Supporting Information

for

NAMI-A Preferentially Reacts with Sp1 protein: Understanding Anti-metastasis Effect of the Drug

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Experimental details

Protein Expression and Purification: Proteins used in this work were obtained according to the literature¹ and were characterized by electrophoresis and electrospray ionization mass spectrometry (ESI-MS).

ESI-MS Analysis of the Reaction of NAMI-A with Sp1: The mass spectrometric analysis was carried out on an Exactive Plusmass spectrometer (Thermo Fisher Scientific). 10 μ M Sp1-zf2 was incubated with 2 molar equivalents of NAMI-A in 100 mM ammonium acetate at 37 °C for 12 h. Then samples were directly infused. Data were processed using XCalibur software (version2.0, Thermo Finnigan).

Fluorescence Measurements: Fluorescence spectra were recorded on a Hitachi F-4600 fluorescence spectrophotometer using a quartz cuvette with the path length of 1 cm. The reactions were conducted on 10 μ M proteins with NAMI-A in 10 mM HEPES buffer.

Electrophoretic Mobility Shift Assay: 20 μ M of Sp1(530-623) was incubated with 10 μ M *surviving* promoter in gel-shift buffer (20 mM HEPES 100 mM NaCl and 500 μ M GSH at pH 7.40) for 0.5 h. Then the complexes were incubated with different concentration of NAMI-A at room temperature for 4 h. Samples were loaded on a native 15% polyacrylamide gel and run in 0.5×Tris-borate buffer. The gel was stained with Ethidium Bromide and visualized under UV.

Circular Dichroism (CD): CD measurements were performed on a Jasco J-810 CD spectrometer using a 1.0 cm path length quartz cuvette. 30 μ M Proteins were incubated with different molar equivalents of NAMI-A in 10 mM phosphate buffer (pH 7.0) at 37 °C for 24 h. Spectra were scanned from 280 nm to 190 nm with a speed of 100 nm·min⁻¹; each spectrum was repeated three times.

NMR Spectroscopy: NMR spectra were recorded on a Bruker Avance 600 MHz spectrometer equipped with TCI CryoProbe. The samples were performed on 0.3 mM ¹⁵N-labeled Sp1-zf2 with 2 molar equivalents of NAMI-A in 20 mM HEPES buffer containing 100 mM NaCl. Data were processed and analyzed using NMRPipe.

References

1. S. Chen, D. Xu, H. Jiang, Z. Xi, P. Zhu and Y. Liu, Angew. Chem. Int. Ed., 2012, **51**, 12258-12262.



Scheme S1. The structure of NAMI-A and the sequence of zinc-finger domain of Sp1.



Figure S1. Fluorescence Characterization of NAMI-A binding to Sp1. (A) The fluorescence spectra of Sp1 after reaction with NAMI-A. (B) Plot of the fluorescence intensity of Sp1 at 350 nm in the reaction of NAMI-A. 10 μ M protein was incubated with NAMI-A at 37 °C for 24 h in 10 mM HEPES, pH 6.80.



Figure S2. Kinetic process of the reaction of Sp1-zf2 with NAMI-A. Time-dependent measurement of the fluorescence intensity of Sp1-zf2 during the reaction with NAMI-A. The reaction was performed on 10 μ M Sp1-zf2 with 2 molar equivalents of NAMI-A at 37 °C.



Figure S3. Characterization the reaction of Sp1-zf2 with NAMI-A. (A) Cationic ion exchange chromatography analysis the reaction of NAMI-A with Sp1-zf2. Sp1-zf2 (46 μ M) was incubated with NAMI-A at 37 °C for 24h in 50 mM HEPES, pH 6.8. (B) Thiol content in Sp1-zf2 measured by DTNB after the reaction with NAMI-A. Reactions were carried out on Sp1-zf2 (100 μ M) with different amount of NAMI-A in 100 mM HEPES buffer (pH 6.8) at 37 °C for 24 h. A 100 μ M DTNB was added to the reaction mixture. The HPLC profiles were recorded on C18 reversed-phase column.



Figure S4. Zinc release from Sp1-zf2 upon the reaction of NAMI-A. Reactions were carried out on Sp1-zf2 (30 μ M) with 2 molar equivalents of NAMI-A at 37 °C.



Figure S5. (A) Overlay of NMR spectra of ¹⁵N-labeled Sp1-zf2 before (blue) and after (red) incubation with 2 molar equivalents of natural isotopic abundance Ru-NCp7. Ru-NCp7 was prepared by the NCp7 reaction with NAMI-A. NMR spectra were recorded at 298 K in 20 mM HEPES buffer (pH 6.8) containing 100 mM NaCl. (B). Overlay of NMR spectra of ¹⁵N-labeled NCp7 before (blue) and after (red) incubation with 2 molar equivalents of natural isotopic abundance Ru- Sp1-zf2. Ru- Sp1-zf2 was prepared by the Sp1-zf2 reaction with NAMI-A. NMR spectra were recorded at 298 K in 20 mM HEPES buffer (pH 6.8) containing 100 mM NaCl.



Figure S6. Characterization the effect of GSH on the reaction of NAMI-A with Sp1. Fluorescence spectra of Sp1 in the reaction with different amount of NAMI-A in the absence or presence of GSH. 10 μ M protein was incubated with NAMI-A in the absence or presence of 1 mM GSH at 37 °C for 24 h in 10 mM HEPES, pH 6.80.



Figure S7. HPLC profiles of Sp1-zf2 in the reaction with NAMI-A. HPLC profiles of Sp1-zf2 in the reaction with NAMI-A In the absence (A) or presence (B) of 1 mM GSH. Sp1-zf2 (100 μ M) was incubated with NAMI-A at 37 °C for 48h in 100 mM HEPES, pH 6.8. HPLC analyses were performed with an Agilent 1200 system equipped with a Kromasil C18 reverse phase column and recorded by UV detection at 280 nm.



Figure S8. ESI-MS spectra of Sp1-zf2 reacted with 2 molar equivalents of NAMI-A in the absence or presence of 5 molar equivalents of GSH at 37 °C for 12h. 10 μ M Sp1-zf2 was incubated with 2 molar equivalents of NAMI-A at 37 °C for 12h in 100 mM NH₄OAc buffer.



Figure S9.Time-dependent measurement of the fluorescence intensity of Sp1-zf2 during the reaction with NAMI-A in the presence of GSH (blue) or with GSH/NAMI-A adducts (red). The reaction was performed on 10 μ M Sp1-zf2 with 4 molar equivalents of NAMI-A at 37 °C.



Figure S10. Time-dependent NMR spectra of Sp1-zf2 in the reaction with NAMI-A. (A) Overlay of NMR spectra of Sp1-zf2 before (red) and after incubation with 2 molar equivalents of NAMI-A at 25 °C for 2 h (blue), 4 h (green), 6 h (gold), 8 h (magenta) and 10 h (black) in 20 mM HEPES buffer containing 100 mM NaCl (pH 6.8). (B) the partial enlarged enlargement of A.



Figure S11. Overlay of NMR spectra of Sp1-zf2 before (red) and after (red) incubation with 2 molar equivalents of NAMI-A at 25 °C for 10 h in 20 mM HEPES buffer containing 100 mM NaCl (pH 6.8) in the presence of 2 mM GSH.



Figure S12. Tricine SDS-PAGE analyses the effect of GSH on the aggregation of Sp1-zf2 induced by NAMI-A. Lanes 1-8: Sp1-zf2 (80μ M) was incubated with different molar equivalents of NAMI-A (0, 0.5, 2, 3, 4, 6, 8 and 10, respectively) at 37 °C for 12h in 100 mM HEPES, pH 6.8. Reactions were performed without (A) or with (B) 1 mM GSH addition.



Figure S13. Characterization the reaction of DNA with NAMI-A. Lane 1: *survivin* promoter; Lanes 2-9: DNA was incubated with different molar equivalents of NAMI-A (2, 4, 6, 8, 10, 15, 20 and 30, respectively).