

# Multifunctional 1,3-diphenylguanidine for carboxylative cyclization of homopropargyl amines with CO<sub>2</sub> under ambient temperature and pressure

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Contents	Page
1. General information	2
2. Preparation of homopropargyl amines	3-9
3. Reaction condition optimization	10-12
4. General procedure for the carboxylative cyclization reaction	12-19
5. Mechanistic studies	20-27
6. Single-crystal X-ray analysis of bicarbonate salt [DPGH][HCO <sub>3</sub> ]	28-33
7. Single-crystal X-ray analysis of complex formed from DPG and AgSbF <sub>6</sub>	34-41
8. NMR and HPLC spectra	42-136
9. References	137

## 1. General information

Reactions were monitored by thin layer chromatography using UV light, I<sub>2</sub> or KMnO<sub>4</sub> to visualize the course of reaction. Purification of reaction products was carried out by flash chromatography on silica gel. Chemical yields refer to pure isolated substances. Infrared (IR) spectra were obtained using a Bruker tensor 27 infrared spectrometer. Chiral HPLC analysis was performed on a LC-20AD instrument using Daicel Chiracel columns at 25 °C and a mixture of HPLC-grade hexane and isopropanol as eluent. Optical rotation was measured using a (JASCO) P-1030 polarimeter equipped with a sodium vapor lamp at 589 nm. <sup>1</sup>H NMR, <sup>13</sup>C NMR and <sup>31</sup>P NMR spectra were obtained using Bruker DPX-300, 400 and 500 MHz Spectrometer. <sup>1</sup>H-<sup>1</sup>H NOESY spectra were obtained using Bruker DPX-500 MHz Spectrometer. Chemical shifts were reported in ppm with TMS as the internal standard. The following abbreviations were used to designate chemical shift multiplicities: s = singlet, d = doublet, t = triplet, q = quartet, h = heptet, m = multiplet, br = broad.

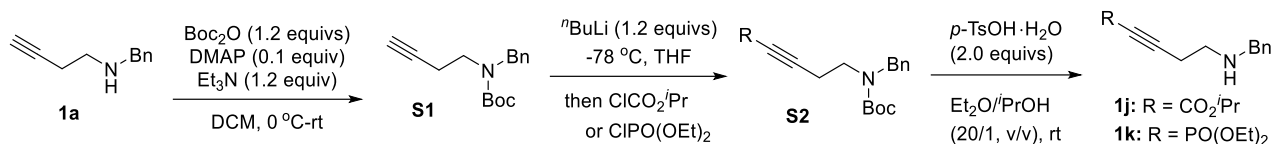
AgSbF<sub>6</sub> (99%) was purchased from Alfa, 1,3-diphenylguanidine was purchased from J&K Scientific. Dichloroethane was purchased from J&K Scientific and used without further purification. The homopropargyl amines **1a-i** and **1l-m** were synthesized according to the literature method.<sup>1</sup>

### List of abbreviation:

Entry	Chemical name	Abbreviation
1	1,3-Diphenylguanidine	DPG
2	1,2,3-Triphenylguanidine	TPG
3	1,1,3,3-Tetramethylguanidine	TMG
4	1,8-Diazabicyclo[5.4.0]undec-7-ene	DBU
5	1,5,7-Triazabicyclo[4.4.0]dec-5-ene	TBD
6	Petroleum ether	PE
7	Diethyl ether	Et <sub>2</sub> O
8	<i>N, N</i> -Dimethyl formamide	DMF
9	Ethyl acetate	EtOAc
10	Dichloroethane	DCE

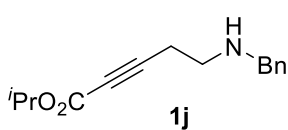
## 2. Preparation of homopropargyl amines

### 2.1. Synthesis of homopropargyl amines **1j** and **1k**.<sup>1-2</sup>

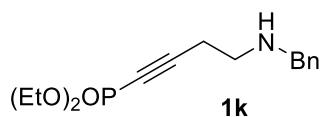


Di-tert-butylidicarbonate (12.0 mmol) was slowly added to a solution of homopropargyl amines (10.0 mmol), DMAP (1.2 mmol, 10 mol%) and  $\text{Et}_3\text{N}$  (12 mmol) in 20 mL  $\text{CH}_2\text{Cl}_2$  at  $0\text{ }^\circ\text{C}$  and the resulting mixture was stirred at ambient temperature for 24 h. Standard extractive work-up followed by silica column chromatography (PE/EtOAc 15:1 to 10:1, v/v) provided tert-butyl benzyl(but-3-yn-1-yl)carbamate **S1** as a pale yellow syrup in 85% yield.

$n\text{-BuLi}$  (1.2 equivs, 2.0 M in hexane) was slowly added at  $-78\text{ }^\circ\text{C}$  to a solution of the compound **S1** obtained above (5.0 mmol) in 15 mL  $\text{Et}_2\text{O}$ . After the mixture had been stirred for 30 min at that temperature, isopropyl carbonochloridate or diethyl phosphorochloridate (6.0 mmol, 1.2 equivs) was introduced, and stirring was continued for another 15 min at  $-78\text{ }^\circ\text{C}$  before the mixture was allowed to reach ambient temperature. After one hour, the reaction was quenched with saturated ammonium chloride aqueous solution, the aqueous layer was extracted with EtOAc, and the combined organic phases were dried over  $\text{Na}_2\text{SO}_4$ . Evaporation of the solvent followed by column chromatography of the residue (PE/EtOAc 10:1 to 4:1, v/v) gave the title compound **1j** or **1k**.

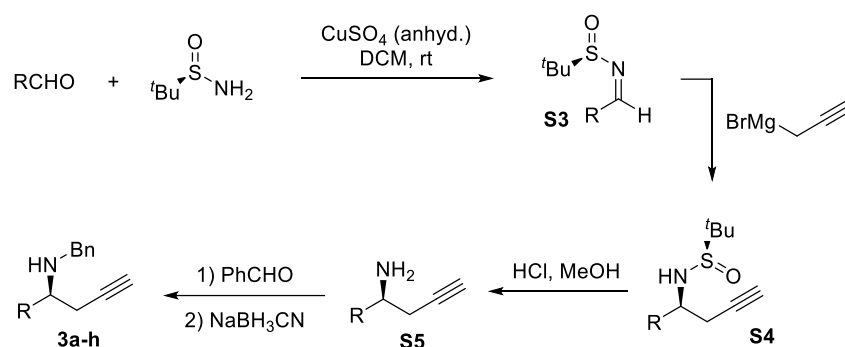


Compound **1j** was prepared in 78% yield according to the general procedure as yellow solid (m.p.  $80\text{-}82\text{ }^\circ\text{C}$ ).  $^1\text{H}$  NMR (400 MHz,  $\text{CDCl}_3$ ):  $\delta$  7.35-7.32 (m, 3H), 7.28-7.26 (m, 2H), 5.12-5.03 (m, 1H), 3.82 (s, 2H), 2.86 (t,  $J = 6.4\text{ Hz}$ , 2H), 2.54 (t,  $J = 6.4\text{ Hz}$ , 2H), 1.63 (s, br, 1H), 1.29 (s, 3H), 1.28 (s, 3H);  $^{13}\text{C}$  NMR (100 MHz,  $\text{CDCl}_3$ ):  $\delta$  153.17, 139.84, 128.42, 128.04, 127.04, 86.64, 74.45, 69.72, 53.24, 46.45, 21.64, 19.96; IR (ATR)  $\nu$  2977.0, 2932.8, 2236.8, 1704.7, 1495.2, 1417.8, 1369.9, 1256.9, 1225.6,  $910.6\text{ cm}^{-1}$ ; HRMS (ESI): Exact mass calcd for  $\text{C}_{15}\text{H}_{20}\text{NO}_2$   $[\text{M}+\text{H}]^+$ : 246.1489, found: 246.1494.



Compound **1k** was prepared in 56% yield according to the general procedure as yellow oil.  $^1\text{H}$  NMR (400 MHz,  $\text{CDCl}_3$ ):  $\delta$  7.34-7.27 (m, 5H), 4.17-4.09 (m, 4H), 3.81 (s, 2H), 2.86 (t,  $J = 6.8$  Hz, 2H), 2.57-2.53 (m, 2H), 1.60 (s, br, 1H), 1.35 (t,  $J = 7.2$  Hz, 6H);  $^{13}\text{C}$  NMR (100 MHz,  $\text{CDCl}_3$ ):  $\delta$  139.76, 128.42, 128.00, 127.07, 100.75 (d,  $J = 52.5$  Hz), 71.71 (d,  $J = 299.5$  Hz), 62.93 (d,  $J = 5.4$  Hz), 53.26, 46.39 (d,  $J = 2,4$  Hz), 20.56 (d,  $J = 4.4$  Hz), 16.05 (d,  $J = 7.0$  Hz);  $^{31}\text{P}$  NMR (122 MHz,  $\text{CDCl}_3$ ):  $\delta$  6.55 (m); IR (ATR)  $\nu$  2982.4, 2203.8, 1716.1, 1568.8, 1453.5, 1367.8, 1251.4, 1163.6, 1099.3, 741.4  $\text{cm}^{-1}$ ; HRMS (ESI): Exact mass calcd for  $\text{C}_{15}\text{H}_{23}\text{NO}_3\text{P}$   $[\text{M}+\text{H}]^+$ : 296.1410, found: 296.1404.

## 2.2. Synthesis of compounds **3a-3h**.<sup>3</sup>



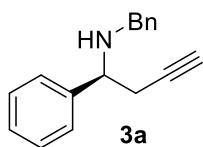
To a solution of (*R*)-*tert*-butanesulfonamide (10.0 mmol) in 20 mL  $\text{CH}_2\text{Cl}_2$  was added anhydrous  $\text{CuSO}_4$  (22.0 mmol) followed by aldehyde (11.0 mmol). The mixture was stirred at room temperature for 12 h. The reaction mixture was filtered through a pad of Celite, and the filter cake was washed well with  $\text{CH}_2\text{Cl}_2$ . The combined organic layers were dried over  $\text{Na}_2\text{SO}_4$  and concentrated give sulfinimine **S3** in almost quantitative yield.

To a solution of the sulfinimine **S3** (10.0 mmol) in 50 mL  $\text{CH}_2\text{Cl}_2$  was added propargyl magnesium bromide ether solution (20.0 mmol) at  $-50$  °C. The mixture was stirred at  $-50$  °C for 2 h and then was allowed to warm to room temperature and stirred overnight. The reaction mixture was quenched with saturated ammonium chloride aqueous solution and the aqueous layer was extracted with  $\text{CH}_2\text{Cl}_2$ . The combined organic layers were dried over  $\text{Na}_2\text{SO}_4$  and concentrated. The residue was purified through silica column chromatography (PE/EtOAc 6/1 to 4/1, v/v) to give sulfinamide **S4** in 75-83% yield.

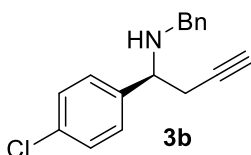


The above sulfinamide **S4** (5.0 mmol) was dissolved in 50 mL MeOH and the solution was cooled to 0 °C. Concentrated hydrochloric acid (11.0 mmol, 2.2 equivs) was added, and the reaction mixture was stirred at 0 °C for 30 minutes. The solvent was evaporated, water was added and acid base work up to give pure amine **S5** in more than 90% yield.

To a mixture of benzaldehydes (4.0 mmol) in 10 mL methanol, the amine **S5** (4.0 mmol) was added. The reaction mixture was stirred at room temperature for 3-4 h, and then sodium cyanoborohydride (3.0 mmol, 0.75 equiv) was added in batches and the mixture was further stirred for another period of 6 h. The reaction was then quenched by the addition of water, and washed with diethyl ether (5 mL× 3). The combined organic phases were washed with saturated aqueous NaCl (10 mL), dried over Na<sub>2</sub>SO<sub>4</sub>, and filtered. Evaporation of the filtrate followed by flash chromatography of the residue (PE/EtOAc 10:1 to 4:1, v/v) gave compounds **3a-3h** successfully.

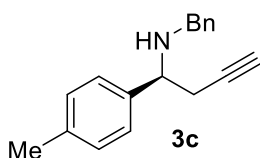


Compound **3a** was prepared in 80% yield according to the general procedure as yellow oil. HPLC analysis (Chiralcel OJ-H, *i*PrOH/hexane = 10/90, 1.0 mL/min, 230 nm; *t<sub>r</sub>* (major) = 10.65 min, *t<sub>r</sub>* (minor) = 13.41 min) gave the isomeric composition of the product: 96% ee; [ $\alpha$ ]<sub>D</sub><sup>25</sup> = -39.9 (c = 1.0, CHCl<sub>3</sub>); <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>):  $\delta$  7.41-7.39 (m, 2H), 7.38-7.34 (m, 2H), 7.32-7.31 (m, 4H), 7.29-7.26 (m, 2H), 3.87 (t, *J* = 6.8 Hz, 1H), 3.72, 3.59 (AB, *J* = 13.2 Hz, 2H), 2.56-2.54 (m, 2H), 2.03-2.01 (m, 1H), 1.88 (s, br, 1H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>):  $\delta$  142.33, 140.15, 128.39, 128.26, 127.98, 127.44, 127.07, 126.80, 81.44, 70.48, 60.58, 51.25, 28.08; IR (ATR)  $\nu$  3290.3, 3061.6, 3026.4, 2918.4, 1602.4, 1493.5, 1355.5, 1201.3, 1073.5, 911.4 cm<sup>-1</sup>; HRMS (ESI): Exact mass calcd for C<sub>17</sub>H<sub>18</sub>N [M+H]<sup>+</sup>: 236.1434, found: 236.1445.

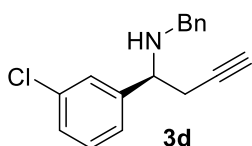


Compound **3b** was prepared in 80% yield according to the general procedure as yellow oil. HPLC analysis (Chiralcel OJ-H, *i*PrOH/hexane = 10/90, 1.0 mL/min, 230 nm; *t<sub>r</sub>* (major) = 9.37 min, *t<sub>r</sub>* (minor) = 10.37 min) gave the isomeric composition of the product: 98% ee; [ $\alpha$ ]<sub>D</sub><sup>25</sup> = -97.5 (c = 1.0, CHCl<sub>3</sub>); <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>):  $\delta$  7.38-7.32 (m, 4H), 7.31-7.29 (m, 2H), 7.27-7.22 (m, 3H), 3.83 (t, *J* = 6.4 Hz, 1H), 3.68, 3.55 (AB, *J* = 13.6 Hz, 2H), 2.51-2.48 (m, 2H), 2.02-2.00 (m, 2H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>):  $\delta$

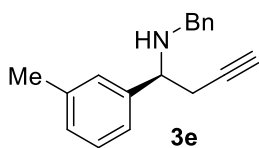
140.95, 140.00, 133.12, 128.64, 128.55, 128.39, 128.01, 126.98, 81.08, 70.74, 60.03, 51.30, 28.13; IR (ATR)  $\nu$  3297.6, 3026.5, 2909.6, 2834.2, 2117.7, 1598.1, 1453.7, 1327.7, 1295.3, 1027.1  $\text{cm}^{-1}$ ; HRMS (ESI): Exact mass calcd for  $\text{C}_{17}\text{H}_{17}\text{ClN}$   $[\text{M}+\text{H}]^+$ : 270.1044, found: 270.1032.



Compound **3c** was prepared in 76% yield according to the general procedure as yellow oil. HPLC analysis (Chiralcel AD-H,  $i$ PrOH/hexane = 1/99, 0.8 mL/min, 230 nm;  $t_r$  (major) = 9.89 min,  $t_r$  (minor) = 11.15 min) gave the isomeric composition of the product: 98% ee;  $[\alpha]_{\text{D}}^{25} = -75.6$  ( $c = 1.0$ ,  $\text{CHCl}_3$ );  $^1\text{H}$  NMR (400 MHz,  $\text{CDCl}_3$ ):  $\delta$  7.31-7.28 (m, 4H), 7.26-7.25 (m, 2H), 7.23-7.20 (m, 1H), 7.16-7.13 (m, 2H), 3.82 (t,  $J = 6.4$  Hz, 1H), 3.69, 3.55 (AB,  $J = 13.2$  Hz, 2H), 2.52-2.49 (m, 2H), 2.33 (s, 3H), 2.04 (s, br, 1H), 1.99-1.97 (m, 1H);  $^{13}\text{C}$  NMR (100 MHz,  $\text{CDCl}_3$ ):  $\delta$  140.31, 139.40, 137.02, 129.12, 128.28, 128.02, 127.00, 126.80, 81.63, 70.38, 60.35, 51.28, 28.20, 21.07; IR (ATR)  $\nu$  3290.0, 3025.3, 2920.1, 1602.7, 1495.1, 1453.3, 1304.9, 1201.0, 1020.2, 908.3  $\text{cm}^{-1}$ ; HRMS (ESI): Exact mass calcd for  $\text{C}_{18}\text{H}_{20}\text{N}$   $[\text{M}+\text{H}]^+$ : 250.1590, found: 250.1594.

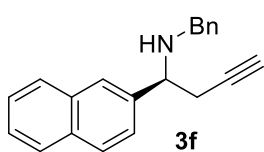


Compound **3d** was prepared in 73% yield according to the general procedure as yellow oil. HPLC analysis (Chiralcel AD-H,  $i$ PrOH/hexane = 1/99, 0.5 mL/min, 230 nm;  $t_r$  (major) = 16.78 min,  $t_r$  (minor) = 18.23 min) gave the isomeric composition of the product: 98% ee;  $[\alpha]_{\text{D}}^{25} = -44.1$  ( $c = 1.0$ ,  $\text{CHCl}_3$ );  $^1\text{H}$  NMR (400 MHz,  $\text{CDCl}_3$ ):  $\delta$  7.42 (s, 1H), 7.35-7.31 (m, 2H), 7.30 (s, 2H), 7.28-7.26 (m, 4H), 3.84 (t,  $J = 6.8$  Hz, 1H), 3.71, 3.58 (AB,  $J = 13.2$  Hz, 2H), 2.58-2.46 (m, 2H), 2.09-1.97 (m, 2H);  $^{13}\text{C}$  NMR (100 MHz,  $\text{CDCl}_3$ ):  $\delta$  144.68, 139.94, 134.37, 129.72, 128.39, 128.01, 127.70, 127.27, 126.98, 125.39, 80.96, 70.82, 60.27, 51.36, 28.06; IR (ATR)  $\nu$  3290.0, 3025.3, 2920.1, 1602.7, 1495.1, 1453.3, 1304.9, 1201.0, 1020.2, 908.3  $\text{cm}^{-1}$ ; HRMS (ESI): Exact mass calcd for  $\text{C}_{17}\text{H}_{17}\text{ClN}$   $[\text{M}+\text{H}]^+$ : 270.1044, found: 270.1039.



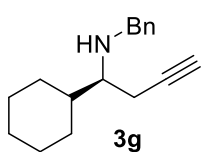
Compound **3e** was prepared in 66% yield according to the general procedure as yellow oil. HPLC analysis (Chiralcel OJ-H,  $i$ PrOH/hexane = 10/90, 1.0 mL/min, 230 nm;  $t_r$  (major) = 7.29 min,  $t_r$  (minor) = 9.29 min) gave the

isomeric composition of the product: 97% ee;  $[\alpha]_{\text{D}}^{25} = -5.7$  ( $c = 0.4$ ,  $\text{CHCl}_3$ );  $^1\text{H NMR}$  (400 MHz,  $\text{CDCl}_3$ ):  $\delta$  7.34-7.28 (m, 4H), 7.24-7.22 (m, 2H), 7.20-7.17 (m, 2H), 7.10 (d,  $J = 7.2$  Hz, 1H), 3.83 (t,  $J = 6.4$  Hz, 1H), 3.71, 3.58 (AB,  $J = 13.2$  Hz, 2H), 2.54-2.52 (m, 2H), 2.36 (s, 3H), 2.02-2.01 (m, 1H), 1.90 (s, br, 1H);  $^{13}\text{C NMR}$  (100 MHz,  $\text{CDCl}_3$ ):  $\delta$  142.31, 140.21, 137.86, 128.23, 128.21, 128.16, 127.93, 127.64, 126.74, 124.12, 81.53, 70.39, 60.59, 51.27, 28.06, 21.37; IR (ATR)  $\nu$  3291.0, 2928.2, 2836.7, 1606.1, 1488.9, 1323.8, 1269.9, 1155.2, 1027.5, 882.7  $\text{cm}^{-1}$ ; HRMS (ESI): Exact mass calcd for  $\text{C}_{18}\text{H}_{20}\text{N}$   $[\text{M}+\text{H}]^+$ : 250.1590, found: 250.1582.



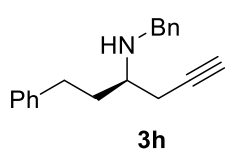
Compound **3f** was prepared in 77% yield according to the general procedure as yellow oil. HPLC analysis (Chiralcel AD-H,  $i\text{PrOH/hexane} = 1/99$ , 0.8 mL/min, 230 nm;  $t_{\text{r}}$  (major) = 11.18 min,  $t_{\text{r}}$  (minor) = 16.02 min) gave the

isomeric composition of the product: 98% ee;  $[\alpha]_{\text{D}}^{25} = -78.0$  ( $c = 1.0$ ,  $\text{CHCl}_3$ );  $^1\text{H NMR}$  (400 MHz,  $\text{CDCl}_3$ ):  $\delta$  7.86-7.82 (m, 4H), 7.56 (d,  $J = 8.4$  Hz, 1H), 7.50-7.44 (m, 2H), 7.34-7.30 (m, 3H), 7.28-7.26 (m, 2H), 4.04 (t,  $J = 6.8$  Hz, 1H), 3.74, 3.61 (AB,  $J = 13.2$  Hz, 2H), 2.64-2.61 (m, 2H), 2.03-1.90 (m, 2H);  $^{13}\text{C NMR}$  (100 MHz,  $\text{CDCl}_3$ ):  $\delta$  140.16, 139.82, 133.29, 133.04, 128.30, 128.27, 128.00, 127.77, 127.60, 126.84, 126.21, 125.95, 125.66, 124.88, 81.44, 70.62, 60.74, 51.32, 28.05; IR (ATR)  $\nu$  3291.1, 3055.8, 3025.2, 2827.3, 1600.7, 1507.1, 1495.0, 1362.1, 1270.4, 1198.9  $\text{cm}^{-1}$ ; HRMS (ESI): Exact mass calcd for  $\text{C}_{21}\text{H}_{20}\text{N}$   $[\text{M}+\text{H}]^+$ : 286.1590, found: 286.1585.



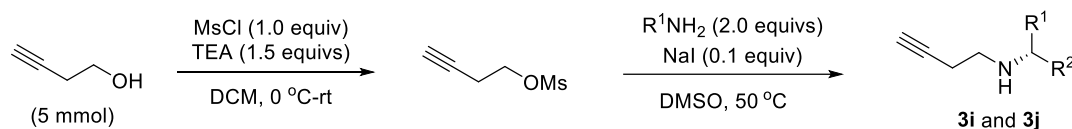
Compound **3g** was prepared in 80% yield according to the general procedure as yellow oil. HPLC analysis (Chiralcel AD-H,  $i\text{PrOH/hexane} = 3/97$ , 0.8 mL/min, 230 nm;  $t_{\text{r}}$  (major) = 5.44 min,  $t_{\text{r}}$  (minor) = 10.84 min) gave the isomeric

composition of the product: 98% ee;  $[\alpha]_{\text{D}}^{25} = +58.2$  ( $c = 0.4$ ,  $\text{CHCl}_3$ );  $^1\text{H NMR}$  (400 MHz,  $\text{CDCl}_3$ ):  $\delta$  7.36-7.35 (m, 2H), 7.33-7.29 (m, 2H), 7.24-7.21 (m, 1H), 3.88, 3.72 (AB,  $J = 13.2$  Hz, 2H), 2.52-2.43 (m, 2H), 2.35-2.30 (m, 1H), 2.03-1.97 (m, 1H), 1.90 (d,  $J = 12.8$  Hz, 1H), 1.73-1.64 (m, 3H), 1.59-1.50 (m, 2H), 1.34-1.11 (m, 4H), 1.06-0.93 (m, 2H);  $^{13}\text{C NMR}$  (100 MHz,  $\text{CDCl}_3$ ):  $\delta$  140.78, 128.26, 128.12, 126.78, 82.23, 69.89, 60.21, 51.49, 40.76, 29.39, 29.29, 26.62, 26.45, 20.49; IR (ATR)  $\nu$  3306.8, 3026.8, 2921.0, 2850.1, 2115.2, 1494.8, 1345.4, 1240.3, 1115.7, 1073.8  $\text{cm}^{-1}$ ; HRMS (ESI): Exact mass calcd for  $\text{C}_{17}\text{H}_{24}\text{N}$   $[\text{M}+\text{H}]^+$ : 242.1903, found: 242.1894.

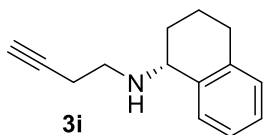


Compound **3h** was prepared in 80% yield according to the general procedure as yellow oil. HPLC analysis (Chiralcel AD-H, *i*PrOH/hexane = 20/80, 0.7 mL/min, 230 nm;  $t_r$  (major) = 6.26 min,  $t_r$  (minor) = 8.75 min) gave the isomeric composition of the product: 98% ee;  $[\alpha]_D^{25} = +21.2$  ( $c = 1.0$ ,  $\text{CHCl}_3$ );  $^1\text{H NMR}$  (400 MHz,  $\text{CDCl}_3$ ):  $\delta$  7.38-7.31 (m, 4H), 7.29-7.27 (m, 3H), 7.19-7.17 (m, 3H), 3.86, 3.74 (AB,  $J = 12.8$  Hz, 2H), 2.82-2.76 (m, 1H), 2.73-2.69 (m, 2H), 2.55-2.48 (m, 1H), 2.40-2.33 (m, 1H), 2.02-2.00 (m, 1H), 1.94-1.80 (m, 2H), 1.58 (s, br, 1H);  $^{13}\text{C NMR}$  (100 MHz,  $\text{CDCl}_3$ ):  $\delta$  142.09, 140.44, 128.34, 128.30, 128.11, 126.88, 125.72, 81.25, 70.42, 54.57, 50.73, 35.70, 32.13, 23.31; IR (ATR)  $\nu$  3292.2, 3025.4, 2922.9, 1602.1, 1494.7, 1453.2, 1116.1, 1071.2, 1028.2, 908.4  $\text{cm}^{-1}$ ; HRMS (ESI): Exact mass calcd for  $\text{C}_{19}\text{H}_{22}\text{N}$   $[\text{M}+\text{H}]^+$ : 264.1747, found: 264.1739.

### 2.3. Preparation of compounds **3i** and **3j**.

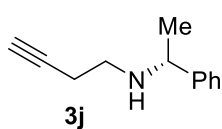


3-Butyn-1-ol (5.0 mmol) was dissolved with in 70 mL dichloromethane, followed by the addition of triethylamine (7.5 mmol) and methanesulfonyl chloride (6.0 mmol) at 0°C. The reaction mixture was allowed to stir at 0°C for 1 hour and then quenched with 1N aqueous HCl (25 mL). The phases were separated and the aqueous layer extracted with dichloromethane (3 x 15 mL). The organic layers were then washed with brine, dried on  $\text{Na}_2\text{SO}_4$  and the solvent was removed under reduced pressure to give the crude mesylate, which was further dissolved in 10 mL DMSO. Then (*R*)-1-aminotetralin and or (*R*)-1-phenylethylamine (10 mmol) and sodium iodide (1.0 mmol) were added, and the reaction mixture was stirred at 50°C for 16 hours. The solution was then cooled to room temperature and diluted with saturated aqueous  $\text{NaHCO}_3$  (50 mL) and ethyl acetate (50 mL). The phases were separated, and the aqueous layer extracted with ethyl acetate (3 x 20 mL), dried with  $\text{Na}_2\text{SO}_4$ , and concentrated under reduced pressure. The residue was subjected to column chromatography using PE/EtOAc (10:1 to 4:1, v/v) as the elution to afford the desired **3i** and **3j**.



Compound **3i** was prepared in 80% yield according to the general procedure as yellow oil. HPLC analysis (Chiralcel AD-H, *i*PrOH/hexane = 5/95, 0.5 mL/min, 230 nm;  $t_r$  (major) = 10.68 min,  $t_r$  (minor) = 9.66 min) gave the

isomeric composition of the product: 98% ee;  $[\alpha]_D^{25} = -9.8$  ( $c = 1.0$ ,  $\text{CHCl}_3$ );  $^1\text{H NMR}$  (400 MHz,  $\text{CDCl}_3$ ):  $\delta$  7.38-7.36 (m, 1H), 7.18-7.14 (m, 2H), 7.09-7.07 (m, 1H), 3.81 (t,  $J = 5.2$  Hz, 1H), 2.95-2.79 (m, 3H), 2.76-2.69 (m, 1H), 2.50-2.36 (m, 2H), 2.03-1.91 (m, 2H), 1.89-1.83 (m, 2H), 1.78-1.69 (m, 1H), 1.58 (s, br, 1H);  $^{13}\text{C NMR}$  (100 MHz,  $\text{CDCl}_3$ ):  $\delta$  138.90, 137.30, 128.96, 128.61, 126.59, 125.65, 82.65, 69.40, 54.86, 45.34, 29.25, 28.28, 19.89, 18.89; IR (ATR)  $\nu$  3292.0, 2930.6, 2855.3, 1488.2, 1320.3, 1271.0, 1197.4, 1035.1, 945.1, 883.2  $\text{cm}^{-1}$ ; HRMS (ESI): Exact mass calcd for  $\text{C}_{14}\text{H}_{18}\text{N}$   $[\text{M}+\text{H}]^+$ : 200.1434, found: 200.1429.



Compound **3j** was prepared in 87% yield according to the general procedure as yellow oil. HPLC analysis (Chiralcel OJ-H, *i*PrOH/hexane = 0.5/99.5, 0.8 mL/min, 230 nm;  $t_r$  (major) = 12.77 min,  $t_r$  (minor) = 16.34 min) gave the isomeric

composition of the product: 97% ee;  $[\alpha]_D^{25} = +43.4$  ( $c = 1.0$ ,  $\text{CHCl}_3$ );  $^1\text{H NMR}$  (400 MHz,  $\text{CDCl}_3$ ):  $\delta$  7.34-7.31 (m, 4H), 7.25-7.22 (m, 1H), 3.80 (q,  $J = 6.4$  Hz, 1H), 2.69-2.57 (m, 2H), 2.36-2.32 (m, 2H), 1.98-1.97 (m, 1H), 1.66 (s, br, 1H), 1.36 (d,  $J = 6.8$  Hz, 3H);  $^{13}\text{C NMR}$  (100 MHz,  $\text{CDCl}_3$ ):  $\delta$  145.37, 128.41, 126.91, 126.54, 82.53, 69.43, 57.68, 45.68, 24.38, 19.59; IR (ATR)  $\nu$  3293.5, 2961.0, 2923.7, 2842.5, 1602.3, 1492.3, 1369.6, 1283.6, 1198.8, 1079.7  $\text{cm}^{-1}$ ; HRMS (ESI): Exact mass calcd for  $\text{C}_{12}\text{H}_{16}\text{N}$   $[\text{M}+\text{H}]^+$ : 174.1277, found: 174.1266.

### 3. Reaction condition optimization

**Table S1.** Carboxylative Cyclization Reaction.

Reaction scheme:  $\text{1a (0.1 mmol)} + \text{CO}_2 \xrightarrow[\text{Solvent (0.2 M), 25 }^\circ\text{C, 12 h}]{[\text{Ag}] (5 \text{ mol\%}), \text{Base (50 mol\%)}} \text{2a}$

Bases screened:

DPG

TPG

TMG

DBU

TBD

Entry	[Ag]	Base	Solvent	Yield (%) <sup>[a]</sup>
1	AgOBz	DPG	DCE	69
2	AgOBz	TPG	DCE	35
3	AgOBz	TMG	DCE	37
4	AgOBz	DBU	DCE	46
5	AgOBz	TBD	DCE	40
6	AgOBz	GBIG	DCE	no reaction
7	AgOAc	DPG	DCE	66
8	AgTFA	DPG	DCE	82
9	AgOTf	DPG	DCE	89
10	AgClO <sub>4</sub> ·H <sub>2</sub> O	DPG	DCE	80
11	AgSbF <sub>6</sub>	DPG	DCE	91
12	AgSbF <sub>6</sub>	DPG	CH <sub>3</sub> CN	37
13	AgSbF <sub>6</sub>	DPG	DMF	6
14	AgSbF <sub>6</sub>	DPG	Acetone	45
15	AgSbF <sub>6</sub>	DPG	Toluene	43
16	--	DPG	DCE	no reaction
17	AgSbF <sub>6</sub>	--	DCE	no reaction

<sup>[a]</sup> Determined by GC-MS with decane as internal standard.

The reaction of *N*-benzyl amine **1a** and CO<sub>2</sub> was undertaken for the evaluation. The reactions were run at 25 °C in 1,2-dichloroethane (DCE), with CO<sub>2</sub> held within a balloon. To our delight, under the catalysis of 5 mol% AgOBz and 50 mol% DPG, the reaction worked well to give the desired 2-oxazinone **2a** in 69% yield (entry 1, Table S1). Next, the performance of other types of organic bases was studied. The use of analogous 1,2,3-triphenylguanidine (TPG) and 1,1,3,3-tetramethylguanidine (TMG) resulted in a greatly diminished 35% and 37% yield for **2a**,

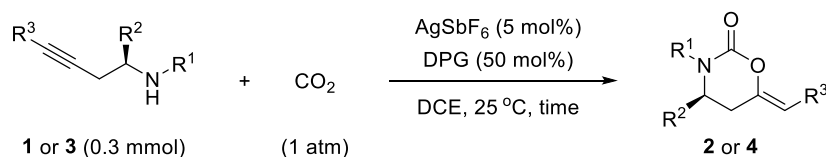
respectively (entries 2-3). The commonly used base, 1,8-diazabicyclo[5.4.0]undec-7-ene (DBU) and 1,5,7-triazabicyclo[4.4.0]dec-5-ene (TBD), showed inferior 46% and 40% yield respectively (entry 4-5). The performance of GBIG<sup>4</sup> (entry 6) was also studied, but no reaction occurred. Considering the metal counterions are of critical importance in impacting the catalytic activity, different silver salts with the combination of DPG was evaluated and AgSbF<sub>6</sub> was found to be the most efficient one, giving **2a** in 91% yield (entries 7-11). Further screening of solvents, including CH<sub>3</sub>CN, DMF, acetone and toluene, failed to improve the result (entries 12-15). In the absence of silver salt or DPG, no reaction occurred at all (entries 16-17).

**Table S2.** The influence of CO<sub>2</sub> pressure and reaction temperature.

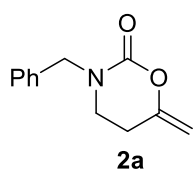
Entry	CO <sub>2</sub> (MPa)	Temp. (°C)	Time (h)	Isolated yield (%)
1	0.1	25	6	47
2	1.0	25	6	59
3	0.1	50	6	62
4	1.0	50	6	91
5	0.1	25	12	91

The influence of CO<sub>2</sub> pressure or reaction temperature was studied. It was found that if the reaction was carried out under higher 50 °C or 10 atm of CO<sub>2</sub>, instead of ambient temperature and pressure, for 6 hours, the reaction yield increased from 47% to 59% and 62% respectively (entries 2-3 vs 1, Table S2). In addition, 91% yield could be obtained under 50 °C and 10 atm of CO<sub>2</sub> simultaneously (entry 4). However, the same result could also be achieved by prolong the reaction time to 12 hours under ambient temperature and pressure (entry 5). These results indicated that, indeed, the higher temperature and pressure could improve the efficiency of carboxylative cyclization to some extent, but the reaction performed under such conditions should be net CO<sub>2</sub> emitter rather than consumer, since the rising of CO<sub>2</sub> pressure and temperature would result indirect production of additional CO<sub>2</sub>. In this context, we prefer to develop the carboxylative cyclization of CO<sub>2</sub> at ambient temperature and pressure, and obviously, under such situation DPG has a distinct advantage than other ones.

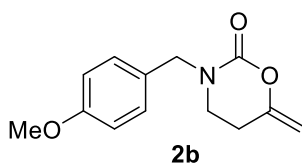
#### 4. General procedure for the carboxylative cyclization reaction.



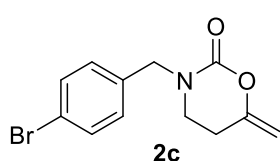
To a 5.0 mL vial were added AgSbF<sub>6</sub> (5.1 mg, 0.015 mmol), DPG (31.7 mg, 0.15 mmol), **1** or **3** (0.3 mmol) and 1.5 mL of DCE, then the resulting solution was stirred under CO<sub>2</sub> atmosphere (1 atm) at 25 °C till full consumption of **1** or **3** by TLC analysis. The residue was directly subjected to column chromatography by using PE/EtOAc (from 4/1 to 2/1, v/v) as the eluent, affording the desired products **2** or **4**. The reaction of homopropargyl amine **21** was performed under 10 atm CO<sub>2</sub> at 50 °C, and IPrAuCl (10 mol%) was used instead of AgSbF<sub>6</sub> (5 mol%).



Product **2a**<sup>5</sup> was obtained in 89% yield as light yellow solid (m.p. 70-72 °C); <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>): δ 7.37-7.33 (m, 2H), 7.31-7.28 (m, 3H), 4.66 (d, *J* = 1.6 Hz, 1H), 4.58 (s, 2H), 4.23 (d, *J* = 1.6 Hz, 1H), 3.21 (t, *J* = 2.0 Hz, 2H), 2.54 (t, *J* = 2.0 Hz, 2H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>): δ 152.58, 151.05, 135.96, 128.62, 127.87, 127.72, 92.67, 52.52, 43.00, 25.97.

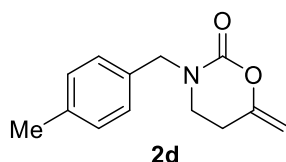


Product **2b**<sup>5</sup> was obtained in 89% yield as light yellow oil; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>): δ 7.24-7.22 (m, 2H), 6.89-6.86 (m, 2H), 4.65 (d, *J* = 1.6 Hz, 1H), 4.51 (s, 2H), 4.22 (d, *J* = 1.6 Hz, 1H), 3.80 (s, 3H), 3.19 (t, *J* = 6.0 Hz, 2H), 2.52 (t, *J* = 6.0 Hz, 2H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>): δ 159.26, 152.68, 151.07, 129.50, 128.14, 114.07, 92.69, 55.25, 52.07, 42.82, 26.12.

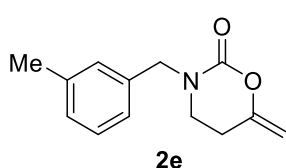


Product **2c**<sup>5</sup> was obtained in 92% yield as light yellow solid (m.p. 65-67 °C); <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>): δ 7.48-7.46 (m, 2H), 7.19-7.17 (m, 2H), 4.68 (d, *J* = 1.6 Hz, 1H), 4.53 (s, 2H), 4.25 (d, *J* = 1.6 Hz, 1H), 3.20 (t, *J* = 6.0 Hz, 2H), 2.55 (t, *J* = 6.0 Hz, 2H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>): δ 152.40, 150.99, 135.11, 131.77, 129.65, 121.72, 92.98, 52.02, 43.17, 25.99.

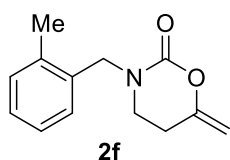




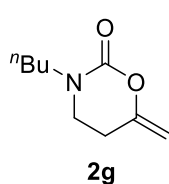
Product **2d** was obtained in 90% yield as light yellow solid (m.p. 77-79 °C);  $^1\text{H}$  NMR (400 MHz,  $\text{CDCl}_3$ ):  $\delta$  7.20-7.18 (m, 2H), 7.16-7.14 (m, 2H), 4.65 (d,  $J = 1.6$  Hz, 1H), 4.54 (s, 2H), 4.22 (d,  $J = 1.6$  Hz, 1H), 3.19 (t,  $J = 6.0$  Hz, 2H), 2.52 (t,  $J = 6.0$  Hz, 2H), 2.34 (s, 3H);  $^{13}\text{C}$  NMR (100 MHz,  $\text{CDCl}_3$ ):  $\delta$  152.67, 151.04, 137.49, 132.96, 129.32, 127.99, 92.58, 52.29, 42.87, 26.05, 21.00; IR (ATR)  $\nu$  2920.0, 1712.3, 1662.3, 1514.7, 1483.4, 1357.0, 1313.4, 1253.7, 1097.3, 999.0  $\text{cm}^{-1}$ ; HRMS (ESI): Exact mass calcd for  $\text{C}_{13}\text{H}_{15}\text{NNaO}_2$   $[\text{M}+\text{Na}]^+$ : 240.0995, found: 240.0989.



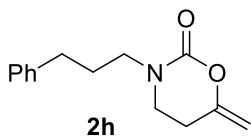
Product **2e** was obtained in 91% yield as light yellow solid (m.p. 71-73 °C);  $^1\text{H}$  NMR (400 MHz,  $\text{CDCl}_3$ ):  $\delta$  7.25-7.21 (m, 1H), 7.12-7.07 (m, 3H), 4.66 (d,  $J = 1.6$  Hz, 1H), 4.54 (s, 2H), 4.23 (d,  $J = 1.6$  Hz, 1H), 3.21 (t,  $J = 6.4$  Hz, 2H), 2.54 (t,  $J = 6.4$  Hz, 2H), 2.34 (s, 3H);  $^{13}\text{C}$  NMR (100 MHz,  $\text{CDCl}_3$ ):  $\delta$  152.66, 151.05, 138.39, 135.92, 128.65, 128.50, 125.00, 92.61, 52.53, 42.97, 26.03, 21.25; IR (ATR)  $\nu$  2919.0, 1711.3, 1607.9, 1483.6, 1354.7, 1298.7, 1254.3, 1107.5, 1053.7, 955.8  $\text{cm}^{-1}$ ; HRMS (ESI): Exact mass calcd for  $\text{C}_{13}\text{H}_{15}\text{NNaO}_2$   $[\text{M}+\text{Na}]^+$ : 240.0995, found: 240.0990.



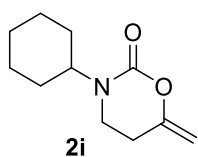
Product **2f** was obtained in 86% yield as light yellow solid (m.p. 80-83 °C);  $^1\text{H}$  NMR (400 MHz,  $\text{CDCl}_3$ ):  $\delta$  7.23-7.18 (m, 3H), 7.17-7.14 (m, 1H), 4.67 (d,  $J = 1.6$  Hz, 1H), 4.62 (s, 2H), 4.24 (d,  $J = 1.6$  Hz, 1H), 3.14 (t,  $J = 6.0$  Hz, 2H), 2.55 (t,  $J = 6.0$  Hz, 2H), 2.31 (s, 3H);  $^{13}\text{C}$  NMR (100 MHz,  $\text{CDCl}_3$ ): 152.56, 150.79, 136.56, 133.43, 130.60, 128.12, 127.78, 126.04, 92.62, 50.41, 42.77, 26.00, 19.00; IR (ATR)  $\nu$  2925.8, 1707.8, 1666.6, 1484.3, 1380.3, 1329.6, 1246.8, 1177.0, 1095.1, 993.1  $\text{cm}^{-1}$ ; HRMS (ESI): Exact mass calcd for  $\text{C}_{13}\text{H}_{15}\text{NNaO}_2$   $[\text{M}+\text{Na}]^+$ : 240.0995, found: 240.0988.



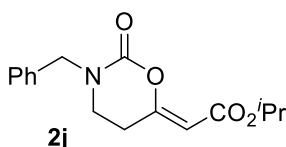
Product **2g**<sup>5</sup> was obtained in 66% yield as light yellow oil;  $^1\text{H}$  NMR (400 MHz,  $\text{CDCl}_3$ ):  $\delta$  4.59 (s, 1H), 4.19 (s, 1H), 3.33 (t,  $J = 7.2$  Hz, 2H), 3.27 (t,  $J = 6.0$  Hz, 2H), 2.56 (t,  $J = 6.4$  Hz, 2H), 1.57-1.50 (m, 2H), 1.35-1.26 (m, 2H), 0.90 (t,  $J = 7.6$  Hz, 3H);  $^{13}\text{C}$  NMR (100 MHz,  $\text{CDCl}_3$ ):  $\delta$  152.78, 150.57, 92.26, 49.37, 43.87, 29.17, 26.20, 19.79, 13.70.



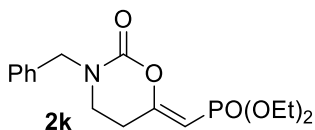
Product **2h** was obtained in 74% yield as light yellow oil;  $^1\text{H}$  NMR (400 MHz,  $\text{CDCl}_3$ ):  $\delta$  7.30-7.26 (m, 2H), 7.22-7.18 (m, 3H), 4.62 (d,  $J = 1.6$  Hz, 1H), 4.22 (d,  $J = 2.0$  Hz, 1H), 3.41 (t,  $J = 7.6$  Hz, 2H), 3.25 (t,  $J = 6.4$  Hz, 2H), 2.65 (t,  $J = 7.6$  Hz, 2H), 2.53 (t,  $J = 6.0$  Hz, 2H), 1.96-1.89 (m, 2H);  $^{13}\text{C}$  NMR (100 MHz,  $\text{CDCl}_3$ ):  $\delta$  152.64, 150.59, 141.13, 128.33, 128.12, 125.89, 92.37, 49.28, 43.87, 32.85, 28.59, 26.07; IR (ATR)  $\nu$  3025.8, 2925.8, 1712.8, 1602.5, 1484.2, 1344.3, 1255.4, 1174.2, 1081.0, 981.4  $\text{cm}^{-1}$ ; HRMS (ESI): Exact mass calcd for  $\text{C}_{14}\text{H}_{17}\text{NNaO}_2$   $[\text{M}+\text{Na}]^+$ : 254.1151, found: 254.1146.



Product **2i** was obtained in 40% yield as light yellow oil;  $^1\text{H}$  NMR (400 MHz,  $\text{CDCl}_3$ ):  $\delta$  4.61 (d,  $J = 1.6$  Hz, 1H), 4.19-4.11 (m, 2H), 3.22 (t,  $J = 6.4$  Hz, 2H), 2.54 (t,  $J = 6.0$  Hz, 2H), 1.82-1.76 (m, 4H), 1.68-1.61 (m, 2H), 1.44-1.34 (m, 4H);  $^{13}\text{C}$  NMR (100 MHz,  $\text{CDCl}_3$ ):  $\delta$  152.60, 150.48, 91.72, 55.73, 38.25, 29.63, 26.51, 25.46, 25.37; IR (ATR)  $\nu$  2927.3, 2854.7, 1707.8, 1658.6, 1481.0, 1373.6, 1347.0, 1254.6, 1092.8, 986.5  $\text{cm}^{-1}$ ; HRMS (ESI): Exact mass calcd for  $\text{C}_{11}\text{H}_{17}\text{NNaO}_2$   $[\text{M}+\text{Na}]^+$ : 218.1151, found: 218.1145.

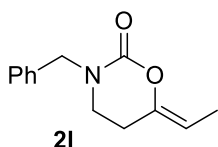


Product **2j** was obtained in 54% yield as light yellow oil;  $^1\text{H}$  NMR (400 MHz,  $\text{CDCl}_3$ ):  $\delta$  7.37-7.33 (m, 2H), 7.32-7.28 (m, 3H), 5.11-5.03 (m, 1H), 5.00 (s, 1H), 4.59 (s, 2H), 3.25 (t,  $J = 6.4$  Hz, 2H), 2.57-2.54 (m, 2H), 1.26 (d,  $J = 6.4$  Hz, 6H);  $^{13}\text{C}$  NMR (100 MHz,  $\text{CDCl}_3$ ):  $\delta$  162.95, 157.65, 149.10, 135.54, 128.76, 128.11, 128.00, 99.20, 67.38, 52.88, 41.89, 30.81, 27.31, 21.77; IR (ATR)  $\nu$  2981.2, 2929.9, 1728.3, 1715.5, 1652.2, 1540.0, 1485.7, 1371.1, 1275.1, 1143.9  $\text{cm}^{-1}$ ; HRMS (ESI): Exact mass calcd for  $\text{C}_{16}\text{H}_{19}\text{NNaO}_4$   $[\text{M}+\text{Na}]^+$ : 312.1206, found: 312.1203. Based on NOE analysis, the Z diastereomer was obtained, for detail see the attached NOE spectrum.



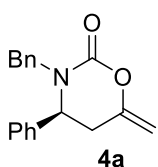
Product **2k** was obtained in 74% yield as light yellow oil;  $^1\text{H}$  NMR (400 MHz,  $\text{CDCl}_3$ ):  $\delta$  7.38-7.31 (m, 3H), 7.30-7.27 (m, 2H), 4.82 (d,  $J = 9.2$  Hz, 1H), 4.57 (s, 2H), 4.24-4.15 (m, 4H), 3.26-3.23 (m, 2H), 2.61-2.58 (m, 2H), 1.36 (t,  $J = 7.2$  Hz, 6H);  $^{13}\text{C}$  NMR (100 MHz,  $\text{CDCl}_3$ ):  $\delta$  159.62 (d,  $J = 4.0$  Hz), 148.90, 135.47, 128.76, 128.02, 95.80, 93.89, 62.04 (d,  $J = 6.0$  Hz), 52.76, 41.85, 27.76 (d,  $J = 15.0$  Hz), 16.24 (d,  $J = 7.0$  Hz);  $^{31}\text{P}$  NMR (122 MHz,  $\text{CDCl}_3$ ):  $\delta$  13.58 (m); IR (ATR)  $\nu$  2981.2, 1727.4, 1651.2,

1482.1, 1391.8, 1349.9, 1238.5, 1185.4, 1095.4, 886.1  $\text{cm}^{-1}$ ; HRMS (ESI): Exact mass calcd for  $\text{C}_{16}\text{H}_{22}\text{NNaO}_3\text{P} [\text{M}+\text{Na}]^+$ : 362.1128, found: 362.1121. Based on NOE analysis, the Z diastereomer was obtained, for detail see the attached NOE spectrum.



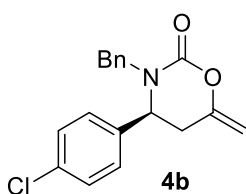
Product **2I** was obtained in 25% yield as light yellow oil;  $^1\text{H}$  NMR (400 MHz,  $\text{CDCl}_3$ ):  $\delta$  7.37-7.33 (m, 2H), 7.31-7.28 (m, 3H), 4.60-4.55 (m, 3H), 3.18 (t,  $J = 6.4$  Hz, 2H), 2.48-2.45 (m, 2H), 1.69-1.66 (m, 3H);  $^{13}\text{C}$  NMR (100 MHz,  $\text{CDCl}_3$ ):  $\delta$  151.62, 145.54, 136.35, 128.73, 128.07, 127.78, 103.01, 52.79, 43.74, 26.54,

9.53; IR (ATR)  $\nu$  2920.4, 2854.0, 1718.0, 1693.4, 1484.0, 1444.2, 1361.0, 1269.8, 1206.3, 1075.9  $\text{cm}^{-1}$ ; HRMS (ESI): Exact mass calcd for  $\text{C}_{13}\text{H}_{15}\text{NNaO}_2 [\text{M}+\text{Na}]^+$ : 240.0995, found: 240.0992. Based on NOE analysis, the Z diastereomer was obtained, for detail see the attached NOE spectrum.



Product **4a** was obtained in 75% yield as light yellow solid (m.p. 65-67  $^\circ\text{C}$ ); HPLC analysis (Chiralcel AD-H,  $i\text{PrOH}$ /hexane = 15/85, 1.0 mL/min, 230 nm;  $t_r$  (major) = 9.95 min,  $t_r$  (minor) = 11.66 min) gave the isomeric composition of the product: 96% ee;  $[\alpha]_{\text{D}}^{25} = +15.8$  ( $c = 1.0$ ,  $\text{CHCl}_3$ );  $^1\text{H}$  NMR (400 MHz,  $\text{CDCl}_3$ ):  $\delta$  7.40-7.35 (m, 3H),

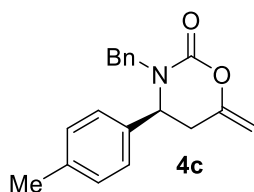
7.33-7.29 (m, 3H), 7.26-7.22 (m, 2H), 7.17-7.15 (m, 2H), 5.30, 3.68 (AB,  $J = 15.2$  Hz, 2H), 4.42 (dd,  $J = 6.4$  Hz, 2.8 Hz, 1H), 4.38 (d,  $J = 262.4$  Hz, 2H), 2.88-2.83 (m, 1H), 2.49 (dd,  $J = 14.4$  Hz, 2.8 Hz, 1H);  $^{13}\text{C}$  NMR (100 MHz,  $\text{CDCl}_3$ ):  $\delta$  151.38, 149.96, 138.58, 136.11, 128.78, 128.66, 128.23, 128.07, 127.76, 126.23, 94.86, 55.94, 50.49, 34.19; IR (ATR)  $\nu$  2925.8, 1694.0, 1495.1, 1444.6, 1367.5, 1281.5, 1108.4, 1031.6, 932.2, 852.8  $\text{cm}^{-1}$ ; HRMS (ESI): Exact mass calcd for  $\text{C}_{18}\text{H}_{17}\text{NNaO}_2 [\text{M}+\text{Na}]^+$ : 302.1151, found: 302.1144.



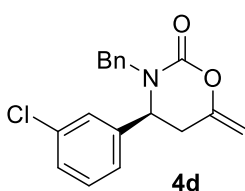
Product **4b** was obtained in 75% yield as white solid (m.p. 77-79  $^\circ\text{C}$ ); HPLC analysis (Chiralcel AD-H,  $i\text{PrOH}$ /hexane = 15/85, 0.8 mL/min, 230 nm;  $t_r$  (major) = 11.27 min,  $t_r$  (minor) = 12.18 min) gave the isomeric composition of the product: 98% ee;  $[\alpha]_{\text{D}}^{25} = -25.9$  ( $c = 1.0$ ,  $\text{CHCl}_3$ );  $^1\text{H}$  NMR (400 MHz,

$\text{CDCl}_3$ ):  $\delta$  7.34-7.29 (m, 5H), 7.22-7.21 (m, 2H), 7.09 (d,  $J = 8.4$  Hz, 2H), 5.27, 3.67 (AB,  $J = 15.2$  Hz, 2H), 4.40 (dd,  $J = 6.0$  Hz, 2.4 Hz, 1H), 4.39 (d,  $J = 260.8$  Hz, 2H), 2.88-2.82 (m, 1H), 2.44 (dd,  $J = 14.0$  Hz, 2.4 Hz, 1H);  $^{13}\text{C}$  NMR (100 MHz,  $\text{CDCl}_3$ ):  $\delta$  151.20, 149.64, 137.20, 135.89, 134.13,

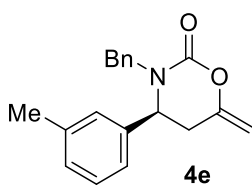
129.03, 128.75, 128.11, 127.91, 127.66, 95.22, 55.47, 50.65, 34.15; IR (ATR)  $\nu$  2920.9, 1712.8, 1661.3, 1490.0, 1425.6, 1358.2, 1299.7, 1269.8, 1147.7, 1029.0  $\text{cm}^{-1}$ ; HRMS (ESI): Exact mass calcd for  $\text{C}_{18}\text{H}_{16}\text{ClNNaO}_2$   $[\text{M}+\text{Na}]^+$ : 336.0762, found: 336.0762.



Product **4c** was obtained in 74% yield as light yellow oil; HPLC analysis (Chiralcel AD-H,  $i$ PrOH/hexane = 5/95, 1.0 mL/min, 230 nm;  $t_r$  (major) = 22.30 min,  $t_r$  (minor) = 29.02 min) gave the isomeric composition of the product: 98% ee;  $[\alpha]_{\text{D}}^{25} = +2.5$  ( $c = 1.0$ ,  $\text{CHCl}_3$ );  $^1\text{H NMR}$  (400 MHz,  $\text{CDCl}_3$ ):  $\delta$  7.35-7.28 (m, 3H), 7.24 (d,  $J = 7.2$  Hz, 2H), 7.18 (d,  $J = 8.0$  Hz, 2H), 7.05 (d,  $J = 7.6$  Hz, 2H), 5.30, 3.66 (AB,  $J = 14.8$  Hz, 2H), 4.38 (dd,  $J = 6.4$  Hz, 2.4 Hz, 1H), 4.38 (d,  $J = 261.2$  Hz, 2H), 2.88-2.82 (m, 1H), 2.47 (dd,  $J = 14.4$  Hz, 2.8 Hz, 1H), 2.36 (s, 3H);  $^{13}\text{C NMR}$  (100 MHz,  $\text{CDCl}_3$ ):  $\delta$  151.46, 150.19, 138.10, 136.28, 135.64, 129.52, 128.70, 128.13, 127.78, 126.24, 94.79, 55.76, 50.44, 34.34, 21.04; IR (ATR)  $\nu$  3028.2, 2916.9, 1713.1, 1663.6, 1514.0, 1495.4, 1427.0, 1359.3, 1261.0, 1074.8  $\text{cm}^{-1}$ ; HRMS (ESI): Exact mass calcd for  $\text{C}_{19}\text{H}_{19}\text{NNaO}_2$   $[\text{M}+\text{Na}]^+$ : 316.1308, found: 316.1301.

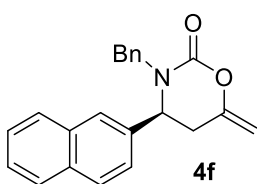


Product **4d** was obtained in 73% yield as white solid (m.p. 85-87  $^{\circ}\text{C}$ ); HPLC analysis (Chiralcel AD-H,  $i$ PrOH/hexane = 15/85, 0.8 mL/min, 230 nm;  $t_r$  (major) = 13.01 min,  $t_r$  (minor) = 15.73 min) gave the isomeric composition of the product: 97% ee;  $[\alpha]_{\text{D}}^{25} = +8.0$  ( $c = 0.1$ ,  $\text{CHCl}_3$ );  $^1\text{H NMR}$  (400 MHz,  $\text{CDCl}_3$ ):  $\delta$  7.37-7.29 (m, 5H), 7.24-7.22 (m, 2H), 7.13 (s, 1H), 7.07-7.03 (m, 1H), 5.31, 3.68 (AB,  $J = 15.2$  Hz, 2H), 4.41 (d,  $J = 264.8$  Hz, 2H), 4.40 (dd,  $J = 6.4$  Hz, 2.4 Hz, 1H), 2.90-2.84 (m, 1H), 2.46 (dd,  $J = 14.4$  Hz, 2.4 Hz, 1H);  $^{13}\text{C NMR}$  (100 MHz,  $\text{CDCl}_3$ ):  $\delta$  151.17, 149.51, 140.82, 135.87, 134.82, 130.15, 128.76, 128.51, 128.12, 127.94, 126.47, 124.38, 95.32, 55.60, 50.76, 34.08; IR (ATR)  $\nu$  2921.0, 1714.8, 1664.9, 1596.5, 1495.0, 1445.2, 1340.5, 1028.4, 971.8, 803.2  $\text{cm}^{-1}$ ; HRMS (ESI): Exact mass calcd for  $\text{C}_{18}\text{H}_{16}\text{ClNNaO}_2$   $[\text{M}+\text{Na}]^+$ : 336.0762, found: 336.0761.



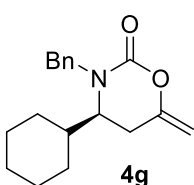
Product **4e** was obtained in 81% yield as light yellow solid (m.p. 90-92 °C); HPLC analysis (Chiralcel AD-H, *i*PrOH/hexane = 10/90, 0.8 mL/min, 230 nm;  $t_r$  (major) = 14.49 min,  $t_r$  (minor) = 17.88 min) gave the isomeric composition of the product: 97% ee;  $[\alpha]_D^{25} = +33.0$  ( $c = 0.5$ , CHCl<sub>3</sub>); <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>):

$\delta$  7.36-7.32 (m, 3H), 7.26-7.23 (m, 3H), 7.14 (d,  $J = 7.6$  Hz, 1H), 6.96-6.95 (m, 2H), 5.32, 3.68 (AB,  $J = 15.2$  Hz, 2H), 4.38 (d,  $J = 262.4$  Hz, 2H), 4.38 (dd,  $J = 6.4$  Hz, 2.8 Hz, 1H), 2.88-2.82 (m, 1H), 2.48 (dd,  $J = 14.0$  Hz, 2.4 Hz, 1H), 2.35 (s, 3H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>):  $\delta$  151.48, 150.11, 138.60, 136.27, 129.03, 128.70, 128.68, 128.11, 127.77, 126.88, 123.38, 94.79, 56.01, 50.56, 34.27, 21.39; IR (ATR)  $\nu$  2950.1, 1709.0, 1692.9, 1657.1, 1453.0, 1435.3, 1104.8, 1076.2, 811.4, 707.9 cm<sup>-1</sup>; HRMS (ESI): Exact mass calcd for C<sub>19</sub>H<sub>19</sub>NNaO<sub>2</sub> [M+Na]<sup>+</sup>: 316.1308, found: 316.1303.



Product **4f** was obtained in 77% yield as light yellow oil; HPLC analysis (Chiralcel AD-H, *i*PrOH/hexane = 5/95, 1.0 mL/min, 230 nm;  $t_r$  (major) = 32.72 min,  $t_r$  (minor) = 38.07 min) gave the isomeric composition of the product: 98% ee;  $[\alpha]_D^{25} = -15.1$  ( $c = 1.0$ , CHCl<sub>3</sub>); <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>):

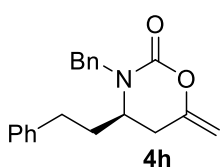
$\delta$  7.88-7.83 (m, 3H), 7.61 (s, 1H), 7.55-7.50 (m, 2H), 7.36-7.29 (m, 3H), 7.27-7.24 (m, 3H), 4.60-4.58 (m, 1H), 5.38, 3.73 (AB,  $J = 15.2$  Hz, 2H), 4.37 (d,  $J = 274.4$  Hz, 2H), 2.94 (dd,  $J = 14.4$  Hz, 6.4 Hz, 1H), 2.62-2.57 (m, 1H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>):  $\delta$  151.56, 150.00, 136.21, 135.97, 133.11, 133.09, 128.95, 128.76, 128.16, 127.91, 127.87, 127.68, 126.61, 126.41, 125.34, 123.89, 95.06, 56.18, 50.67, 34.26; IR (ATR)  $\nu$  3028.1, 2919.3, 1712.3, 1601.0, 1508.4, 1445.8, 1367.2, 1269.9, 1206.1, 1074.9 cm<sup>-1</sup>; HRMS (ESI): Exact mass calcd for C<sub>22</sub>H<sub>19</sub>NNaO<sub>2</sub> [M+Na]<sup>+</sup>: 352.1308, found: 352.1311.



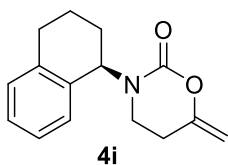
Product **4g** was obtained in 75% yield as light yellow solid (m.p. 67-69 °C); HPLC analysis (Chiralcel AD-H, *i*PrOH/hexane = 5/95, 1.0 mL/min, 205 nm;  $t_r$  (major) = 22.04 min,  $t_r$  (minor) = 29.46 min) gave the isomeric composition of the product: 97% ee;  $[\alpha]_D^{25} = +30.4$  ( $c = 0.5$ , CHCl<sub>3</sub>); <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>):  $\delta$  7.35-7.32

(m, 2H), 7.30-7.27 (m, 3H), 5.34, 3.94 (AB,  $J = 15.2$  Hz, 2H), 4.41 (d,  $J = 194.8$  Hz, 2H), 2.98 (t,  $J = 6.4$  Hz, 1H), 2.56 (d,  $J = 15.2$  Hz, 1H), 2.31 (dd,  $J = 14.8$  Hz, 4.8 Hz, 1H), 1.80 (d,  $J = 10.8$  Hz, 2H), 1.75-1.62 (m, 4H), 1.22-1.16 (m, 2H), 1.13-0.92 (m, 3H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>):  $\delta$

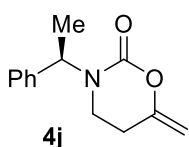
152.23, 151.48, 136.52, 128.67, 127.94, 127.73, 92.75, 56.81, 52.55, 40.72, 30.19, 28.97, 28.90, 26.01, 25.91; IR (ATR)  $\nu$  2934.7, 2852.7, 1704.7, 1659.6, 1424.9, 1339.2, 1208.1, 1135.1, 1073.7, 1028.0  $\text{cm}^{-1}$ ; HRMS (ESI): Exact mass calcd for  $\text{C}_{18}\text{H}_{23}\text{NNaO}_2$   $[\text{M}+\text{Na}]^+$ : 308.1621, found: 308.1622.



Product **4h** was obtained in 71% yield as light yellow oil; HPLC analysis (Chiralcel AD-H,  $^i\text{PrOH}/\text{hexane} = 10/90$ , 0.8 mL/min, 205 nm;  $t_r$  (major) = 21.78 min,  $t_r$  (minor) = 19.08 min) gave the isomeric composition of the product: 98% ee;  $[\alpha]_{\text{D}}^{25} = -13.8$  ( $c = 1.0$ ,  $\text{CHCl}_3$ );  $^1\text{H}$  NMR (400 MHz,  $\text{CDCl}_3$ ):  $\delta$  7.32-7.28 (m, 3H), 7.27-7.21 (m, 3H), 7.14-7.10 (m, 4H), 5.07, 3.94 (AB,  $J = 14.8$  Hz, 2H), 4.49 (d,  $J = 208.8$  Hz, 2H), 3.23-3.18 (m, 1H), 2.71-2.64 (m, 1H), 2.51-2.45 (m, 3H), 1.93-1.86 (m, 2H);  $^{13}\text{C}$  NMR (100 MHz,  $\text{CDCl}_3$ ):  $\delta$  151.06, 150.96, 140.20, 136.42, 128.63, 128.58, 128.17, 128.07, 127.73, 126.28, 94.14, 51.15, 50.79, 32.89, 31.87, 29.91; IR (ATR)  $\nu$  3026.6, 2925.1, 1712.5, 1661.3, 1602.6, 1495.3, 1351.5, 1282.2, 1227.8, 1028.9  $\text{cm}^{-1}$ ; HRMS (ESI): Exact mass calcd for  $\text{C}_{20}\text{H}_{21}\text{NNaO}_2$   $[\text{M}+\text{Na}]^+$ : 330.1464, found: 330.1466.



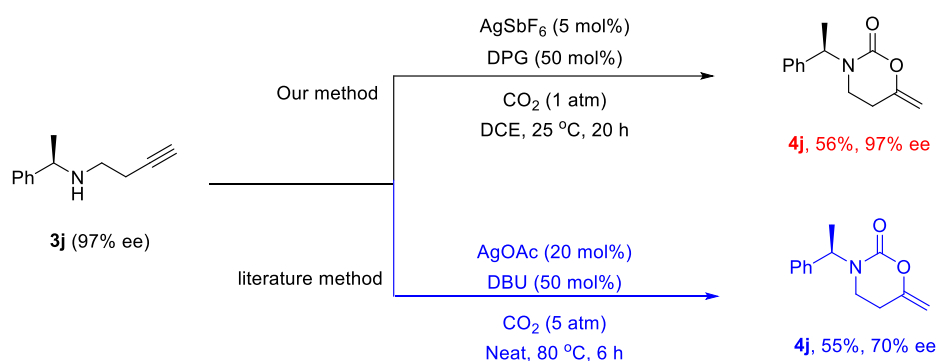
Product **4i** was obtained in 82% yield as light yellow solid (m.p. 89-90  $^{\circ}\text{C}$ ); HPLC analysis (Chiralcel OJ-H,  $^i\text{PrOH}/\text{hexane} = 15/85$ , 0.8 mL/min, 230 nm;  $t_r$  (major) = 12.43 min,  $t_r$  (minor) = 11.62 min) gave the isomeric composition of the product: 98% ee;  $[\alpha]_{\text{D}}^{25} = -34.9$  ( $c = 1.0$ ,  $\text{CHCl}_3$ );  $^1\text{H}$  NMR (400 MHz,  $\text{CDCl}_3$ ):  $\delta$  7.18-7.15 (m, 3H), 7.12-7.10 (m, 1H), 5.68-5.64 (m, 1H), 4.67 (d,  $J = 1.6$  Hz, 1H), 4.22-4.21 (m, 1H), 3.12-3.05 (m, 1H), 2.97-2.91 (m, 1H), 2.83-2.73 (m, 2H), 2.49 (t,  $J = 6.0$  Hz, 2H), 2.16-2.10 (m, 1H), 2.01-1.94 (m, 1H), 1.86-1.72 (m, 2H);  $^{13}\text{C}$  NMR (100 MHz,  $\text{CDCl}_3$ ):  $\delta$  152.84, 151.42, 138.68, 133.81, 129.36, 127.13, 126.86, 126.24, 92.03, 55.48, 39.14, 29.31, 27.16, 26.39, 21.57; IR (ATR)  $\nu$  2942.0, 1715.8, 1652.2, 1475.5, 1361.1, 1337.2, 1257.8, 1230.9, 1083.6, 957.1  $\text{cm}^{-1}$ ; HRMS (ESI): Exact mass calcd for  $\text{C}_{15}\text{H}_{17}\text{NNaO}_2$   $[\text{M}+\text{Na}]^+$ : 266.1151, found: 266.1144.



Product **4j** was obtained in 56% yield as light yellow oil; HPLC analysis (Chiralcel OJ-H,  $^i\text{PrOH}/\text{hexane} = 15/85$ , 0.8 mL/min, 230 nm;  $t_r$  (major) = 15.89 min,  $t_r$  (minor) = 13.95 min) gave the isomeric composition of the product: 97% ee;  $[\alpha]_{\text{D}}^{25} = +68.5$

( $c = 1.0$ ,  $\text{CHCl}_3$ );  $^1\text{H}$  NMR (400 MHz,  $\text{CDCl}_3$ ):  $\delta$  7.30-7.27 (m, 3H), 7.25-7.20 (m, 2H), 5.68 (q,  $J = 7.2$  Hz, 1H), 4.33 (d,  $J = 176.0$  Hz, 2H), 3.08-3.02 (m, 1H), 2.79-2.73 (m, 1H), 2.43-2.37 (m, 1H), 2.34-2.27 (m, 1H), 1.48 (d,  $J = 7.2$  Hz, 3H);  $^{13}\text{C}$  NMR (100 MHz,  $\text{CDCl}_3$ ):  $\delta$  152.60, 150.91, 139.23, 128.52, 127.67, 127.14, 92.06, 53.59, 37.88, 26.20, 15.36; IR (ATR)  $\nu$  2976.4, 1707.4, 1480.7, 1378.8, 1357.6, 1258.2, 1231.5, 1126.3, 1039.2, 983.7  $\text{cm}^{-1}$ ; HRMS (ESI): Exact mass calcd for  $\text{C}_{13}\text{H}_{15}\text{NNaO}_2$   $[\text{M}+\text{Na}]^+$ : 240.0995, found: 240.0987.

Notably, the synthesis of chiral 2-oxazinones from optically active homopropargyl amines is not as trivial as it first appears. If the reaction was run under previous reported condition of high temperature and pressure,<sup>6</sup> the desired chiral 2-oxazinones might be obtained in diminished ee values, as exemplified by the synthesis of **4j** from chiral amine **3j**.

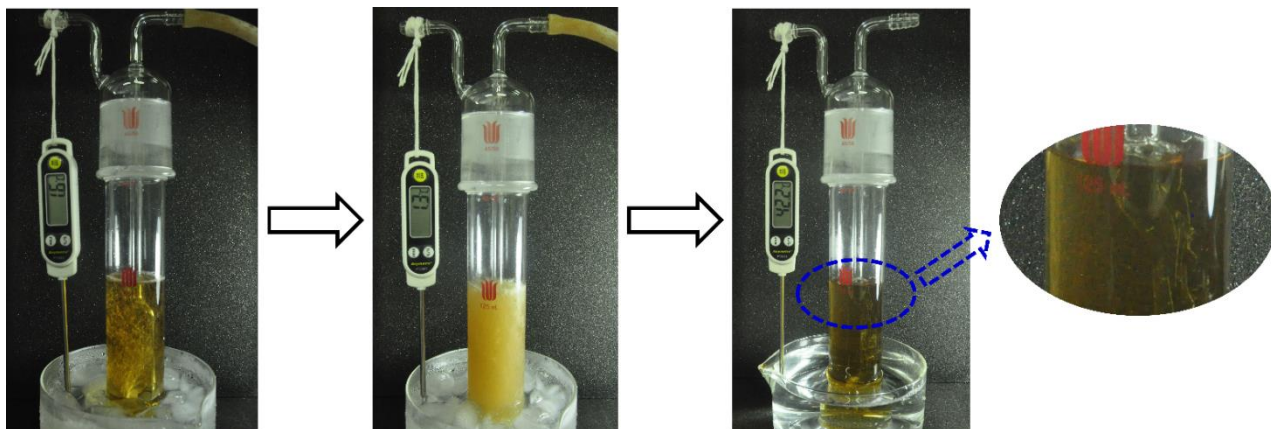


## 5. Mechanistic studies

The superiority that DPG exhibited in the Ag(I)-catalyzed carboxylative cyclization of both *N*-aryl propargylanilines and *N*-alkyl homopropargyl amines is very intriguing. To gain more insight into the role of DPG, a variety of experiments including  $^1\text{H}$  NMR,  $^{13}\text{C}$  NMR, HRMS as well as X-ray analysis were conducted.

### 5.1. The study of interaction between DPG and $\text{CO}_2$ .

Initially, the detail for the trapping and releasing of  $\text{CO}_2$  by DPG was studied. As far as we know, the corresponding reaction between DPG and  $\text{CO}_2$  has never been documented.<sup>4</sup> A general procedure for the capture of  $\text{CO}_2$  was as follows: DPG (633 mg, 3 mmol) was stirred in DCE (10 mL) under a continuous stream of  $\text{CO}_2$  (15 mL/min) for 2 h at  $0^\circ\text{C}$ . Then, deposited solid was quickly filtered off and washed with cold DCE ( $3\times 5$  mL) to give the complex as a white powder (600 mg, 78%) with high purity. Notably, the carboxylation process was reversible. As shown in Figure S1, although the precipitate formed in  $0^\circ\text{C}$  could be isolated via a quick filtration, it would be gradually disappeared with the release of  $\text{CO}_2$  bubbles if raising the solution temperature above  $25^\circ\text{C}$ .

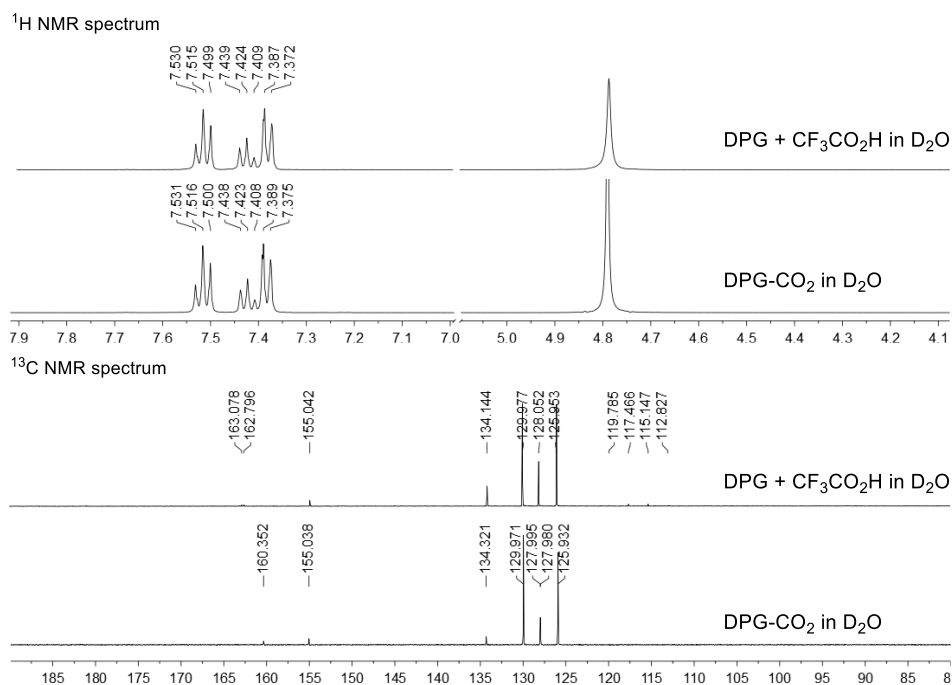


**Figure S1.** The capture and release of  $\text{CO}_2$ . (Note, in order to give a clearer phenomenon, the  $\text{CO}_2$  release process was performed under around  $40^\circ\text{C}$ .)

Then, the structure of the thus obtained complex was analyzed. Initially, NMR studies were performed, but we tried in vain to characterize its structure in common organic deuterated solvents, such as  $\text{CD}_2\text{Cl}_2$ ,  $\text{CDCl}_3$ ,  $\text{CD}_3\text{CN}$ ,  $\text{DMSO-}d_6$ ,  $\text{THF-}d_8$ , and only spectra assigned to the DPG structure was obtained due to the release of  $\text{CO}_2$ . Fortunately, in  $\text{D}_2\text{O}$  the complex could be successfully



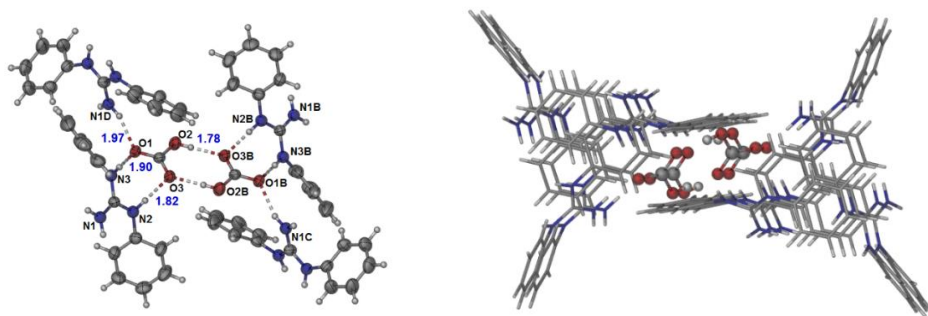
characterized, with the data shown below:  $^1\text{H}$  NMR (500 MHz,  $\text{D}_2\text{O}$ ):  $\delta$  7.53-7.50 (m, 4H), 7.44-7.40 (m, 2H), 7.39-7.37 (m, 4H);  $^{13}\text{C}$  NMR (125 MHz,  $\text{D}_2\text{O}$ ):  $\delta$  160.35, 155.04, 134.32, 129.97, 128.00, 127.98, 125.93; IR (ATR)  $\nu$  1645, 1582, 1544, 1495, 1385, 1242, 750, 689  $\text{cm}^{-1}$ .



**Figure S2.** NMR spectrum comparison.

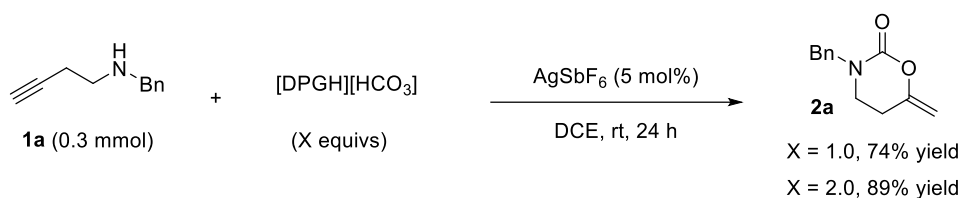
Then, the NMR data of the thus obtained complex in  $\text{D}_2\text{O}$  with that of protonated DPG,  $[\text{DPGH}][\text{CF}_3\text{CO}_2]$ , was compared with the results shown in Figure S2. Obviously, the  $^1\text{H}$  and  $^{13}\text{C}$  NMR data were consistent with the DPG portion of the complex being simply protonated DPG. And the new peak at 160.35 ppm in the  $^{13}\text{C}$  NMR spectrum of the complex was consistent with bicarbonate anion. These results revealed that a bicarbonate salt  $[\text{DPGH}][\text{HCO}_3^-]$  might be formed.

In order to figure out the exact structure of the complex,  $\text{CO}_2$  was diffused into the solvent of DPG in THF/ $\text{Et}_2\text{O}$  (1/1, v/v) under  $-20\text{ }^\circ\text{C}$ , and finally a single crystal of the bicarbonate adduct  $[\text{DPGH}][\text{HCO}_3^-]$  was obtained (CCDC-1907983). X-ray crystallography shows that a centrosymmetric dimer was formed by the “anti-electrostatic” hydrogen-bonding between oxygen atoms of the bicarbonate anion  $[\text{HCO}_3^-]$ , with  $\text{H}\cdots\text{O}$  contact distances of 1.78 Å. In each monomer, the bicarbonate anion associated with the cation  $[\text{DPGH}]^+$  through three hydrogen bonds between the oxygen and nitrogen atoms, with  $\text{N-H}\cdots\text{O}$  contact distances of 1.82, 1.90 and 1.97 Å (Figure S3, left). In addition, the cationic stacks flank the anionic cluster in a close-packed arrangement (right).



**Figure S3.** The X-Ray crystal structure of bicarbonate salt [DPGH][HCO<sub>3</sub>].

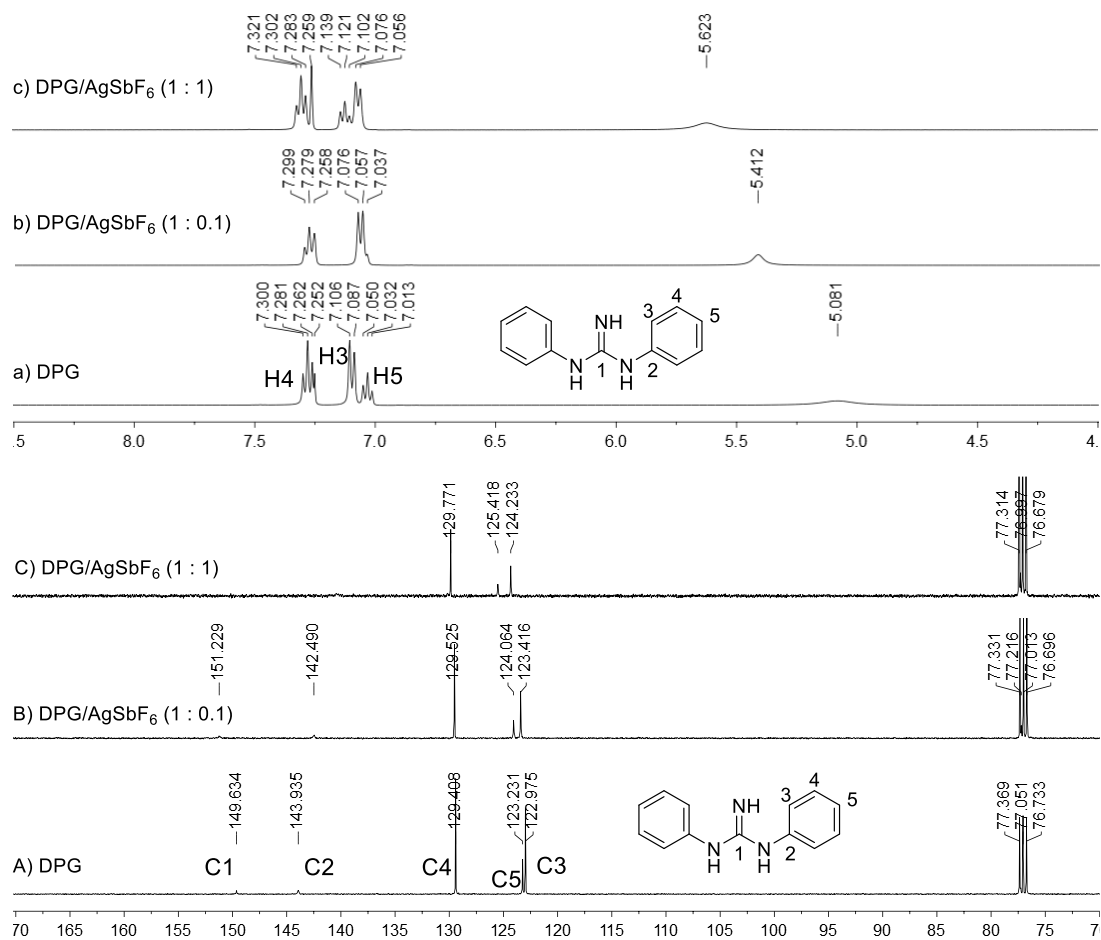
Then the reaction of homopropargyl amine with the bicarbonate adduct was performed. The treatment of homopropargyl amine **1a** with bicarbonate salt [DPGH][HCO<sub>3</sub>] in the presence of 5 mol% of AgSbF<sub>6</sub> under N<sub>2</sub> atmosphere at room temperature gave the desired 2-oxazinone **2a** in 74% and 89% yield respectively, by using 1.0 or 2.0 equivalent of bicarbonate salt. These results further demonstrated that [DPGH][HCO<sub>3</sub>] could release CO<sub>2</sub> effectively during the reaction.



## 5.2 The study of interaction between DPG and AgSbF<sub>6</sub>

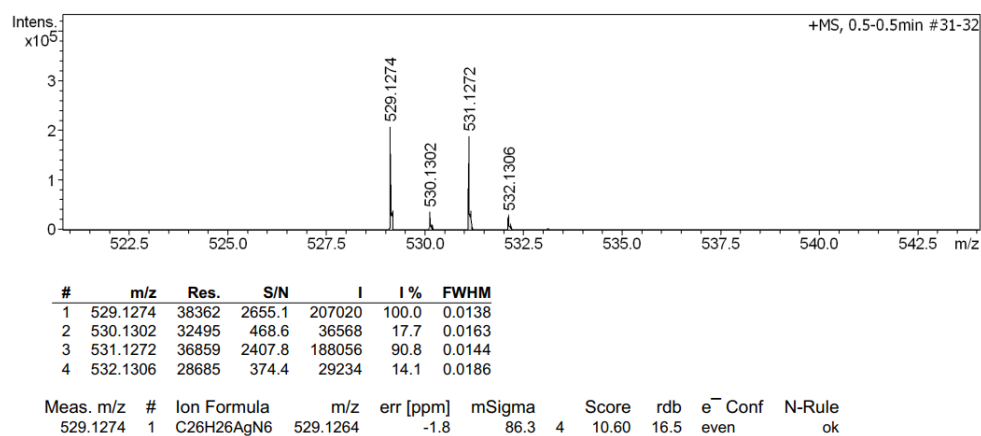
At first the interaction of DPG with AgSbF<sub>6</sub> was studied by NMR analysis, which was conducted in air using CDCl<sub>3</sub> as the solvent. The general procedure was as follows: to a NMR tube were added the DPG (0.1 mmol) and CDCl<sub>3</sub> (0.5 mL), followed by the addition of AgSbF<sub>6</sub> (10 mol% to 100 mol%). Then the tube was shaken vigorously and quickly subjected to NMR analysis at 25 °C.

Obvious changes were immediately observed when AgSbF<sub>6</sub> was added to the solution of DPG in CDCl<sub>3</sub>. <sup>1</sup>H NMR showed that, with the amount of AgSbF<sub>6</sub> increased from 0.1 equiv to 1.0 equiv, the characteristic peaks corresponding to the proton at C5 position of DPG, changed gradually from 7.03 ppm to 7.06 and 7.12 ppm. Meanwhile, the signal of proton on the nitrogen atom changed from 5.08 ppm to 5.41 and 5.62 ppm gradually. Obvious changes could also be observed for the <sup>13</sup>C NMR analysis (Figure S4). When 0.1 equiv of AgSbF<sub>6</sub> was added, the characteristic peaks of C1, C2 and C5 changed from 149.63, 143.94 and 123.23 ppm to 151.23, 142.49 and 124.06 ppm respectively. Further increase the amount of AgSbF<sub>6</sub> to 1.0 equiv, the characteristic peak of C5 shifted to 125.42 ppm and the signals of C1 and C2 might be too weak to be detected. These observations indicated that DPG might coordinate to AgSbF<sub>6</sub>.



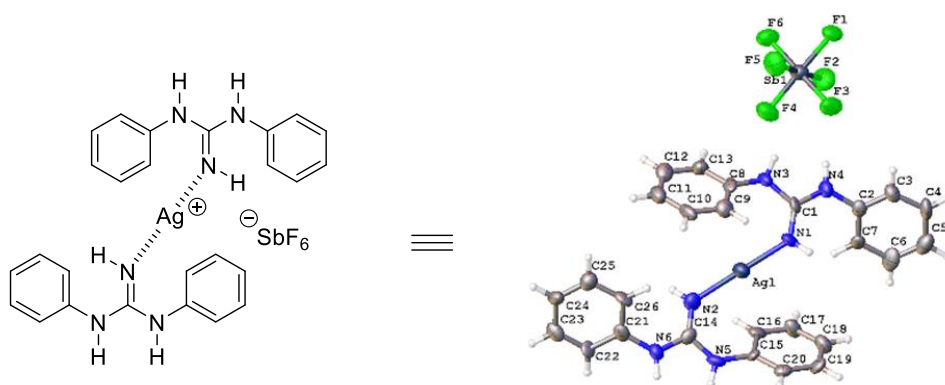
**Figure S4.**  $^1\text{H}$  NMR and  $^{13}\text{C}$  NMR spectra of DPG with 0.1 and 1.0 equiv of  $\text{AgSbF}_6$ .

To get more information about the possible binding interaction, HRMS analysis of the DPG-Ag(I) complex with different molecular ratio ( $\text{AgSbF}_6$  : DPG = 1:1, 1:2 and 1:10) was conducted respectively, and in all cases a signal at  $m/z$  529.1274 was observed as shown in Figure S5, consistent with the 1/2 complex cation,  $[(\text{DPG})_2 + \text{Ag}]^+$ .



**Figure S5.** HRMS analysis of the DPG-Ag(I) complex.

Fortunately, we obtained a single crystal of the complex derived from DPG and  $\text{AgSbF}_6$  upon crystallization of the 1/2 mixture of  $\text{AgSbF}_6$  and DPG from  $\text{CD}_2\text{Cl}_2$  (CCDC-1894496). X-ray diffraction study revealed that the DPG served as neutral monodentate ligand and bound to the silver center via a head-to-head fashion (Figure S6). These results in combination with HRMS analysis, further cast light on the coordination fashion of DPG to  $\text{AgSbF}_6$ .



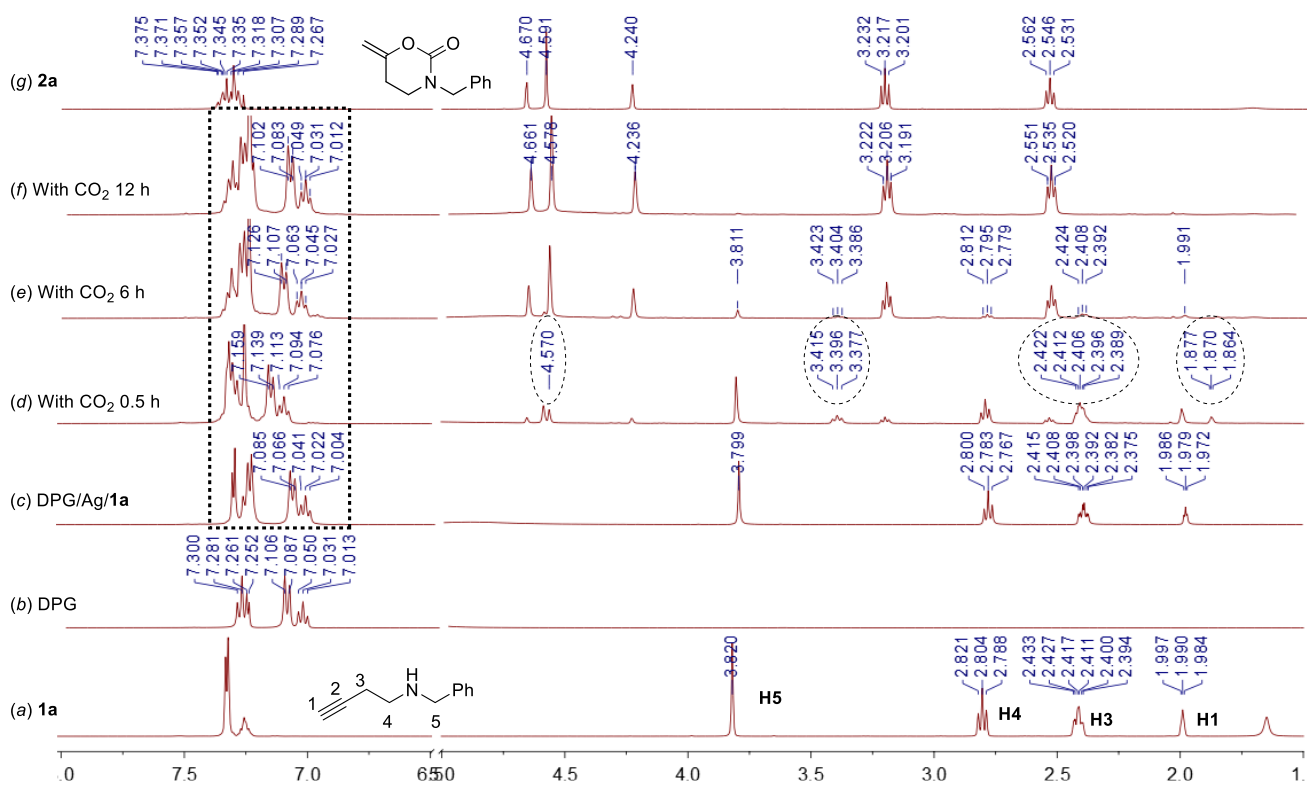
**Figure S6.** the single crystal of DPG-Ag(I) complex.

### 5.3. NMR study of the reaction process.

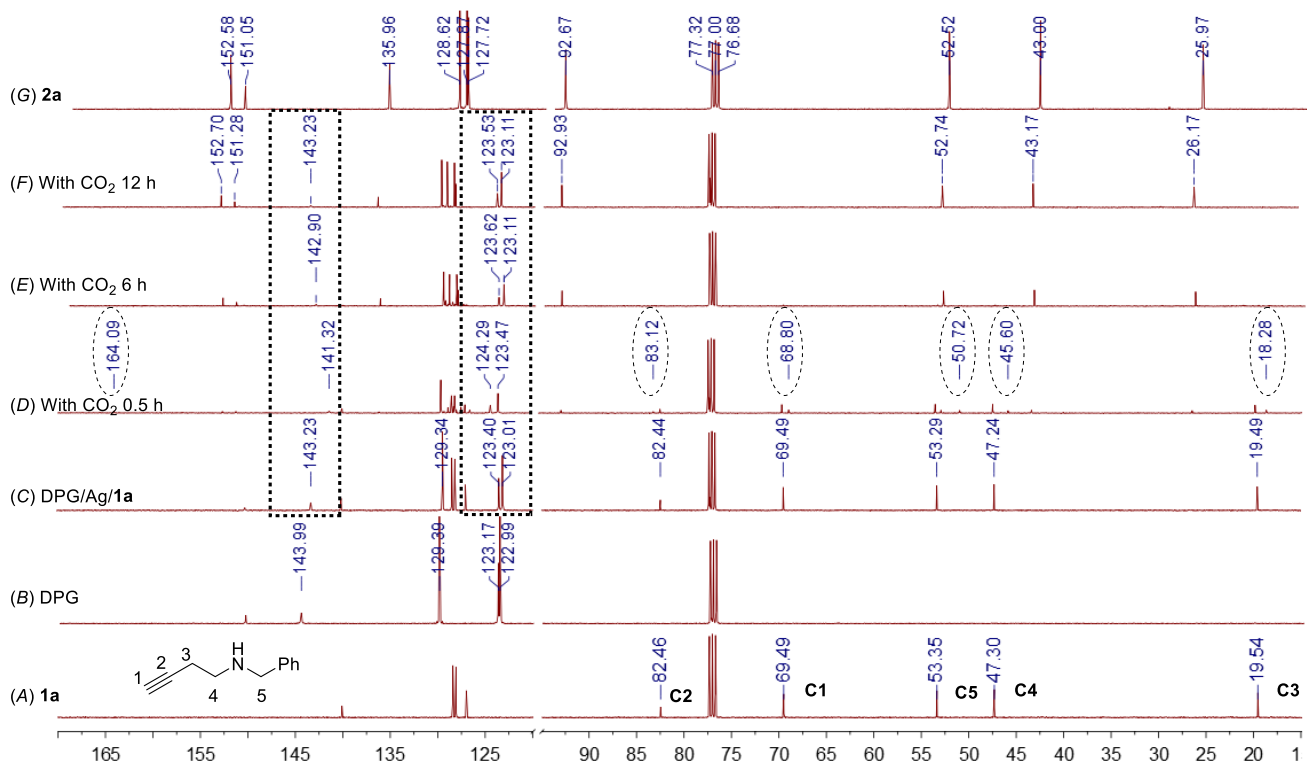
To get more information about the reaction course and to understand the role of DPG during the reaction,  $^1\text{H}$  and  $^{13}\text{C}$  NMR studies of the reaction process were carried out based on the reaction of homopropargyl amine **1a** under 1 atm of  $\text{CO}_2$  at  $25\text{ }^\circ\text{C}$  in  $\text{CDCl}_3$ , in the presence of 10 mol%  $\text{AgSbF}_6$  and 100 mol% DPG.

As shown in Figure S7, when **1a** was added to the mixture of  $\text{AgSbF}_6$  and DPG, the characteristic peaks assigned to H1, H3, H4 and H5 of **1a** upfield shifted from 1.99, 2.41, 2.80, 3.82 ppm to 1.98, 2.39, 2.78 and 3.80 ppm respectively (c vs a). Meanwhile,  $^{13}\text{C}$  NMR spectrum revealed the signals assigned to C3, C4 and C5 shifted from 19.54, 47.30 and 53.35 ppm to 19.49, 47.24 and 53.29 ppm respectively. A slightly upfield shift of the signals assigned to C1 and C2 of the C-C triple bond was also observed (C vs A). These observations indicated that both the alkyne and amine moiety of **1a** might interact with the DPG-Ag(I) complex, which was helpful for suppressing the side intramolecular hydroamination reaction.<sup>7</sup>

1. <sup>1</sup>H NMR analysis



2. <sup>13</sup>C NMR analysis

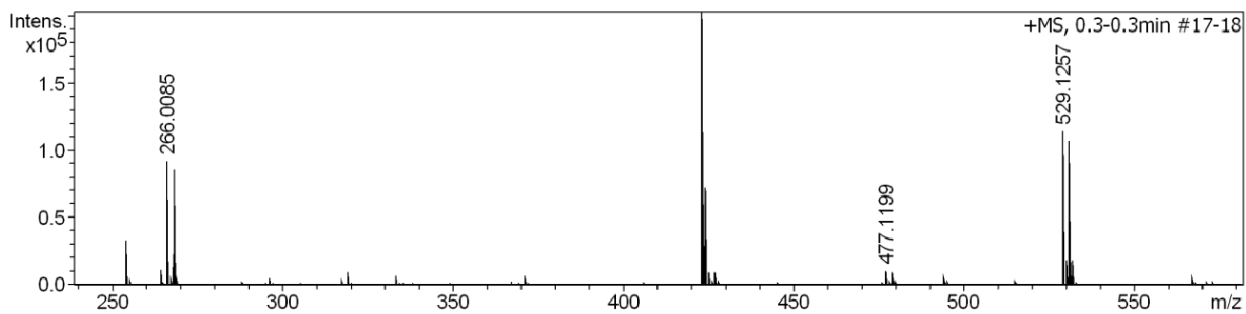


**Figure S7.** <sup>1</sup>H and <sup>13</sup>C NMR study in CDCl<sub>3</sub>. (a, A) **1a** (0.1 mmol). (b, B) DPG (0.1 mmol). (c, C) AgSbF<sub>6</sub> (0.01 mmol), DPG (0.1 mmol) and **1a** (0.1 mmol). (d-f, D-F) Under CO<sub>2</sub> atmosphere 0.5, 6.0 and 12 h respectively. (g, G) **2a** (0.1 mmol).

Subsequently, the above reaction mixture was subjected to CO<sub>2</sub> atmosphere with the reaction process monitored over time and notable changes were observed (Figure S7, d-f & D-F). Within half an hour, apart from the detection of 2-oxazinone **2a**, some new signals appeared at both <sup>1</sup>H NMR (δ = 4.57, 3.40, 2.41 and 1.87 ppm) and <sup>13</sup>C NMR (δ = 164.09, 83.12, 68.80, 50.72, 45.60 and 18.28 ppm) spectrum (d vs c; D vs C, outside the dashed boxes part), which might be attributed to the carbamate formed via the reaction of **1a** with CO<sub>2</sub>. After 6 hours, the signals attribute to **1a** and carbamic intermediate became weak gradually and the full conversion of **1a** to **2a** was observed within 12 hours (e-f; E-F). Notably, during the reaction course, the character peaks belong to DPG also shifted distinctly (the dashed boxes part). In the first half an hour, the characteristic signals of <sup>1</sup>H NMR shifted from 7.07 and 7.02 ppm to 7.15 and 7.09 ppm respectively, meanwhile the signals of <sup>13</sup>C NMR also shifted from 143.23, 123.40 and 123.01 ppm to 141.32, 124.29 and 123.47 ppm obviously (d vs c; D vs C). Then, as the reaction proceeded, the character peaks of DPG shifted back to its original state gradually (d-f vs c; D-F vs C), which indicated that there should be interactions between DPG and the carbamate intermediate.

#### 5.4. HRMS analysis of the complex derived from DPG, AgSbF<sub>6</sub> and **1a**.

In order to get more information of the interactions among AgSbF<sub>6</sub>, DPG and homopropargyl amine **1a**, the HRMS analysis was conducted (Figure S8). When a CH<sub>2</sub>Cl<sub>2</sub> solution of 1:10:10 mixture of AgSbF<sub>6</sub>, DPG and **1a** was subjected to the HRMS analysis, the signals of [**1a**+Ag]<sup>+</sup> at 266.0087, [(DPG)<sub>2</sub>+Ag]<sup>+</sup> at m/z 529.1257, as well as [DPG+**1a**+Ag]<sup>+</sup> at m/z 477.1199 could be detected respectively. This information further confirmed the interactions between AgSbF<sub>6</sub> with DPG and **1a**, and also suggested the formation of 1/1/1 complex of AgSbF<sub>6</sub>, DPG and **1a**.



Meas. m/z	#	Ion Formula	m/z	err [ppm]	mSigma	Score	rdb	e <sup>-</sup> Conf	N-Rule	
266.0085	1	<b>C11H13AgN</b>	<b>266.0093</b>	<b>3.2</b>	<b>62.7</b>	<b>1</b>	<b>100.00</b>	<b>5.5</b>	<b>even</b>	<b>ok</b>
	1	C9H14AgNNa	266.0069	-5.9	49.9	1	100.00	2.5	even	ok
477.1199	1	C9H22AgN16O	477.1208	1.8	100.0	1	100.00	6.5	even	ok
	2	C23H30AgO4	477.1190	-2.1	146.8	2	6.79	8.5	even	ok
	3	<b>C24H26AgN4</b>	<b>477.1203</b>	<b>0.7</b>	<b>174.1</b>	<b>3</b>	<b>1.41</b>	<b>13.5</b>	<b>even</b>	<b>ok</b>
	1	C11H27AgN10NaO3	477.1211	2.4	101.4	1	100.00	2.5	even	ok
529.1257	1	C10H26AgN14O5	529.1256	-0.2	12.2	1	100.00	4.5	even	ok
	2	C11H22AgN18O	529.1269	2.3	18.9	2	48.93	9.5	even	ok
	3	C25H30AgN2O4	529.1251	-1.1	73.1	3	14.17	11.5	even	ok
	4	<b>C26H26AgN6</b>	<b>529.1264</b>	<b>1.4</b>	<b>84.9</b>	<b>4</b>	<b>8.24</b>	<b>16.5</b>	<b>even</b>	<b>ok</b>
	1	C9H23AgN18NaO	529.1245	-2.2	8.0	1	68.52	6.5	even	ok
	2	C12H31AgN8NaO7	529.1259	0.3	14.9	2	100.00	0.5	even	ok
	3	C13H27AgN12NaO3	529.1272	2.9	20.9	3	42.57	5.5	even	ok
	4	C24H27AgN6Na	529.1240	-3.2	79.1	4	6.45	13.5	even	ok
	5	C28H31AgNaO2	529.1267	1.9	86.5	5	7.19	12.5	even	ok

**Figure S8.** HRMS analysis.

## 6. Single-crystal X-ray analysis of bicarbonate salt [DPGH][HCO<sub>3</sub>].

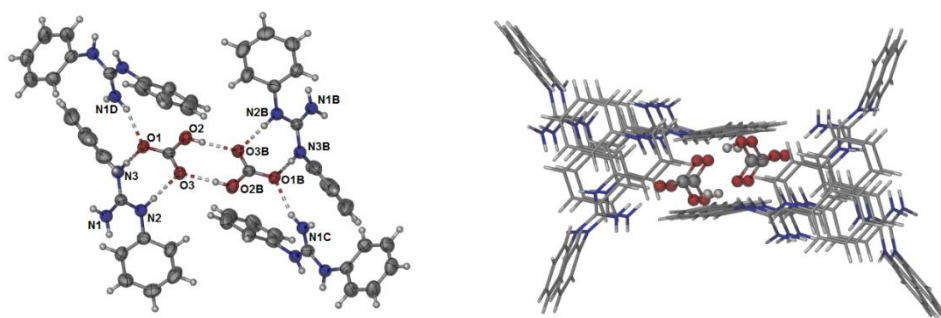


Table S3. Crystal data and structure refinement for CCDC-1907983.

Identification code	CCDC-1907983	
Empirical formula	C <sub>14</sub> H <sub>15</sub> N <sub>3</sub> O <sub>3</sub>	
Formula weight	273.29	
Temperature	193(2) K	
Wavelength	0.71073 Å	
Crystal system	Trigonal	
Space group	R -3 :H	
Unit cell dimensions	a = 34.5947(14) Å	a = 90°.
	b = 34.5947(14) Å	b = 90°.
	c = 10.7422(5) Å	g = 120°.
Volume	11133.8(10) Å <sup>3</sup>	
Z	18	
Density (calculated)	0.734 Mg/m <sup>3</sup>	
Absorption coefficient	0.053 mm <sup>-1</sup>	
F(000)	2592	
Crystal size	0.160 x 0.140 x 0.110 mm <sup>3</sup>	
Theta range for data collection	2.014 to 25.000°.	
Index ranges	-41<=h<=31, -32<=k<=41, -12<=l<=12	
Reflections collected	12749	
Independent reflections	3789 [R(int) = 0.0794]	
Completeness to theta = 25.242°	84.4 %	
Absorption correction	Semi-empirical from equivalents	
Max. and min. transmission	0.7456 and 0.5782	
Refinement method	Full-matrix least-squares on F <sup>2</sup>	
Data / restraints / parameters	3789 / 0 / 183	
Goodness-of-fit on F <sup>2</sup>	1.015	
Final R indices [I>2sigma(I)]	R1 = 0.0784, wR2 = 0.1955	
R indices (all data)	R1 = 0.1169, wR2 = 0.2122	
Extinction coefficient	0.0017(4)	
Largest diff. peak and hole	0.198 and -0.167 e.Å <sup>-3</sup>	



Table S4. Atomic coordinates ( $\times 10^4$ ) and equivalent isotropic displacement parameters ( $\text{\AA}^2 \times 10^3$ ) for d8v19290.  $U(\text{eq})$  is defined as one third of the trace of the orthogonalized  $U_{ij}$  tensor.

	x	y	z	$U(\text{eq})$
O(1)	4446(1)	4055(1)	1381(2)	53(1)
O(2)	4998(1)	4464(1)	96(2)	59(1)
O(3)	4650(1)	4768(1)	1071(2)	58(1)
N(1)	3482(1)	4340(1)	3996(2)	59(1)
N(2)	4197(1)	4694(1)	3190(2)	54(1)
N(3)	3691(1)	4013(1)	2432(2)	51(1)
C(1)	3781(1)	4346(1)	3233(2)	48(1)
C(2)	4424(1)	5020(1)	4111(3)	51(1)
C(3)	4705(1)	5458(1)	3737(3)	67(1)
C(4)	4964(1)	5779(1)	4575(4)	77(1)
C(5)	4943(1)	5673(1)	5818(3)	72(1)
C(6)	4666(1)	5245(1)	6208(3)	69(1)
C(7)	4406(1)	4917(1)	5354(3)	59(1)
C(8)	3270(1)	3645(1)	2131(2)	53(1)
C(9)	3237(1)	3224(1)	2087(3)	68(1)
C(10)	2845(2)	2867(1)	1712(3)	85(1)
C(11)	2481(1)	2903(1)	1409(3)	83(1)
C(12)	2505(1)	3314(2)	1492(3)	82(1)
C(13)	2906(1)	3690(1)	1828(3)	64(1)
C(14)	4682(1)	4428(1)	879(2)	47(1)

Table S5. Bond lengths [ $\text{\AA}$ ] and angles [ $^\circ$ ] for d8v19290.

O(1)-C(14)	1.252(3)
O(2)-C(14)	1.335(3)
O(2)-H(2A)	0.8400
O(3)-C(14)	1.252(3)
N(1)-C(1)	1.312(4)
N(1)-H(1A)	0.8800
N(1)-H(1B)	0.8800
N(2)-C(1)	1.338(4)
N(2)-C(2)	1.408(4)
N(2)-H(2)	0.8800
N(3)-C(1)	1.346(3)
N(3)-C(8)	1.411(4)
N(3)-H(3)	0.8800
C(2)-C(7)	1.375(4)
C(2)-C(3)	1.389(4)
C(3)-C(4)	1.361(5)

C(3)-H(3A)	0.9500
C(4)-C(5)	1.376(5)
C(4)-H(4)	0.9500
C(5)-C(6)	1.367(5)
C(5)-H(5)	0.9500
C(6)-C(7)	1.384(4)
C(6)-H(6)	0.9500
C(7)-H(7)	0.9500
C(8)-C(13)	1.383(5)
C(8)-C(9)	1.404(5)
C(9)-C(10)	1.360(5)
C(9)-H(9)	0.9500
C(10)-C(11)	1.365(7)
C(10)-H(10)	0.9500
C(11)-C(12)	1.385(6)
C(11)-H(11)	0.9500
C(12)-C(13)	1.394(5)
C(12)-H(12)	0.9500
C(13)-H(13)	0.9500
C(14)-O(2)-H(2A)	109.5
C(1)-N(1)-H(1A)	120.0
C(1)-N(1)-H(1B)	120.0
H(1A)-N(1)-H(1B)	120.0
C(1)-N(2)-C(2)	127.9(2)
C(1)-N(2)-H(2)	116.1
C(2)-N(2)-H(2)	116.1
C(1)-N(3)-C(8)	127.7(2)
C(1)-N(3)-H(3)	116.2
C(8)-N(3)-H(3)	116.2
N(1)-C(1)-N(2)	121.5(2)
N(1)-C(1)-N(3)	123.0(3)
N(2)-C(1)-N(3)	115.5(2)
C(7)-C(2)-C(3)	118.6(3)
C(7)-C(2)-N(2)	122.8(3)
C(3)-C(2)-N(2)	118.4(2)
C(4)-C(3)-C(2)	121.0(3)
C(4)-C(3)-H(3A)	119.5
C(2)-C(3)-H(3A)	119.5
C(3)-C(4)-C(5)	119.9(3)
C(3)-C(4)-H(4)	120.0
C(5)-C(4)-H(4)	120.0
C(6)-C(5)-C(4)	120.1(3)
C(6)-C(5)-H(5)	120.0
C(4)-C(5)-H(5)	120.0
C(5)-C(6)-C(7)	120.0(3)

C(5)-C(6)-H(6)	120.0
C(7)-C(6)-H(6)	120.0
C(2)-C(7)-C(6)	120.4(3)
C(2)-C(7)-H(7)	119.8
C(6)-C(7)-H(7)	119.8
C(13)-C(8)-C(9)	120.2(3)
C(13)-C(8)-N(3)	122.6(3)
C(9)-C(8)-N(3)	117.1(3)
C(10)-C(9)-C(8)	118.6(4)
C(10)-C(9)-H(9)	120.7
C(8)-C(9)-H(9)	120.7
C(9)-C(10)-C(11)	122.3(4)
C(9)-C(10)-H(10)	118.8
C(11)-C(10)-H(10)	118.8
C(10)-C(11)-C(12)	119.6(3)
C(10)-C(11)-H(11)	120.2
C(12)-C(11)-H(11)	120.2
C(11)-C(12)-C(13)	119.7(4)
C(11)-C(12)-H(12)	120.2
C(13)-C(12)-H(12)	120.2
C(8)-C(13)-C(12)	119.5(4)
C(8)-C(13)-H(13)	120.3
C(12)-C(13)-H(13)	120.3
O(1)-C(14)-O(3)	125.2(2)
O(1)-C(14)-O(2)	116.8(2)
O(3)-C(14)-O(2)	117.9(2)

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Symmetry transformations used to generate equivalent atoms:

Table S6. Anisotropic displacement parameters ( $\text{\AA}^2 \times 10^3$ ) for d8v19290. The anisotropic displacement factor exponent takes the form:  $-2p^2[h^2 a^*2U^{11} + \dots + 2 h k a^* b^* U^{12}]$

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	U11	U22	U33	U23	U13	U12
O(1)	56(1)	38(1)	54(1)	1(1)	7(1)	15(1)
O(2)	56(1)	47(1)	71(1)	6(1)	20(1)	24(1)
O(3)	62(1)	43(1)	59(1)	2(1)	22(1)	18(1)
N(1)	45(1)	54(2)	59(1)	-14(1)	10(1)	10(1)
N(2)	49(1)	49(1)	50(1)	-4(1)	13(1)	13(1)
N(3)	44(1)	46(1)	53(1)	-7(1)	3(1)	14(1)
C(1)	49(2)	42(2)	44(1)	-3(1)	4(1)	17(1)
C(2)	42(2)	50(2)	55(2)	-4(1)	2(1)	18(1)
C(3)	63(2)	48(2)	66(2)	6(1)	-10(2)	10(2)

C(4)	66(2)	50(2)	86(2)	2(2)	-22(2)	8(2)
C(5)	60(2)	66(2)	77(2)	-22(2)	-20(2)	22(2)
C(6)	69(2)	68(2)	55(2)	-8(2)	-10(2)	22(2)
C(7)	59(2)	47(2)	62(2)	4(1)	2(2)	20(2)
C(8)	60(2)	50(2)	38(1)	-2(1)	6(1)	18(1)
C(9)	83(2)	47(2)	58(2)	-1(1)	-7(2)	20(2)
C(10)	101(3)	48(2)	70(2)	4(2)	-20(2)	10(2)
C(11)	78(3)	62(2)	65(2)	-6(2)	-6(2)	1(2)
C(12)	64(2)	102(3)	63(2)	-19(2)	-10(2)	28(2)
C(13)	55(2)	67(2)	61(2)	-14(2)	-6(2)	24(2)
C(14)	49(2)	42(2)	46(1)	-6(1)	2(1)	19(1)

Table S7. Hydrogen coordinates ( $\times 10^4$ ) and isotropic displacement parameters ( $\text{\AA}^2 \times 10^3$ )

	x	y	z	U(eq)
H(2A)	5110	4711	-274	88
H(1A)	3553	4568	4495	71
H(1B)	3209	4109	4009	71
H(2)	4347	4722	2502	65
H(3)	3923	4025	2051	62
H(3A)	4716	5534	2883	80
H(4)	5160	6075	4302	92
H(5)	5120	5898	6405	86
H(6)	4652	5173	7066	83
H(7)	4215	4620	5629	71
H(9)	3483	3188	2313	82
H(10)	2823	2583	1661	102
H(11)	2213	2648	1142	100
H(12)	2249	3339	1321	98
H(13)	2930	3976	1848	77

Table S8. Torsion angles [ $^\circ$ ] for d8v19290.

C(2)-N(2)-C(1)-N(1)	21.2(5)
C(2)-N(2)-C(1)-N(3)	-159.8(3)
C(8)-N(3)-C(1)-N(1)	10.0(5)
C(8)-N(3)-C(1)-N(2)	-168.9(3)
C(1)-N(2)-C(2)-C(7)	42.8(5)
C(1)-N(2)-C(2)-C(3)	-142.2(3)
C(7)-C(2)-C(3)-C(4)	1.3(5)
N(2)-C(2)-C(3)-C(4)	-173.9(3)
C(2)-C(3)-C(4)-C(5)	-1.6(6)

C(3)-C(4)-C(5)-C(6)	1.0(6)
C(4)-C(5)-C(6)-C(7)	-0.2(6)
C(3)-C(2)-C(7)-C(6)	-0.5(5)
N(2)-C(2)-C(7)-C(6)	174.5(3)
C(5)-C(6)-C(7)-C(2)	0.0(5)
C(1)-N(3)-C(8)-C(13)	48.2(4)
C(1)-N(3)-C(8)-C(9)	-135.0(3)
C(13)-C(8)-C(9)-C(10)	1.5(5)
N(3)-C(8)-C(9)-C(10)	-175.3(3)
C(8)-C(9)-C(10)-C(11)	-1.6(5)
C(9)-C(10)-C(11)-C(12)	-0.7(6)
C(10)-C(11)-C(12)-C(13)	3.1(6)
C(9)-C(8)-C(13)-C(12)	0.8(5)
N(3)-C(8)-C(13)-C(12)	177.5(3)
C(11)-C(12)-C(13)-C(8)	-3.1(5)

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Symmetry transformations used to generate equivalent atoms:

Table S9. Hydrogen bonds for d8v19290 [Å and °].

D-H...A	d(D-H)	d(H...A)	d(D...A)	<(DHA)
N(3)-H(3)...O(1)	0.88	1.90	2.780(3)	174.5
N(2)-H(2)...O(3)	0.88	1.82	2.701(3)	178.8
N(1)-H(1B)...O(1)#1	0.88	1.97	2.789(3)	155.0
O(2)-H(2A)...O(3)#2	0.84	1.78	2.623(3)	179.4
N(3)-H(3)...O(1)	0.88	1.90	2.780(3)	174.5
N(2)-H(2)...O(3)	0.88	1.82	2.701(3)	178.8
N(1)-H(1B)...O(1)#1	0.88	1.97	2.789(3)	155.0
O(2)-H(2A)...O(3)#2	0.84	1.78	2.623(3)	179.4
O(2)-H(2A)...O(3)#2	0.84	1.78	2.623(3)	179.4
N(1)-H(1B)...O(1)#1	0.88	1.97	2.789(3)	155.0
N(2)-H(2)...O(3)	0.88	1.82	2.701(3)	178.8
N(3)-H(3)...O(1)	0.88	1.90	2.780(3)	174.5
O(2)-H(2A)...O(3)#2	0.84	1.78	2.623(3)	179.4
N(1)-H(1B)...O(1)#1	0.88	1.97	2.789(3)	155.0
N(2)-H(2)...O(3)	0.88	1.82	2.701(3)	178.8
N(3)-H(3)...O(1)	0.88	1.90	2.780(3)	174.5

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Symmetry transformations used to generate equivalent atoms:

#1 -y+2/3,x-y+1/3,z+1/3      #2 -x+1,-y+1,-z

## 7. Single-crystal X-ray analysis of complex formed from DPG and AgSbF<sub>6</sub>.

Single crystal of C<sub>26</sub>H<sub>26</sub>AgF<sub>6</sub>N<sub>6</sub>Sb was selected on a 'Bruker APEX-II CCD' diffractometer. The crystal was kept at 170.0 K during data collection. Using Olex2, the structure was solved with the ShelXS structure solution program using Direct Methods and refined with the ShelXL refinement package using Least Squares minimisation. Crystal Data for C<sub>26</sub>H<sub>26</sub>AgF<sub>6</sub>N<sub>6</sub>Sb (M = 766.15 g/mol): orthorhombic, space group Pna2<sub>1</sub> (no. 33), a = 14.7369(6) Å, b = 31.5086(13) Å, c = 5.9773(2) Å, V = 2775.49(19) Å<sup>3</sup>, Z = 4, T = 170.0 K,  $\mu(\text{GaK}\alpha) = 9.462 \text{ mm}^{-1}$ , D<sub>calc</sub> = 1.833 g/cm<sup>3</sup>, 22100 reflections measured ( $5.76^\circ \leq 2\theta \leq 109.906^\circ$ ), 4443 unique ( $R_{\text{int}} = 0.1534$ ,  $R_{\text{sigma}} = 0.1249$ ) which were used in all calculations. The final R<sub>1</sub> was 0.0639 ( $I > 2\sigma(I)$ ) and wR<sub>2</sub> was 0.1822 (all data). Number of restraints - 1, number of constraints - unknown.

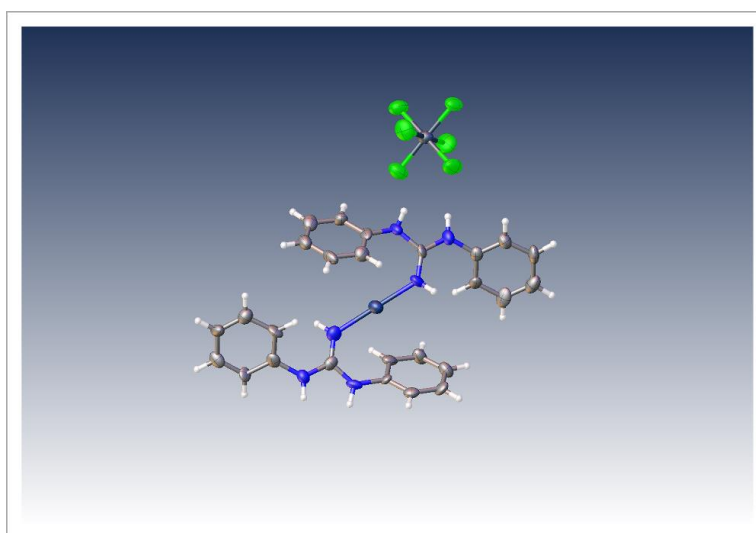


Table S10. Crystal data and structure refinement for CCDC-1894496.

Identification code	CCDC-1894496	
Empirical formula	C <sub>26</sub> H <sub>26</sub> Ag F <sub>6</sub> N <sub>6</sub> Sb	
Formula weight	766.15	
Temperature	170.0 K	
Wavelength	1.34139 Å	
Crystal system	Orthorhombic	
Space group	Pna2 <sub>1</sub>	
Unit cell dimensions	a = 14.7369(6) Å	$\alpha = 90^\circ$ .
	b = 31.5086(13) Å	$\beta = 90^\circ$ .
	c = 5.9773(2) Å	$\gamma = 90^\circ$ .

Volume	2775.49(19) Å <sup>3</sup>
Z	4
Density (calculated)	1.833 Mg/m <sup>3</sup>
Absorption coefficient	9.462 mm <sup>-1</sup>
F(000)	1504
Crystal size	0.03 x 0.02 x 0.01 mm <sup>3</sup>
Theta range for data collection	2.880 to 54.953°.
Index ranges	-15<=h<=17, -38<=k<=35, -7<=l<=4
Reflections collected	22100
Independent reflections	4443 [R(int) = 0.1534]
Completeness to theta = 53.594°	99.9 %
Absorption correction	Semi-empirical from equivalents
Max. and min. transmission	0.7508 and 0.4138
Refinement method	Full-matrix least-squares on F <sup>2</sup>
Data / restraints / parameters	4443 / 1 / 361
Goodness-of-fit on F <sup>2</sup>	1.001
Final R indices [I>2sigma(I)]	R1 = 0.0639, wR2 = 0.1336
R indices (all data)	R1 = 0.1557, wR2 = 0.1822
Absolute structure parameter	0.01(3)
Extinction coefficient	n/a
Largest diff. peak and hole	0.912 and -1.434 e.Å <sup>-3</sup>

Table S11. Atomic coordinates ( $\times 10^4$ ) and equivalent isotropic displacement parameters ( $\text{\AA}^2 \times 10^3$ )

for mj19077\_0m.  $U(\text{eq})$  is defined as one third of the trace of the orthogonalized  $U^{ij}$  tensor.

	x	y	z	$U(\text{eq})$
Sb(1)	-12684(1)	-3742(1)	8989(2)	40(1)
F(1)	-13600(9)	-3814(4)	11130(30)	55(4)
F(2)	-12860(9)	-3157(3)	9050(30)	71(4)
F(3)	-11816(11)	-3703(5)	11260(40)	77(6)
F(4)	-11748(10)	-3672(5)	6930(30)	70(5)
F(5)	-12537(9)	-4327(4)	8910(30)	70(4)
F(6)	-13560(10)	-3779(5)	6700(40)	66(5)
Ag(1)	-7678(1)	-3756(1)	6563(3)	45(1)
N(1)	-8470(13)	-3668(6)	9370(40)	43(6)
N(2)	-6865(14)	-3844(6)	3740(50)	47(6)
N(3)	-9860(12)	-3926(5)	7990(30)	41(5)
N(4)	-9898(13)	-3423(6)	10750(30)	44(5)
N(5)	-5519(12)	-3576(5)	5020(30)	35(5)
N(6)	-5465(12)	-4079(5)	2330(30)	39(5)
C(1)	-9361(14)	-3668(6)	9470(30)	32(5)
C(2)	-9600(14)	-3094(6)	12220(40)	34(6)
C(3)	-9991(15)	-3046(6)	14190(40)	42(6)
C(4)	-9775(17)	-2720(8)	15680(40)	47(7)
C(5)	-9080(20)	-2443(7)	15080(50)	53(8)
C(6)	-8683(18)	-2486(8)	13020(60)	50(7)
C(7)	-8931(12)	-2805(6)	11560(40)	35(5)
C(8)	-9537(15)	-4243(6)	6540(50)	44(6)
C(9)	-8782(16)	-4505(6)	7090(40)	45(6)
C(10)	-8519(16)	-4800(7)	5460(50)	49(8)
C(11)	-8960(17)	-4840(7)	3440(50)	51(7)
C(12)	-9691(17)	-4587(7)	2990(40)	49(7)
C(13)	-9967(15)	-4286(7)	4510(40)	42(6)
C(14)	-5996(15)	-3835(6)	3700(40)	35(5)
C(15)	-5835(12)	-3272(5)	6650(40)	28(5)
C(16)	-6580(13)	-3002(7)	6000(40)	41(6)
C(17)	-6855(16)	-2703(7)	7510(40)	42(6)



C(18)	-6458(16)	-2675(7)	9610(40)	44(7)
C(19)	-5742(15)	-2932(6)	10130(40)	41(6)
C(20)	-5448(14)	-3228(6)	8630(40)	37(5)
C(21)	-5765(14)	-4400(7)	740(40)	38(6)
C(22)	-5349(14)	-4430(6)	-1310(40)	32(5)
C(23)	-5605(15)	-4754(6)	-2740(40)	42(6)
C(24)	-6277(16)	-5043(7)	-2130(50)	44(7)
C(25)	-6698(17)	-5014(7)	-140(40)	45(7)
C(26)	-6433(13)	-4699(6)	1470(40)	40(5)

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Table S12. Bond lengths [Å] and angles [°] for mj19077\_0m.

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Sb(1)-F(1)	1.874(14)
Sb(1)-F(2)	1.862(11)
Sb(1)-F(3)	1.87(2)
Sb(1)-F(4)	1.862(16)
Sb(1)-F(5)	1.855(12)
Sb(1)-F(6)	1.884(19)
Ag(1)-N(1)	2.06(2)
Ag(1)-N(2)	2.09(3)
N(1)-C(1)	1.31(3)
N(2)-C(14)	1.28(3)
N(3)-C(1)	1.41(3)
N(3)-C(8)	1.41(3)
N(4)-C(1)	1.35(2)
N(4)-C(2)	1.43(3)
N(5)-C(14)	1.34(3)
N(5)-C(15)	1.44(2)
N(6)-C(14)	1.37(3)
N(6)-C(21)	1.46(3)
C(2)-C(3)	1.32(3)
C(2)-C(7)	1.40(3)
C(3)-C(4)	1.39(3)
C(4)-C(5)	1.40(3)
C(5)-C(6)	1.37(4)
C(6)-C(7)	1.38(3)
C(8)-C(9)	1.42(3)
C(8)-C(13)	1.38(3)
C(9)-C(10)	1.40(3)

C(10)-C(11)	1.38(3)
C(11)-C(12)	1.37(3)
C(12)-C(13)	1.38(3)
C(15)-C(16)	1.44(3)
C(15)-C(20)	1.32(3)
C(16)-C(17)	1.37(3)
C(17)-C(18)	1.39(3)
C(18)-C(19)	1.37(3)
C(19)-C(20)	1.36(3)
C(21)-C(22)	1.37(3)
C(21)-C(26)	1.43(3)
C(22)-C(23)	1.38(3)
C(23)-C(24)	1.39(3)
C(24)-C(25)	1.34(3)
C(25)-C(26)	1.44(3)
F(1)-Sb(1)-F(6)	89.7(6)
F(2)-Sb(1)-F(1)	90.3(6)
F(2)-Sb(1)-F(3)	90.9(8)
F(2)-Sb(1)-F(6)	88.9(7)
F(3)-Sb(1)-F(1)	90.3(8)
F(3)-Sb(1)-F(6)	179.8(8)
F(4)-Sb(1)-F(1)	178.2(7)
F(4)-Sb(1)-F(2)	89.9(7)
F(4)-Sb(1)-F(3)	88.0(7)
F(4)-Sb(1)-F(6)	92.1(8)
F(5)-Sb(1)-F(1)	89.0(6)
F(5)-Sb(1)-F(2)	178.7(6)
F(5)-Sb(1)-F(3)	90.2(7)
F(5)-Sb(1)-F(4)	90.9(6)
F(5)-Sb(1)-F(6)	90.1(7)
N(1)-Ag(1)-N(2)	179.4(9)
C(1)-N(1)-Ag(1)	126.8(19)
C(14)-N(2)-Ag(1)	126(2)
C(1)-N(3)-C(8)	128.3(18)
C(1)-N(4)-C(2)	125.8(19)
C(14)-N(5)-C(15)	129.3(17)
C(14)-N(6)-C(21)	127.4(18)
N(1)-C(1)-N(3)	119.7(19)
N(1)-C(1)-N(4)	128(2)

N(4)-C(1)-N(3)	112.4(18)
C(3)-C(2)-N(4)	120(2)
C(3)-C(2)-C(7)	119(2)
C(7)-C(2)-N(4)	121(2)
C(2)-C(3)-C(4)	124(2)
C(3)-C(4)-C(5)	118(2)
C(6)-C(5)-C(4)	119(3)
C(5)-C(6)-C(7)	122(3)
C(6)-C(7)-C(2)	119(2)
N(3)-C(8)-C(9)	122(2)
C(13)-C(8)-N(3)	117(2)
C(13)-C(8)-C(9)	120(2)
C(10)-C(9)-C(8)	116(2)
C(11)-C(10)-C(9)	123(2)
C(12)-C(11)-C(10)	119(2)
C(11)-C(12)-C(13)	120(3)
C(12)-C(13)-C(8)	121(2)
N(2)-C(14)-N(5)	122(2)
N(2)-C(14)-N(6)	125(2)
N(5)-C(14)-N(6)	113.3(19)
C(16)-C(15)-N(5)	117(2)
C(20)-C(15)-N(5)	122.3(19)
C(20)-C(15)-C(16)	120(2)
C(17)-C(16)-C(15)	117(2)
C(16)-C(17)-C(18)	121(2)
C(19)-C(18)-C(17)	120(2)
C(20)-C(19)-C(18)	120(2)
C(15)-C(20)-C(19)	122(2)
C(22)-C(21)-N(6)	120(2)
C(22)-C(21)-C(26)	122(2)
C(26)-C(21)-N(6)	118(2)
C(21)-C(22)-C(23)	119(2)
C(22)-C(23)-C(24)	121(2)
C(25)-C(24)-C(23)	121(2)
C(24)-C(25)-C(26)	121(2)
C(21)-C(26)-C(25)	116(2)

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Symmetry transformations used to generate equivalent atoms:

Table S13. Anisotropic displacement parameters ( $\text{\AA}^2 \times 10^3$ ) for mj19077\_0m. The anisotropic displacement factor exponent takes the form:  $-2 \text{ h}^2 \text{ a}^* 2 \text{ U}11 + \dots + 2 \text{ h k a}^* \text{ b}^* \text{ U}12$  ]

	U11	U22	U33	U23	U13	U12
Sb(1)	33(1)	39(1)	48(1)	4(1)	0(1)	0(1)
F(1)	41(8)	78(9)	45(10)	12(7)	6(7)	-13(7)
F(2)	85(11)	37(6)	92(12)	-3(8)	8(11)	-11(6)
F(3)	44(10)	132(15)	55(12)	1(12)	-12(10)	-8(9)
F(4)	49(10)	99(13)	62(12)	8(9)	12(9)	2(8)
F(5)	78(11)	50(7)	81(12)	-1(8)	4(11)	24(6)
F(6)	44(10)	87(10)	67(15)	2(9)	-7(10)	-11(8)
Ag(1)	34(1)	50(1)	50(2)	-8(1)	7(1)	-2(1)
N(1)	26(11)	70(13)	34(16)	0(10)	11(9)	-2(9)
N(2)	42(13)	59(12)	41(15)	-4(12)	-7(12)	6(9)
N(3)	30(10)	43(10)	49(13)	-17(9)	16(9)	-9(8)
N(4)	45(13)	43(11)	43(13)	-13(9)	9(10)	-1(9)
N(5)	18(10)	35(9)	50(13)	8(9)	6(8)	-1(8)
N(6)	38(12)	46(11)	34(13)	-3(9)	8(9)	0(9)
C(1)	38(13)	38(12)	18(13)	-2(9)	10(9)	1(9)
C(2)	33(13)	35(12)	33(16)	-1(10)	-4(10)	12(9)
C(3)	44(14)	46(13)	35(18)	1(12)	8(13)	7(10)
C(4)	58(18)	54(16)	31(15)	-14(12)	0(12)	20(13)
C(5)	60(20)	44(14)	60(20)	-15(14)	-17(14)	9(13)
C(6)	64(17)	31(11)	56(19)	-7(12)	-13(16)	24(15)
C(7)	20(11)	46(12)	39(14)	-10(12)	-7(12)	9(9)
C(8)	50(15)	38(12)	45(16)	-18(12)	18(14)	-29(11)
C(9)	50(16)	46(13)	38(17)	6(11)	2(12)	-3(11)
C(10)	40(16)	34(12)	70(20)	-15(12)	28(14)	-2(11)
C(11)	50(17)	49(14)	50(20)	-14(13)	18(15)	-9(13)
C(12)	63(19)	47(14)	38(17)	3(12)	10(14)	-14(12)
C(13)	28(12)	53(14)	47(18)	-9(13)	-3(11)	0(10)
C(14)	38(13)	33(11)	34(14)	3(11)	-11(12)	0(9)
C(15)	23(10)	29(10)	33(13)	-10(10)	-3(11)	-9(8)
C(16)	24(12)	63(14)	37(15)	7(12)	6(11)	-13(10)
C(17)	35(14)	56(13)	35(16)	5(12)	1(12)	18(11)
C(18)	42(16)	46(13)	43(17)	1(11)	13(12)	9(11)
C(19)	40(15)	33(12)	51(17)	-17(11)	-8(12)	-10(10)
C(20)	27(12)	37(11)	47(17)	3(11)	1(12)	-11(9)
C(21)	29(14)	46(13)	39(15)	0(11)	-2(11)	19(10)

C(22)	46(14)	36(11)	15(13)	10(10)	4(11)	-3(9)
C(23)	45(14)	34(12)	46(17)	7(11)	10(12)	16(11)
C(24)	27(15)	48(14)	60(20)	-20(13)	4(12)	-1(11)
C(25)	47(17)	42(13)	46(18)	-3(12)	-6(13)	3(11)
C(26)	34(13)	49(13)	38(14)	12(12)	18(13)	-4(10)

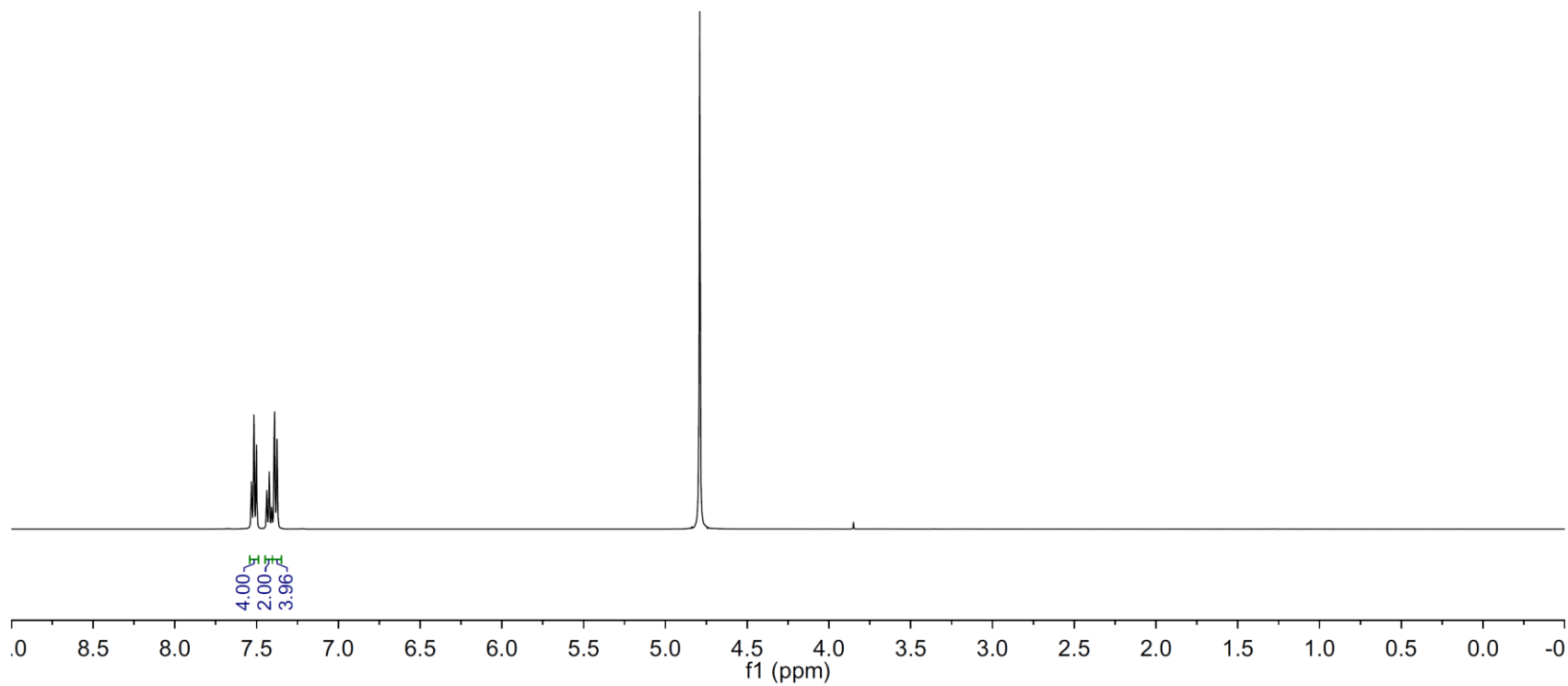
Table S14. Hydrogen coordinates ( $\times 10^4$ ) and isotropic displacement parameters ( $\text{\AA}^2 \times 10^3$ )

	x	y	z	U(eq)
H(1)	-8180	-3627	10643	52
H(2)	-7144	-3892	2462	57
H(3)	-10327	-4030	8725	49
H(4)	-10486	-3469	10673	52
H(5A)	-4926	-3594	4881	41
H(6A)	-4876	-4039	2429	47
H(3A)	-10444	-3244	14624	50
H(4A)	-10092	-2688	17057	57
H(5)	-8877	-2228	16086	64
H(6)	-8225	-2291	12584	60
H(7)	-8653	-2828	10132	42
H(9)	-8474	-4480	8475	54
H(10)	-8016	-4980	5765	58
H(11)	-8758	-5042	2370	61
H(12)	-10010	-4620	1616	59
H(13)	-10461	-4105	4151	51
H(16)	-6866	-3030	4586	49
H(17)	-7326	-2511	7121	51
H(18)	-6682	-2478	10683	52
H(19)	-5447	-2905	11538	50
H(20)	-4954	-3407	9020	45
H(22)	-4893	-4233	-1735	39
H(23)	-5318	-4780	-4153	50
H(24)	-6439	-5264	-3137	52
H(25)	-7178	-5204	198	54
H(26)	-6685	-4691	2936	48

7.534  
7.531  
7.527  
7.516  
7.514  
7.504  
7.500  
7.498  
7.440  
7.438  
7.435  
7.423  
7.419  
7.410  
7.408  
7.405  
7.392  
7.389  
7.385  
7.378  
7.375  
7.373

gxt-ge-140 H

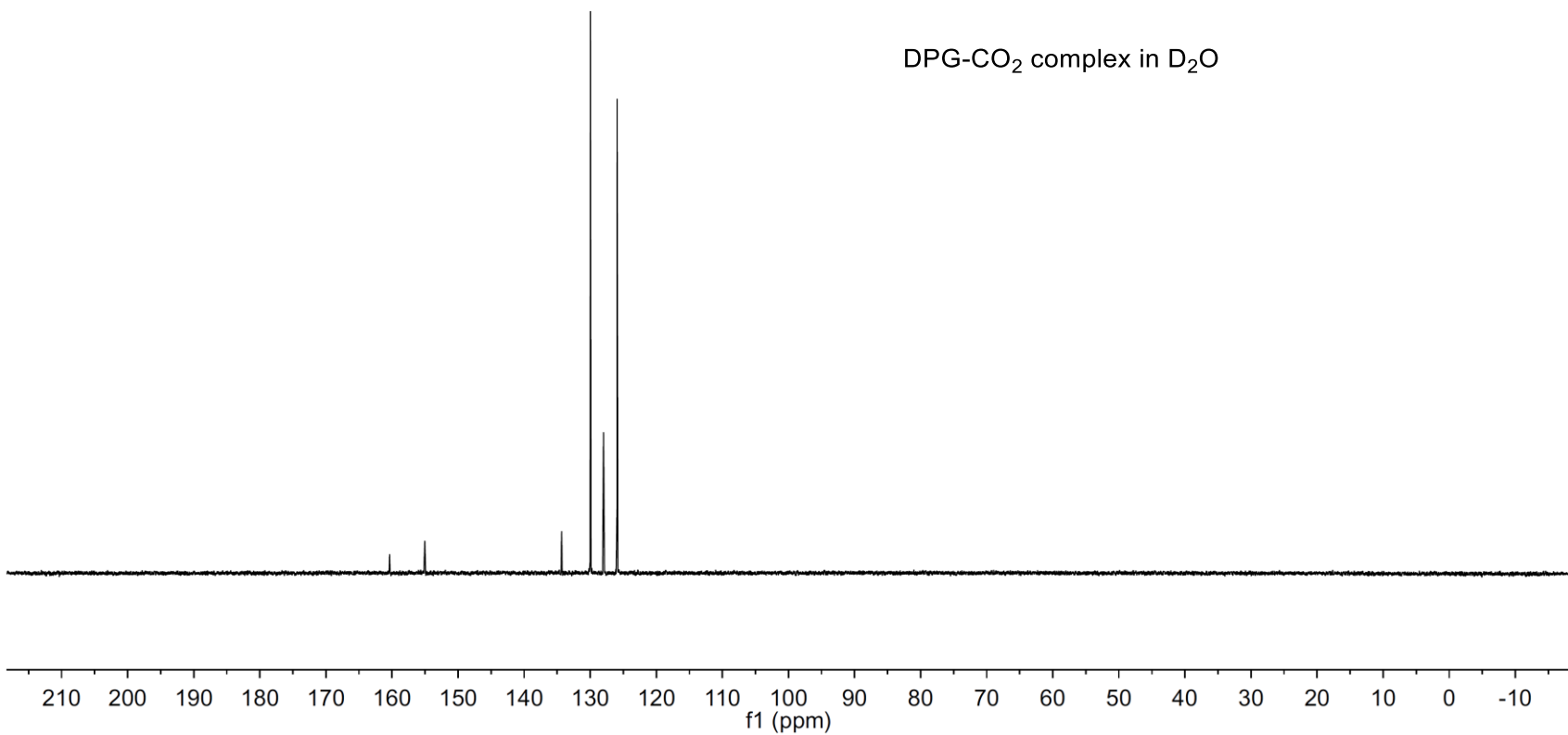
DPG-CO<sub>2</sub> complex in D<sub>2</sub>O

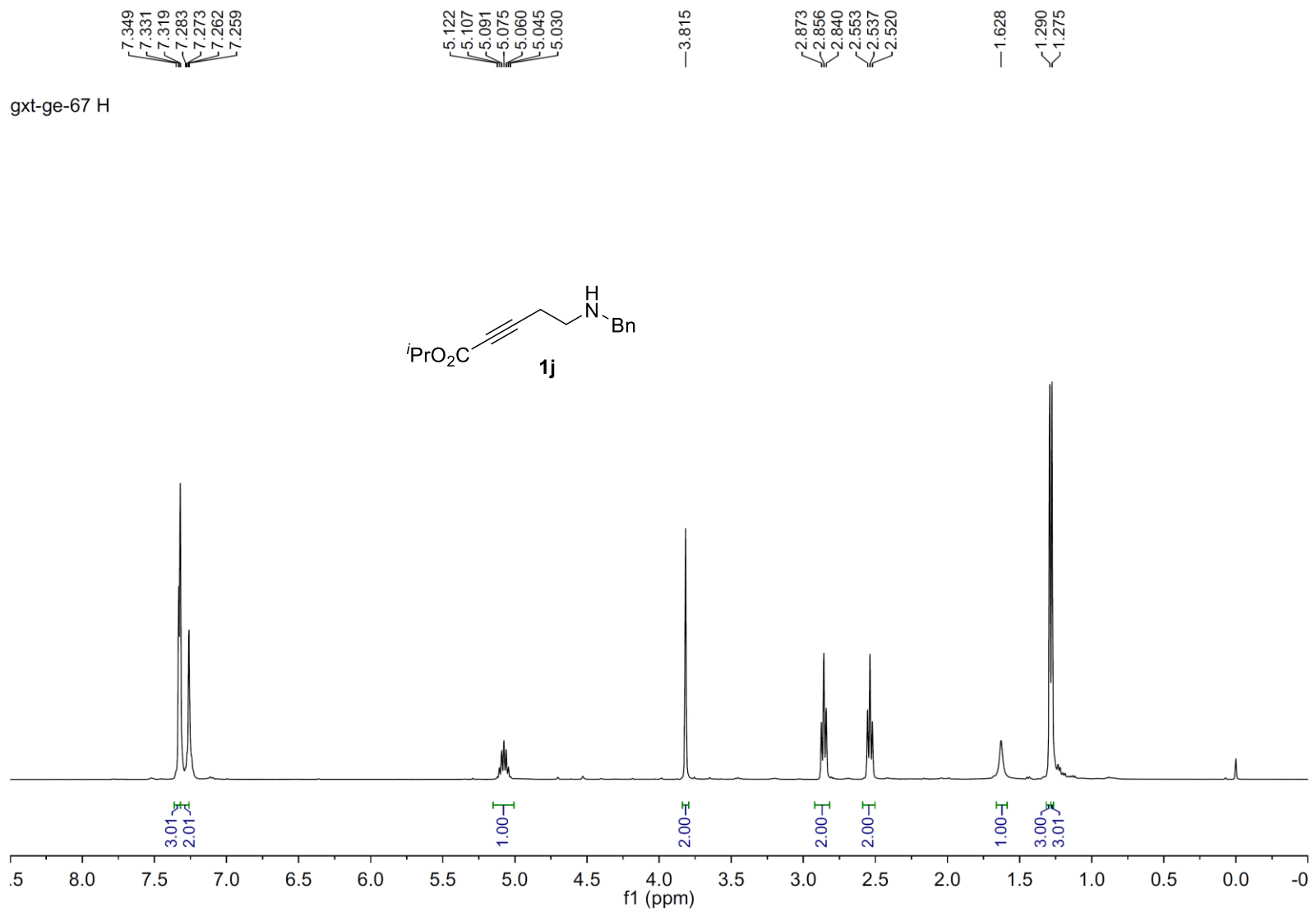


gxt-ge-140 C

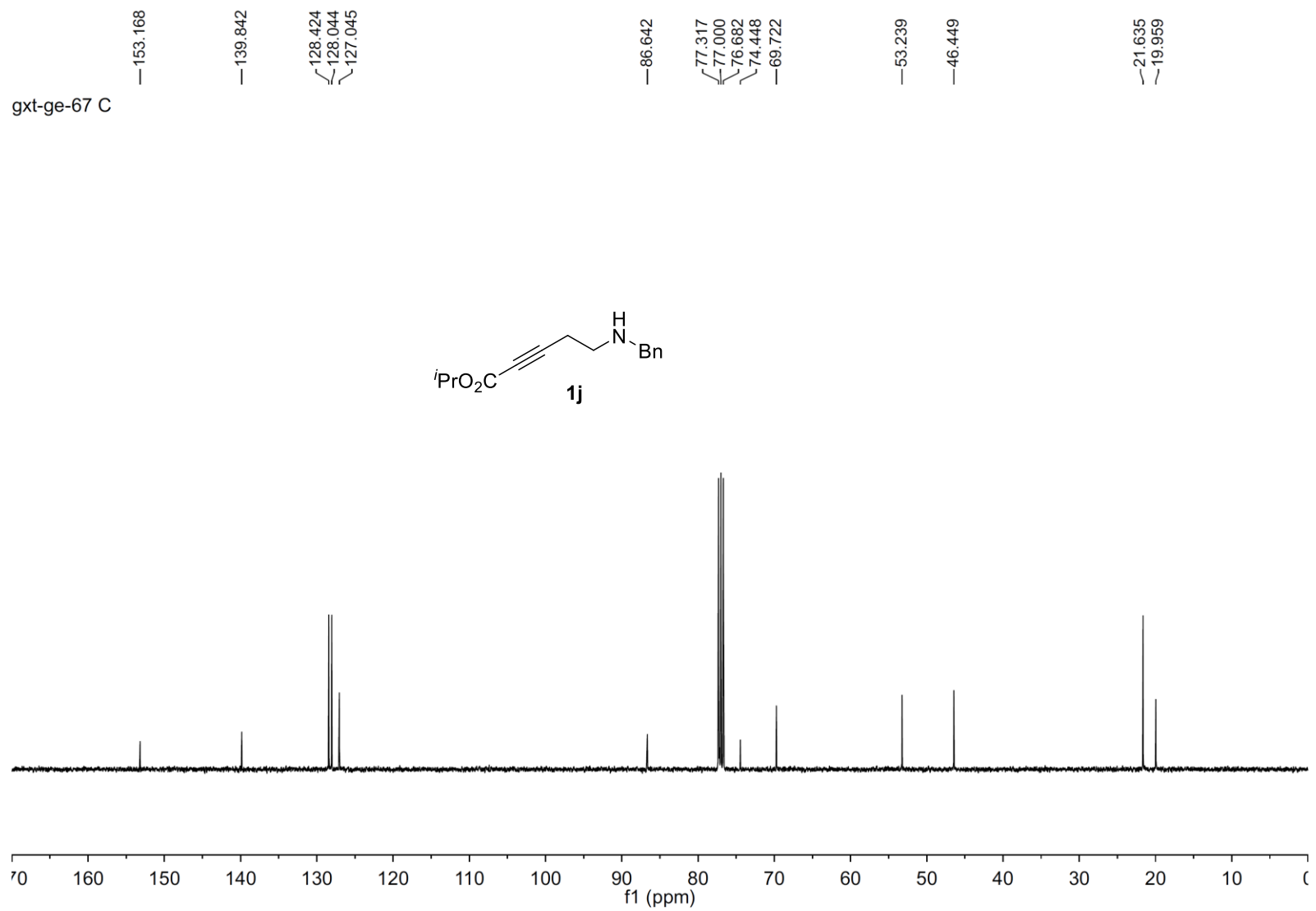
— 160.351  
— 155.038  
/ 134.321  
/ 129.971  
/ 127.995  
/ 127.980  
/ 125.932

DPG-CO<sub>2</sub> complex in D<sub>2</sub>O

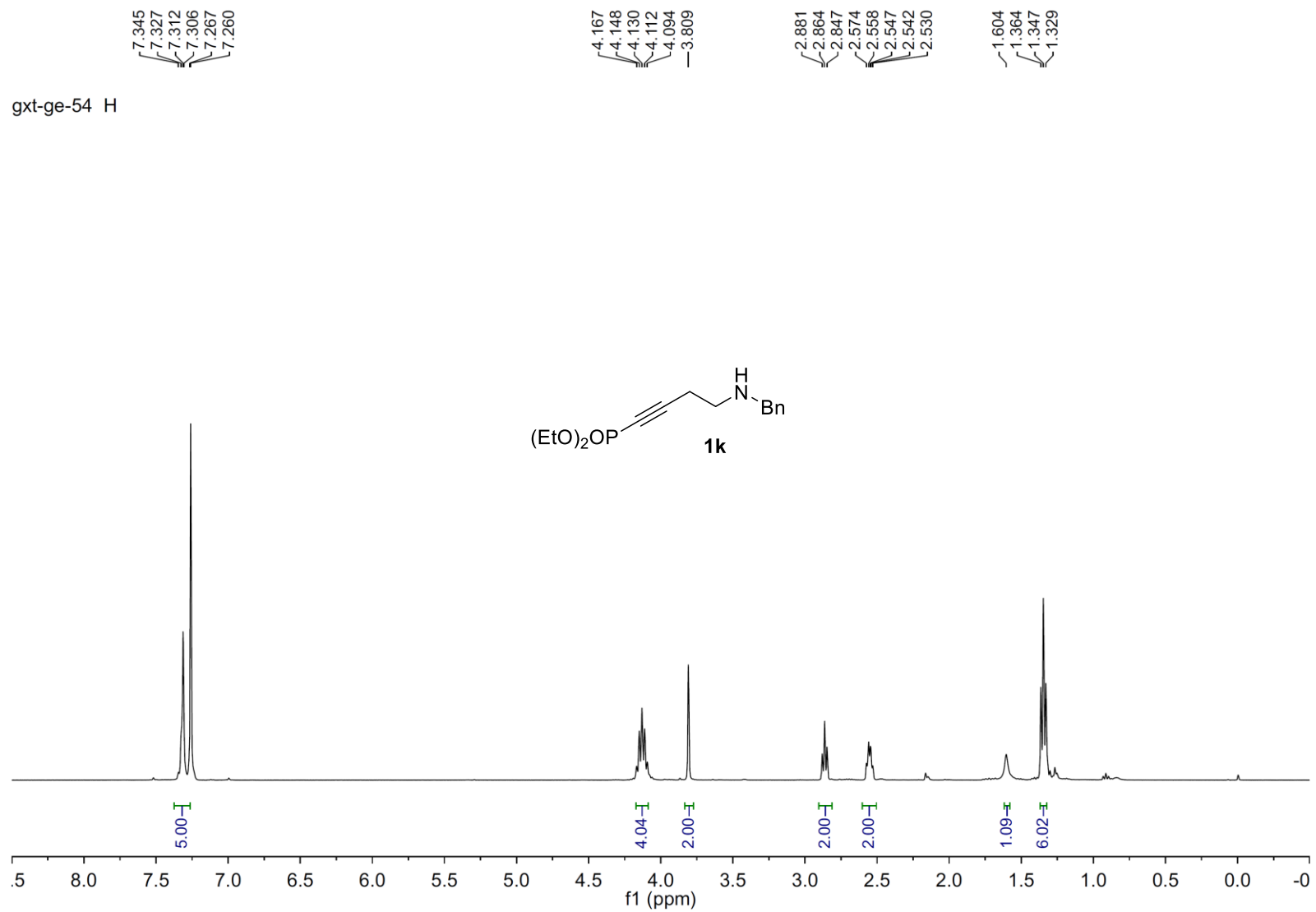




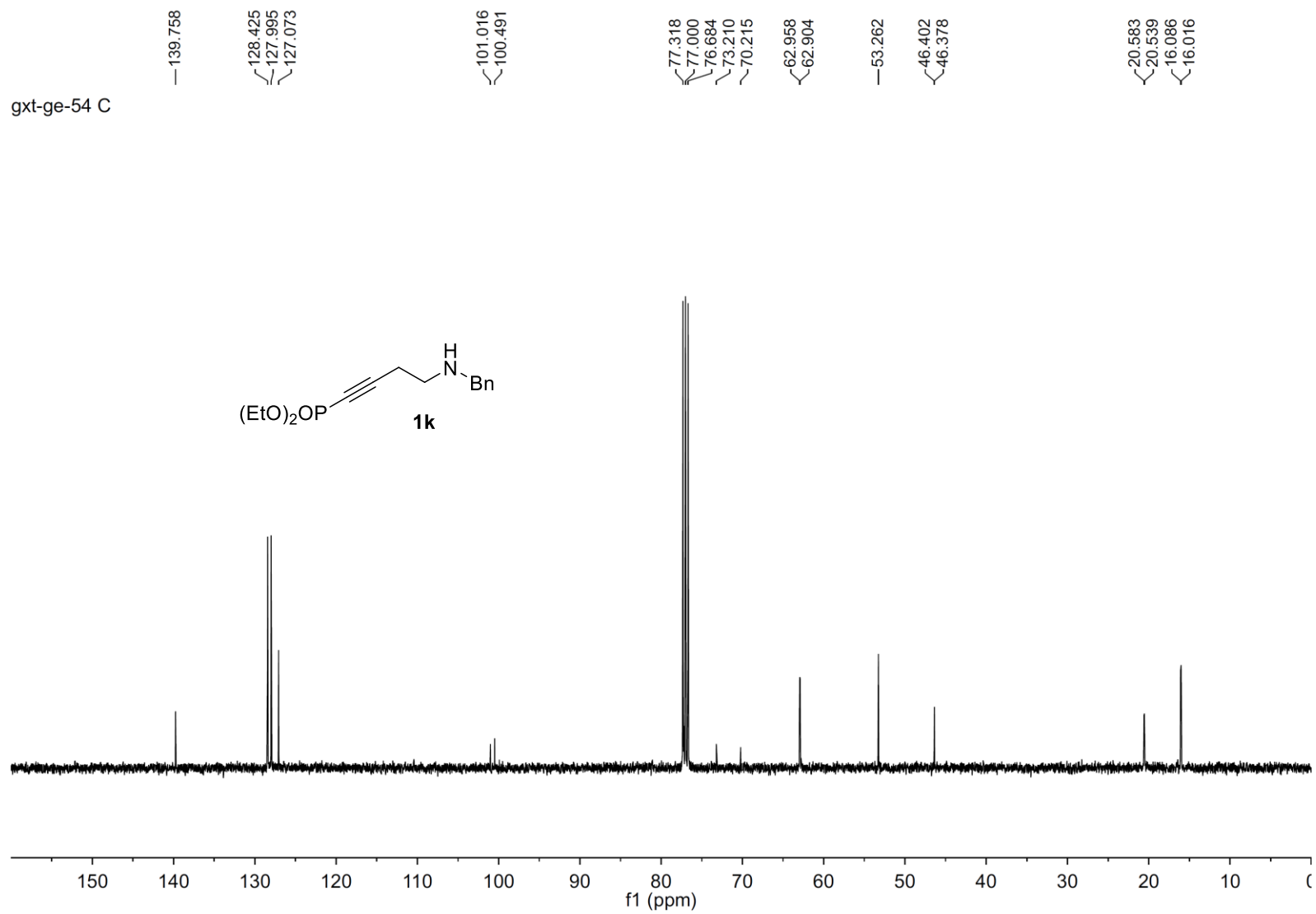




gxt-ge-54 H

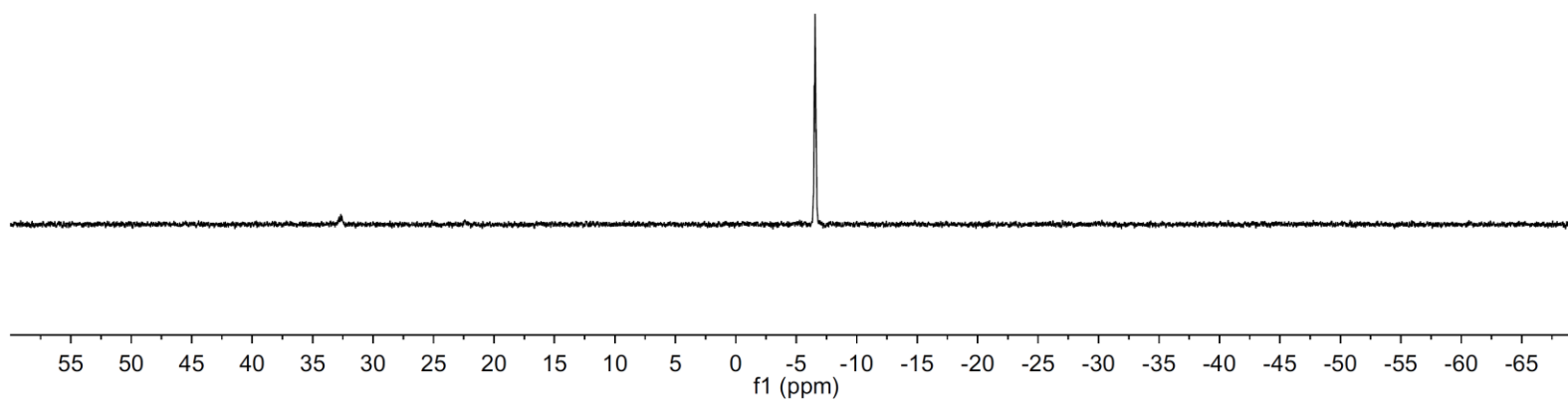
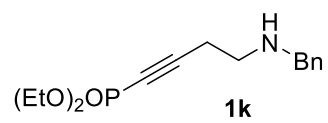


gxt-ge-54 C



gxt-ge-54 P

6.450  
6.484  
6.517  
6.552  
6.586  
6.621  
6.653

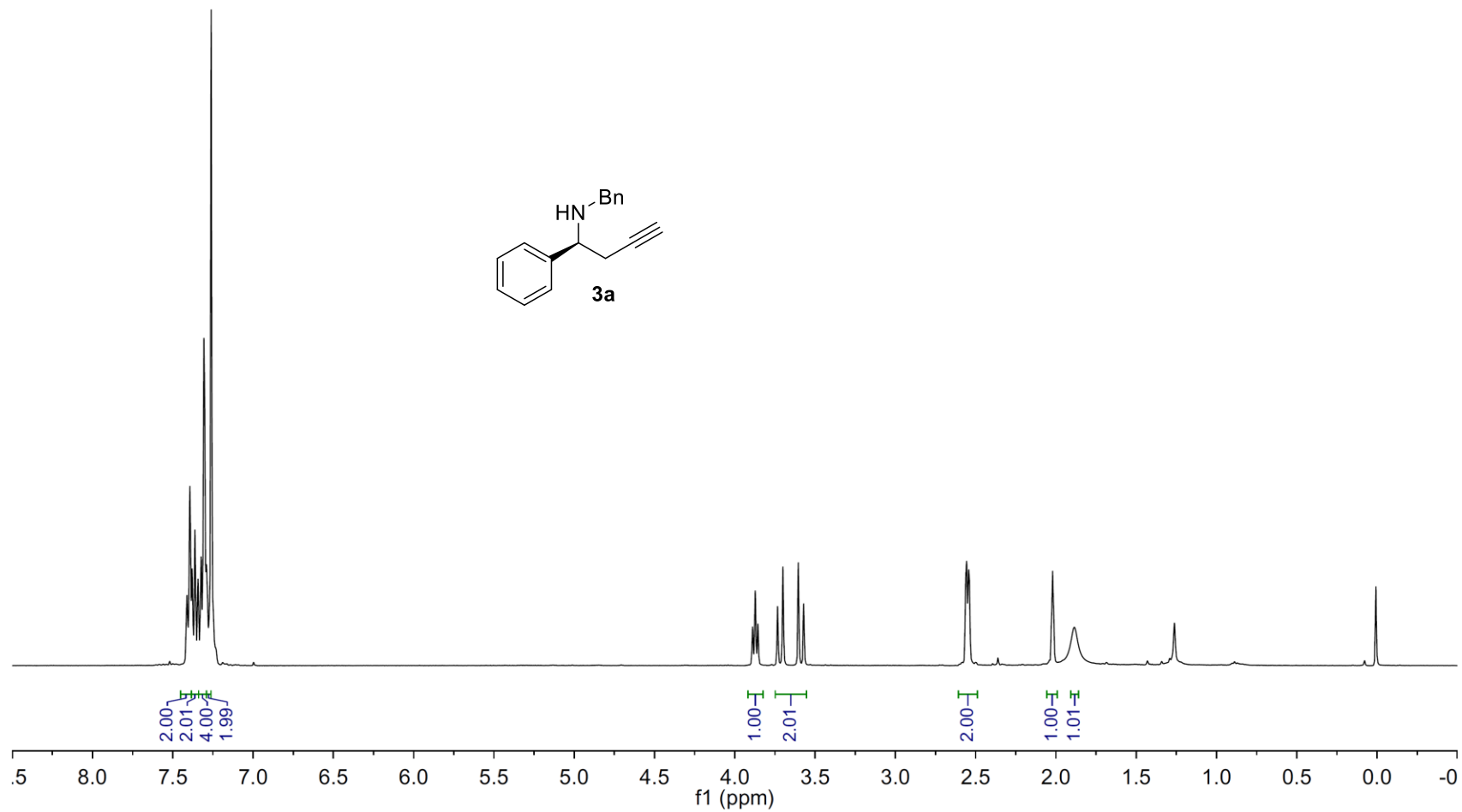


7.411  
7.393  
7.380  
7.362  
7.343  
7.324  
7.306  
7.291  
7.260

3.889  
3.873  
3.856  
3.733  
3.700  
3.604  
3.571

2.562  
2.556  
2.547  
2.543  
2.540  
2.536  
2.028  
2.023  
2.020  
2.014  
1.885

gxt-gd-108 H



gxt-gd-108 C

—142.334  
—140.149

128.386  
128.259  
127.981  
127.438  
127.070  
126.804

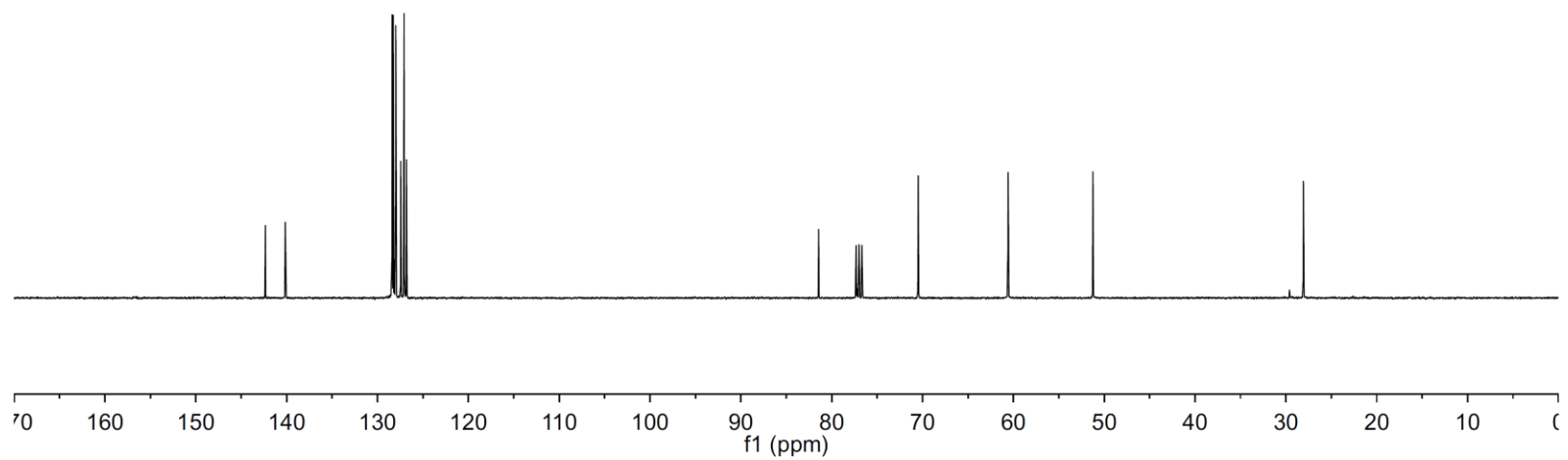
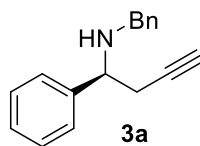
81.440  
77.318  
77.000  
76.682

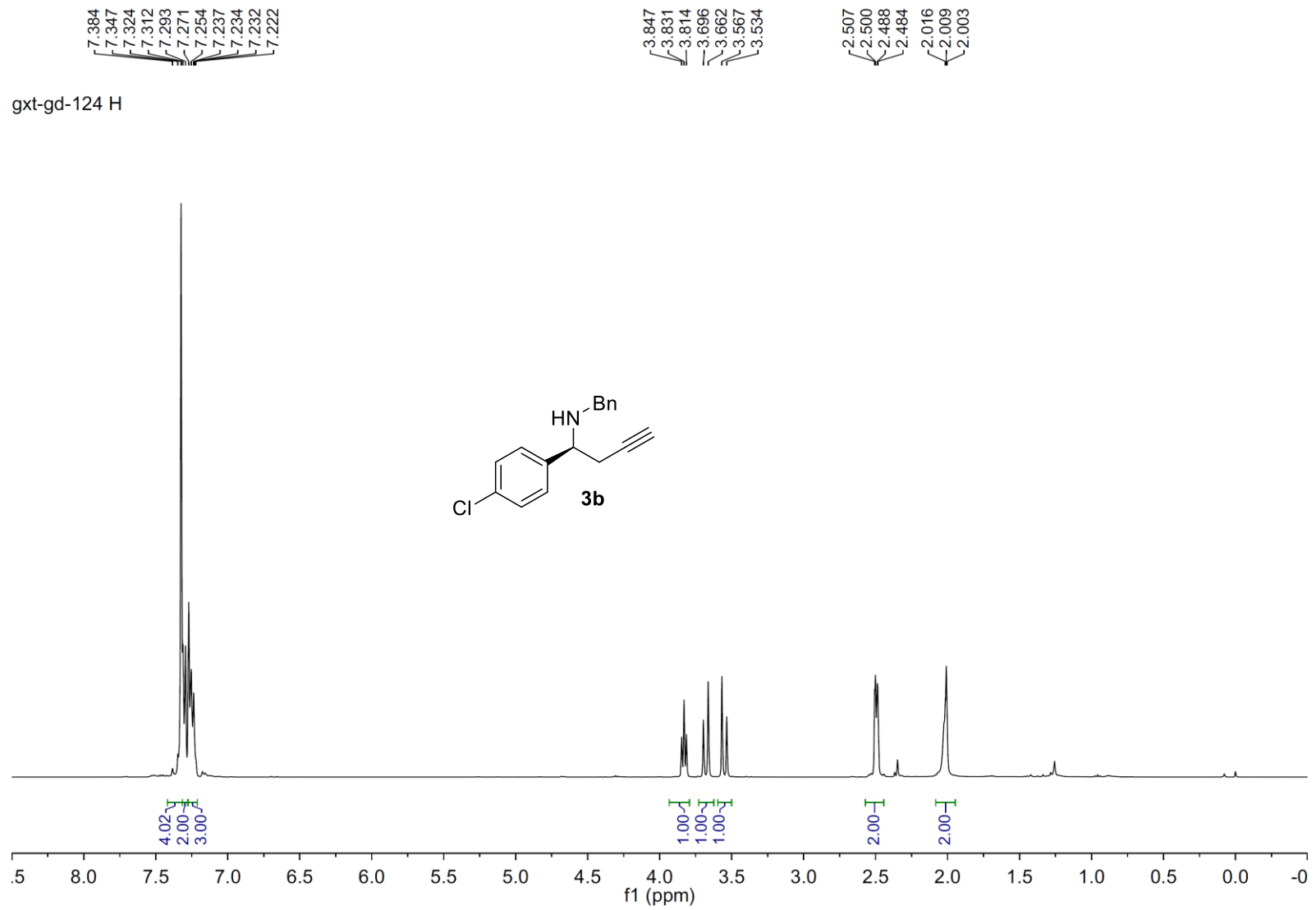
—70.482

—60.585

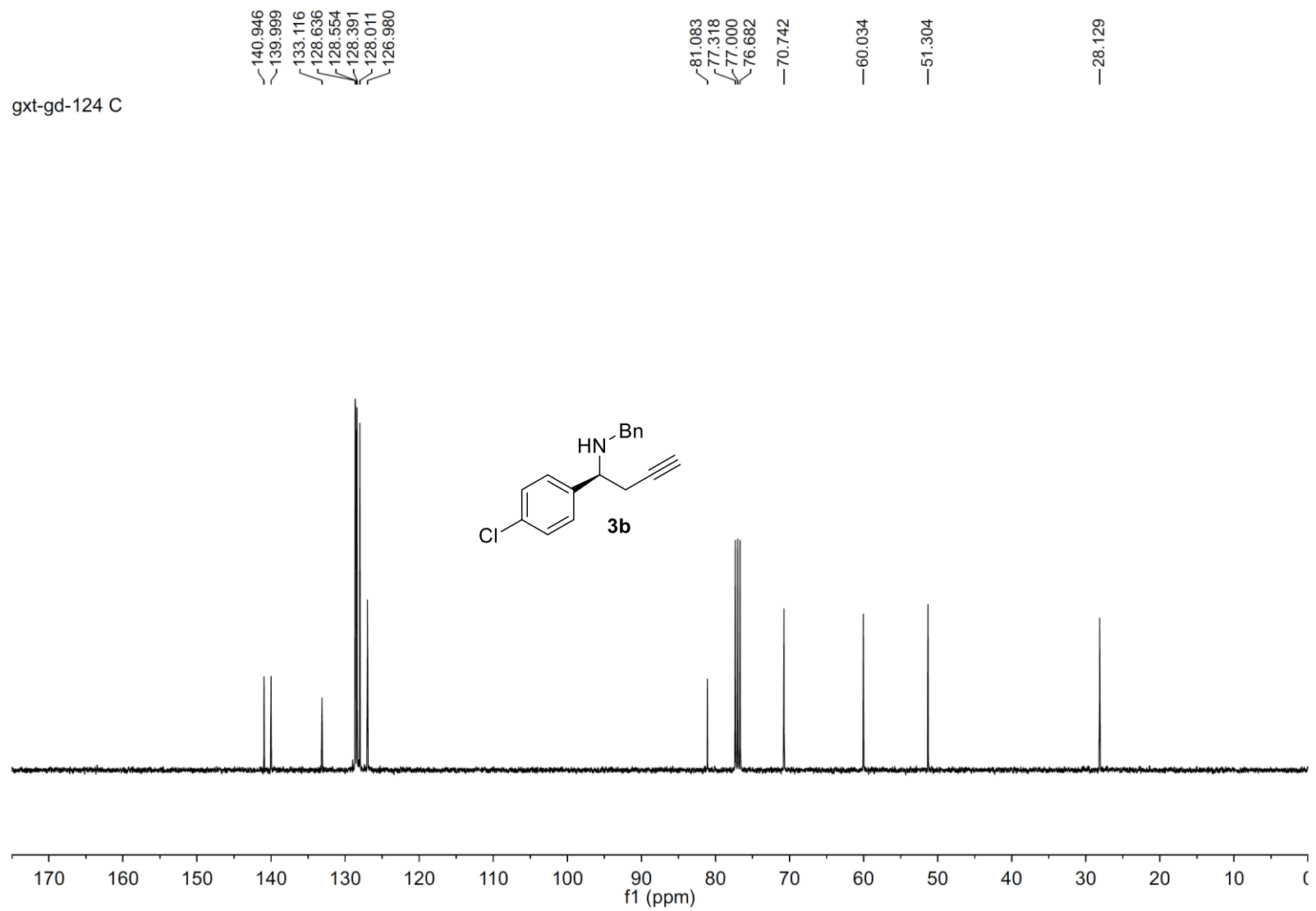
—51.247

—28.081

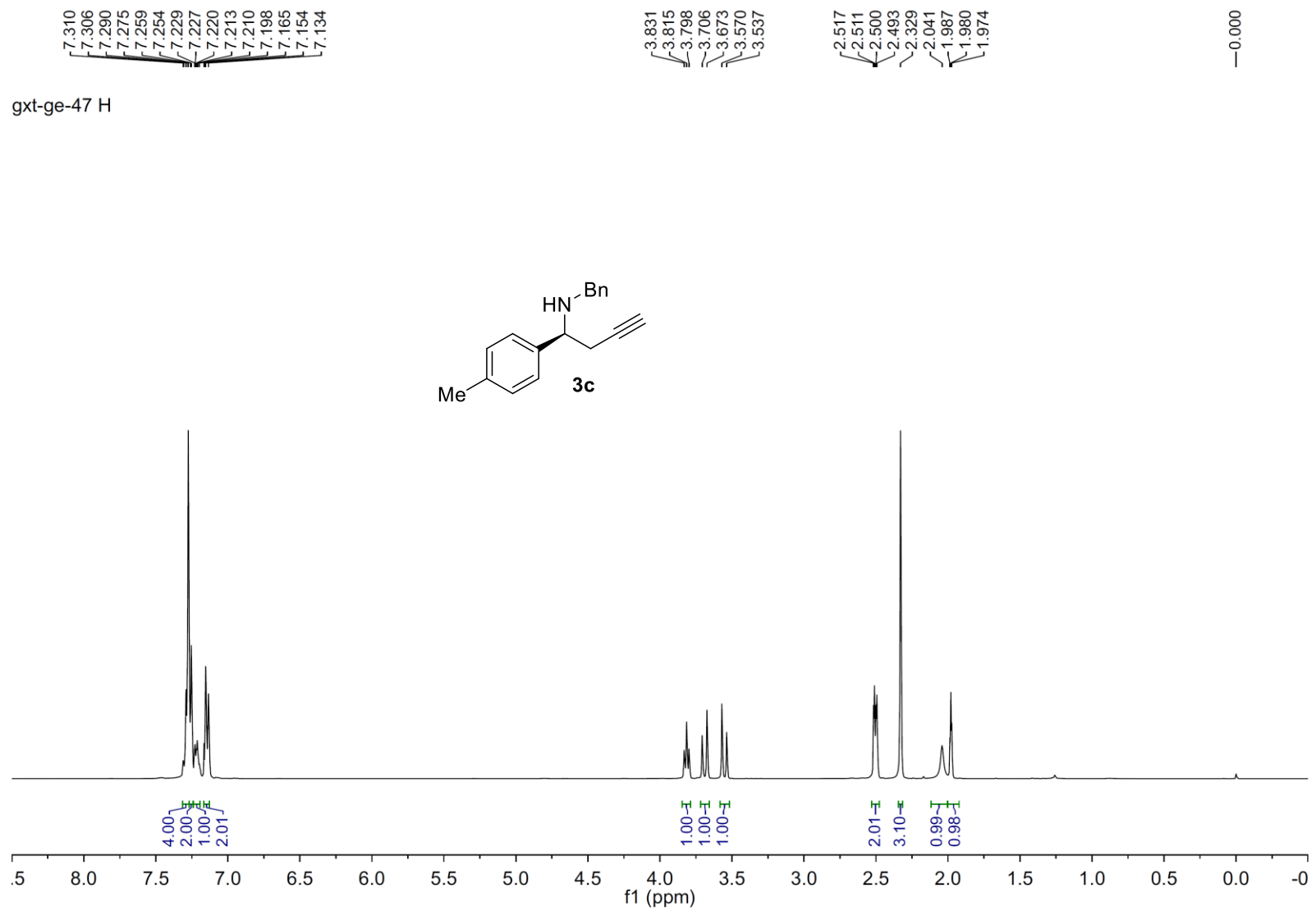




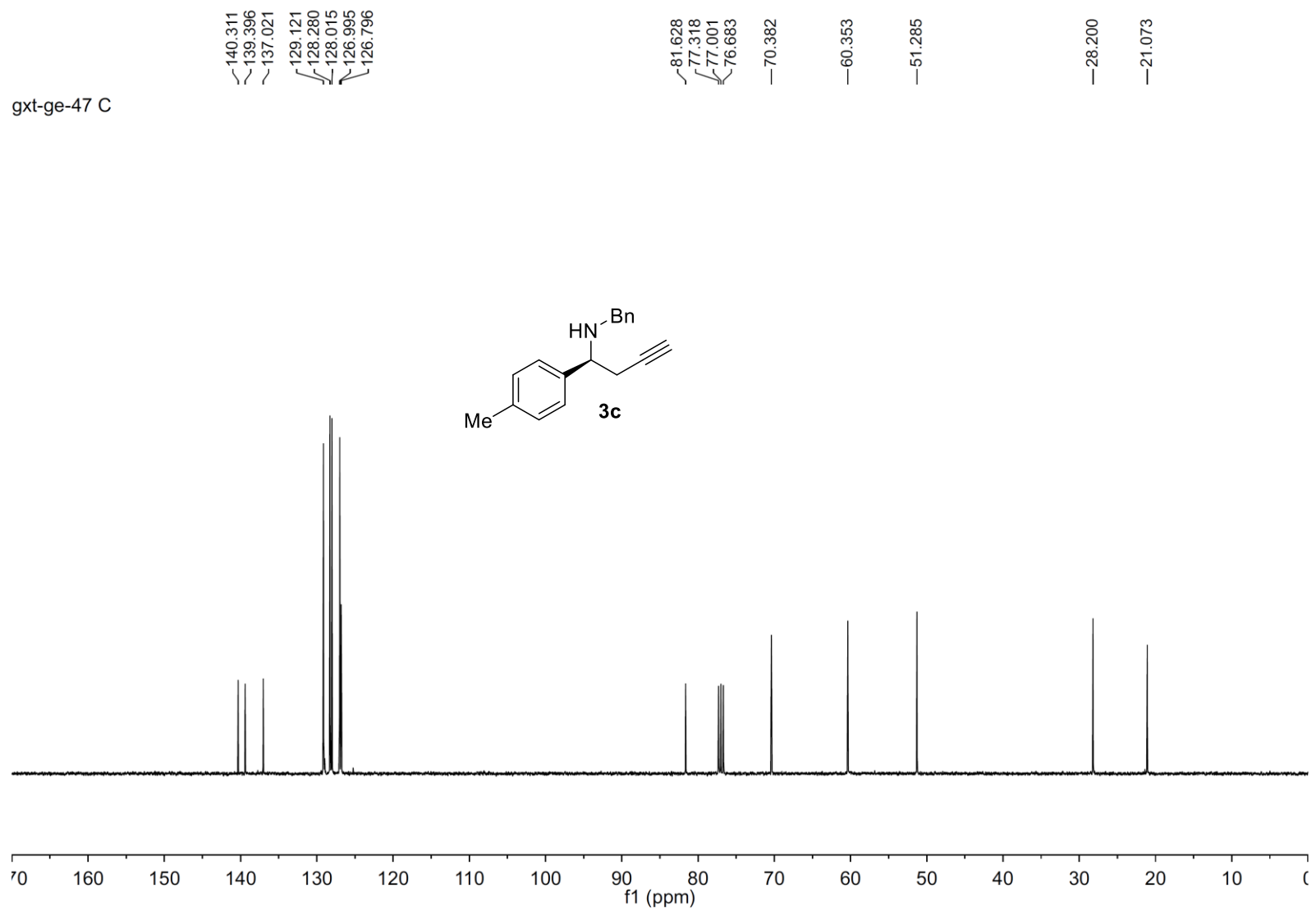
gxt-gd-124 C

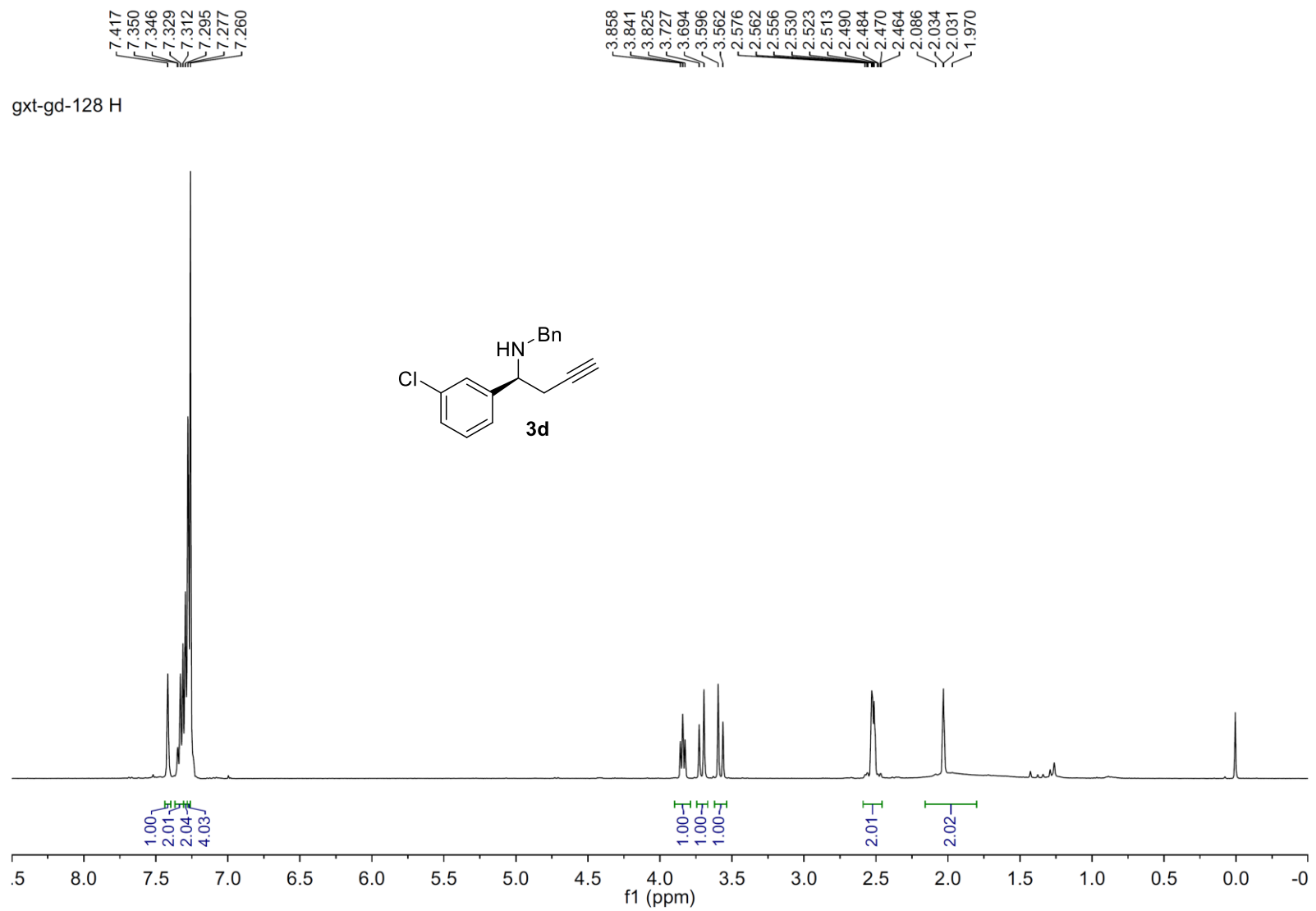




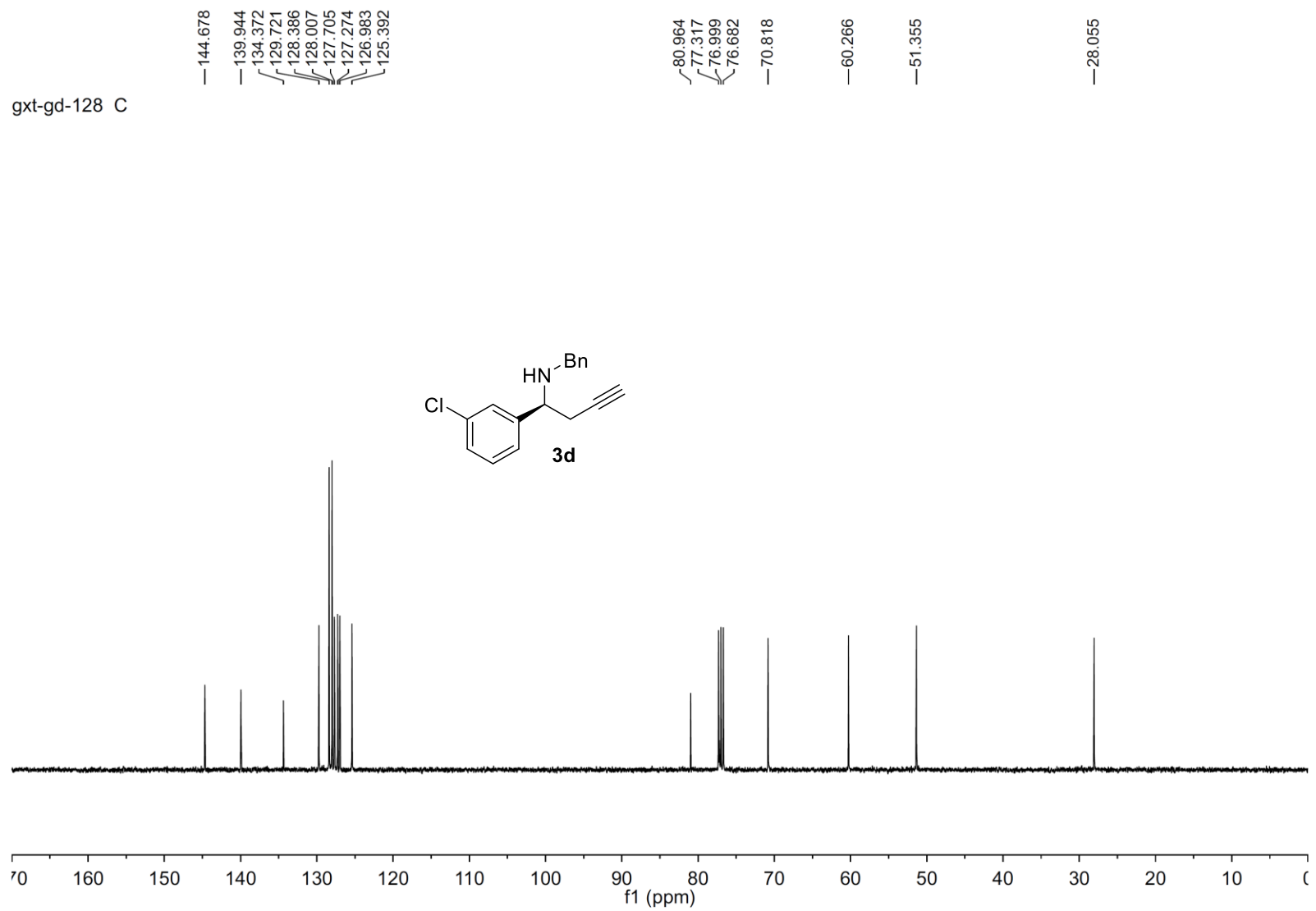


gxt-ge-47 C





gxt-gd-128 C

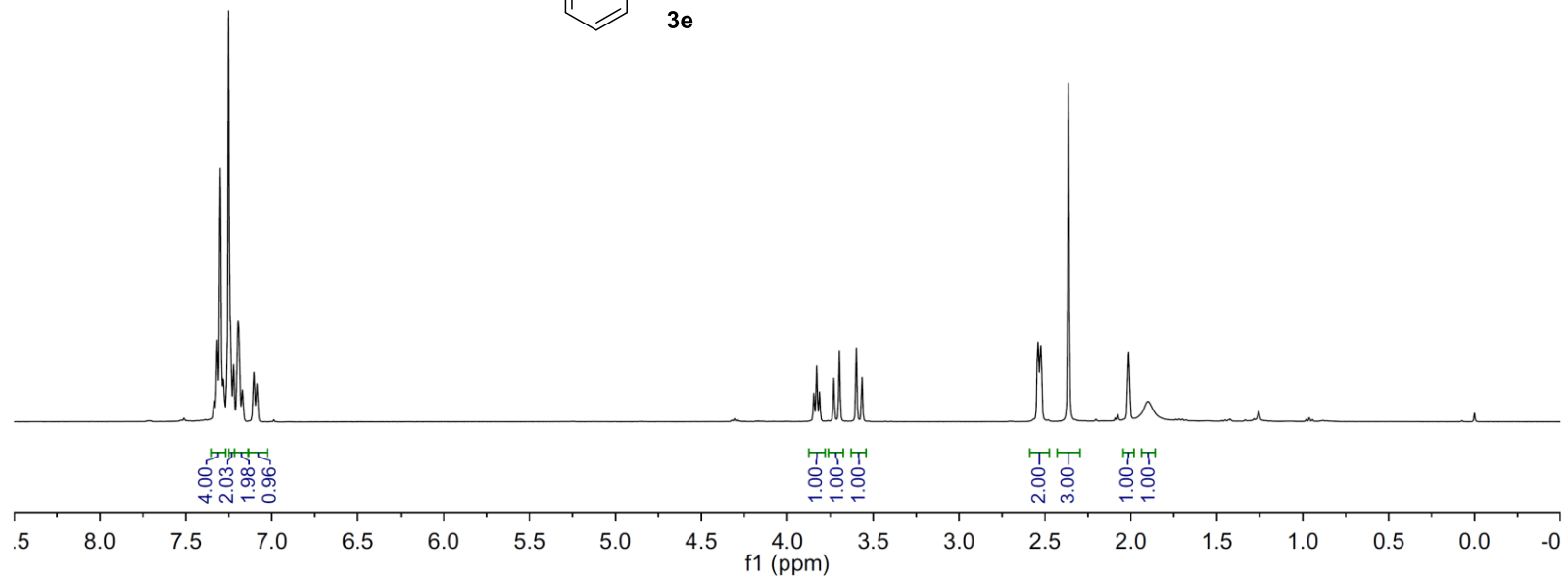
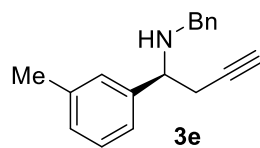


7.336  
7.318  
7.300  
7.283  
7.252  
7.241  
7.223  
7.196  
7.171  
7.105  
7.087

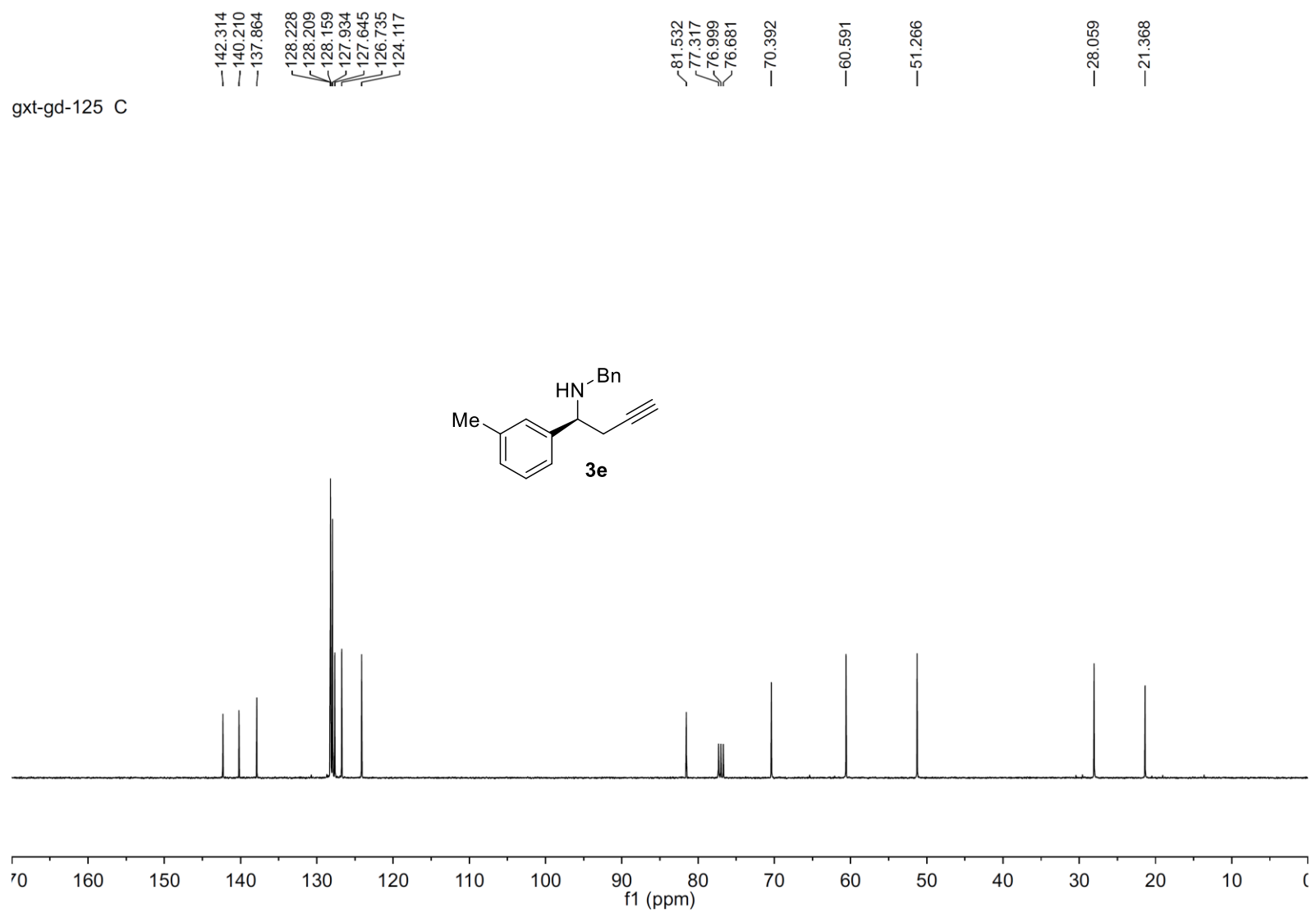
3.845  
3.829  
3.812  
3.729  
3.696  
3.598  
3.565

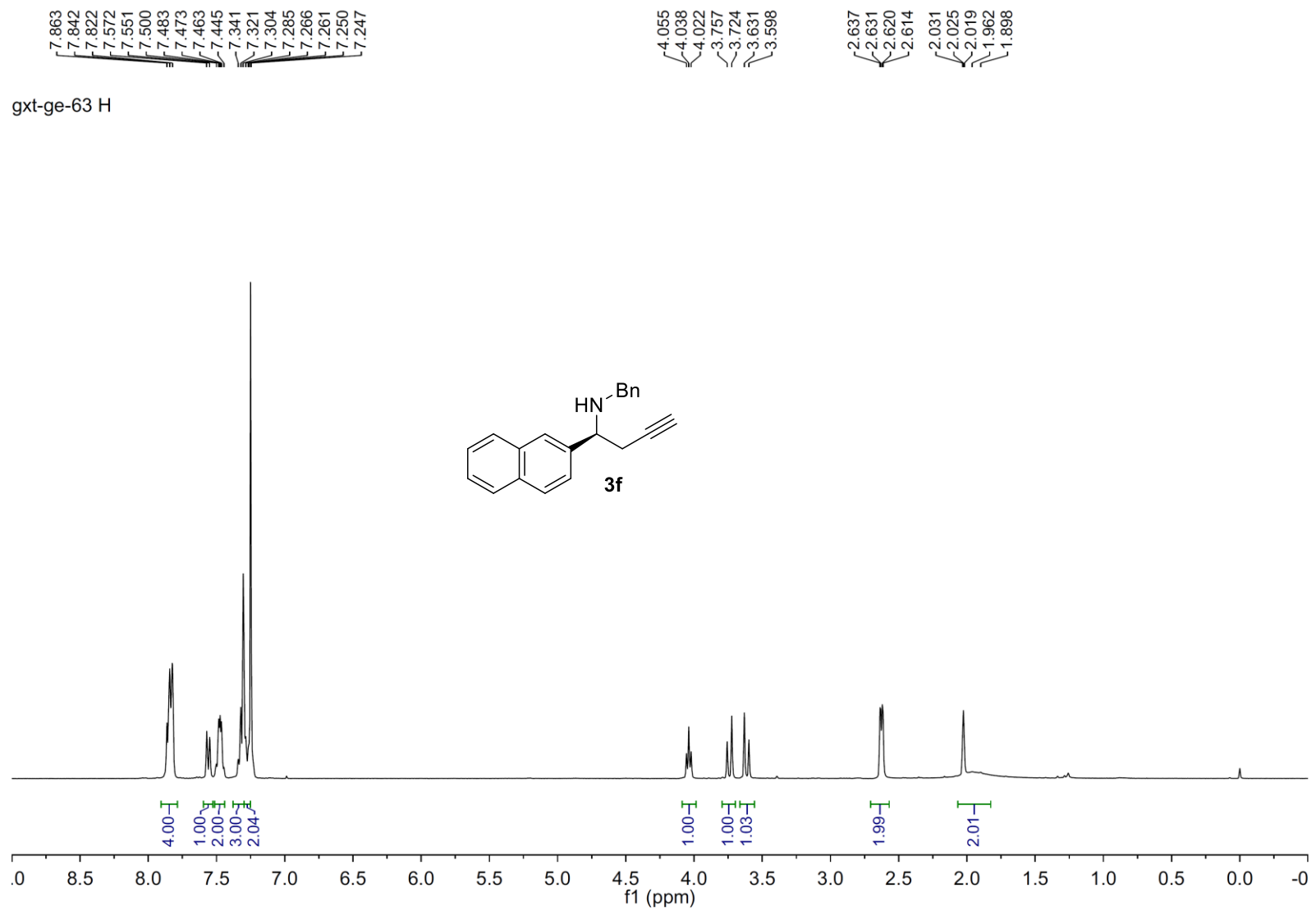
2.542  
2.537  
2.529  
2.525  
2.521  
2.363  
2.022  
2.017  
2.012  
2.007  
1.901

gxt-gd-125 H

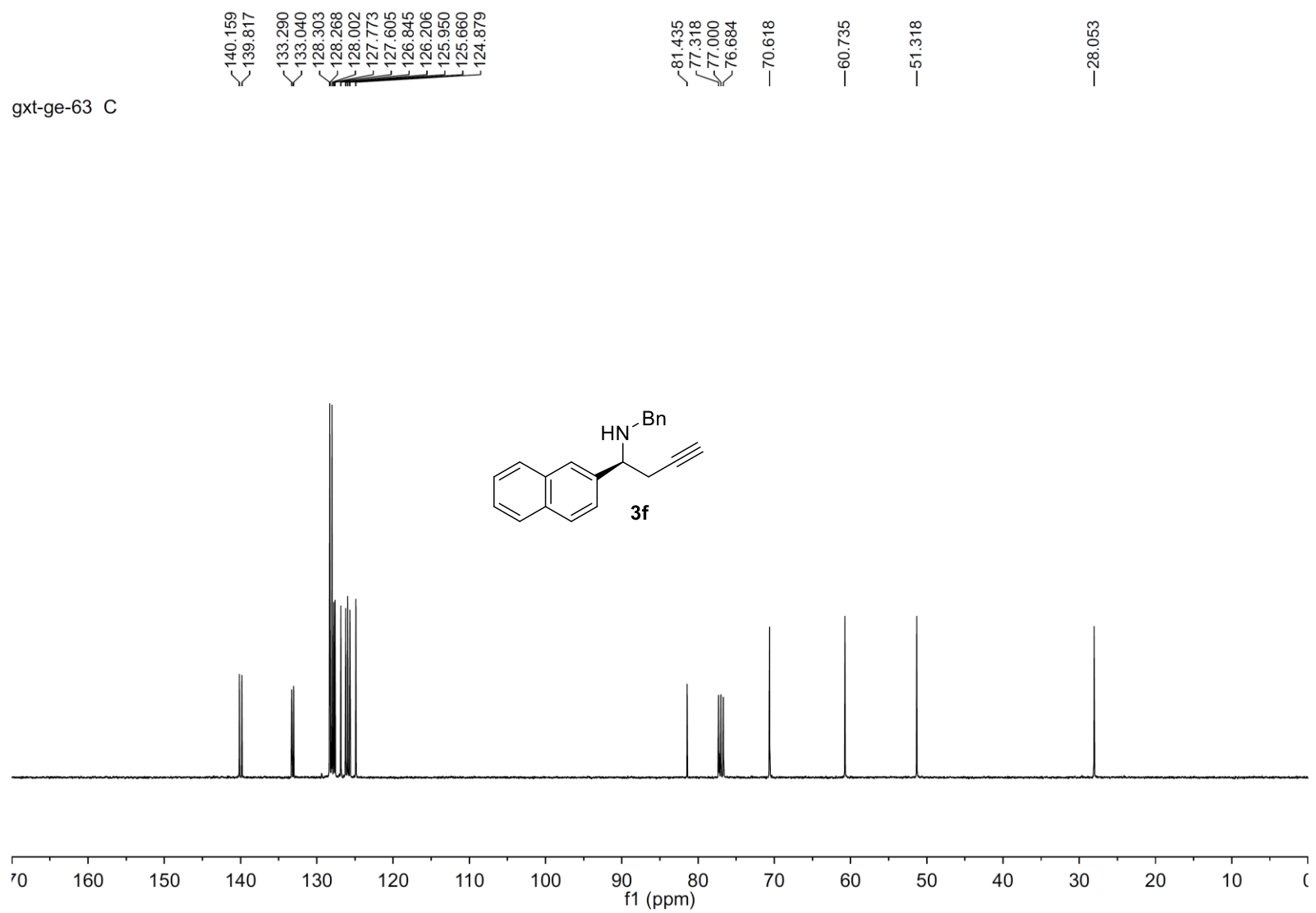


gxt-gd-125 C



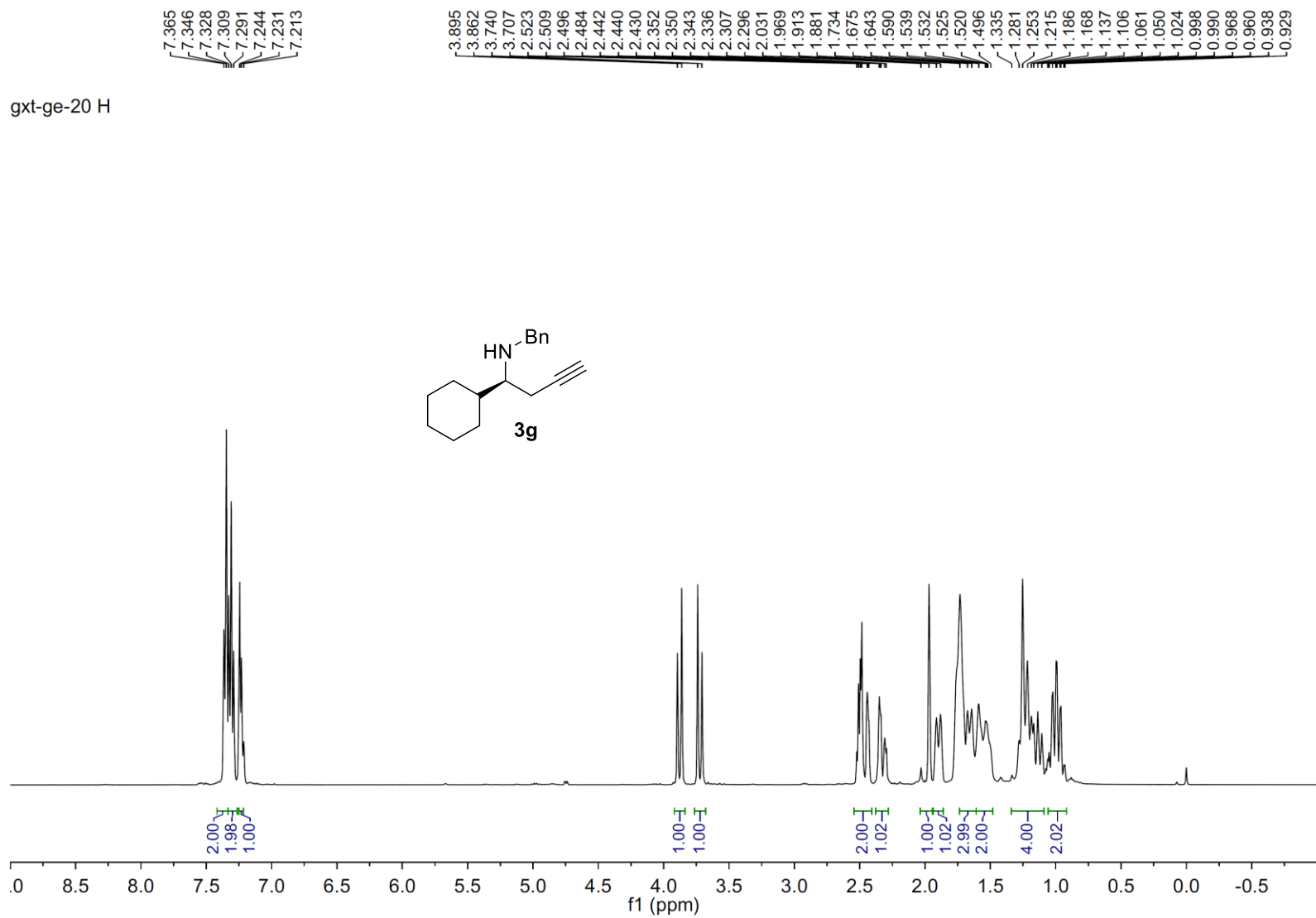


gxt-ge-63 C

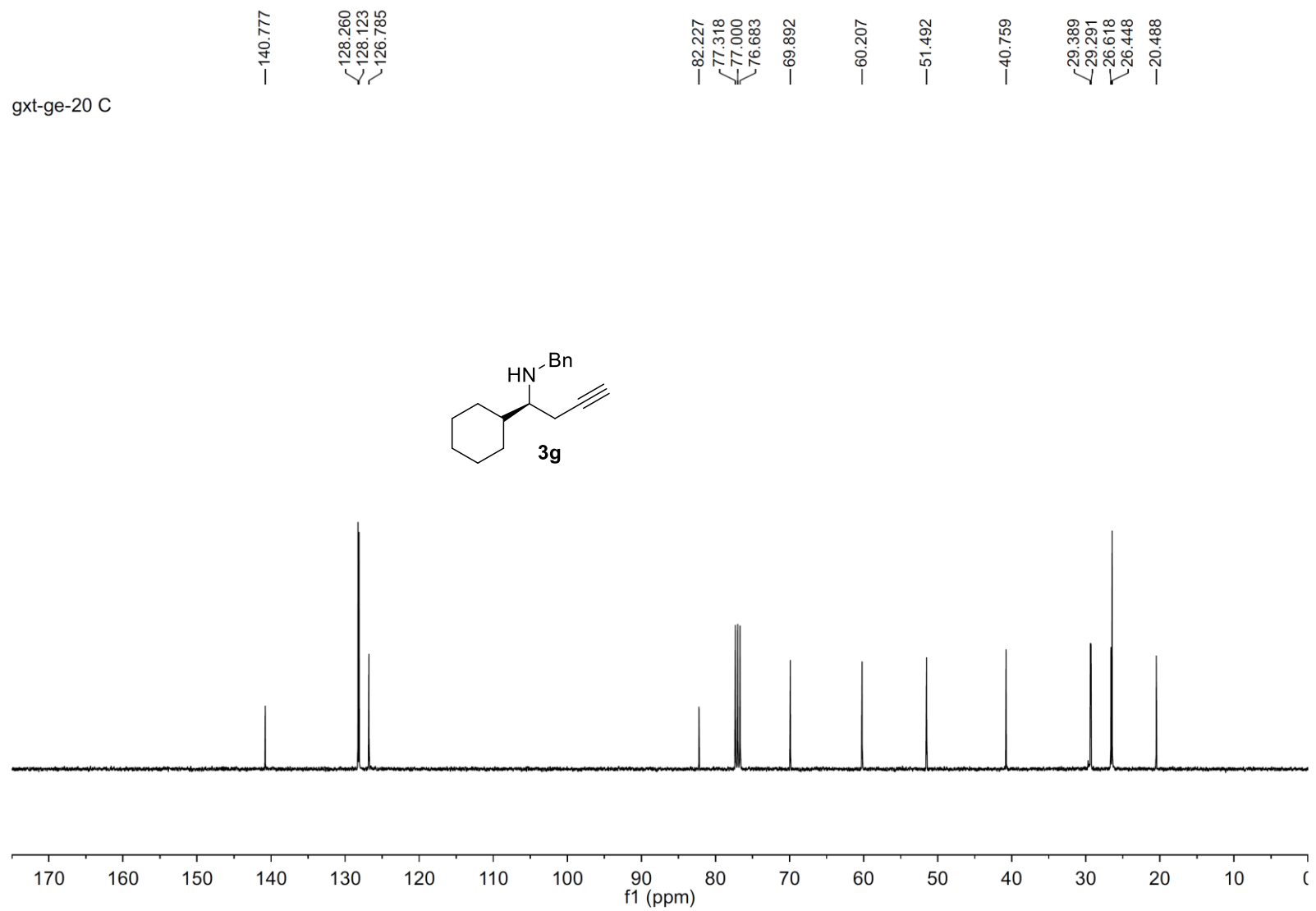


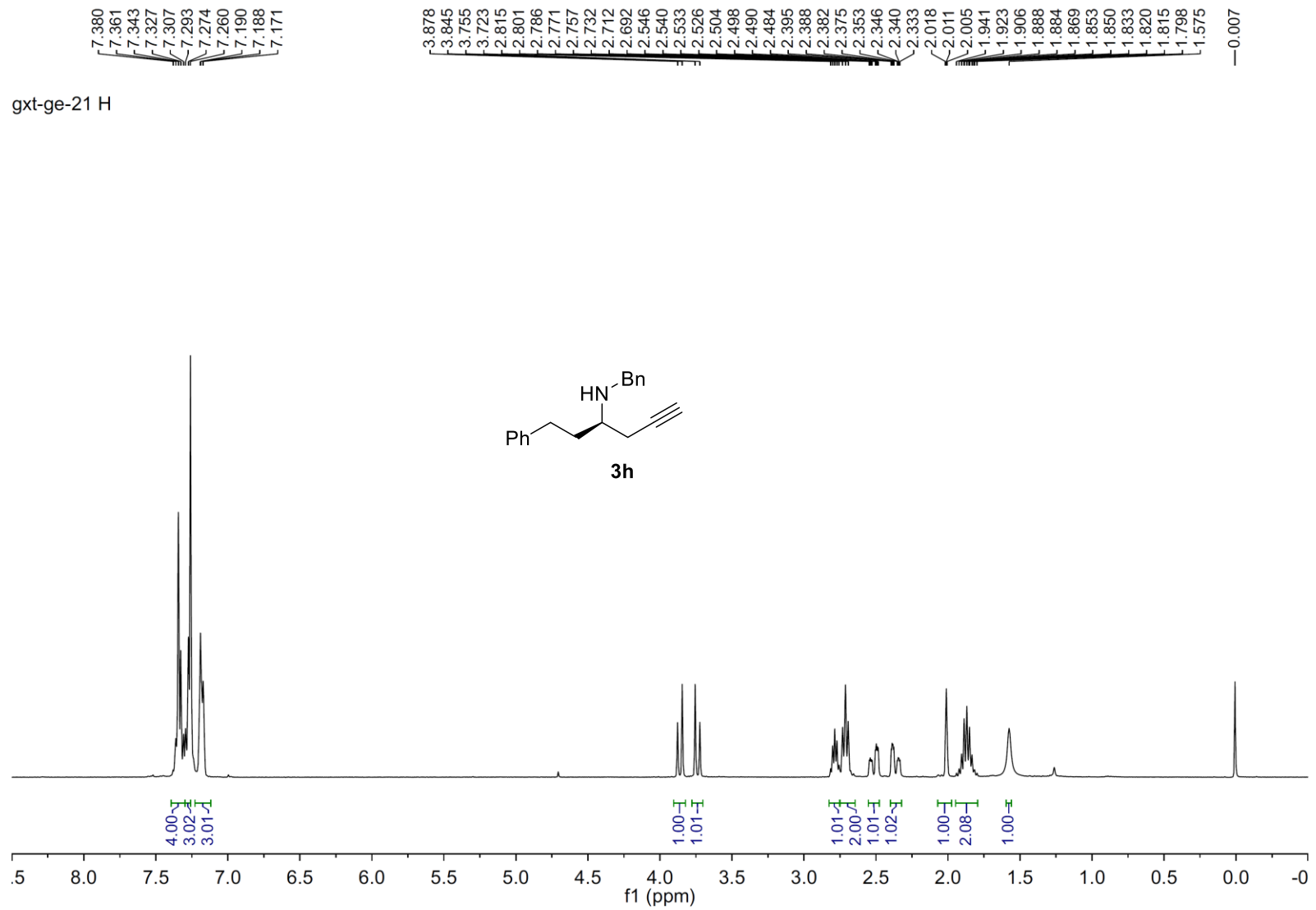


gxt-ge-20 H



gxt-ge-20 C





gxt-ge-21 C

142.090  
140.440

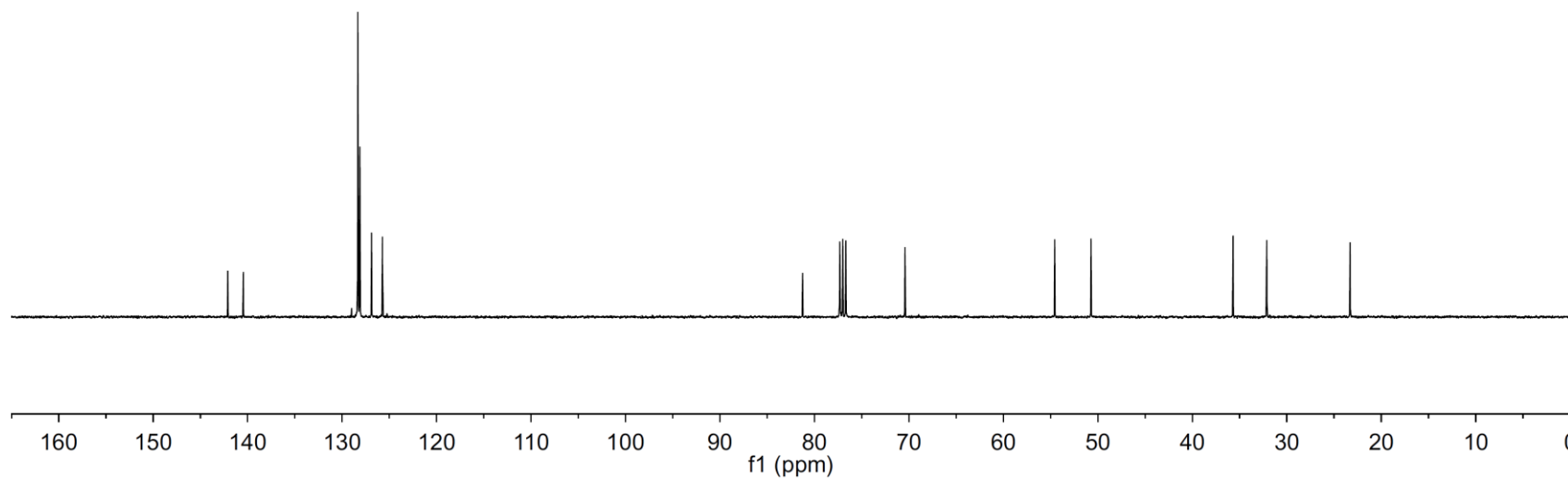
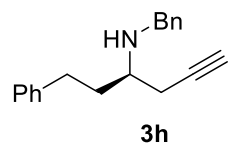
128.335  
128.301  
128.114  
126.881  
125.723

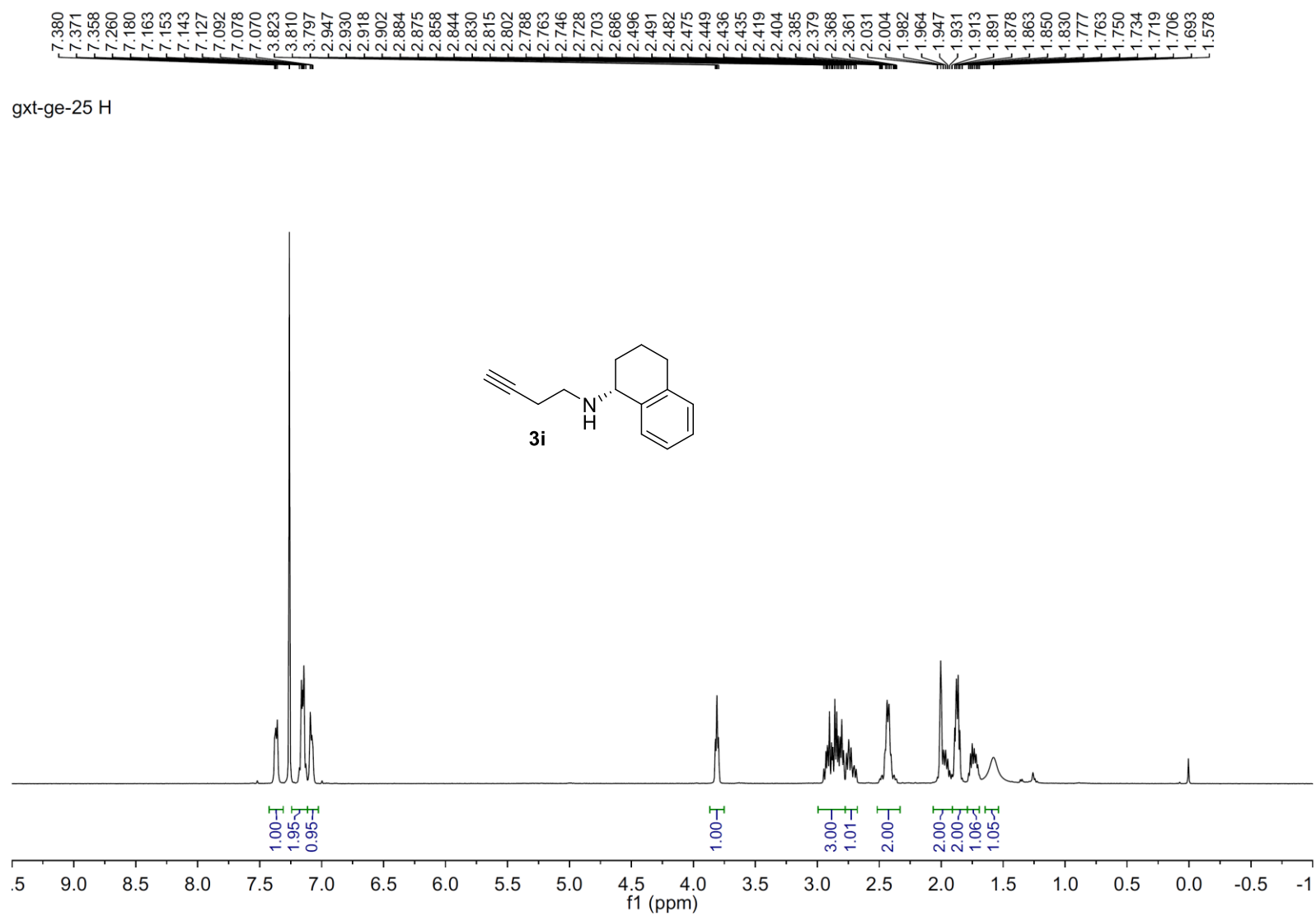
81.251  
77.318  
77.000  
76.683  
70.420

54.574  
50.733

35.697  
32.129

23.311





gxt-ge-25 C

138.903  
137.300

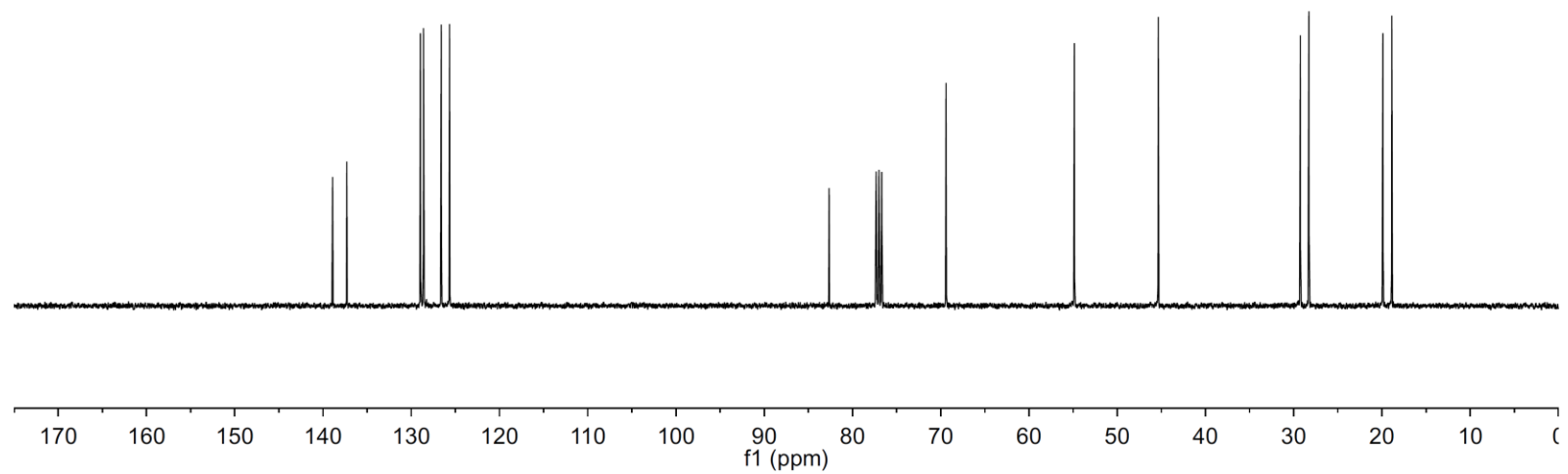
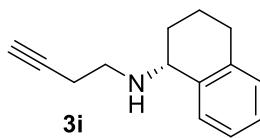
128.964  
128.609  
126.590  
125.653

82.651  
77.318  
77.000  
76.683  
69.395

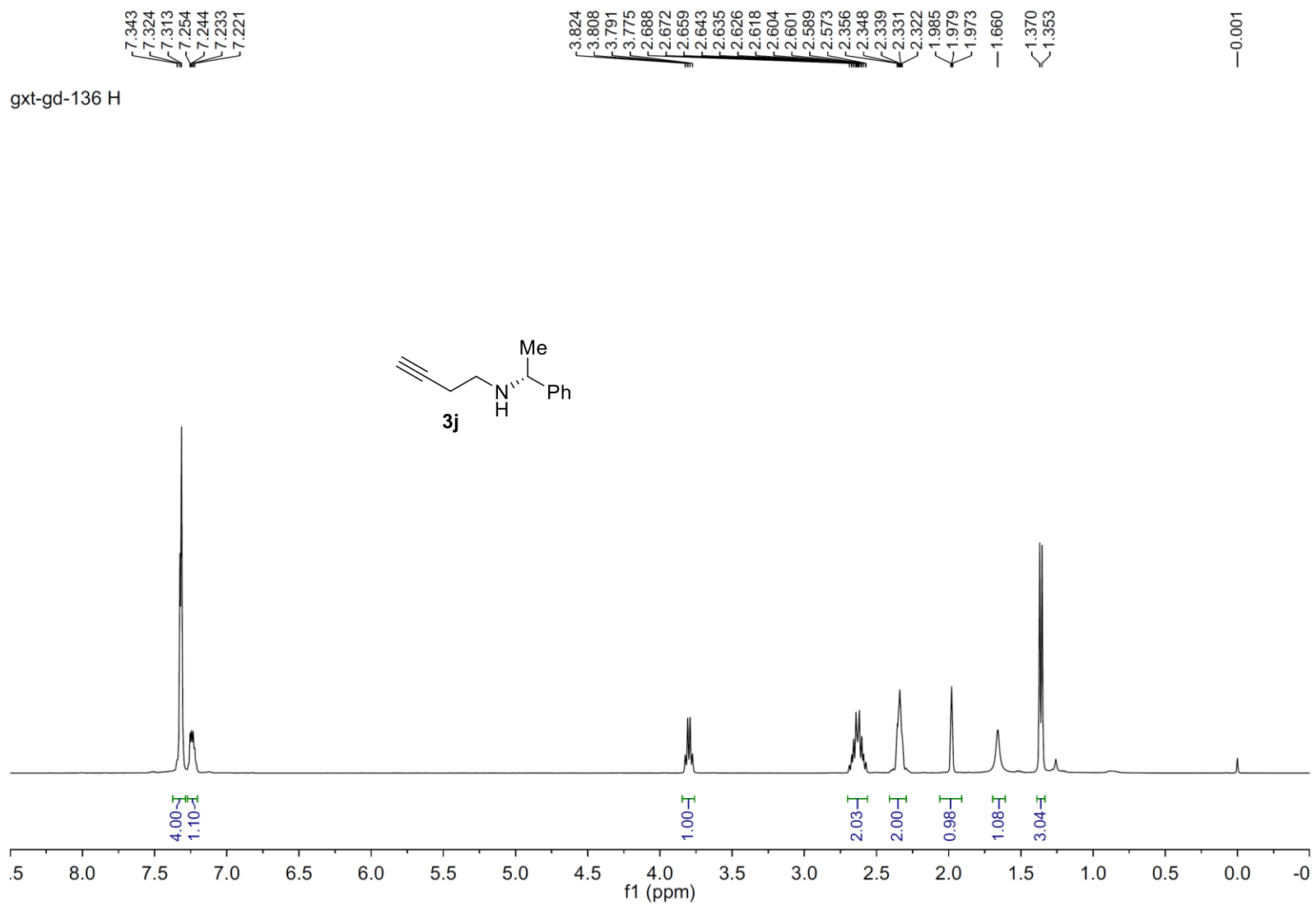
54.859  
45.344

29.252  
28.284

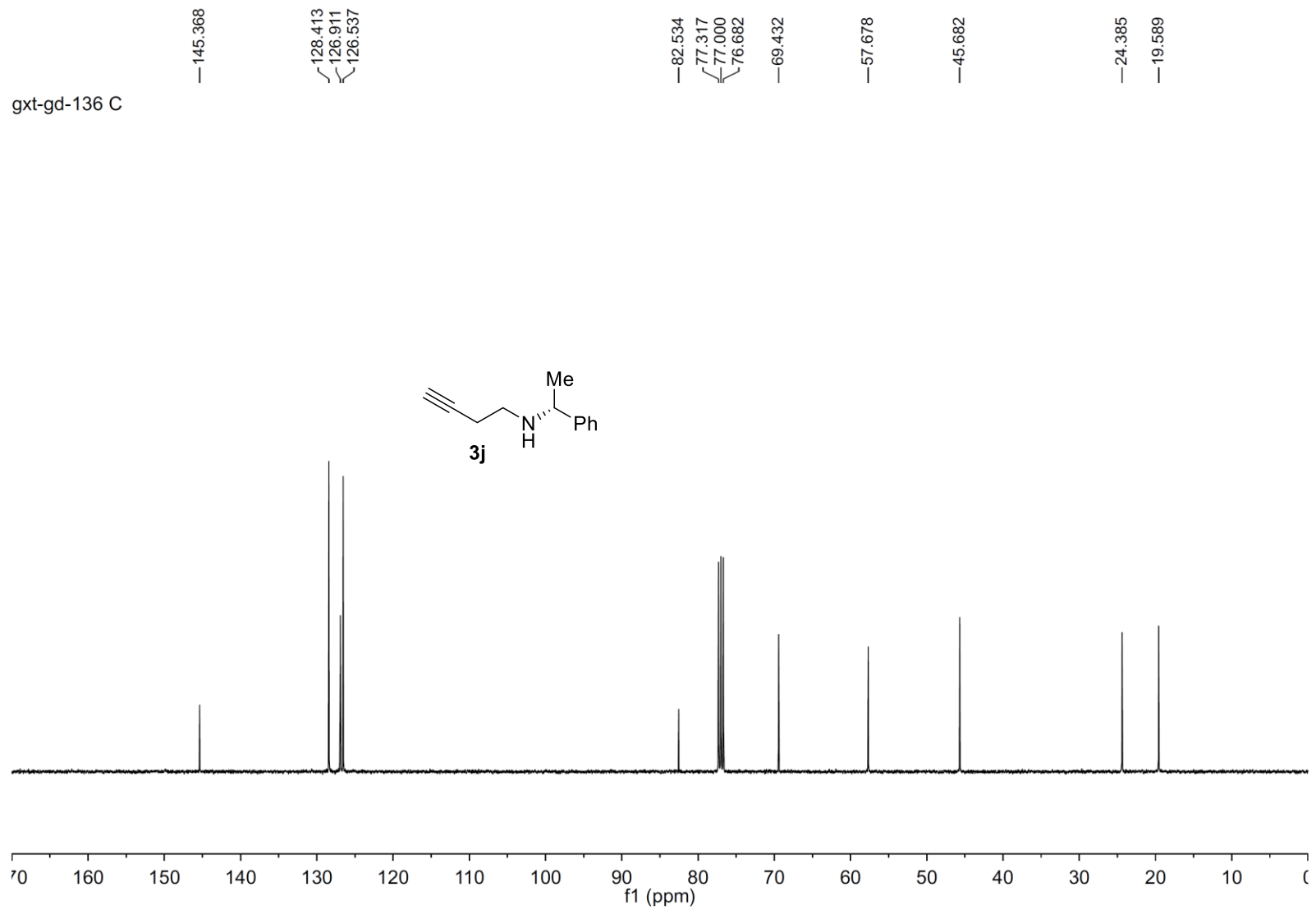
19.893  
18.890



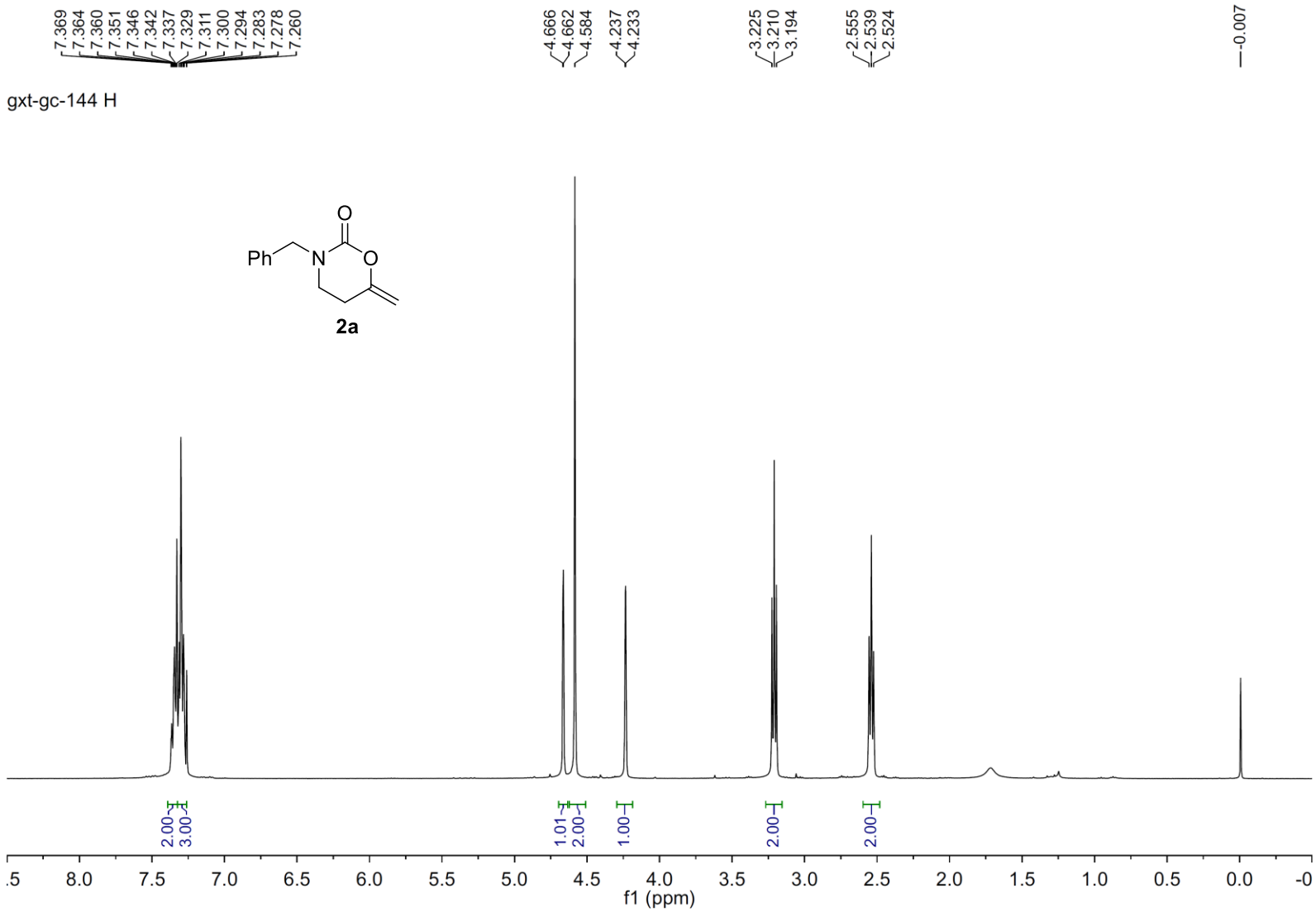
gxt-gd-136 H

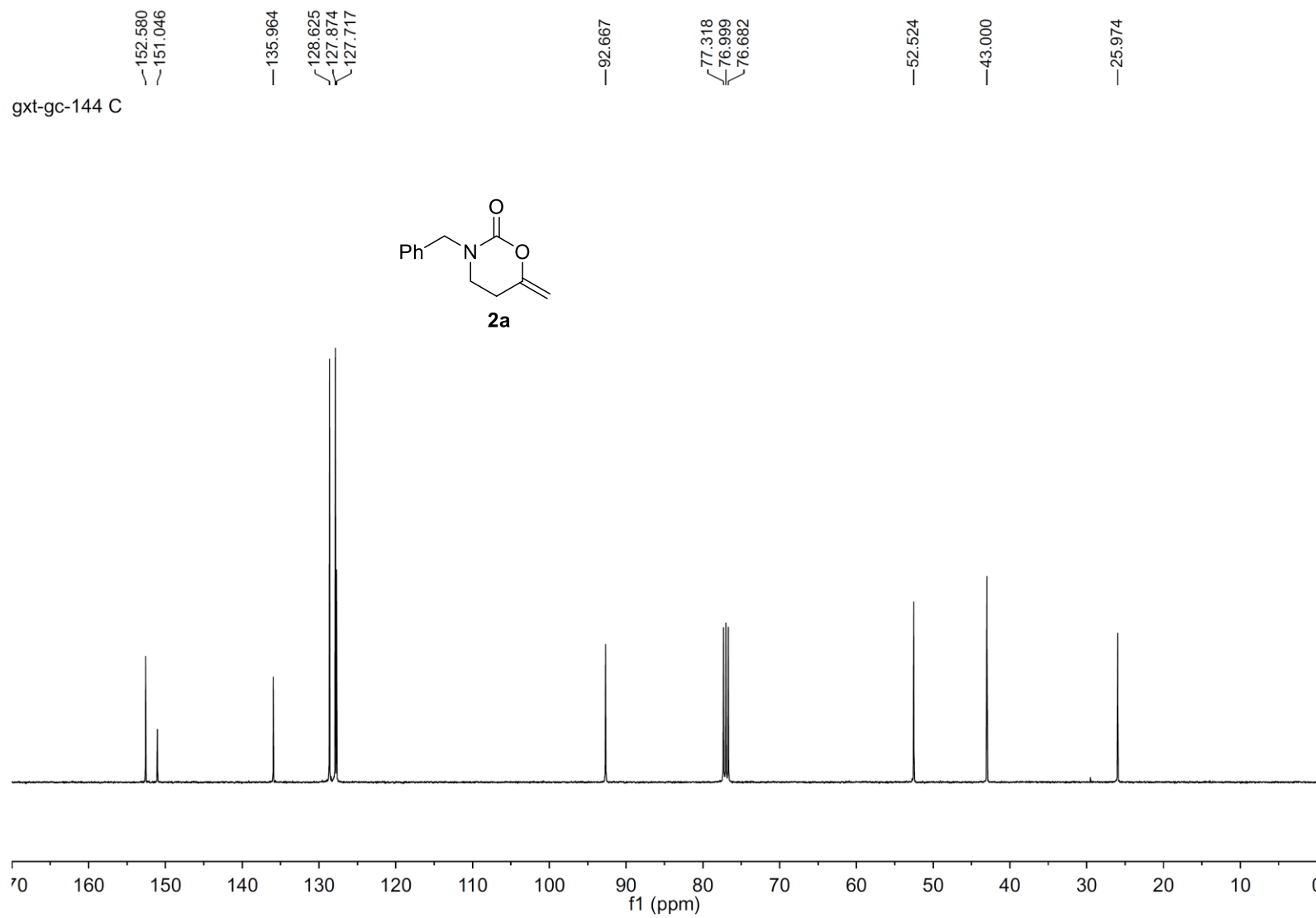


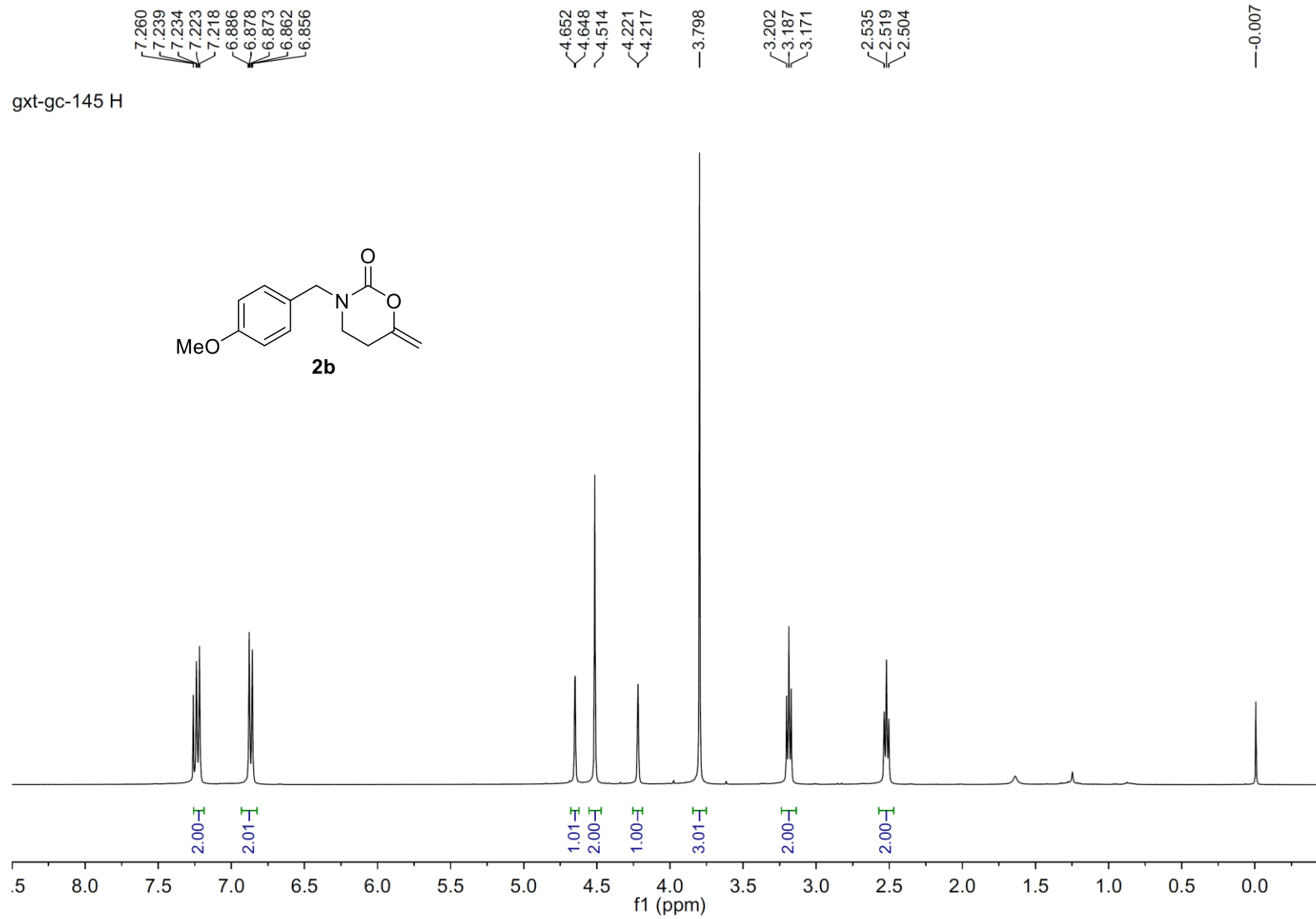
gxt-gd-136 C

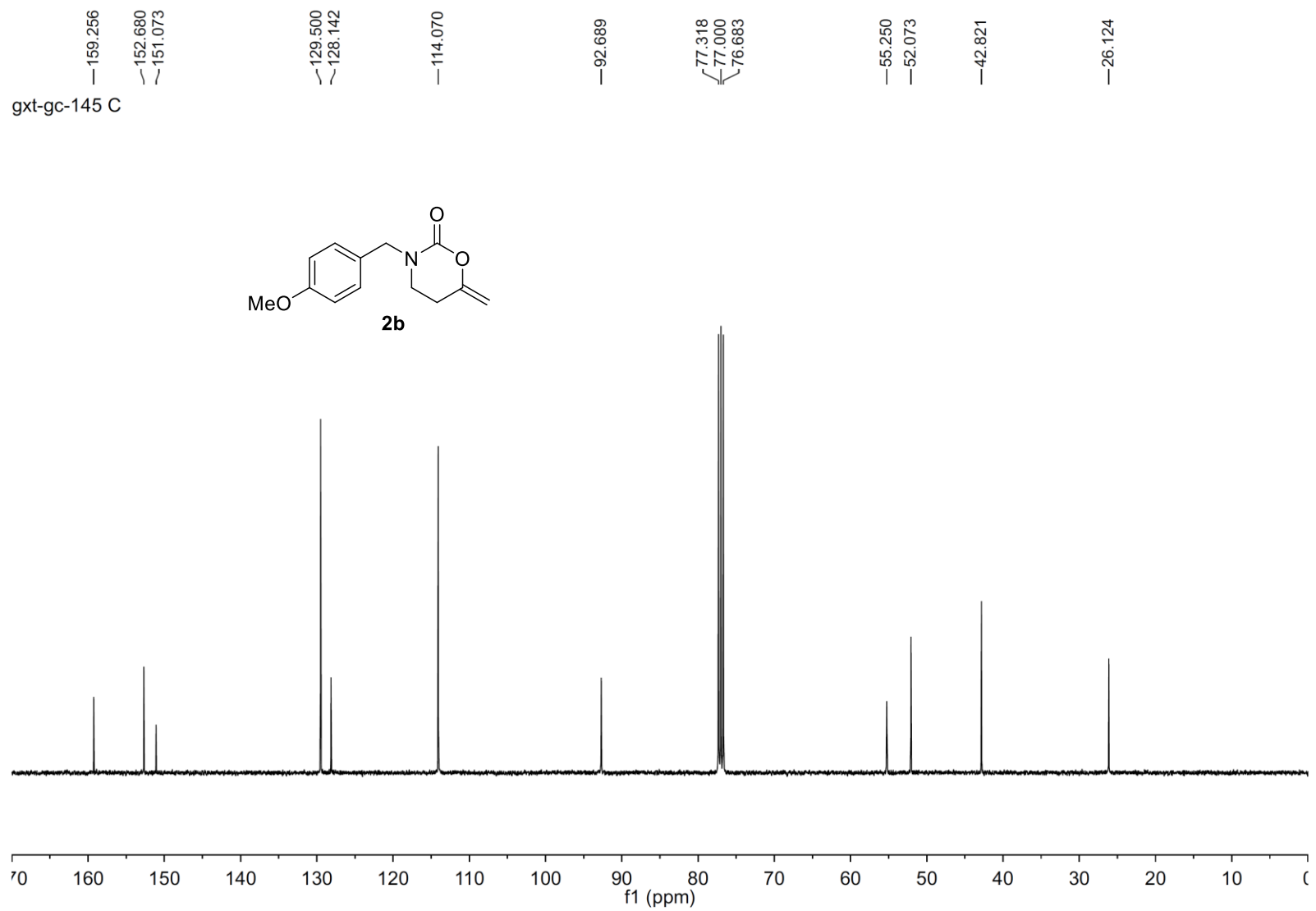


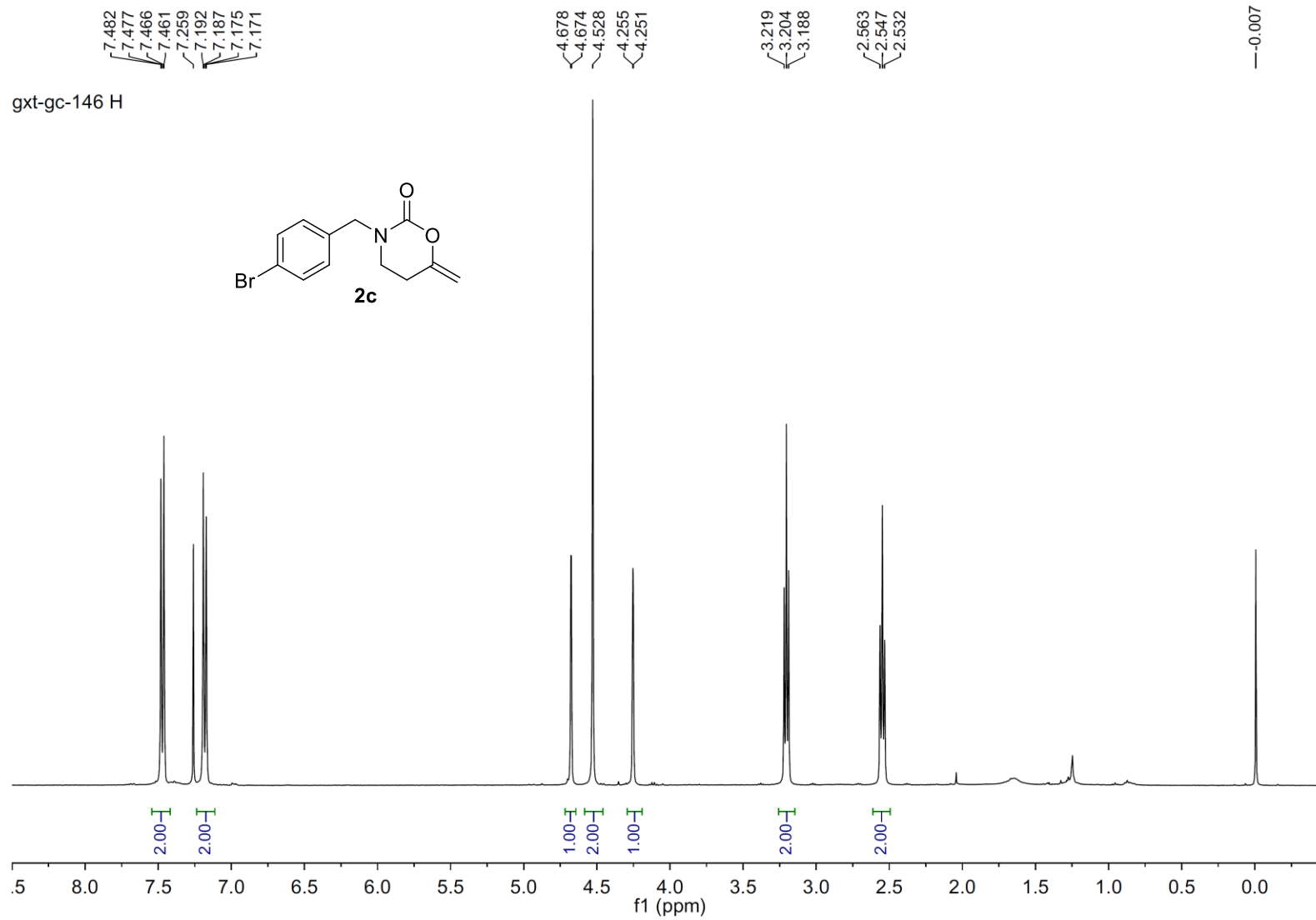


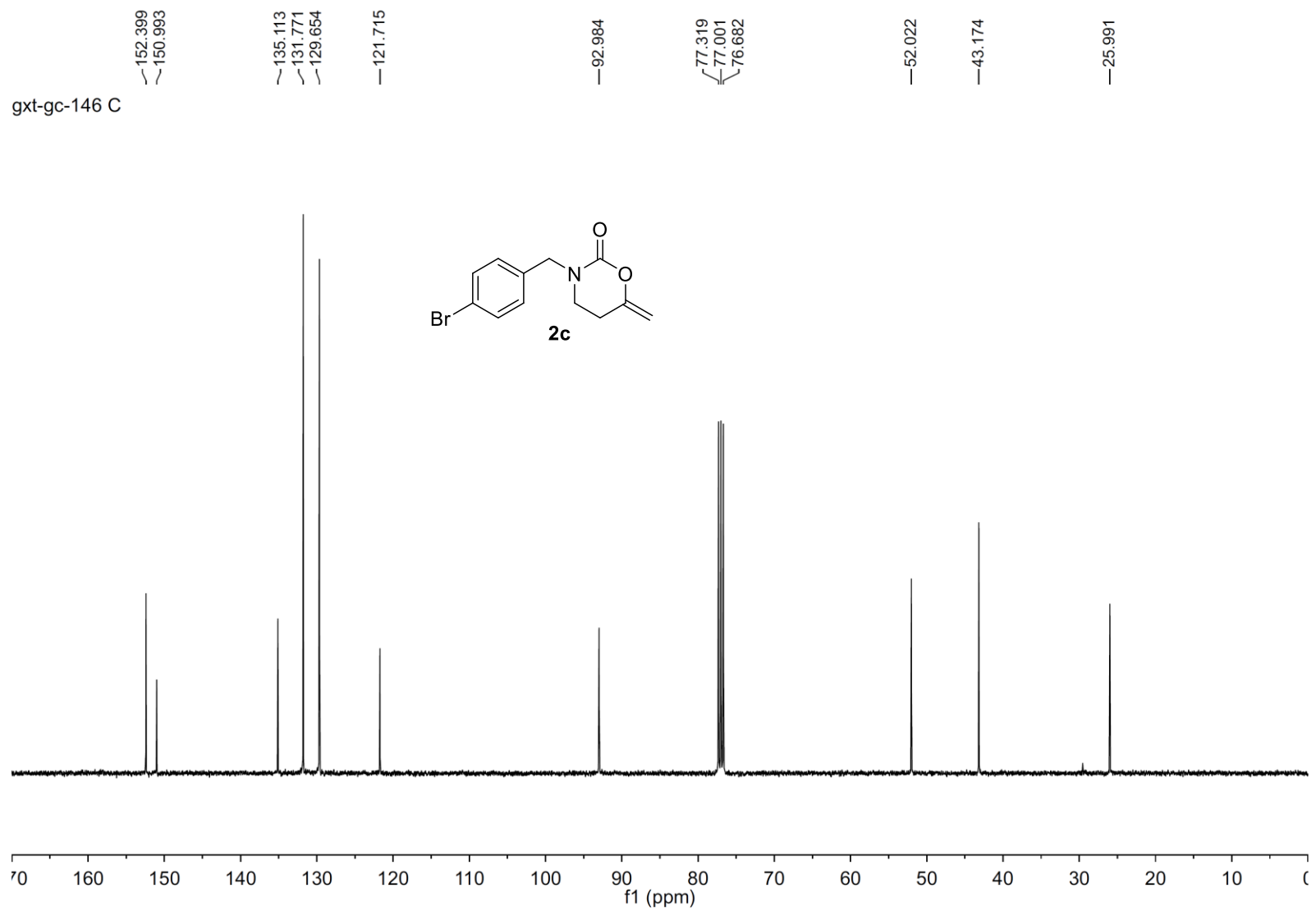












gxt-gd-36 H

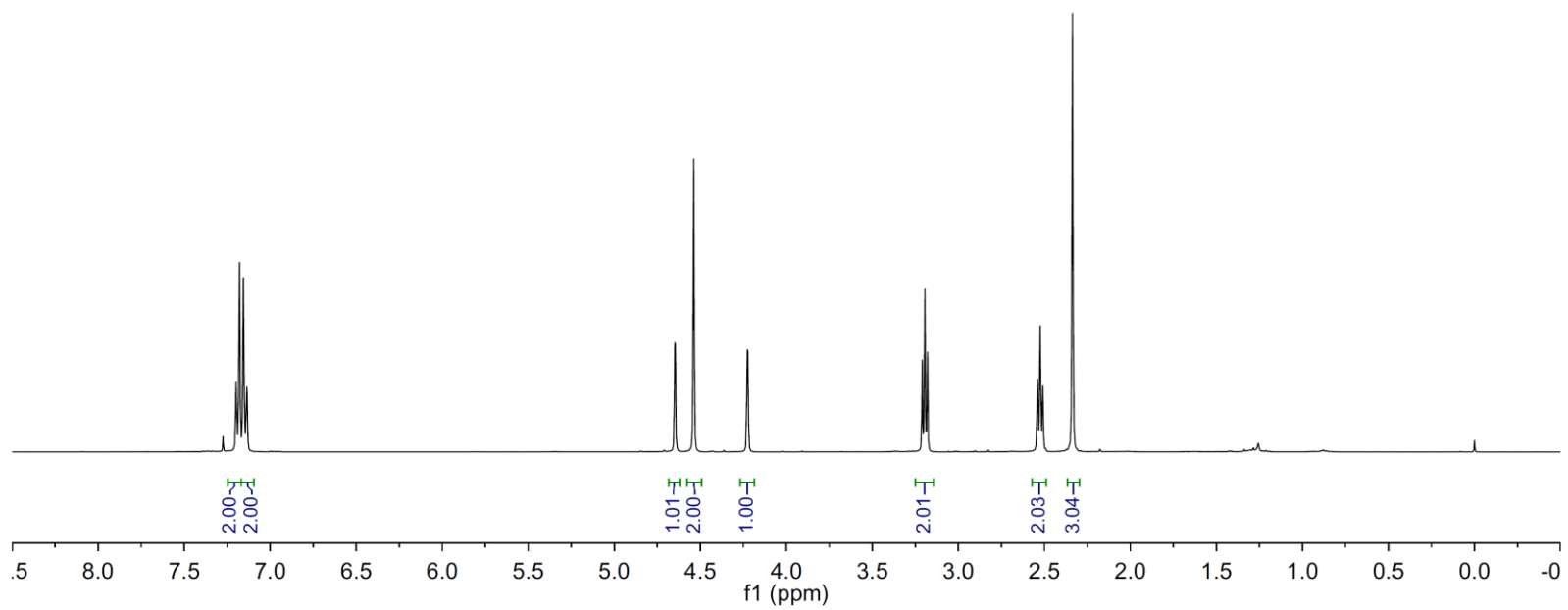
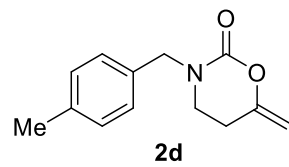
7.274  
7.198  
7.178  
7.156  
7.136

4.648  
4.644  
4.538  
4.227  
4.223

3.210  
3.194  
3.179

2.540  
2.524  
2.509  
2.336

0.000



gxt-gd-36 C

152.668  
151.043

137.491  
132.965  
129.318  
127.993

92.584

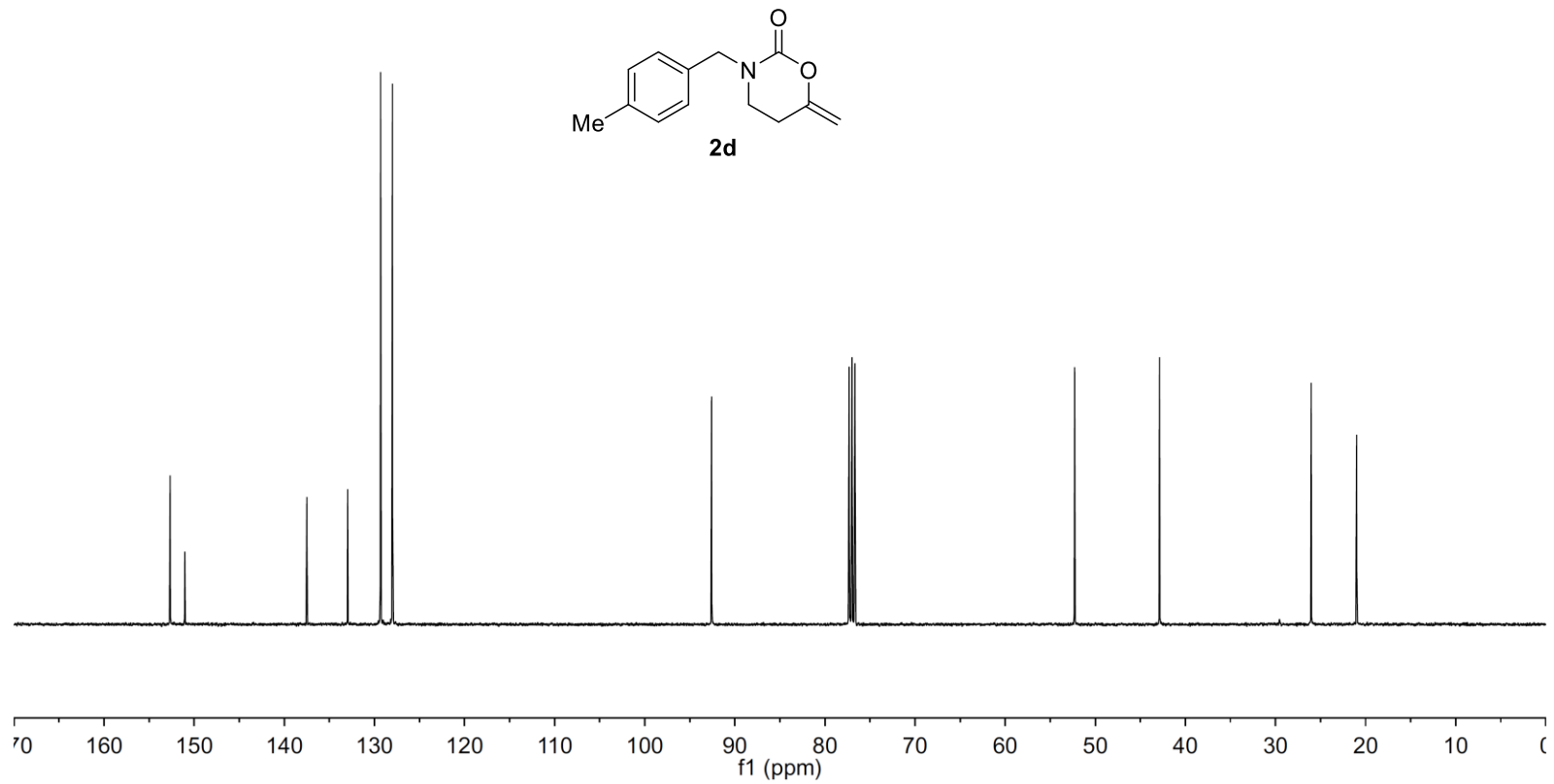
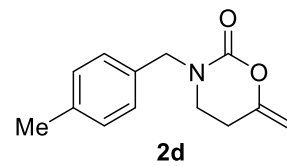
77.319  
77.001  
76.683

52.293

42.874

26.048

20.999





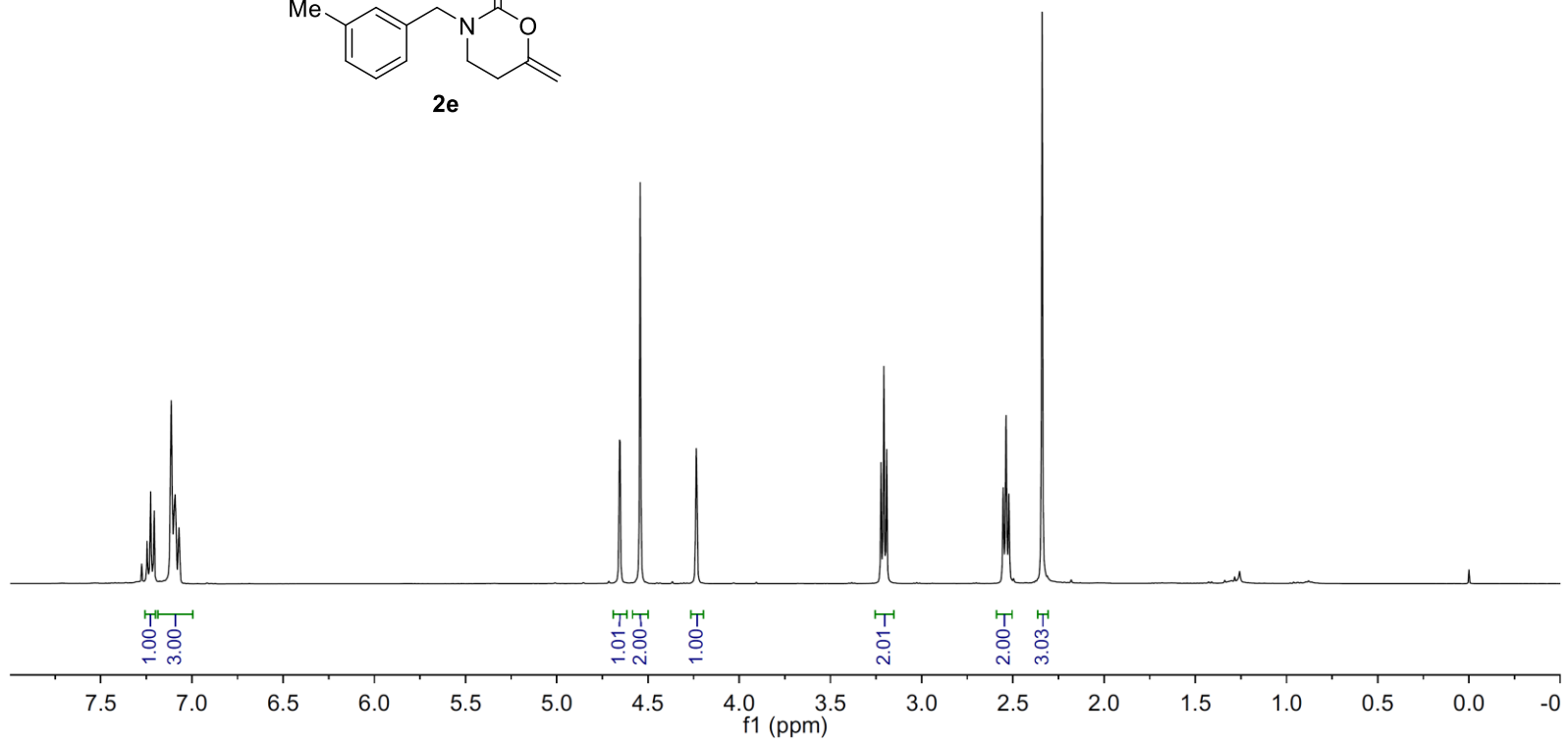
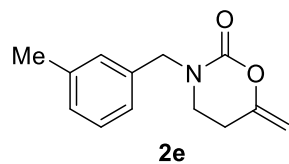
7.275  
7.246  
7.226  
7.213  
7.206  
7.115  
7.092  
7.071

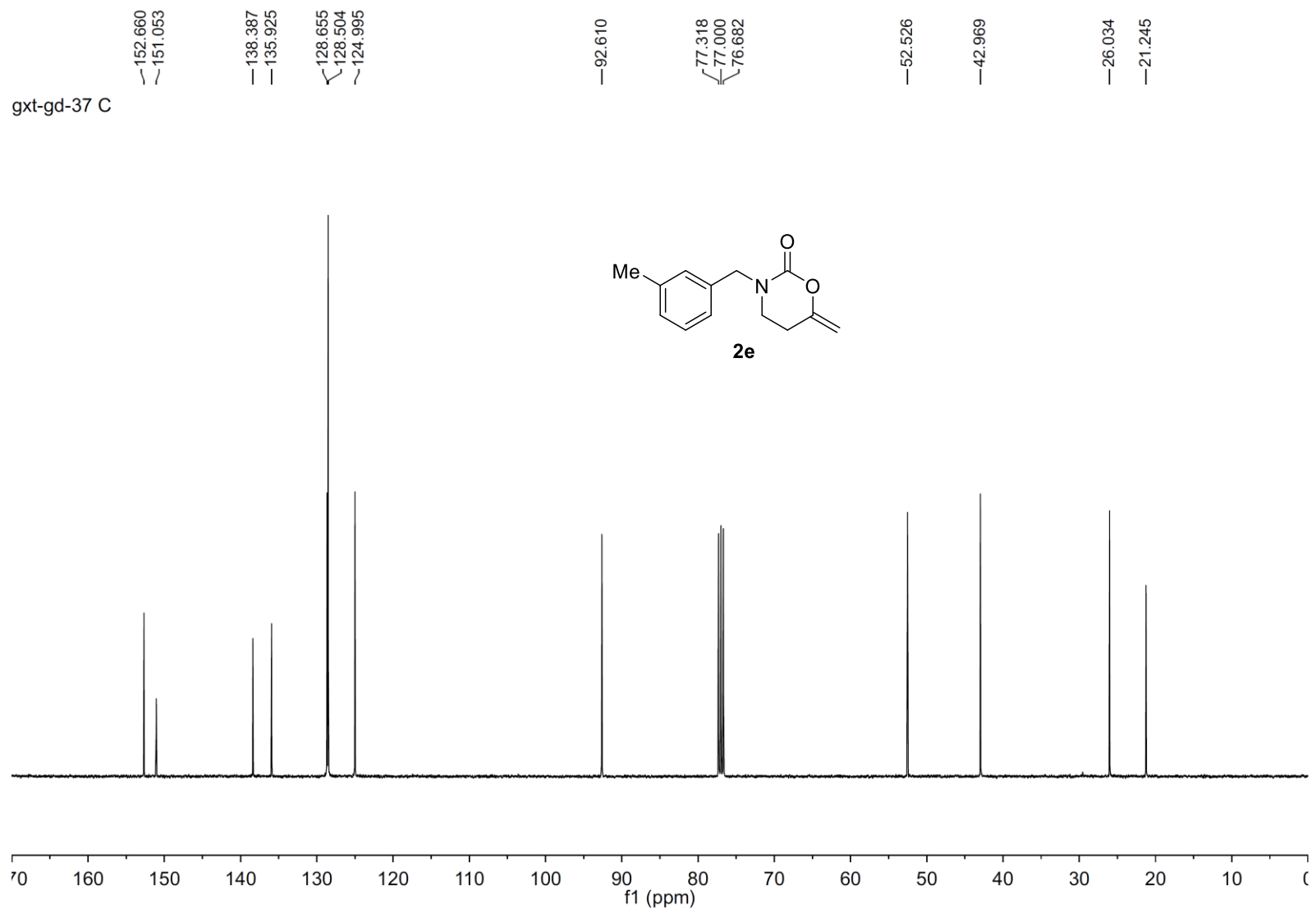
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4.653  
4.543  
4.236  
4.232

3.223  
3.207  
3.191

2.553  
2.537  
2.522  
2.340

gxt-gd-37 H





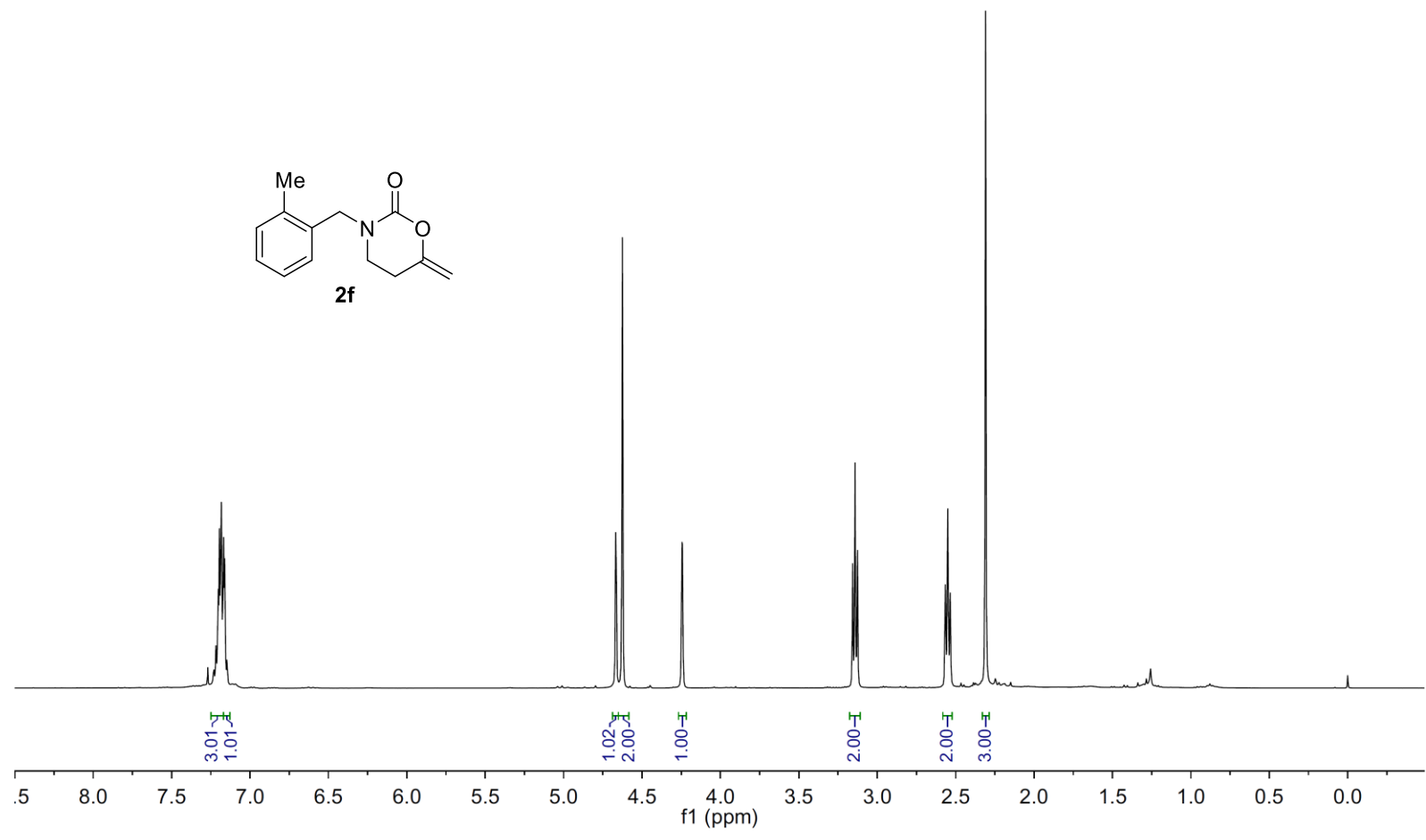
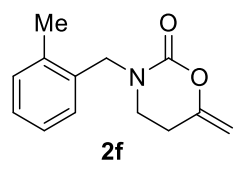
7.234  
7.229  
7.218  
7.209  
7.201  
7.195  
7.182  
7.168  
7.162  
7.148  
7.143

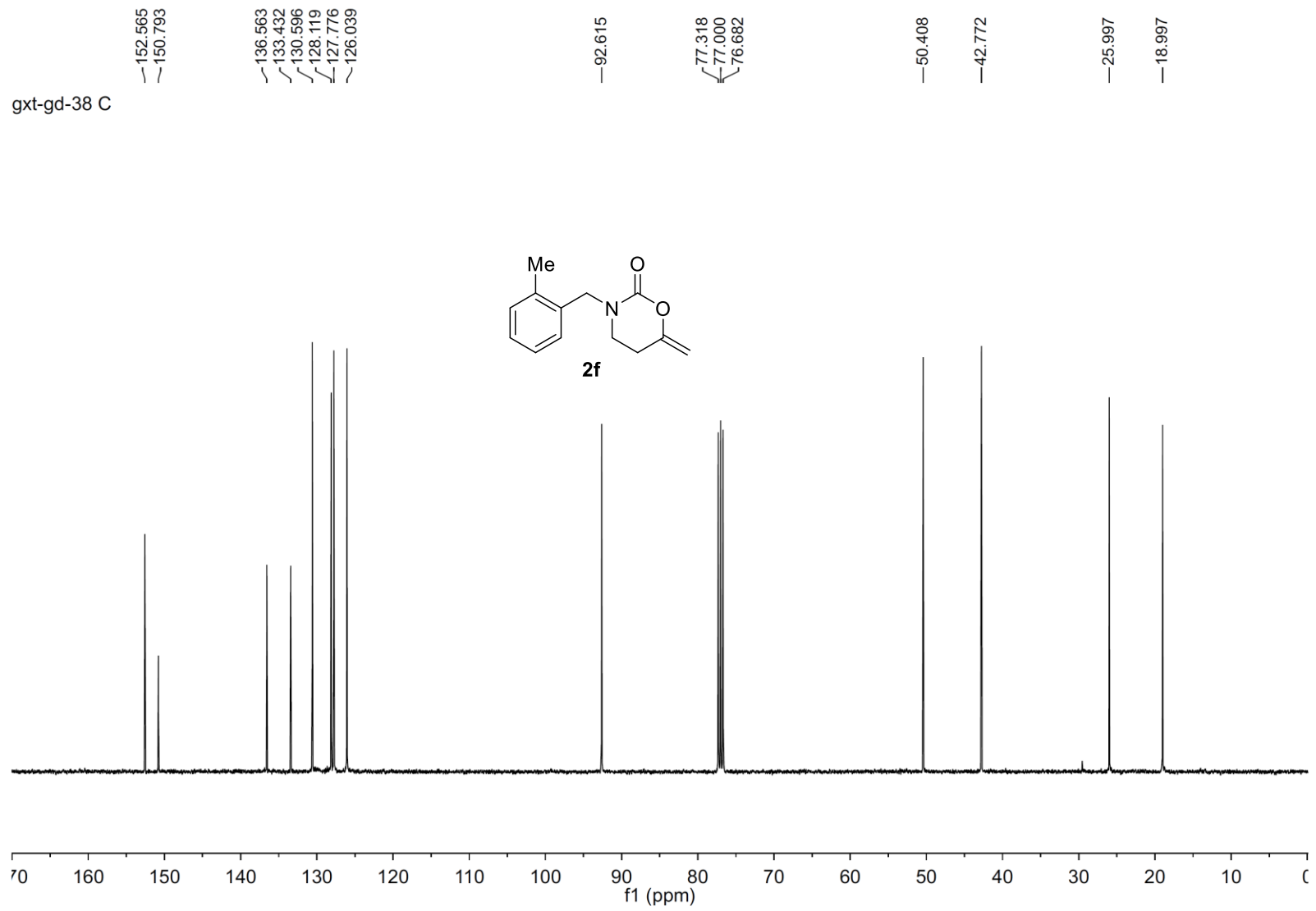
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4.664  
4.625  
4.246  
4.242

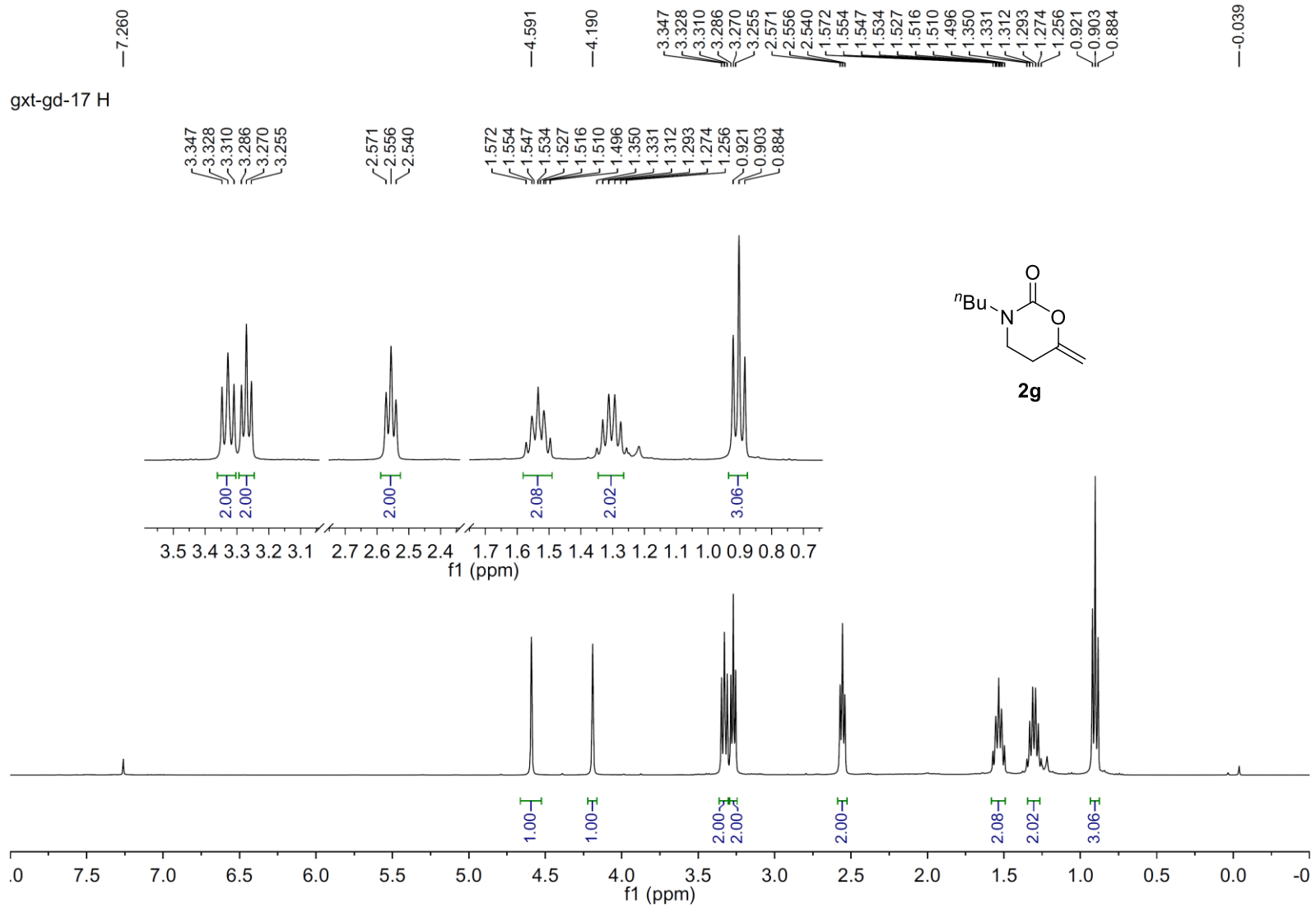
3.157  
3.141  
3.126

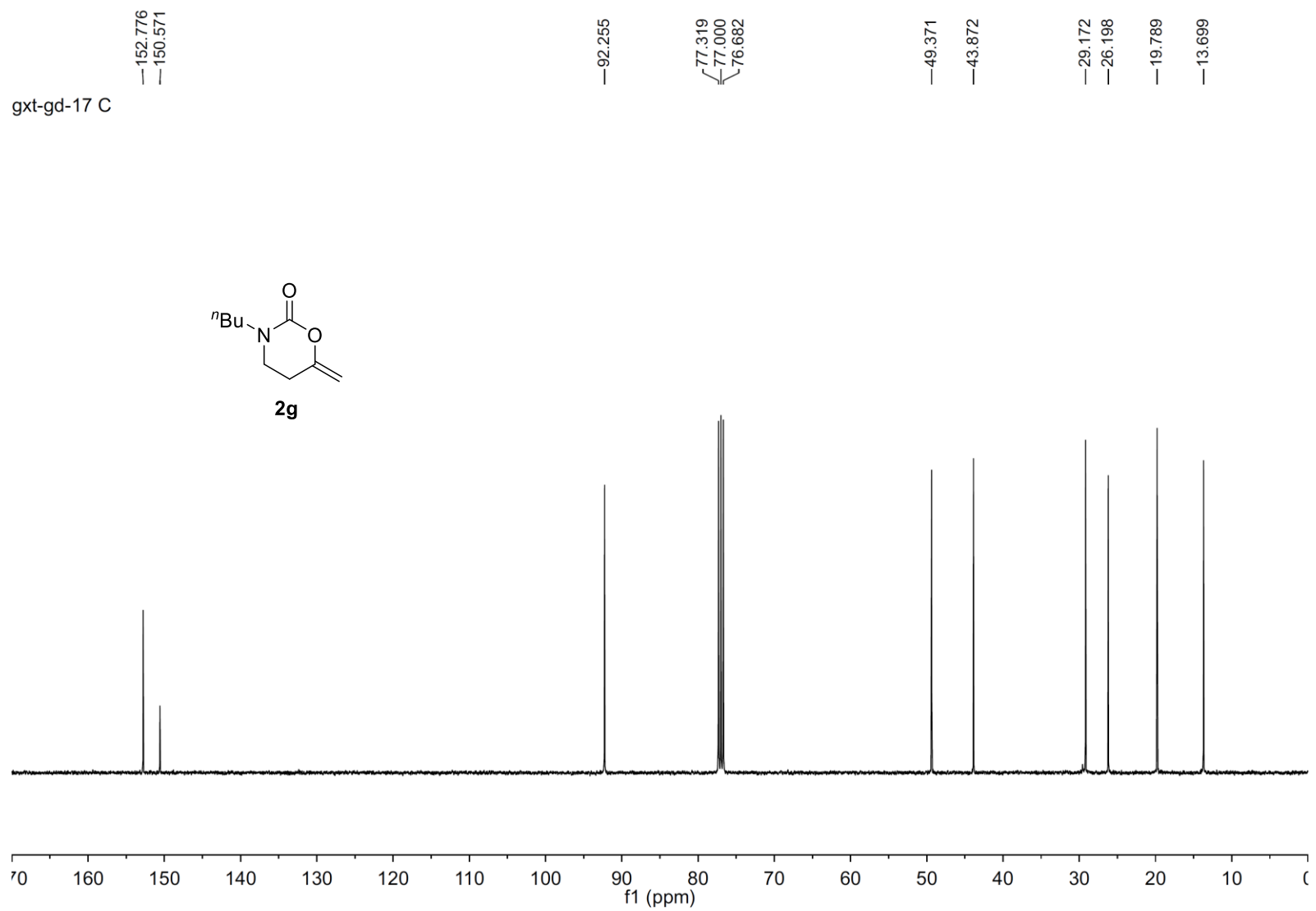
2.566  
2.550  
2.535  
2.309

gxt-gd-38 H









7.298  
7.278  
7.270  
7.260  
7.220  
7.200  
7.195  
7.183  
7.175

4.627  
4.623  
4.218  
4.213

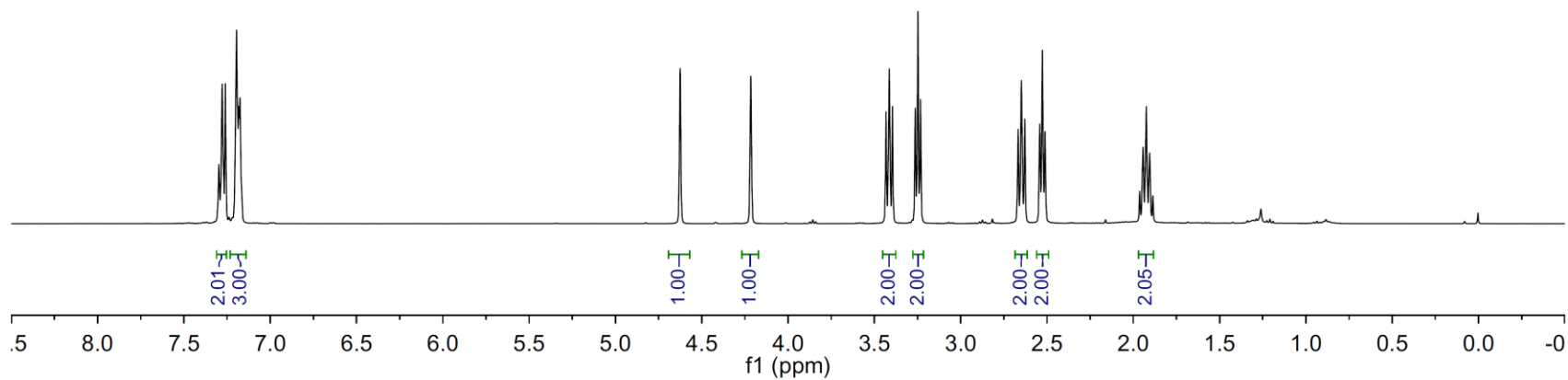
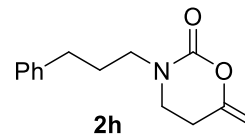
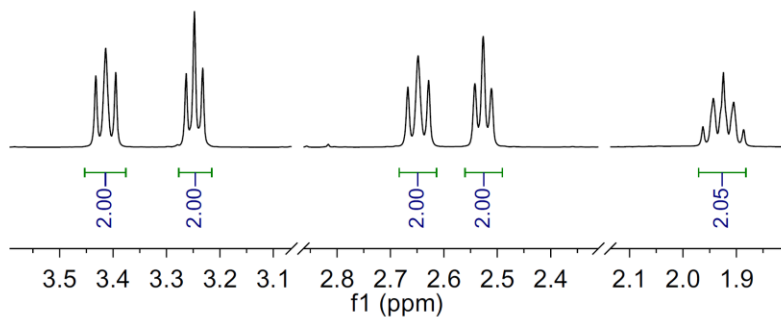
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3.414  
3.395  
3.263  
3.248  
3.232  
2.668  
2.648  
2.629  
2.542  
2.526  
2.511  
1.963  
1.947  
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1.940  
1.930  
1.924  
1.909  
1.905  
1.886

gxt-gd-19 H

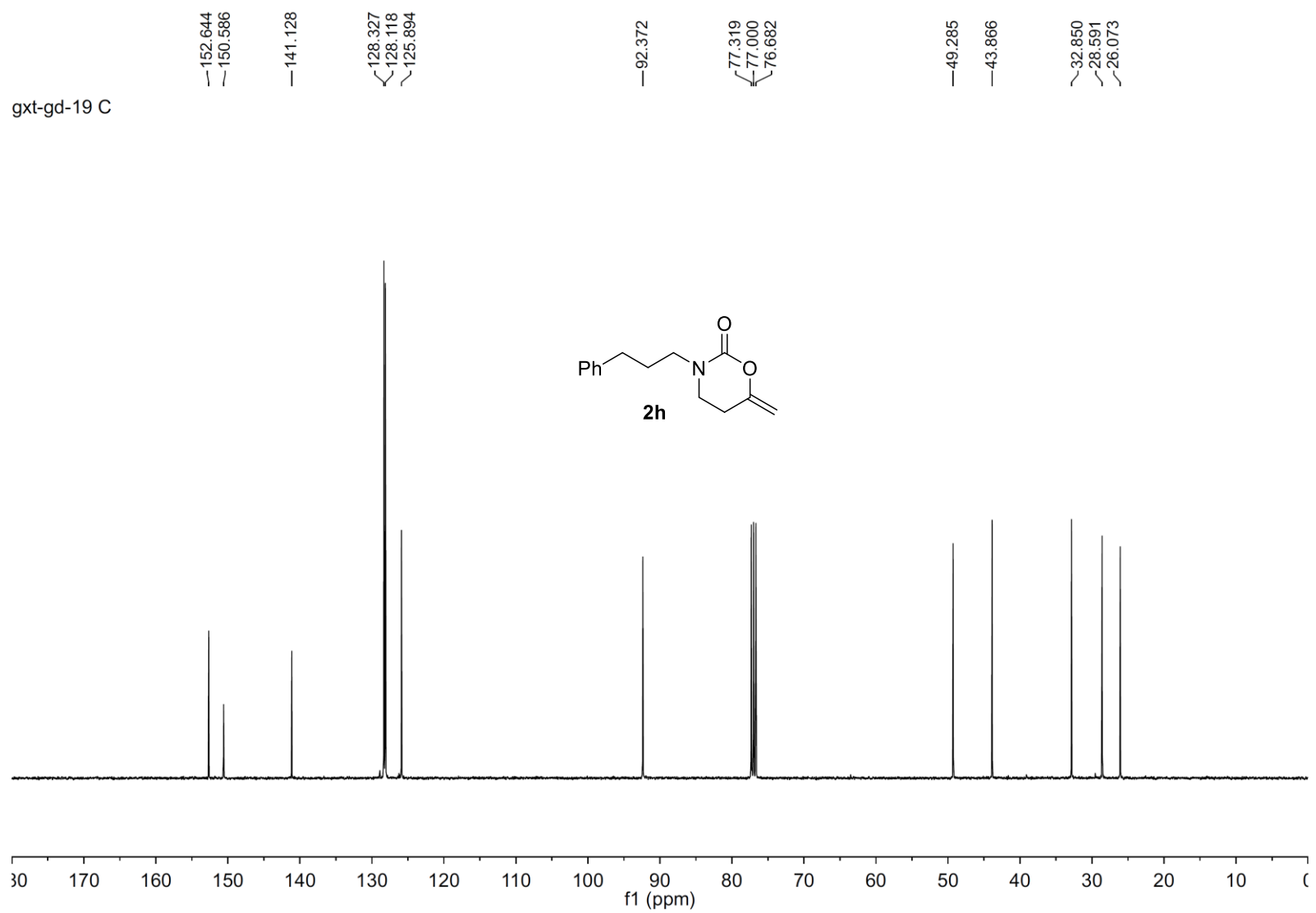
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3.414  
3.395  
3.263  
3.248  
3.232

2.668  
2.648  
2.629  
2.542  
2.526  
2.511

1.947  
1.943  
1.940  
1.930  
1.924  
1.909  
1.905

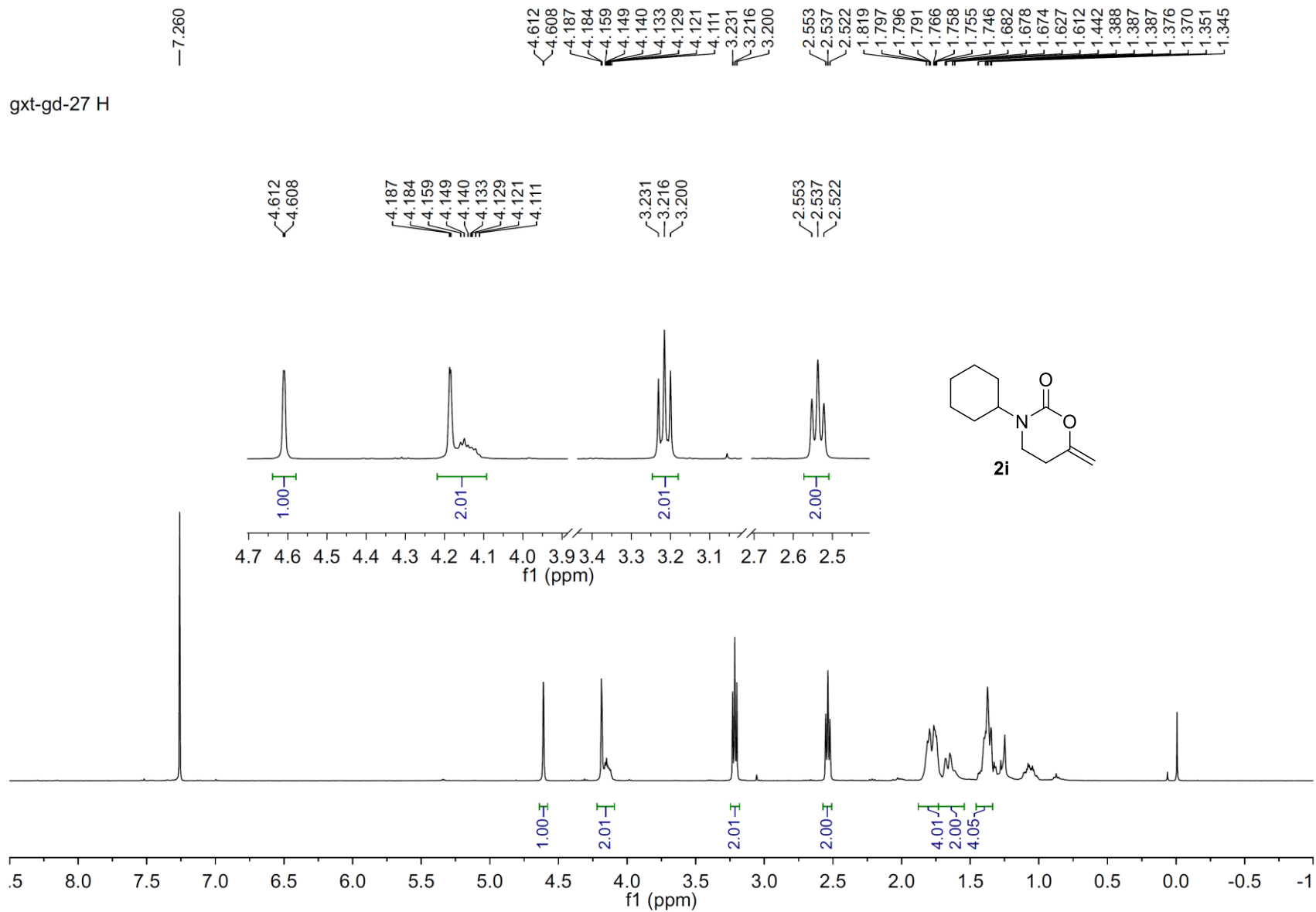


gxt-gd-19 C

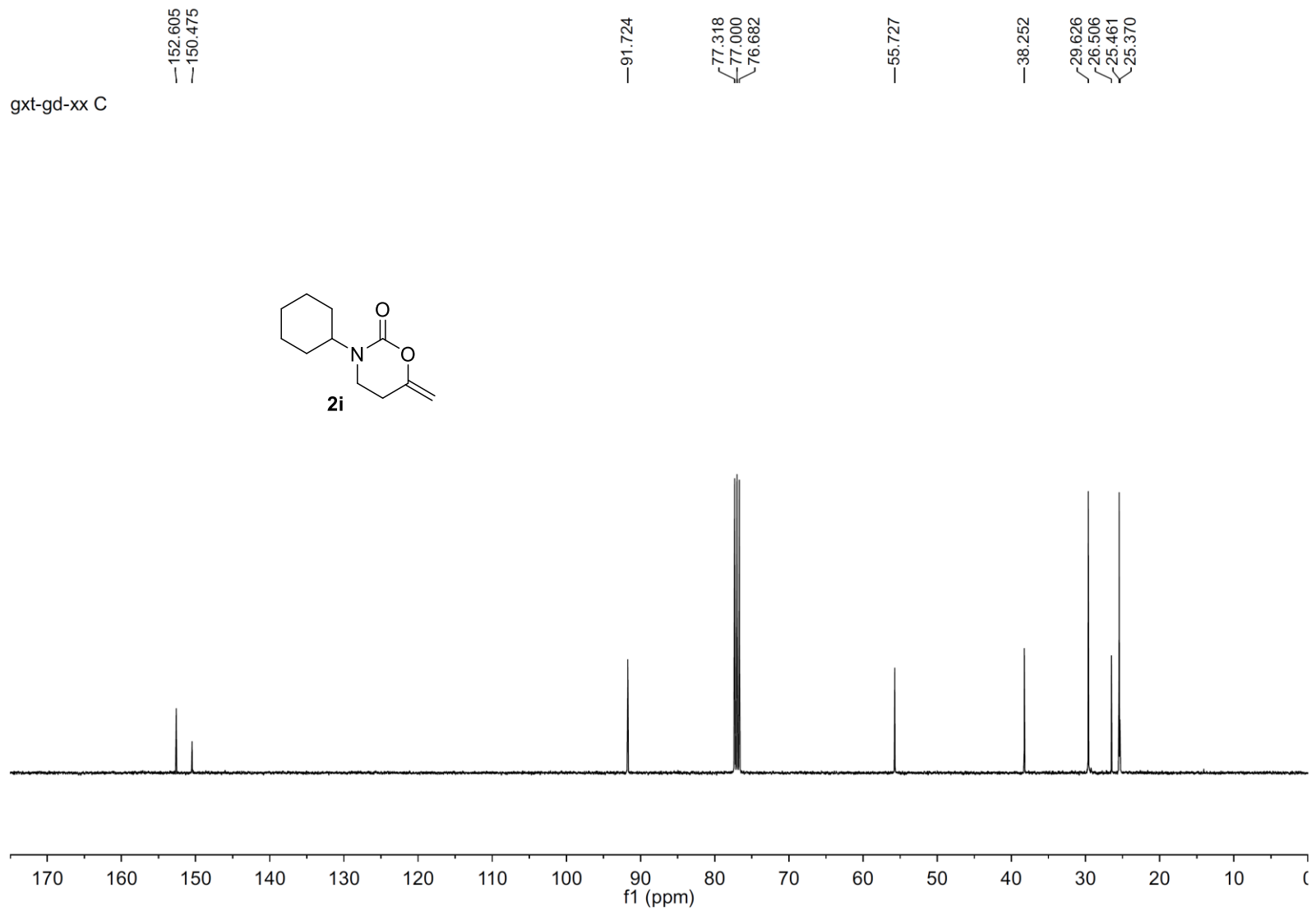
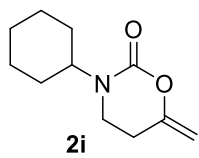




gxt-gd-27 H



gxt-gd-xx C



7.370  
7.365  
7.360  
7.352  
7.347  
7.342  
7.337  
7.331  
7.322  
7.318  
7.307  
7.302  
7.291  
7.287  
7.283  
7.260

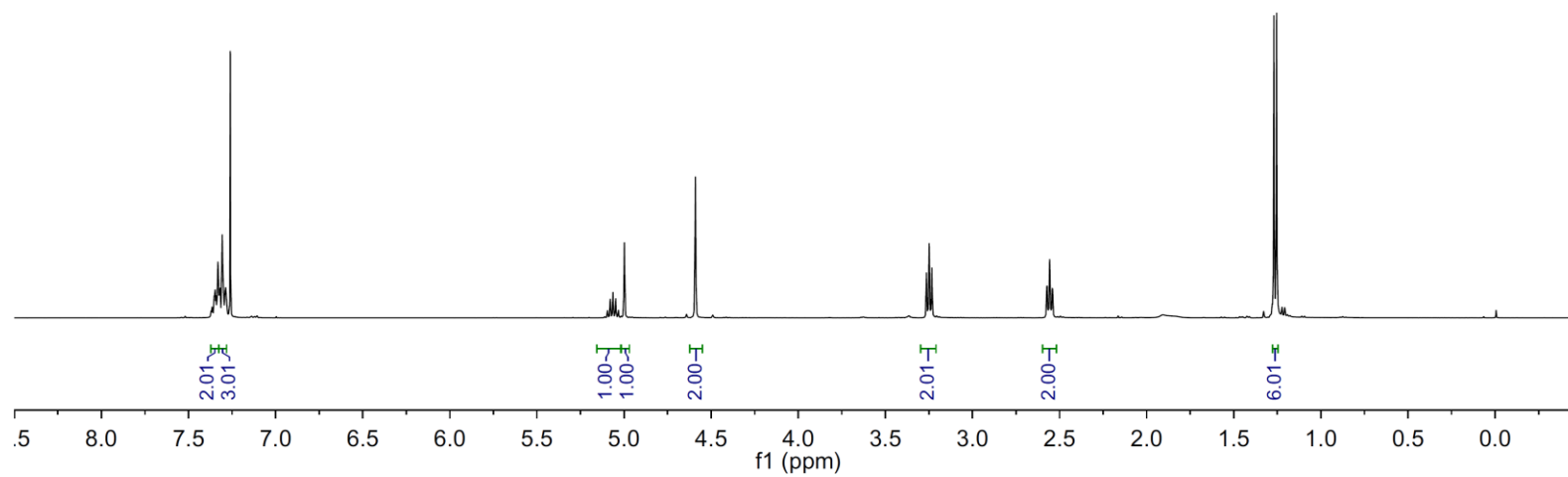
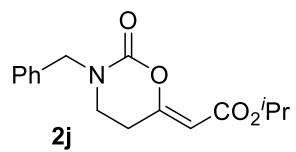
5.111  
5.095  
5.081  
5.079  
5.066  
5.063  
5.050  
5.048  
5.032  
4.998  
4.590

3.264  
3.249  
3.233

2.574  
2.571  
2.557  
2.555  
2.542  
2.540

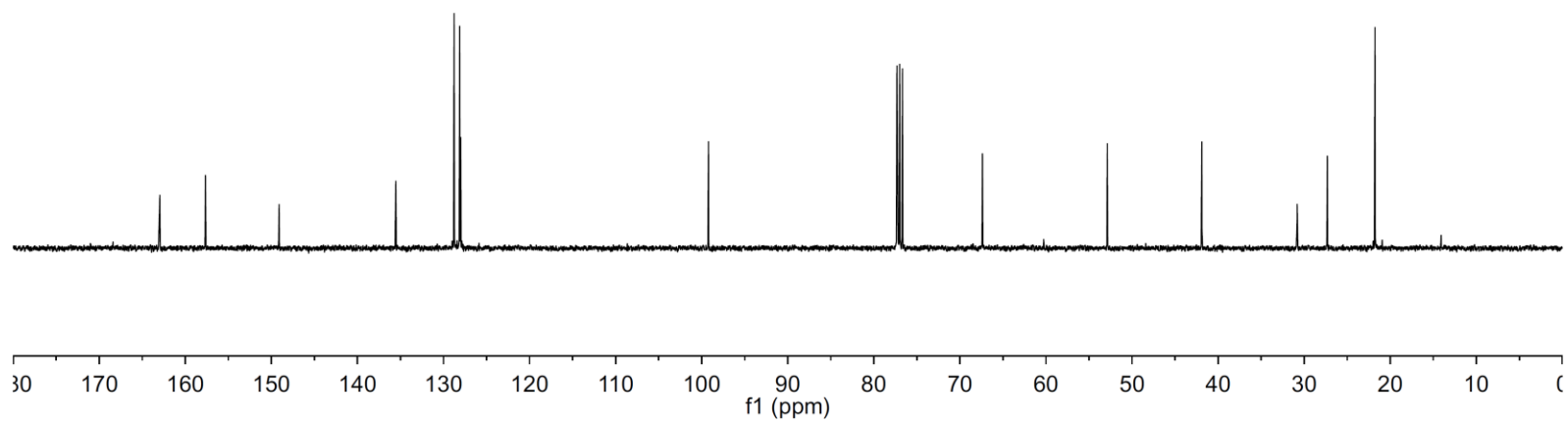
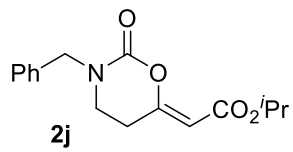
1.270  
1.254

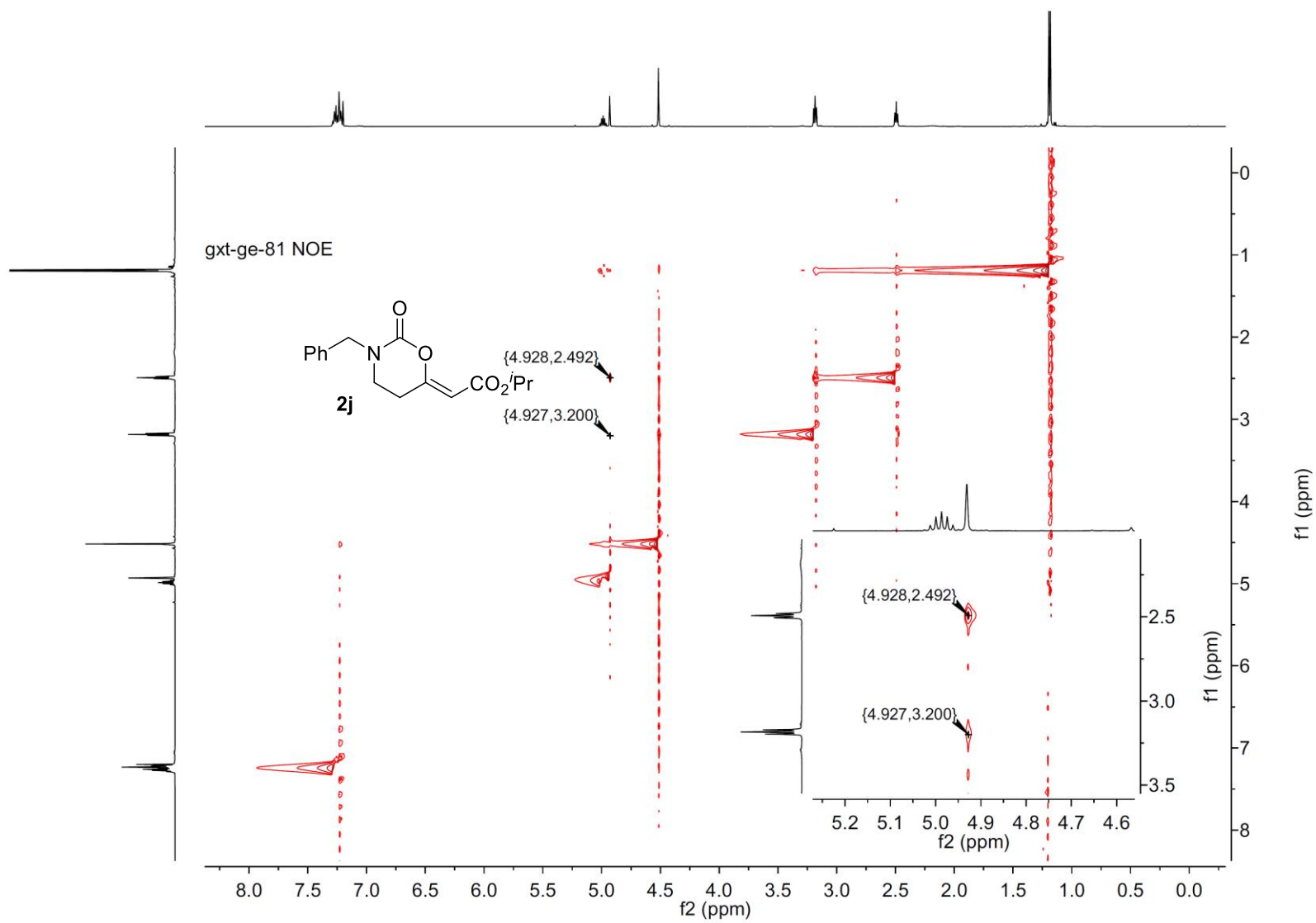
gxt-ge-81 H



gxt-ge-81 C

—162.952  
—157.652  
—149.101  
—135.537  
—128.761  
—128.113  
—127.996  
—99.205  
—77.318  
—77.001  
—76.681  
—67.378  
—52.877  
—41.893  
—30.810  
—27.309  
—21.772





v

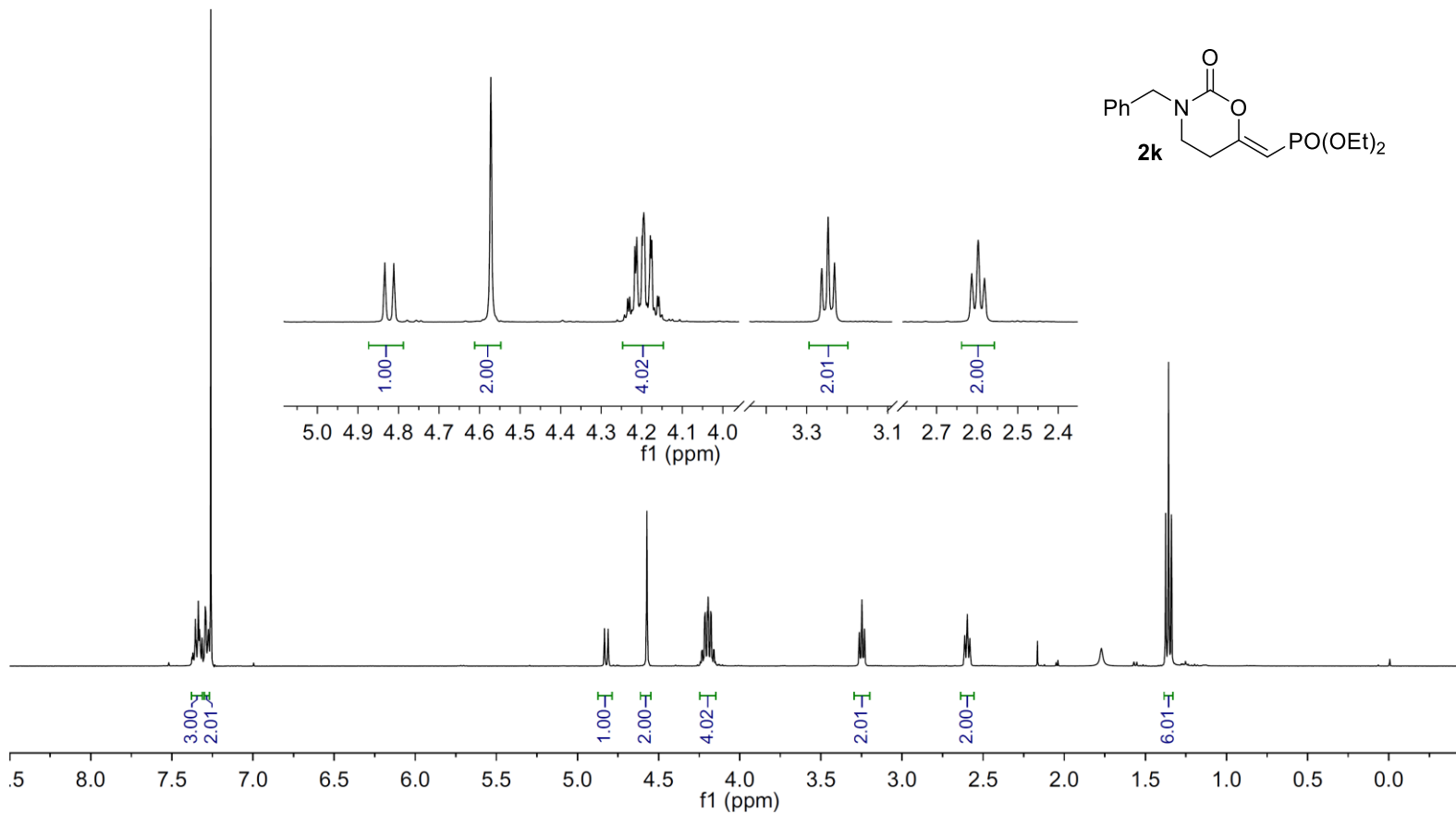
7.377  
7.371  
7.367  
7.361  
7.355  
7.341  
7.337  
7.330  
7.325  
7.321  
7.314  
7.302  
7.295  
7.290  
7.275  
7.271  
7.260

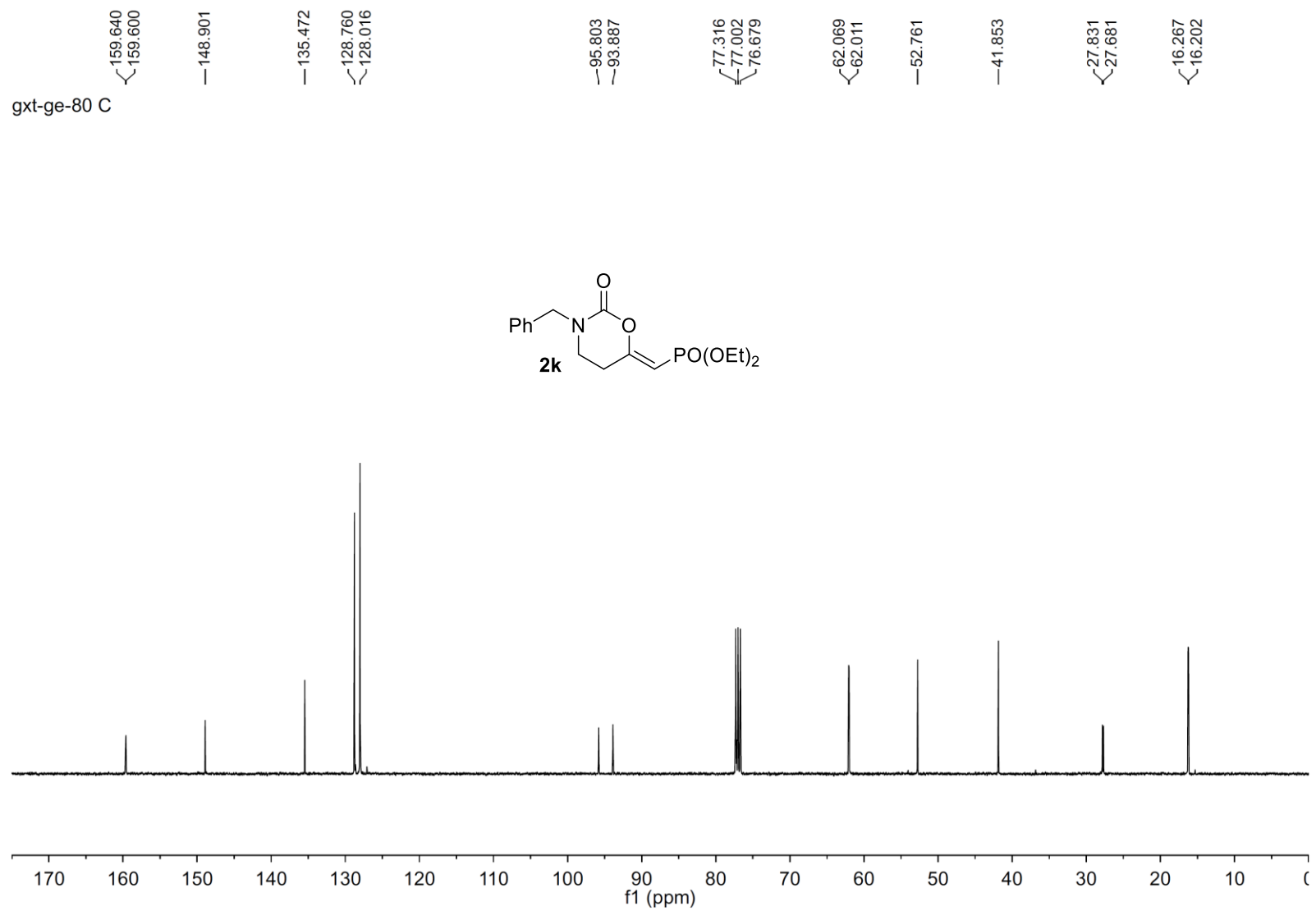
4.834  
4.811  
4.572  
4.242  
4.235  
4.230  
4.228  
4.217  
4.212  
4.199  
4.195  
4.193  
4.179  
4.175  
4.161  
4.158  
4.152  
4.150  
3.264  
3.262  
3.247  
3.232  
3.231  
2.613  
2.611  
2.599  
2.597  
2.594  
2.582

1.375  
1.358  
1.340

gxt-ge-80 H

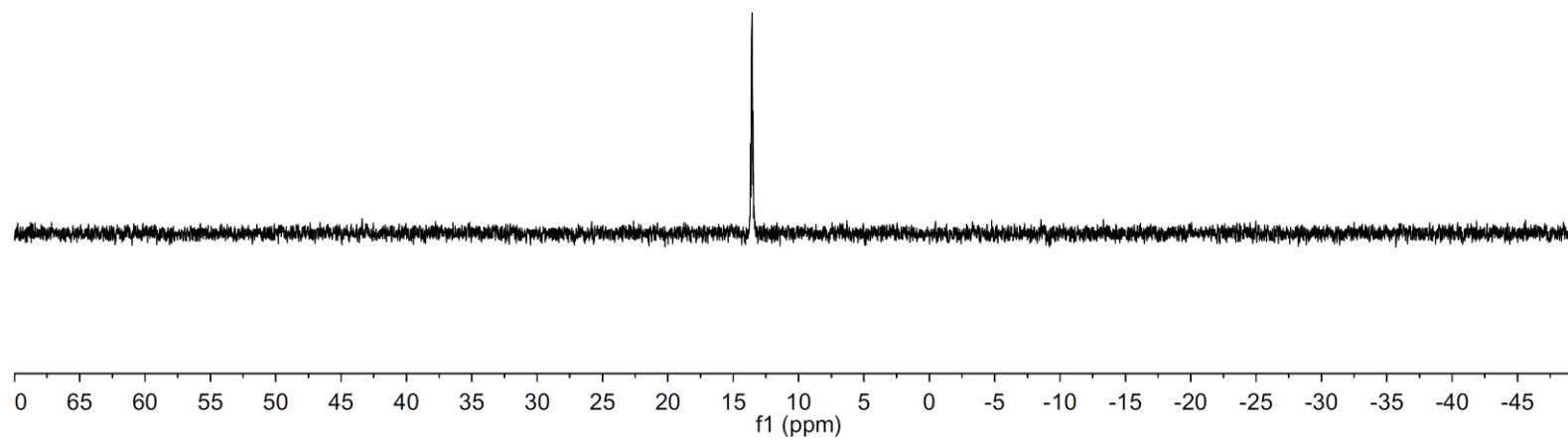
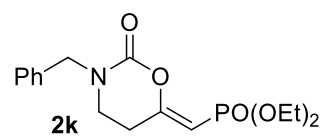
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4.811  
4.572  
4.242  
4.235  
4.230  
4.228  
4.217  
4.212  
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4.195  
4.193  
4.179  
4.175  
4.161  
4.158  
4.152  
4.150  
3.264  
3.262  
3.247  
3.232  
3.231  
2.613  
2.611  
2.599  
2.597  
2.594  
2.582



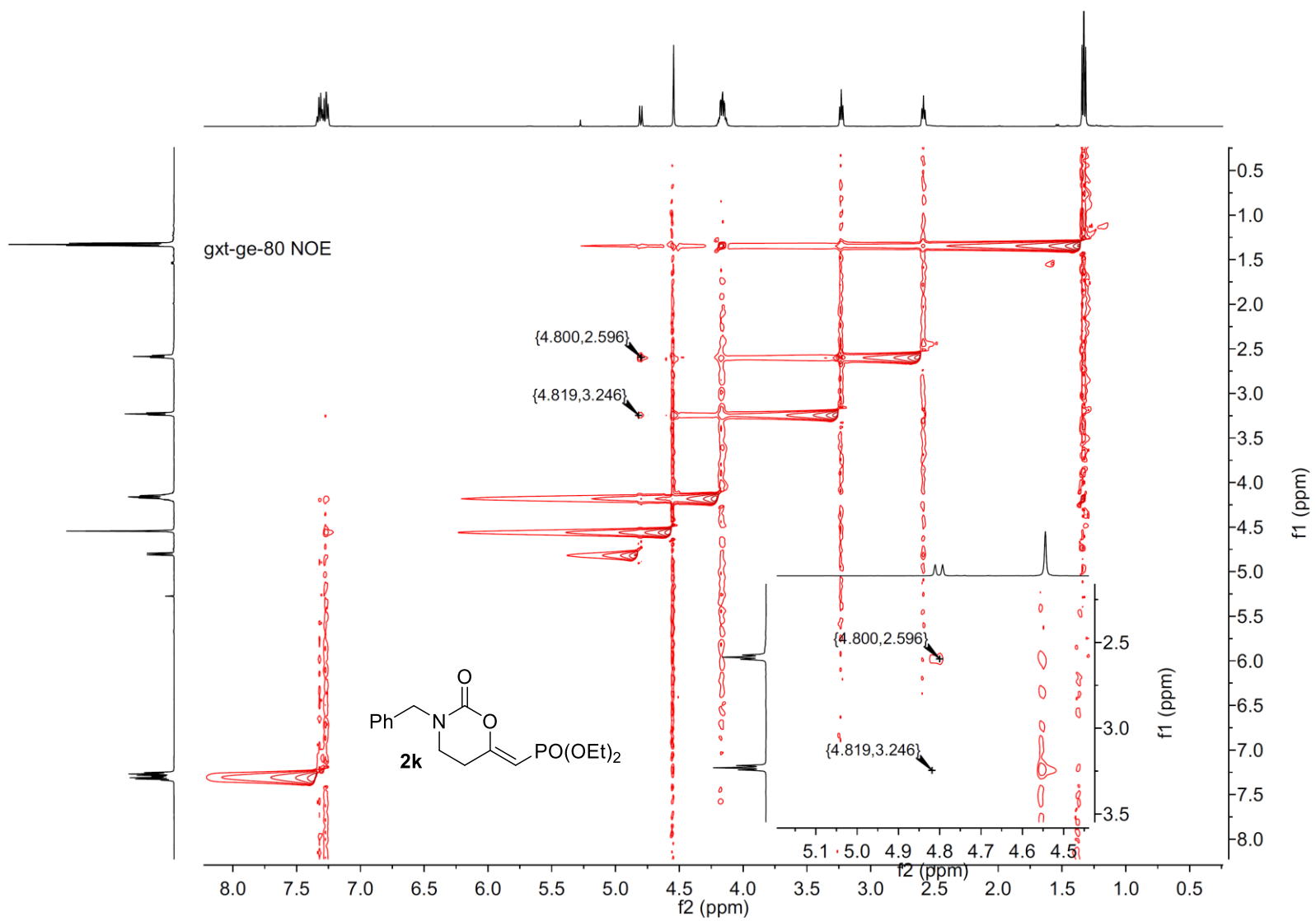


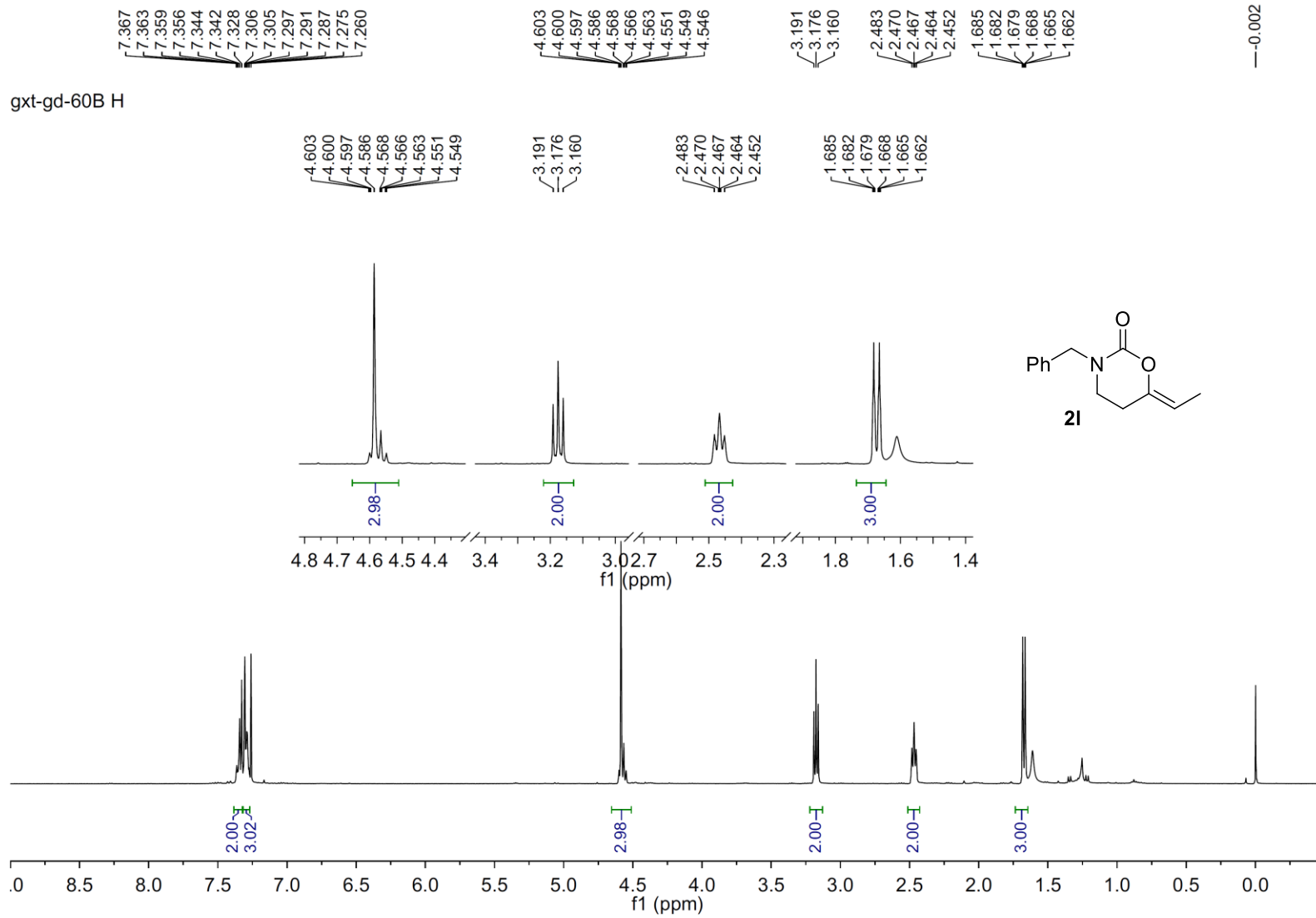
gxt-ge-80 P

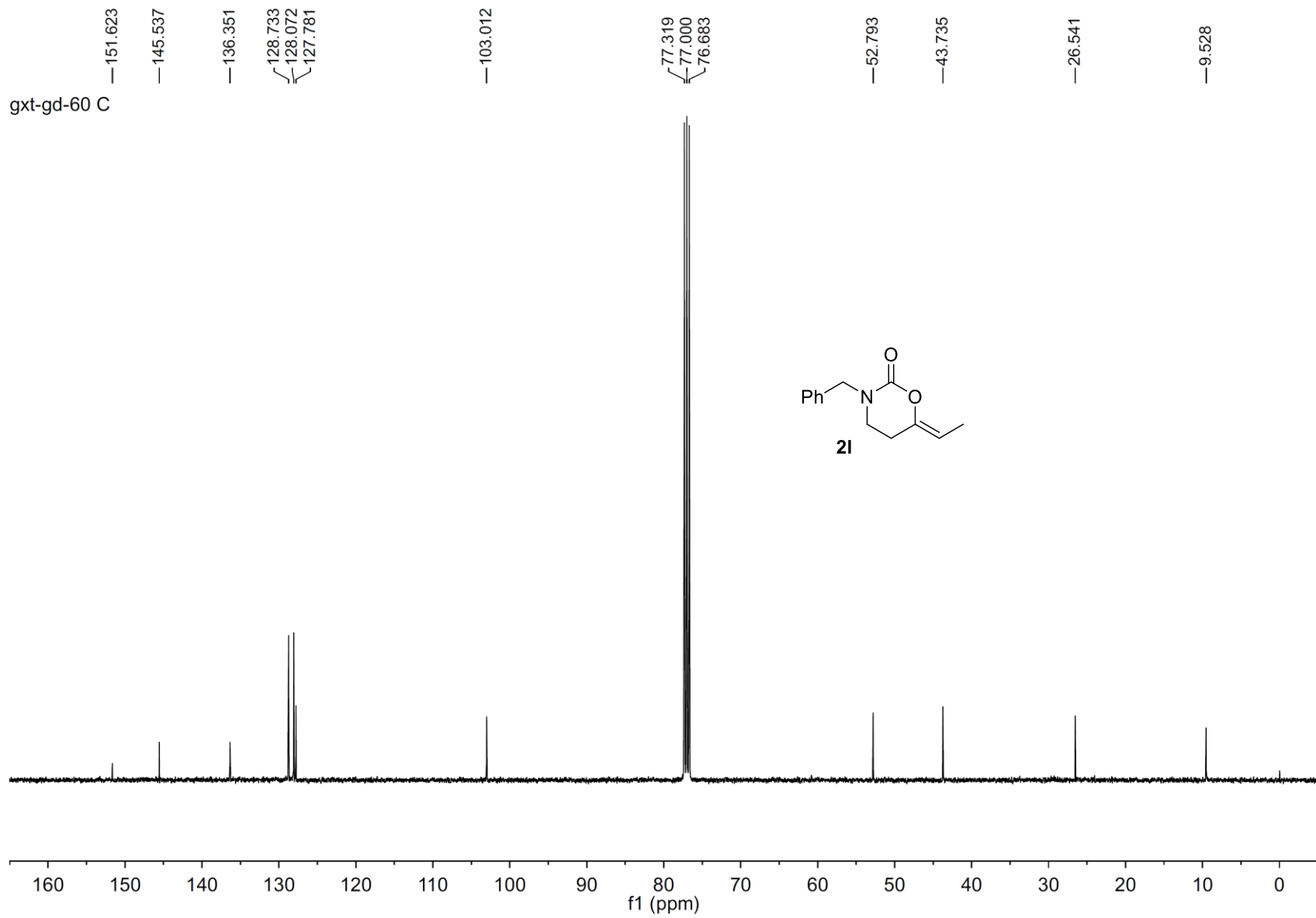
13.680  
13.613  
13.551  
13.486

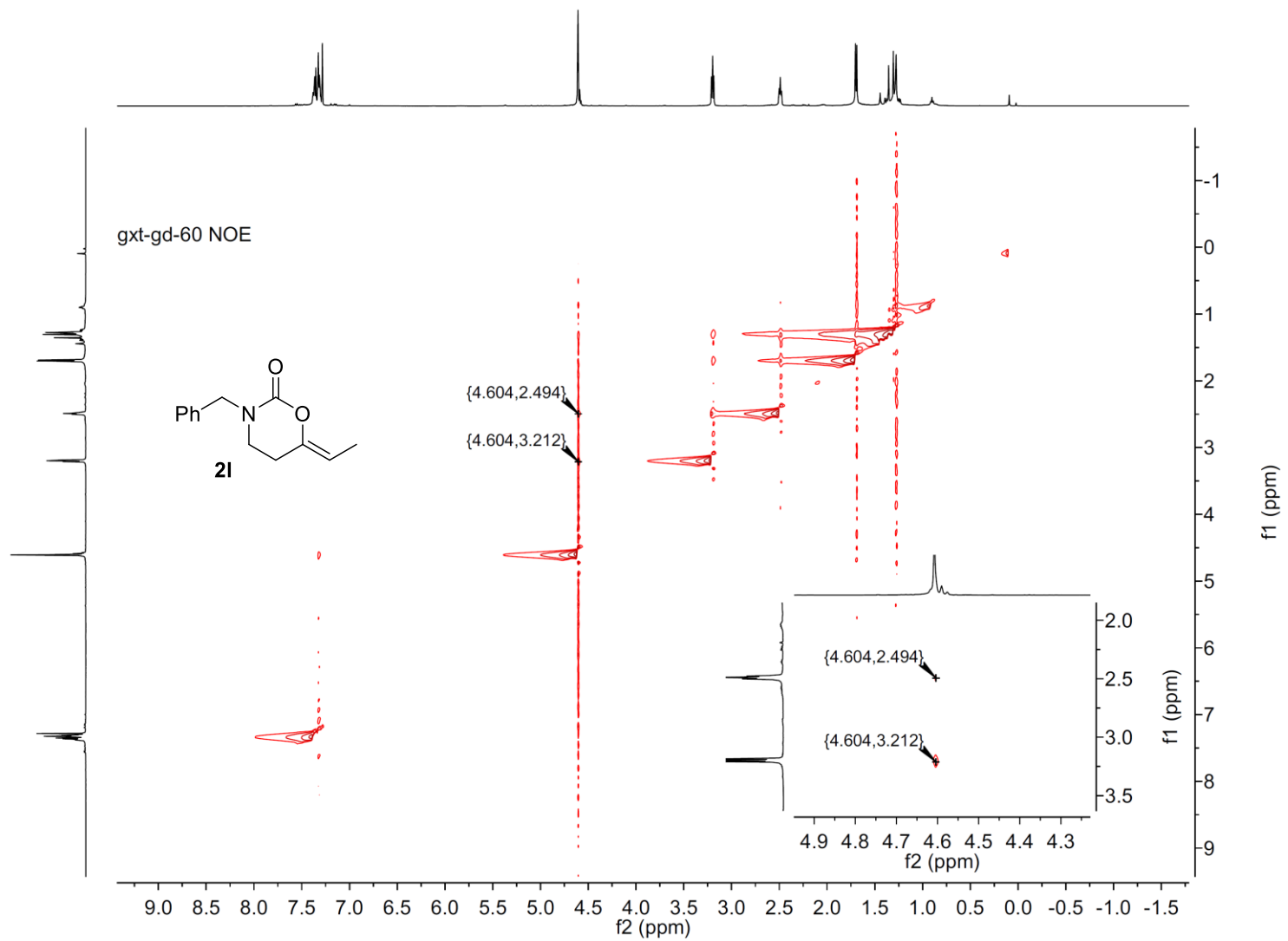










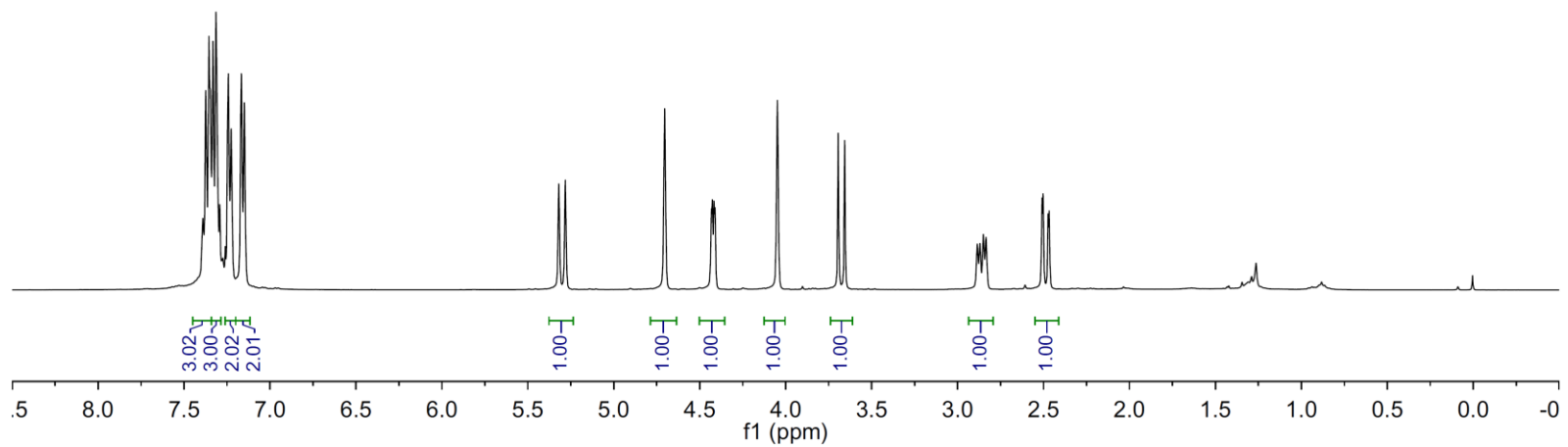
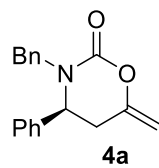


7.395  
7.390  
7.374  
7.355  
7.348  
7.332  
7.314  
7.309  
7.293  
7.260  
7.246  
7.241  
7.225  
7.170  
7.165  
7.150

5.320  
5.282

4.704  
4.433  
4.426  
4.417  
4.410  
4.048  
3.694  
3.657  
2.885  
2.880  
2.870  
2.866  
2.854  
2.850  
2.845  
2.834  
2.830  
2.509  
2.502  
2.473  
2.467

gxt-gd-26 H



151.378  
149.965

138.577  
136.111  
128.782  
128.658  
128.227  
128.071  
127.761  
126.229

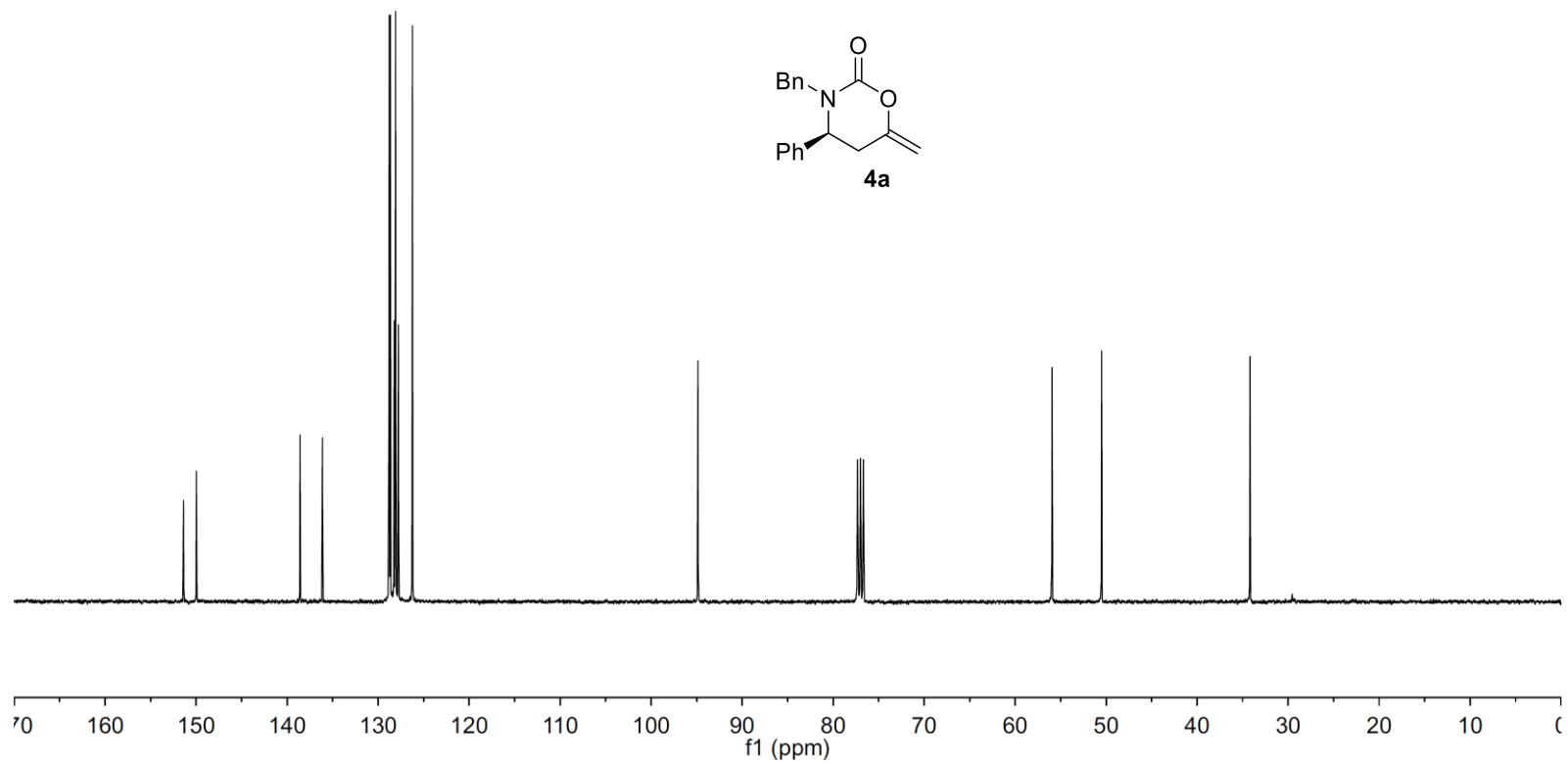
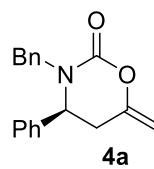
94.856

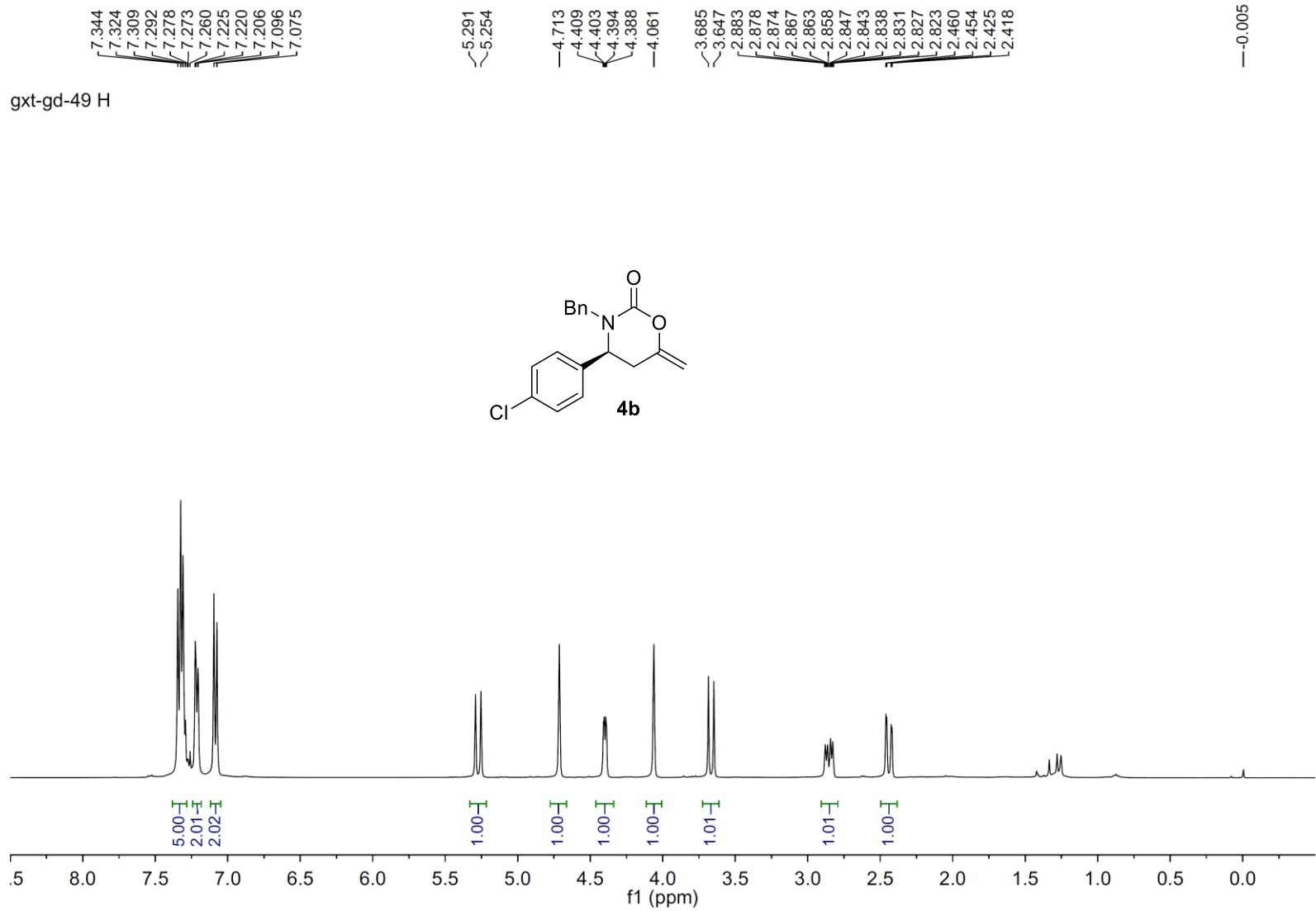
77.320  
77.000  
76.683

55.941  
50.493

34.192

gxt-gd-26 C





gxt-gd-49 C

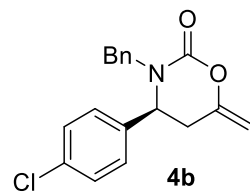
151.196  
149.645  
137.203  
135.894  
134.131  
129.032  
128.754  
128.112  
127.912  
127.660

95.215

77.319  
77.000  
76.683

55.468  
50.650

34.152



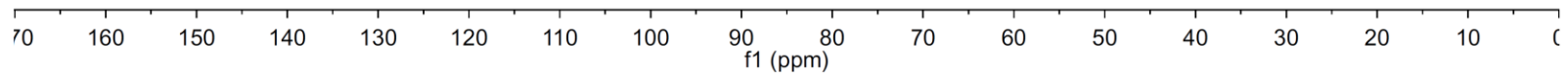
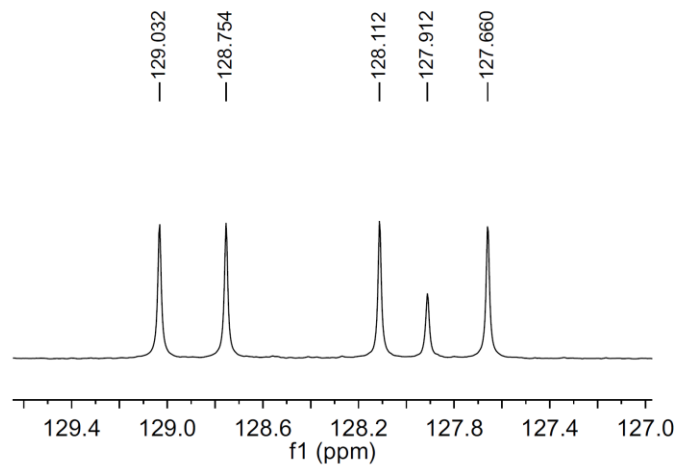
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128.754

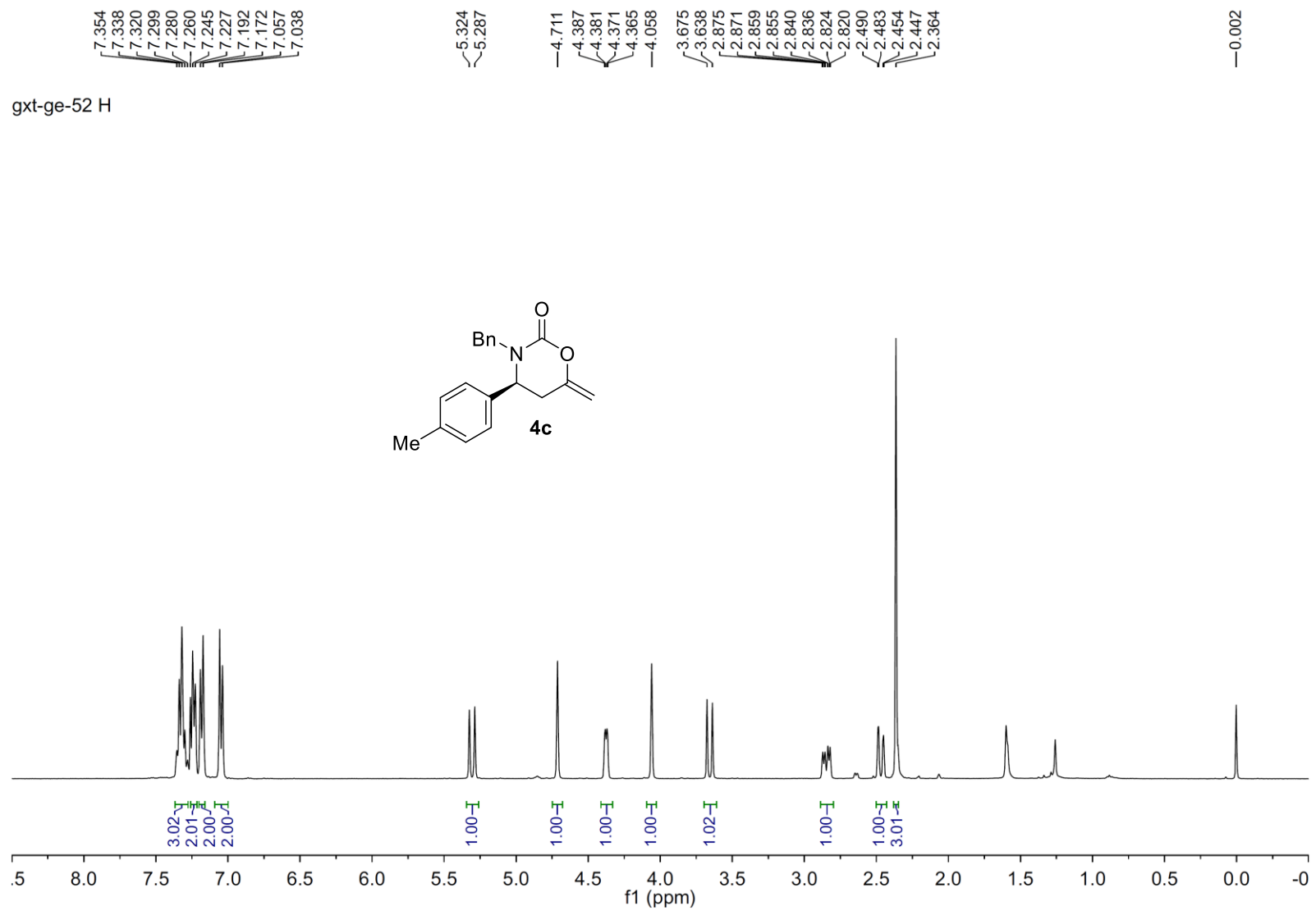
128.112

127.912

127.660







gxt-ge-52 C

151.462  
150.187

138.098  
136.276  
135.636  
129.518  
128.701  
128.130  
127.775  
126.241

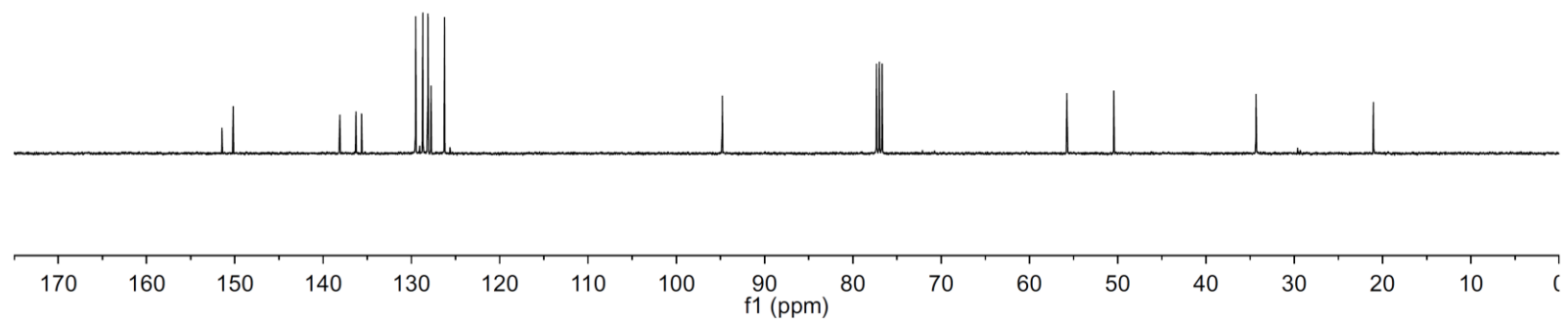
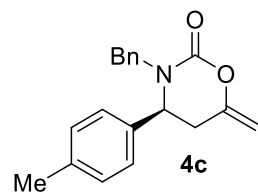
94.785

77.318  
77.000  
76.682

55.757  
50.444

34.336

21.042



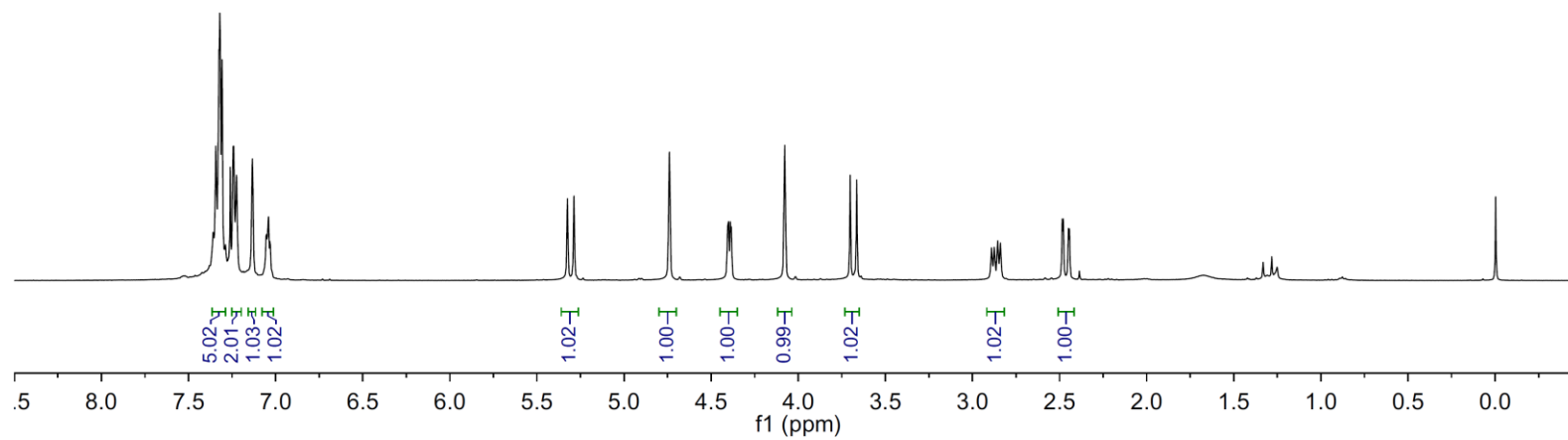
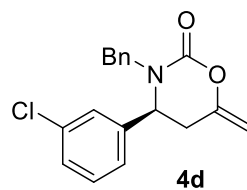
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7.359  
7.343  
7.326  
7.320  
7.317  
7.307  
7.288  
7.260  
7.245  
7.239  
7.225  
7.134  
7.066  
7.056  
7.052  
7.051  
7.046  
7.041  
7.034  
7.029

5.325  
5.287

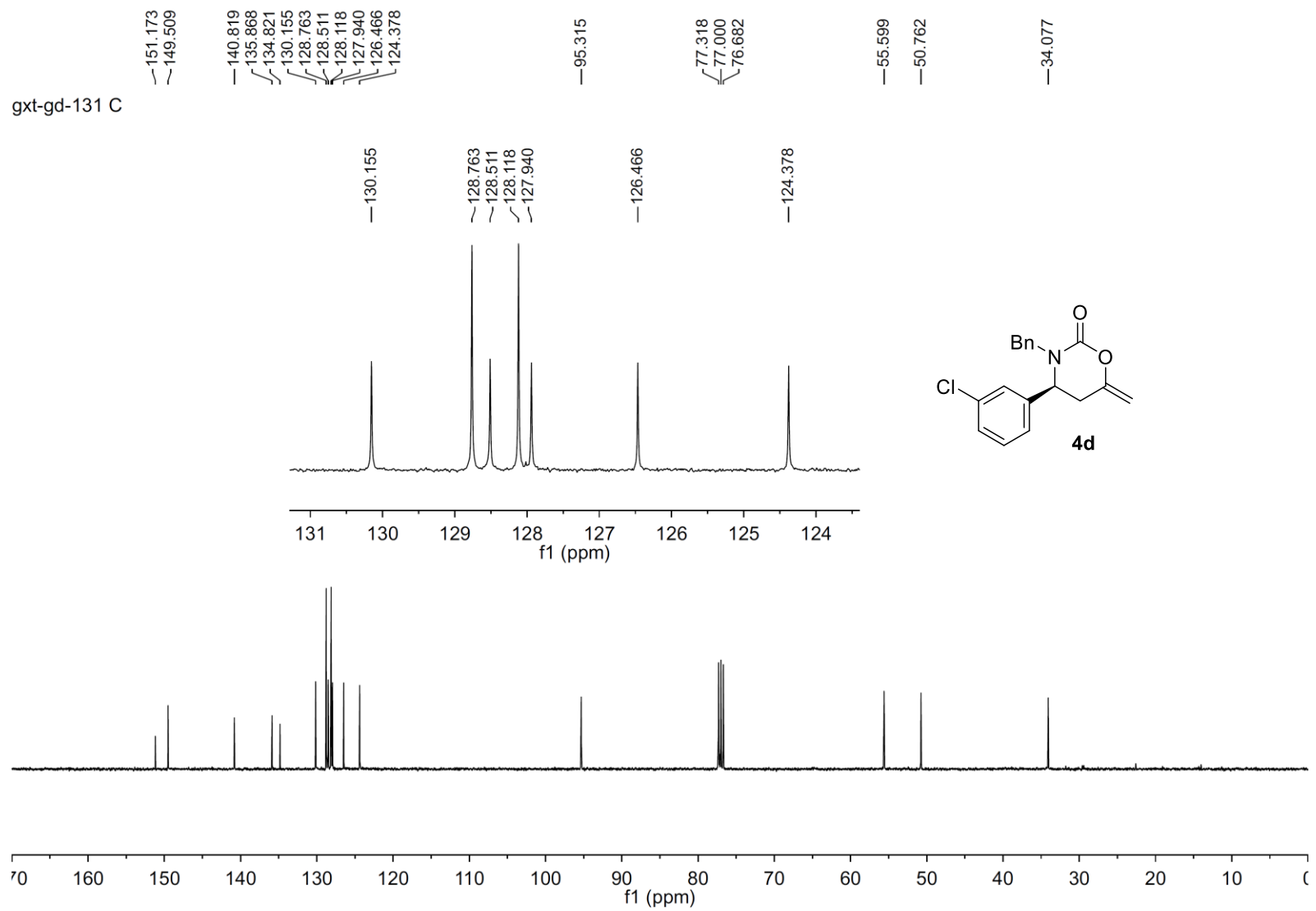
4.739  
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4.400  
4.390  
4.384  
4.077

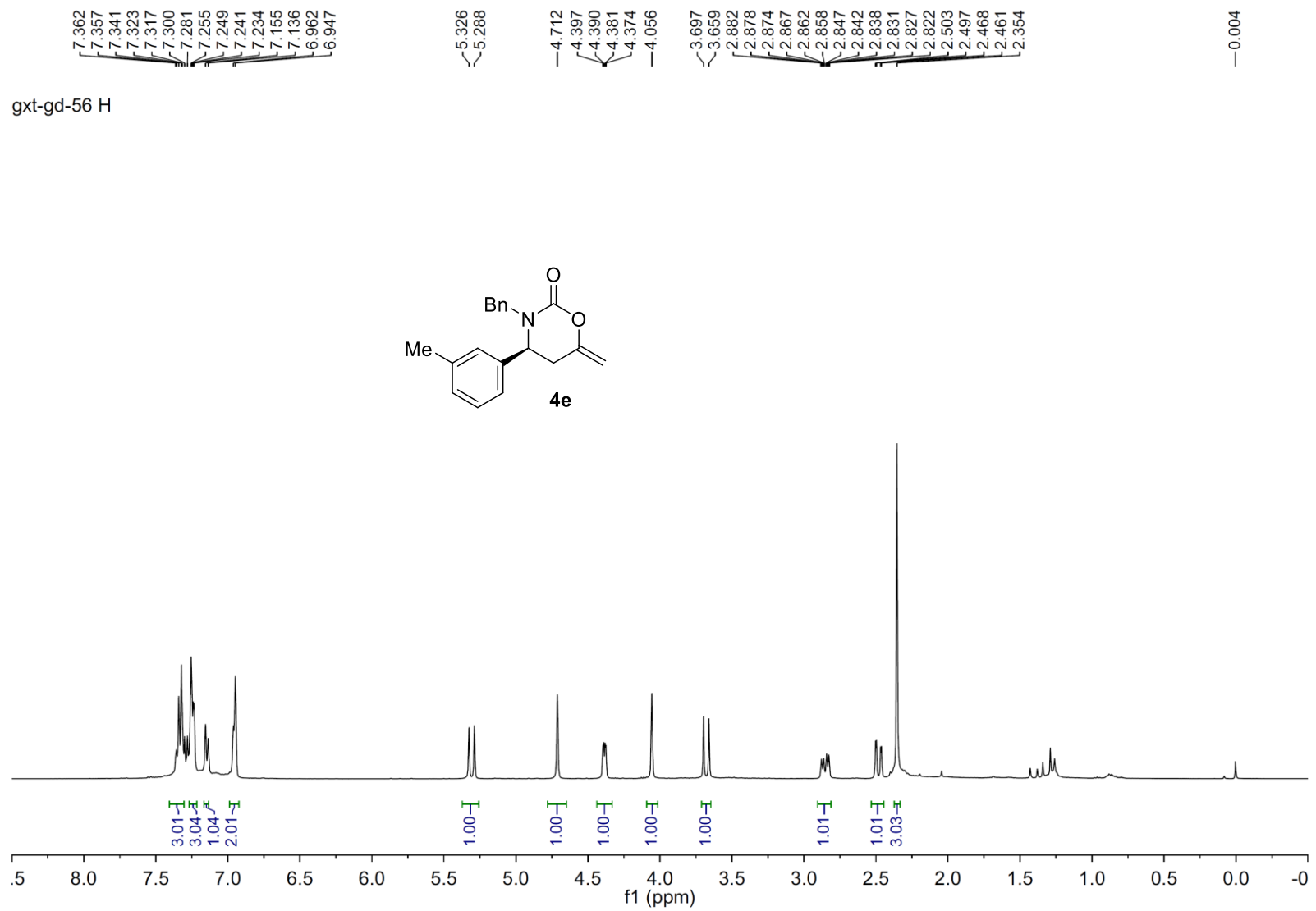
3.702  
3.664  
2.895  
2.886  
2.879  
2.875  
2.871  
2.859  
2.855  
2.851  
2.844  
2.839  
2.835  
2.485  
2.479  
2.449  
2.443

gxt-gd-57 H

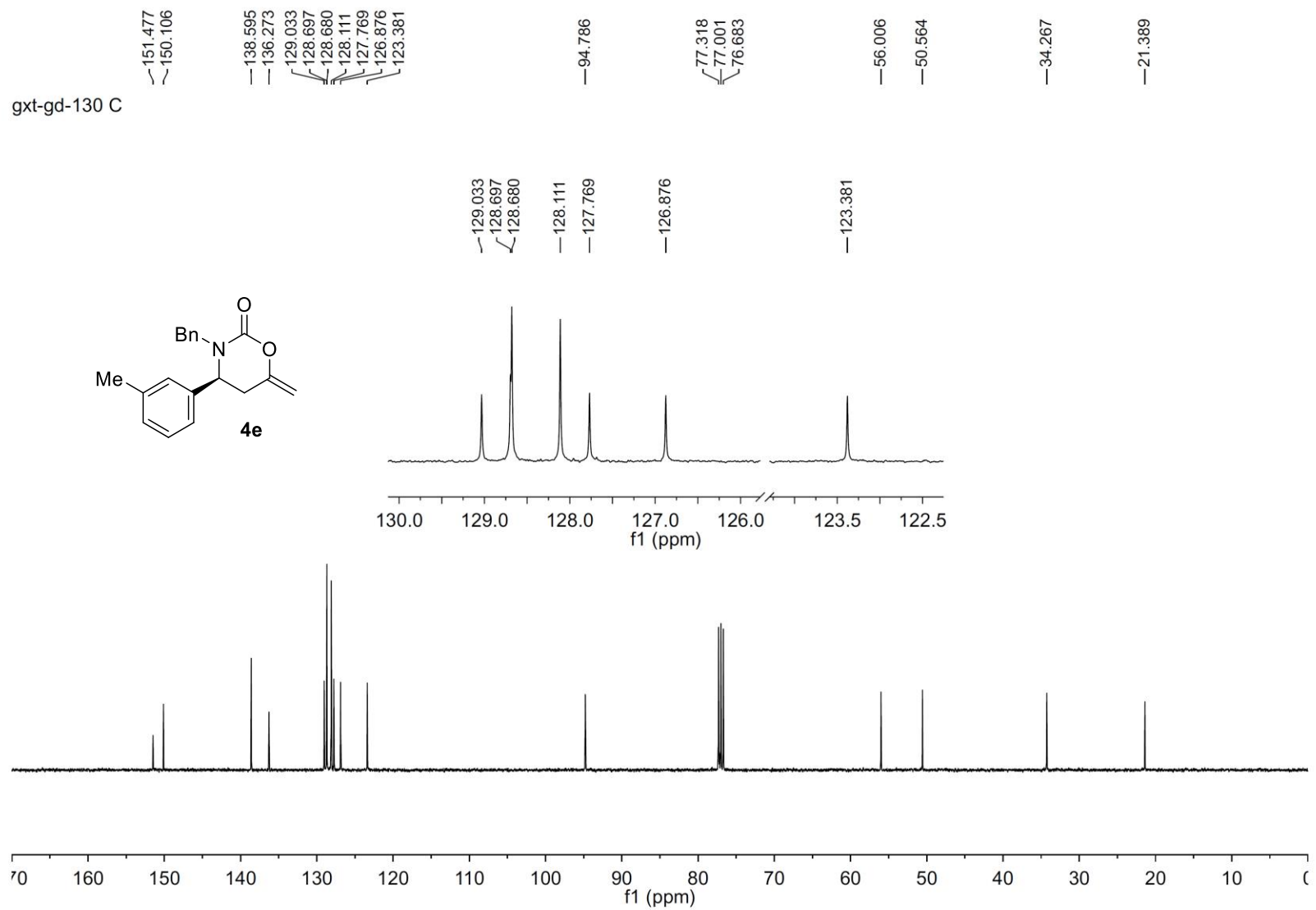
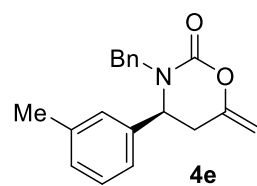


gxt-gd-131 C





gxt-gd-130 C



7.875  
7.854  
7.828  
7.609  
7.554  
7.537  
7.526  
7.515  
7.499  
7.362  
7.346  
7.328  
7.309  
7.292  
7.274  
7.260  
7.242

5.402  
5.364

4.714  
4.601  
4.595  
4.583  
4.578

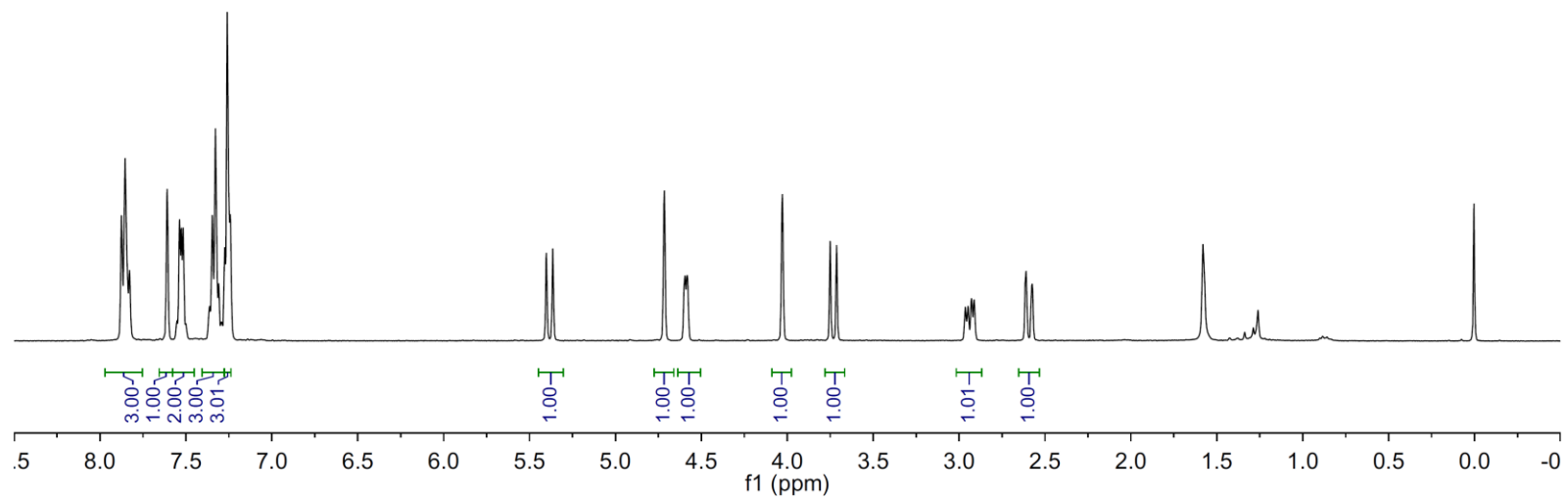
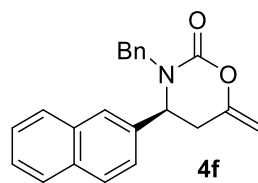
4.028

3.750  
3.713

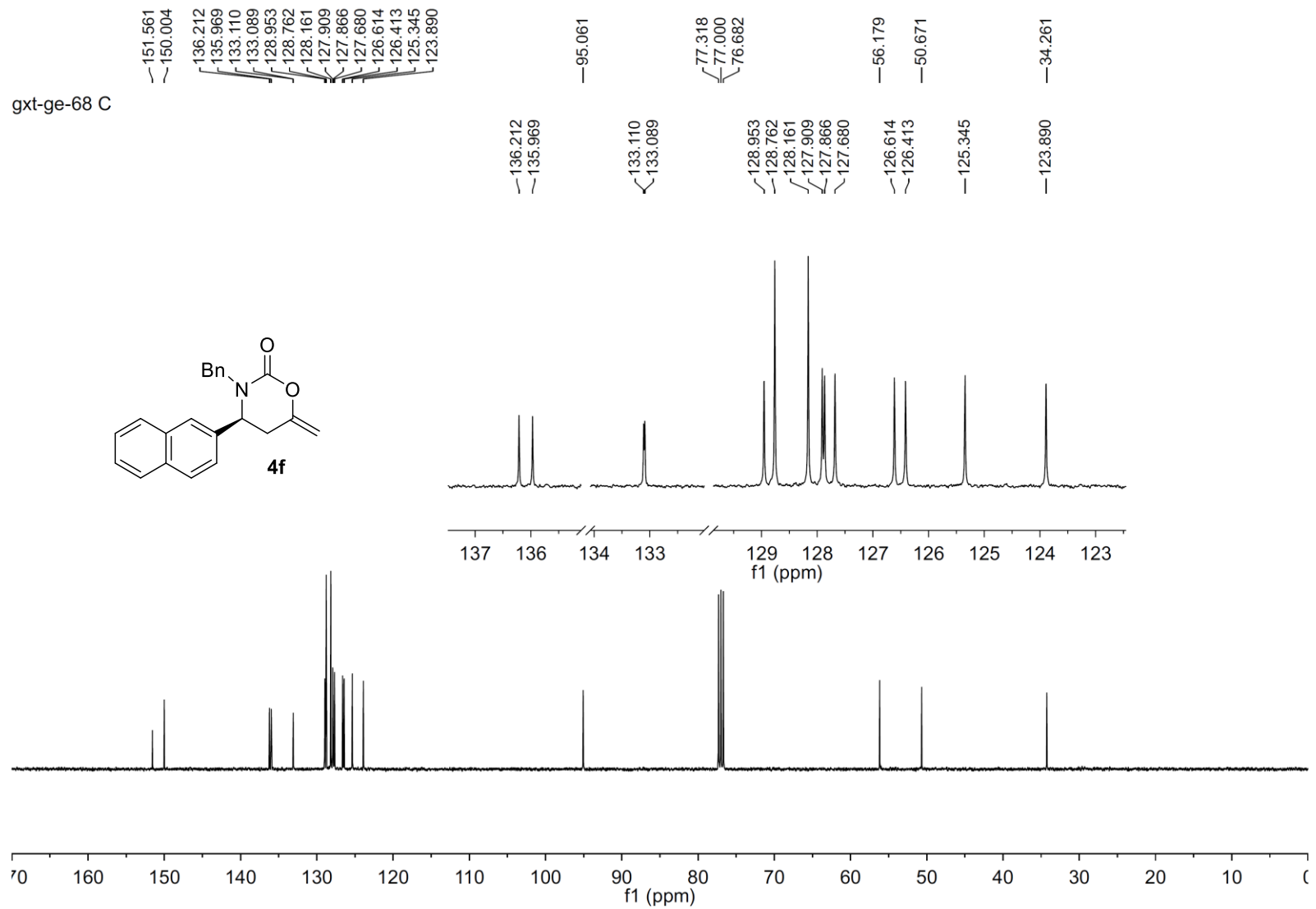
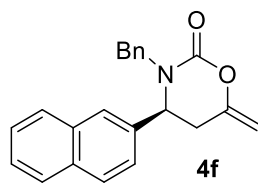
2.964  
2.948  
2.928  
2.912  
2.615  
2.608  
2.579  
2.573

0.003

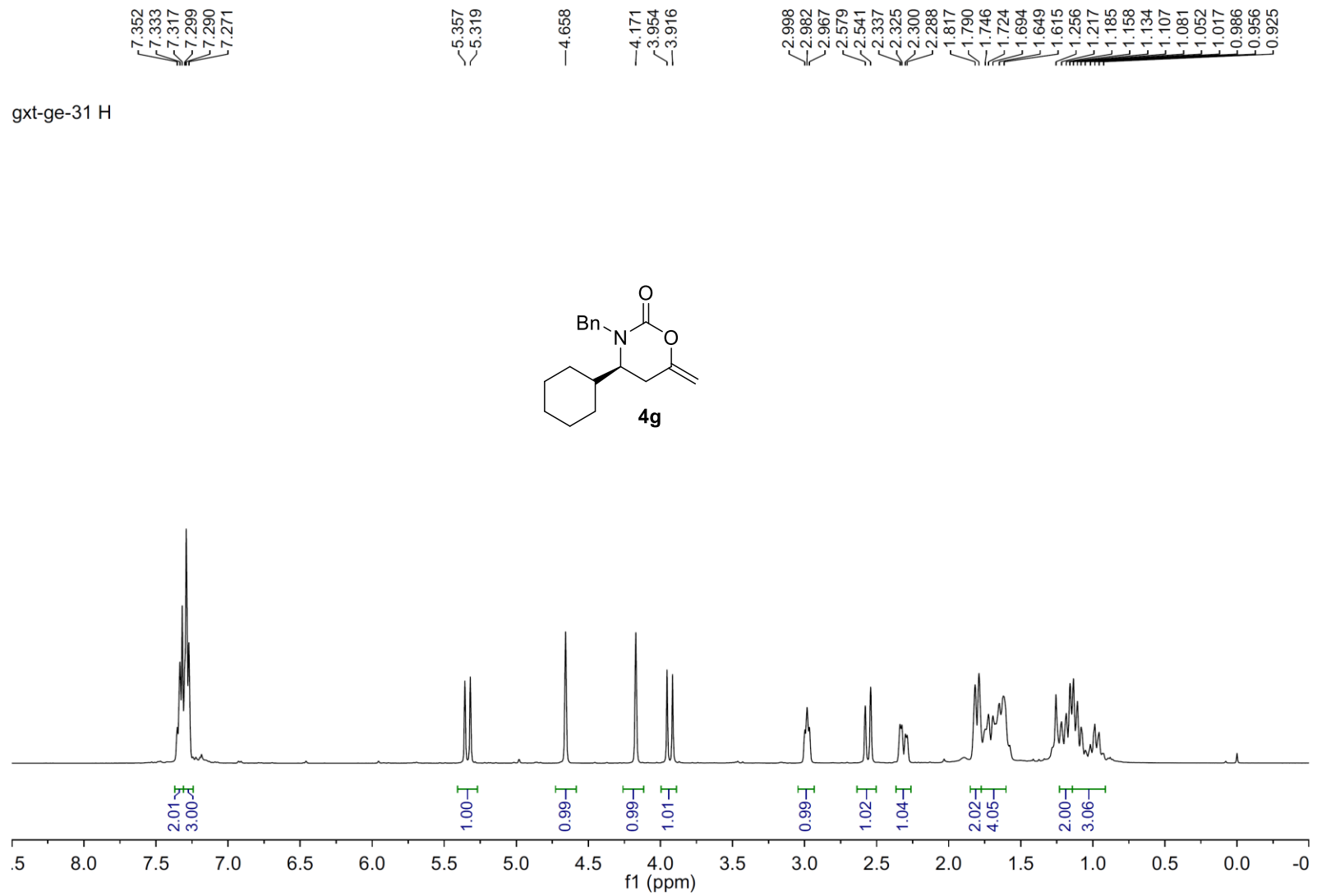
gxt-ge-68 H

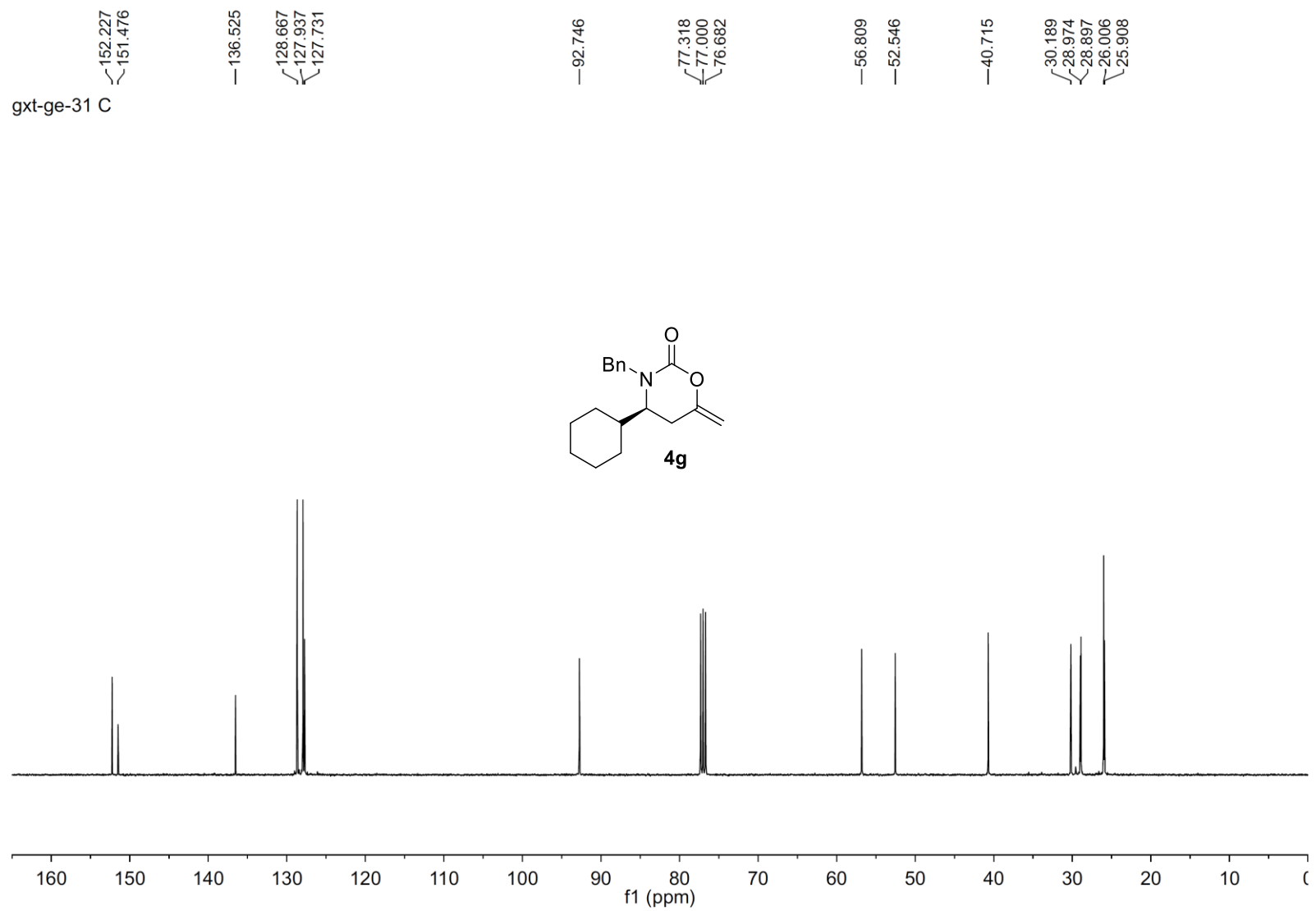


gxt-ge-68 C

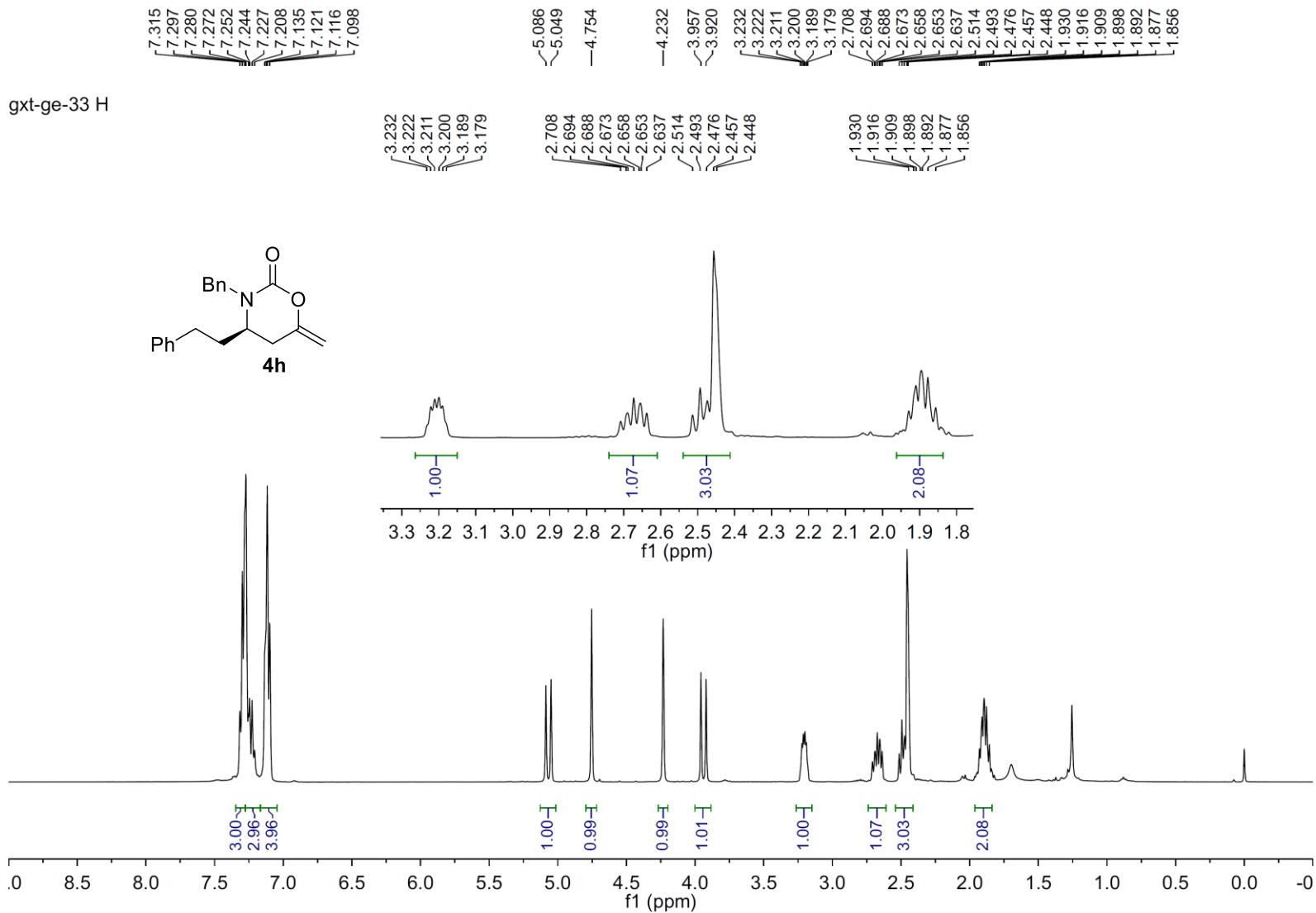
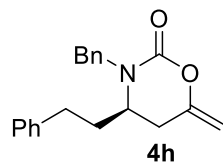


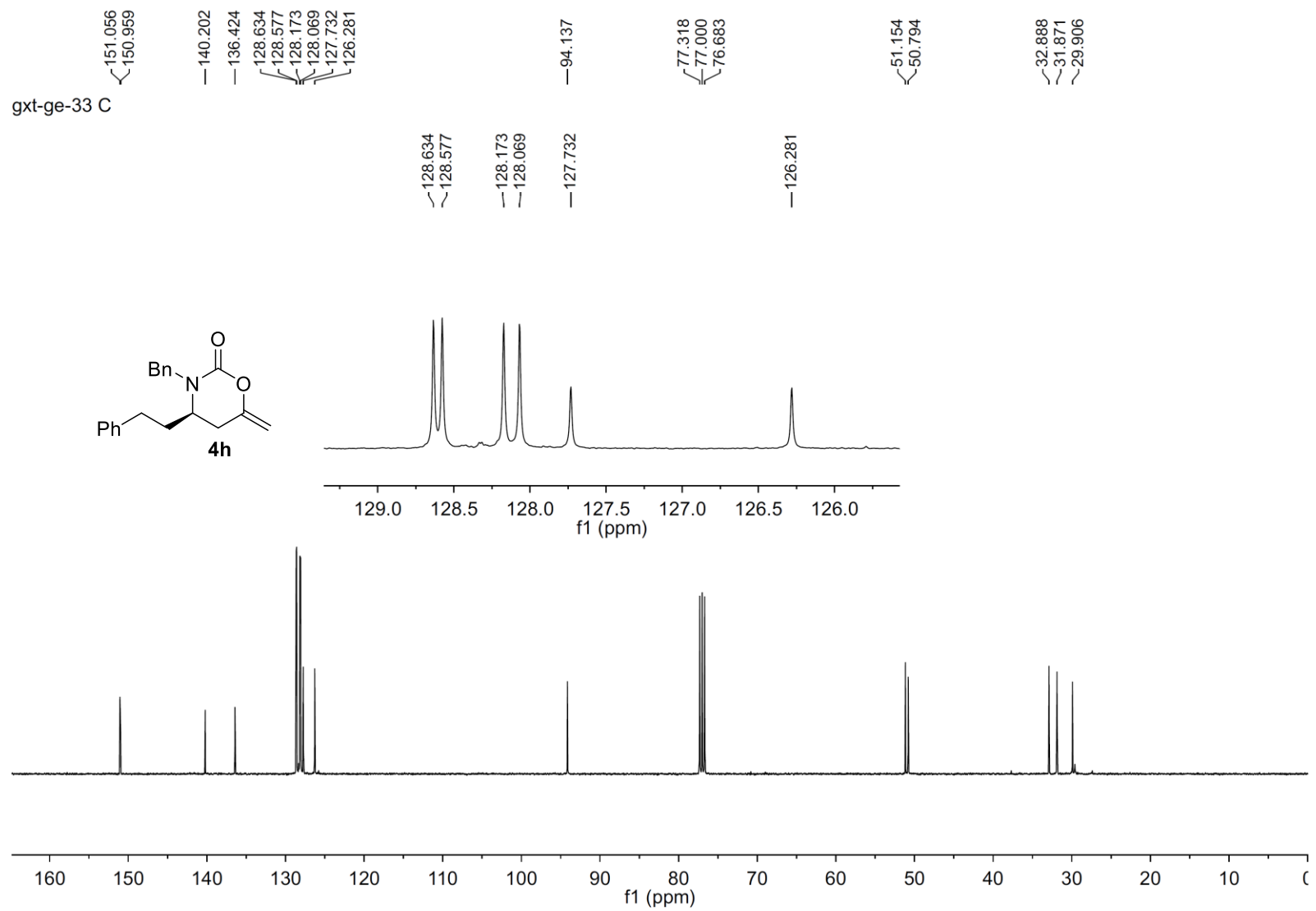




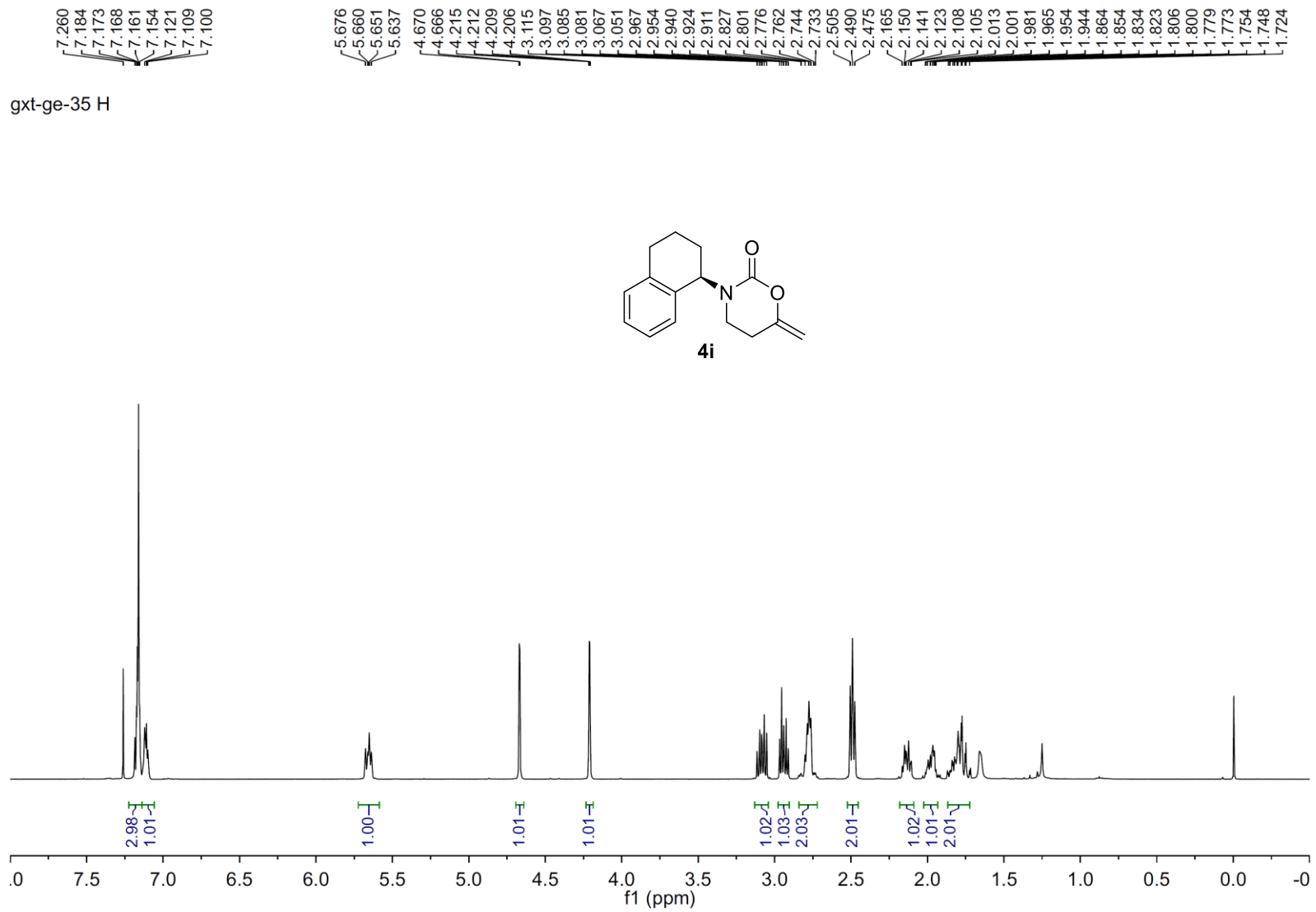


gxt-ge-33 H





gxt-ge-35 H



gxt-ge-35 C

152.836  
151.415

138.681  
133.809  
129.355  
127.133  
126.863  
126.244

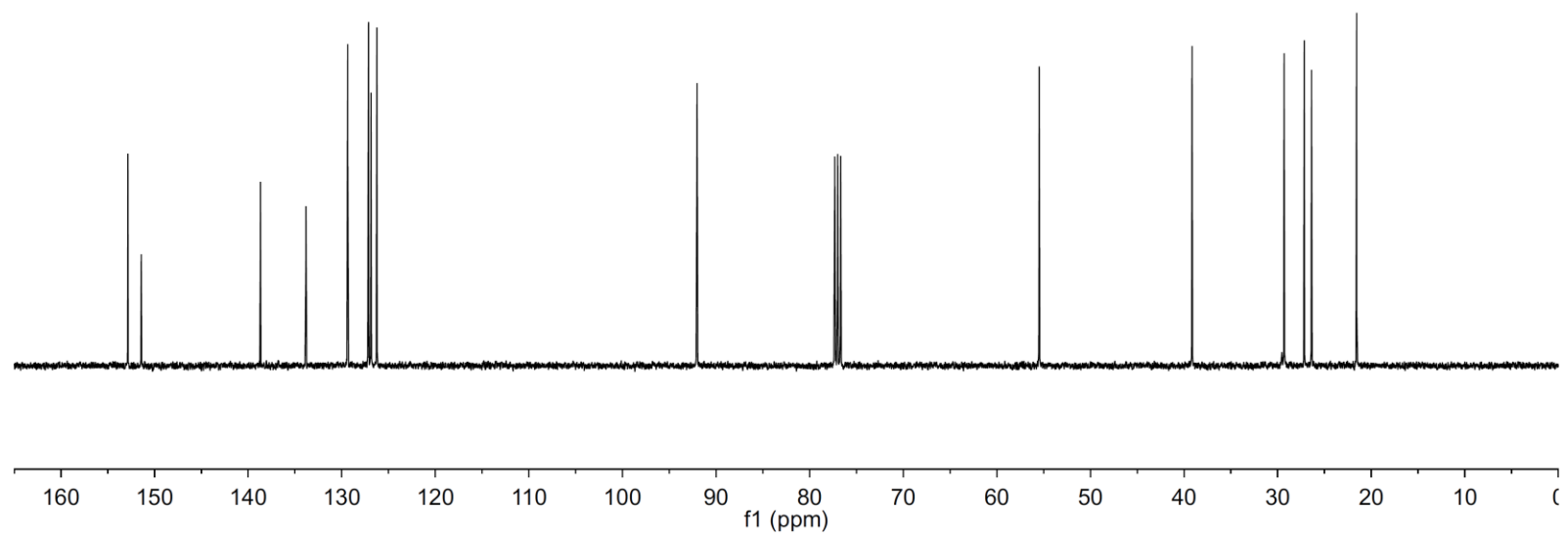
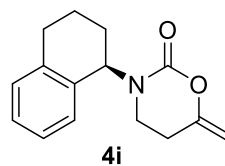
92.029

77.319  
77.002  
76.682

55.478

39.143

29.312  
27.165  
26.388  
21.566



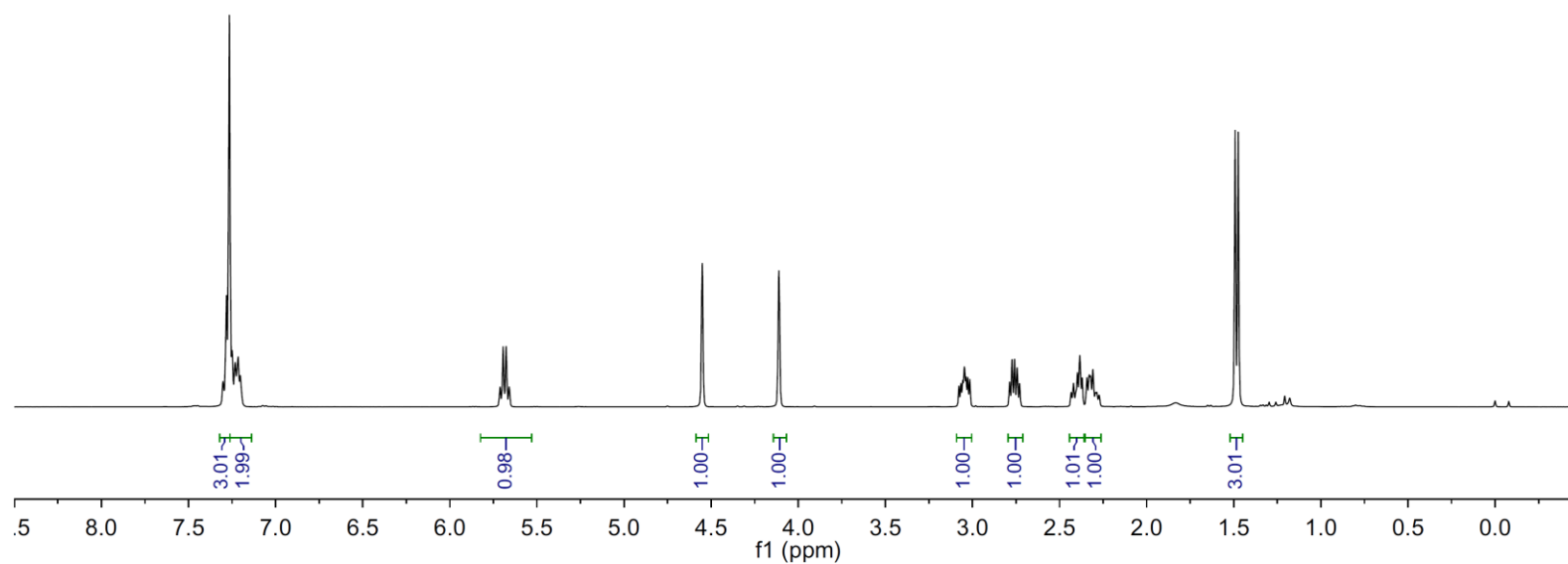
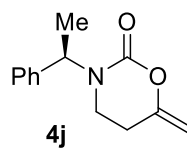
7.302  
7.282  
7.266  
7.250  
7.235  
7.231  
7.214  
7.202

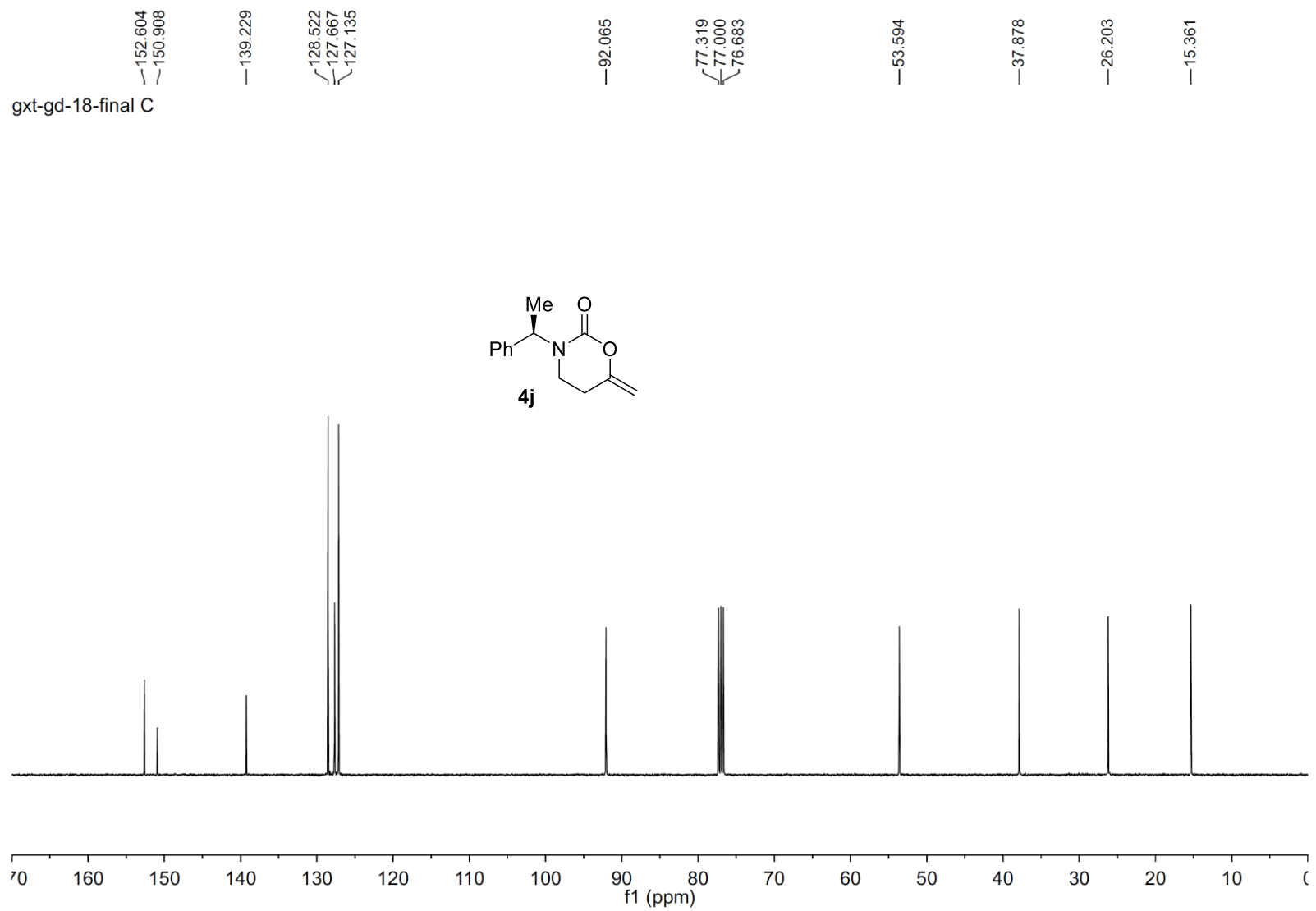
5.712  
5.694  
5.676  
5.658

4.551


4.111  
3.077  
3.066  
3.056  
3.048  
3.044  
3.036  
3.026  
3.015  
2.787  
2.773  
2.758  
2.743  
2.729  
2.434  
2.420  
2.407  
2.398  
2.384  
2.370  
2.343  
2.330  
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2.293  
2.285  
2.272  
1.492  
1.474

gxt-gd-18-final H








**Analysis Report**

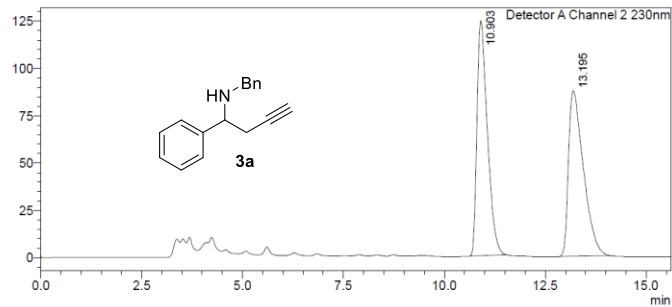
## &lt;Sample Information&gt;

Sample Name : gxt-gd-22-rac-objh-90-10-1.0-  
 Sample ID :  
 Data Filename : gxt-gd-22-rac-objh-90-10-1.0-4.lcd  
 Method Filename : gxt-20170531.lcm  
 Batch Filename :  
 Vial # : 1-1  
 Injection Volume : 20 uL  
 Date Acquired : 2018/6/10 17:42:02  
 Date Processed : 2018/6/10 17:57:40

Sample Type : Unknown  
 Acquired by : System Administrator  
 Processed by : System Administrator

## &lt;Chromatogram&gt;

mV




## &lt;Peak Table&gt;

Detector A Channel 2 230nm

Peak#	Ret. Time	Area	Height	Conc.
1	10.903	2229255	124118	49.819
2	13.195	2245429	87402	50.181
Total		4474685	211521	

D:\Data\GXT\20180518\gxt-gd-22-rac-objh-90-10-1.0-4.lcd


**Analysis Report**

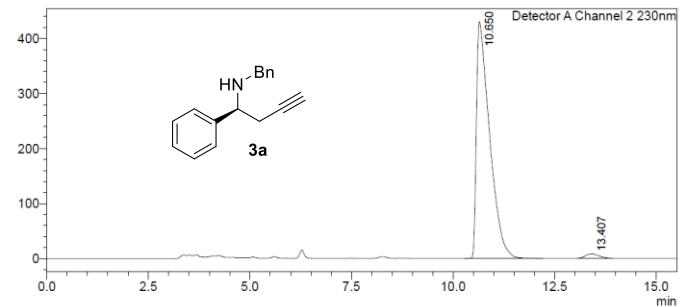
## &lt;Sample Information&gt;

Sample Name : gxt-gd-22-asy-objh-90-10-1.0-  
 Sample ID :  
 Data Filename : gxt-gd-22-asy-objh-90-10-1.0-1.lcd  
 Method Filename : gxt-20170531.lcm  
 Batch Filename :  
 Vial # : 1-1  
 Injection Volume : 20 uL  
 Date Acquired : 2018/6/10 17:59:27  
 Date Processed : 2018/6/10 18:14:58

Sample Type : Unknown  
 Acquired by : System Administrator  
 Processed by : System Administrator

## &lt;Chromatogram&gt;

mV



## &lt;Peak Table&gt;

Detector A Channel 2 230nm

Peak#	Ret. Time	Area	Height	Conc.
1	10.650	9762775	429955	98.163
2	13.407	182660	8039	1.837
Total		9945435	437994	

D:\Data\GXT\20180518\gxt-gd-22-asy-objh-90-10-1.0-1.lcd

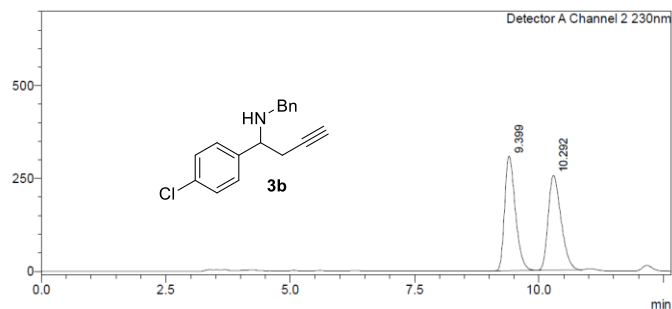
SHIMADZU LabSolutions Analysis Report

## &lt;Sample Information&gt;

Sample Name : gxt-gd-44-rac-ojh-90-10-1.0-  
 Sample ID :  
 Data Filename : gxt-gd-44-rac-ojh-90-10-1.0-1.lcd  
 Method Filename : gxt-20170531.lcm  
 Batch Filename :  
 Vial # : 1-1 Sample Type : Unknown  
 Injection Volume : 20 uL  
 Date Acquired : 2018/6/10 18:16:18 Acquired by : System Administrator  
 Date Processed : 2018/6/10 18:28:58 Processed by : System Administrator

## &lt;Chromatogram&gt;

mV



## &lt;Peak Table&gt;

Detector A Channel 2 230nm

Peak#	Ret. Time	Area	Height	Conc.
1	9.399	4623457	308332	50.184
2	10.292	4589612	254918	49.816
Total		9213069	563250	

D:\Data\GXT\20180518\gxt-gd-44-rac-ojh-90-10-1.0-1.lcd

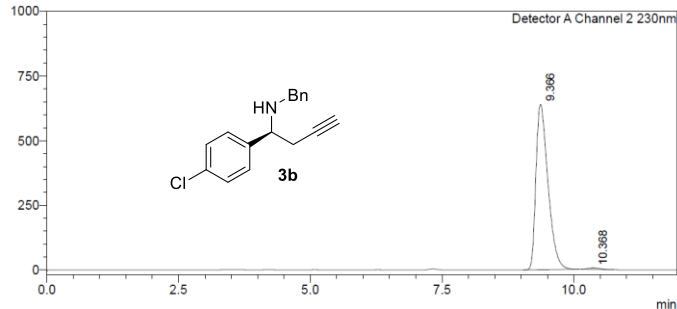
SHIMADZU LabSolutions Analysis Report

## &lt;Sample Information&gt;

Sample Name : gxt-gd-44-asy-ojh-90-10-1.0-  
 Sample ID :  
 Data Filename : gxt-gd-44-asy-ojh-90-10-1.0-3.lcd  
 Method Filename : gxt-20170531.lcm  
 Batch Filename :  
 Vial # : 1-1 Sample Type : Unknown  
 Injection Volume : 20 uL  
 Date Acquired : 2018/6/10 19:00:01 Acquired by : System Administrator  
 Date Processed : 2018/6/10 19:11:59 Processed by : System Administrator

## &lt;Chromatogram&gt;

mV



## &lt;Peak Table&gt;

Detector A Channel 2 230nm

Peak#	Ret. Time	Area	Height	Conc.
1	9.366	10163952	640057	99.076
2	10.368	94836	5856	0.924
Total		10258788	645913	

D:\Data\GXT\20180518\gxt-gd-44-asy-ojh-90-10-1.0-3.lcd

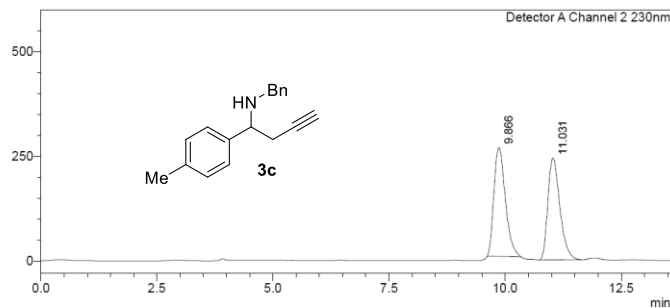
SHIMADZU LabSolutions Analysis Report

## &lt;Sample Information&gt;

Sample Name : gxt-ge-47-asy-adh-99-1-0-8-  
 Sample ID :  
 Data Filename : gxt-ge-47-asy-adh-99-1-0-8-1.lcd  
 Method Filename : gxt-20170531.lcm  
 Batch Filename :  
 Vial # : 1-1 Sample Type : Unknown  
 Injection Volume : 20 uL  
 Date Acquired : 2018/7/14 16:04:18 Acquired by : System Administrator  
 Date Processed : 2018/7/14 16:17:53 Processed by : System Administrator

## &lt;Chromatogram&gt;

mV



## &lt;Peak Table&gt;

Detector A Channel 2 230nm

Peak#	Ret. Time	Area	Height	Conc.
1	9.866	4464318	260757	50.499
2	11.031	4376114	243543	49.501
Total		8840432	504301	

D:\Data\GXT\20180518\gxt-ge-47-asy-adh-99-1-0-8-1.lcd

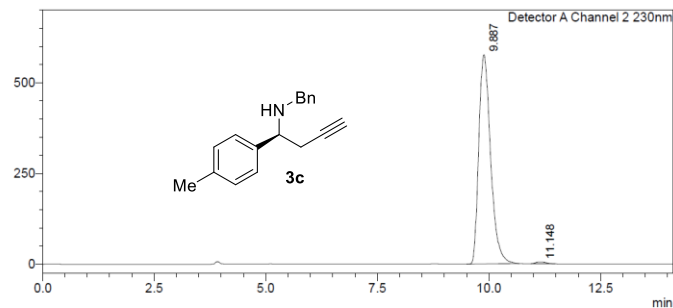
SHIMADZU LabSolutions Analysis Report

## &lt;Sample Information&gt;

Sample Name : gxt-ge-47-rac-adh-99-1-0-8-  
 Sample ID :  
 Data Filename : gxt-ge-47-rac-adh-99-1-0-8-1.lcd  
 Method Filename : gxt-20170531.lcm  
 Batch Filename :  
 Vial # : 1-1 Sample Type : Unknown  
 Injection Volume : 20 uL  
 Date Acquired : 2018/7/14 15:48:30 Acquired by : System Administrator  
 Date Processed : 2018/7/14 16:02:38 Processed by : System Administrator

## &lt;Chromatogram&gt;

mV



## &lt;Peak Table&gt;

Detector A Channel 2 230nm

Peak#	Ret. Time	Area	Height	Conc.
1	9.887	10620679	575942	99.243
2	11.148	81056	5080	0.757
Total		10701736	581022	

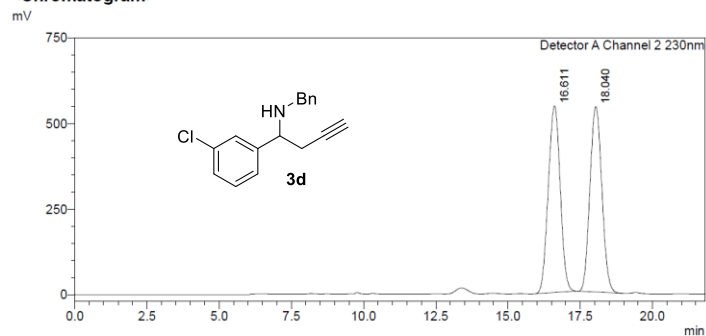
D:\Data\GXT\20180518\gxt-ge-47-rac-adh-99-1-0-8-1.lcd

SHIMADZU LabSolutions Analysis Report

## &lt;Sample Information&gt;

Sample Name : gxt-gd-53-rac-adh-99-1-0.5-  
 Sample ID :  
 Data Filename : gxt-gd-53-rac-adh-99-1-0.5-1.lcd  
 Method Filename : gxt-20170531.lcm  
 Batch Filename :  
 Vial # : 1-1 Sample Type : Unknown  
 Injection Volume : 20 uL  
 Date Acquired : 2018/6/11 18:19:13 Acquired by : System Administrator  
 Date Processed : 2018/6/11 18:41:03 Processed by : System Administrator

## &lt;Chromatogram&gt;



## &lt;Peak Table&gt;

Detector A Channel 2 230nm				
Peak#	Ret. Time	Area	Height	Conc.
1	16.611	15708599	545195	50.099
2	18.040	15646773	542532	49.901
Total		31355373	1087727	

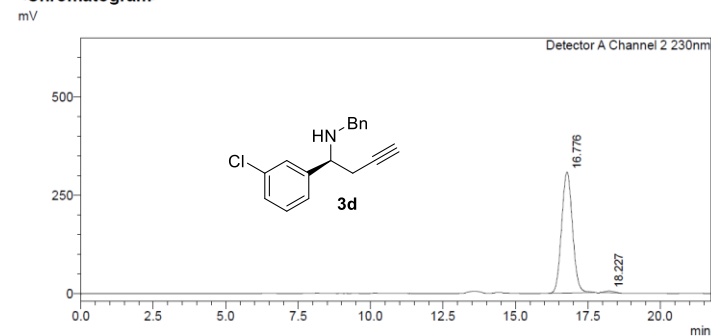
D:\Data\GXT\20180518\gxt-gd-53-rac-adh-99-1-0.5-1.lcd

SHIMADZU LabSolutions Analysis Report

## &lt;Sample Information&gt;

Sample Name : gxt-gd-53-asy-adjh-99-1-0.5-  
 Sample ID :  
 Data Filename : gxt-gd-53-asy-adjh-99-1-0.5-1.lcd  
 Method Filename : gxt-20170531.lcm  
 Batch Filename :  
 Vial # : 1-1 Sample Type : Unknown  
 Injection Volume : 20 uL  
 Date Acquired : 2018/6/11 17:56:11 Acquired by : System Administrator  
 Date Processed : 2018/6/11 18:23:38 Processed by : System Administrator

## &lt;Chromatogram&gt;



## &lt;Peak Table&gt;

Detector A Channel 2 230nm				
Peak#	Ret. Time	Area	Height	Conc.
1	16.776	8051327	307443	98.798
2	18.227	97965	4651	1.202
Total		8149292	312094	

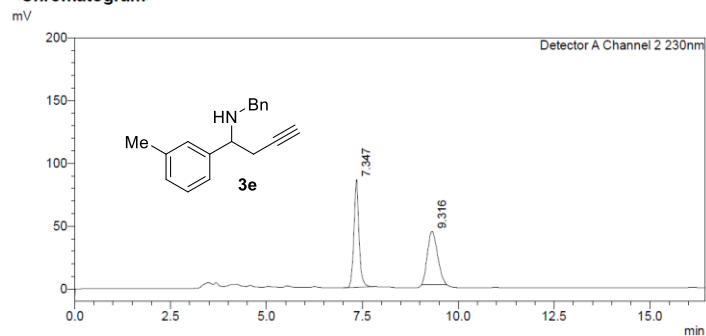
D:\Data\GXT\20180518\gxt-gd-53-asy-adjh-99-1-0.5-1.lcd

SHIMADZU LabSolutions Analysis Report

## &lt;Sample Information&gt;

Sample Name : gxt-gd-52-rac-objh-90-10-1.0-  
 Sample ID :  
 Data Filename : gxt-gd-52-rac-objh-90-10-1.0-3.lcd  
 Method Filename : gxt-20170531.lcm  
 Batch Filename :  
 Vial # : 1-1 Sample Type : Unknown  
 Injection Volume : 20 uL  
 Date Acquired : 2018/6/11 14:34:00 Acquired by : System Administrator  
 Date Processed : 2018/6/11 14:54:53 Processed by : System Administrator

## &lt;Chromatogram&gt;



## &lt;Peak Table&gt;

Detector A Channel 2 230nm				
Peak#	Ret. Time	Area	Height	Conc.
1	7.347	792799	85846	50.349
2	9.316	781813	42548	49.651
Total		1574611	128393	

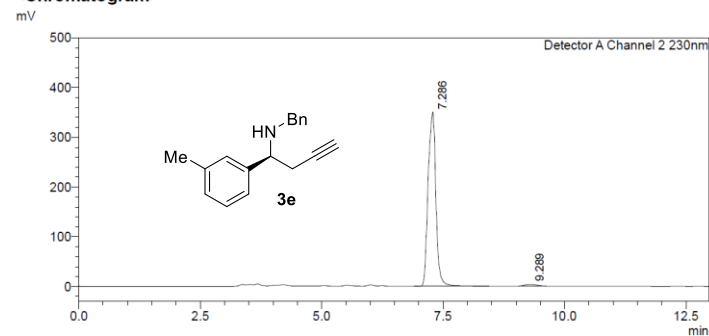
D:\Data\GXT\20180518\gxt-gd-52-rac-objh-90-10-1.0-3.lcd

SHIMADZU LabSolutions Analysis Report

## &lt;Sample Information&gt;

Sample Name : gxt-gd-52-asy-objh-90-10-1.0-  
 Sample ID :  
 Data Filename : gxt-gd-52-asy-objh-90-10-1.0-1.lcd  
 Method Filename : gxt-20170531.lcm  
 Batch Filename :  
 Vial # : 1-1 Sample Type : Unknown  
 Injection Volume : 20 uL  
 Date Acquired : 2018/6/11 14:51:44 Acquired by : System Administrator  
 Date Processed : 2018/7/15 17:32:22 Processed by : System Administrator


## &lt;Chromatogram&gt;



## &lt;Peak Table&gt;

Detector A Channel 2 230nm				
Peak#	Ret. Time	Area	Height	Conc.
1	7.286	3891120	350426	98.489
2	9.289	59699	3341	1.511
Total		3950819	353766	

D:\Data\GXT\20180518\gxt-gd-52-asy-objh-90-10-1.0-1.lcd

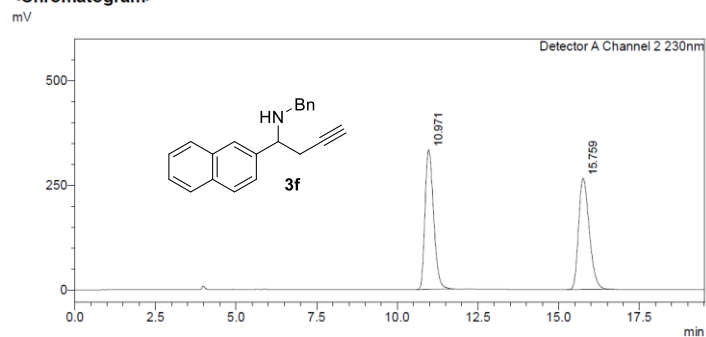

**Analysis Report**

## &lt;Sample Information&gt;

Sample Name : gxt-ge-63-rac-adh-99-1-0.8-  
 Sample ID :  
 Data Filename : gxt-ge-63-rac-adh-99-1-0.8-4.lcd  
 Method Filename : gxt-20170531.lcm  
 Batch Filename :  
 Vial # : 1-1  
 Injection Volume : 20 uL  
 Date Acquired : 2018/7/15 16:51:56  
 Date Processed : 2018/7/15 17:11:29

Sample Type : Unknown  
 Acquired by : System Administrator  
 Processed by : System Administrator


## &lt;Chromatogram&gt;



## &lt;Peak Table&gt;

Peak#	Ret. Time	Area	Height	Conc.
1	10.971	6212166	334440	50.145
2	15.759	6176124	265984	49.855
Total		12388290	600423	

D:\Data\GXT\20180518\gxt-ge-63-rac-adh-99-1-0.8-4.lcd

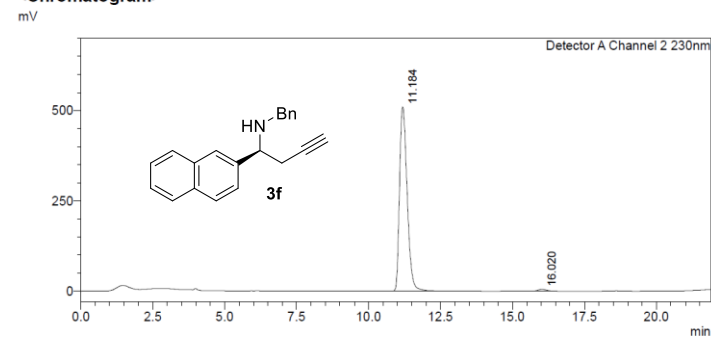

**Analysis Report**

## &lt;Sample Information&gt;

Sample Name : gxt-ge-63-asy-adh-99-1-0.8-  
 Sample ID :  
 Data Filename : gxt-ge-63-asy-adh-99-1-0.8-4.lcd  
 Method Filename : gxt-20170531.lcm  
 Batch Filename :  
 Vial # : 1-1  
 Injection Volume : 20 uL  
 Date Acquired : 2018/7/15 16:27:34  
 Date Processed : 2018/7/15 17:15:16

Sample Type : Unknown  
 Acquired by : System Administrator  
 Processed by : System Administrator


## &lt;Chromatogram&gt;



## &lt;Peak Table&gt;

Peak#	Ret. Time	Area	Height	Conc.
1	11.184	9387603	509846	99.063
2	16.020	88827	4894	0.937
Total		9476430	514740	

D:\Data\GXT\20180518\gxt-ge-63-asy-adh-99-1-0.8-4.lcd

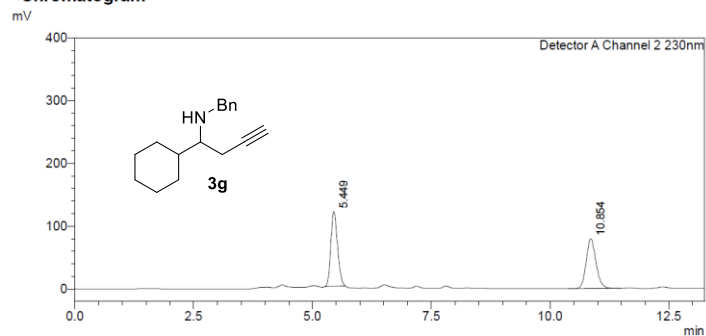

**Analysis Report**

## &lt;Sample Information&gt;

Sample Name : gxt-ge-20-rac-adh-97-3-0.8-  
 Sample ID :  
 Data Filename : gxt-ge-20-rac-adh-97-3-0.8-1.lcd  
 Method Filename : gxt-20170531.lcm  
 Batch Filename :  
 Vial # : 1-1  
 Injection Volume : 20 uL  
 Date Acquired : 2018/6/8 20:06:17  
 Date Processed : 2018/6/8 20:23:45

Sample Type : Unknown  
 Acquired by : System Administrator  
 Processed by : System Administrator

## &lt;Chromatogram&gt;




## &lt;Peak Table&gt;

Detector A Channel 2 230nm

Peak#	Ret. Time	Area	Height	Conc.
1	5.449	1167688	119250	50.490
2	10.854	1145007	79499	49.510
Total		2312696	198748	

D:\Data\GXT\20180518\gxt-ge-20-rac-adh-97-3-0.8-1.lcd

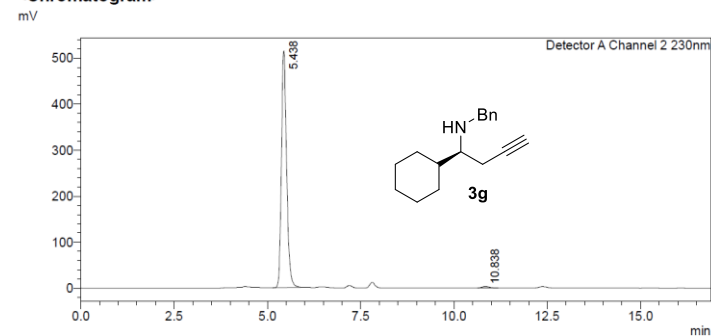

**Analysis Report**

## &lt;Sample Information&gt;

Sample Name : gxt-ge-20-asy-adh-97-3-0.8-  
 Sample ID :  
 Data Filename : gxt-ge-20-asy-adh-97-3-0.8-1.lcd  
 Method Filename : gxt-20170531.lcm  
 Batch Filename :  
 Vial # : 1-1  
 Injection Volume : 20 uL  
 Date Acquired : 2018/6/8 20:21:14  
 Date Processed : 2018/6/8 20:38:07

Sample Type : Unknown  
 Acquired by : System Administrator  
 Processed by : System Administrator

## &lt;Chromatogram&gt;



## &lt;Peak Table&gt;

Detector A Channel 2 230nm

Peak#	Ret. Time	Area	Height	Conc.
1	5.438	4994617	513707	99.063
2	10.838	47243	3401	0.937
Total		5041861	517109	

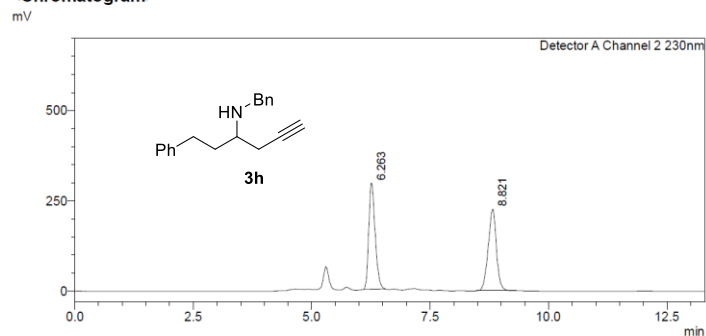
D:\Data\GXT\20180518\gxt-ge-20-asy-adh-97-3-0.8-1.lcd

SHIMADZU LabSolutions Analysis Report

## &lt;Sample Information&gt;

Sample Name : gxt-ge-21-rac-adh-80-20-0.7  
 Sample ID :  
 Data Filename : gxt-ge-21-rac-adh-80-20-0.8.lcd  
 Method Filename : gxt-20170531.lcm  
 Batch Filename :  
 Vial # : 1-1 Sample Type : Unknown  
 Injection Volume : 20 uL  
 Date Acquired : 2018/6/8 15:57:24 Acquired by : System Administrator  
 Date Processed : 2018/6/8 16:23:48 Processed by : System Administrator

## &lt;Chromatogram&gt;



## &lt;Peak Table&gt;

Peak#	Ret. Time	Area	Height	Conc.
1	6.263	2750170	293399	50.155
2	8.821	2733141	225926	49.845
Total		5483312	519325	

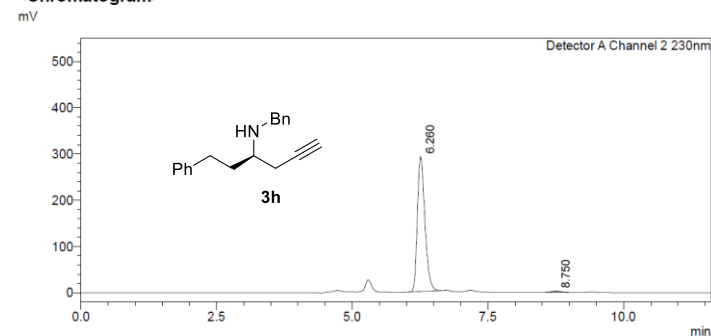
D:\Data\GXT\20180518\gxt-ge-21-rac-adh-80-20-0.8.lcd

SHIMADZU LabSolutions Analysis Report

## &lt;Sample Information&gt;

Sample Name : gxt-ge-21-asy-adh-80-20-0.7-second  
 Sample ID :  
 Data Filename : gxt-ge-21-asy-adh-80-20-0.7-second1.lcd  
 Method Filename : gxt-20170531.lcm  
 Batch Filename :  
 Vial # : 1-1 Sample Type : Unknown  
 Injection Volume : 20 uL  
 Date Acquired : 2018/6/8 16:31:45 Acquired by : System Administrator  
 Date Processed : 2018/6/8 16:45:25 Processed by : System Administrator

## &lt;Chromatogram&gt;



## &lt;Peak Table&gt;

Peak#	Ret. Time	Area	Height	Conc.
1	6.260	2795856	292020	98.718
2	8.750	36318	3291	1.282
Total		2832174	295311	

D:\Data\GXT\20180518\gxt-ge-21-asy-adh-80-20-0.7-second1.lcd

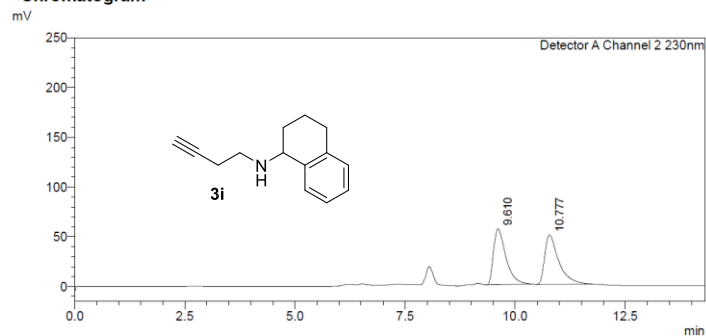


SHIMADZU LabSolutions Analysis Report

## &lt;Sample Information&gt;

Sample Name : gxt-ge-25-rac-adh-95-5-0.5-third-  
 Sample ID :  
 Data Filename : gxt-ge-25-rac-adh-95-5-0.5-third-1.lcd  
 Method Filename : gxt-20170531.lcm  
 Batch Filename :  
 Vial # : 1-1 Sample Type : Unknown  
 Injection Volume : 20 uL  
 Date Acquired : 2018/6/9 10:18:43 Acquired by : System Administrator  
 Date Processed : 2018/6/9 10:36:32 Processed by : System Administrator

## &lt;Chromatogram&gt;



## &lt;Peak Table&gt;

Detector A Channel 2 230nm

Peak#	Ret. Time	Area	Height	Conc.
1	9.610	1099207	56163	50.097
2	10.777	1094941	49649	49.903
Total		2194149	105812	

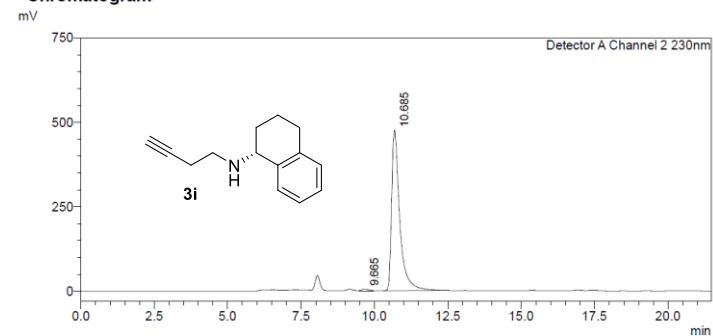
D:\Data\GXT\20180518\gxt-ge-25-rac-adh-95-5-0.5-third-1.lcd

SHIMADZU LabSolutions Analysis Report

## &lt;Sample Information&gt;

Sample Name : gxt-ge-25-asy-adh-95-5-0.5-third-  
 Sample ID :  
 Data Filename : gxt-ge-25-asy-adh-95-5-0.5-third-1.lcd  
 Method Filename : gxt-20170531.lcm  
 Batch Filename :  
 Vial # : 1-1 Sample Type : Unknown  
 Injection Volume : 20 uL  
 Date Acquired : 2018/6/9 10:34:33 Acquired by : System Administrator  
 Date Processed : 2018/6/9 11:00:03 Processed by : System Administrator

## &lt;Chromatogram&gt;



## &lt;Peak Table&gt;

Detector A Channel 2 230nm

Peak#	Ret. Time	Area	Height	Conc.
1	9.665	75121	5226	0.812
2	10.685	9174318	476109	99.188
Total		9249439	481334	

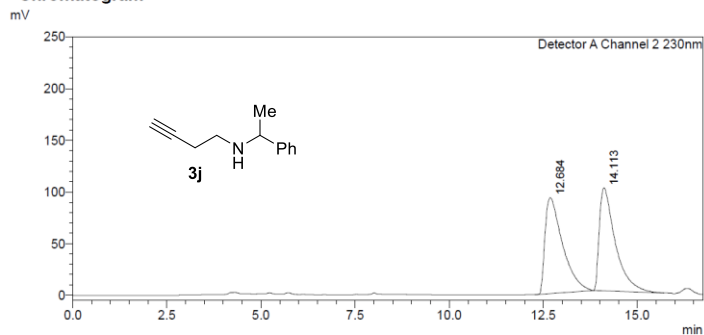
D:\Data\GXT\20180518\gxt-ge-25-asy-adh-95-5-0.5-third-1.lcd

SHIMADZU LabSolutions Analysis Report

## &lt;Sample Information&gt;

Sample Name : gxt-gd-136-rac-ojh-99.5-0.5-0.8-  
 Sample ID :  
 Data Filename : gxt-gd-136-rac-ojh-99.5-0.5-0.8-1.lcd  
 Method Filename : gxt-20170531.lcm  
 Batch Filename :  
 Vial # : 1-1 Sample Type : Unknown  
 Injection Volume : 20 uL  
 Date Acquired : 2018/6/9 16:51:31 Acquired by : System Administrator  
 Date Processed : 2018/6/9 17:08:17 Processed by : System Administrator

## &lt;Chromatogram&gt;



## &lt;Peak Table&gt;

Detector A Channel 2 230nm

Peak#	Ret. Time	Area	Height	Conc.
1	12.684	3004552	92785	50.568
2	14.113	2937019	99409	49.432
Total		5941571	192194	

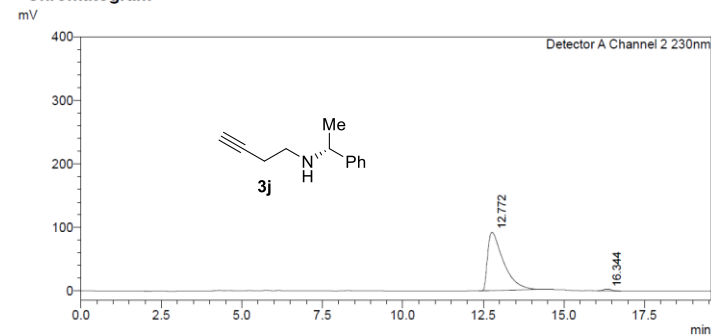
D:\Data\GXT\20180518\gxt-gd-136-rac-ojh-99.5-0.5-0.8-1.lcd

SHIMADZU LabSolutions Analysis Report

## &lt;Sample Information&gt;

Sample Name : gxt-gd-136-asy-ojh-99.5-0.5-0.8-  
 Sample ID :  
 Data Filename : gxt-gd-136-asy-ojh-99.5-0.5-0.8-2.lcd  
 Method Filename : gxt-20170531.lcm  
 Batch Filename :  
 Vial # : 1-1 Sample Type : Unknown  
 Injection Volume : 20 uL  
 Date Acquired : 2018/6/9 16:31:01 Acquired by : System Administrator  
 Date Processed : 2018/6/9 17:03:19 Processed by : System Administrator

## &lt;Chromatogram&gt;



Detector A Channel 2 230nm

Peak#	Ret. Time	Area	Height	Conc.
1	12.772	3143226	91929	98.567
2	16.344	45704	2456	1.433
Total		3188929	94386	

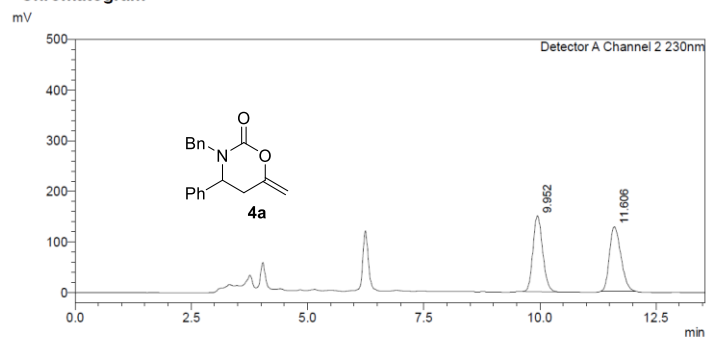
D:\Data\GXT\20180518\gxt-gd-136-asy-ojh-99.5-0.5-0.8-2.lcd

SHIMADZU LabSolutions Analysis Report

## &lt;Sample Information&gt;

Sample Name : gxt-gd-109-rac-adh-85-15-1.0  
 Sample ID :  
 Data Filename : gxt-gd-109-rac-adh-85-15-1.0.lcd  
 Method Filename : gxt-20170531.lcm  
 Batch Filename :  
 Vial # : 1-1 Sample Type : Unknown  
 Injection Volume : 20 uL  
 Date Acquired : 2018/5/18 14:46:16 Acquired by : System Administrator  
 Date Processed : 2018/5/18 16:29:53 Processed by : System Administrator

## &lt;Chromatogram&gt;



## &lt;Peak Table&gt;

Detector A Channel 2 230nm				
Peak#	Ret. Time	Area	Height	Conc.
1	9.952	2175245	149744	50.001
2	11.606	2175126	127011	49.999
Total		4350371	276755	

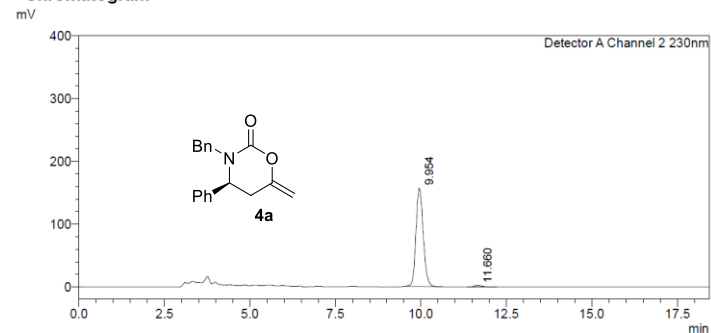
D:\Data\GXT\20180518\gxt-gd-109-rac-adh-85-15-1.0.lcd

SHIMADZU LabSolutions Analysis Report

## &lt;Sample Information&gt;

Sample Name : gxt-gd-109-asy-adh-85-15-1.0  
 Sample ID :  
 Data Filename : gxt-gd-109-asy-adh-85-15-1.0.lcd  
 Method Filename : gxt-20170531.lcm  
 Batch Filename :  
 Vial # : 1-1 Sample Type : Unknown  
 Injection Volume : 20 uL  
 Date Acquired : 2018/5/18 15:03:06 Acquired by : System Administrator  
 Date Processed : 2018/5/18 16:21:36 Processed by : System Administrator

## &lt;Chromatogram&gt;



## &lt;Peak Table&gt;

Detector A Channel 2 230nm				
Peak#	Ret. Time	Area	Height	Conc.
1	9.954	2317992	156613	98.254
2	11.660	41183	2431	1.746
Total		2359174	159044	

D:\Data\GXT\20180518\gxt-gd-109-asy-adh-85-15-1.0.lcd

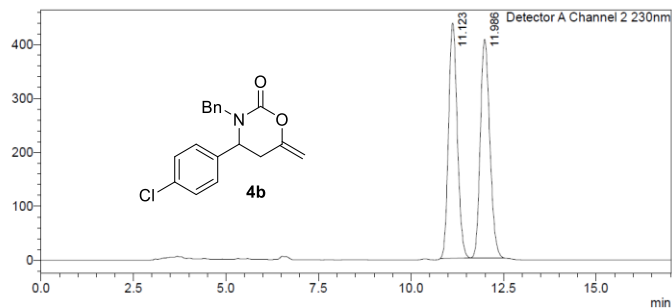
SHIMADZU LabSolutions Analysis Report

## &lt;Sample Information&gt;

Sample Name : gxt-gd-129-rac-adh-85-15-1.0-3  
 Sample ID :  
 Data Filename : gxt-gd-129-rac-adh-85-15-1.0-3.lcd  
 Method Filename : gxt-20170531.lcm  
 Batch Filename :  
 Vial # : 1-1 Sample Type : Unknown  
 Injection Volume : 20 uL  
 Date Acquired : 2018/5/19 15:28:28 Acquired by : System Administrator  
 Date Processed : 2018/5/19 15:45:30 Processed by : System Administrator

## &lt;Chromatogram&gt;

mV



## &lt;Peak Table&gt;

Detector A Channel 2 230nm

Peak#	Ret. Time	Area	Height	Conc.
1	11.123	7166656	436051	49.782
2	11.986	7229294	405070	50.218
Total		14395950	841121	

D:\Data\GXT\20180518\gxt-gd-129-rac-adh-85-15-1.0-3.lcd

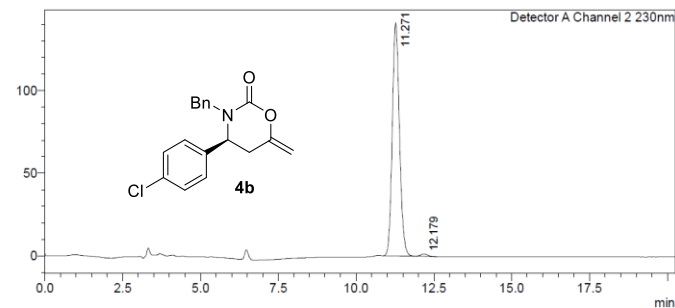
SHIMADZU LabSolutions Analysis Report

## &lt;Sample Information&gt;

Sample Name : gxt-gd-129-asy-adh-85-15-0.8-1  
 Sample ID :  
 Data Filename : gxt-gd-129-asy-adh-85-15-0.8-1.lcd  
 Method Filename : gxt-20170531.lcm  
 Batch Filename :  
 Vial # : 1-1 Sample Type : Unknown  
 Injection Volume : 20 uL  
 Date Acquired : 2018/5/19 9:37:01 Acquired by : System Administrator  
 Date Processed : 2018/5/19 9:57:17 Processed by : System Administrator

## &lt;Chromatogram&gt;

mV



## &lt;Peak Table&gt;

Detector A Channel 2 230nm

Peak#	Ret. Time	Area	Height	Conc.
1	11.271	2273484	140763	98.998
2	12.179	23003	1467	1.002
Total		2296487	142230	

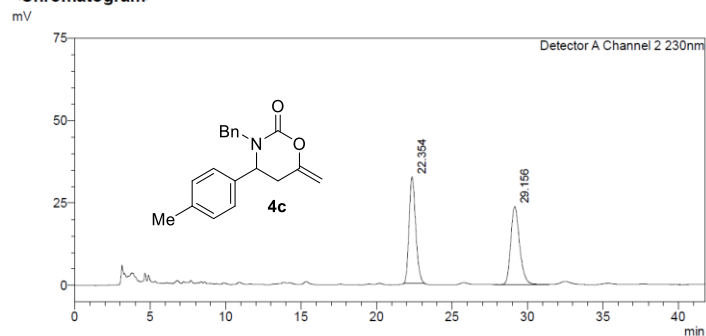
D:\Data\GXT\20180518\gxt-gd-129-asy-adh-85-15-0.8-1.lcd

SHIMADZU LabSolutions Analysis Report

## &lt;Sample Information&gt;

Sample Name : gxt-ge-53-chanwu-rac-adh-95-5-1.0-  
 Sample ID :  
 Data Filename : gxt-ge-53-chanwu-rac-adh-95-5-1.0-2.lcd  
 Method Filename : gxt-20170531.lcm  
 Batch Filename :  
 Vial # : 1-1 Sample Type : Unknown  
 Injection Volume : 20 uL  
 Date Acquired : 2018/7/1 13:37:14 Acquired by : System Administrator  
 Date Processed : 2018/7/1 14:19:00 Processed by : System Administrator

## &lt;Chromatogram&gt;



## &lt;Peak Table&gt;

Detector A Channel 2 230nm				
Peak#	Ret. Time	Area	Height	Conc.
1	22.354	983950	32312	50.442
2	29.156	966697	23656	49.558
Total		1950647	55968	

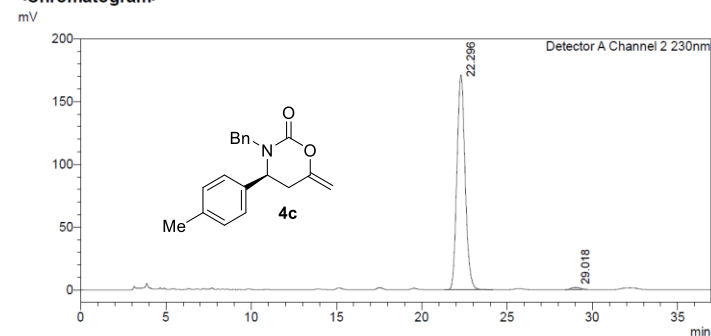
D:\Data\GXT\20180518\gxt-ge-53-chanwu-rac-adh-95-5-1.0-2.lcd

SHIMADZU LabSolutions Analysis Report

## &lt;Sample Information&gt;

Sample Name : gxt-ge-53-chanwu-asy-adh-95-5-1.0-  
 Sample ID :  
 Data Filename : gxt-ge-53-chanwu-asy-adh-95-5-1.0-1.lcd  
 Method Filename : gxt-20170531.lcm  
 Batch Filename :  
 Vial # : 1-1 Sample Type : Unknown  
 Injection Volume : 20 uL  
 Date Acquired : 2018/7/1 14:20:06 Acquired by : System Administrator  
 Date Processed : 2018/7/1 14:57:04 Processed by : System Administrator

## &lt;Chromatogram&gt;



## &lt;Peak Table&gt;

Detector A Channel 2 230nm				
Peak#	Ret. Time	Area	Height	Conc.
1	22.296	5736455	171183	98.952
2	29.018	60754	1703	1.048
Total		5797208	172886	

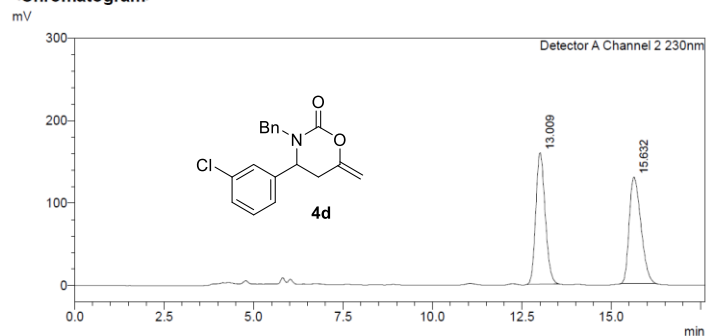
D:\Data\GXT\20180518\gxt-ge-53-chanwu-asy-adh-95-5-1.0-1.lcd

SHIMADZU LabSolutions Analysis Report

## &lt;Sample Information&gt;

Sample Name : gxt-gd-131-rac-adh-85-15-0.8-  
 Sample ID :  
 Data Filename : gxt-gd-131-rac-adh-85-15-0.8-1.lcd  
 Method Filename : gxt-20170531.lcm  
 Batch Filename :  
 Vial # : 1-1 Sample Type : Unknown  
 Injection Volume : 20 uL  
 Date Acquired : 2018/6/11 19:51:52 Acquired by : System Administrator  
 Date Processed : 2018/6/11 20:09:29 Processed by : System Administrator

## &lt;Chromatogram&gt;



## &lt;Peak Table&gt;

Detector A Channel 2 230nm				
Peak#	Ret. Time	Area	Height	Conc.
1	13.009	2909273	159564	49.618
2	15.632	2954066	129187	50.382
Total		5863339	288751	

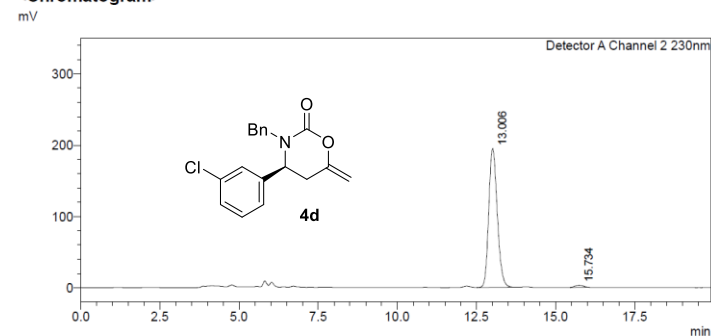
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SHIMADZU LabSolutions Analysis Report

## &lt;Sample Information&gt;

Sample Name : gxt-gd-131-asy-adh-85-15-0.8-  
 Sample ID :  
 Data Filename : gxt-gd-131-asy-adh-85-15-0.8-2.lcd  
 Method Filename : gxt-20170531.lcm  
 Batch Filename :  
 Vial # : 1-1 Sample Type : Unknown  
 Injection Volume : 20 uL  
 Date Acquired : 2018/6/11 20:10:58 Acquired by : System Administrator  
 Date Processed : 2018/6/11 20:30:53 Processed by : System Administrator

## &lt;Chromatogram&gt;



## &lt;Peak Table&gt;

Detector A Channel 2 230nm				
Peak#	Ret. Time	Area	Height	Conc.
1	13.006	3661449	194900	98.537
2	15.734	54344	3003	1.463
Total		3715793	197904	

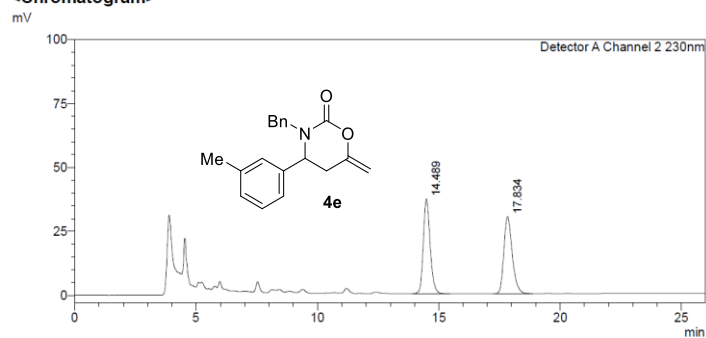
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SHIMADZU LabSolutions Analysis Report

## &lt;Sample Information&gt;

Sample Name : gxt-gd-56-rac-adh-90-10-0.8-  
 Sample ID :  
 Data Filename : gxt-gd-56-rac-adh-90-10-0.8-1.lcd  
 Method Filename : gxt-20170531.lcm  
 Batch Filename :  
 Vial # : 1-1 Sample Type : Unknown  
 Injection Volume : 20 uL  
 Date Acquired : 2018/7/15 18:22:07 Acquired by : System Administrator  
 Date Processed : 2018/7/15 18:48:07 Processed by : System Administrator

## &lt;Chromatogram&gt;



## &lt;Peak Table&gt;

Peak#	Ret. Time	Area	Height	Conc.
1	14.489	722771	37139	50.004
2	17.834	722651	30118	49.996
Total		1445422	67257	

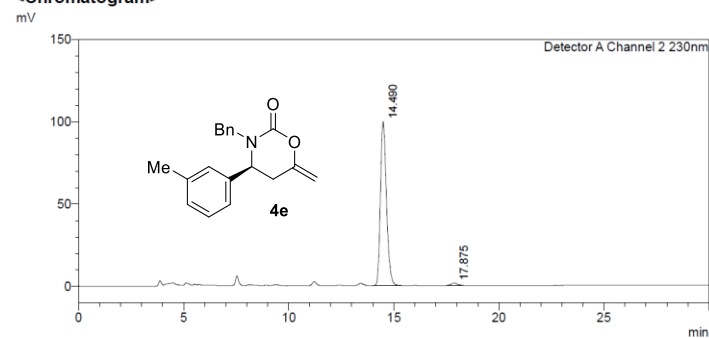
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SHIMADZU LabSolutions Analysis Report

## &lt;Sample Information&gt;

Sample Name : gxt-gd-56-asy-adh-90-10-0.8-  
 Sample ID :  
 Data Filename : gxt-gd-56-asy-adh-90-10-0.8-1.lcd  
 Method Filename : gxt-20170531.lcm  
 Batch Filename :  
 Vial # : 1-1 Sample Type : Unknown  
 Injection Volume : 20 uL  
 Date Acquired : 2018/7/15 18:49:32 Acquired by : System Administrator  
 Date Processed : 2018/7/15 19:19:33 Processed by : System Administrator

## &lt;Chromatogram&gt;



## &lt;Peak Table&gt;

Peak#	Ret. Time	Area	Height	Conc.
1	14.490	2002005	99648	98.499
2	17.875	30510	1408	1.501
Total		2032515	101056	

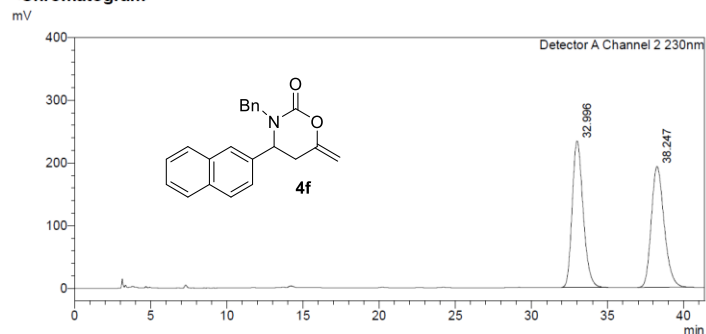
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SHIMADZU LabSolutions Analysis Report

## &lt;Sample Information&gt;

Sample Name : gxt-ge-68-chanwu-rac-adh-95-5-1.0-  
 Sample ID :  
 Data Filename : gxt-ge-68-chanwu-rac-adh-95-5-1.0-1.lcd  
 Method Filename : gxt-20170531.lcm  
 Batch Filename :  
 Vial # : 1-1 Sample Type : Unknown  
 Injection Volume : 20 uL  
 Date Acquired : 2018/7/1 11:31:29 Acquired by : System Administrator  
 Date Processed : 2018/7/1 12:12:53 Processed by : System Administrator

## &lt;Chromatogram&gt;



## &lt;Peak Table&gt;

Detector A Channel 2 230nm				
Peak#	Ret. Time	Area	Height	Conc.
1	32.996	11669008	233713	50.661
2	38.247	11364673	192854	49.339
Total		23033681	426567	

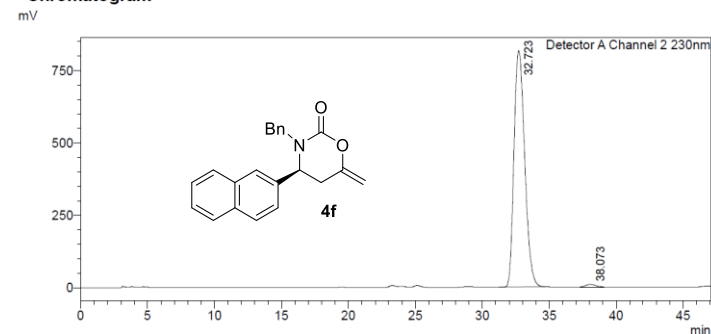
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SHIMADZU LabSolutions Analysis Report

## &lt;Sample Information&gt;

Sample Name : gxt-ge-68-chanwu-asy-adh-95-5-1.0-  
 Sample ID :  
 Data Filename : gxt-ge-68-chanwu-asy-adh-95-5-1.0-1.lcd  
 Method Filename : gxt-20170531.lcm  
 Batch Filename :  
 Vial # : 1-1 Sample Type : Unknown  
 Injection Volume : 20 uL  
 Date Acquired : 2018/7/1 12:14:08 Acquired by : System Administrator  
 Date Processed : 2018/7/1 13:01:13 Processed by : System Administrator

## &lt;Chromatogram&gt;



## &lt;Peak Table&gt;

Detector A Channel 2 230nm				
Peak#	Ret. Time	Area	Height	Conc.
1	32.723	45615020	816287	98.997
2	38.073	462150	8982	1.003
Total		46077169	825269	

D:\Data\GXT\20180518\gxt-ge-68-chanwu-asy-adh-95-5-1.0-1.lcd



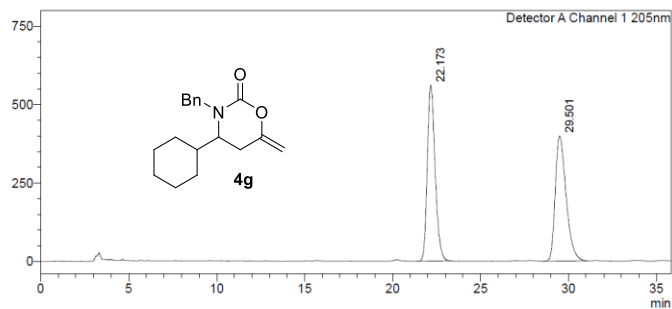
SHIMADZU LabSolutions Analysis Report

## &lt;Sample Information&gt;

Sample Name : gxt-ge-31-chanwu-rac-adh-95-5-1.0-  
 Sample ID :  
 Data Filename : gxt-ge-31-chanwu-rac-adh-95-5-1.0-1.lcd  
 Method Filename : gxt-20170531.lcm  
 Batch Filename :  
 Vial # : 1-1 Sample Type : Unknown  
 Injection Volume : 20 uL  
 Date Acquired : 2018/7/1 15:45:31 Acquired by : System Administrator  
 Date Processed : 2018/7/14 15:22:54 Processed by : System Administrator

## &lt;Chromatogram&gt;

mV



## &lt;Peak Table&gt;

Detector A Channel 1 205nm

Peak#	Ret. Time	Area	Height	Conc.
1	22.173	17318551	561344	50.421
2	29.501	17029373	398078	49.579
Total		34347925	959422	

D:\Data\GXT\20180518\gxt-ge-31-chanwu-rac-adh-95-5-1.0-1.lcd

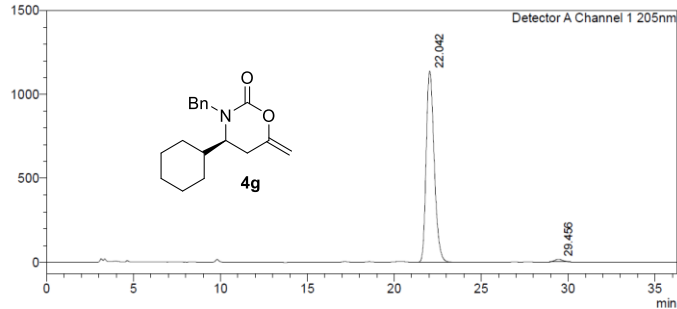
SHIMADZU LabSolutions Analysis Report

## &lt;Sample Information&gt;

Sample Name : gxt-ge-31-chanwu-asy-adh-95-5-1.0-  
 Sample ID :  
 Data Filename : gxt-ge-31-chanwu-asy-adh-95-5-1.0-1.lcd  
 Method Filename : gxt-20170531.lcm  
 Batch Filename :  
 Vial # : 1-1 Sample Type : Unknown  
 Injection Volume : 20 uL  
 Date Acquired : 2018/7/1 16:22:53 Acquired by : System Administrator  
 Date Processed : 2018/7/14 15:25:31 Processed by : System Administrator

## &lt;Chromatogram&gt;

mV




## &lt;Peak Table&gt;

Detector A Channel 1 205nm

Peak#	Ret. Time	Area	Height	Conc.
1	22.042	36766813	1139500	98.545
2	29.456	542713	15557	1.455
Total		37309526	1155056	

D:\Data\GXT\20180518\gxt-ge-31-chanwu-asy-adh-95-5-1.0-1.lcd



# Analysis Report

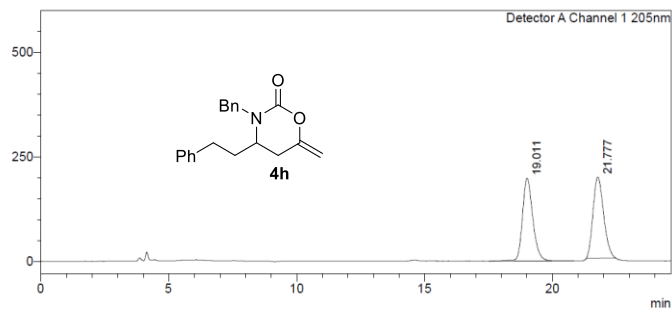
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Sample Name : gxt-ge-34-rac-adh-90-10-0.8-  
 Sample ID :  
 Data Filename : gxt-ge-34-rac-adh-90-10-0.8-2.lcd  
 Method Filename : gxt-20170531.lcm  
 Batch Filename :  
 Vial # : 1-1  
 Injection Volume : 20 uL  
 Date Acquired : 2018/7/15 20:17:23  
 Date Processed : 2018/7/15 20:42:03

Sample Type : Unknown  
 Acquired by : System Administrator  
 Processed by : System Administrator

**<Chromatogram>**


mV

**<Peak Table>**

Detector A Channel 1 205nm

Peak#	Ret. Time	Area	Height	Conc.
1	19.011	5571414	197753	49.943
2	21.777	5584072	194267	50.057
Total		11155486	392020	

D:\Data\GXT\20180518\gxt-ge-34-rac-adh-90-10-0.8-2.lcd



# Analysis Report

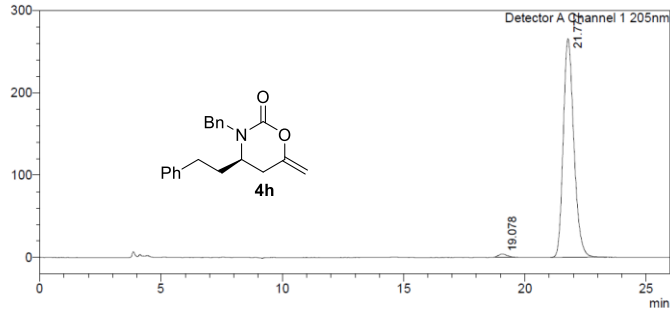
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 Sample ID :  
 Data Filename : gxt-ge-34-asy-adh-90-10-0.8-2.lcd  
 Method Filename : gxt-20170531.lcm  
 Batch Filename :  
 Vial # : 1-1  
 Injection Volume : 20 uL  
 Date Acquired : 2018/7/15 20:43:34  
 Date Processed : 2018/7/15 21:09:33

Sample Type : Unknown  
 Acquired by : System Administrator  
 Processed by : System Administrator

**<Chromatogram>**


mV

**<Peak Table>**

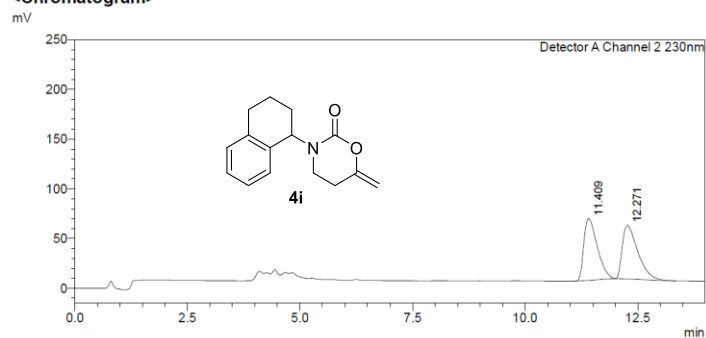
Detector A Channel 1 205nm

Peak#	Ret. Time	Area	Height	Conc.
1	19.078	79563	3436	0.976
2	21.777	8069762	265384	99.024
Total		8149325	268820	

D:\Data\GXT\20180518\gxt-ge-34-asy-adh-90-10-0.8-2.lcd



**Analysis Report**
**<Sample Information>**

Sample Name : gxt-ge-35-rac-ogh-85-15-0.8-  
 Sample ID :  
 Data Filename : gxt-ge-35-rac-ogh-85-15-0.8-1.lcd  
 Method Filename : gxt-20170531.lcm  
 Batch Filename :  
 Vial # : 1-1                      Sample Type : Unknown  
 Injection Volume : 20 uL  
 Date Acquired : 2018/7/15 22:28:39              Acquired by : System Administrator  
 Date Processed : 2018/7/15 22:42:40              Processed by : System Administrator

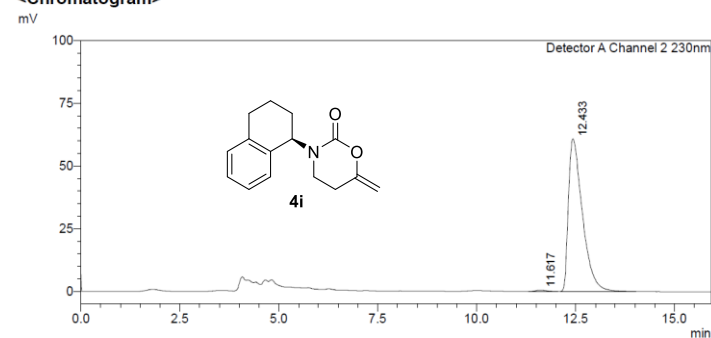
**<Chromatogram>****<Peak Table>**

Detector A Channel 2 230nm				
Peak#	Ret. Time	Area	Height	Conc.
1	11.409	1286051	62105	49.876
2	12.271	1292456	54032	50.124
Total		2578507	116136	

D:\Data\GXT\20180518\gxt-ge-35-rac-ogh-85-15-0.8-1.lcd


**Analysis Report**
**<Sample Information>**

Sample Name : gxt-ge-35-asy-ogh-85-15-0.8-  
 Sample ID :  
 Data Filename : gxt-ge-35-asy-ogh-85-15-0.8-1.lcd  
 Method Filename : gxt-20170531.lcm  
 Batch Filename :  
 Vial # : 1-1                      Sample Type : Unknown  
 Injection Volume : 20 uL  
 Date Acquired : 2018/7/15 22:44:13              Acquired by : System Administrator  
 Date Processed : 2018/7/15 23:00:08              Processed by : System Administrator

**<Chromatogram>****<Peak Table>**

Detector A Channel 2 230nm				
Peak#	Ret. Time	Area	Height	Conc.
1	11.617	11883	637	0.786
2	12.433	1499239	60894	99.214
Total		1511122	61531	

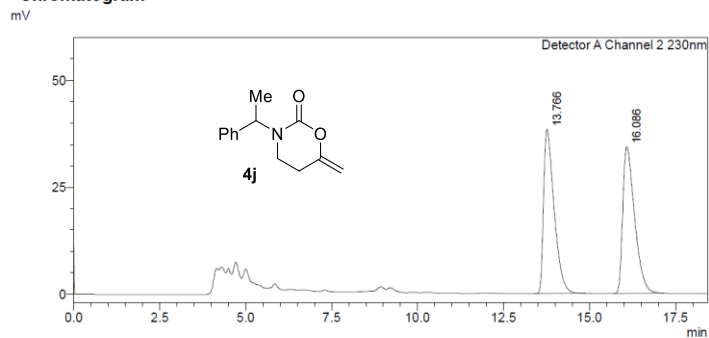
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SHIMADZU LabSolutions Analysis Report

## &lt;Sample Information&gt;

Sample Name : gxt-gd-141-rac-ojh-85-15-0.8-  
 Sample ID :  
 Data Filename : gxt-gd-141-rac-ojh-85-15-0.8-1.lcd  
 Method Filename : gxt-20170531.lcm  
 Batch Filename :  
 Vial # : 1-1 Sample Type : Unknown  
 Injection Volume : 20 uL  
 Date Acquired : 2018/6/11 21:22:10 Acquired by : System Administrator  
 Date Processed : 2018/6/11 21:40:37 Processed by : System Administrator

## &lt;Chromatogram&gt;



## &lt;Peak Table&gt;

Detector A Channel 2 230nm				
Peak#	Ret. Time	Area	Height	Conc.
1	13.766	817478	38339	49.831
2	16.086	823029	34288	50.169
Total		1640507	72627	

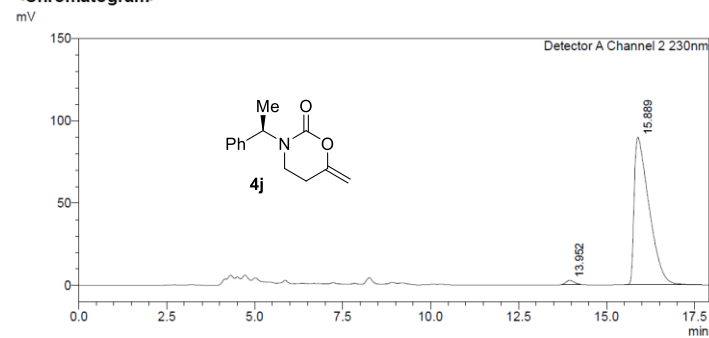
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SHIMADZU LabSolutions Analysis Report

## &lt;Sample Information&gt;

Sample Name : gxt-gd-141-asy-ojh-85-15-0.8-  
 Sample ID :  
 Data Filename : gxt-gd-141-asy-ojh-85-15-0.8-1.lcd  
 Method Filename : gxt-20170531.lcm  
 Batch Filename :  
 Vial # : 1-1 Sample Type : Unknown  
 Injection Volume : 20 uL  
 Date Acquired : 2018/6/11 21:42:28 Acquired by : System Administrator  
 Date Processed : 2018/6/11 22:00:23 Processed by : System Administrator

## &lt;Chromatogram&gt;



## &lt;Peak Table&gt;

Detector A Channel 2 230nm				
Peak#	Ret. Time	Area	Height	Conc.
1	13.952	40699	2495	1.520
2	15.889	2636145	89529	98.480
Total		2676844	92024	

D:\Data\GXT\20180518\gxt-gd-141-asy-ojh-85-15-0.8-1.lcd

## 8. References

- [1] P. Quinodoz, A. Quelhas, K. Wright, B. Drouillat, J. Marrot and F. Couty, *Eur. J. Org. Chem.*, 2017, 2621–2626.
- [2] (a) A. Fürstner, O. Guth, A. Düffels, G. Seidel, M. Liebl, B. Gabor and R. Mynott, *Chem. Eur. J.*, 2001, **7**, 4811–4820; (b) Y. Hirai, T. Terada, T. Yamazaki and T. Momose, *J. Chem. Soc., Perkin Trans. 1*, 1992, **0**, 509–516.
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