% threshold. For DMSO removal a vacuum concentrator RVC 2-25 CDPLUS with an Alpha 2-4 LDplus freeze-dryer was used. Precatalysts **18** and **19** were prepared according to literature procedure.¹

3 Analytical dehomologation experiments

3.1 Calibration method for reducing sugars

Stock solutions (100 mM) in H₂O of the synthesized carbohydrates **1a**, **3** and **4a** (*vide infra*) as well as erythritol were prepared and aliquots of 40 μ L were taken out, combined and evaporated to dryness. Then, a solution of *O*-methylhydroxyl amine hydrochloride in pyridine (400 μ L, 40 mg/mL) was added and heated to 70 °C for 1 h. Subsequently, a solution of DMAP in pyridine (400 μ L, 1.5 mg/mL) as well as BSTFA (400 μ L, + 1% TMSCl) were added and heated to 70 °C for another 2 h. To this mixture, a solution of methylbenzoate (MB) in EtOAc (800 μ L, 2.5 mM) was added to reach following concentrations: MB 1.00 mM, all other analytes 5.00 mM. Then, 800 μ L were taken out and diluted with a solution of MB in EtOAc (200 μ L, 1.00 mM) leading to a concentration of all analytes of 4.00 mM and keeping the concentration of MB at 1.00 mM. From these, 500 μ L were taken out and diluted in a ratio of 1:1 with a solution of methylbenzoate in EtOAc (1.00 mM), halving the concentration of the analytes while maintaining a concentration of MB of 1.00 mM. This step was repeated three times to generate a total of ten samples with concentrations of 5.00, 4.00, 2.50, 2.00, 1.25, 1.00, 0.63, 0.50, 0.31 and 0.25 mM, which were filtered *via* syringe filter and submitted to analysis by GC.

3.2 General procedure for the dehomologation reaction and analysis thereof



Stock solutions were prepared for all reactions in DMSO of the sugars, the precatalysts and chalcone. A significant dependence on the grain size of the used K₂CO₃ on the reaction outcome was observed. If not noted otherwise, granulated K₂CO₃ (Alfa Aesar, product number A16625) was used. If fine, powdered K₂CO₃ is used, the formed products are not stable in the reaction mixture leading to substantially lower recovery of identified products. A decline in detectable carbohydrate-based substrates and products over time was given throughout the majority of all conducted experiments (see 3.3 to 3.10 for a selection of the timecourses that have been performed. Decomposition of the formed products and not reacted substrates is very likely, given the forcing conditions. The formation of a defined side product, which would explain the loss of material, was not observed *via* TLC, NMR, HPLC-MS or GC (upon derivatization). For

representative values, 20 minutes was chosen as the usually ideal time point for comparison of different reaction parameters.

Transformation with NHCs

The base (0.20, 0.08 or 0.04 equiv.) was charged into a microwave vial. Appropriate amounts of the stock solutions of benzylated-sugar (0.16 mmol, 1.00 equiv.), the precatalyst (0.25, 0.10 or 0.05 equiv.) and the chalcone (2.0 equiv.), as well as the respective solvent (anh., totaling: 2.0 mL) were added. The microwave vial was capped and the reaction mixture was set under Ar atmosphere *via* Schlenk technique. After stirring for 2 min at rt a t0 sample was taken to later correct for the actual initial benzyl hexose concentration. The reaction mixture was heated in a heating block at the temperature and time indicated with samples of roughly 0.2 ml being taken at several time points and were directly processed further or stored at -18 °C.

Processing of aliquots and derivatisation

Exactly 125 μ l from each sample were transferred to an Eppendorf vial, a solution of erythritol in H₂O (60 μ l, 100 mM, 0.012 mmol) was added (as silylation standard) and volatiles were removed in the vacuum concentrator (0.7 mbar, 26.5 °C, ~ 2 h). For each of the samples obtained in this manner the following procedure was applied:

The residue was transferred in MeCN:H₂O=1:4 (2×0.5 mL), onto a LiChrolut® RP18 (100 mg). The vial was once rinsed with MeCN (0.2 ml) onto the column and eluted with MeCN:H₂O 1:4 (0.5 ml). The obtained eluent was concentrated *in vacuo*. Following a modified literature procedure², a solution of *O*-methylhydroxylamin in pyridine (anh., 200 µl, 40 mg/ml) was added to the residue and heated to 70 °C for 1 h. Next, a solution of DMAP in pyridine (anh., 200 µl, 1.5 mg/ml) was added as well as BSTFA (+1% TMSCl, 200 µl). The mixture was stirred at 70 °C for another 2 h. Upon cooling to rt, EtOAc (400 µl) was added, resulting in a maximum concentration of starting material derived components of 10 mM and 6 mM of erythritol. 600 µl were taken out and diluted with a solution of methylbenzoate in EtOAc (600 µl, 2 mM). The sample was filtered *via* syringe filter and subjected to GC analysis. Thereby, a peak area each for methyl benzoate, the silylated erythritol, lactone and the two sugar species (as the sum of the two respective peaks) was obtained. The measured concentration of the analysts to the area of the methyl benzoate with the calibration curve. Finally, the values were corrected for the actual concentration in the t0-sample to consider minor weighing errors.

GC-Chromatogramm after 20 min:



3.3 Time resolved dehomologation of 3-O-Bn-glucose 1a with catalyst 2



Reaction conditions: 1a/2/K₂CO₃/5 = 1.00 : 0.25 : 0.20 : 2.00 (molar ratio)



3.4 Time resolved dehomologation of 3-O-Bn-mannose 1b with catalyst 2



Reaction conditions: **1b/2**/K₂CO₃/**5** = 1.00 : 0.25 : 0.20 : 2.00 (molar ratio)



3.5 Screening of reaction conditions with 3-O-Bn-Glucose 1a as starting material



Reaction conditions: 1a/2/base/5 = 1.00 : 0.25 : 0.20 : 2.00 (molar ratio), 20 min. GC yields based on calibrated GC. ^areactions performed under μ W irradiation. Powdered K₂CO₃ was used in all cases except entry 5 & 6.

Entry	Deviation from standard conditions	1a [%]	3 [%]	4a [%]	Sum [%]
1	none	5	13	56	73
2	Solvent: MeCN ^a	0	4	40	45
3	Solvent: DMF ^a	0	12	56	69
4	Solvent: EtOH ^a	0	4	13	17
5	Base: Li ₂ CO ₃	46	34	16	96
6	Base: DBU	96	0	1	97
7	T = 90 °C / 320 min	30	5	39	74
8	T = 110 °C / 80 min	2	5	33	39
9	T = 150 °C / 10 min	8	4	22	33

Table 1 Screening of reaction conditions

3.6 Time resolved dehomologation of 3-O-Bn-mannose 1b with catalyst 16



Reaction conditions: 1b/16/K₂CO₃/5 = 1.00 : 0.25 : 0.20 : 2.00 (molar ratio)



3.7 Time resolved dehomologation of 3-O-Bn-glucose 1a with catalyst 17



Reaction conditions: **1a/17**/K₂CO₃(powdered)/**5** = 1.00 : 0.25 : 0.20 : 2.00 (molar ratio)



3.8 Time resolved dehomologation of 3-*O*-Bn-mannose 1b with catalyst 16 – different catalyst loading



Reaction conditions: **1b/16**/K₂CO₃/**5** = 1.00 : 0.10 : 0.08 : 2.00 (molar ratio)



Reaction conditions: **1b/16**/K₂CO₃/**5** = 1.00 : 0.05 : 0.04 : 2.00 (molar ratio)



3.9 Time resolved dehomologation of 3-*O*-Bn-glucose 1a with catalyst 17 – different catalyst loading



Reaction conditions: **1a/17**/K₂CO₃(powdered)/**5** = 1.00 : 0.10 : 0.08 : 2.00 (molar ratio)



Reaction conditions: $1a/17/K_2CO_3(powdered)/5 = 1.00 : 0.05 : 0.04 : 2.00$ (molar ratio)



3.10 Time resolved dehomologation of 2-O-Bn-arabinose 3 with catalyst 16



Reaction conditions: 3/16/K₂CO₃/5 = 1.00 : 0.25 : 0.20 : 2.00 (molar ratio)



4 Synthesis and characterisation *via* preparative dehomologation experiments

4.1 1-O-Acetyl-2,3-O-isopropylidene-L-erythrose 7b via preparative dehomologation



Total amounts of 3,4-*O*-isopropylidene L-arabinose **6** (612 mg, 3.2 mmol, 1.0 equiv.), 3,4,5-trimethylthiazolium iodide **2** (205 mg, 0.8 mmol, 0.25 equiv.), K_2CO_3 (89.0 mg, 0.64 mmol, 0.2 equiv.) and chalcone **5** (668 mg, 3.2 mmol, 1.0 equiv.) were equally split into two 20 ml microwave vials and dissolved in MeCN (20 ml each). The vial was three times purged with Ar *via* Schlenk technique. The suspension was heated under microwave irradiation for 15 min at 130 °C (very high absorption). After cooling to rt, TLC showed nearly complete reaction. The solvent was evaporated, and the residue was taken up in DCM and purification of the crude mixture was achieved by column chromatography using SiO₂ (90 g) and a gradient of EtOAc in LP from 15-50%. The product **7a** containing fractions were united, concentrated under reduced pressure for acetylation to yield ~800 mg of material.

Acetylation of lactol 7a:

About 800 mg of erythrose acetonide **7a** were dissolved in pyridine (1.5 ml). Ac₂O (1 ml) was added portionwise under cooling with an ice bath due to exothermic reactions. Then a small portion of DMAP was added. After 3h of stirring, Ac₂O was quenched with MeOH (1 ml). EtOAc (10 ml) were added and the mixture was washed with ice cold HCl solution (1N), until the mixture had turned acidic (monitored *via* pH-paper). The organic layer was washed with NaHCO₃ solution (sat.) until the solution was basic. The organic layer was dried with brine and Na₂SO₄. The solvent was evaporated (90 mbar, 50 °C) yielding a highly viscous liquid. After 16 h, brown crystals (372 mg, 53%) were observed, which was confirmed to be the acetylation product of erythrose acetonide **7b** by NMR.

m.p. 63.8-64.2 °C (EtOAc).

 $\mathbf{R}_{\mathbf{f}}$ (LP:EtOAc = 7:3, anisaldehyde stain): 0.41.

Optical rotation $[\alpha]_D{}^{20} = +108$ (c = 1.0, CHCl₃), (lit.³ $[\alpha]_D{}^{20} = -104.5$, D-form, c = 1.0, MeOH).

¹**H** NMR (400 MHz, CDCl₃) δ 6.16 (s, 1H, H1), 4.86 (dd, J = 5.9, 3.7 Hz, 1H, H3), 4.65 (d, J = 5.9 Hz, 1H, H2), 4.11 (d, J = 10.5 Hz, 1H, H4a), 3.98 (dd, J = 10.5, 3.7 Hz, 1H, H4b), 2.05 (s, 3H, COCH₃), 1.47, 1.32 (2 × s, 2 × 1H, C(CH₃)₂).

¹³C NMR (101 MHz, CDCl₃) δ 169.7 (<u>C</u>OCH₃), 113.0 (<u>C</u>(CH₃)₂), 101.6 (C1), 84.8 (C2), 79.7 (C3), 73.8 (C4), 26.4, 25.0 (2 × C(<u>C</u>H₃)₂), 21.2 (CO<u>C</u>H₃).

GC-MS $t_R = 4.24 \text{ min}$, main fragments 188 (4.1, M-Me⁺), 187 (49, M-Me⁺), 143 (49), 101 (81), 85 (100), 59 (47).

Spectral data in accordance with literature.³





4,6-Benzylidene-D-glucose **8** (268 mg, 1.00 mmol, 1.00 equiv.), 3,4,5-trimethylthiazol-3-ium iodide **2** (63.8 mg, 0.25 mmol, 0.25 equiv.), K_2CO_3 (27.6 mg, 0.20 mmol, 0.20 equiv.) and chalcone **5** (416 mg, 2.00 mmol, 2.00 equiv.) were weighted into a microwave vial and MeCN (12.5 ml) was added. The vial was flushed with Ar *via* Schlenk technique and heated in the microwave for 15 min at 130 °C. Reaction monitoring at this point indicated complete conversion of starting material to a less polar spot, staining differently (starting material with anisaldehyde, product only with KMnO₄). Reaction mixture was evaporated to dryness and taken up in H₂O. The aq. layer was extracted with Et₂O (2 x 50 ml) and evaporated to dryness, yielding 150 mg of a yellow, high viscous oil containing lactone **9a**. This was taken up in pyridine (1.5 ml) and Ac₂O (0.6 ml) added. A small portion of DMAP was added and it was

stirred at rt overnight. The next day, MeOH (0.5 ml) was added to quench excessive reagent. The mixture was diluted with DCM and washed with ice cold 1N HCl, sat. aq. NaHCO₃ and brine, dried over MgSO₄ and solvent evaporated, yielding 100 mg of a yellow-orange oil, according to NMR the desired compound in 80-90% purity. Product was purified *via* flash column chromatography (5 g SiO₂, LP/EtOAc 2:1, 5 ml fractions). Fractions containing the product were pooled and solvent evaporated affording the target compound as a colourless oil (77 mg), that primarily contained the target compound **9b** (59 mg, 41%) as a mixture containing an impurity of 23% EtOAc according to ¹H-NMR.

 $\mathbf{R}_{\mathbf{f}}$ (LP:EtOAc = 1:1, potassium permanganate stain): 0.51.

¹**H NMR** (**400 MHz**, **CDCl**₃) δ 5.40 (ddt, *J* = 6.6, 4.8, 1.6 Hz, 1H, H3), 4.48 (dd, *J* = 11.0, 4.8 Hz, 1H, H4a), 4.34 (dt, *J* = 11.0, 1.2 Hz, 1H, H4b), 2.84 (dd, *J* = 18.4, 6.7 Hz, 1H, H2a), 2.58 (dt, *J* = 18.4, 1.3 Hz, 1H, H2b), 2.07 (s, 2H, COCH₃).

¹³C NMR (101 MHz, CDCl₃) δ 174.6 (C1), 170.4 (<u>C</u>OCH₃), 73.1 (C4), 69.9 (C3), 34.6 (C2), 20.9 (CO<u>C</u>H₃).

Spectral data in accordance with literature.⁴

4.3 2-O-Benzyl-D-arabinose 3 via preparative dehomologation



To avoid upscaling problems 3-*O*-benzyl-D-mannose **1b** (270 mg, 1.00 mmol, 1.00 equiv.), chalcone **5** (417 mg, 2.00 mmol, 2.00 equiv.), dipp thiazolium precatalyst **16** (104 mg, 0.25 mmol, 0.25 equiv.) and K_2CO_3 (27.6 mg, 0.20 mmol, 0.20 equiv.) were partitioned in five identical batches in five 2-5 ml microwave vials. Then, MeCN (2.5 mL) was added, the vial was three times purged with Ar *via* Schlenk technique and the reaction mixture was heated in a heating block at 130 °C for 40 minutes (as after 20 min still significant amounts of starting

material were present according to TLC), when TLC indicated nearly full conversion. A small sample was drawn and derivatized as described in **3.2**, indicating the formation of 78% 2-*O*-benzyl-D-arabinose. The solvent was removed under reduced pressure and the residue was distributed between H₂O (40 ml) and Et₂O (50 ml) and the aqueous phase was washed with Et₂O (2×50 ml). The aqueous phase was lyophilized overnight yielding 211 mg. Purification of the crude mixture was achieved by column chromatography using SiO₂ (12 g) and a gradient of MeOH in DCM from 5-20% and afforded the target compound as a yellowish crystals (161.4 mg), that primarily contained 2-*O*-benzyl-arabionse **3** (152 mg, 63%) as a mixture containing an impurity of 6% lactone **4a** according to ¹H-NMR.

4.4 3,5-*O*-Diacetyl-2-deoxy-D-ribono-1,4-lactone 4b *via* preparative dehomologation



3-*O*-benzyl-D-glucose **1a** (1.35 g, 5.00 mmol, 1.00 equiv.), chalcone **5** (2.08 g, 10.0 mmol, 2.00 equiv.), triphenyl triazolium precatalyst **17** (470 mg, 1.25 mmol, 0.25 equiv.) and K₂CO₃ (powdered, 138 mg, 1.00 mmol, 0.20 equiv.) were weighted into a microwave vial. Then, MeCN (20 mL) was added, the vial was three times purged with Ar *via* Schlenk technique and the reaction mixture was heated under microwave irradiation to 130 °C for 20 min (absorbance high). Then, reaction monitoring *via* TLC (DCM/MeOH 5:1; starting material anisaldehyde stain; product KMnO₄ stain) indicated full conversion of the starting material to the desired product. It was evaporated to dryness, taken up in H₂O (approx. 100 ml), extracted with Et₂O

 $(2 \times 200 \text{ ml})$, the aq. layer was evaporated again to obtain ca. 1.00 g of a crude mixture. This was taken up in pyridine (10 ml), Ac₂O (2.83 ml) added and after the addition of DMAP (spatula) stirred overnight at rt to allow full conversion to the peracetylated compound, as observed with TLC (DCM/MeOH 5:1; LP/EtOAc 1:1). Acetylation was worked up by quenching with MeOH (5 ml), extraction with DCM against 1N HCl (ice cold), sat. aq. NaHCO₃ and brine. The organic layer was dried over MgSO₄ and evaporated to obtain ca. 1.00 g of a dark yellow – orange liquid, according to the NMR the desired product (80-90% pure).

Purification of the crude mixture was achieved by flash column chromatography using SiO_2 (45 g) and a gradient of EtOAc in DCM from 0-100% and afforded the target compound **4b** as a colourless oil (590 mg, 55%) and pure according to ¹H-NMR.

 $\mathbf{R}_{\mathbf{f}}$ (LP:EtOAc = 7:3, potassium permanganate stain): 0.52.

Optical rotation $[\alpha]_D^{20} = -5.3$ (c = 1.25, EtOH), (lit.⁵: -5.3, c = 1.25, EtOH).

¹**H NMR (400 MHz, CDCl**₃) δ 5.26 (dt, *J* = 7.5, 1.9 Hz, 1H, H3), 4.65 (td, *J* = 3.5, 1.7 Hz, 1H, H4), 4.36 (dd, *J* = 12.3, 3.4 Hz, 1H, H5a), 4.26 (dd, *J* = 12.3, 3.6 Hz, 1H, H5b), 2.98 (dd, *J* = 18.7, 7.5 Hz, 1H, H2a), 2.59 (dd, *J* = 18.7, 2.1 Hz, 1H, H2b), 2.09, 2.07 (2 × s, 2 × 3H, 2 × COCH₃).

¹³C NMR (101 MHz, CDCl₃) δ 173.9 (C1), 170.4, 170.1 (2 × <u>C</u>OCH₃), 82.1 (C4), 71.2 (C3), 63.4 (C5), 34.9 (C2), 20.9, 20.7 (2 × CO<u>C</u>H₃).

Spectral data in accordance with literature.⁶

4.5 3,5-*O*-Diacetyl-2-deoxy-D-ribono-1,4-lactone 4a *via* preparative dehomologation from 3-*O*-(4-nitrophenyl)-D-glucose 10



3-O-(4-nitrophenyl)-D-glucose **10** (0.300 g, 1.00 mmol, 1.00 equiv.), 3-ethyl-4,5dimethylthiazolium bromide **40** (44 mg, 0.20 mmol, 0.20 equiv.), K₂CO₃ (28 mg, 0.20 mmol, 0.20 equiv.) and chalcone **5** (414 mg, 2.00 mmol, 2.00 equiv.) were added to a microwave vial and heated at 130 °C for 15 min under microwave irradiation. This led to full conversion to the desired product and therefore the solvent was evaporated. Purification of the crude mixture was achieved by flash column chromatography using SiO₂ (90 g) and a gradient of MeOH in EtOAc from 0-10% and afforded the target compound **4a** as a slightly yellow oil (70 mg, 53%) containing 6% of *p*-nitrophenol as an impurity according to ¹H-NMR.

5 Synthesis and characterization of substrates and reference materials



5.1 3-O-Benzyl-1,2:5,6-di-O-isopropylidene-α-D-glucofuranose 21

Diacetone glucose 20 (20.42 g, 78.5 mmol, 1.0 equiv.) was dissolved in dry DMF (80 mL) and a continuous flow of argon was applied to the reaction flask. The reaction mixture was stirred and cooled in an ice bath, before NaH (9.22 g, 60 % in paraffin oil, 231 mmol, 2.9 equiv.) was added portionwise and the formation of H₂ gas was observed. After stirring at 0 °C for 20 min, benzyl bromide (11.2 mL, 94.1 mmol, 1.2 equiv.) was added, so that the reaction temperature did not rise above 20 °C. During that process, a white solid material was formed and more DMF (10 mL) was added, so that the reaction mixture could be stirred more efficiently. The reaction mixture was allowed to reach rt and TLC (LP:EtOAc 6:1) confirmed the clean formation of the target compound 21. Foaming occurred, as MeOH (10 mL) was added carefully to quench the reaction. The mixture was distributed between aqueous NH₄Cl (10 %, 300 mL) and Et₂O (300 mL) and the aqueous phase was then extracted with Et₂O (2×150 mL, 1×100 mL). The pooled organic phases were washed with water $(3 \times 150 \text{ mL})$, brine $(1 \times 150 \text{ mL})$, dried over Na₂SO₄ and filtered. The solvent was evaporated and crude material was subsequently distributed between acetonitrile (200 mL) and n-hexane (4×50 mL), to remove the paraffin oil, which has been introduced with the NaH. The combined hexane phases were extracted with acetonitrile $(2 \times 50 \text{ mL})$ and the collected acetonitrile fractions were evaporated to obtain target product **21** as a viscous syrup (27.12 g, 98%), pure according to ¹H-NMR.

 $\mathbf{R}_{\mathbf{f}}$ (LP:EtOAc = 4:1, anisaldehyde stain): 0.31.

Optical rotation $[\alpha]_D{}^{20} = -28$ (c = 1.0 in EtOH), (lit.⁷ $[\alpha]_D{}^{28} = -28.3$, c = 1.3, EtOH).

¹**H** NMR (400 MHz, CDCl₃) δ 7.38 – 7.26 (m, 5H, PhH), 5.90 (d, *J* = 3.7 Hz, 1H, H1), 4.69 (d, *J* = 11.8 Hz, 1H, Ph-C<u>H</u>H), 4.64 (d, *J* = 11.9 Hz, 1H, Ph-CH<u>H</u>), 4.59 (d, *J* = 3.7 Hz, 1H, H2), 4.38 (dt, *J* = 7.7, 6.0 Hz, 1H, H5), 4.16 (dd, *J* = 7.7, 3.1 Hz, 1H, H4), 4.12 (dd, *J* = 8.6, 6.2 Hz, 1H, H6a), 4.03 (d, *J* = 3.1 Hz, 1H, H3), 4.01 (dd, *J* = 8.6, 5.9 Hz, 1H, H6b), 1.50, 1.43, 1.38, 1.31 (4 × s, 4 × 3H, 4 × CH₃).

¹³C NMR (101 MHz, CDCl₃) δ 137.7 (PhC1), 128.5 (PhC2/6 or PhC3/5), 127.9 (PhC4), 127.7 (PhC2/6 or PhC3/5), 111.9, 109.1 (2 × <u>C</u>(CH₃)₂), 105.4 (C1), 82.7 (C2), 81.8 (C3), 81.4 (C4), 72.6 (C5), 72.5 (Ph-CH₂), 67.5 (C6), 27.0, 26.9, 26.4, 25.6 (4 × CH₃).

Spectral data in accordance with literature.⁸

5.2 3-O-Benzyl-D-glucose 1a



Procedure:

3-*O*-benzyl-1,2:5,6-di-*O*-isopropylidene- α -D-glucofuranose **21** (6.13 g, 17.5 mmol, 1 equiv.) was stirred in H₂O (31 ml) and heated to 70 °C. Amberlyst 15 (hydrogen form, 12.3 g) was first washed with water (3x 80 mL) and then added to the reaction mixture. TLC (3:2 EtOAc/LP for the starting material, 6:1 DCM/MeOH for the product) showed after approx. 4.5 h complete reaction. The liquid phase was vacuum-filtered over Celite[®] resulting in 4 fractions of each 100 mL containing product. The remaining residue was washed with water. The united aqueous fractions were extracted once with Et₂O (250 mL) to remove lipophilic traces. The aqueous solution was then lyophilized yielding the target compound **1a** as a colourless solid (4.20 g, 89%).

m.p. 133.7-136.6 °C (H₂O), (lit.⁹ 135-137 °C).

 $\mathbf{R}_{\mathbf{f}}$ (DCM:MeOH = 4:1, anisaldehyde stain): 0.46.

Optical rotation $[\alpha]_D{}^{20} = +42$ (c = 1.0, H₂O, 12 h), (lit.⁷ $[\alpha]_D{}^{29} = +41.3$, c = 0.9, H₂O).

¹**H** NMR (400 MHz, DMSO-*d*₆) beta-anomer: δ 7.41 (d, *J* = 7.0 Hz, 2H, PhH2/H6), 7.31 (t, *J* = 7.3 Hz, 2H, PhH3/H5), 7.24 (t, *J* = 7.3 Hz, 1H, PhH4), 6.68 (d, *J* = 6.4 Hz, 1H, OH1), 5.09 – 5.04 (m, 2H, OH2, OH4), 4.81 (d, *J* = 11.7 Hz, 1H, Ph-C<u>H</u>H), 4.77 (d, *J* = 11.7 Hz, 1H, Ph-CH<u>H</u>), 4.52 (t, *J* = 5.8 Hz, 1H, OH6), 4.33 (app. t, *J* = 7.0 Hz, 1H, H1), 3.68 (ddd, *J* = 11.7, 5.5, 2.1 Hz, 1H, H6a), 3.45 (dt, *J* = 11.8, 6.0 Hz, 1H, H6b), 3.28 – 3.17 (m, 2H, H4, H3), 3.16 – 3.03 (m, 2H, H5, H2).

¹³C NMR (101 MHz, CDCl₃) beta-anomer: δ 139.7 (PhC1), 127.9 (PhC3/C5), 127.4 (PhC2/C6), 127.0 (PhC4), 96.9 (C1), 85.3 (C3), 76.7 (C5), 74.8 (C2), 73.6 (Ph-<u>C</u>H₂), 69.9 (C4), 61.1 (C6).

HRMS (ESI⁺) Exact mass calc. for $C_{13}H_{18}O_6Na$ [M+Na]⁺: 293.0996, found: 293.0995 (- 0.11 ppm).

Spectral data in accordance with literature.⁸

5.3 Methyl 6-O-(tert-butyldimethylsilyl)-α-D-mannopyranoside 23



Procedure adapted from literature protocol. ¹⁰

Methyl- α -D-mannopyranoside **22** (10.0 g, 51.5 mmol, 1.00 equiv.) was dispersed in MeCN (anh., 180 ml) and DABCO (6.64 g, 59.6 mmol, 1.15 equiv.) was added at rt. The mixture was cooled to 5 °C *via* ice bath. TBDMSCl (8.54 g, 56.6 mmol, 1.10 equiv.) was added dropwise and the mixture was stirred for 18 h and when TLC (DCM:MeOH 6:1) indicated still incomplete conversion further DABCO (1.44 g, 12.9 mmol, 0.25 equiv.) was added at rt. Again, the mixture was cooled to 5 °C *via* ice bath, and then TBDMSCl (1.55 g, 10.3 mmol, 0.20 equiv.) was added dropwise. The mixture was stirred for 1 h when TLC (DCM:MeOH 6:1) showed complete conversion. MeOH (12.5 ml, 6 equiv.) was added and the solution was stirred for 30 min and concentrated *in vacuo*. The residue was taken up in H₂O (200 ml) and extracted with DCM (600 ml) until no further product was detected in the extract using TLC. Then the organic layer was dried with Na₂SO₄, concentrated *in vacuo* and crystallized overnight. The crude yield was 15.65 g (98.5%) of a colourless solid. An aliquot of 7.30 g of crude product was purified *via* column chromatography using SiO₂ (180 g) and a gradient of MeOH in DCM from 0-20% and afforded the target compound **23** as a colourless solid (4.86 g, 82%) and pure according to ¹H-NMR.

m.p. 102.8 – 103.1 °C (DCM), (lit.¹¹ 98-100 °C).

 $\mathbf{R}_{\mathbf{f}}$ (DCM:MeOH = 7:1, anisaldehyde stain): 0.60.

Optical rotation $[\alpha]_D^{20} = +49.0$ (c 1.0, CHCl₃), (lit.¹² $[\alpha]_D^{23} = +57$, c 0.3, CHCl₃).

¹**H** NMR (600 MHz, MeOD) δ 4.61 (d, J = 1.6 Hz, 1H, H1), 3.98 (dd, J = 11.1, 2.0 Hz, 1H, H6a), 3.81 - 3.74 (m, 2H, H2, H6b), 3.64 (dd, J = 9.1, 3.5 Hz, 1H, H3), 3.53 (t, J = 9.5 Hz, 1H, H4), 3.51 - 3.45 (m, 1H, H5), 3.36 (s, 3H, OMe), 0.92 (s, 9H, C(CH₃)₃), 0.10 (d, J = 2.1 Hz, 6H, Si(CH₃)₂).

¹³C NMR (101 MHz, MeOD) δ 102.6 (C1), 75.0 (C5), 72.8 (C3), 72.0 (C2), 68.8 (C4), 64.6 (C6), 55.0 (OMe), 26.4 (C(<u>C</u>H₃)₃), 19.2 (Si(CH₃)₂), -5.1 (<u>C</u>(CH₃)₃).

HRMS (ESI⁺) Exact mass calc. for $C_{13}H_{28}O_6SiNa$ [M+Na]⁺: 331.1547, found: 331.1566 (5.51 ppm).

Spectral data in accordance with literature.¹¹

5.4 Methyl 6-O-(tert-butyldimethylsilyl)-3-O-benzyl-α-D-mannopyranoside 24



Methyl 6-*O*-(*tert*-butyldimethylsilyl)- α -D-mannopyranoside **23** (10.0 g, 32.4 mmol, 1.00 equiv.), Ag₂O (8.26 g, 35.7 mmol, 1.10 equiv.) and 2-aminoethyl diphenylborinate (730 mg, 3.2 mmol, 0.1 equiv.) were dispersed in MeCN (325 ml, anh.). The flask was purged with Ar, benzylbromide (8.32 g, 48.6 mmol, 1.5 equiv.) was added slowly and the solution was heated to 40 °C. The mixture was stirred vigorously for 28 h when TLC (LP:EtOAc = 7:3, anisaldehyde stain) indicated complete conversion. The suspension was filtered through Celite[®], rinsed with DCM (400 ml) until no further product was detected in the extract using TLC (LP:EtOAc = 7:3, anisaldehyde stain). The yellow solution was dried over Na₂SO₄ and concentrated *in vacuo*. Purification of the crude mixture was achieved by column chromatography using SiO₂ (300 g) and a gradient of EtOAc in LP from 5-25% and afforded the target compound **24** as a colourless solid (9.12 g, 71%) and pure according to ¹H-NMR.

m.p. 84.3 – 85.5 °C (EtOAc).

 $\mathbf{R}_{\mathbf{f}}$ (LP:EtOAc = 1:1, anisaldehyde stain): 0.57.

Optical rotation $[\alpha]_D^{20} = +30.4$ (c 1.0, CHCl₃), (lit.¹³: +34.6, c = 0.1, CHCl₃).

¹**H** NMR (600 MHz, MeOD) δ 7.43 (d, *J* = 7.0 Hz, 2H, PhH2/H6), 7.32 (t, *J* = 7.5 Hz, 2H, PhH3/H5), 7.26 (t, *J* = 7.4 Hz, 1H, PhH4), 4.71 (d, *J* = 11.8 Hz, 1H, PhC<u>H</u>H), 4.64 (d, *J* = 11.3 Hz, 1H, PhCH<u>H</u>), 4.63 (s, 1H, H1), 3.98 (dd, *J* = 11.1, 2.0 Hz, 1H, H6a), 3.94 (dd, *J* = 3.2, 1.8 Hz, 1H, H2), 3.78 (dd, *J* = 11.1, 6.7 Hz, 1H, H6b), 3.70 (t, *J* = 9.7 Hz, 1H, H4), 3.56 (dd, *J* = 9.3, 3.3 Hz, 1H, H3), 3.51 (ddd, *J* = 9.5, 6.7, 1.9 Hz, 1H, H5), 3.35 (s, 3H, OMe), 0.92 (s, 9H, C(CH₃)₃), 0.10 (d, *J* = 2.1 Hz, 6H, Si(CH₃)₂).

¹³C NMR (101 MHz, MeOD) δ 139.98 (PhC1), 129.26 (PhC3/C5), 129.06 (PhC2/C6), 128.59 (PhC4), 102.57 (C1), 80.54 (C3), 75.03 (C5), 72.65 (Ph<u>C</u>H₂), 68.88 (C2), 67.85 (C4), 64.60 (C6), 55.05 (OMe), 26.41 (C(CH₃)₃), 19.23 (Si(CH₃)₂), -5.12 (C(CH₃)₃).

HRMS (ESI⁺) Exact mass calc. for $C_{20}H_{34}O_6SiNa$ [M+Na]⁺: 421.2017, found: 421.2035 (4.34 ppm).

Spectral data in accordance with literature.¹⁴

5.5 Methyl 3-*O*-benzyl-α-D-mannopyranoside 25



Methyl 3-*O*-Bn-6-TBDMS-mannopyranoside **24** (5.00 g, 12.5 mmol, 1.00 equiv.) and tetrabutylammonium fluoride solution (14.3 ml, 14.3 mmol, 1.14 equiv., 1 M in THF) in THF (71 ml, anh.) were left stirring at rt under Ar. After 1 h TLC (DCM:MeOH = 6:1, anisaldehyde stain) indicated complete conversion and the reaction mixture was concentrated *in vacuo*. The crude yield was 3.34 g of a colourless oil. Purification of the crude mixture was achieved by column chromatography using solid loading (10 g SiO₂) on SiO₂ (90 g) and a gradient of MeOH in DCM from 5-25% and afforded the target compound **25** as a colourless oil (2.49 g, 70%) and pure according to ¹H-NMR.

 $\mathbf{R}_{\mathbf{f}}$ (DCM:MeOH = 7:1, anisaldehyde stain): 0.58.

Optical rotation $[\alpha]_D^{20} = +35.9$ (c = 1.0, EtOH), (lit.¹⁵ +35.2, c = 1, EtOH).

¹**H** NMR (600 MHz, MeOD) δ 7.43 (d, J = 7.0 Hz, 2H, PhH2/H6), 7.33 (t, J = 7.5 Hz, 2H, PhH3/H5), 7.26 (t, J = 7.3 Hz, 1H, PhH4), 4.72 (d, J = 11.8 Hz, 1H, PhC<u>H</u>H), 4.66 – 4.63 (m, 1H, H1, PhCH<u>H</u>), 3.95 (dd, J = 3.2, 1.9 Hz, 1H, H2), 3.84 (dd, J = 11.8, 2.3 Hz, 1H, 6a), 3.76

(t, J = 9.7 Hz, 1H, H4), 3.71 (dd, J = 11.8, 6.0 Hz, 1H, 6b), 3.57 (dd, J = 9.4, 3.3 Hz, 1H, H3), 3.50 (ddd, J = 9.8, 6.0, 2.2 Hz, 1H, H5), 3.36 (s, 3H, OMe).

¹³C NMR (101 MHz, MeOD) δ 138.60 (PhC1), 127.87 (PhC3/C5), 127.67 (PhC2/C6), 127.20 (PhC4), 101.26 (C1), 78.97 (C3), 73.16 (C5), 71.27 (PhCH₂), 67.60 (C2), 66.28 (C4), 61.54 (C6), 53.80 (OMe).

HRMS (ESI⁺) Exact mass calc. for $C_{14}H_{20}O_6Na$ [M+Na]⁺: 307.1152, found: 307.1168 (5.26 ppm).

Spectral data in accordance with literature.¹⁶

5.6 1,2,4,6-Tetra-*O*-acetyl-3-*O*-benzyl-α-D-mannopyranose 27



Procedure adapted from literature protocol.¹⁵

The acetylation step was performed separately from the acetolysis step, as the exothermic reaction of the first step leads to partial debenzylation under acetolysis conditions. Methyl 3-*O*-benzyl-mannopyranoside **25** (2.00 g, 7.04 mmol, 1.00 equiv.) was dissolved in pyridine (20.4 ml). Acetic anhydride (6.0 ml, 63 mmol, 1.1 equiv.) and DMAP (43.0 mg 0.352 mmol, 0.05 equiv.) were added carefully and the mixture was stirred at rt for 2 h, when TLC (LP:EA = 7:3) indicated full conversion. MeOH (2.6 ml) was added and the mixture was diluted with EtOAc (200 ml). The solution was transferred into a separating funnel and was washed with 1N HCl (400 ml) until TLC showed no more pyridine in the organic phase. After extracting the 1N HCl phase again with EtOAc, the combined organic phases washed with aq. NaHCO₃ (100 ml) and brine (50 ml). The organic layer was concentrated in vacuo yielding the intermediate **26** as a colourless oil (2.75 g) as crude, which was used without further purification.

An aliquot of intermediate **26** (2.65 g, 6.46 mmol, 1.00 equiv.) in Ac₂O:AcOH:H₂SO₄ (23.2 ml, 50:20:0.5 v/v) was stirred at rt for 5 h when TLC (LP:EtOAc = 4:3) indicated full conversion. The solution was dropwise added to ice water (150 ml) for better heat distribution until the excess of Ac₂O was hydrolyzed. Then NaOAc (790 mg) was added to quench the H₂SO₄. The product was extracted with EtOAc (3 × 100 ml) using TLC as indicator for completion. The

combined organic phases were washed with aq. NaHCO₃ and dried first with brine and, afterwards, with Na₂SO₄ and concentrated *in vacuo*. Purification of the crude mixture was achieved by column chromatography using SiO₂ (90 g) and a gradient of EtOAc in LP from 15-40% and afforded the target compound **27** as a colourless oil (2.57 g, 91%), pure according to ¹H-NMR.

 $\mathbf{R}_{\mathbf{f}}$ (LP:EtOAc = 1:1, anisaldehyde stain): 0.45.

Optical rotation $[\alpha]_D^{20} = +4.7$ (c = 0.9, CHCl₃), (lit.¹⁵ +0.20, c ~1, CHCl₃).

¹**H** NMR (400 MHz, MeOD) δ 7.37 – 7.26 (m, 5H, Ph), 6.03 (d, J = 1.8 Hz, 1H, H1), 5.37 (dd, J = 3.2, 2.1 Hz, 1H, H2), 5.23 (appt. t, J = 10.1 Hz, 1H, H4), 4.65 (d, J = 11.8 Hz, 1H, PhC<u>H</u>H), 4.48 (d, J = 11.8 Hz, 1H, PhCH<u>H</u>), 4.22 (dd, J = 12.5, 4.9 Hz, 1H, H6a), 4.03 (dd, J = 12.1, 2.3 Hz, 1H, H6b), 4.03 – 3.97 (m, 1H, H5), 3.97 (dd, J = 9.8, 3.4 Hz, 1H, H3), 2.14, 2.13, 2.04, 2.01 (4 × s, 4 × 3H, 4 × COCH₃).

¹³C NMR (101 MHz, MeOD) δ 172.4, 171.5, 171.4, 169.9 (4 × s, 4 × <u>C</u>OCH₃), 139.1 (s, PhC1), 129.4 (d, PhC3/C5), 129.1 (d, PhC2/C6), 128.9 (d, PhC4), 92.3 (d, C1), 75.9 (d, C3), 72.8 (t, PhCH₂), 72.0 (d, C5), 68.8 (d, C2), 68.2 (d, C4), 63.4 (t, C6), 20.8, 20.7, 20.64, 20.61 (4 × q, 4 × CO<u>C</u>H₃).

HRMS (ESI⁺) calc. for C₂₁H₂₆O₁₀Na [M+Na]⁺: 461.1418, found: 461.1444 (-5.51 ppm).

Spectral data in accordance with literature.¹⁷

5.7 3-*O*-Benzyl-α-D-mannose 1b



The 1,2,4,6-tetra-*O*-acetyl-3-*O*-benzyl-mannopyranoside **27** (600 mg, 1.37 mmol, 1.00 equiv.) was dissolved in MeOH (5 ml, anh.). NaOMe (0.23 ml, 0.05 equiv., 30% solution in MeOH) was added and the pH was checked to be ~10. The solution was stirred at room temperature for 3 h, when full conversion was observed by TLC (DCM/MeOH = 10:1, anisaldehyde stain). Acidic ion exchange resin (Amberlyst 15, 0.6 g) was added and the mixture was stirred for a few minutes. After neutralization (checked using pH paper) the resin was filtered off, washed with methanol, the filtrate was dried using Na₂SO₄ and concentrated *in vacuo*. Purification of the crude mixture was achieved by column chromatography using SiO₂ (12 g) and a gradient

of MeOH in DCM from 5-25% and afforded the target compound **1b** as a colourless oil (188 mg, 51%) and pure according to ¹H-NMR.

 $\mathbf{R}_{\mathbf{f}}$ (CHCl₃:MeOH:H₂O = 7:3:0.5, anisaldehyde stain): 0.45.

Optical rotation $[\alpha]_D^{20} = -0.80$ (c = 1.0, MeOH, 12 h), (lit.¹⁸ -16, c = 1.0, CHCl₃; lit.¹⁵ +3.0, c = 1, CHCl₃).

¹**H NMR** (**400 MHz**, **MeOD**) α- and β-isomer were obtained in a ratio of ~ 7:3. δ 7.45 (d, J = 6.8 Hz, 2H, PhH2/H6), 7.33 (t, J = 7.3 Hz, 2H, PhH3/H5), 7.27 (d, J = 7.1 Hz, 1H, PhH4), 5.08 (d, J = 1.8 Hz, 0.7H, α H1), 4.74 (d, J = 11.6 Hz, 1H, PhC<u>H</u>H), 4.70 (d, J = 0.9 Hz, 0.3H, β H1), 4.66 (d, J = 11.7 Hz, 1H, PhCH<u>H</u>), 3.99 (d, J = 2.3 Hz, 0.3H, β H2), 3.96 (dd, J = 3.0, 2.0 Hz, 0.7H, α H2), 3.86 (dd, J = 11.8, 2.4 Hz, 0.3H, β H6a), 3.83 – 3.73 (m, 3.4H, α H4, β H4, α H5, α H6a, α H6b, H6b β), 3.69 (dd, J = 8.9, 3.0 Hz, 0.7H, α H3), 3.38 (dd, J = 9.4, 3.1 Hz, 0.3H, β H3), 3.24 (ddd, J = 9.6, 5.8, 2.4 Hz, 1H, β H5).

¹³C NMR (101 MHz, MeOD) α-anomer: δ 140.1 (PhC1), 129.3 (PhC3/C5), 129.1 (PhC2/6), 128.6 (PhC4), 95.8 (C1), 80.3 (C3), 74.1 (C5), 72.7 (PhCH₂), 69.9 (C2), 67.8 (C4), 63.0 (C6). β-anomer: δ 140.1 (PhC1), 129.3 (PhC3/C5), 129.1 (PhC2/C6), 128.6 (PhC4), 95.6 (C1), 82.9 (C3), 78.1 (C4 or C5), 72.4 (Ph-CH₂), 70.0 (C2), 67.4 (C4 or C5), 63.0 (C6).

HRMS (ESI+) calc. for C₁₃H₁₈O₆Na [M+Na]+: 293.0996, found: 293.1009 (-4.43 ppm).

5.8 2-O-Benzyl-D-arabinose 3¹⁹



3-*O*-Benzyl- α -D-glucose **1a** (5.00 g, 18.5 mmol, 1.00 equiv.) was dissolved in water (15 mL). sodium periodate (4.16 g, 19.4 mmol, 1.05 equiv.) was dissolved in water (15 mL), under heating in a water bath, then rapidly cooled in an ice bath and added to the glucose solution. The mixture was cooled in an ice bath instantly for about 1 min and then stirred at rt for 15 min. A white solid precipitated, and the reaction mixture turned solid. The formed precipitate was then collected *via* vacuum filtration and washed with cold water, yielding 4 g of intermediate **28**. MeOH (50 mL) was added to the filtrate and it was placed in the refrigerator for 3h, where further precipitation occurred, which was again collected *via* vacuum filtration, yielding another

2.5 g of **28**. Both parts of **28** were combined, dissolved in MeOH (28 mL), mixed with NEt₃ (9 mL) and stirred overnight. After saponification, a precipitate formed, which was filtered off and washed with MeOH. The filtrate was concentrated under reduced pressure. Purification of the crude mixture was achieved first by column chromatography using solid loading (Celite[®], 10 g) on SiO₂ (90 g) and a gradient of MeOH in DCM from 6-21%, collecting the product fractions. After solvent removal these were refluxed in EtOAc (4 ml). After cooling down the precipitate was collected by vacuum filtration and washed with EtOAc, yielding the target compound **3** as a slightly beige solid (2.71 g, 61%).

m.p. 111.3-112.1 °C (EtOAc), (lit.¹⁹ 111-113 °C).

 $\mathbf{R}_{\mathbf{f}}$ (DCM:MeOH = 9:1, anisaldehyde stain): 0.29.

Optical rotation $[\alpha]_D^{20} = -71.8$ (c = 1.0, H₂O, 12 h), (lit.¹⁹ -73, c = 2.1, H₂O).

¹**H** NMR (400 MHz, MeOD) the isomers were obtained in a ratio of furanosides : α-pyranoside : β-pyranoside ~ 37:30:33. δ 9.02 – 8.77 (m, 5H, PhH), 6.82, 6.81 (d, J = 2.2 Hz, 0.37H, αf H1, βf H1), 6.71 (d, J = 3.3 Hz, 0.3H, αp H1), 6.45 – 6.17 (m, 2H, PhCH₂), 6.11 (d, J = 6.8 Hz, 0.33H, βp H1), 5.73 (dd, J = 7.0, 6.3 Hz, 0.1H, CHOH), 5.61 – 5.47, 5.45 – 5.35, 5.33 – 5.28, 5.27 – 5.14, 5.14 – 5.08, 5.06 – 5.00 (6 × m, 5.14H, CH(OH)).

¹³**C NMR (101 MHz, MeOD)** δ 140.3, 140.0, 139.4 (PhC1), 129.3, 129.3, 129.2, 129.2, 129.1, 129.1, 128.9, 128.8, 128.7, 128.7, 128.5 (PhC2-6), 101.9 (αf C1), 98.5 (βp C1), 92.8 (αp C1), 91.9, 85.7, 84.4, 83.9, 81.7, 78.3, 76.7, 75.4, 75.1, 73.8, 73.7, 73.2, 72.8, 70.5, 69.8, 69.8, 66.5, 64.6, 63.7, 62.9 (C2-C5, Ph<u>C</u>H₂).

HRMS (ESI⁺) Exact mass calc. for $C_{12}H_{16}O_5Na$ [M+Na]⁺: 263.0890, found: 263.0899 (3.27 ppm).

Hydrogenation (H₂, Pd/C, MeOH) led to clean conversion to D-arabinose.

5.9 2-Deoxy-D-ribono-1,4-lactone 4a²⁰



2-Deoxy-D-ribose **29** (2.00 g, 14.9 mmol, 1.00 equiv.) was dissolved in H_2O (12 ml) in a round bottom flask. To this solution, Br_2 (4.0 ml, 77.6 mmol, 5.20 equiv.) was added, which led to the

formation of a biphasic system. The flask was sealed and stirring was continued at rt for 24 h, when TLC (CHCl₃:MeOH:H₂O = 7:3:0.5, starting material staining with anisaldehyde, product with KMnO₄ solution) showed complete conversion to the desired product. Ag₂CO₃ was added until the supernatant decolourized, which led to the formation of gas and a yellow-white precipitate. The reaction mixture was filtered through a bed of Celite[®] and the solvent was lyophilized. Purification of the crude mixture was achieved by column chromatography using SiO₂ (30 g) and a gradient of MeOH in EtOAc from 0-10% and afforded the target compound **4a** as a colourless oil (1.50 g, 76%) and pure according to ¹H-NMR.

 \mathbf{R}_{f} (DCM:MeOH = 6:1, KMnO₄ stain): 0.48.

Optical rotation $[\alpha]_D^{20} = +4.7$ (c = 1.0, MeOH), (lit.²⁰ +3.05, c = 1.14, MeOH).

¹**H NMR (400 MHz, MeOD)** δ 4.43 (dt, *J* = 6.7, 2.2 Hz, 1H, H3), 4.37 (q, *J* = 3.2 Hz, 1H, H4), 3.76 (dd, *J* = 12.4, 3.2 Hz, 1H, H5a), 3.69 (dd, *J* = 12.4, 3.6 Hz, 1H, H5b), 2.91 (dd, *J* = 18.0, 6.7 Hz, 1H, H2a), 2.37 (dd, *J* = 18.0, 2.4 Hz, 1H, H2b).

¹³C NMR (101 MHz, MeOD) δ 177.2 (C1), 88.8 (C4), 68.3 (C3), 61.1 (C5), 37.8 (C2).

Spectral data in accordance with literature.²¹

5.10 3,4-O-Isopropylidene L-arabinose 6²²



A suspension of L-arabinose **30** (5.00 g, 33.8 mmol, 1.00 equiv.) in DMF (anh., 20 mL) was prepared. Dimethoxypropane (10.5 g, 101 mmol, 3.00 equiv.) and pTsOH·H₂O (96 mg, 0.5 mmol, 0.02 equiv.) were added while stirring at rt. After 160 minutes the starting material was fully converted to a less polar spot (TLC (LP/EtOAc = 1:2 and CHCl₃/MeOH/H₂O = 7:3:0.5, anisaldehyde stain). triethylamine (0.17 g, 1.70 mmol, 0.05 equiv.) was added and the mixture was stirred overnight. The next day, the clear solution was co-evaporated with toluene (5 × 40 mL). The residue was dried under high vacuum. Purification of the crude mixture was achieved by column chromatography using SiO₂ (90 g) and a gradient of EtOAc in LP from

20-100% and afforded the target compound **6** as a colourless solid (4.67 g, 73%) and pure according to ¹H-NMR.

m.p. 124.4-124.5 °C (EtOAc), (lit.²³ 110 °C).

 $\mathbf{R}_{\mathbf{f}}$ (LP:EtOAc = 1:3, anisaldehyde stain): 0.22.

Optical rotation $[\alpha]_D^{20} = +118$ (c = 1.0, H₂O, 12 h), (lit.²³ +128.8, c = 1.0, H₂O).

¹**H** NMR (400 MHz, MeOD) The anomeric ratio is according to ¹H-NMR α : β = 0.17 :0.83. δ 5.01 (d, *J* = 3.3 Hz, 1H, β H1), 4.37 (d, *J* = 7.9 Hz, 1H, α H1), 4.27 – 4.20 (m, 1H, β H4), 4.21 – 4.16 (m, 2H, β H3 + α H4), 4.17 – 4.14 (m, 1H, α + β H5a), 4.00 (t, *J* = 6.5 Hz, 1H, α H3), 3.85 – 3.76 (m, 2H, α + β H5b), 3.62 (dd, *J* = 7.0, 3.3 Hz, 1H, β H2), 3.41 (t, *J* = 7.6 Hz, 1H, α H2), 1.48 (s, 3H, CH₃), 1.34 (s, 3H, CH₃).

¹³C NMR (101 MHz, MeOD) δ 110.7 (α <u>C</u>(CH₃)₂), 109.9 (β <u>C</u>(CH₃)₂), 97.7 (α C1), 93.7 (β C1), 80.5 (α C3), 77.4 (β C3), 75.6 (α C2), 75.0 (α C4), 74.6 (β C4), 71.7 (β C2), 64.0 (α C5), 59.9 (β C5), 28.4 (α C(<u>C</u>H₃)₂), 26.4 (β C(<u>C</u>H₃)₂).

Spectral data is in accordance with literature.²⁴

5.11 3-*O*-(4-Nitrophenyl)-1,2:5,6-di-*O*-isopropylidene-α-D-glucofuranose 31



p-Fluoronitrobenzene was distilled by Kugelrohr distillation at 100 °C and 0.35 mbar for about 30 minutes to give a clear liquid from a brown liquid.

A vial with diacetone glucose **20** (1000 mg, 3.84 mmol, 1.10 equiv.) was evacuated and purged with Ar three times. Then, anhydrous THF (17.5 mL) and p-NO₂PhF (492 mg, 3.49 mmol, 1.00 equiv.) were added via syringe. The reaction mixture was cooled in an ice bath and KHMDS (anh., 0.7 M solution in toluene, 7.68 mL, 3.84 mmol, 1.10 equiv.) was added dropwise to the cooled reaction mixture. After 2.5 h, a first TLC analysis (LP/EtOAc 4:1) showed the formation of a new, less polar spot with still both starting material and *p*-NO₂PhF left. The reaction was stirred overnight with no further reaction having occurred according to

TLC. Aq. NaHCO₃ (sat., 5 mL) was added. After 15 minutes brine (30 mL) was added and the reaction mixture was extracted with EtOAc (2 \times 70 mL, 1 \times 20 mL). The combined organic layers were dried over Na₂SO₄ and the solvent was evaporated to give the desired product as crude material as yellow liquid. Purification of the crude mixture was achieved by column chromatography using SiO₂ (90 g) and a gradient of EtOAc in LP from 9-20% and afforded the target compound **31** as colourless crystals (1418 mg, 97%) and pure according to ¹H-NMR.

m.p. 134-136 °C (CHCl₃), (lit.²⁵ 136-137 °C).

 $\mathbf{R}_{\mathbf{f}}$ (LP:EtOAc = 4:1, anisaldehyde stain): 0.22.

Optical rotation $[\alpha]_D{}^{20} = -35.6$ (c = 1.0, CHCl₃), (lit.²⁶ $[\alpha]_D{}^{21} = -45$, c = 1.0, CHCl₃).

¹**H NMR (400 MHz, CDCl₃)** δ 8.26 – 8.20 (m, 2H, PhH3/H5), 7.11 – 7.05 (m, 2H, PhH2/H6), 5.95 (d, *J* = 3.8 Hz, 1H, H1), 4.82 (d, *J* = 3.0 Hz, 1H, H3), 4.57 (d, *J* = 3.9 Hz, 1H, H2), 4.41 (ddd, *J* = 8.3, 6.0, 5.1 Hz, 1H, H5), 4.29 (dd, *J* = 8.3, 3.0 Hz, 1H, H4), 4.15 (dd, *J* = 8.7, 6.0 Hz, 1H, H6a), 4.09 (dd, *J* = 8.7, 5.0 Hz, 1H, H6b), 1.56, 1.43, 1.32, 1.29 (4×s, 4×3H, 2×C(CH₃)₂) ppm.

¹³C NMR (101 MHz, CDCl₃) δ 162.1 (PhC1), 142.4 (PhC4), 126.1 (PhC3/C5), 115.6 (PhC2/C6), 112.6 (<u>C</u>(CH₃)₂(O1/O2)), 109.6 (<u>C</u>(CH₃)₂(O5/O6)), 105.4 (C1), 82.4 (C2), 80.8 (C3), 80.5 (C4), 72.0 (C5), 67.4 (C6), 27.1, 26.8, 26.4, 25.3 (2×C(<u>C</u>H₃)₂) ppm.

Spectral data in accordance with literature.²⁶

5.12 3-O-(4-Nitrophenyl)-D-glucose 10



Amberlyst 15 (hydrogen form, 3.57 g) was first washed with water (3x 50 mL) and then added to the reaction mixture. 3-O-(4-Nitrophenyl)-1,2:5,6-di-O-isopropylidene- α -D-glucofuranose **31** (1.39 g, 3.64 mmol, 1.00 equiv.) was stirred in H₂O (14 ml) and heated to 70 °C. TLC
(4:1 DCM/MeOH) showed after approx. 2.5 h complete reaction. The reaction mixture was filtered over Celite in a glassinter funnel and washed several times with water. The pooled aqueous fractions (~70 mL) were washed with Et₂O (2 x 10 mL) whereby most of the *p*-NO₂PhF from the reaction before was removed. Then, it was evaporated *in vacuo* to give the product **10** as yellow powder (556 mg, 51%) due to small leftover impurities of *p*-NO₂PhF. According to NMR analysis the obtained product **10** was pure.

m.p. 172-175 °C (H₂O). **R**_f (DCM:MeOH = 4:1, anisaldehyde stain): 0.50.

Optical rotation $[\alpha]_D^{20} = +51.2$ (c = 1.0, H₂O, 12 h).

¹**H** NMR (400 MHz, D₂O) α- and β-isomer were obtained in a ratio of ~ 4:6. (400 MHz, D₂O) δ 8.24 (d, J = 9.1 Hz, 2H, α,β PhH3/H5), 7.27 (d, J = 9.1 Hz, 2H, α,β PhH2/H6), 5.33 (d, J = 3.7 Hz, 0.4H, α H1), 4.82 (d, J = 8.1 Hz, 0.6H, β H1), 4.81 – 4.73 (m, 0.4H, α H3), 4.60 (t, J = 9.1 Hz, 0.6H, β H3), 4.03 – 3.94 (m, 0.4H, α H5), 3.93 (dd, J = 12.3, 2.0 Hz, 0.6H, β H6a), 3.90 – 3.70 (m, 2.8H, α H2, α H4, β H4, α H6a, α H6b, β H6b), 3.62 (ddd, J = 10.0, 5.6, 2.1 Hz, 0.6H, β H5), 3.57 (s, 0.6H, β H2) ppm.

¹³C NMR (101 MHz, D₂O) δ 165.9, 165.7 (α,β PhC1), 142.1, 142.0 (α,β PhC4), 126.7 (α,β PhC3/C5), 117.03 (β PhC2/C6), 116.95 (α PhC2/C6), 96.3 (β C1), 92.8 (α C1), 84.3 (β C3), 81.9 (α C3), 76.2 (β C5), 74.1 (β C2), 71.9 (α C5), 71.5 (α C2), 69.7, 69.6 (α, β C4), 61.2 (β C6), 61.0 (α C6) ppm.

HRMS (ESI⁻) Exact mass calc. for C₁₄H₁₈NO₁₀ [M+COO]⁻: 360.0936, found: 360.0964 (-7.78 ppm).

6 Synthesis and characterization of precatalysts

6.1 General procedure for the synthesis of cyclohepta[*d*]thiazole-2-thiones 34, 35, 36, 37 and 38 (procedure A)²⁷



A solution of the respective aniline (1.00 equiv.) in DMSO (0.5 ml/mmol) was treated with aqueous NaOH solution (20 N, 1.00 equiv.). At 0 °C CS₂ (1.00 equiv.) was added dropwise and stirred for 1 h at room temperature. Then, 2-bromocycloheptanone **33** (1.00 equiv.) was added slowly at 0 °C, which led to a colour change of the solution and the mixture was stirred for 20 h at rt. H₂O (1 mL/mmol) was added, the mixture was stirred for 10 min at 0 °C and the supernatant solution was decanted. Depending on the substrate either a slurry or a precipitate remained, that was suspended in EtOH (1 mL/mmol). Concentrated HCl (0.05 mL/mmol, 0.58 equiv.) was added dropwise and the mixture was heated to reflux for 1 h. After cooling to room temperature, the mixture was placed in the fridge, where crystals precipitated. The solid was collected by suction filtration, washed with small amounts of *n*-pentane before being dried by first purging with air and then under reduced pressure on the rotary evaporator. Purification by column chromatography was performed when products were not already obtained pure according to ¹H-NMR.

6.2 General procedure for the synthesis of cyclohepta[d]thiazole precatalysts 12, 13, 14, 15 and 16 (procedure B)²⁷



A solution or suspension of the thione (1.00 equiv.) in glacial acetic acid (4.1 ml/mmol) was cooled with a water bath, then H_2O_2 (30%, aqueous solution, 3.30 equiv.) was added dropwise and the reaction mixture was stirred for 1 h. The volatile components were removed under reduced pressure and the residue was dissolved in MeOH (0.69 ml/mmol). It was cooled to 0 °C and a mixture of sodium perchlorate monohydrate (4.10 equiv.) in MeOH/H₂O = 2/1 (3.61 ml/mmol) was added and the solution was stirred for 30 min. H₂O (50 mL) was added and the mixture was extracted three times with DCM. The combined organic layers were dried over Na₂SO₄ and the solvent was removed under reduced pressure. Purification was performed by column chromatography or trituration when required.

6.3 3,4,5-Trimethylthiazol-3-ium iodide 2²⁸



4,5-Dimethylthiazole **39** (2.15 g, 19.0 mmol, 1.00 equiv.) was dissolved in MeOH (12.1 ml), iodomethane (22.9 g, 162 mmol, 8.5 equiv.) was added and the solution was stirred in a closed flask at rt for 4 days, leading to the formation of a first slightly yellow, then brown solution with a white precipitate. EtOAc (50 mL) was added to the solution to decrease the solubility of the already precipitating product. The formed precipitate was filtered, washed with EtOAc (30 ml) and was consequently dried on a suction filter. Remaining solvent was evaporated *in vacuo* (6 mbar, 40°C) yielding the target compound **2** slightly yellow crystalline solid (4.51 g, 93%) pure according to ¹H-NMR.

m.p. 227.8-228.9 °C (EtOAc), (lit.²⁹: 228 - 230 °C).

¹**H NMR (400 MHz, DMSO-***d*₆) δ 9.94 (d, *J* = 0.9 Hz, 1H, H2), 4.08 (d, *J* = 0.8 Hz, 3H, N-CH₃), 2.49 (d, *J* = 0.8 Hz, 3H, CH₃), 2.40 (d, *J* = 0.9 Hz, 3H, CH₃).

¹³C NMR (101 MHz, DMSO-*d*₆) δ 155.6 (C2), 142.1 (C4), 132.3 (C5), 40.4 (N-CH₃), 12.0,11.1 (2 × CH₃).

HRMS (**ESI**⁺) Exact mass calc. for $C_6H_{10}NS$ [M-I]⁺: 128.0543, found: 128.0534 (7.11 ppm). Spectral data in accordance with literature.³⁰

6.4 3-Ethyl-4,5-dimethylthiazol-3-ium bromide 40²⁸



4,5-Dimethylthiazole **39** (1.0 mL, 8.80 mmol, 1.00 equiv.) was dissolved in 1-butanol (1.0 mL). Then, EtBr (925 μ L, 13.2 mmol, 1.50 equiv.) was added and the vial was closed. The reaction vial was heated to 110 °C and stirred for 20 minutes. Then, the vial was allowed to cool down to room temperature. Two layers formed of which the lower one contained beige floating solid,

which was collected and washed with EtOAc (~30 mL) to give the product **40** as a white powder (443 mg, 24%), pure according to NMR analysis.

m.p.: 162-164 °C (BuOH/EtOAc)

¹**H NMR** (400 MHz, MeOD) δ 9.97 (s, 1H, H2), 4.55 (q, *J* = 7.3 Hz, 2H, CH₂), 2.60 (s, 3H, CH₃*), 2.56 (s, 3H, CH₃*), 1.63 (t, *J* = 7.3 Hz, 3H, *CH*₃-CH₂) ppm.

¹³C NMR (101 MHz, MeOD) δ 155.2 (C2), 143.5 (C4), 135.1 (C5), 50.3 (CH₂), 14.9 (CH₃-CH₂), 12.3 (CH₃*), 11.4 (CH₃*) ppm.

HRMS (ESI⁺) Exact mass calc. for $C_7H_{12}NS$ [M-Br]⁺: 142.0685, found: 142.0697 (8.11 ppm). Spectral data in accordance with literature.²⁸

6.5 2-Bromocycloheptanone 33³¹



Under Ar atmosphere cycloheptanone **32** (5.00 g, 44.6 mmol, 1.00 equiv.) was dissolved in DCM (anh., 11 ml) in a three-neck round bottom flask. *p*-Toluenesulfonic acid monohydrate (0.85 g, 4.5 mmol, 0.10 equiv.) was added in one portion and *N*-bromosuccinimide (7.93 g, 44.6 mmol, 1.00 equiv.) was added in several small portions while cooling with ice-water, preventing the reaction from exceeding 5 °C. After slowly warming to rt, the reaction mixture was stirred for 20 h at this temperature, when GC-MS (2.74 min: starting material with m/z = 112.09, 4.24 min: product with m/z = 190.0 and 192.0, 5.56 min: dibromo-product with m/z = 267.94, 269.90, 271.88) and TLC (LP:EtOAc = 10:1, cerium molybdate stain) indicated dominant product formation. The reaction was diluted by adding LP (25 ml) and was stirred for 5-10 min to improve precipitation of the formed succinimide.

The beige solid was filtered off and the filtrate was washed with aqueous Na₂S₂O₃ (satd., 2×75 ml), aqueous NaHCO₃ (satd., 2×75 ml) and brine (2×75 ml), dried over Na₂SO₄ and evaporated yielding 8.08 g (95%) of a slightly yellowish liquid. Analysis by ¹H-NMR confirmed predominant product formation with 12% of what is assumed to be the 2,7-dibrominated product. Distillation at 106 °C, 7 mbar afforded product **33** as a yellowish liquid (6.48 g, 76%) pure according to ¹H-NMR.

b.p. 106 °C (7 mbar).

Rf (LP/EtOAc 10:1, cerium molybdate stain): 0.61

GC-MS $t_R = 4.24$ min, main fragments 192 (1.5, M⁺), 190 (1.5, M⁺), 111 (45), 93 (46), 55 (100).

¹**H NMR (400 MHz, CDCl**₃) δ 4.37 (dd, *J* = 9.6, 5.1 Hz, 1H, H2), 2.92 – 2.80 (m, 1H, H7a), 2.56 – 2.42 (m, 2H, H7b), 2.42 – 2.28 (m, 1H, H3a), 2.13 – 1.86 (m, 4H, H3b, H4a, H6a), 1.81 – 1.64 (m, 2H, H5a), 1.63 – 1.47 (m, 4H, H5b, H6b), 1.44 – 1.31 (m, 2H, H4b).

¹³C NMR (101 MHz, CDCl₃) δ 206.4 (C1), 53.9 (C2), 39.5 (C7), 34.4 (C3), 29.7 (C5), 27.0 (C4), 25.1 (C6).

Spectral data in accordance with literature.³²

6.6 3-Phenyl-3,4,5,6,7,8-hexahydro-2*H*-cyclohepta[*d*]thiazole-2-thione 34



Thione **34** was prepared according to general **procedure A** from aniline (487 mg, 5.23 mmol, 1.00 equiv.). It was further purified using column chromatography (SiO₂, 12 g, EtOAc in LP = 5-50%) yielding a bright beige solid (228 mg, 17%) pure according to ¹H-NMR.

m.p. 197.2-197.6 °C. (EtOAc).

 $\mathbf{R}_{\mathbf{f}}$ (LP:EtOAc = 5:1, potassium permanganate stain): 0.27.

¹**H NMR (400 MHz, CDCl**₃) δ 7.59 – 7.53 (m, 2H, PhH3/H5), 7.52 – 7.45 (m, 1H, PhH4), 7.22 (m, 2H, PhH2/H6), 2.66 – 2.53 (m, 2H, H8), 2.42 – 2.19 (m, 2H, H4), 1.87 – 1.73 (m, 4H, H6, H7), 1.65 – 1.45 (m, 2H, H5).

¹³C NMR (101 MHz, CDCl₃) δ 187.3 (C2), 141.8 (C3a), 138.8 (Ph C1), 130.0 (Ph C3/C5), 129.6 (Ph C4), 128.5 (Ph C2/C6), 124.3 (C8a), 30.8 (C6 or C7), 29.8 (C4), 27.2 (C8 & C6 or C7), 27.1 (C8 & C6 or C7), 26.0 (C5).

HRMS (ESI⁺) calc. for $C_{14}H_{16}NS_2$ [M+H]⁺: 262.0719, found: 262.0725 (- 2.32 ppm).

6.7 3-Phenyl-5,6,7,8-tetrahydro-4*H*-cyclohepta[*d*]thiazol-3-ium perchlorate 12



Thiazol-3-ium perchlorate **12** was prepared according to general **procedure B** from thione **34** (200 mg, 0.77 mmol, 1.00 equiv.). The product was not obtained pure after precipitation and purification *via* column chromatography with SiO₂ (12 g) and a gradient of MeOH in DCM from 0-20% and a second column using 5-10% afforded the target compound **12** as a brown oil (97 mg, 39%) and pure according to ¹H-NMR.

 $\mathbf{R}_{\mathbf{f}}$ (DCM:MeOH = 9:1, potassium permanganate stain): 0.28.

¹**H NMR (400 MHz, CDCl**₃) δ 9.54 (s, 1H, H2), 7.71 – 7.57 (m, 3H, PhH3/H5, PhH4), 7.54 – 7.48 (m, 2H, PhH2/H6), 3.13 – 3.01 (m, 2H, H8), 2.78 – 2.69 (m, 2H, H4), 2.02 – 1.83 (m, 4H, H6 & H7), 1.76 – 1.65 (m, 2H, H5).

¹³C NMR (101 MHz, CDCl₃) δ 154.4 (C2), 148.5 (C3a), 140.3 (C8a), 136.9 (PhC1), 131.9 (PhC4), 130.6 (PhC3/C5), 126.2 (PhC2/C6), 30.8 (C6), 28.2 (C8), 28.0 (C4), 26.4 (C7), 25.2 (C5).

HRMS (ESI⁺) calc. for $C_{14}H_{16}NS$ [M-ClO₄]⁺: 230.0998, found: 230.0998 (± 0.00 ppm).

6.8 3-(4-Methoxyphenyl)-3,4,5,6,7,8-hexahydro-2H-cyclohepta[d]thiazole-2-thione 35



3-(4-Methoxyphenyl)-3,4,5,6,7,8-hexahydro-2*H*-cyclohepta[*d*]thiazole-2-thione **35** was prepared according to general **procedure A** from 4-methoxyaniline (645 mg, 5.23 mmol, 1.00 equiv.) yielding a beige solid (619 mg, 41%) pure product according to ¹H-NMR directly after precipitation.

m.p. 119.2-120.1 °C (H₂O).

 $\mathbf{R}_{\mathbf{f}}$ (LP:EtOAc = 5:1, permanganate stain): 0.23.

¹**H NMR (400 MHz, CDCl**₃) δ 7.12, 7.03 (2 × d, *J* = 8.5 Hz, 2 × 2H, PhH2/H6 & PhH3/H5), 3.85 (s, 3H, OCH₃), 2.64 – 2.57 (m, 2H, H8), 2.34 – 2.26 (m, 2H, H4), 1.82 – 1.71 (m, 4H, H6 & H7), 1.65 – 1.53 (m, 2H, H5).

¹³C NMR (101 MHz, CDCl₃) δ 187.3 (C2), 160.0 (PhC4), 142.1 (C3a), 131.2 (PhC1), 129.4, 115.0 (PhC2/C6, PhC3/C5), 123.8 (C8a), 55.5 (OCH₃), 30.7 (C6 or C7), 29.7 (C4), 27.1, 27.0 (C8 & C6 or C7), 25.8 (C5).

HRMS (ESI⁺) calc. for $C_{15}H_{18}NOS_2$ [M+H]⁺: 292.0824, found: 292.0829 (- 1.63 ppm).

6.9 3-(4-Methoxyphenyl)-5,6,7,8-tetrahydro-4H-cyclohepta[d]thiazol-3-ium perchlorate 13



3-(4-Methoxyphenyl)-5,6,7,8-tetrahydro-4*H*-cyclohepta[*d*]thiazol-3-ium perchlorate **13** was prepared according to general **procedure B** from **36** (475 mg, 1.63 mmol, 1.00 equiv.) yielding the target compound **13** (512 mg, 87%) pure according to ¹H-NMR.

 $\mathbf{R}_{\mathbf{f}}$ (DCM:MeOH = 9:1, permanganate stain): 0.36.

¹**H NMR (400 MHz, CDCl₃)** δ 9.50 (s, 1H, H2), 7.43, 7.05 (2×d, *J* = 8.6 Hz, 2×2H, PhH2/H6 & PhH3/H5), 3.87 (s, 3H, OCH₃), 3.18 – 2.96 (m, 2H, H8), 2.83 – 2.68 (m, 2H, H4), 1.96 – 1.81 (m, 4H, H6 & H7), 1.71 – 1.61 (m, 2H, H5).

¹³C NMR (101 MHz, CDCl₃) δ 161.8 (PhC4), 154.4 (C2), 148.7 (C3a), 139.9 (C8a), 129.5 (PhC1), 127.5, 115.5 (PhC2/C6 & PhC3/C5), 56.0 (OCH₃), 30.8 (C6), 28.2 (C8), 27.9 (C4), 26.5 (C7), 25.2 (C5).

HRMS (ESI⁺) calc. for C₁₅H₁₈NOS [M-ClO₄]⁺: 260.1104, found: 260.1119 (- 5.99 ppm).

6.10 3-(4-Nitrophenyl)-5,6,7,8-tetrahydro-4H-cyclohepta[d]thiazol-3-ium perchlorate 14



3-(4-Nitrophenyl)-3,4,5,6,7,8-hexahydro-2*H*-cyclohepta[*d*]thiazole-2-thione **36** was prepared according to general **procedure A** from 4-nitroaniline (723 mg, 5.23 mmol, 1.00 equiv.). It was further purified using column chromatography yielding an orange solid (361.0 mg), however still containing an unidentified impurity (¹H-NMR). The material was used without further purification in the next step.

 $\mathbf{R}_{\mathbf{f}}$ (LP:EtOAc = 5:1, permanganate stain): 0.33.

¹**H NMR (400 MHz, CDCl**₃) δ 8.54 – 8.36, 7.52 – 7.40 (2 × m, 2 × 2H, PhH2/H6, PhH3/H5), 2.63 (dd, *J* = 15.8, 9.2 Hz, 2H, H8), 2.31 (td, *J* = 11.4, 5.8 Hz, 2H, H4), 1.90 – 1.75 (m, 4H, H6, H7), 1.64 (dt, *J* = 16.9, 5.4 Hz, 2H, H5).

¹³C NMR (101 MHz, CDCl₃) δ 187.4 (s, C2), 148.0, 143.9 (2×s, PhC1 & PhC4), 140.6 (s, H3a), 130.1 (d, PhC2/C6), 125.5 (s, H8a), 125.2 (d, PhC3/C5), 30.5 (t, C6), 29.7 (t, C4), 27.1 (t, C8), 26.9 (t, C7), 25.8 (t, C5).

HRMS (ESI⁺) calc. for $C_{14}H_{15}N_2O_2S_2$ [M+H]⁺: 307.0568, found: 307.0568 (± 0.00 ppm).



3-(4-Nitroxyphenyl)-5,6,7,8-tetrahydro-4*H*-cyclohepta[*d*]thiazol-3-ium perchlorate **14** was prepared according to general **procedure B** from the crude mixture of **36** (170.6 mg, stoichiometric calculations assuming a content of 1.63 mmol, 1.00 equiv.). Purification was achieved through trituration of the obtained crude oil with Et₂O resulting in the precipitation of the target compound **13** which was washed with CHCl₃ to yield a beige solid (60 mg, 29%) which were pure according to ¹H-NMR.

m.p. 173-175 °C (MeOH).

 $\mathbf{R}_{\mathbf{f}}$ (DCM:MeOH = 9:1, permanganate stain): 0.36.

¹**H** NMR (600 MHz, MeOD) δ 10.08 (s, 1H, H2), 8.58 (d, *J* = 9.0 Hz, 2H, PhH2/H6 or PhH3/H5), 7.92 (d, *J* = 9.0 Hz, 2H, PhH2/H6 or PhH3/H5), 3.23 – 3.11 (m, 2H, H8), 2.90 – 2.74 (m, 2H, H4), 2.05 – 1.97 (m, 2H, H6), 1.94 – 1.88 (m, 2H, H7), 1.81 – 1.73 (m, 2H, H5).

¹³C NMR (151 MHz, MeOD) δ 149.6 (s, PhC1 or PhC4), 148.6 (s, C3a), 141.3 (s, PhC1 or PhC4), 140.1 (s, C8a), 127.8, 125.2 (2×d, PhC2/C6 & PhC3/C5), 30.3 (t, C6), 27.3 (t, C4), 27.1 (t, C8), 26.1 (t, C7), 24.6 (t, C5). C2 is not visible, presumably due to neighboring D exchange resulting in broadening of the signal.

HRMS (ESI⁺) calc. for C₁₇H₂₂NO₂ [M-ClO₄]⁺: 275.0849, found: 275.0848 (- 0.36 ppm).

6.11 3-Mesityl-3,4,5,6,7,8-hexahydro-2*H*-cyclohepta[*d*]thiazole-2-thione 37²⁷



Thione **37** was prepared according to general **procedure A** from 2,4,6-trimethylaniline (707 mg, 5.23 mmol, 1.00 equiv.) yielding a beige solid (596 mg, 38%) pure according to 1 H-NMR directly after precipitation.

m.p. 135.3-137.0 °C (*n*-pentane).

 $\mathbf{R}_{\mathbf{f}}$ (LP:EtOAc = 5:1, potassium permanganate stain): 0.33.

¹**H NMR (400 MHz, CDCl**₃) δ 7.00 (s, 2H, Ph H3/5), 2.66 – 2.60 (m, 2H, H8), 2.33 (s, 3H, PhC<u>H</u>₃(C4)), 2.24 – 2.16 (m, 2H, H4), 2.01 (s, 6H, PhCH₃(C2/C6)), 1.85-1.74 (m, 4H, H6 & H7), 1.62-1.53 (m, 2H, H5).

¹³C NMR (101 MHz, CDCl₃) δ 185.4 (C2), 140.8 (C3a), 139.6 (PhC4), 135.7 (PhC1), 134.2 (Ar C2/6), 129.7 (Ar C3/C5), 124.5 (C8a), 31.1 (C6 or C7), 28.9 (C4), 27.5 (C8), 27.4 (C6 or C7), 26.5 (C5), 21.4 (PhC4-CH₃), 17.8 (PhC2/C6-CH₃).

HRMS (ESI⁺) calc. for $C_{17}H_{22}NO_2$ [M+H]⁺: 304.1188, found: 304.1203 (- 4.93 ppm).

Spectral data in accordance with literature.²⁷

6.12 3-Mesityl-5,6,7,8-tetrahydro-4*H*-cyclohepta[*d*]thiazol-3-ium perchlorate 15²⁷



Thiazol-3-ium perchlorate **15** was prepared according to general **procedure B** from thione **37** (700 mg, 2. 31 mmol, 1.00 equiv.) yielding a beige-brown solid (776 mg, 90%) pure according to ¹H-NMR directly upon precipitation.

m.p. 135.6-135.7 °C (DCM).

 $\mathbf{R}_{\mathbf{f}}$ (DCM:MeOH = 9:1, potassium permanganate stain): 0.43.

¹**H NMR (400 MHz, CDCl**₃) δ 9.61 (s, 1H, H2), 7.06 (s, 2H, Ph-H3/H5), 3.28 – 3.04 (m, 2H, H8), 2.58 – 2.48 (m, 2H, H4), 2.37 (s, 3H, PhCH₃(C4)), 2.00 – 1.93 (m, 8H, PhCH₃(C2/5), H6), 1.91 – 1.82 (m, 2H, H7), 1.70 – 1.61 (m, 2H, H5).

¹³C NMR (101 MHz, CDCl₃) δ 155.5 (C2), 148.0 (C3a), 142.2 (PhC4), 141.1 (C8a), 134.1 (PhC2/C6), 132.9 (PhC1), 130.3 (PhC3/C5), 30.9 (C6), 28.3 (C8), 27.0 (C4), 26.8 (C7), 25.7 (C5), 21.3 (PhCH₃(C4)), 17.4 (PhCH₃(C2/C6)).

HRMS (ESI⁺) calc. for $C_{17}H_{22}NS^+$ [M-ClO₄]⁺: 272.1467, found: 272.1479 (-4.22 ppm).

Spectral data in accordance with literature.²⁷

6.13 **3**-(2,6-Diisopropylphenyl)-3,4,5,6,7,8-hexahydro-2*H*-cyclohepta[*d*]thiazole-2-thione 38²⁷



Thione **38** was prepared according to general **procedure A** from 2,6-diisopropylaniline (928 mg, 5.23 mmol, 1.00 equiv.) yielding a slightly yellow solid (972 mg, 54%) pure according to ¹H-NMR directly after precipitation.

m.p. 165.0-165.1 °C (*n*-pentane).

 $\mathbf{R}_{\mathbf{f}}$ (LP:EtOAc = 10:1, potassium permanganate stain): 0.31.

¹**H NMR (400 MHz, CDCl3)** δ 7.47 (t, *J* = 7.8 Hz, 1H, PhH4), 7.29 (d, *J* = 7.8 Hz, 2H, PhH3/H5), 2.68 – 2.62 (m, 2H, H8), 2.46 (sept, *J* = 6.8 Hz, 2H, Ph-CH), 2.26 – 2.14 (m, 2H, H4), 1.88 – 1.70 (m, 4H, H6 & H7), 1.59 – 1.50 (m, 2H, H5), 1.28 (d, *J* = 6.8 Hz, 6H, CH(CH₃)₂), 1.13 (d, *J* = 6.9 Hz, 6H, CH(CH₃)₂).

¹³C NMR (101 MHz, CDCl₃) δ 186.8 (C2), 146.5 (PhC2/C6), 141.9 (C3a), 133.8 (PhC1), 130.4 (PhC4), 124.7 (PhC3/C5), 124.2 (C8a), 31.1 (C6), 29.5 (C4), 28.9 (<u>C</u>H(CH₃)₂), 27.6 (C8), 27.3 (C7), 26.4 (C5), 24.5, 24.0 (2 × CH(<u>C</u>H₃)₂).

HRMS (**ESI**⁺) calc. for C₂₀H₂₈NS₂ [M+H]⁺: 346.1658, found: 346.1675 (- 5.11 ppm).

Spectral data in accordance with literature.²⁷

6.14 3-(2,6-Diisopropylphenyl)-5,6,7,8-tetrahydro-4*H*-cyclohepta[*d*]thiazol-3-ium perchlorate 16²⁷



3-(2,6-Diisopropylphenyl)-5,6,7,8-tetrahydro-4*H*-cyclohepta[*d*]thiazol-3-ium perchlorate **16** was prepared according to general **procedure B** from **38** (700 mg, 2.03 mmol, 1.00 equiv.) yielding a beige foam (696 mg, 83%) of the target compound pure according to ¹H-NMR directly upon evaporation.

m.p. 96.7-97.2 °C (DCM).

 $\mathbf{R}_{\mathbf{f}}$ (DCM:MeOH = 9:1, potassium permanganate stain): 0.49.

¹**H NMR (400 MHz, CDCl**₃) δ 9.65 (s, 1H, H2), 7.59 (t, *J* = 7.9 Hz, 1H, Ph H4), 7.35 (d, *J* = 7.9 Hz, 2H, Ph H3/H5), 3.22 – 3.14 (m, 1H, H8), 2.59 – 2.46 (m, 1H, H4), 2.05 (sept, *J* = 6.8 Hz, 2H, C<u>H</u>(CH₃)₂), 1.99 – 1.92 (m, 1H, H6), 1.90 (q, *J* = 4.7, 4.1 Hz, 1H, H7), 1.62 (pent, *J* = 5.1 Hz, 1H, H5), 1.16 (d, *J* = 6.9 Hz, 6H, C<u>H</u>₃-CH).

¹³C NMR (101 MHz, CDCl₃) δ 155.4 (C2), 148.8 (C3a), 145.0 (PhC2/C6), 141.2 (C8a), 132.6 (PhC4), 132.2 (PhC1), 125.3 (PhC3/C5), 30.9 (C6), 28.9 (<u>C</u>H(CH₃)₂), 28.3 (C8), 27.4 (C4), 26.7 (C7), 25.6 (C5), 24.9, 23.3 (2 × CH(<u>C</u>H₃)₂).

HRMS (**ESI**⁺) calc. for C₁₀H₂₈NS [M-ClO₄]⁺: 314.1937, found: 314.1951 (-4.61 ppm).

Spectral data in accordance with literature.²⁷

6.15 1,3,4-Triphenyl-1,2,4-triazol-1-ium perchlorate 17³³



Benzanilide 41 (4.00 g, 20.3 mmol, 1.00 equiv.) was dissolved in toluene (20 ml) in a flask equipped with a reflux condenser with dry tube (CaCl₂). The suspension was stirred for several minutes and then thionyl chloride (7.24 g, 60.8 mmol, 3.00 equiv.) was added at once with a syringe. Then the reaction mixture was heated to 80 °C and stirred there overnight with the solution turning yellow. The reflux condenser was removed, and a distillation head was added. The solvent was evaporated (up to 5 mbar at 60 °C). a yellow solution remained, which solidified upon cooling to rt. THF (anh., 20 ml) was added with the solution turning yellow again. NEt₃ (4.24 ml, 30.4 mmol, 1.5 equiv.) was added slowly and phenylhydrazine (1.99 ml, 20.3 mmol, 1.1 equiv.) was added in a way to keep the temperature under 40 °C. A colourless solid precipitated and the yellow colour got more intense. The suspension was stirred at rt for 3 h, when reaction monitoring via TLC (LP/EtOAc 9:1) showed complete conversion of starting material and formation of a new, not smearing spot. The reagents and the solvent were removed in vacuo (40 °C, 40 mbar) and a thick, honey like, orange-yellow coloured residue remained with the crude NMR being reported in 9.6 and matching intermediate 42. The residue was treated with 2% HOAc (36 ml) and heated up to 70°C under stirring for 30 min, until a homogenous suspension was observed. The precipitate was vacuum filtered and washed with H₂O and MeOH. After washing with MeOH, a slightly yellow solid (2.80 g, 48%) was observed.

Formic acid (8.46 ml, 224 mmol, 23 equiv.) and Ac₂O (16.6 ml, 175 mmol, 18 equiv.) were mixed and heated up to 60 °C for 15 min. After cooling to rt, the intermediate **42** (2.80 g, 9.74 mmol, 1.00 equiv.) was added portionwise under cooling with a water bath over 10 min. The solution was stirred for 20 h. The solvent was evaporated (60 °C, up to 45 mbar) and HClO₄

solution (35%, 16.8 ml) was added. A solid precipitated and the rm was stirred for 30 min. After adding 5 ml H₂O the reaction mixture was stirred for 10 min. The precipitate was vacuum filtered and washed with H₂O and MeOH. The target compound **17** was obtained as a slightly beige solid (2.50 g, 65%, over 2 steps: 31%) pure according to ¹H-NMR.

m.p. 232-233 °C (MeOH), (lit.³³ 220 °C).

¹**H** NMR (400 MHz, DMSO-*d*₆) δ 11.35 (s, 1H, H2), 8.09 (d, *J* = 7.9 Hz, 2H, PhH), 7.79 (t, *J* = 7.7 Hz, 3H, PhH), 7.73 – 7.67, 7.66 – 7.61, 7.57 – 7.51 (3 × m, 7H; 2H; 6H, PhH).

¹³C NMR (101 MHz, DMSO-*d*₆) δ 153.29, 143.18, 134.87, 132.37, 132.07, 131.57, 130.92, 130.46, 130.25, 129.33, 129.27, 126.58, 122.39, 120.66.

HRMS (ESI⁺) Exact mass calc. for $C_{20}H_{16}N_3$ [M-ClO₄]⁺: 298.1361, found: 298.1344 (5.49 ppm).

Spectral data in accordance with literature.³³

7 NMR spectra of dehomologation products

7.1 1-O-Acetyl-2,3-O-isopropylidene-L-erythrose 7b

1H NMR (CDCl3, 400 MHz)





13C NMR (CDCl3, 101 MHz)

(3S)-β-acetoxy-γ-butyrolactone 9b 7.2

1H NMR (CDCl3, 400 MHz)



7.3 3,5-O-Diacetyl-2-deoxy-D-ribono-1,4-lactone 4b

1H NMR (CDCl3, 400 MHz)



8 NMR spectra of sugar substrates

8.1 NMR-spectra of isolated 3-*O*-benzyl-1,2:5,6-di-*O*-isopropylidene-α-D-

glucofuranose 21









NMR-spectra of isolated 3-O-benzyl-D-glucose 1a 8.2

1H NMR (DMSO, 400 MHz)

13C NMR (DMSO, 101 MHz)



8.3 NMR-spectra of isolated methyl 6-O-(tert-butyldimethylsilyl)-α-D-

mannopyranoside 23

1H NMR (MeOD, 600 MHz)



13C NMR (MeOD, 101 MHz)



8.4 NMR-spectra of isolated methyl 6-*O*-(*tert*-butyldimethylsilyl)-3-*O*-benzyl-α-Dmannopyranoside 24

1H NMR (MeOD, 600 MHz)



13C NMR (MeOD, 101 MHz)



8.5 NMR-spectra of isolated methyl 3-*O*-benzyl-α-D-mannopyranoside 25



13C NMR (MeOD, 101 MHz)









8.7 NMR-spectra of isolated 3-*O*-benzyl-α-D-mannose 1b

1H NMR (MeOD, 400 MHz)





8.8 NMR-spectra of isolated 2-O-benzyl-D-arabinose 3

1H NMR (MeOD, 400 MHz)



13C NMR (MeOD, 101 MHz)



8.9 NMR-spectra of isolated 2-deoxy-D-ribono-1,4-lactone 4a

1H NMR (MeOD, 400 MHz)



8.10 NMR-spectra of isolated 3,4-O-isopropylidene L-arabinose 6

1H NMR (MeOD, 400 MHz)





8.11 NMR-spectra of isolated 3-O-(4-nitrophenyl)-1,2:5,6-di-O-isopropylidene-α-D-

glucofuranose 31

1H NMR (CDCl3, 400.13 MHz)



8.12 NMR-spectra of isolated 3-O-(4-nitrophenyl)-D-glucose 10

1H NMR (D2O, 400.13 MHz)



9 NMR spectra of precatalysts

9.1 NMR-spectra of isolated 3,4,5-trimethylthiazol-3-ium iodide 2

1H NMR (DMSO, 400 MHz)



9.2 NMR-spectra of isolated 3-Ethyl-4,5-dimethylthiazol-3-ium iodide 40

1H NMR (MeOD, 400 MHz)



1H NMR (CDCl3, 400 MHz) **7.26** CDCI **4.39 4.39 5.35 5.49 5.35 5.49 5.29** Br 1.00_H 1.06^{-1} 1.09 2.99 .82 1.13 5.0 f1 (ppm) 4.5 2.5 0.0 10.0 9.5 9.0 8.5 8.0 7.5 7.0 6.5 6.0 5.5 4.0 3.5 3.0 2.0 1.5 1.0 0.5 13C NMR (CDCl3, 101 MHz) 77.48 CDCl3 77.16 CDCl3 76.84 CDCl3 - 206.45 - 53.86 39.55 34.41 29.73 26.97 25.13 Br

9.3 NMR-spectra of isolated 2-bromocycloheptanone 33

130 120 110 100 f1 (ppm)

220 210

200 190

180 170

160 150 140 130

70

60 50

90 80

40 30

20

0

10

9.4 NMR-spectra of isolated 3-phenyl-3,4,5,6,7,8-hexahydro-2*H*-cyclohepta[*d*]thiazole-

2-thione 34

1H NMR (CDCI3, 400 MHz)



) 110 100 f1 (ppm)

9.5 NMR-spectra of isolated 3-phenyl-5,6,7,8-tetrahydro-4*H*-cyclohepta[*d*]thiazol-3ium perchlorate 12

1H NMR (CDCl3, 400 MHz)



13C NMR (CDCl3, 101 MHz)



9.6 NMR-spectra of isolated 3-(4-methoxyphenyl)-3,4,5,6,7,8-hexahydro-2Hcyclohepta[d]thiazole-2-thione 35

1H NMR (CDCl3, 400.13 MHz)



9.7 NMR-spectra of isolated 3-(4-methoxyphenyl)-5,6,7,8-tetrahydro-4Hcyclohepta[d]thiazol-3-ium perchlorate 13

1H NMR (CDCl3, 400.13 MHz)


9.8 NMR-spectra of crude 3-(4-nitroxyphenyl)-3,4,5,6,7,8-hexahydro-2H-

cyclohepta[d]thiazole-2-thione 36



9.9 NMR-spectra of isolated 3-(4-nitrophenyl)-5,6,7,8-tetrahydro-4*H*-

cyclohepta[d]thiazol-3-ium perchlorate 14

1H NMR (MeOD, 600.15 MHz)



9.10 NMR-spectra of isolated 3-mesityl-3,4,5,6,7,8-hexahydro-2H-cyclohepta[d]thiazole-

2-thione 37



13C NMR (CDCl3, 101 MHz)



9.11 NMR-spectra of isolated 3-mesityl-5,6,7,8-tetrahydro-4*H*-cyclohepta[*d*]thiazol-3-

ium perchlorate 15



13C NMR (CDCl3, 101 MHz)



9.12 NMR-spectra of isolated 3-(2,6-diisopropylphenyl)-3,4,5,6,7,8-hexahydro-2H-

cyclohepta[d]thiazole-2-thione 38







9.13 NMR-spectra of isolated 3-(2,6-diisopropylphenyl)-5,6,7,8-tetrahydro-4*H*cyclohepta[*d*]thiazol-3-ium perchlorate 16

1H NMR (CDCl3, 400 MHz)





9.14 NMR-spectra of crude N,N'-diphenylbenzohydrazonamide 42

1H NMR (CDCl3, 400 MHz)



1H NMR (DMSO, 400 MHz)



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