# Photo-controlled Chirality Transfer and FRET Effects based on Pseudo[3]rotaxane

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Table of Contents

I. Experimental section

II. Figures

Figure S1. <sup>1</sup>H NMR (400 MHz) spectrum of 4 in CDCl<sub>3</sub> at 25 °C.

Figure S2. <sup>13</sup>C NMR (101 MHz) spectrum of 4 in CDCl<sub>3</sub> at 25 °C.

Figure S3. <sup>1</sup>H NMR (400 MHz) spectrum of 5 in CDCl<sub>3</sub> at 25 °C.

Figure S4. <sup>13</sup>C NMR (101 MHz) spectrum of 5 in CDCl<sub>3</sub> at 25 °C.

Figure S5. <sup>1</sup>H NMR spectra (400 MHz,  $CDCl_3/CD_3CN = 2:1$ , 298 K, [(S)-2] = [1] = 1 mM) of (a) free guest (S)-2, (b) an equimolar mixture of (S)-2 and 1, and (c) free crown ether host 1. for the full proton labeling, see Scheme 1.

Figure S6. Selected regions of the ROESY 2D NMR spectrum (400 MHz, CDCl<sub>3</sub>/CD<sub>3</sub>CN = 2:1) of (*R*)-2@1.

Figure S7. Selected regions of the ROESY 2D NMR spectrum (400 MHz, CDCl<sub>3</sub>/CD<sub>3</sub>CN = 2:1) of (*S*)-2@1.

Figure S8. ESI-MS (low resolution) spectrum of a) (*R*)-2@1, b) (*S*)-2@1 in CHCl<sub>3</sub>/CH<sub>3</sub>CN (2:1) at 298 K. The peak at m/z = 946.6 is assigned to the  $[(R/S)-2@1]^{2+}$ .

Figure S9. DOSY-NMR spectrum (400 MHz, CDCl<sub>3</sub>/CD<sub>3</sub>CN = 2:1, 298 K) of 1 (1 mM), plotted using the log values of the diffusion constant.

Figure S10. DOSY-NMR spectrum (400 MHz,  $CDCl_3/CD_3CN = 2:1$ , 298 K) of (*R*)-2 (1 mM), plotted using the log values of the diffusion constant.

Figure S11. DOSY-NMR spectrum (400 MHz,  $CDCl_3/CD_3CN = 2:1$ , 298 K) of (S)-2 (1mM), plotted using the log values of the diffusion constant.

Figure S12. DOSY-NMR spectrum (400 MHz,  $CDCl_3/CD_3CN = 2:1, 298$  K) of (*R*)-2@1 (1 mM), plotted using the log values of the diffusion constant.

Figure S13. DOSY-NMR spectrum (400 MHz, CDCl<sub>3</sub>/CD<sub>3</sub>CN = 2:1, 298 K) of (*S*)-2@1 (1 mM), plotted using the log values of the diffusion constant.

Figure S14. Fluorescence spectra of supramolecular assembly of 1 and (R)-2 ( $6.0 \times 10^{-5}$  M) with

the addition of 1 ( $\lambda_{ex} = 305$  nm).

Figure S15. Nonlinear least-squares analysis of the emission (F,  $\lambda = 365$  nm) of a) (*R*)-2@1, b) (*S*)-2@1 to calculate the complex binding constant a) K = 5.2 × 10<sup>4</sup> M<sup>-1</sup>, b) K = 4.9 × 10<sup>4</sup> M<sup>-1</sup>.

Figure S16. Molecular energy minimization structure through molecular modulation of 2@1.

Figure S17. Circular dichroism spectra of (R)-2, (S)-2, 1 at 0.01m M.

Figure S18. a) Circular dichroism spectra of (R/S)-2@1 after irritated at 365 nm for 10 h, b) Circular dichroism spectra of a) after heated at 100°C for 10 h under N<sub>2</sub>.

Figure S19. Circular dichroism spectra of (R/S)-2@1 from 250 to 350 nm.

Figure S20. Circular dichroism spectra of (*R/S*)-2 from 360 to 430 nm.

Figure S21. Normalized induced circular dichroism spectra of a), (S)-2/(R)-2=1/9, 2/8, 4.5/5.5.

b), (S)-2/(R)-2=9/1, 5.75/4.25, 6/4. in CHCl<sub>3</sub>/CH<sub>3</sub>CN at 298 k ([1] = [(R)-2 + (S)-2] = 0.3 mM).

Scheme S22. The model molecules  $1_{mod}$  and  $2_{mod}$  of host and guest.

Table S1. Important electronic excited states and their NTOs analysis for (R)-2@1 and (R)-2<sub>mod</sub>.

Figure S23. The direction of rotation and signal of  $1_{mod}$ - $D_{2h}$ ,  $1_{mod}$ - $D_2$  and its' enantiomer  $1_{mod}$ - $D_{2-E}$ .

Table S2. Important electronic excited states and their NTOs analysis for  $1_{mod}$ - $D_{2h}$ ,  $1_{mod}$ - $D_2$  and  $1_{mod}$ - $D_{2-E}$ .

Figure S24. The transitions of (a) the CT in  $S_2$  and (b) the main LE in  $S_4$ .

Figure S25. The MOs participated in the transition of (*R*)-2@1.

Figure S26. The MOs participated in the transition of (*R*)-2mod.

Figure S27. The MOs participated in the transition of  $1_{mod}$ - $D_{2h}$ ,  $1_{mod}$ - $D_2$  and  $1_{mod}$ - $D_{2-E}$ . REFERENCES

### **Experimental Procedures**

All the reagents and solvents were commercially available and used as received unless otherwise specified purification. Compounds, 3<sup>a</sup>, were prepared according to the literatures procedure. Column chromatography was performed on 200-300 mesh silica gel.



Scheme S1. Synthetic route of (*R/S*)-2.

**Preparation of 4.** Complex **3** (309 mg, 1.2 mmol), 1,1'-Bi-2-naphthol (114.5 mg, 0.4 mmol), and K<sub>2</sub>CO<sub>3</sub> (550 mg, 4 mmol) were suspended in anhydrous MeCN (20 mL). The reaction mixture was heated to reflux under argon atmosphere and stirred for 24 h. After cooling down to r. t., the reaction mixture was filtered off and washed with CH<sub>2</sub>Cl<sub>2</sub> (100 mL). The filtrate was concentrated under reduced pressure, which was further purified by column chromatography (SiO2, hexane : ethyl acetate = 6:1) to yield the product 4. (199 mg, 82%). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>, 298 K)  $\delta$ : 9.86 (s, 2H), 7.91 (d, *J* = 9.0 Hz, 2H), 7.83 (d, *J* = 8.1 Hz, 2H), 7.77 (d, *J* = 8.7 Hz, 4H), 7.40 (d, *J* = 9.0 Hz, 2H), 7.34–7.28 (m, 2H), 7.20 (q, *J* = 8.6 Hz, 4H), 6.74 (d, *J* = 8.6 Hz, 4H), 4.18–4.02 (m, 4H), 4.00–3.90 (m, 2H), 3.56–3.40 (m, 4H). <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>)  $\delta$ : 190.96 , 164.32 , 154.69, 134.41 , 132.12 , 130.00 , 129.74 , 129.68, 128.20 , 126.56 , 125.74 , 124.02 , 121.17 , 116.29 , 114.95 , 69.68 , 67.74 , 60.64. m/z calcd for C<sub>42</sub>H<sub>39</sub>O<sub>6</sub><sup>+</sup>: 639.2747 (M+H<sup>+</sup>), found:639.2743.

**Preparation of 5 and** (*R/S*)-2. A solution of 4 (1277.5 mg, 2.0 mmol) and benzylamine (437.35 mg, 4.0 mmol) in EtOH (50 mL) was heated under reflux for 12 h. After the reaction mixture was cooled to r.t., NaBH<sub>4</sub> (0.76 g, 20.0 mmol) was added to this solution in small portions, and the reaction mixture stirred at r.t. for another 12 h. The solvent was evaporated under reduced pressure and the residue was partitioned between  $CH_2Cl_2$  (50 mL) and water (50 mL). The aqueous layer was further washed with  $CH_2Cl_2$  (3 × 100 mL). The organic phases were combined and dried over anhydrous MgSO<sub>4</sub>. Filtration, followed by evaporation

gave a white solid which was subjected to column chromatography (SiO<sub>2</sub>,  $CH_2Cl_2$ : MeOH = 100:3). Pure 5 was obtained as a white solid (1.26 g, 80%). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$ 7.89 (d, J = 9.0 Hz, 2H), 7.82 (d, J = 8.1 Hz, 2H), 7.38 (d, J = 9.0 Hz, 2H), 7.34–7.11 (m, 20H), 6.66 (d, J = 8.4 Hz, 4H), 4.01 (dt, J = 9.3, 5.9 Hz, 2H), 3.90 (dt, J = 13.7, 6.6 Hz, 2H), 3.76 (s, 4H), 3.70 (s, 4H), 3.45 (t, J = 6.2 Hz, 4H), 1.98 (s, 2H), 1.61-1.43 (m, 4H), 1.31 (qq, J= 13.3, 6.5 Hz, 4H). <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>)  $\delta$ : 158.27, 154.66, 140.39, 134.36, 132.12, 129.58, 129.50, 129.48, 128.65, 128.44, 128.13, 127.21, 126.43, 125.69, 123.83, 69.50, 67.20, 53.72, 53.18, 52.71, 26.06, 25.61. m/z calcd for C<sub>56</sub>H<sub>58</sub>N<sub>2</sub>O<sub>6</sub><sup>2+</sup>: 411.2199 (M  $+ 2H^+$ ), found:411.2195. Then compound 5 (100 mg, 0.127 mmol) was dissolved in dry CH<sub>2</sub>Cl<sub>2</sub> (20 mL) and TFA (0.19 mL, 0.3 mmol) TFA (0.32 mL, 5.0 mmol) was added at room tempeature. After the solution stirred for 2 h under argon atmosphere, the solvent was removed under vacuum. The residue was dissolved in MeOH, and then saturated NH<sub>4</sub>PF<sub>6</sub> (20 mL, aq) was added and stirred for several minutes to yield a yellow creamy solid. The residue was extracted by CH<sub>2</sub>Cl<sub>2</sub>. The organic layer was washed by H<sub>2</sub>O for three times and dried over anhydrous Na<sub>2</sub>SO<sub>4</sub>, and thesolvent was removed under vacuum to give the compound (R/S)-2 (123 mg, 90%). <sup>1</sup>H NMR (400 MHz, CD<sub>3</sub>CN)  $\delta$  8.01 (d, J = 8.9 Hz, 2H), 7.91 (d, J = 7.9 Hz, 2H), 7.58–7.42 (m, 12H), 7.34 (dd, J = 17.3, 7.9 Hz, 6H), 7.21 (t, J = 7.2 Hz, 2H), 7.04 (d, J = 8.2 Hz, 2H), 6.79 (d, J = 8.0 Hz, 4H), 4.20 (d, J = 16.7 Hz, 8H), 4.14–3.97 (m, 4H), 1.64–1.47 (m, 8H), 1.39 (d, J = 12.0 Hz, 8H). <sup>13</sup>C NMR (101 MHz, CD<sub>3</sub>CN)  $\delta$  211.12, 203.99, 160.76, 155.26, 134.74, 132.61, 131.31, 130.98, 130.56, 130.17, 130.10, 129.91, 128.85, 127.13, 125.65, 124.43, 122.81, 120.79, 116.48, 115.55, 69.74, 68.04, 61.99, 51.99, 51.86, 50.32, 28.10 m/z calcd for  $(2-2PF_6)^{2+}$ : 411.2199, found: 411.2195.

## Measurements

**NMR spectroscopy.** <sup>1</sup>H and <sup>13</sup>C NMR spectra and 2D ROESY were recorded on a Brucker AV400 spectrometer.

**Fluorescence spectroscopy.** Steady-state fluorescence spectra were recorded in a conventional quartz cell (light path 10 mm) on a Varian Cary Eclipse equipped with a Varian Cary single-cell peltier accessory to control temperature.

**UV/Vis spectroscopy.** UV/Vis spectra and the optical transmittance were recorded in a quartz cell (light path 10 mm) on a Shimadzu UV-3600 spectrophotometer equipped with a PTC-348WI temperature controller.

**ESI-MS spectroscopy.** Electrospray ionization mass spectra (ESI-MS) were measured by Agilent 6520 Q-TOF-MS.

**CD spectroscopy.** CD spectra were recorded on a BioLogic MOS500 spectropolarimeter in a quartz cell of 10 mm light path.

**Switch experiments.** The switch experiments were carried out using a photochemical reaction apparatus with a 500W Hg lamp.

**Theoretical calculation section.** To further understand the mechanism of the surprising ICD signals, density functional theory (DFT) and time-dependent density functional theory (TD-DFT) calculations were performed by Gaussian16.<sup>[b]</sup> The geometry optimization were carried out at B3LYP-GD3<sup>[c-d]</sup> / 6-31G(d,p) <sup>[e]</sup> level with symmetry constraint. TD-DFT and natural transition orbitals (NTOs) were calculated using the M06-2X-GD3<sup>[d,f]</sup> and 6-311G(d,p)<sup>[e]</sup> basis set in chloroform using the integral equation formalism polarizable continuum model (IEF-PCM).<sup>[g]</sup>As a comparison, model molecules  $1_{mod}$  and  $2_{mod}$  (Scheme.S22) were also calculated.

# Figures



Figure S1. <sup>1</sup>H NMR (400 MHz) spectrum of 4 in CDCl<sub>3</sub> at 25 °C.









Figure S4. <sup>13</sup>C NMR (101 MHz) spectrum of 5 in CDCl<sub>3</sub> at 25 °C.



Figure S5. <sup>1</sup>H NMR spectra (400 MHz,  $CDCl_3/CD_3CN = 2:1, 298$  K, [(S)-2] = [1] = 1 mM ) of (a) free guest (S)-2, (b) an equimolar mixture of (S)-2 and 1, and (c) free crown ether host 1. for the full proton labeling, see Scheme 1.



Figure S6. Selected regions of the ROESY 2D NMR spectrum (400 MHz,  $CDCl_3/CD_3CN = 2:1$ ) of (*R*)-2@1.



Figure S7. Selected regions of the ROESY 2D NMR spectrum (400 MHz,  $CDCl_3/CD_3CN = 2:1$ ) of (S)-2@1.



Figure S8. ESI-MS (low resolution) spectrum of a) (*R*)-2@1, b) (*S*)-2@1 in CHCl<sub>3</sub>/CH<sub>3</sub>CN (2:1) at 298 K. The peak at m/z = 946.6 is assigned to the  $[(R/S)-2@1]^{2+}$ .



**Figure S9.** DOSY-NMR spectrum (400 MHz, CDCl<sub>3</sub>/CD<sub>3</sub>CN = 2:1, 298 K) of **1** (1 mM), plotted using the log values of the diffusion constant.



**Figure S10.** DOSY-NMR spectrum (400 MHz,  $CDCl_3/CD_3CN = 2:1, 298$  K) of (*R*)-2 (1 mM), plotted using the log values of the diffusion constant.



**Figure S11.** DOSY-NMR spectrum (400 MHz, CDCl<sub>3</sub>/CD<sub>3</sub>CN = 2:1, 298 K) of (*S*)-2 (1mM), plotted using the log values of the diffusion constant.



Figure S12. DOSY-NMR spectrum (400 MHz, CDCl<sub>3</sub>/CD<sub>3</sub>CN = 2:1, 298 K) of (*R*)-2@1 (1 mM), plotted using the log values of the diffusion constant.



Figure S13. DOSY-NMR spectrum (400 MHz, CDCl<sub>3</sub>/CD<sub>3</sub>CN = 2:1, 298 K) of (S)-2@1 (1 mM), plotted using the log values of the diffusion constant.



**Figure S14.** Fluorescence spectra of supramolecular assembly of **1** and (*R*)-**2** ( $6.0 \times 10^{-5}$  M) with the addition of **1** ( $\lambda_{ex} = 305$  nm).



**Figure S15.** Nonlinear least-squares analysis of the emission (F,  $\lambda = 365$  nm) of a) (*R*)-2@1, b) (*S*)-2@1 to calculate the complex binding constant a)  $K = 5.2 \times 10^4$  M<sup>-1</sup>, b)  $K = 4.9 \times 10^4$  M<sup>-1</sup>.



Figure S16. The optimized structure the assembly (R)-2@1<sup>[h]</sup>.



Figure S17. Circular dichroism spectra of (R)-2, (S)-2, 1 at 0.01m M.



**Figure S18.** a) Circular dichroism spectra of (*R/S*)-2@1 after irritated at 365 nm for 10 h, b) Circular dichroism spectra of a) after heated at 100°C for 10 h under  $N_2$ .



Figure S19. Circular dichroism spectra of (*R/S*)-2@1 from 250 to 350 nm.



Figure S20. Circular dichroism spectra of (R/S)-2 from 360 to 430 nm.



Figure S21. Normalized induced circular dichroism spectra of a), (S)-2/(R)-2=1/9, 2/8, 4.5/5.5. b), (S)-2/(R)-2=9/1, 5.75/4.25, 6/4. in CHCl<sub>3</sub>/CH<sub>3</sub>CN at 298 k ([1] = [(R)-2 + (S)-2] = 0.3 mM).



Scheme S22. The model molecules  $1_{mod}$  and  $2_{mod}$  of host and guest.

According to the simulation of electronic circular dichroism (ECD) spectrum by TD-DFT, we found that the signal of the excited state  $S_2$  was opposite to the signal in the shorterwavelength region for the assembly (R)-2(a)1, which was consistent with the experimental results. The NTOs calculation shown that the excited state S2 was related to the transition local excitation in anthracene and charge-transfer excitation from guest to host (Table.S1 and Figure.S25). To be noted, there is a signal of  $S_2$  whose direction is the same as the signal in shorter-wavelength region, but it is not shown in experiment. We think it is the result of vibration racemization. When the benzene ring is perpendicular to the anthracene ring, the chromophore is D<sub>2h</sub> symmetry, and there is no CD signal due to no chirality. When the benzene ring rotated along the anthracene ring, the symmetry plane disappears and the symmetry of the chromophore group become D<sub>2</sub>. The D<sub>2</sub> chromophore has CD signals accompanying its enantiomer has opposite CD signals.(Table.S2 and Figure.S23). Based on the Franck-Condon principle, the rate of the electron transition is much larger than the rate of molecular vibration. If the probability of molecular vibration to the two opposite directions is equal, the CD signal of local excitation will not be observed. However, for the excited state S<sub>2</sub> of the assembly, the vibration racemization will not occur at CT when the signal of LE disappear, but the strength of signal will decrease sharply in S2. Moreover, we notice that the symmetry of the phase of the occupied orbital of the CT in  $S_2$  is not similar with that of main transition in  $S_4$  (Figure.S24), which may be the result of the selectivity of the orbital

symmetry in transition, and it is also the reason of that the exhibited ICD signals in anthracene were opposite to that of (S)-2 or (R)-2 in binaphthyl.

	<b>2</b> <sub>mod</sub> .								
Complex	Electronic				Assignment				
	transition	λ	Energy	R <sub>vel</sub>	(H=HOMO,				
					L=LUMO)				
(R)-2@1	$S_0 \rightarrow S_1$	376.95nm	3.2891eV	20.5426	(0.70)H-1→L				
	$S_0 \rightarrow S_2$	315.01nm	3.9359eV	-1.2265	(0.44)H-11→L				
					(0.17)H-4→L				
					$(0.48)\text{H-1}{\rightarrow}\text{L+2}$				
	$S_0 \rightarrow S_3$	298.82nm	4.1492eV	39.7861	(-0.26)H-2→L+3				
					(0.63)H→L+1				
					(0.11)H→L+21				
	S <sub>0</sub> →S <sub>4</sub>	288.20nm	4.3020eV	66.7046	(-0.15)H-6→L+1				
					(0.13)H-3→L+3				
					(-0.11)H-3→L+17				
					(-0.38)H-2→L+1				
					(-0.11)H-2→L+21				
					(0.50)H→L+3				
					(0.16)H→L+17				
( <b>R</b> )-2 <sub>mod</sub>	$S_0 \rightarrow S_1$	296.62nm	4.1798eV	-12.0321	(-0.35)H-1→L+1				
					(0.57)H→L				
					(-0.12)H→L+3				
	$S_0 \rightarrow S_2$	289.33nm	4.2852eV	75.9631	(-0.15)H-3→L				
					(-0.10)H-3→L+3				
					(0.13)H-2→L+1				
					(-0.12)H-2→L+2				
					(-0.38)H-1→L				
					(0.11)H-1→L+3				
					$(0.50)H\rightarrow L+1$				
					$(0.14)H\rightarrow L+2$				

Table S1. Important electronic excited states and their NTOs analysis for (R)-2@1 and (R)-



Figure S23. The direction of rotation and signal of  $1_{mod}$ - $D_{2h}$ ,  $1_{mod}$ - $D_2$  and its' enantiomer  $1_{mod}$ - $D_{2-E}$ .

	inou 2 E					
	Electronic				Assignment	
Complex	transition	λ	Energy	R <sub>vel</sub>	(H=HOMO,	
					L=LUMO)	
	$S_0 \rightarrow S_1$	363.86nm	3.4075eV	0	(0.70)H→L	
					(0.47)H-1→L	
$1_{mod}$ - $D_{2h}$	$S_0 \rightarrow S_2$	310.16nm	3.9974eV	0	(0.49)H→L+1	
					(-0.14)H→L+5	
	$S_0 \rightarrow S_1$	366.76nm	3.3806eV	7.5272	(0.70)H→L	
$1_{mod}$ -D <sub>2</sub>					(0.47)H-1→L	
	$S_0 \rightarrow S_2$	310.45nm	3.9937 eV	1.3258	(0.50)H→L+1	
					(-0.14)H→L+5	
1 <sub>mod</sub> -D <sub>2-E</sub>	$S_0 \rightarrow S_1$	366.76nm	3.3806eV	-7.5272	(0.70)H→L	
					(0.47)H-1→L	
	$S_0 \rightarrow S_2$	310.45nm	3.9937 eV	-1.3258	(0.50)H→L+1	
					(-0.14)H→L+5	

Table S2. Important electronic excited states and their NTOs analysis for  $1_{mod}$ - $D_{2h}$ ,  $1_{mod}$ - $D_{2}$ and  $1_{mod}$ - $D_{2-E}$ .



Figure S24. The transitions of (a) the CT in  $S_2$  and (b) the main LE in  $S_4$ .



Figure S25. The MOs participated in the transition of (*R*)-2@1.



Figure S26. The MOs participated in the transition of (*R*)-2mod.



Figure S27. The MOs participated in the transition of  $1_{mod}$ - $D_{2h}$ ,  $1_{mod}$ - $D_2$  and  $1_{mod}$ - $D_{2-E}$ .

### References

[a] W. Jiang, K. Nowosinski, N. L. Löw, E. V. Dzyuba, F. Klautzsch, A. Schäfer, J. Huuskonen, K. Rissanen and C. A. Schalley, J. Am. Chem. Soc., 2012, 134, 1860-1868.

[b]. M. J. Frisch, G. W. Trucks, H. B. Schlegel, G. E. Scuseria, M. A. Robb, J. R. Cheeseman, G. Scalmani, V. Barone, G. A. Petersson, H. Nakatsuji, X. Li, M. Caricato, A. V. Marenich, J. Bloino, B. G. Janesko, R. Gomperts, B. Mennucci, H. P. Hratchian, J. V. Ortiz, A. F. Izmaylov, J. L. Sonnenberg, D. Williams-Young, F. Ding, F. Lipparini, F. Egidi, J. Goings, B. Peng, A. Petrone, T. Henderson, D. Ranasinghe, V. G. Zakrzewski, J. Gao, N. Rega, G. Zheng, W. Liang, M. Hada, M. Ehara, K. Toyota, R. Fukuda, J. Hasegawa, M. Ishida, T. Nakajima, Y. Honda, O. Kitao, H. Nakai, T. Vreven, K. Throssell, J. A. Montgomery, Jr., J. E. Peralta, F. Ogliaro, M. J. Bearpark, J. J. Heyd, E. N. Brothers, K. N. Kudin, V. N. Staroverov,

T. A. Keith, R. Kobayashi, J. Normand, K. Raghavachari, A. P. Rendell, J. C. Burant, S. S. Iyengar, J. Tomasi, M. Cossi, J. M. Millam, M. Klene, C. Adamo, R. Cammi, J. W. Ochterski, R. L. Martin, K. Morokuma, O. Farkas, J. B. Foresman, and D. J. Fox, Gaussian 16, Revision A.03, Gaussian, Inc., Wallingford, CT, 2016.

[c]. (a) A. D. Becke, J. Cherm. Phys., 1993, 98, 5648-5652; (b) A. D. Becke, J. Chem. Phys., 1993, 98, 1372-1377; (c) K. Raghavachari, Theor. Chem. Acc., 2000, 103, 361-363.

[d]. (a) S. Grimme, J. Antony, S. Ehrlich and H. Krieg, *J. Chem. Phys.*, 2010, **132**, 154104; (b) S. Grimme, *J. Comput. Chem.*, 2006, **27**, 1787-1799.

[e]. P. C. Hariharan and J. A. Pople, Theor. Chim. Acta, 1973, 28, 213-222.

[f]. (a) Y. Zhao and D. G. Truhlar, *Acc. Chem. Res.*, 2008, **41**, 157-167; bY. Zhao and D. G. Truhlar, *Theor. Chem. Acc.*, 2008, **120**, 215-241.

[g]. (a) B. Mennucci and J. Tomasi, *J. Chem. Phys.*, 1997, **106**, 5151-5158; (b) V. Barone and M. Cossi, *J. Phys. Chem. A*, 1998, **102**, 1995-2001.

[h]. C. Y.Legault, CYLview, 1.0b, Université de Sherbrooke, Sherbrooke, Quebec, Canada, 2009, http://www.cylview.org.