

Electronic supporting information

***N*-Protecting Group Tuning the Enantioselectivity in Strecker Reactions of Trifluoromethyl Ketimines to Synthesize Quaternary α - Trifluoromethyl Amino Nitriles by Ion Pair Catalysis**

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I. General information

All solvents used in the reaction were commercial available without further purification. Purification of reaction products was carried out by flash chromatography using silica gel (300 – 400 mesh). TLC was visualized by UV fluorescence (254 nm) or KMnO_4 , phosphomolybdic acid. The ^1H NMR spectra were recorded on a Bruker 400 or an Agilent 400 (400 MHz). All chemical shifts (δ) were given in ppm. Data were reported as follows: chemical shift, integration, multiplicity (s = single, d = doublet, t = triplet, q = quartet, br = broad, m = multiplet) and coupling constants (Hz). ^{13}C NMR spectra were recorded on a Bruker 400 (100 MHz) or an Agilent 500 (126 MHz). ^{19}F NMR spectra were recorded on an Agilent 400 (376 MHz). ^{31}P NMR spectra were recorded on an Agilent 400 (163 MHz). High-resolution mass spectra (HRMS) were recorded on a Bruker APEXIII 7.0 Tesla ESI – FT mass spectrometer or an Agilent TOF/LC-MS 1260-6230 mass spectrometer. Melting points were determined on a SGW X-4 melting point and were uncorrected. Optical rotations were measured on an Anton Paar MCP 5500 Polarimeter at $\lambda = 589$ nm. IR spectra were recorded on a Perkin-Elmer 983G instrument. All bifunctional phosphine catalysts were synthesized according to procedures reported previously.^{1,2}

II. General experimental procedure

2.1 General procedure for the preparation of ketimines

To a solution of corresponding trifluoromethyl ketone (2.0 mmol, 1.0 equiv) in toluene (5.0 mL) was added *N*-Boc-imino-(triphenyl)-phosphorane (4.0 mmol, 2.0 equiv.). The system was stirred at reflux until the completion of reaction (monitored by TLC). Then the system was cooled down to room temperature and the solvent was removed in vacuum. The residue was purified by silica gel column chromatography to give corresponding *N*-Boc ketimines. The *N*-PMP ketimines were prepared according to the reported literature.^{3,7}

2.2 General procedure for the asymmetric Strecker reaction

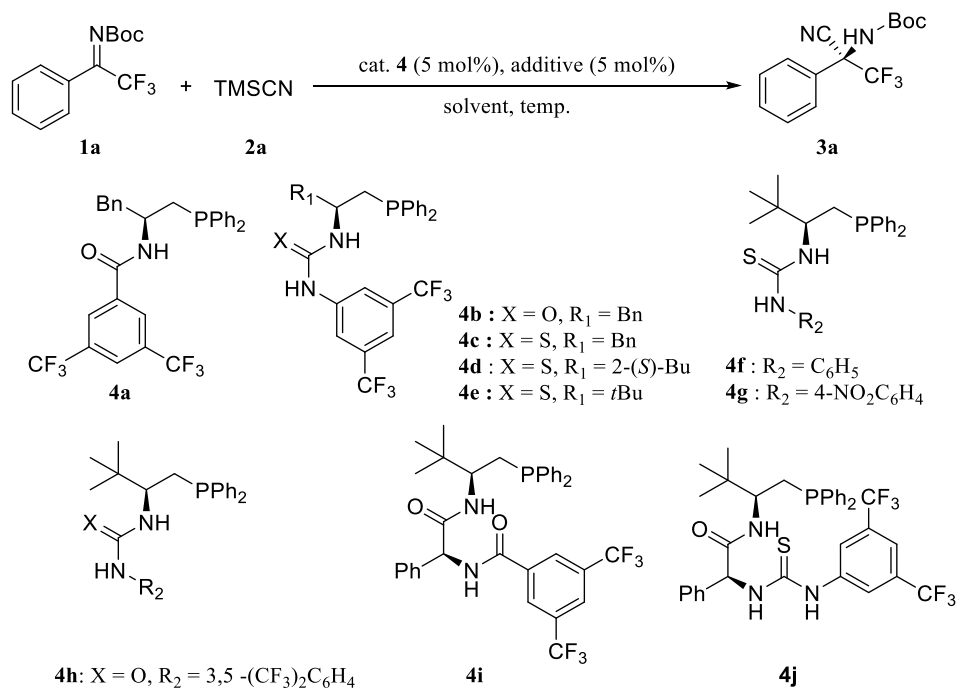
General procedure for the asymmetric Strecker reaction of *N*-PMP ketimines

To a solution of organophosphine catalyst (0.005 mmol, 5 mol%) in dichloromethane (1.0 mL) was added methyl acrylate (0.005 mmol, 5 mol%). The system was stirred at room temperature for ten minutes, then TMSCN (0.2 mmol) was added. Five minutes later the system was cooled to -20 °C and after another ten minutes corresponding ketimine (0.1 mmol) was added. After the completion of reaction (monitored by TLC) the system was allowed to warm to room temperature and the solvent was removed in vacuum. The residue was purified by silica gel column chromatography to give the pure product.

General procedure for the asymmetric Strecker reaction of *N*-Boc ketimines

To a solution of organophosphine catalyst (0.005 mmol, 5 mol%) in dichloromethane (1.0 mL) was added methyl acrylate (0.005 mmol, 5 mol%). The system was stirred at room temperature for ten minutes and then corresponding ketimine (0.1 mmol) was added. Ten minutes later the system was cooled to -72 °C and after another ten minutes TMSCN (0.2 mmol) was added. After the completion of reaction (monitored by TLC) the system was allowed to warm to room temperature and the solvent was removed in vacuum. The residue was purified by silica gel column chromatography to give the pure product.

2.3 Optimization of the reaction conditions for the reaction of *N*-Boc ketimine^a



Entry	Cat.	Temp./°C	Solvent	Additive	t/h	yield/% ^b	ee/% ^c
1	4a	20	DCM	Methyl acrylate	0.5	85	0
2	4b	20	DCM	Methyl acrylate	0.5	86	3
3	4c	20	DCM	Methyl acrylate	0.5	87	2
4	4d	20	DCM	Methyl acrylate	0.5	91	19
5	4e	20	DCM	Methyl acrylate	0.5	93	29
6	4f	20	DCM	Methyl acrylate	0.5	90	16
7	4g	20	DCM	Methyl acrylate	0.5	90	18
8	4h	20	DCM	Methyl acrylate	0.5	93	23
9	4e	0	DCM	Methyl acrylate	1	89	43
10	4e	-20	DCM	Methyl acrylate	1	94	60
11	4e	-60	DCM	Methyl acrylate	2	89	70
12	4e	-72	DCM	Methyl acrylate	2	91	85
13	4e	-72	Toluene	Methyl acrylate	2	88	46
14	4e	-72	Et ₂ O	Methyl acrylate	2	89	39
15 ^d	4e	-72	DCM	Methyl acrylate	3	90	82
16	4i	-72	DCM	Methyl acrylate	3	86	35
17	4j	-72	DCM	Methyl acrylate	3	89	30

^a Reaction conditions: 0.10 mmol **1a**, 0.20 mmol **2**, methyl acrylate (5 mol%), chiral phosphine (5 mol%), DCM (1.0 mL). ^b Isolated yield. ^c The ee value was determined by chiral HPLC analysis. ^d Chiral phosphine (1 mol%) was used.

2.4 Procedure for transformations of α -trifluoromethyl- α -amino nitriles

2.4.1 Procedure for the preparation of **5a**

Hydrogen chloride gas was introduced into a solution of **3'a** (15.1 mg, 0.05 mmol, 85% ee) in methanol (1.0 mL) at 0 °C until the starting material consumed. The resulting

mixture was stirred overnight and then the mixture was concentrated under reduced pressure, and saturated aqueous sodium hydrogen carbonate solution was added to adjust pH 9. The aqueous phase was extracted with dichloromethane (3×1.0 mL), and then the combined organic layers were washed with water and brine. Anhydrous Na_2SO_4 was added to the mixture, followed by a filtration. The filtrate was concentrated under reduced pressure and the residue was purified by silica gel column chromatography (petroleum ether/ethyl acetate = 3/1) to give **5a** (9.0 mg) in 90% yield as colorless oil.

2.4.2 Procedure for the preparation of **5b**⁴

To a stirred solution of **3'a** (30.0 mg, 0.1 mmol, 85% ee) in acetone (2.0 mL) was added Na_2CO_3 (53.0 mg, 0.5 mmol). Then 30% H_2O_2 aq. (2.0 mL) was added dropwise to the suspension at 0 °C. The resulting mixture was stirred at room temperature overnight, then water was added and the resulting mixture was extracted with ethyl acetate (3×5.0 mL). The combined organic layers were washed with water and brine and dried over anhydrous Na_2SO_4 . The filtrate was concentrated under reduced pressure, and the resulting residue was purified by silica gel column chromatography (petroleum ether/ethyl acetate = 3/1) to give **5b** (28.0 mg) in 88% yield as colorless oil.

2.4.3 Procedure for the preparation of **5c**

To a stirred solution of **3'a** (300.2 mg, 1.0 mmol, 85% ee) and $\text{NiCl}_2 \cdot 6\text{H}_2\text{O}$ (237.7 mg, 1.0 mmol) in methanol (18.0 mL) was added NaBH_4 (264.8 mg, 7.0 mmol) in portions at 0 °C over 10 minutes. The resulting mixture was stirred for 3h at 0 °C until **3'a** completely disappeared (monitored by TLC). The reaction was quenched by the addition of saturated NH_4Cl aqueous solution (2.0 mL), and the resulting mixture was stirred at room temperature until the generation of gas ceased. Anhydrous Na_2SO_4 was added to the mixture, followed by a filtration with diatomite. The filtrate was concentrated under reduced pressure and the residue was purified by silica gel column chromatography (petroleum ether/ethyl acetate = 5/1-1/1) to give **5c** (278.9 mg) in 92% yield as a white solid.

2.4.4 Procedure for the preparation of **5d**

To a stirred solution of 4-bromobenzoic acid (48.2 mg, 0.24 mmol) in CH_2Cl_2 (10.0 mL) were added HBTU (113.8 mg, 0.3 mmol) and DIPEA (51.6 mg, 0.4 mmol) successfully at 0 °C. Then **5c** (60.8 mg, 0.2 mmol, 85% ee) was added and the reaction mixture was stirred at room temperature and monitored by TLC. After 6h, the resulting solution was poured into 10.0 mL of H_2O , and extracted with dichloromethane (3×10.0 mL). The combined organic layers were washed with water and brine, dried over anhydrous Na_2SO_4 . The filtrate was concentrated under reduced pressure, and the resulting residue was purified by silica gel column chromatography (petroleum ether/ethyl acetate = 4/1) to give **5d** (89.9 mg) in 92% yield as a white solid.

2.4.5 Procedure for the preparation of **5e**

To a stirred solution of **5c** (60.8 mg, 0.2 mmol, 85% ee) in CH_2Cl_2 (5.0 mL) was added 4-nitrophenyl isothiocyanate (43.2 mg, 0.24 mmol) and the resulting mixture was stirred for 5h (monitored by TLC). Then the solvent was removed in vacuum and the

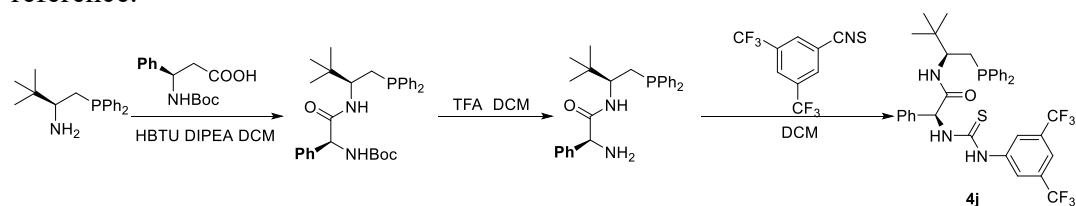
residue was purified by silica gel column chromatography (petroleum ether/ethyl acetate = 5/1) to give **5e** (91.0 mg) in 94% yield as a yellow solid.

2.4.6 Procedure for the preparation of **5f**⁵

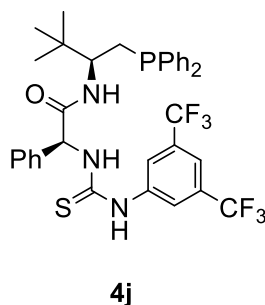
To the solution of **5c** (30.0 mg, 0.1 mmol) in toluene (1.0 mL) was added DBU (0.003 mmol). The mixture was heated until the starting material consumed. Then it was cooled to room temperature and concentrated under reduced pressure. The residue was purified by fast silica gel column chromatography (ethyl acetate / methanol = 20/1) to afford product **5f** in 90% yield as a pale yellow solid.

2.5 Procedure for the synthesis of the bifunctional catalyst **4j**

The dipeptide-based organophosphine **4i** was prepared according to the reported reference.²



(S)-2-(3-(3,5-bis(trifluoromethyl)phenyl)thioureido)-*N*-((*S*)-1-(diphenylphosphanyl)-3,3-dimethylbutan-2-yl)-2-phenylacetamide (**4j**)



White solid, mp 190-192 °C; ¹H NMR (CDCl₃, 400 MHz) δ 9.07 (s, 1H), 8.82-8.80 (m, 1H), 7.88 (s, 2H), 7.62-7.61 (m, 2H), 7.44-7.37 (m, 5H), 7.24 (m, 5H), 7.15-7.13 (m, 6H), 6.44-6.42 (d, J=7.2Hz, 1H), 5.95-5.93 (m, 1H), 3.66-3.58 (m, 1H), 2.41-2.37 (m, 1H), 0.79 (s, 9H); ¹³C NMR (CDCl₃, 100 MHz) δ 181.41, 172.51, 140.05, 139.21, 139.08, 136.99, 136.84, 135.49, 133.74, 133.54, 131.82, 131.64, 131.14, 130.80, 129.28, 129.23, 128.86, 128.77, 128.71, 128.45, 128.39, 128.38, 128.33, 128.17, 124.07, 121.68, 117.79, 62.17, 56.09, 35.46, 31.23, 31.09, 29.69, 26.13.; ¹⁹F NMR (CDCl₃, 376 MHz) δ -63.0; ³¹P NMR (CDCl₃, 162 MHz) δ -22.5; IR (Neat): 3290, 2965, 1653, 1538, 1473, 1382, 1278, 1180, 1131, 738, 695; HRMS (ESI): calcd for [M+H]⁺ (C₃₅H₃₅N₃O₆PS)⁺ requires 690.2064; found 690.2137; [α]_D^{26.2} = +28.5 (c = 1.0, CHCl₃).

III. Experimental data

Experimental data of 3a-3n^{6a,6b}

(R)-3,3,3-trifluoro-2-((4-methoxyphenyl)amino)-2-phenylpropanenitrile (3a)

White solid; ¹H NMR (CDCl₃, 400 MHz) δ 7.76-7.78 (m, 2H), 7.45-7.51 (m, 3H), 6.71 (d, *J* = 8.8 Hz, 2H), 6.60 (d, *J* = 8.8 Hz, 2H), 4.32 (s, 1H), 3.70 (s, 3H); ¹³C NMR (CDCl₃, 126 MHz) δ 155.3, 134.6, 130.7, 129.7, 129.1, 128.0, 122.3 (q, ¹*J*_{CF} = 286.4 Hz), 120.0, 114.9, 114.5, 66.3 (q, ²*J*_{CF} = 30.0 Hz), 55.4; ¹⁹F NMR (CDCl₃, 376 MHz) δ -76.5 (s, 3F); enantiomeric excess: 91%, determined by HPLC (Chiralpak OD, hexane/*i*-PrOH 90/10, flow rate 0.7 mL/min; *t*_{major} = 8.9 min, *t*_{minor} = 11.0 min, λ = 254 nm); [α]_D^{28.0} = -68.7 (*c* = 1.0, CHCl₃).

(R)-3,3,3-trifluoro-2-(4-fluorophenyl)-2-((4-methoxyphenyl)amino)propanenitrile (3b)

Yellow oil; ¹H NMR (CDCl₃, 400 MHz) δ 7.67-7.70 (m, 2H), 7.07-7.11 (m, 2H), 6.65 (d, *J* = 9.2 Hz, 2H), 6.53 (d, *J* = 9.2 Hz, 2H), 4.24 (s, 1H), 3.64 (s, 3H); ¹³C NMR (CDCl₃, 126 MHz) δ 165.0, 163.0, 155.4, 134.3, 130.2, 130.1, 125.5, 125.4, 122.2 (q, ¹*J*_{CF} = 285.9 Hz), 120.2, 116.4, 116.3, 114.8, 114.6, 65.8 (q, ²*J*_{CF} = 30.2 Hz), 55.4; ¹⁹F NMR (CDCl₃, 376 MHz) δ -76.7 (s, 3F), -109.6 (s, 1F); enantiomeric excess: 88%, determined by HPLC (Chiralpak AD-H, hexane/*i*-PrOH 95/5, flow rate 0.7 mL/min; *t*_{major} = 11.9 min, *t*_{minor} = 13.8 min, λ = 254 nm); [α]_D^{28.3} = -82.9 (*c* = 1.0, CHCl₃).

(R)-2-(4-chlorophenyl)-3,3,3-trifluoro-2-((4-methoxyphenyl)amino)propanenitrile (3c)

Yellow oil; ¹H NMR (CDCl₃, 400 MHz) δ 7.62-7.64 (m, 2H), 7.36-7.39 (m, 2H), 6.65 (d, *J* = 8.8 Hz, 2H), 6.52 (d, *J* = 8.8 Hz, 2H), 4.24 (s, 1H), 3.64 (s, 3H); ¹³C NMR (CDCl₃, 126 MHz) δ 155.4, 137.1, 134.1, 129.5, 128.3, 122.1 (q, ¹*J*_{CF} = 286.6 Hz), 120.1, 114.6, 65.6 (q, ²*J*_{CF} = 30.2 Hz), 55.4; ¹⁹F NMR (CDCl₃, 376 MHz) δ -76.6 (s, 3F); enantiomeric excess: 90%, determined by HPLC (Chiralpak AD-H, hexane/*i*-PrOH 95/5, flow rate 0.7 mL/min; *t*_{major} = 11.8 min, *t*_{minor} = 13.2 min, λ = 254 nm); [α]_D^{28.5} = -75.5 (*c* = 1.0, CHCl₃).

(R)-2-(3-chlorophenyl)-3,3,3-trifluoro-2-((4-methoxyphenyl)amino)propanenitrile (3d)

White solid, mp: 78-79 °C; ¹H NMR (CDCl₃, 400 MHz) δ 7.76 (s, 1H), 7.66 (d, *J* = 8.4 Hz, 1H), 7.46-7.49 (m, 1H), 7.40 (t, *J* = 8.0 Hz, 1H), 6.72 (d, *J* = 8.8 Hz, 2H), 6.60 (d, *J* = 8.8 Hz, 2H), 4.31 (s, 1H), 3.71 (s, 3H); ¹³C NMR (CDCl₃, 126 MHz) δ 155.5, 135.4, 134.1, 131.8, 131.1, 130.4, 128.2, 126.3, 122.1 (q, ¹*J*_{CF} = 284.3 Hz), 120.1, 114.6, 66.0 (q, ²*J*_{CF} = 29.9 Hz), 55.4; ¹⁹F NMR (CDCl₃, 376 MHz) δ -76.2 (s, 3F); IR (Neat): 3343, 2836, 2359, 1513, 1241, 1206, 1180, 1035, 823, 788; enantiomeric excess: 89%, determined by HPLC (Chiralpak AD-H, hexane/*i*-PrOH 90/10, flow rate 1.0 mL/min; *t*_{major} = 5.4 min, *t*_{minor} = 6.0 min, λ = 254 nm); [α]_D²⁵ = -111.7 (*c* = 1.0, CHCl₃).

(R)-2-(2-chlorophenyl)-3,3,3-trifluoro-2-((4-methoxyphenyl)amino)propanenitrile (3e)

Colorless oil; ¹H NMR (CDCl₃, 400 MHz) δ 7.86 (d, *J* = 7.6 Hz, 1H), 7.48-7.50 (m, 1H), 7.38-7.43 (m, 1H), 7.31-7.35 (m, 1H), 6.71-6.75 (m, 2H), 6.58-6.62 (m, 2H), 4.52 (s, 1H), 3.71 (s, 3H); ¹³C NMR (CDCl₃, 126 MHz) δ 155.0, 134.3, 133.5, 132.7, 131.8, 131.5, 127.5, 126.2, 122.6 (q, ¹*J*_{CF} = 285.9 Hz), 119.0, 114.6, 64.6 (q, ²*J*_{CF} = 30.9 Hz), 55.4; ¹⁹F NMR (CDCl₃, 376 MHz) δ -74.7 (s, 3F); IR (Neat): 3355, 2927, 1513, 1468, 1239, 1208, 1178, 1036, 762; HRMS (EI): calcd for [C₁₆H₁₂ClF₃N₂O] requires 340.0590; found 340.0587; enantiomeric excess: 90%, determined by

HPLC (Chiralpak AD-H, hexane/i-PrOH 90/10, flow rate 1.0 mL/min; $t_{\text{major}} = 6.5$ min, $t_{\text{minor}} = 7.5$ min, $\lambda = 254$ nm); $[\alpha]_{\text{D}}^{25} = -240.7$ ($c = 0.5$, CHCl_3).

(R)-2-(4-bromophenyl)-3,3,3-trifluoro-2-((4-methoxyphenyl)amino)propanenitrile (3f)

Yellow oil; $^1\text{H NMR}$ (CDCl_3 , 400 MHz) δ 7.52-7.57 (m, 4H), 6.65 (d, $J = 8.8$ Hz, 2H), 6.52 (d, $J = 8.8$ Hz, 2H), 4.24 (s, 1H), 3.64 (s, 3H); $^{13}\text{C NMR}$ (CDCl_3 , 126 MHz) δ 155.4, 134.1, 132.4, 129.7, 128.8, 125.4, 122.0 (q, $^1J_{\text{CF}} = 286.6$ Hz), 120.1, 114.6, 66.0 (q, $^2J_{\text{CF}} = 30.2$ Hz), 55.4; $^{19}\text{F NMR}$ (CDCl_3 , 376 MHz) δ -76.5 (s, 3F); enantiomeric excess: 90%, determined by HPLC (Chiralpak AD-H, hexane/i-PrOH 95/5, flow rate 0.7 mL/min; $t_{\text{major}} = 12.8$ min, $t_{\text{minor}} = 14.0$ min, $\lambda = 254$ nm); $[\alpha]_{\text{D}}^{28.9} = -60.3$ ($c = 1.0$, CHCl_3).

(R)-3,3,3-trifluoro-2-((4-methoxyphenyl)amino)-2-(p-tolyl)propanenitrile (3g)

Yellow oil; $^1\text{H NMR}$ (CDCl_3 , 400 MHz) δ 7.55-7.57 (m, 2H), 7.18-7.20 (m, 2H), 6.63 (d, $J = 8.8$ Hz, 2H), 6.53 (d, $J = 8.8$ Hz, 2H), 4.21 (s, 1H), 3.63 (s, 3H), 2.32 (s, 3H); $^{13}\text{C NMR}$ (CDCl_3 , 126 MHz) δ 155.2, 141.0, 134.7, 129.8, 127.9, 126.7, 122.4 (q, $^1J_{\text{CF}} = 286.3$ Hz), 120.0, 115.1, 114.5, 66.2 (q, $^2J_{\text{CF}} = 30.1$ Hz), 55.4, 21.2; $^{19}\text{F NMR}$ (CDCl_3 , 376 MHz) δ -76.7 (s, 3F); enantiomeric excess: 92%, determined by HPLC (Chiralpak AD-H, hexane/i-PrOH 95/5, flow rate 0.7 mL/min; $t_{\text{major}} = 12.1$ min, $t_{\text{minor}} = 15.5$ min, $\lambda = 254$ nm); $[\alpha]_{\text{D}}^{28.7} = -63.4$ ($c = 1.0$, CHCl_3).

(R)-3,3,3-trifluoro-2-((4-methoxyphenyl)amino)-2-(m-tolyl)propanenitrile (3h)

Yellow oil; $^1\text{H NMR}$ (CDCl_3 , 400 MHz) δ 7.44-7.50 (m, 2H), 7.21-7.29 (m, 2H), 6.64 (d, $J = 8.8$ Hz, 2H), 6.54 (d, $J = 8.8$ Hz, 2H), 4.21 (s, 1H), 3.63 (s, 3H), 2.33 (s, 3H); $^{13}\text{C NMR}$ (CDCl_3 , 126 MHz) δ 155.2, 139.1, 134.7, 131.5, 129.6, 129.0, 128.5, 125.1, 122.3 (q, $^1J_{\text{CF}} = 286.4$ Hz), 120.0, 115.0, 114.5, 66.4 (q, $^2J_{\text{CF}} = 30.1$ Hz), 55.4, 21.5; $^{19}\text{F NMR}$ (CDCl_3 , 376 MHz) δ -76.4 (s, 3F); enantiomeric excess: 91%, determined by HPLC (Chiralpak AD-H, hexane/i-PrOH 95/5, flow rate 0.7 mL/min; $t_{\text{major}} = 9.3$ min, $t_{\text{minor}} = 10.7$ min, $\lambda = 254$ nm); $[\alpha]_{\text{D}}^{29.0} = -79.9$ ($c = 1.0$, CHCl_3).

(R)-3,3,3-trifluoro-2-(4-methoxyphenyl)-2-((4-methoxyphenyl)amino)propanenitrile (3i)

Yellow oil; $^1\text{H NMR}$ (CDCl_3 , 400 MHz) δ 7.59 (d, $J = 8.4$ Hz, 2H), 6.88 (d, $J = 8.8$ Hz, 2H), 6.64 (d, $J = 8.0$ Hz, 2H), 6.54 (d, $J = 8.0$ Hz, 2H), 4.21 (s, 1H), 3.76 (s, 3H), 3.63 (s, 3H); $^{13}\text{C NMR}$ (CDCl_3 , 126 MHz) δ 161.3, 155.2, 134.7, 129.4, 122.4 (q, $^1J_{\text{CF}} = 286.2$ Hz), 121.3, 120.1, 115.1, 114.5 (2C), 65.9 (q, $^2J_{\text{CF}} = 30.0$ Hz), 55.4 (2C); $^{19}\text{F NMR}$ (CDCl_3 , 376 MHz) δ -76.9 (s, 3F); enantiomeric excess: 90%, determined by HPLC (Chiralpak AD-H, hexane/i-PrOH 80/20, flow rate 1.0 mL/min; $t_{\text{major}} = 6.0$ min, $t_{\text{minor}} = 7.0$ min, $\lambda = 254$ nm); $[\alpha]_{\text{D}}^{28.8} = -66.2$ ($c = 1.0$, CHCl_3).

(R)-3,3,3-trifluoro-2-((4-methoxyphenyl)amino)-2-(4-(trifluoromethyl)phenyl)propanenitrile (3j)

Yellow oil; $^1\text{H NMR}$ (CDCl_3 , 400 MHz) δ 7.52-7.57 (m, 4H), 6.65 (d, $J = 8.8$ Hz, 2H), 6.52 (d, $J = 8.8$ Hz, 2H), 4.24 (s, 1H), 3.64 (s, 3H); $^{13}\text{C NMR}$ (CDCl_3 , 126 MHz) δ 155.5, 133.9, 133.7, 133.0 (q, $^2J_{\text{CF}} = 33.1$ Hz), 128.7, 126.2 (q, $^3J_{\text{CF}} = 3.7$ Hz), 123.5 (q, $^1J_{\text{CF}} = 273.2$ Hz), 122.1 (q, $^1J_{\text{CF}} = 286.8$ Hz), 120.1, 114.7, 114.4, 66.0 (q, $^2J_{\text{CF}} = 30.1$ Hz), 55.4; $^{19}\text{F NMR}$ (CDCl_3 , 376 MHz) δ -63.0 (s, 3F), -76.3 (s, 3F); enantiomeric excess: 86%, determined by HPLC (Chiralpak PC-2, hexane/i-PrOH 98/2, flow rate 0.7 mL/min; $t_{\text{major}} = 10.6$ min, $t_{\text{minor}} = 9.6$ min, $\lambda = 254$ nm); $[\alpha]_{\text{D}}^{29.2} = -74.7$ ($c = 1.0$, CHCl_3).

(R)-3,3,3-trifluoro-2-((4-methoxyphenyl)amino)-2-(thiophen-2-yl)propanenitrile (3k)

Yellow oil; ¹H NMR (CDCl₃, 400 MHz) δ 7.40-7.45 (m, 2H), 7.00-7.02 (m, 1H), 6.67-6.72 (m, 4H), 4.17 (s, 1H), 3.67 (s, 3H); ¹³C NMR (CDCl₃, 126 MHz) δ 156.1, 134.2, 133.4, 130.2, 129.5, 127.2, 121.9 (q, ¹J_{CF} = 287.0 Hz), 121.7, 114.5, 114.3, 64.0 (q, ²J_{CF} = 30.1 Hz), 55.4; ¹⁹F NMR (CDCl₃, 376 MHz) δ -76.6 (s, 3F); enantiomeric excess: 86%, determined by HPLC (Chiralpak AD-H, hexane/i-PrOH 95/5, flow rate 0.7 mL/min; t_{major} = 13.8 min, t_{minor} = 16.1 min, λ = 254 nm); [α]_D^{29.1} = -35.7 (c = 1.0, CHCl₃).

(R)-3,3,3-trifluoro-2-((4-fluorophenyl)amino)-2-phenylpropanenitrile (3l)

White solid; ¹H NMR (CDCl₃, 400 MHz) δ 7.67-7.68 (m, 2H), 7.39-7.44 (m, 3H), 6.76-6.80 (m, 2H), 6.49-6.52 (m, 2H), 4.41 (s, 1H); ¹³C NMR (CDCl₃, 126 MHz) δ 158.3 (d, ¹J_{CF} = 242 Hz), 137.3, 130.9, 129.3, 129.2, 127.9, 122.2 (q, ¹J_{CF} = 286.4 Hz), 119.2 (d, ³J_{CF} = 7.9 Hz), 115.9 (d, ²J_{CF} = 22.8 Hz), 114.6, 65.8 (q, ²J_{CF} = 30.2 Hz); ¹⁹F NMR (CDCl₃, 376 MHz) δ -76.6 (s, 3F), -121.7 (s, 1F); enantiomeric excess: 90%, determined by HPLC (Chiralpak OJ, hexane/i-PrOH 90/10, flow rate 1.0 mL/min; t_{major} = 10.3 min, t_{minor} = 7.2 min, λ = 254 nm); [α]_D^{27.9} = -91.5 (c = 1.0, CHCl₃).

(R)-2-((4-bromophenyl)amino)-3,3,3-trifluoro-2-phenylpropanenitrile (3m)

White solid; ¹H NMR (CDCl₃, 400 MHz) δ 7.64-7.66 (m, 2H), 7.40-7.44 (m, 3H), 7.17 (d, J = 8.8 Hz, 2H), 6.39 (d, J = 8.8 Hz, 2H), 4.55 (s, 1H); ¹³C NMR (CDCl₃, 126 MHz) δ 140.3, 132.1, 131.0, 129.4, 128.9, 127.7, 122.1 (q, ¹J_{CF} = 286.6 Hz), 118.7, 114.4, 114.3, 65.2 (q, ²J_{CF} = 30.5 Hz). ¹⁹F NMR (CDCl₃, 376 MHz) δ -76.7 (s, 3F); enantiomeric excess: 82%, determined by HPLC (Chiralpak OJ, hexane/i-PrOH 95/5, flow rate 0.7 mL/min; t_{major} = 25.2 min, t_{minor} = 19.0 min, λ = 254 nm); [α]_D^{28.0} = -79.7 (c = 1.0, CHCl₃).

(R)-3,3,3-trifluoro-2-phenyl-2-(p-tolylamino)propanenitrile (3n)

White solid; ¹H NMR (CDCl₃, 400 MHz) δ 7.68-7.69 (m, 2H), 7.39-7.40 (m, 3H), 6.87 (d, J = 8.4 Hz, 2H), 6.42 (d, J = 8.4 Hz, 2H), 4.40 (s, 1H), 2.14 (s, 3H); ¹³C NMR (CDCl₃, 126 MHz) δ 138.8, 131.3, 130.7, 129.7, 129.6, 129.2, 127.9, 122.3 (q, ¹J_{CF} = 286.3 Hz), 117.4, 114.8, 65.6 (q, ²J_{CF} = 30.1 Hz), 20.5; ¹⁹F NMR (CDCl₃, 376 MHz) δ -76.7 (s, 3F); enantiomeric excess: 89%, determined by HPLC (Chiralpak OJ, hexane/i-PrOH 90/10, flow rate 1.0 mL/min; t_{major} = 9.3 min, t_{minor} = 7.0 min, λ = 254 nm); [α]_D^{28.1} = -106.2 (c = 1.0, CHCl₃).

Experimental data of 3'a-3'm

Tert-butyl (S)-(1-cyano-2,2,2-trifluoro-1-phenylethyl)carbamate (3'a)

White solid, mp: 81-82 °C; ¹H NMR (CDCl₃, 400 MHz) δ 7.64-7.66 (m, 2H), 7.47-7.48 (m, 3H), 6.17 (br, 1H), 1.31 (s, 9H); ¹³C NMR (CDCl₃, 100 MHz) δ 152.7, 130.7, 130.3, 128.9, 126.6, 121.7 (q, ¹J_{CF} = 284.7 Hz), 113.8, 83.2, 62.4 (q, ²J_{CF} = 31.3 Hz), 27.7; ¹⁹F NMR (CDCl₃, 376 MHz) δ -75.7 (s, J = 3F); IR (Neat): 3244, 3147, 2980, 2933, 2855, 1717, 1492, 1454, 1370, 1250, 1192, 1156; HRMS (ESI): calcd for [M-H]⁺ (C₁₄H₁₄O₂N₂F₃)⁺ requires 299.1013; found 299.1005; enantiomeric excess: 85%, determined by HPLC (Chiralpak AD-H, hexane/i-PrOH 95/5, flow rate 0.6 mL/min; t_{major} = 16.1 min, t_{minor} = 11.4 min, λ = 254 nm); [α]_D²⁴ = 35.4 (c = 1.0, CHCl₃).

Tert-butyl (S)-(1-cyano-2,2,2-trifluoro-1-(3-fluorophenyl)ethyl)carbamate (3'b)

Colorless oil; ¹H NMR (CDCl₃, 400 MHz) δ 7.44-7.48 (m, 2H), 7.35-7.38 (m, 1H), 7.17-7.22 (m,

1H), 6.41 (br, 1H), 1.34 (s, 9H); **¹³C NMR** (CDCl₃, 100 MHz) δ 162.7 (d, ¹J_{CF} = 247.2 Hz), 152.7, 133.1, 130.6 (d, ³J_{CF} = 8.2 Hz), 122.6, 122.5, 121.5 (q, ¹J_{CF} = 284.9 Hz), 117.5 (d, ²J_{CF} = 21.0 Hz), 114.1 (d, ²J_{CF} = 24.7 Hz), 113.5, 83.6, 62.0 (q, ²J_{CF} = 31.0 Hz), 27.7; **¹⁹F NMR** (CDCl₃, 376 MHz) δ -72.2 (s, 3F), -107.5 (s, 1F); **IR** (Neat): 3244, 3147, 2981, 1719, 1598, 1492, 1371, 1249, 1201, 1156; **HRMS** (ESI): calcd for [M-H]⁻ (C₁₄H₁₃O₂N₂F₄)⁻ requires 317.0919; found 317.0915; enantiomeric excess: 79%, determined by HPLC (Chiralpak AD-H, hexane/i-PrOH 98/2, flow rate 0.4 mL/min; t_{major} = 36.0 min, t_{minor} = 27.3 min, λ = 254 nm); [α]_D²⁵ = 47.6 (c = 0.5, CHCl₃).

Tert-butyl (S)-(1-cyano-2,2,2-trifluoro-1-(4-fluorophenyl)ethyl)carbamate (3'c)

Colorless oil; **¹H NMR** (CDCl₃, 400 MHz) δ 7.62-7.65 (dd, J = 8.4 Hz, 4.8 Hz, 2H), 7.17 (t, J = 8.4 Hz, 2H), 5.92 (br, 1H), 1.35 (s, 9H); **¹³C NMR** (CDCl₃, 100 MHz) δ 163.5 (d, ¹J_{CF} = 249.6 Hz), 152.4, 128.8 (d, ³J_{CF} = 8.7 Hz), 126.4, 121.6 (q, ¹J_{CF} = 284.4 Hz), 116.1 (d, ²J_{CF} = 22.2 Hz), 113.7, 83.5, 61.9 (q, ²J_{CF} = 31.0 Hz), 27.8; **¹⁹F NMR** (CDCl₃, 376 MHz) δ -76.1 (s, 3F), -110.2 (s, 1F); **IR** (Neat): 3246, 3148, 2981, 1719, 1605, 1512, 1395, 1371, 1251, 1191; **HRMS** (ESI): calcd for [M-H]⁻ (C₁₄H₁₃O₂N₂F₄)⁻ requires 317.0919; found 317.0915; enantiomeric excess: 89%, determined by HPLC (Chiralpak AD-H, hexane/i-PrOH 98/2, flow rate 0.4 mL/min; t_{major} = 31.3 min, t_{minor} = 24.0 min, λ = 254 nm); [α]_D²⁵ = 52.4 (c = 0.5, CHCl₃).

Tert-butyl (S)-(1-cyano-1-(3,5-difluorophenyl)-2,2,2-trifluoroethyl)carbamate (3'd)

White solid, mp: 96-98 °C; **¹H NMR** (CDCl₃, 400 MHz) δ 7.21-7.23 (m, 2H), 6.93-6.98 (m, 1H), 6.23 (br, 1H), 1.37 (s, 9H); **¹³C NMR** (CDCl₃, 100 MHz) δ 163.5 (dd, ¹J_{CF} = 249.7 Hz, ³J_{CF} = 12.4 Hz), 152.6, 134.7, 121.3 (q, ¹J_{CF} = 284.7 Hz), 113.1, 110.5 (d, ²J_{CF} = 28.0 Hz), 110.5 (d, ³J_{CF} = 11.7 Hz), 106.2 (t, ²J_{CF} = 24.9 Hz), 84.0, 61.9 (q, ²J_{CF} = 31.1 Hz), 27.8; **¹⁹F NMR** (CDCl₃, 376 MHz) δ -71.9 (s, 3F), -103.4 (s, 1F); **IR** (Neat): 3242, 3142, 2975, 1731, 1624, 1608, 1491, 1442, 1370, 1254, 1212, 1187, 1122; **HRMS** (ESI): calcd for [M-H]⁻ (C₁₄H₁₂O₂N₂F₅)⁻ requires 335.0824; found 317.0818; enantiomeric excess: 83%, determined by HPLC (Chiralpak AD-H, hexane/i-PrOH 95/5, flow rate 0.6 mL/min; t_{major} = 8.9 min, t_{minor} = 8.2 min, λ = 254 nm); [α]_D²⁵ = 52.5 (c = 0.5, CHCl₃).

Tert-butyl (S)-(1-(2-chlorophenyl)-1-cyano-2,2,2-trifluoroethyl)carbamate (3'e)

Colorless oil; **¹H NMR** (CDCl₃, 400 MHz) δ 7.81-7.83 (m, 1H), 7.37-7.48 (m, 3H), 5.74 (br, 1H), 1.37 (s, 9H); **¹³C NMR** (CDCl₃, 100 MHz) δ 152.5, 132.6, 131.4, 130.9, 127.3, 126.8, 122.1 (q, ¹J_{CF} = 285.9 Hz), 113.4, 83.1, 62.6 (q, ²J_{CF} = 32.0 Hz), 27.8; **¹⁹F NMR** (CDCl₃, 376 MHz) δ -73.9 (s, 3F); **IR** (Neat): 3242, 3144, 2979, 1720, 1433, 1370, 1248, 1191, 1157; **HRMS** (ESI): calcd for [M+NH₄]⁺ (C₁₄H₁₈O₂N₃ClF₃)⁺ requires 352.1034.0623; found 352.1034; enantiomeric excess: 85%, determined by HPLC (Chiralpak AD-H, hexane/i-PrOH 90/10, flow rate 1.0 mL/min; t_{major} = 10.5 min, t_{minor} = 5.6 min, λ = 254 nm); [α]_D²⁵ = 0.47 (c = 1.0, CHCl₃).

Tert-butyl (S)-(1-(3-chlorophenyl)-1-cyano-2,2,2-trifluoroethyl)carbamate (3'f)

Colorless oil; **¹H NMR** (CDCl₃, 400 MHz) δ 7.64 (s, 1H), 7.57 (d, J = 8.0 Hz, 1H), 7.40-7.49 (m, 2H), 6.33 (br, 1H), 1.34 (s, 9H); **¹³C NMR** (CDCl₃, 100 MHz) δ 152.7, 135.1, 132.8, 130.6, 130.1, 126.8, 125.0, 121.5 (q, ¹J_{CF} = 284.9 Hz), 113.4, 83.7, 62.0 (q, ²J_{CF} = 31.8 Hz), 27.7; **¹⁹F NMR** (CDCl₃, 376 MHz) δ -72.0 (s, 3F); **IR** (Neat): 3245, 3147, 2980, 1718, 1478, 1370, 1249, 1193, 1156; **HRMS** (ESI): calcd for [M-H]⁻ (C₁₄H₁₃O₂N₂ClF₃)⁻ requires 333.0623; found 333.0622; enantiomeric excess: 72%, determined by HPLC (Chiralpak AD-H, hexane/i-PrOH 98/2, flow rate

0.4 mL/min; $t_{\text{major}} = 38.6$ min, $t_{\text{minor}} = 33.2$ min, $\lambda = 254$ nm); $[\alpha]_{\text{D}}^{25} = 36.4$ ($c = 0.5$, CHCl_3).

Tert-butyl (S)-(1-(4-chlorophenyl)-1-cyano-2,2,2-trifluoroethyl)carbamate (3'g)

White solid, mp: 108-110 °C; $^1\text{H NMR}$ (CDCl_3 , 400 MHz) δ 7.59 (d, $J = 7.6$ Hz, 2H), 7.46 (d, $J = 8.8$ Hz, 2H), 6.35 (br, 1H), 1.34 (s, 9H); $^{13}\text{C NMR}$ (CDCl_3 , 100 MHz) δ 152.7, 136.7, 129.1, 128.0, 121.5 (q, $^1J_{\text{CF}} = 284.7$ Hz), 113.5, 83.6, 62.0 (q, $^2J_{\text{CF}} = 31.3$ Hz), 27.8; $^{19}\text{F NMR}$ (CDCl_3 , 376 MHz) δ -72.0 (s, 3F); **IR** (Neat): 3241, 3142, 2980, 1717, 1493, 1369, 1248, 1193, 1156; **HRMS** (ESI): calcd for $[\text{M-H}]^-$ ($\text{C}_{14}\text{H}_{13}\text{O}_2\text{N}_2\text{ClF}_3$) requires 333.0623; found 333.0618; enantiomeric excess: 83%, determined by HPLC (Chiralpak AD-H, hexane/i-PrOH 91/9, flow rate 0.7 mL/min; $t_{\text{major}} = 6.7$ min, $t_{\text{minor}} = 6.2$ min, $\lambda = 214$ nm); $[\alpha]_{\text{D}}^{25} = 43.3$ ($c = 0.5$, CHCl_3).

Tert-butyl (S)-(1-(3-bromophenyl)-1-cyano-2,2,2-trifluoroethyl)carbamate (3'h)

White solid, mp: 118-120 °C; $^1\text{H NMR}$ (CDCl_3 , 400 MHz) δ 7.79 (s, 2H), 7.62 (t, $J = 8.0$ Hz, 2H), 7.36 (t, $J = 8.0$ Hz, 1H), 6.38 (br, 1H), 1.33 (s, 9H); $^{13}\text{C NMR}$ (CDCl_3 , 100 MHz) δ 152.7, 133.6, 130.4, 129.6, 125.4, 123.0, 121.4 (q, $^1J_{\text{CF}} = 285.0$ Hz), 113.4, 83.7, 61.9 (q, $^2J_{\text{CF}} = 31.0$ Hz), 27.7; $^{19}\text{F NMR}$ (CDCl_3 , 376 MHz) δ -72.1 (s, 3F); **IR** (Neat): 3245, 3147, 2980, 1719, 1476, 1370, 1249, 1193, 1156; **HRMS** (ESI): calcd for $[\text{M-H}]^-$ ($\text{C}_{14}\text{H}_{13}\text{O}_2\text{N}_2\text{BrF}_3$) requires 377.0118; found 377.0112; enantiomeric excess: 68%, determined by HPLC (Chiralpak AD-H, hexane/i-PrOH 98/2, flow rate 0.6 mL/min; $t_{\text{major}} = 27.1$, $t_{\text{minor}} = 25.2$, $\lambda = 254$ nm); $[\alpha]_{\text{D}}^{25} = 25.9$ ($c = 0.5$, CHCl_3).

Tert-butyl (S)-(1-(4-bromophenyl)-1-cyano-2,2,2-trifluoroethyl)carbamate (3'i)

White solid, mp: 102-104 °C; $^1\text{H NMR}$ (CDCl_3 , 400 MHz) δ 7.62 (d, $J = 8.8$ Hz, 2H), 7.52 (d, $J = 8.4$ Hz, 2H), 6.23 (br, 1H), 1.34 (s, 9H); $^{13}\text{C NMR}$ (CDCl_3 , 100 MHz) δ 152.6, 132.2, 129.9, 128.3, 125.0, 121.5 (q, $^1J_{\text{CF}} = 284.4$ Hz), 113.5, 83.6, 62.2 (q, $^2J_{\text{CF}} = 31.2$ Hz), 27.8; $^{19}\text{F NMR}$ (CDCl_3 , 376 MHz) δ -72.1 (s, 3F); **IR** (Neat): 3244, 3146, 2980, 1718, 1489, 1370, 1248, 1194, 1156, 1076; **HRMS** (ESI): calcd for $[\text{M-H}]^-$ ($\text{C}_{14}\text{H}_{13}\text{O}_2\text{N}_2\text{BrF}_3$) requires 377.0107; found 377.0114; enantiomeric excess: 70%, determined by HPLC (Chiralpak AD-H, hexane/i-PrOH 98/2, flow rate 0.4 mL/min; $t_{\text{major}} = 27.8$ min, $t_{\text{minor}} = 23.9$ min, $\lambda = 254$ nm); $[\alpha]_{\text{D}}^{25} = 33.5$ ($c = 0.5$, CHCl_3).

Tert-butyl (S)-(1-cyano-2,2,2-trifluoro-1-(3-nitrophenyl)ethyl)carbamate (3'j)

Colorless oil; $^1\text{H NMR}$ (CDCl_3 , 400 MHz) δ 8.51 (s, 1H), 8.38 (d, $J = 8.0$ Hz, 1H), 8.05 (d, $J = 8.0$ Hz, 1H), 7.73 (t, $J = 8.0$ Hz, 1H), 6.20 (br, 1H), 1.36 (s, 9H); $^{13}\text{C NMR}$ (CDCl_3 , 100 MHz) δ 152.5, 148.6, 133.1, 132.8, 130.3, 125.4, 121.8, 120.9 (q, $^1J_{\text{CF}} = 285.0$ Hz), 113.1, 84.1, 61.9 (q, $^2J_{\text{CF}} = 30.6$ Hz), 27.8; $^{19}\text{F NMR}$ (CDCl_3 , 376 MHz) δ -72.1 (s, 3F); **IR** (Neat): 3320, 3147, 2981, 2938, 1722, 1538, 1458, 1352, 1250, 1197; **HRMS** (ESI): calcd for $[\text{M-H}]^-$ ($\text{C}_{14}\text{H}_{13}\text{O}_4\text{N}_3\text{F}_3$) requires 344.0864; found 344.0863; enantiomeric excess: 70%, determined by HPLC (Chiralpak AS-H, hexane/i-PrOH 95/5, flow rate 0.6 mL/min; $t_{\text{major}} = 17.1$ min, $t_{\text{minor}} = 15.6$ min, $\lambda = 254$ nm); $[\alpha]_{\text{D}}^{25} = 25.4$ ($c = 0.5$, CHCl_3).

Tert-butyl (S)-(1-cyano-2,2,2-trifluoro-1-(p-tolyl)ethyl)carbamate (3'k)

White solid, mp: 74-75 °C; $^1\text{H NMR}$ (CDCl_3 , 400 MHz) δ 7.51 (d, $J = 8.0$ Hz, 2H), 7.27 (d, $J = 8.0$ Hz, 2H), 5.96 (br, 1H), 2.39 (s, 3H), 1.34 (s, 9H); $^{13}\text{C NMR}$ (CDCl_3 , 100 MHz) δ 152.6, 140.6, 129.6, 127.6, 126.5, 121.8 (q, $^1J_{\text{CF}} = 284.2$ Hz), 113.9, 83.1, 62.2 (q, $^2J_{\text{CF}} = 31.1$ Hz), 27.8, 21.1; $^{19}\text{F NMR}$ (CDCl_3 , 376 MHz) δ -75.9 (s, 3F); **IR** (Neat): 3244, 3146, 2980, 1715, 1394, 1369, 1251,

1210, 1185, 1158; **HRMS** (ESI): calcd for $[M+NH_4]^+$ ($C_{15}H_{21}F_3N_3O_2$)⁺ requires 332.1580; found 332.1581; enantiomeric excess: 84%, determined by HPLC (Chiralpak AD-H, hexane/i-PrOH 90/10, flow rate 1.0 mL/min; t_{major} = 5.0 min, t_{minor} = 4.6 min, λ = 254 nm); $[\alpha]_D^{25}$ = 25.4 (c = 1.0, $CHCl_3$).

Tert-butyl (S)-(1-cyano-2,2,2-trifluoro-1-(thiophen-2-yl)ethyl)carbamate (3'l)

White solid; mp: 61-62 °C; **¹H NMR** ($CDCl_3$, 400 MHz) δ 7.43-7.45 (m, 2H), 7.06-7.08 (m, 1H), 6.56 (br, 1H), 1.38 (s, 9H); **¹³C NMR** ($CDCl_3$, 100 MHz) δ 152.9, 134.2, 129.3, 128.2, 127.2, 111.4 (q, $^1J_{CF}$ = 284.9 Hz), 113.2, 83.6, 59.2 (q, $^2J_{CF}$ = 32.8 Hz), 27.8; **¹⁹F NMR** ($CDCl_3$, 376 MHz) δ -76.3 (s, 3F); **IR** (Neat): 3146, 2981, 2359, 1716, 1370, 1251, 1216, 1188, 1157; **HRMS** (ESI): calcd for $[M+NH_4]^+$ ($C_{12}H_{17}F_3N_3O_2S$)⁺ requires 324.0988; found 324.0988; enantiomeric excess: 77%, determined by HPLC (Chiralpak AD-H, hexane/i-PrOH 95/5, flow rate 0.8 mL/min; t_{major} = 12.7 min, t_{minor} = 10.0 min, λ = 220 nm); $[\alpha]_D^{25}$ = 14.2 (c = 1.0, $CHCl_3$).

Benzyl (S)-(1-cyano-2,2,2-trifluoro-1-phenylethyl)carbamate (3'm)

White solid, mp: 91-92 °C; 88% yield; **¹H NMR** ($CDCl_3$, 400 MHz) δ 7.62-7.60 (m, 2H), 7.48-7.40 (m, 3H), 7.30-7.19 (m, 5H), 6.29 (br, 1H), 5.12-5.03 (m, 2H); **¹³C NMR** ($CDCl_3$, 100 MHz) δ 153.6, 134.8, 130.6, 129.9, 129.1, 128.6, 128.3, 126.7, 121.8 (q, $^1J_{CF}$ = 284.5 Hz), 113.5, 68.4, 62.4 (q, $^2J_{CF}$ = 31.2 Hz); **¹⁹F NMR** ($CDCl_3$, 376 MHz) δ -100.4 (s, 3F); **IR** (Neat): 3301, 3035, 1724, 1525, 1454, 1258, 1005, 742; **HRMS** (ESI): calcd for $[M+Na]^+$ ($C_{17}H_{13}F_3N_2O_2Na$)⁺ requires 357.0821; found 357.0824; enantiomeric excess: 49%, determined by HPLC (Chiralpak OD-H, hexane/i-PrOH 90/10, flow rate 1.0 mL/min; t_{major} = 9.0 min, t_{minor} = 7.0 min, λ = 254 nm); $[\alpha]_D^{25}$ = 25.1 (c = 1.0, $CHCl_3$).

Experimental data of 5a-5f

(S)-2-amino-3,3,3-trifluoro-2-phenylpropanenitrile (5a)^{6a}

Colorless oil; **¹H NMR** ($CDCl_3$, 400 MHz) δ 7.78 (d, J = 8.0 Hz, 2H), 7.45-7.52 (m, 3H), 2.31 (br, 2H); **¹³C NMR** ($CDCl_3$, 100 MHz) δ 131.0, 130.8, 129.0, 127.2, 122.5 (q, $^1J_{CF}$ = 283.0 Hz), 117.9, 61.2 (q, $^2J_{CF}$ = 31.6 Hz); **¹⁹F NMR** ($CDCl_3$, 376 MHz) δ -78.5 (s, 3F); **HRMS** (ESI): calcd for $[M+H]^+$ ($C_9H_8F_3N_2$)⁺ requires 201.0634 ; found 201.0635 ; enantiomeric excess: 85%, determined by HPLC (Chiralpak AD-H, hexane/i-PrOH 95/5, flow rate 0.8 mL/min; t_{major} = 12.3 min, t_{minor} = 11.6 min, λ = 220 nm); $[\alpha]_D^{25}$ = 0.20 (c = 0.5, $CHCl_3$).

Tert-butyl (S)-(3-amino-1,1,1-trifluoro-3-oxo-2-phenylpropan-2-yl)carbamate (5b)

Colorless oil; **¹H NMR** ($CDCl_3$, 400 MHz) δ 7.39-7.49 (m, 5H), 6.30 (br, 1H), 5.88 (br, 2H), 1.32 (s, 9H); **¹³C NMR** ($CDCl_3$, 100 MHz) δ 167.7, 153.2, 132.9, 129.1, 128.8, 127.0, 123.9 (q, $^1J_{CF}$ = 285.9 Hz), 81.0, 67.9 (q, $^2J_{CF}$ = 27.5 Hz), 28.0; **¹⁹F NMR** ($CDCl_3$, 376 MHz) δ -70.5 (s, 3F); **IR** (Neat): 3354, 2980, 1698, 1600, 1480, 1425, 1369, 1255, 1161, 1052; **HRMS** (ESI): calcd for $[M+H]^+$ ($C_{14}H_{18}O_3N_2F_3$)⁺ requires 319.1264; found 319.1267; enantiomeric excess: 85%, determined by HPLC (Chiralpak PC-2, hexane/i-PrOH 90/10, flow rate 1.0 mL/min; t_{major} = 13.5 min, t_{minor} = 9.4 min, λ = 220 nm); $[\alpha]_D^{25}$ = 30.7 (c = 0.5, $CHCl_3$).

Tert-butyl (S)-(3-amino-1,1,1-trifluoro-2-phenylpropan-2-yl)carbamate (5c)

White solid, mp: 70-71 °C; **¹H NMR** ($CDCl_3$, 400 MHz) δ 7.31-7.45 (m, 5H), 5.71 (br, 1H), 3.64 (d, J = 13.6 Hz 1H), 3.27 (d, J = 13.2 Hz, 1H), 1.39 (s, 11H); **¹³C NMR** ($CDCl_3$, 100 MHz) δ 154.0,

136.1, 128.5, 128.3, 126.3, 126.0 (q, $^1J_{\text{CF}} = 285.6$ Hz), 80.5, 65.1 (q, $^2J_{\text{CF}} = 25.0$ Hz), 46.7, 28.1; ^{19}F NMR (CDCl₃, 376 MHz) δ -70.7 (s, 3F); enantiomeric excess: 85%, determined by HPLC (Chiralpak IC, hexane/i-PrOH 90/10, flow rate 1.0 mL/min; $t_{\text{major}} = 7.7$ min, $t_{\text{minor}} = 8.5$ min, $\lambda = 254$ nm); $[\alpha]_{\text{D}}^{25} = 20.8$ ($c = 1.0$, CHCl₃).

Tert-butyl (S)-(3-(4-bromobenzamido)-1,1,1-trifluoro-2-phenylpropan-2-yl)carbamate (5d)

White solid, mp: 48-49 °C; ^1H NMR (CDCl₃, 400 MHz) δ 8.11 (br, 1H), 7.42-7.51 (m, 6H), 7.31-7.38 (m, 3H), 5.86 (br, 1H), 4.79 (br, 1H), 3.94 (d, $J = 14.0$ Hz, 1H), 1.46 (s, 9H); ^{13}C NMR (CDCl₃, 100 MHz) δ 166.6, 155.4, 133.0, 132.7, 131.6, 129.0, 128.6, 128.5, 127.2, 126.1, 124.9 (q, $^1J_{\text{CF}} = 284.2$ Hz), 81.7, 65.8 (q, $^2J_{\text{CF}} = 25.6$ Hz), 43.8, 28.2; ^{19}F NMR (CDCl₃, 376 MHz) δ -76.2 (s, 3F); IR (Neat): 3300, 2978, 1713, 1651 1591, 1258, 1157, 1011 843; HRMS (ESI): calcd for $[\text{M}+\text{H}]^+$ (C₂₁H₂₃BrF₃N₂O₃)⁺ requires 487.0839; found 487.0843; enantiomeric excess: 85%, determined by HPLC (Chiralpak PC-2, hexane/i-PrOH 90/10, flow rate 1.0 mL/min; $t_{\text{major}} = 8.6$ min, $t_{\text{minor}} = 6.7$ min, $\lambda = 254$ nm); $[\alpha]_{\text{D}}^{25} = -57.9$ ($c = 1.0$, CHCl₃).

Tert-butyl (S)-(1,1,1-trifluoro-3-(3-(4-nitrophenyl)thioureido)-2-phenylpropan-2-yl)carbamate (5e)

Yellow solid, mp: 72-74 °C ; ^1H NMR (CDCl₃, 400 MHz) δ 8.89 (br, 1H), 8.61 (br, 1H), 7.97 (d, $J = 8.0$ Hz, 2H), 7.44-7.48 (m, 5H), 6.73 (br, 2H), 5.64 (br, 1H), 5.42 (br, 1H), 3.96 (d, $J = 12.4$ Hz, 1H), 1.31 (s, 9H); ^{13}C NMR (CDCl₃, 100 MHz) δ 179.8, 155.5, 144.4, 142.3, 132.2, 129.4, 128.7, 127.7, 125.0, 124.6 (q, $^1J_{\text{CF}} = 284.4$ Hz), 123.0, 82.4, 66.1 (q, $^2J_{\text{CF}} = 21.1$ Hz), 49.6, 27.9; ^{19}F NMR (CDCl₃, 376 MHz) δ -77.6 (s, 3F); IR (Neat): 3280, 2979, 1712, 1596, 1344, 1260, 1015, 853, 703; HRMS (ESI): calcd for $[\text{M}-\text{H}]^-$ (C₂₁H₂₄F₃N₄O₄S)⁺ requires 485.1466; found 485.1465; enantiomeric excess: 86%, determined by HPLC (Chiralpak ADH, hexane/i-PrOH 90/10, flow rate 1.0 mL/min; $t_{\text{major}} = 23.8$ min, $t_{\text{minor}} = 30.2$ min, $\lambda = 254$ nm); $[\alpha]_{\text{D}}^{25} = 13.1$ ($c = 1.0$, CHCl₃).

(S)-4-phenyl-4-(trifluoromethyl)imidazolidin-2-one (5f)^{5,6b}

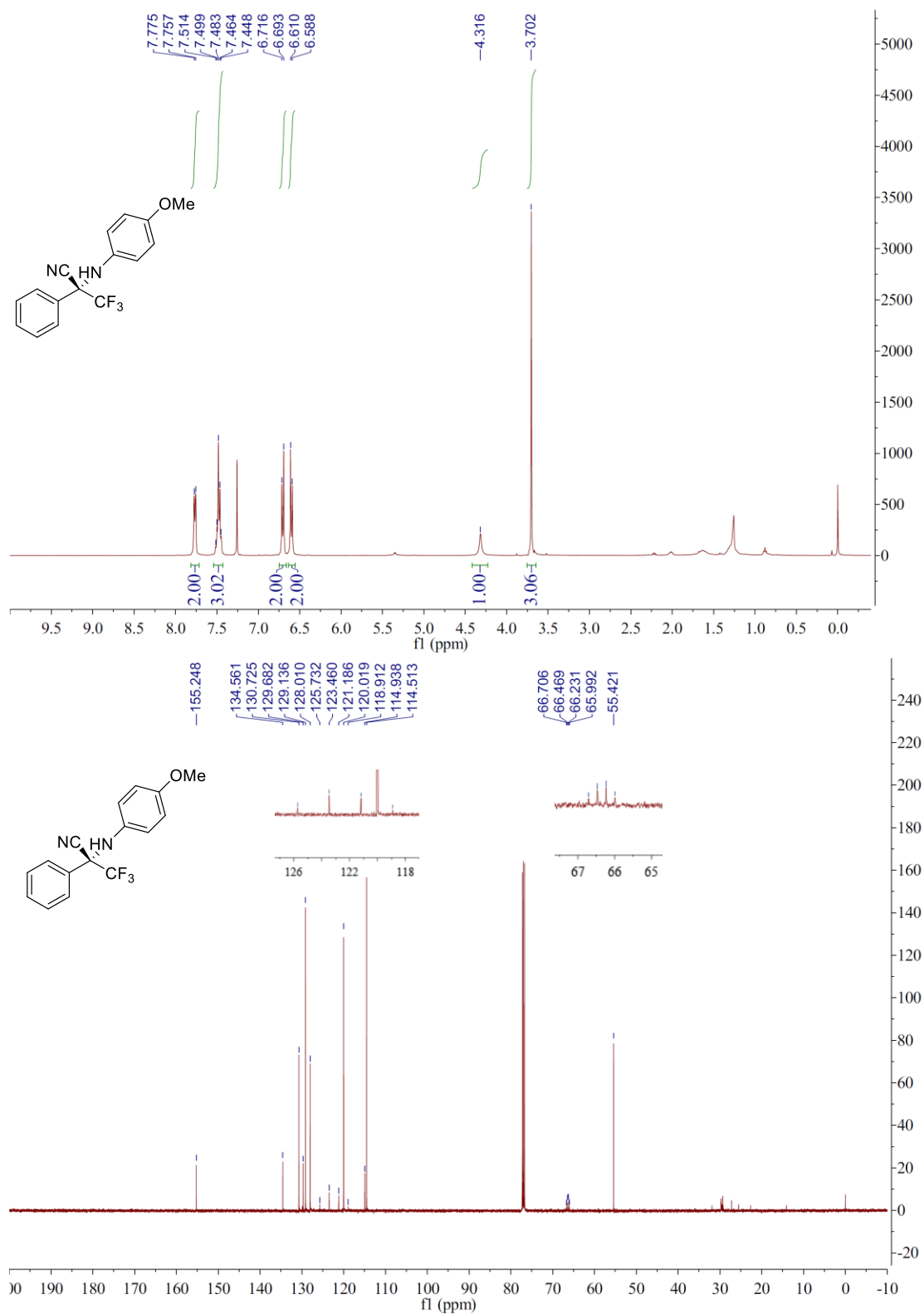
White solid; ^1H NMR (DMSO-*d*₆, 400 MHz) δ 8.31 (br, 1H), 7.42-7.53 (m, 5H), 6.79 (br, 1H), 4.03 (d, $J = 10.4$ Hz, 1H), 3.68 (d, $J = 11.2$ Hz, 1H); ^{13}C NMR (DMSO-*d*₆, 100 MHz) δ 161.6, 137.5, 129.2, 128.9, 127.2, 126.3 (q, $^1J_{\text{CF}} = 284.2$ Hz), 64.8 (q, $^2J_{\text{CF}} = 28.0$ Hz), 48.0; ^{19}F NMR (CDCl₃, 376 MHz) δ -74.8 (s, 3F); enantiomeric excess: 88%, determined by HPLC (Chiralpak OD-H, hexane/i-PrOH 85/15, flow rate 1.0 mL/min; $t_{\text{major}} = 6.2$ min, $t_{\text{minor}} = 6.9$ min, $\lambda = 254$ nm).

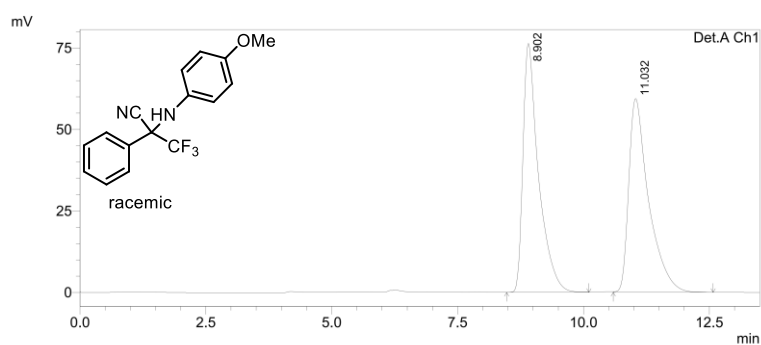
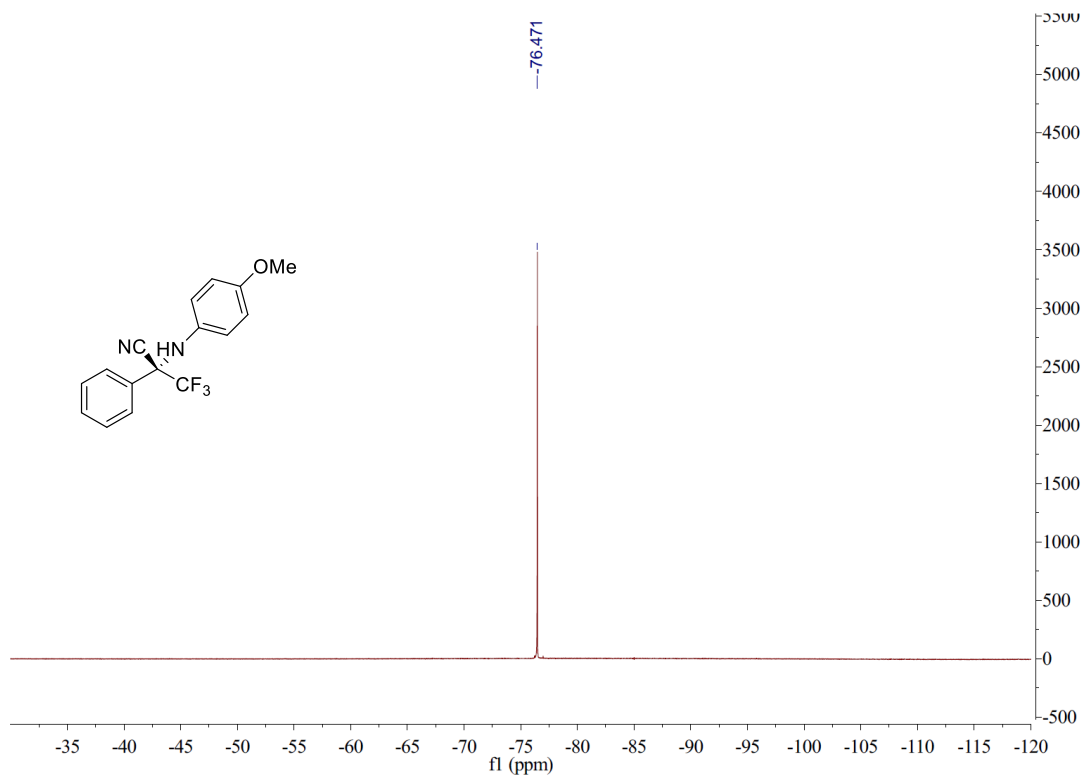
IV. References

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- 7 (a) Fetyukhin et al, *Journal of Organic Chemistry USSR* (English Translation), 1977, **13**, 244; *Zhurnal Organicheskoi Khimii*, 1977, **13**, 271; (b) N. V. Mel'nikhenko and M. V. Vovk, *Russian Journal of Organic Chemistry*, 2008, **44**, 149.

V. Copies of NMR and HPLC spectra

Copies of NMR and HPLC spectra for 3a-3n

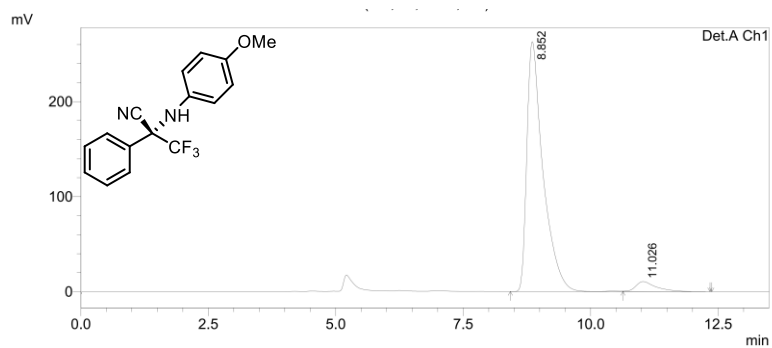




1 Det.A Ch1/254nm

PeakTable

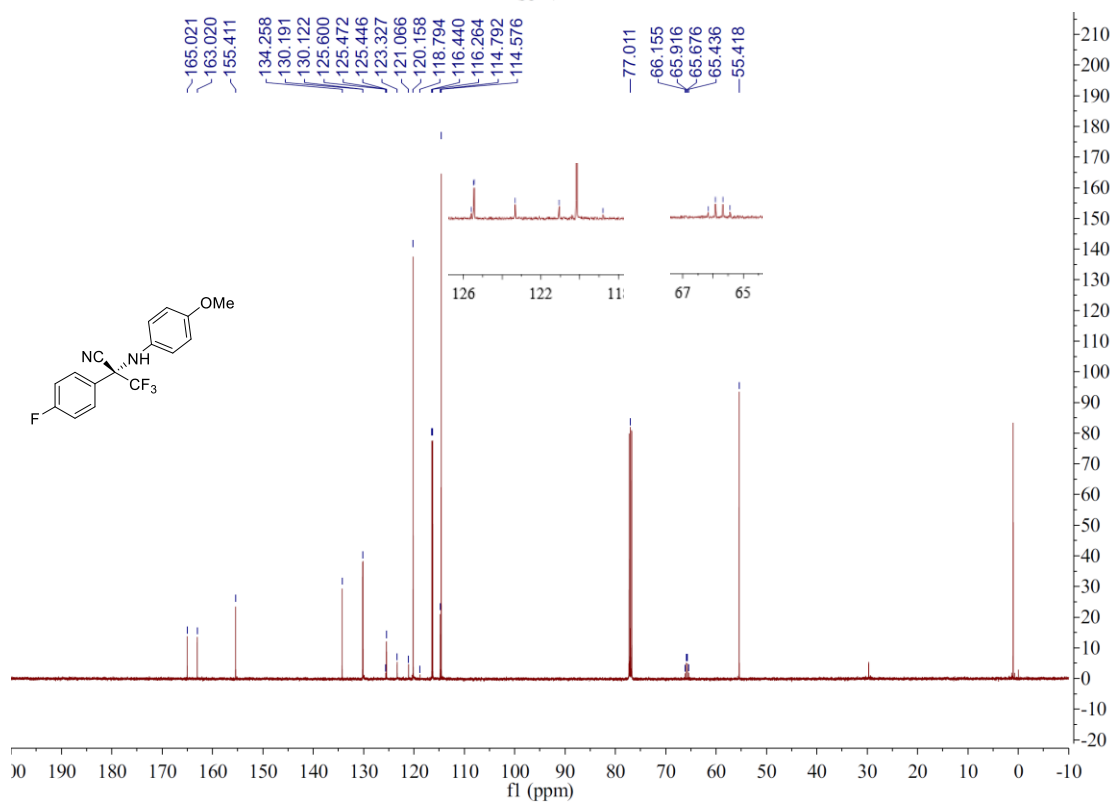
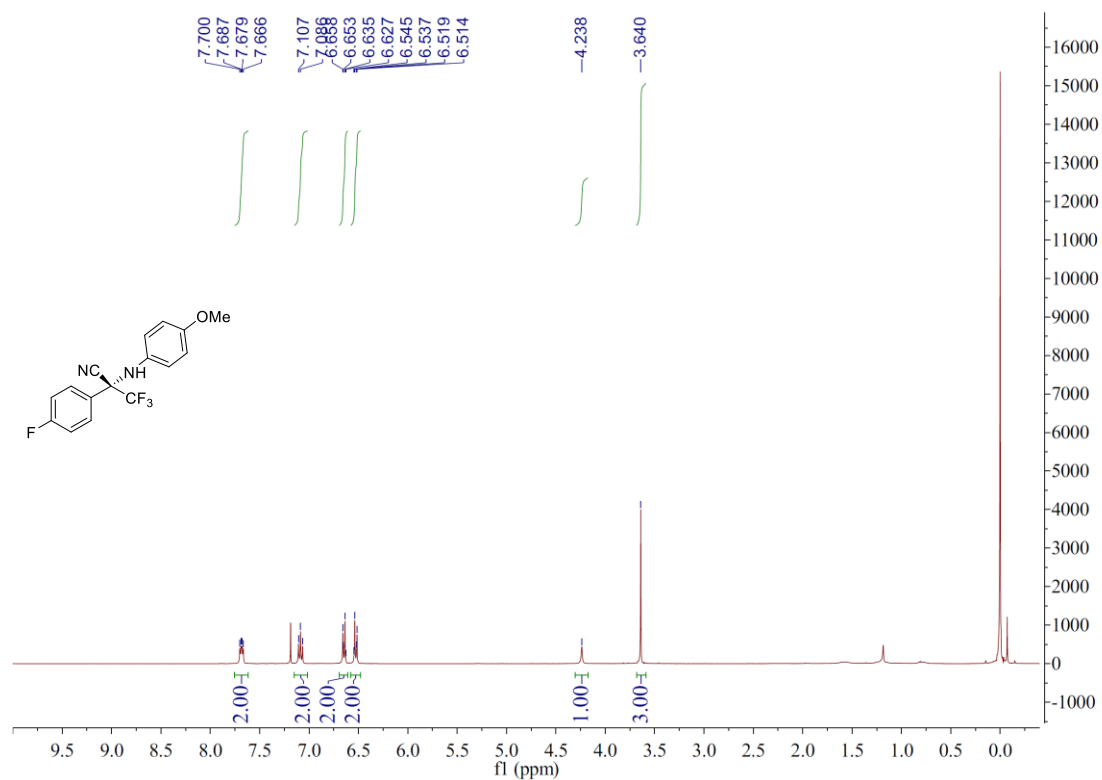
Peak#	Ret. Time	Area	Height	Area %	Height %
1	8.902	1640839	76376	50.113	56.234
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Total		3274280	135820	100.000	100.000

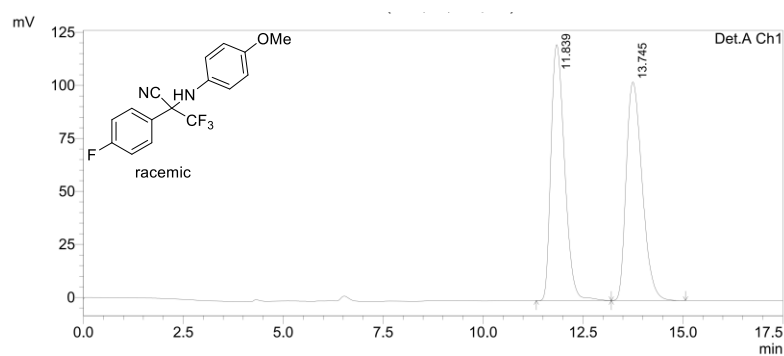
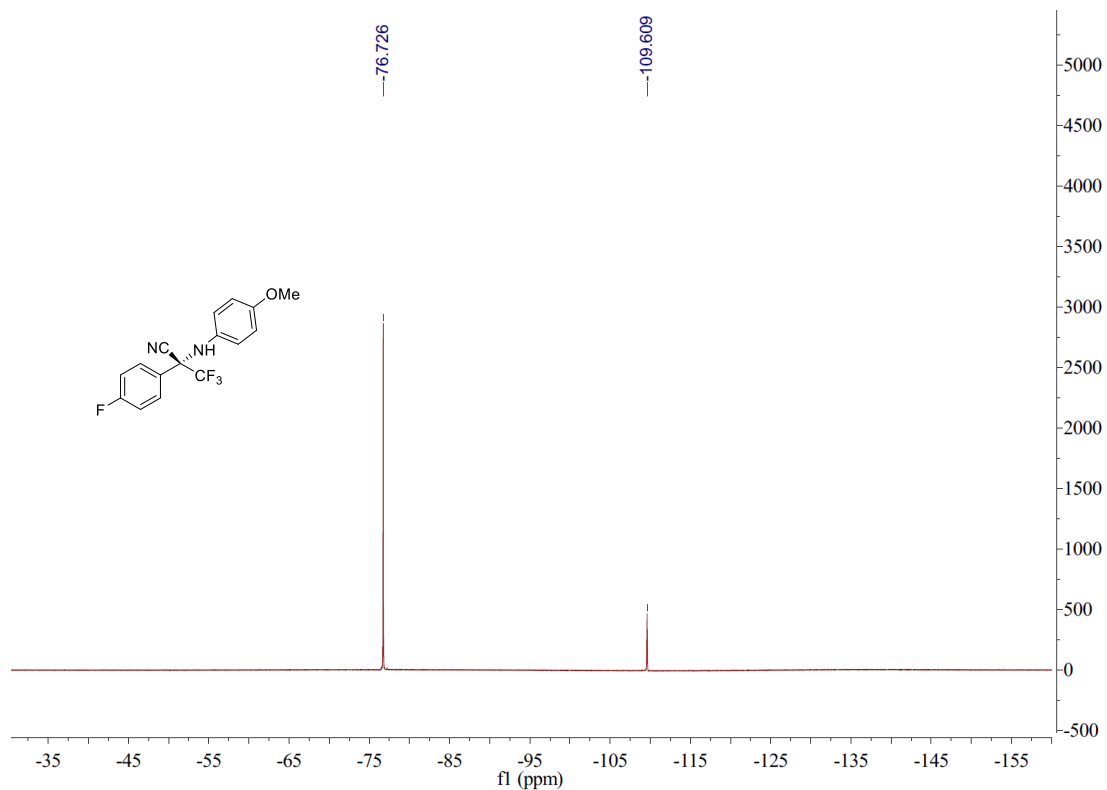


1 Det.A Ch1/254nm

PeakTable

Peak#	Ret. Time	Area	Height	Area %	Height %
1	8.852	5802988	262879	95.269	96.154
2	11.026	288163	10515	4.731	3.846
Total		6091151	273395	100.000	100.000

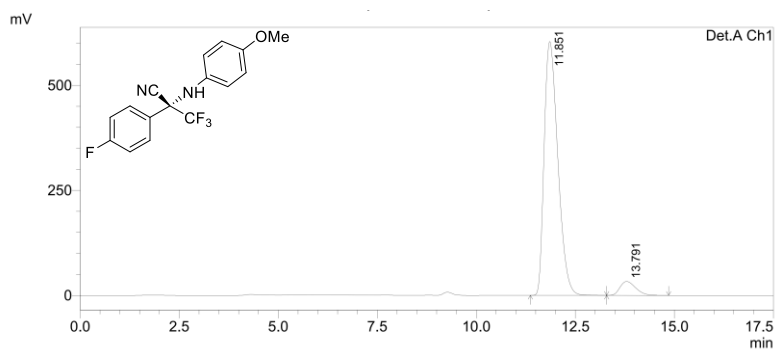




1 Det.A Ch1/254nm

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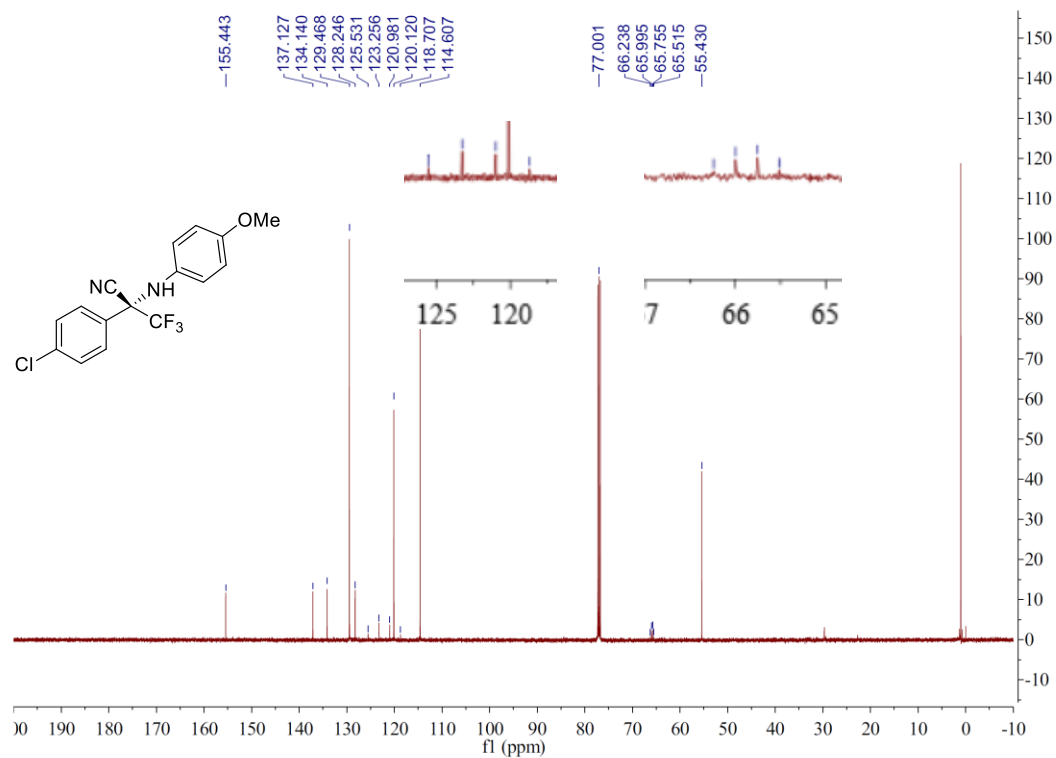
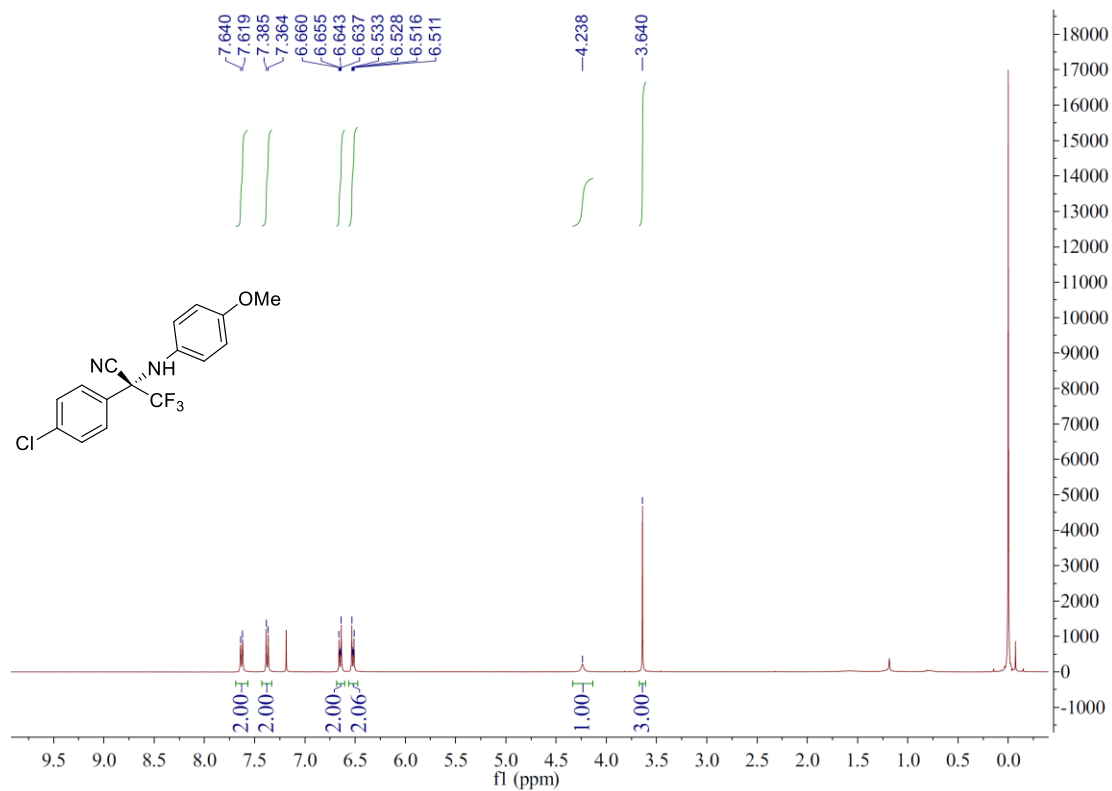
Peak#	Ret. Time	Area	Height	Area %	Height %
1	11.839	2858688	120701	49.942	53.947
2	13.745	2865352	103040	50.058	46.053
Total		5724040	223741	100.000	100.000

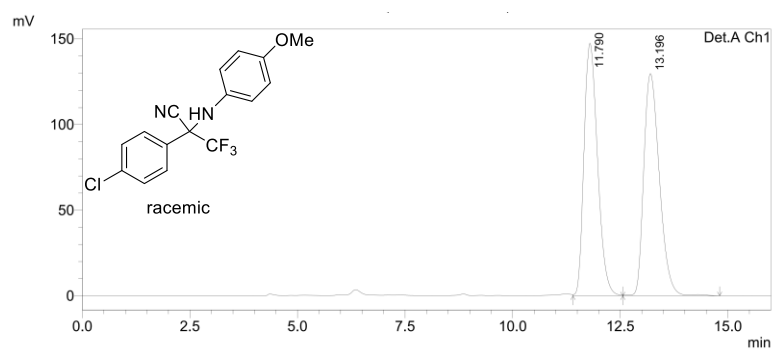
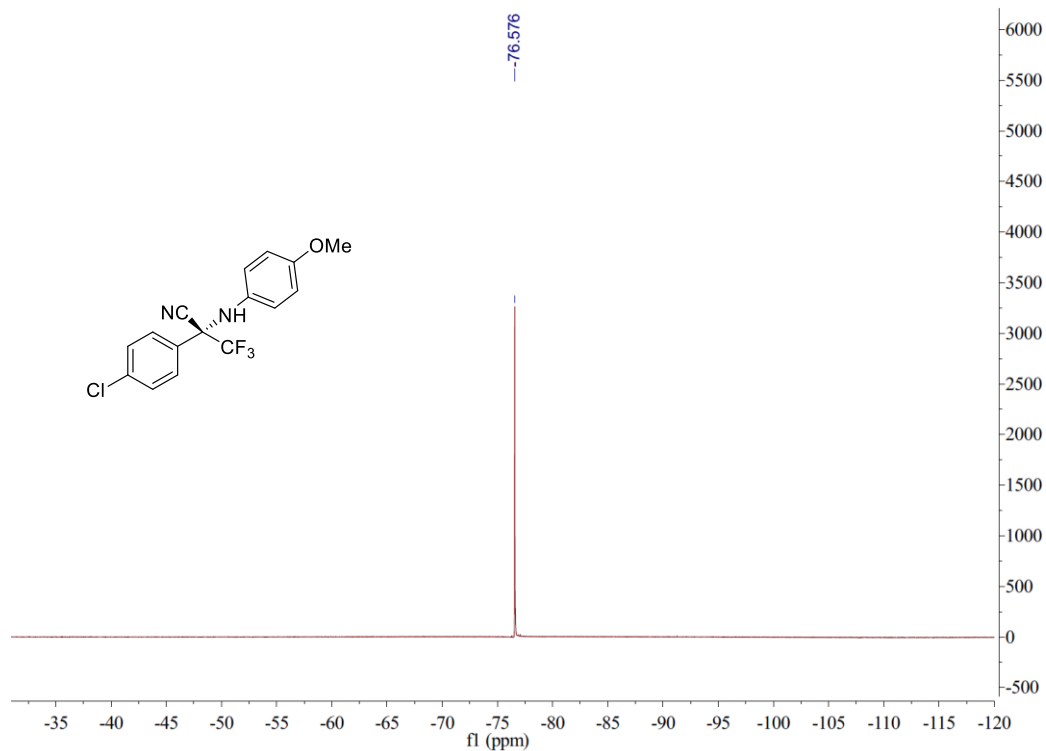


1 Det.A Ch1/254nm

PeakTable

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1	11.851	14499596	604132	94.099	94.813
2	13.791	909326	33050	5.901	5.187
Total		15408923	637182	100.000	100.000

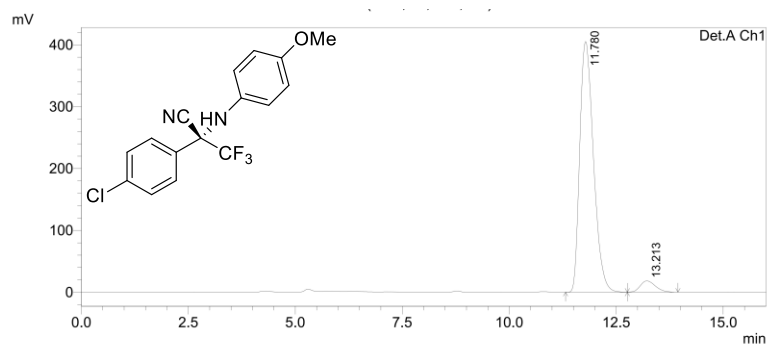




1 Det.A Ch1/254nm

PeakTable

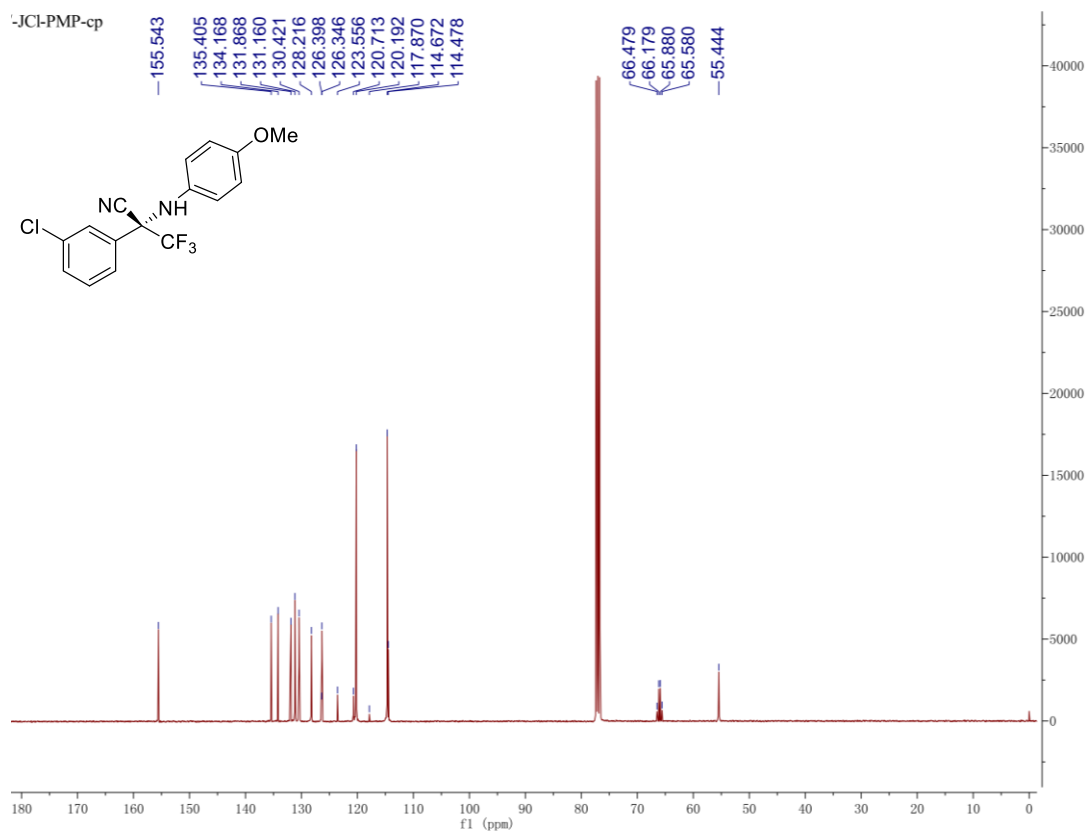
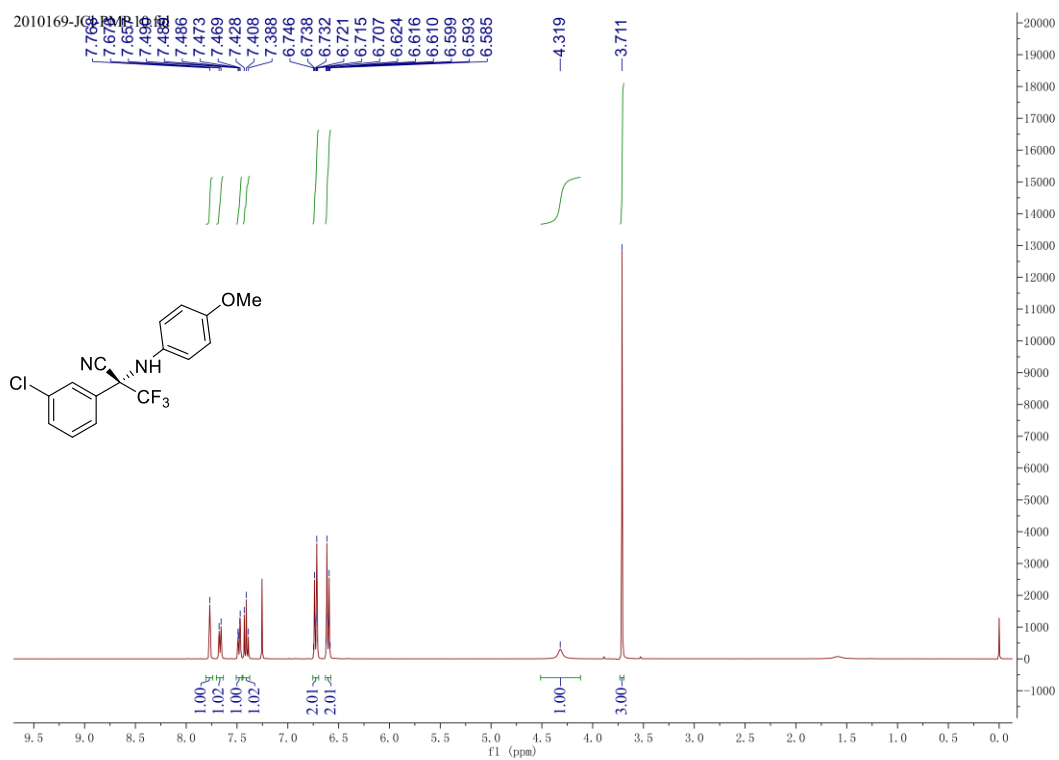
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1	11.790	3240784	147474	49.737	53.195
2	13.196	3275056	129757	50.263	46.805
Total		6515840	277231	100.000	100.000



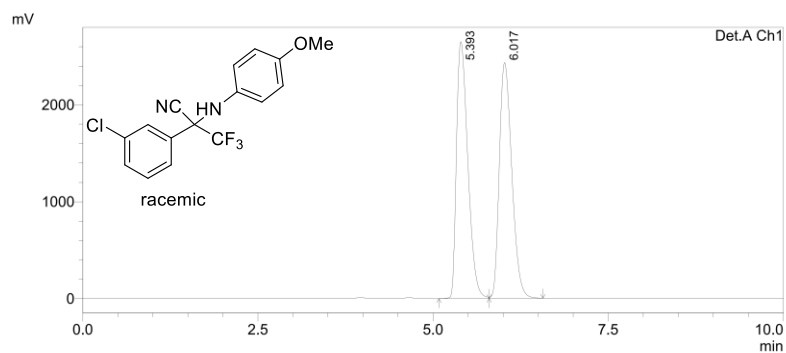
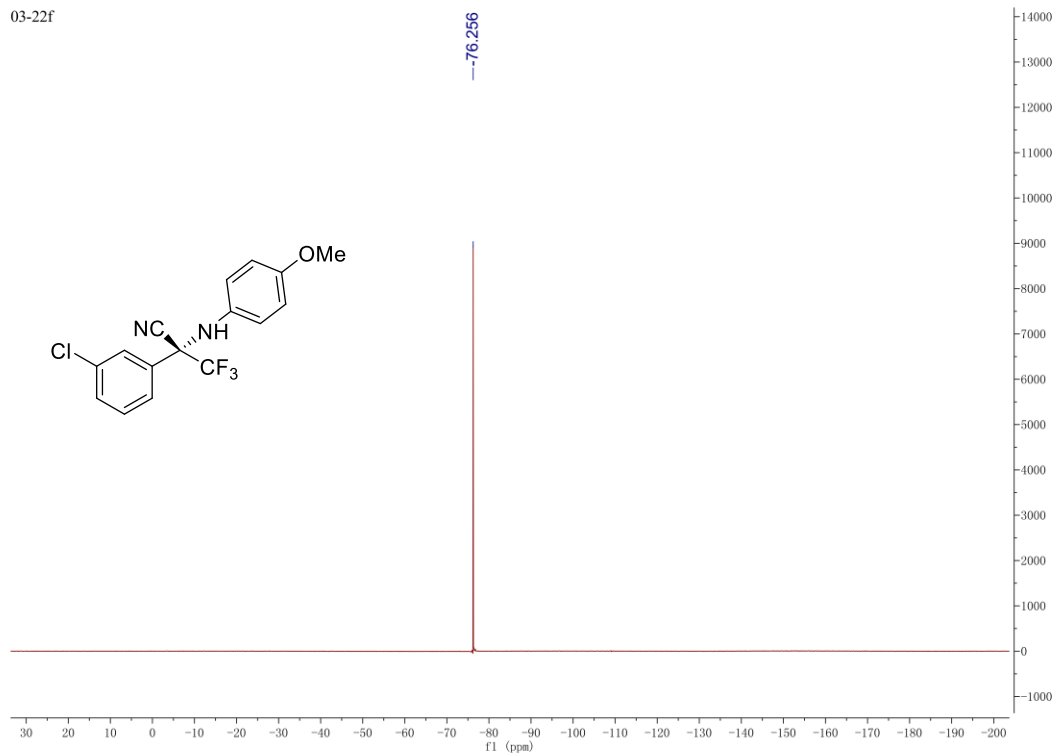
1 Det.A Ch1/254nm

PeakTable

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1	11.780	9046621	405320	95.249	95.699
2	13.213	451249	18218	4.751	4.301
Total		9497870	423538	100.000	100.000



03-22f

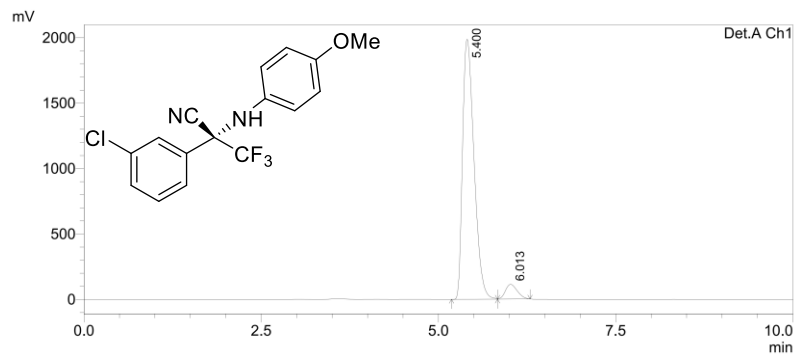


1 Det.A Ch1/254nm

PeakTable

Detector A Ch1 254nm

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1	5.393	29846374	2650647	49.692	52.148
2	6.017	30215811	2432307	50.308	47.852
Total		60062184	5082954	100.000	100.000

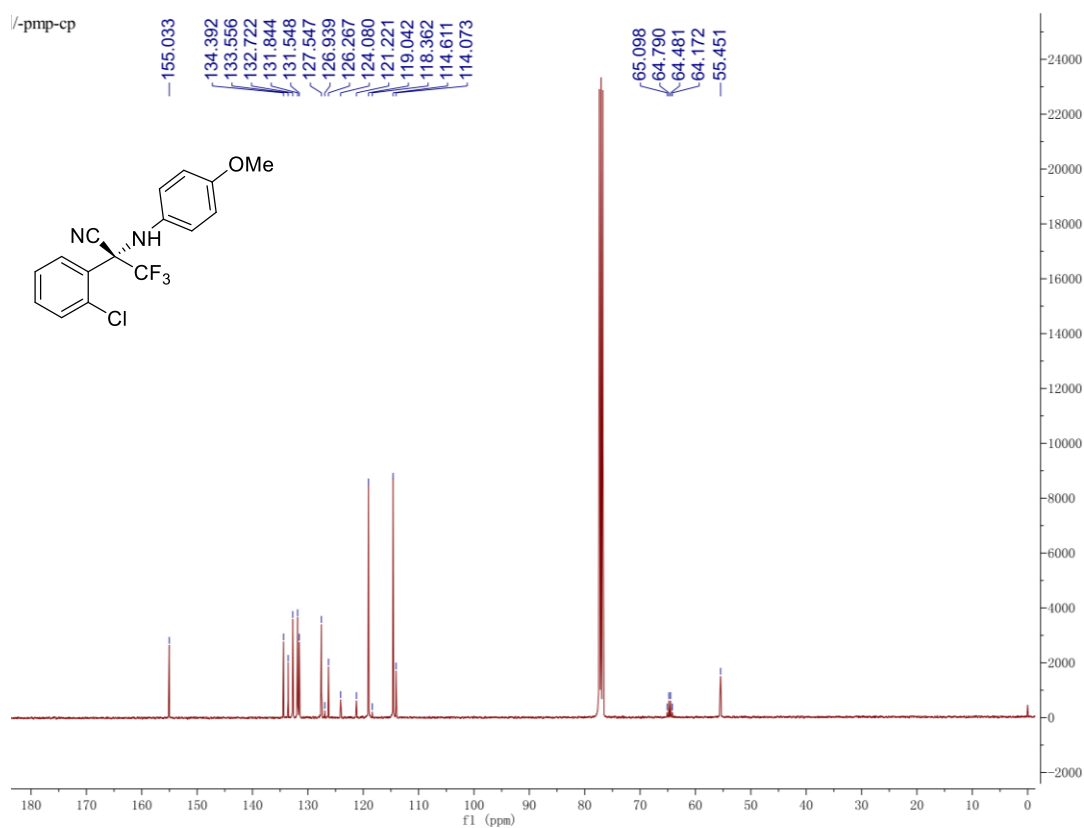
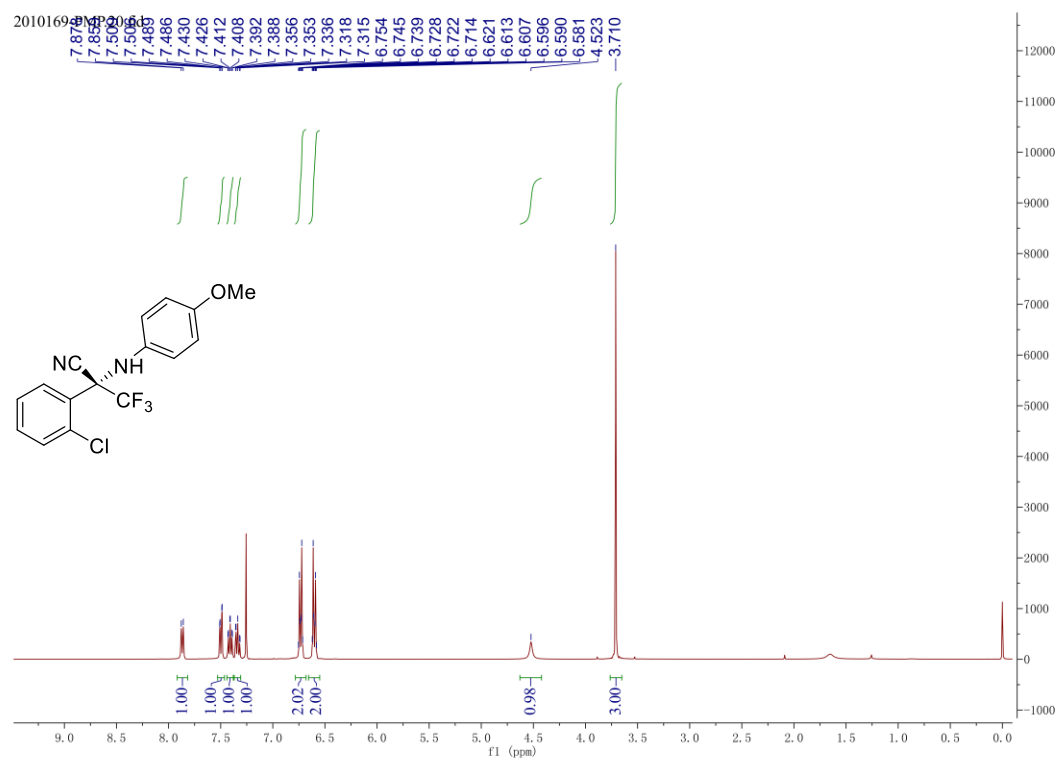


1 Det.A Ch1/254nm

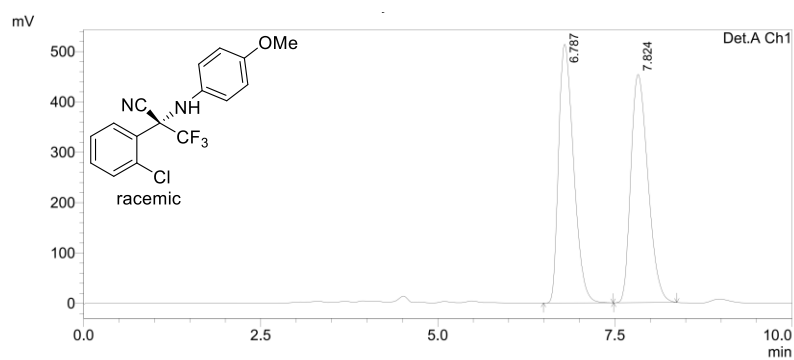
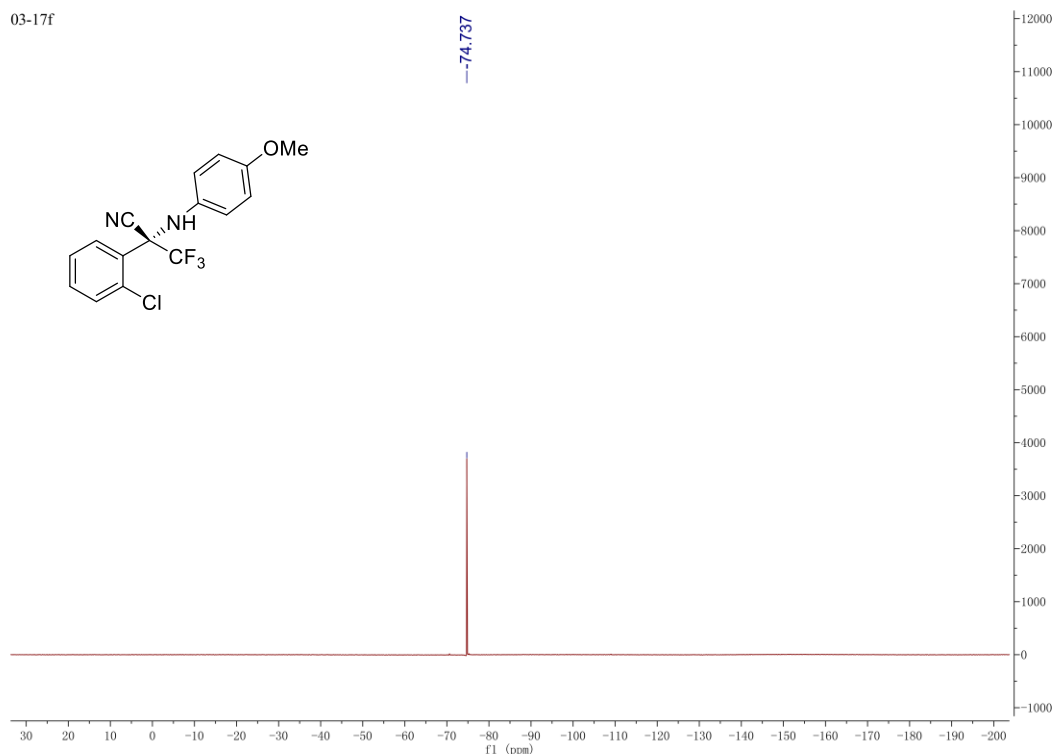
PeakTable

Detector A Ch1 254nm

Peak#	Ret. Time	Area	Height	Area %	Height %
1	5.400	22296505	1986464	94.450	94.638
2	6.013	1310269	112540	5.550	5.362
Total		23606774	2099004	100.000	100.000



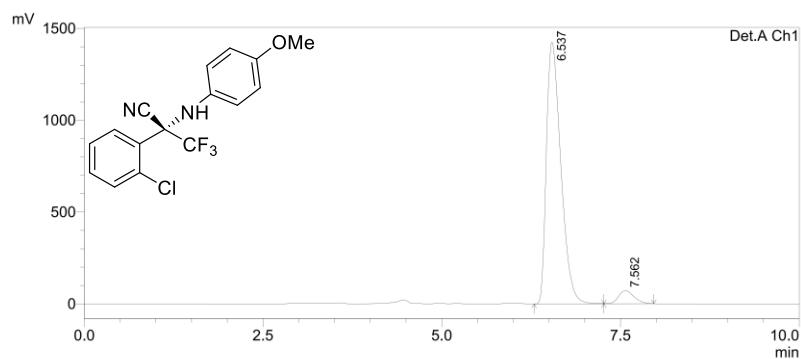
03-17f



1 Det.A Ch1/254nm

PeakTable

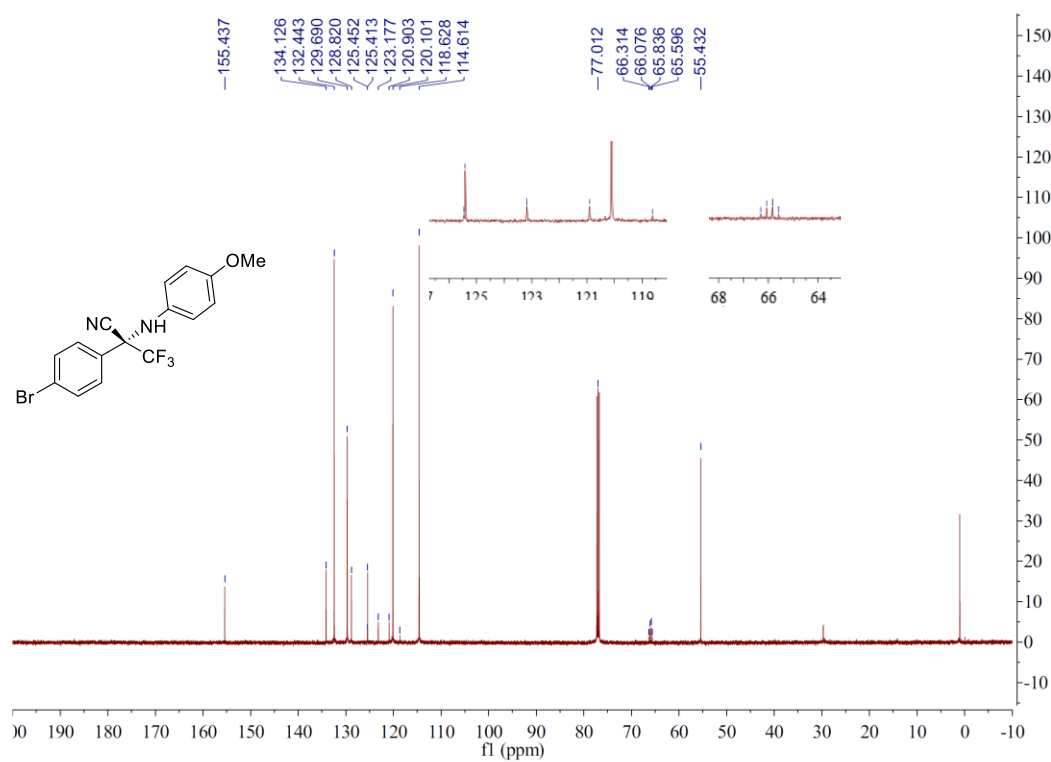
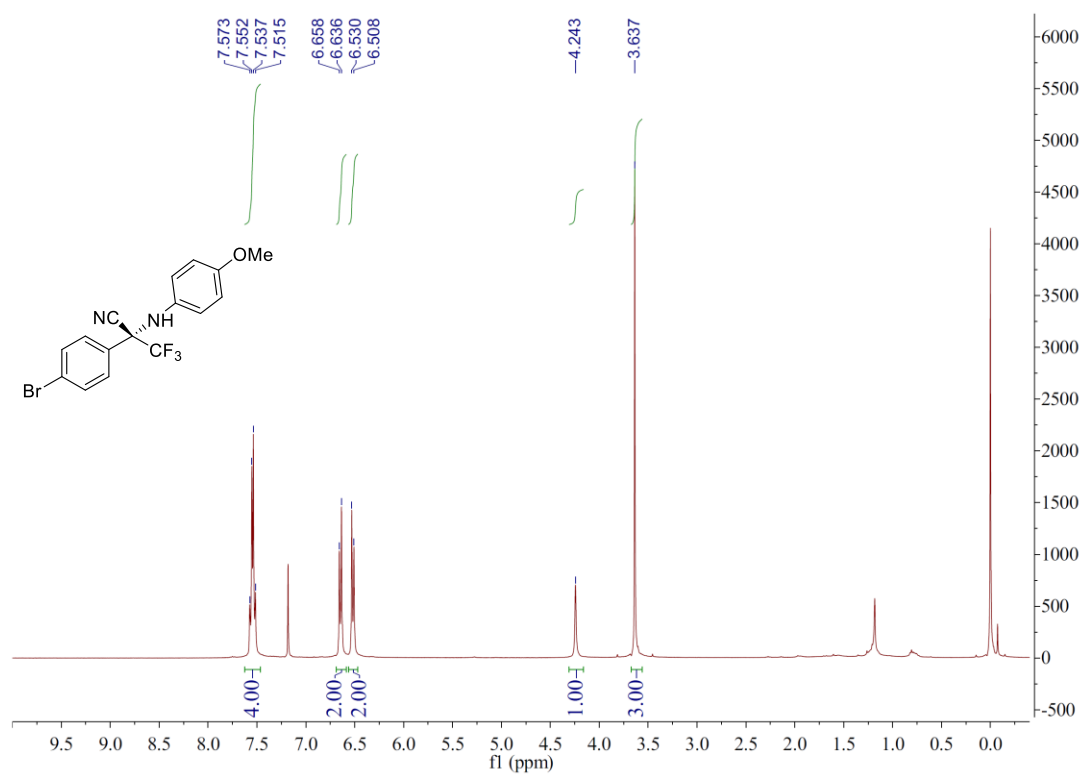
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1	6.787	7591456	513917	49.924	53.141
2	7.824	7614598	453165	50.076	46.859
Total		15206054	967082	100.000	100.000

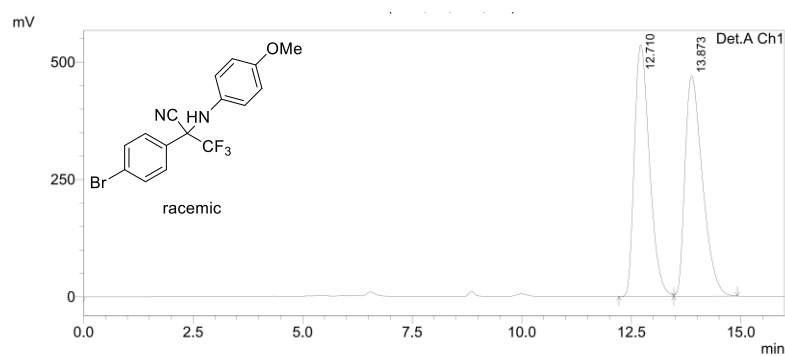
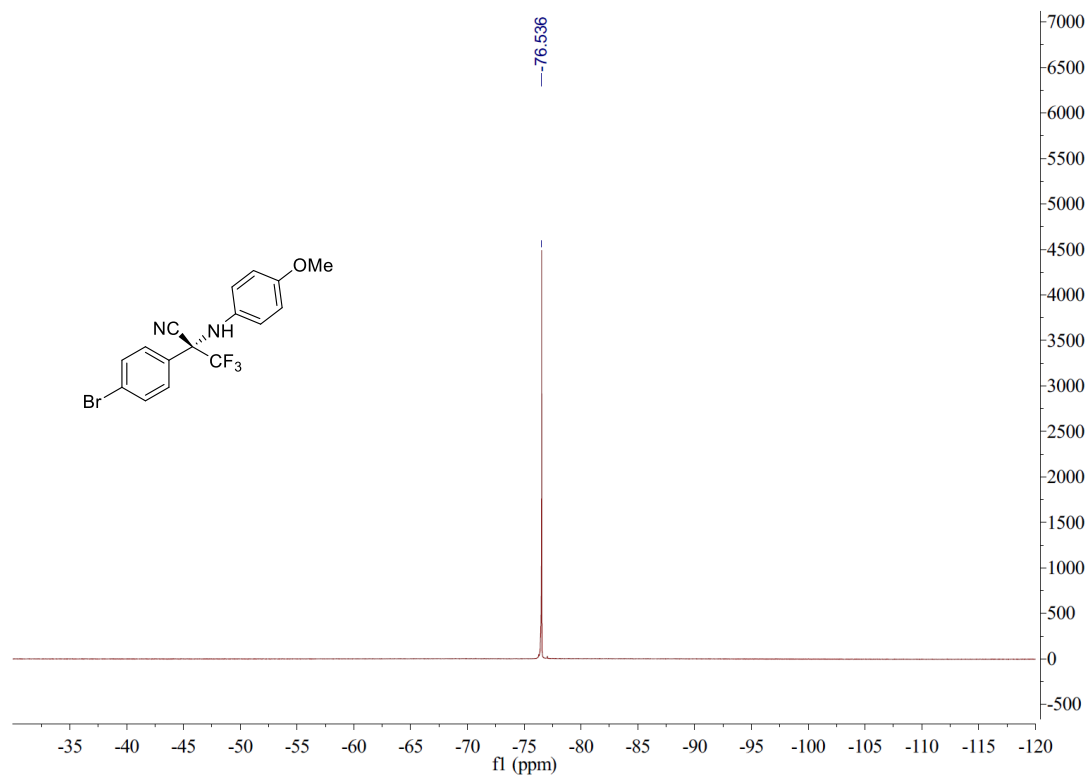


1 Det.A Ch1/254nm

PeakTable

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1	6.537	20024440	1426112	94.921	95.244
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Total		21095980	1497318	100.000	100.000

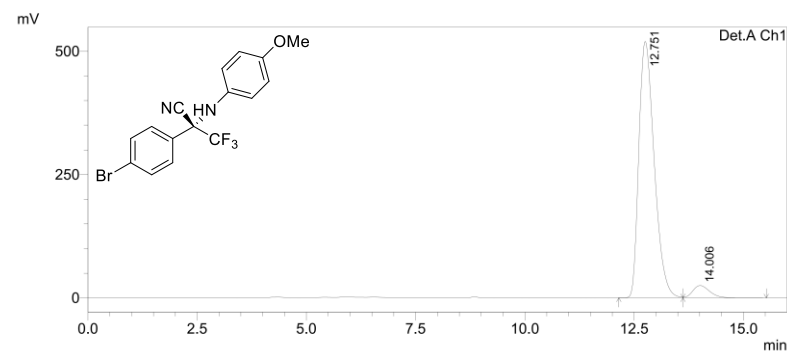




1 Det.A Ch1/254nm

PeakTable

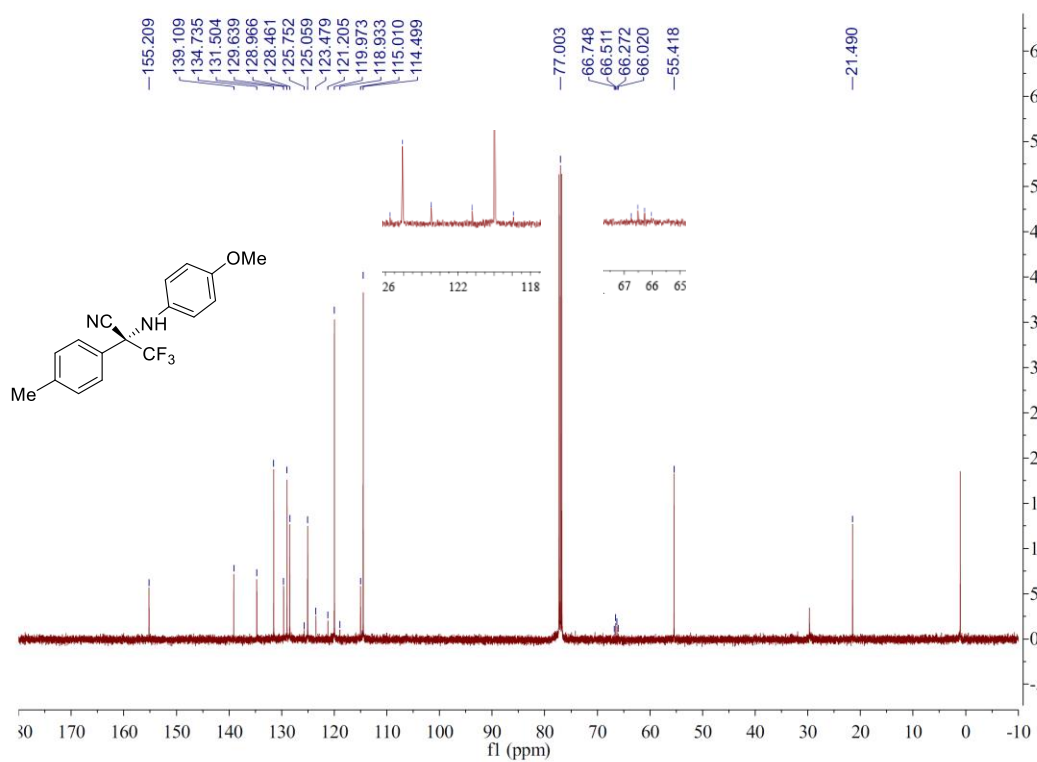
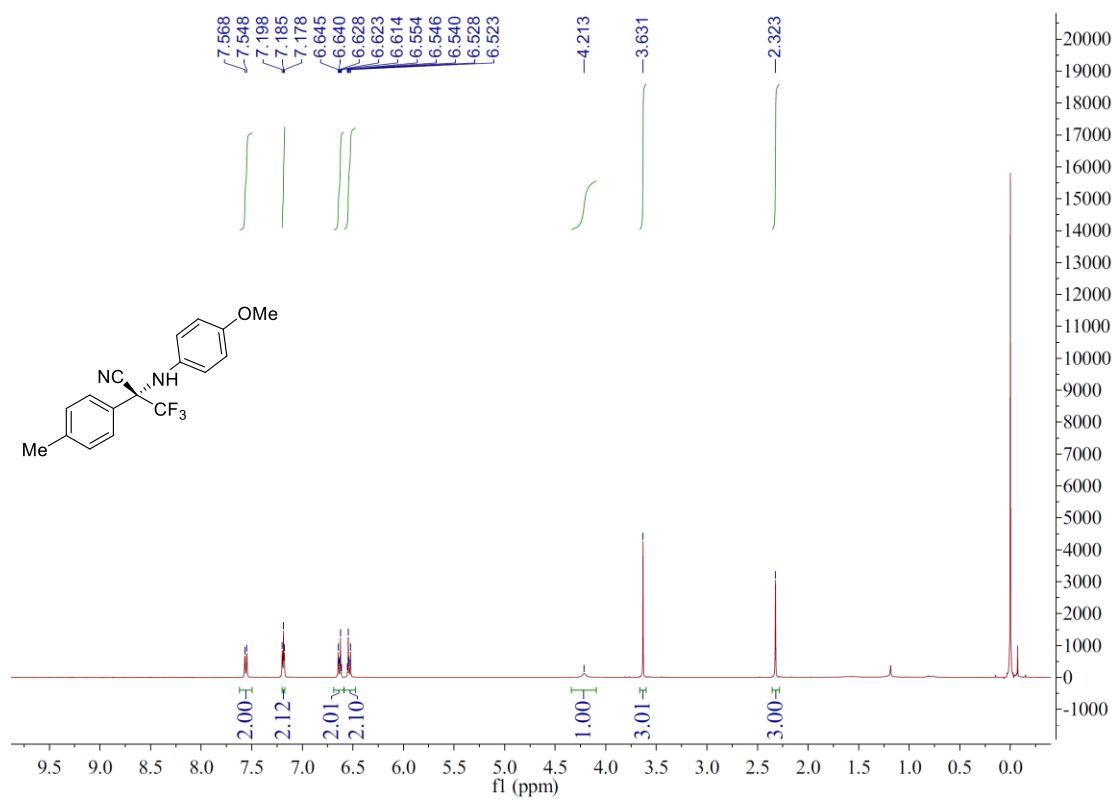
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1	12.710	12880066	536272	49.702	53.345
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Total		25914668	1005283	100.000	100.000

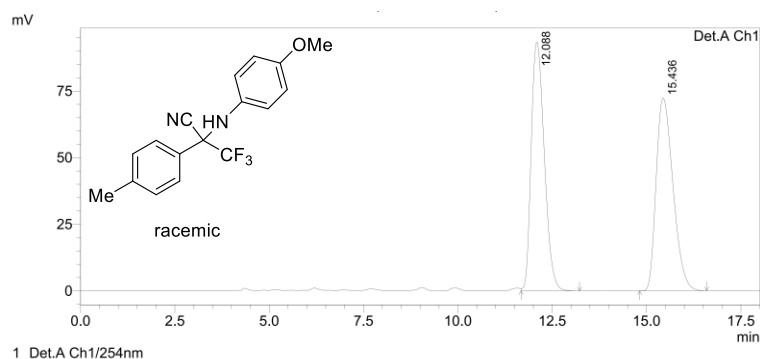
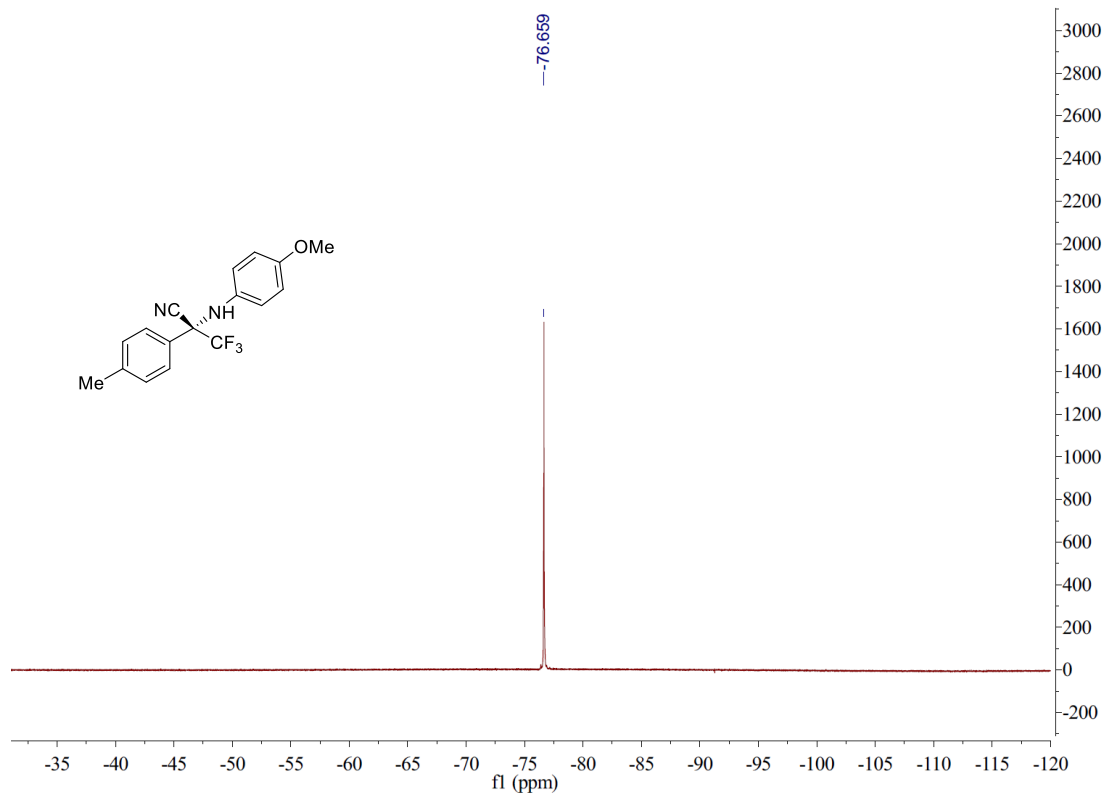


1 Det.A Ch1/254nm

PeakTable

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1	12.751	12739425	519876	94.961	95.408
2	14.006	675980	25021	5.039	4.592
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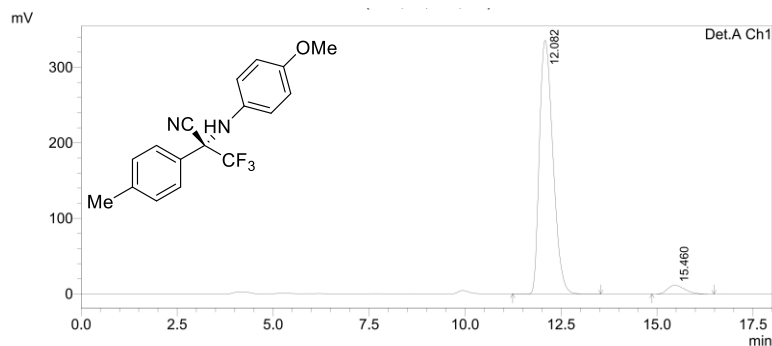




1 Det.A Ch1/254nm

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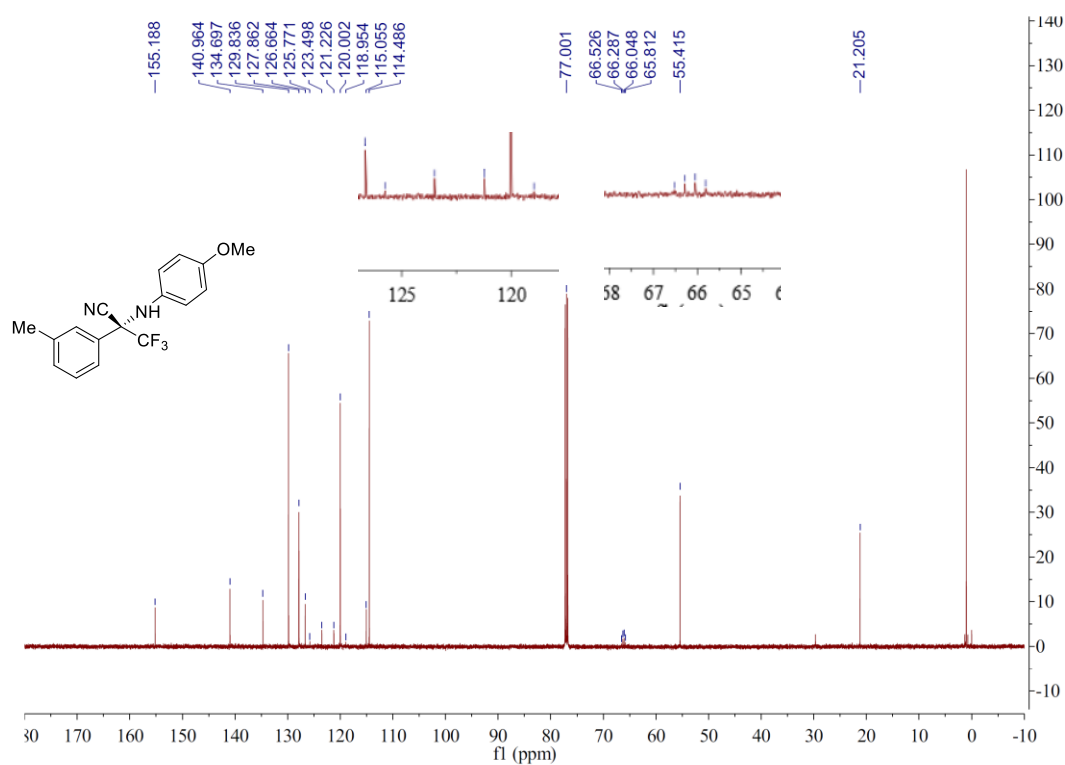
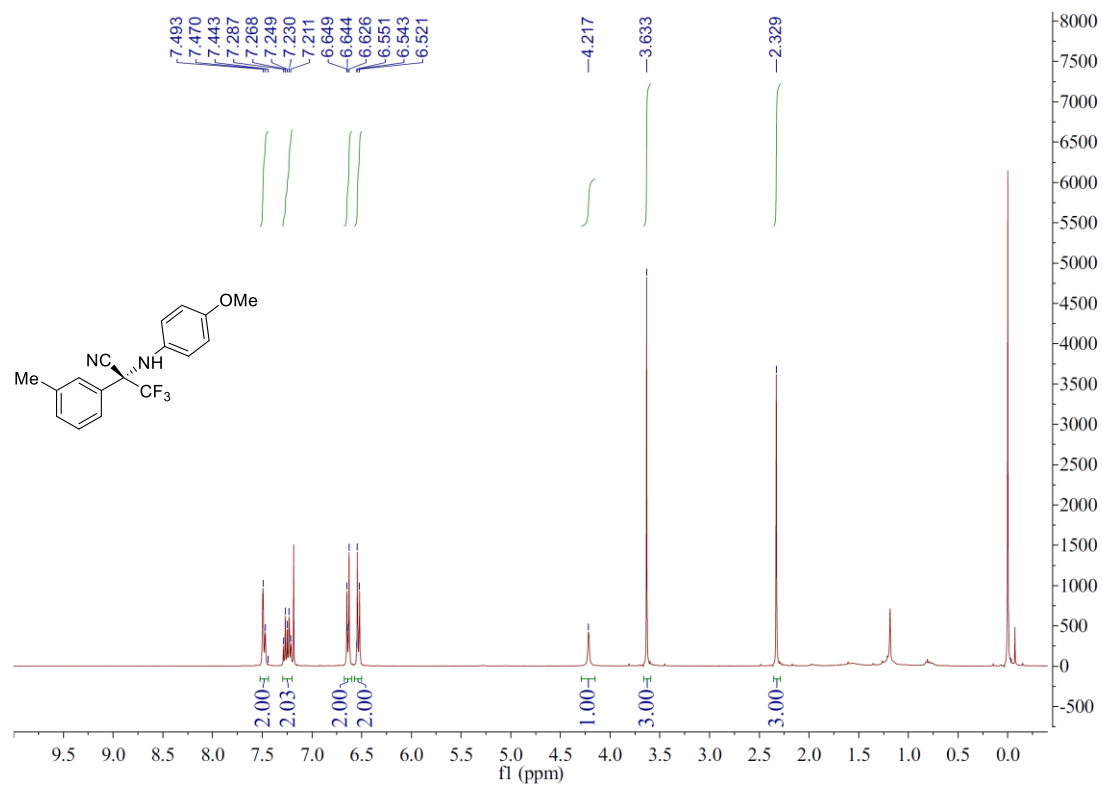
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1	12.088	2255617	93437	50.047	56.353
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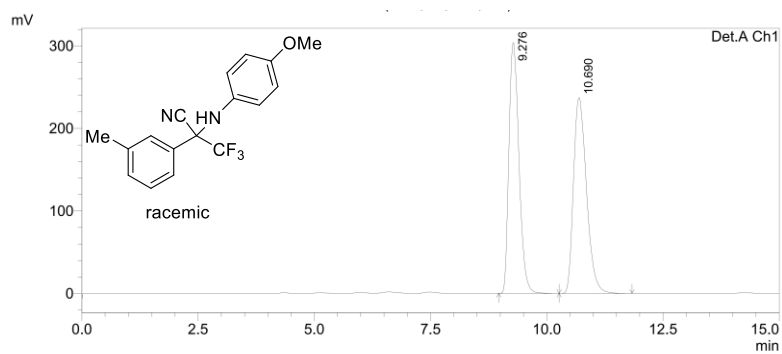
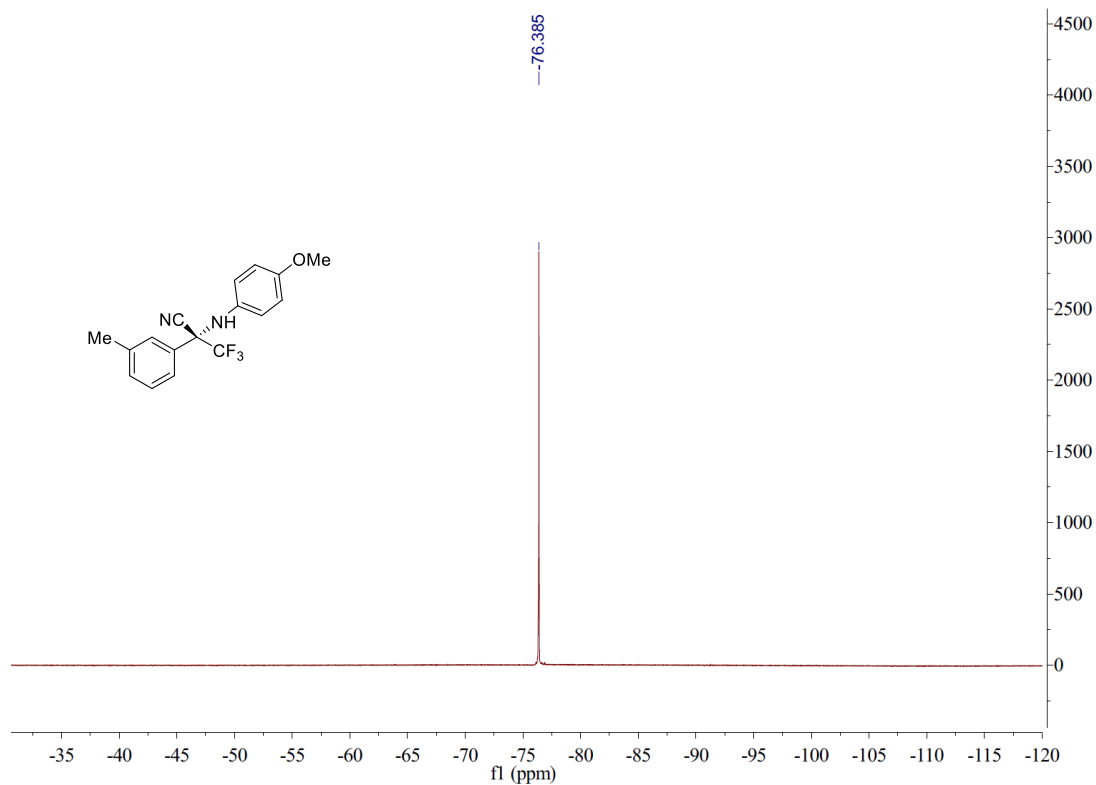


1 Det.A Ch1/254nm

PeakTable

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2	15.460	357297	11559	4.162	3.327
Total		8585026	347409	100.000	100.000

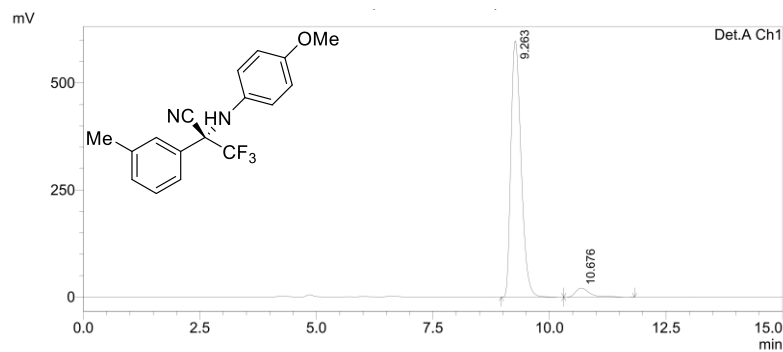




1 Det.A Ch1/254nm

PeakTable

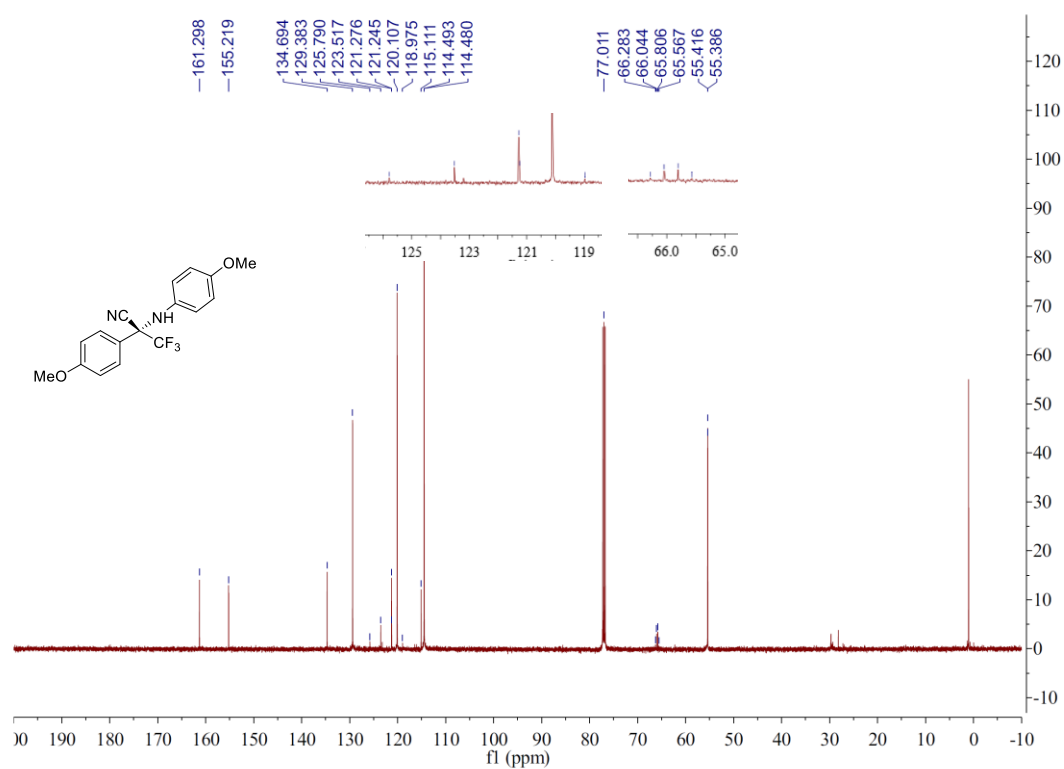
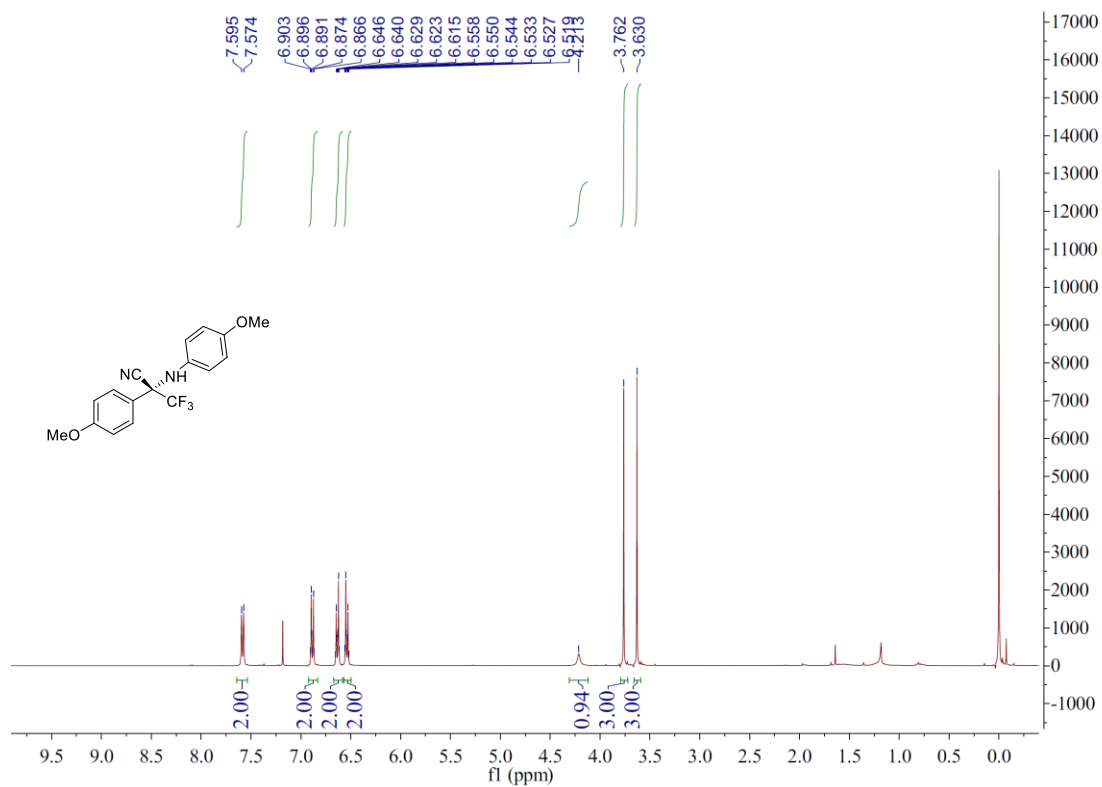
Detector A Ch1 254nm					
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1	9.276	4569402	303896	49.698	56.158
2	10.690	4624961	237244	50.302	43.842
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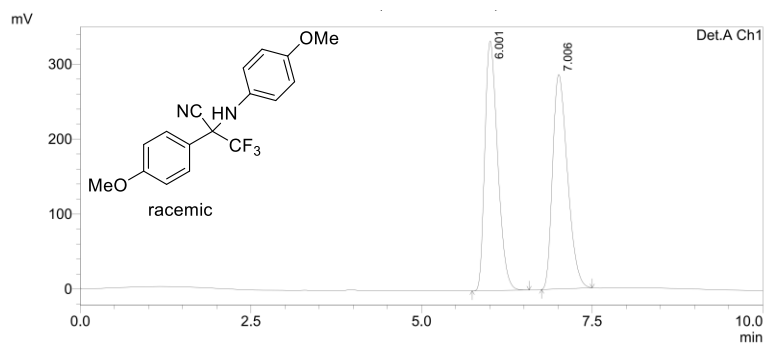
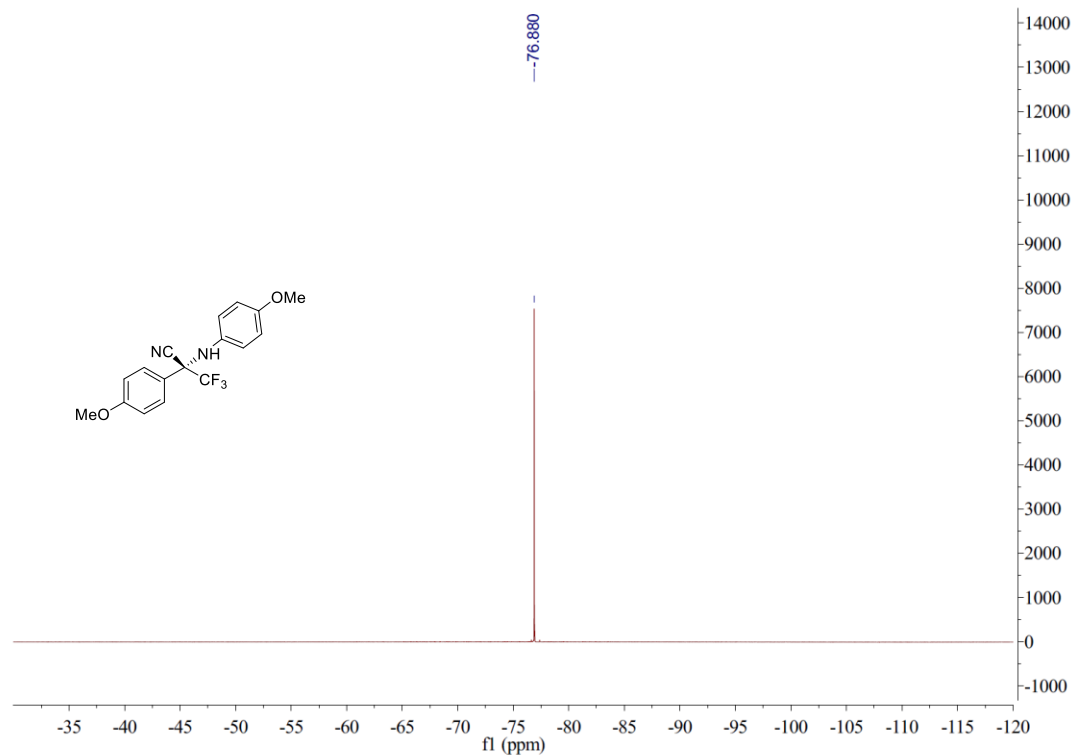


1 Det.A Ch1/254nm

PeakTable

Detector A Ch1 254nm					
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1	9.263	9101589	598514	95.392	96.622
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Total		9541284	619437	100.000	100.000

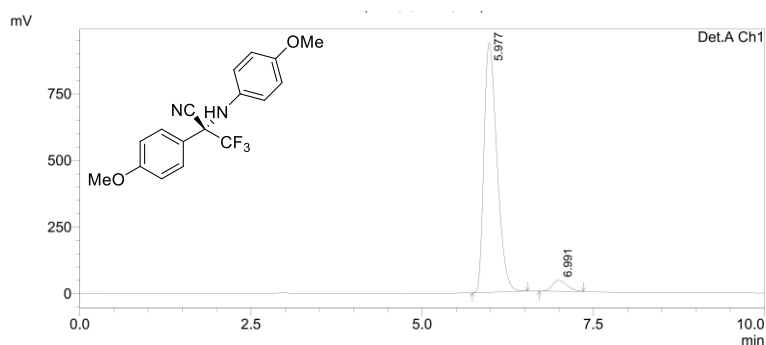




1 Det.A Ch1/254nm

PeakTable

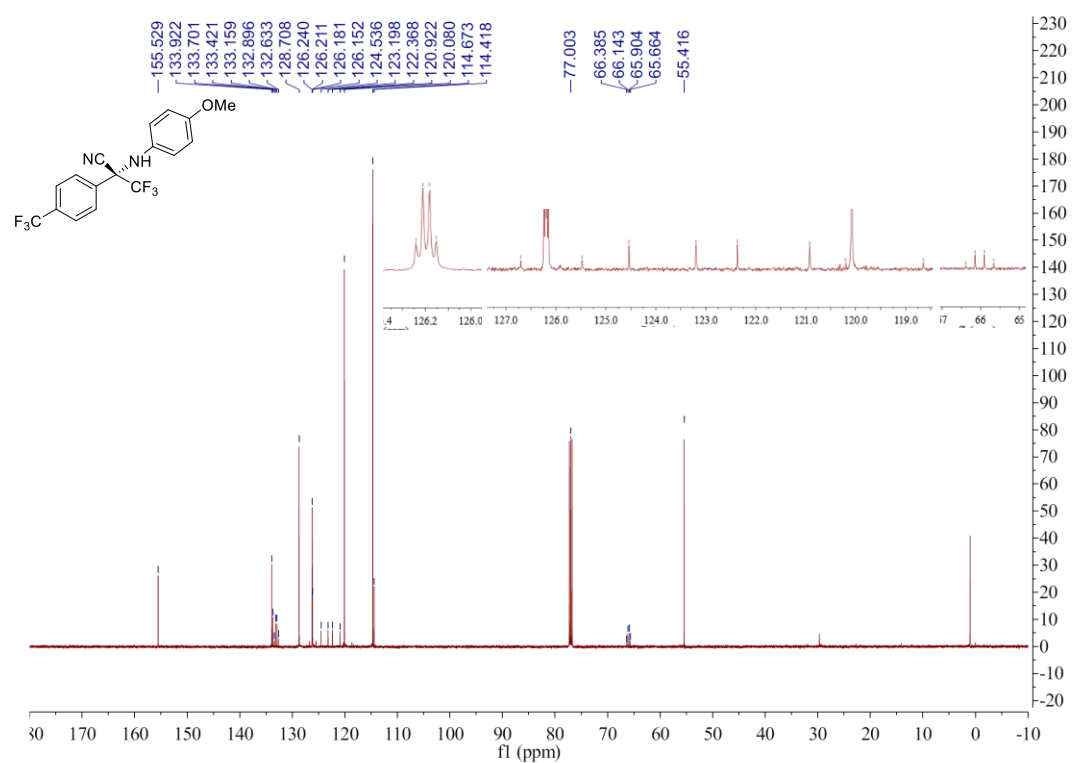
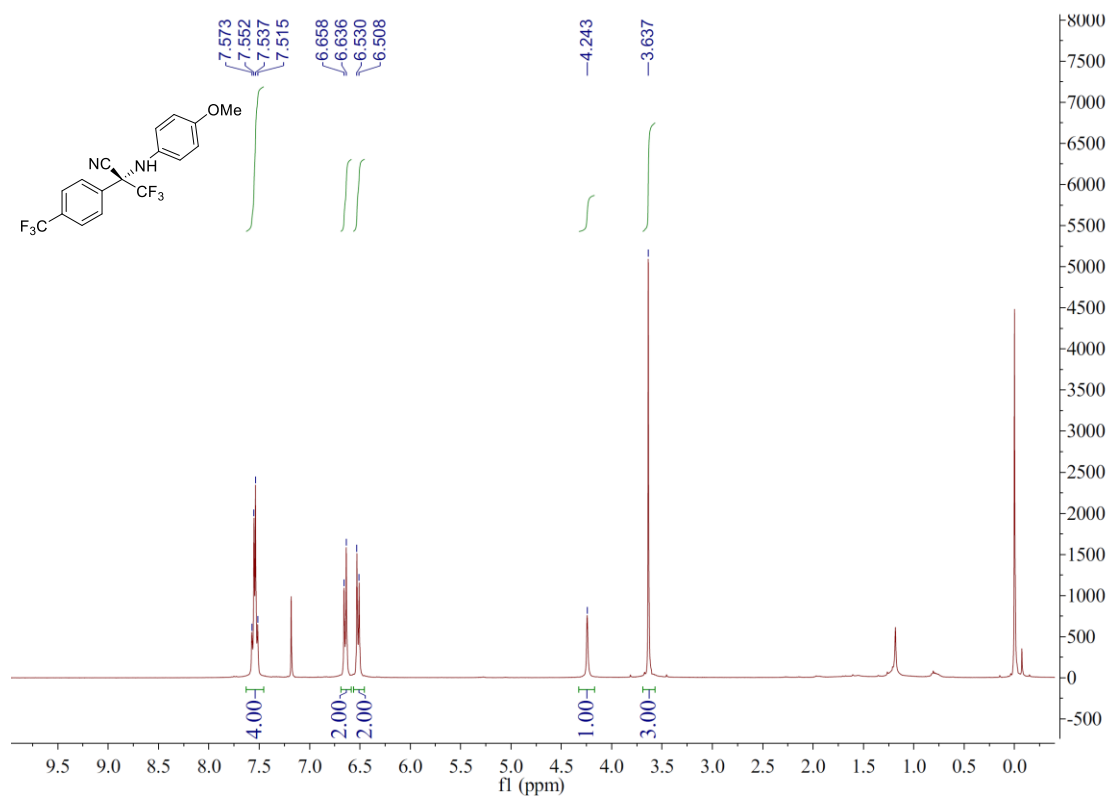
Peak#	Ret. Time	Area	Height	Area %	Height %
1	6.001	4334083	333015	49.977	53.786
2	7.006	4338000	286132	50.023	46.214
Total		8672083	619147	100.000	100.000

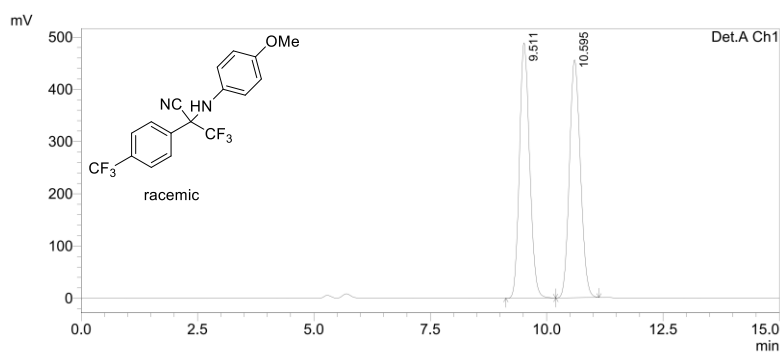
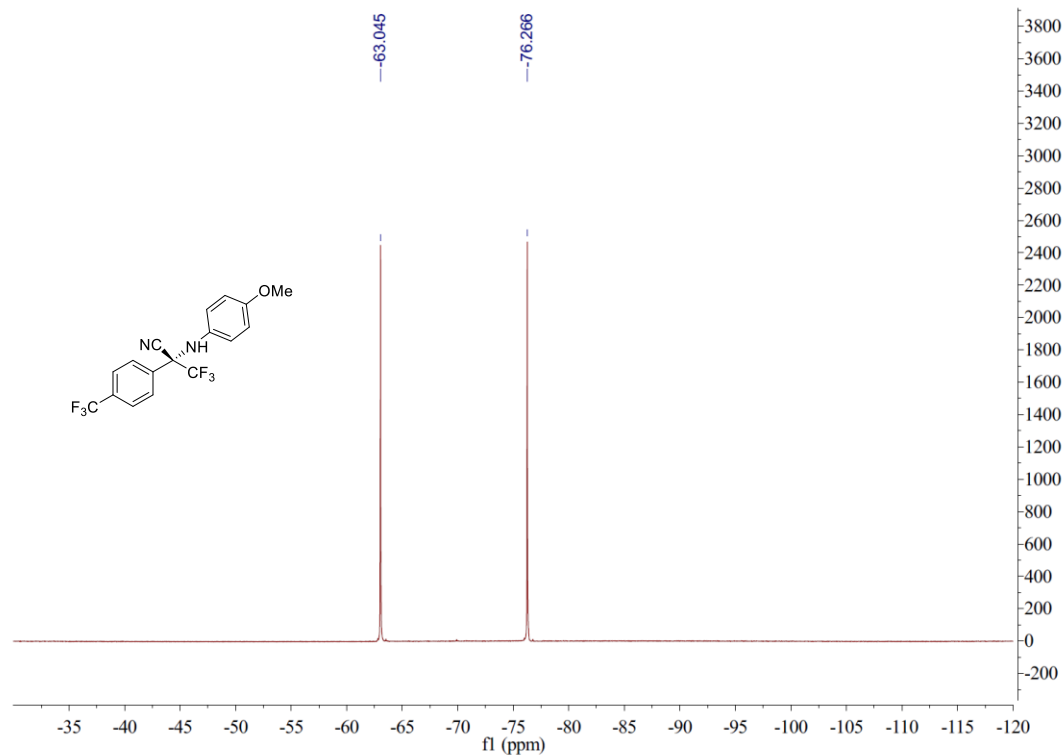


1 Det.A Ch1/254nm

PeakTable

Peak#	Ret. Time	Area	Height	Area %	Height %
1	5.977	12319889	935441	95.376	95.827
2	6.991	597334	40738	4.624	4.173
Total		12917223	976179	100.000	100.000



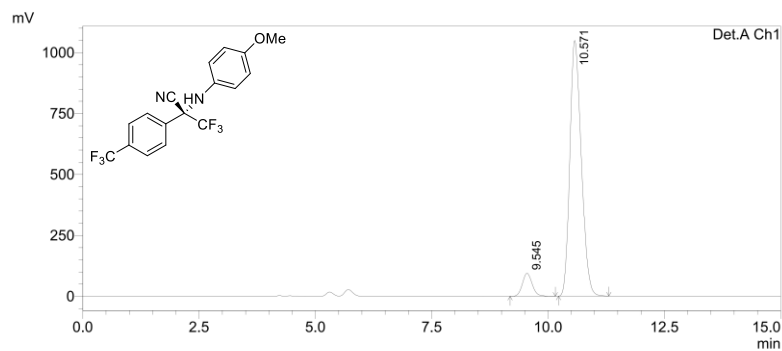


1 Det.A Ch1/254nm

PeakTable

Detector A Ch1 254nm

Peak#	Ret. Time	Area	Height	Area %	Height %
1	9.511	7417456	488545	49.775	51.766
2	10.595	7484436	455209	50.225	48.234
Total		14901892	943754	100.000	100.000

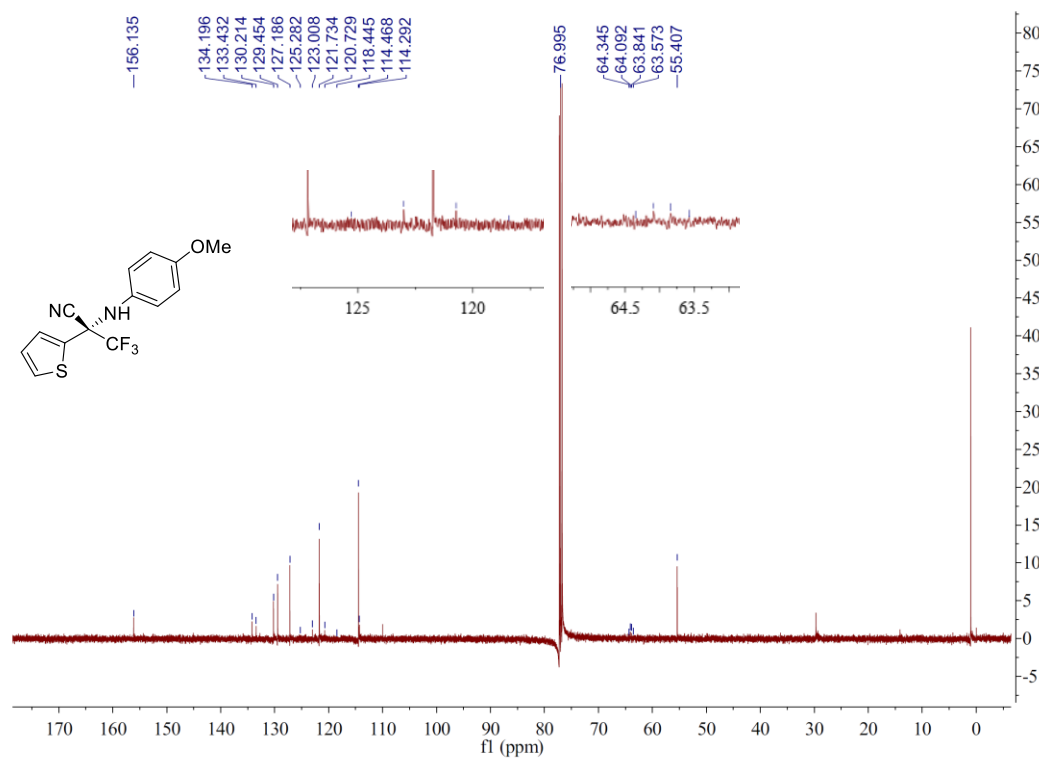
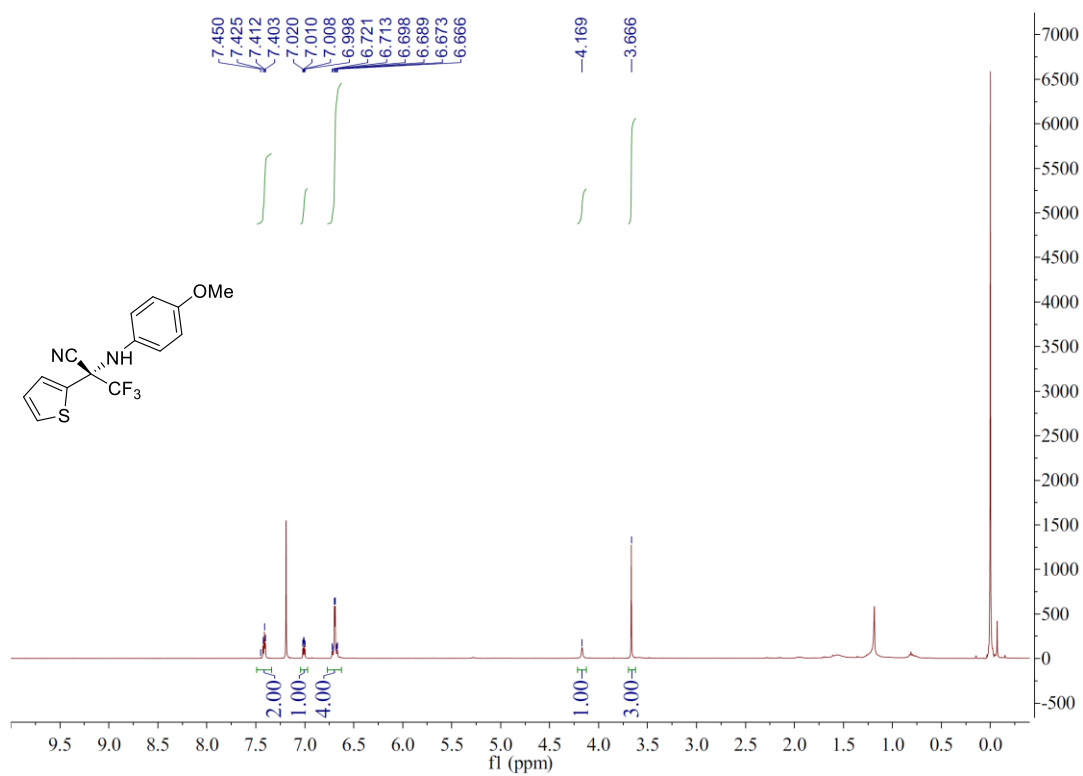


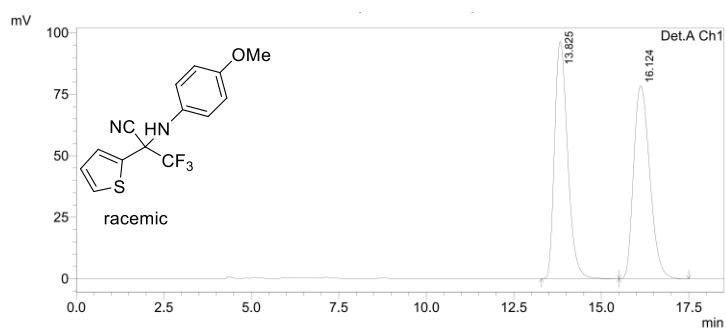
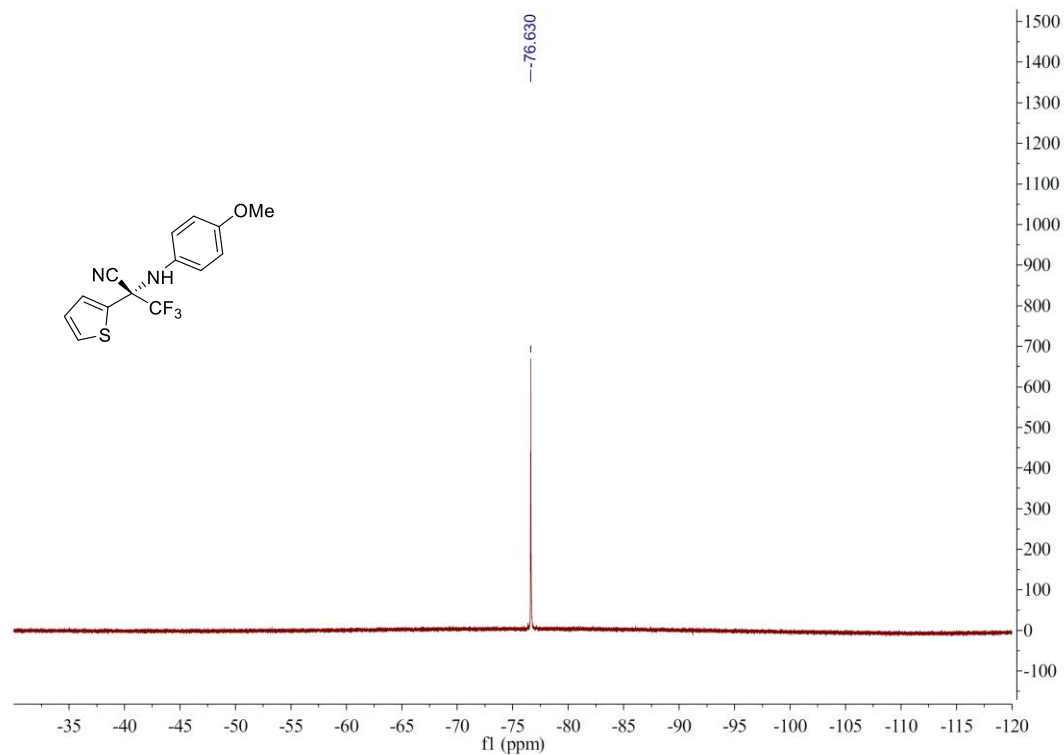
1 Det.A Ch1/254nm

PeakTable

Detector A Ch1 254nm

Peak#	Ret. Time	Area	Height	Area %	Height %
1	9.545	1390496	94677	7.205	8.280
2	10.571	17908298	1048827	92.795	91.720
Total		19298794	1143504	100.000	100.000

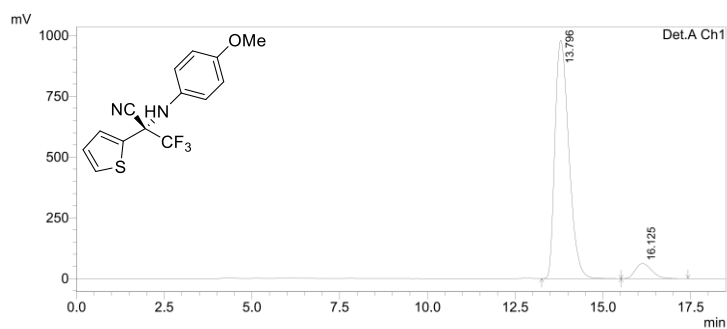




1 Det.A Ch1/254nm

PeakTable

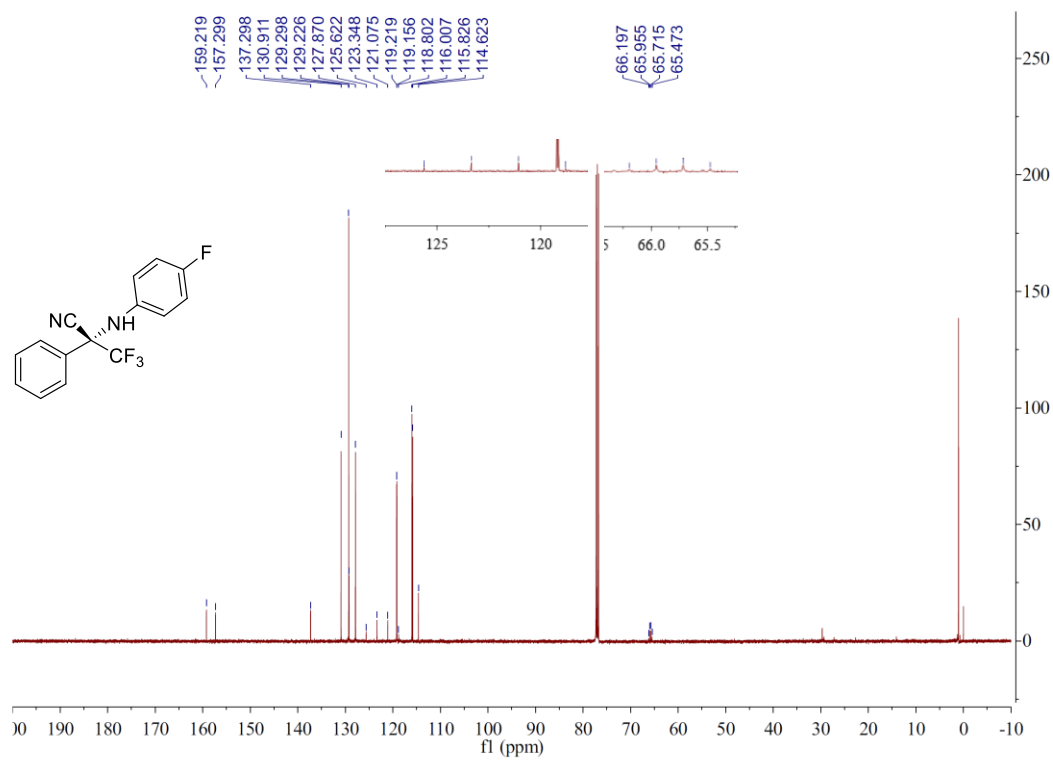
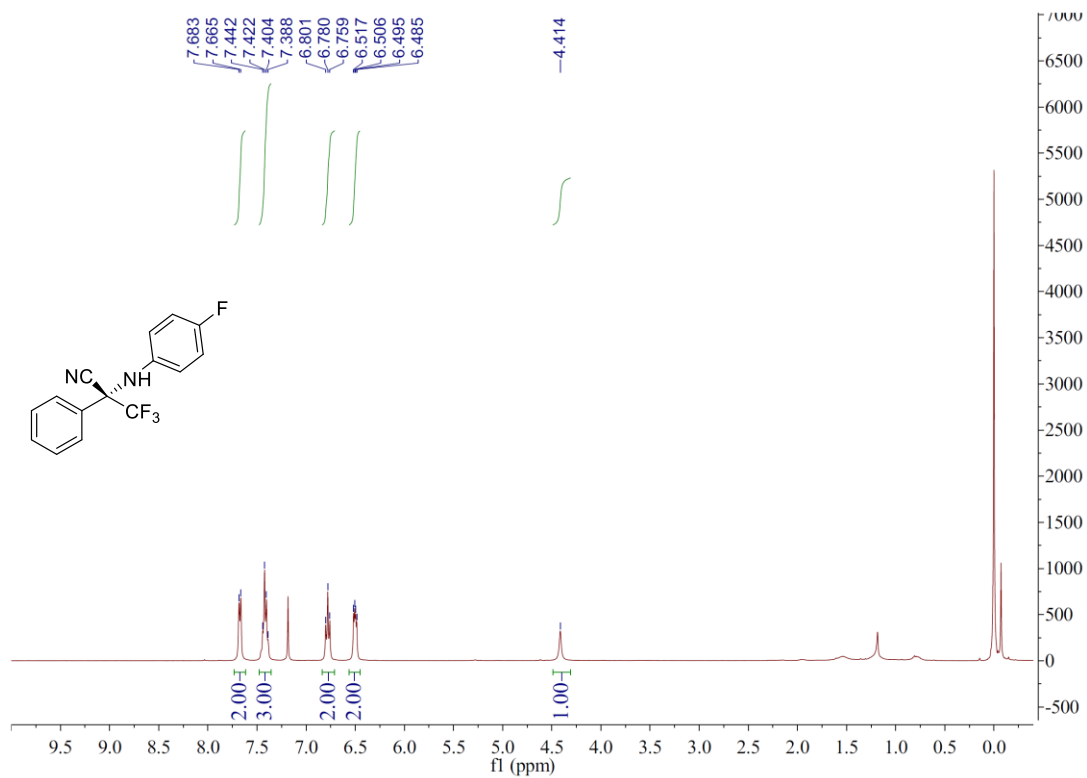
Peak#	Ret. Time	Area	Height	Area %	Height %
1	13.825	2506689	96429	49.894	55.153
2	16.124	2517294	78409	50.106	44.847
Total		5023983	174839	100.000	100.000

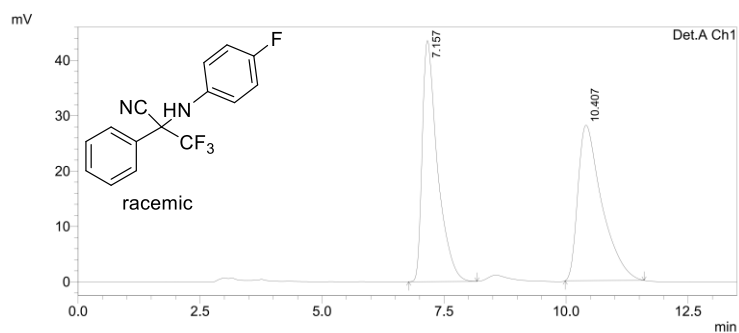
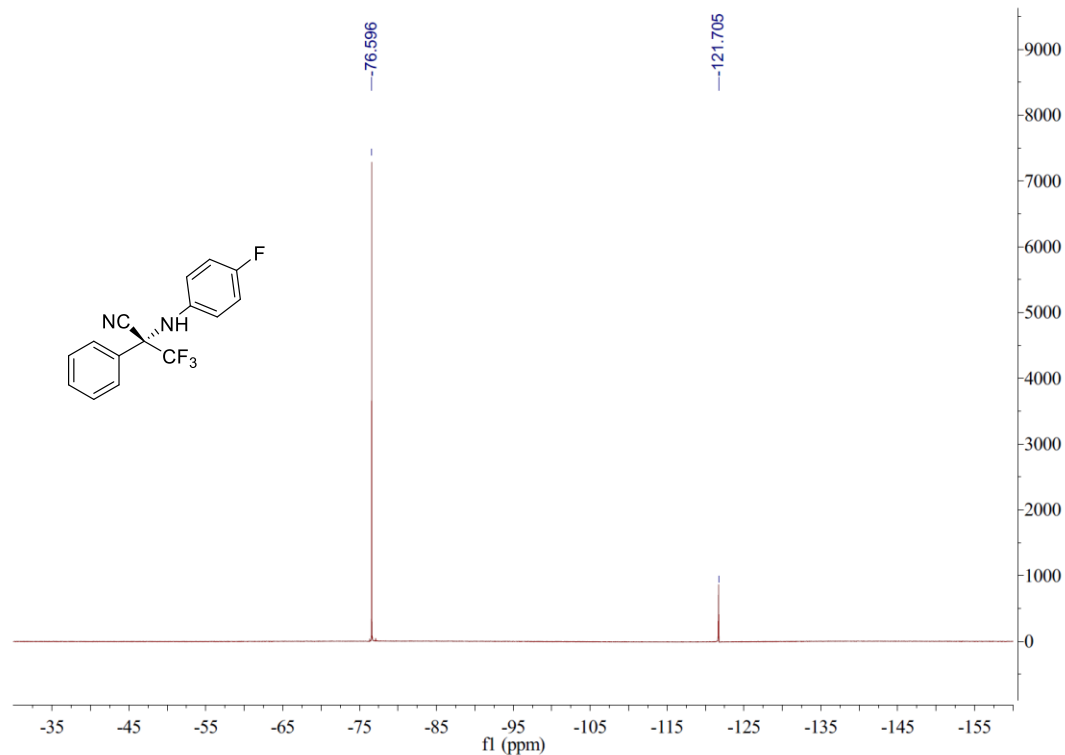


1 Det.A Ch1/254nm

PeakTable

Peak#	Ret. Time	Area	Height	Area %	Height %
1	13.796	27021081	980862	93.176	94.094
2	16.125	1978838	61562	6.824	5.906
Total		28999919	1042424	100.000	100.000

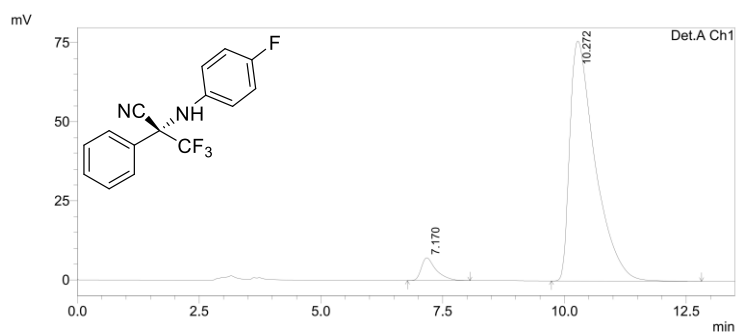




1 Det.A Ch1/254nm

PeakTable

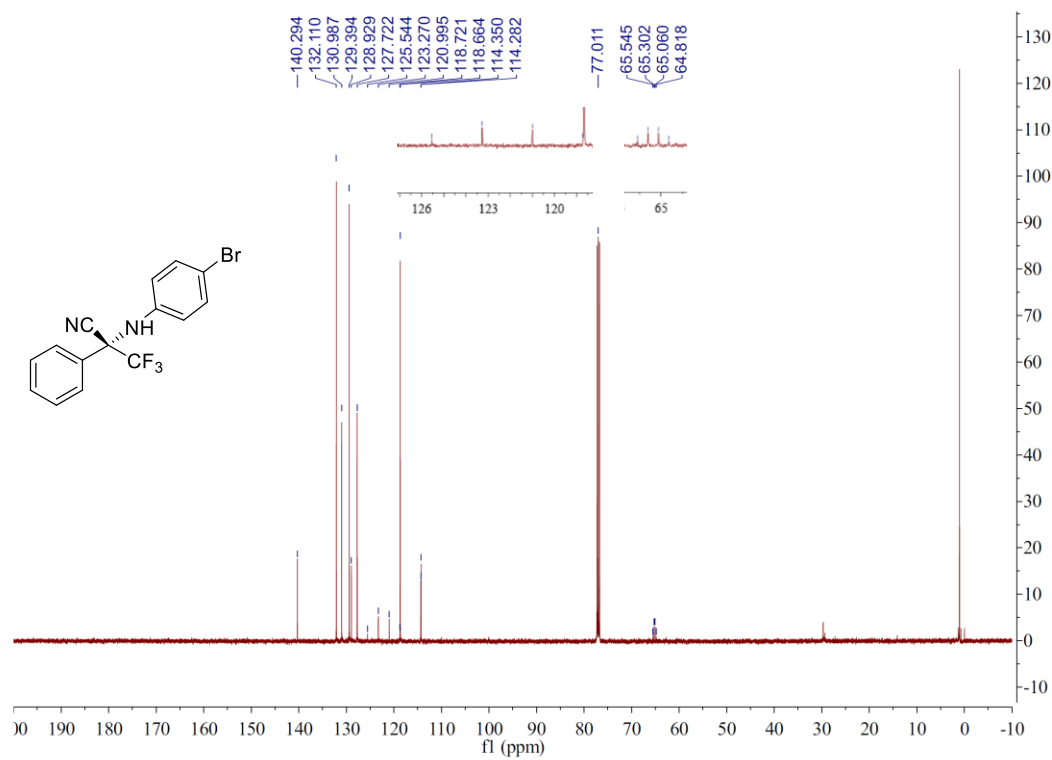
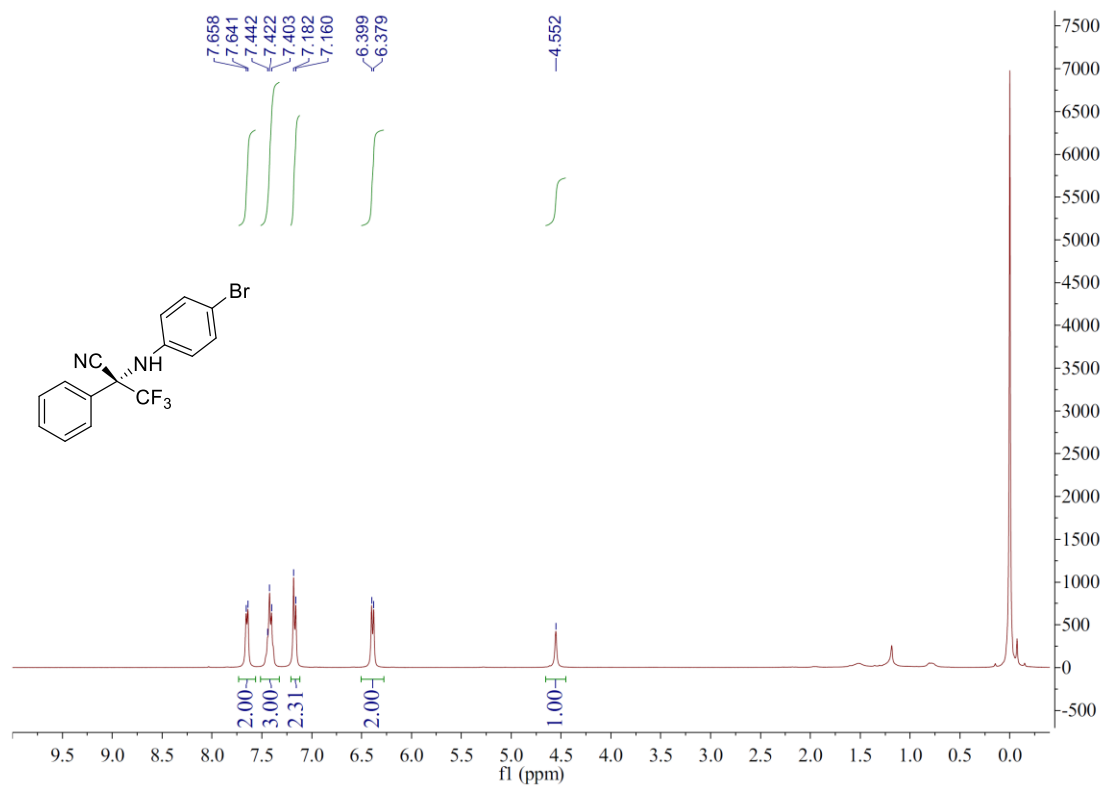
Peak#	Ret. Time	Area	Height	Area %	Height %
1	7.157	944140	43542	49.578	60.802
2	10.407	960208	28070	50.422	39.198
Total		1904349	71612	100.000	100.000

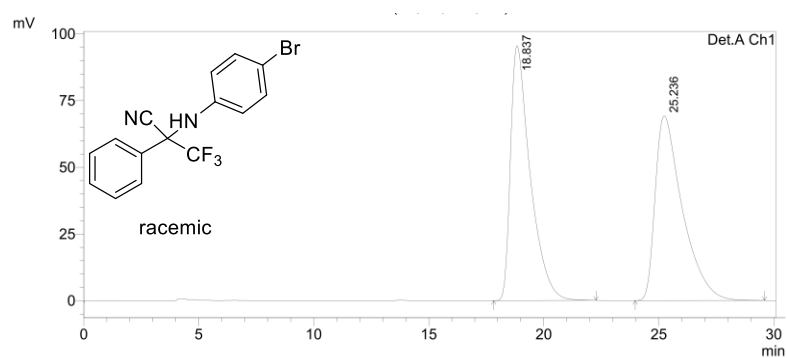
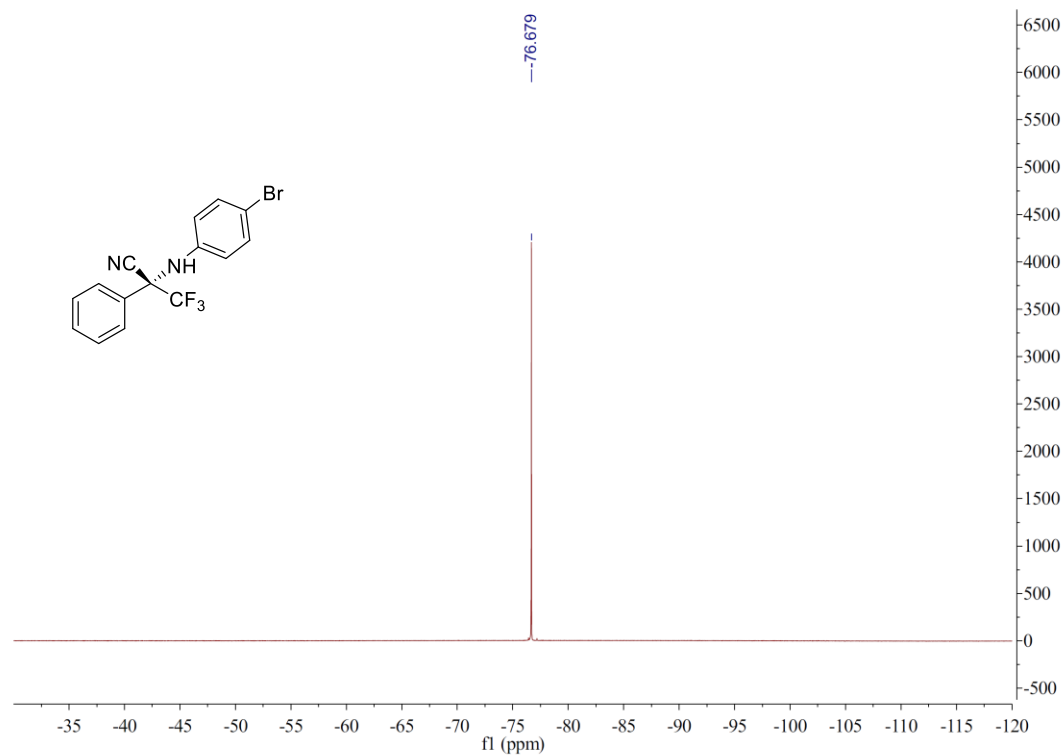


1 Det.A Ch1/254nm

PeakTable

Peak#	Ret. Time	Area	Height	Area %	Height %
1	7.170	151659	7200	5.240	8.671
2	10.272	2742313	75836	94.760	91.329
Total		2893972	83036	100.000	100.000

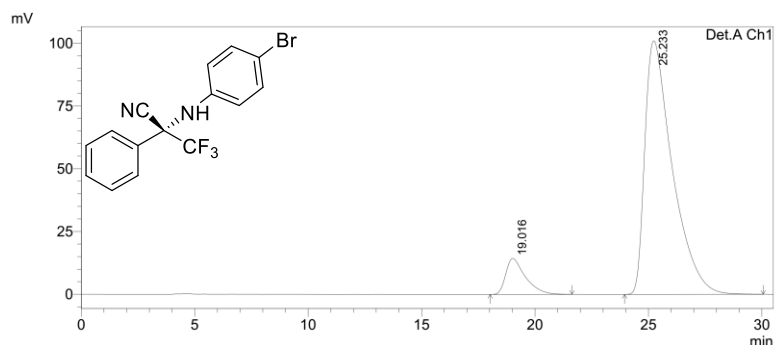




1 Det.A Ch1/254nm

PeakTable

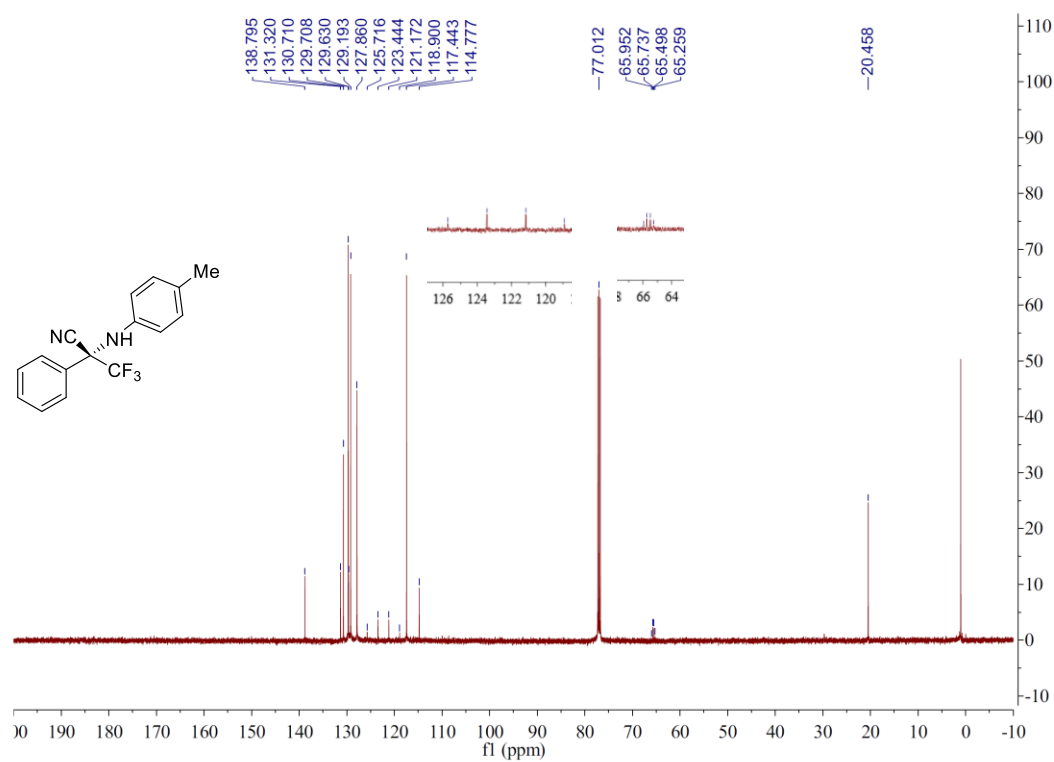
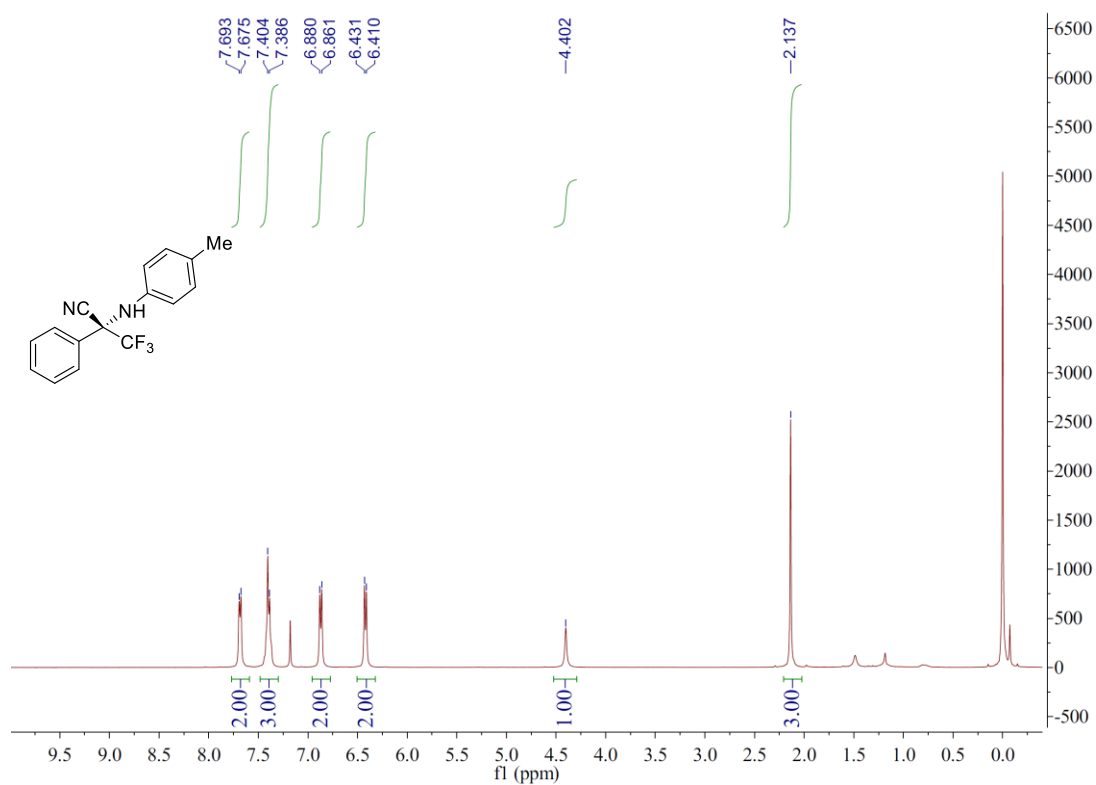
Detector A Ch1 254nm					
Peak#	Ret. Time	Area	Height	Area %	Height %
1	18.837	5684862	95514	49.972	57.981
2	25.236	5691280	69218	50.028	42.019
Total		11376142	164731	100.000	100.000

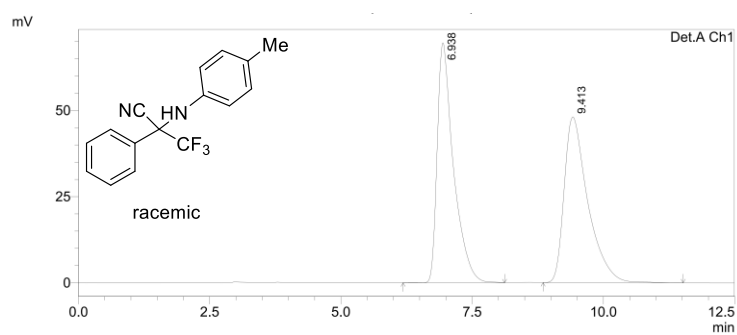
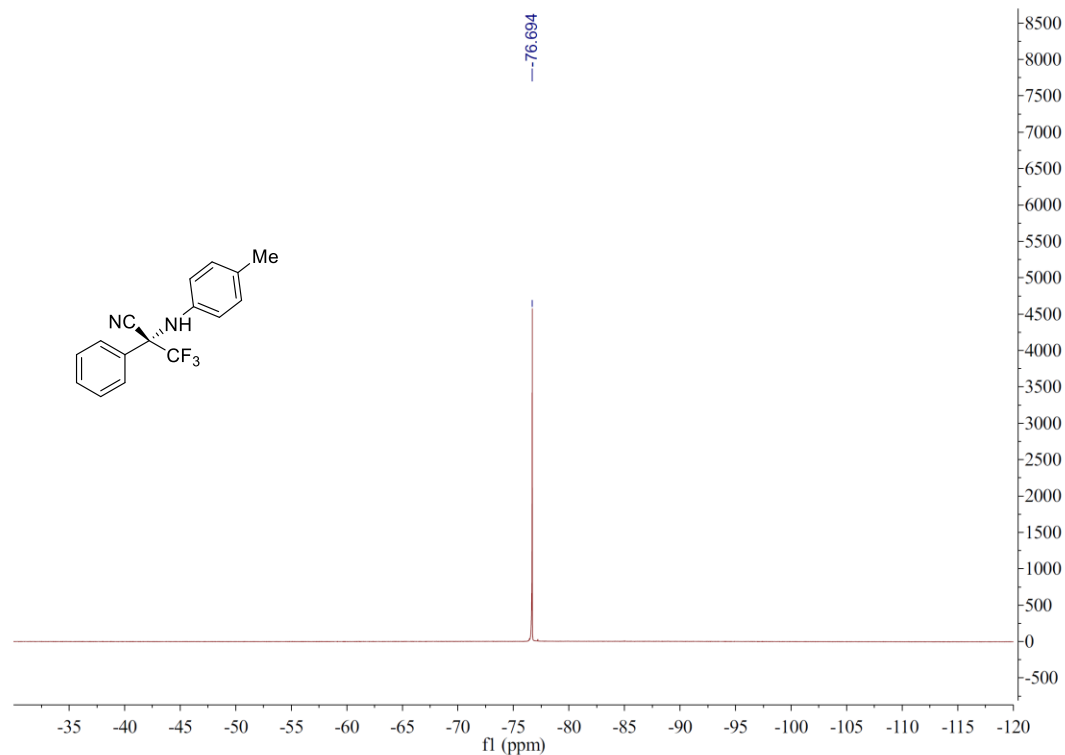


1 Det.A Ch1/254nm

PeakTable

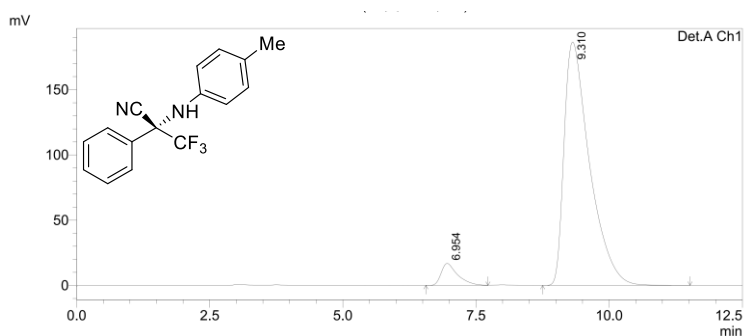
Detector A Ch1 254nm					
Peak#	Ret. Time	Area	Height	Area %	Height %
1	19.016	859340	14318	9.154	12.414
2	25.233	8528152	101013	90.846	87.586
Total		9387493	115331	100.000	100.000





1 Det.A Ch1/254nm

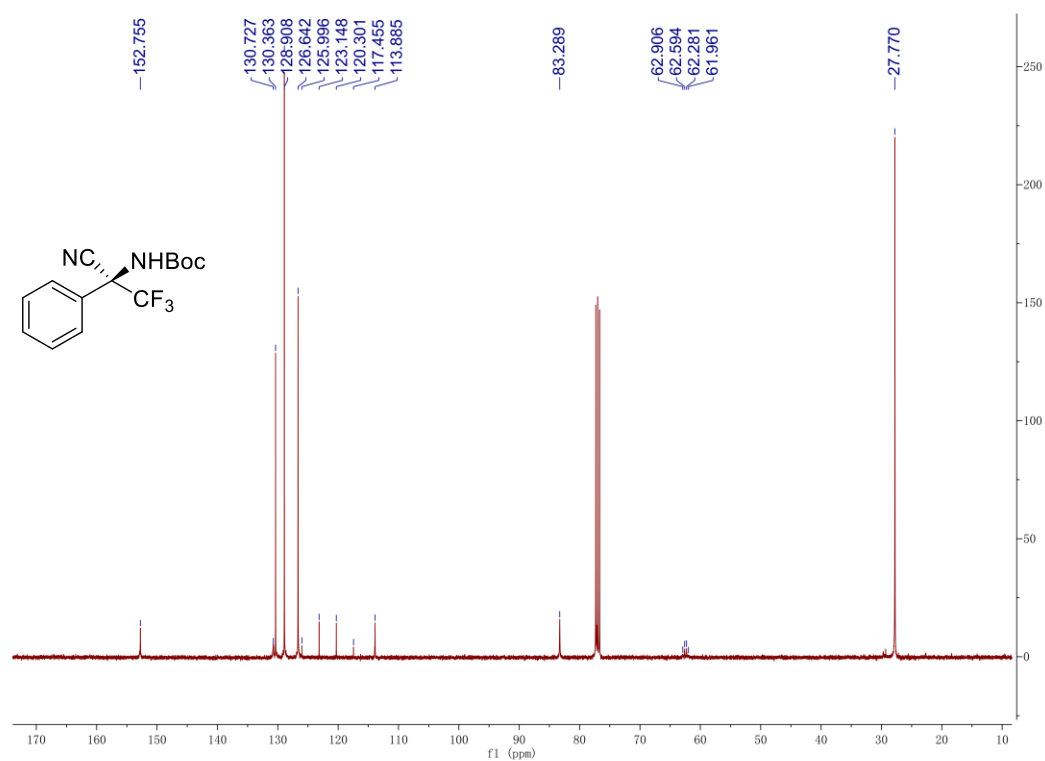
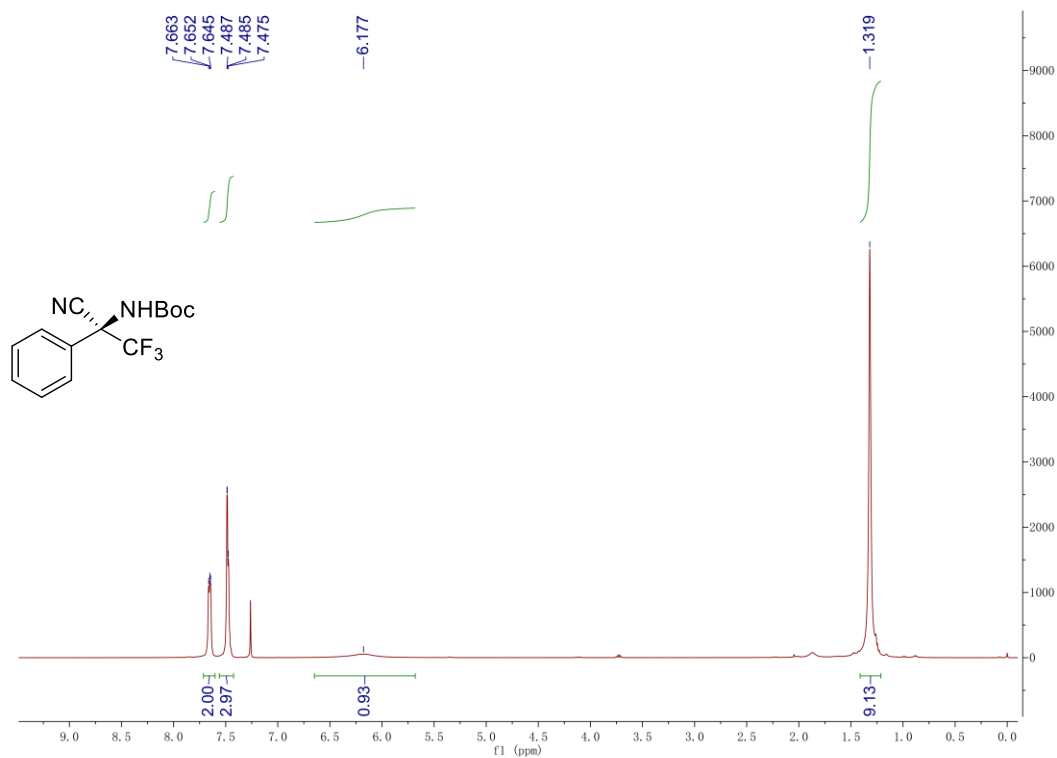
PeakTable					
Peak#	Ret. Time	Area	Height	Area %	Height %
1	6.938	1505337	69618	49.935	59.156
2	9.413	1509241	48067	50.065	40.844
Total		3014578	117685	100.000	100.000

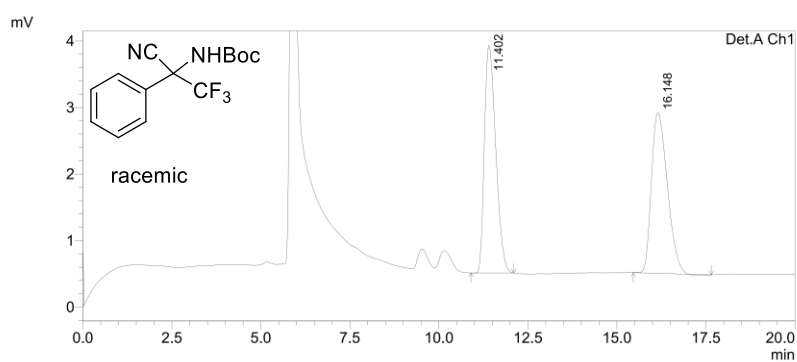
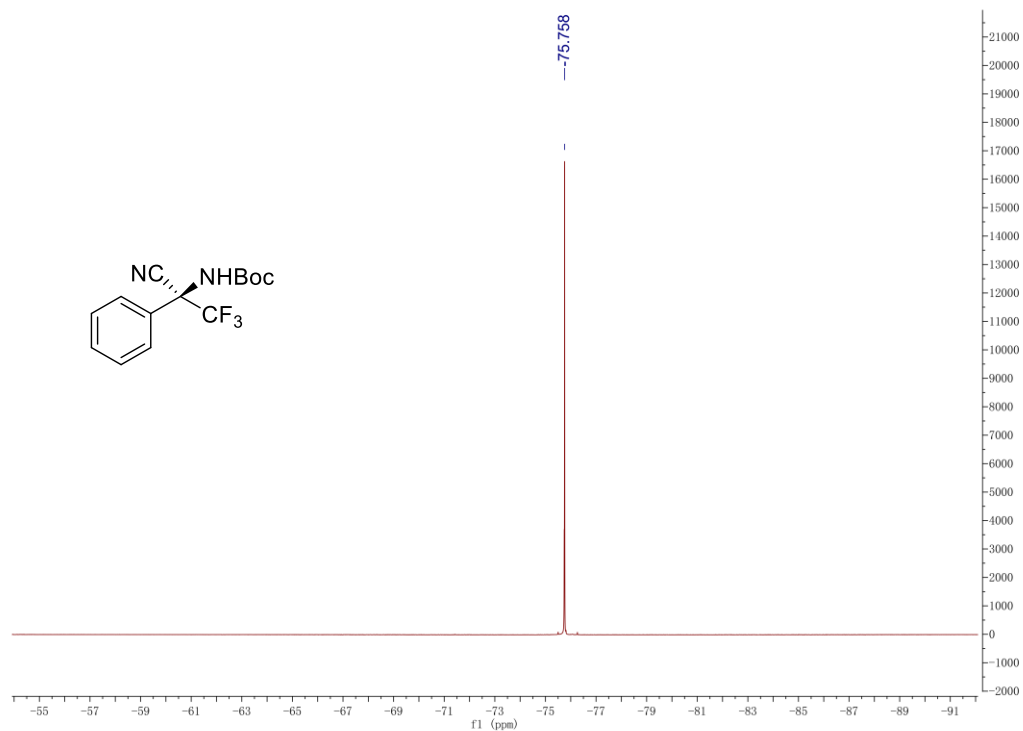


1 Det.A Ch1/254nm

PeakTable					
Peak#	Ret. Time	Area	Height	Area %	Height %
1	6.954	363581	16839	5.525	8.282
2	9.310	6217539	186492	94.475	91.718
Total		6581120	203331	100.000	100.000

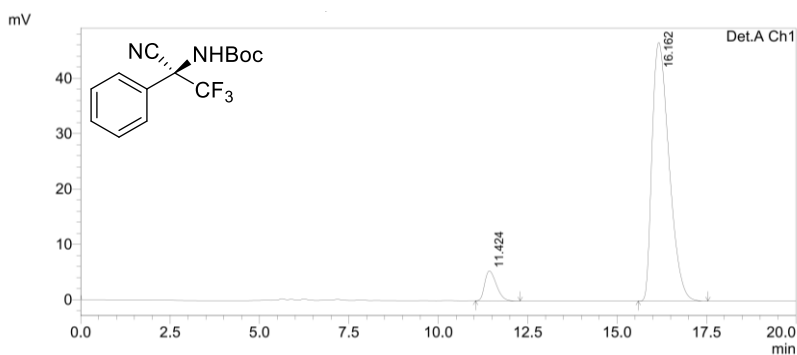
Copies of NMR and HPLC spectra for 3'a-3'm





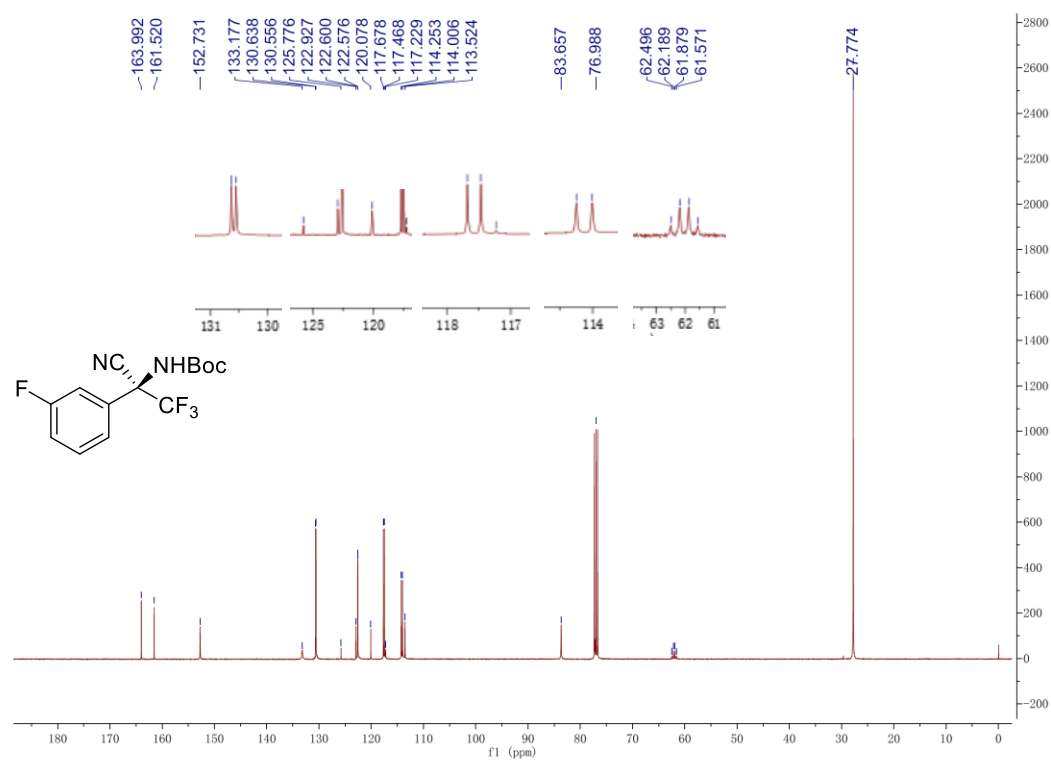
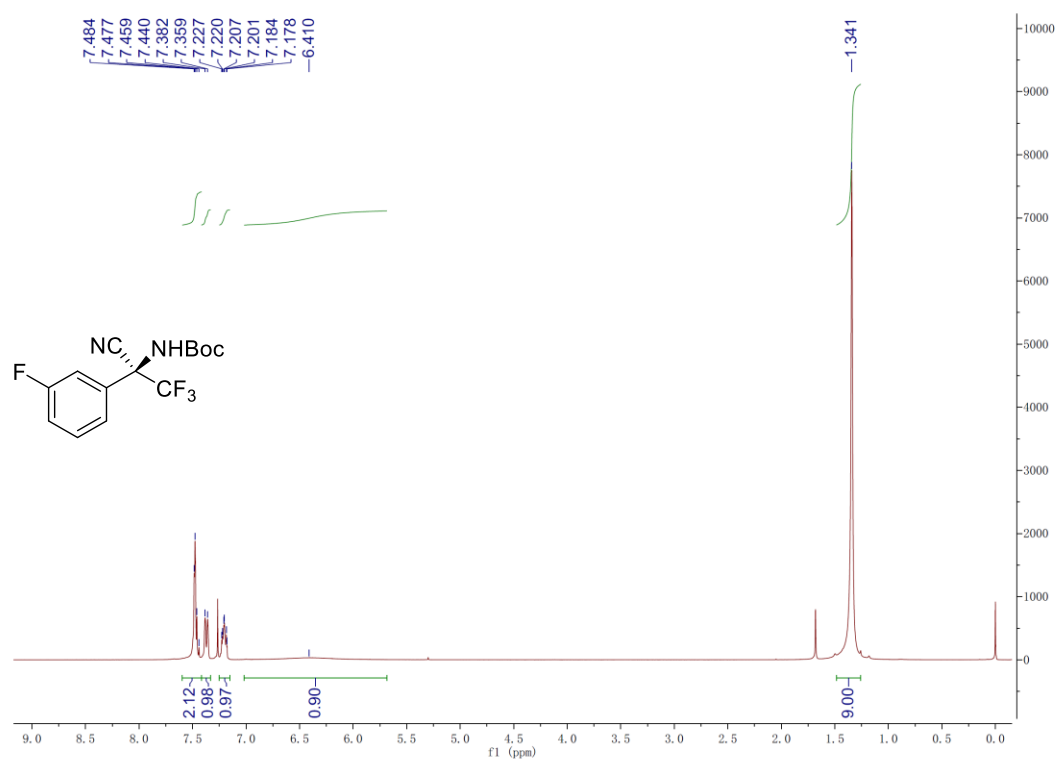
Detector A Ch1 254nm

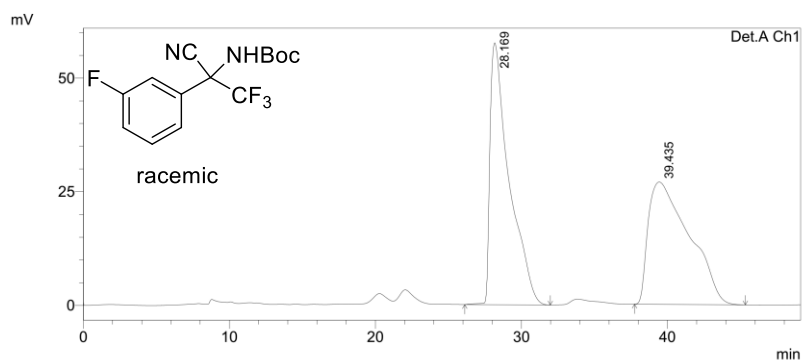
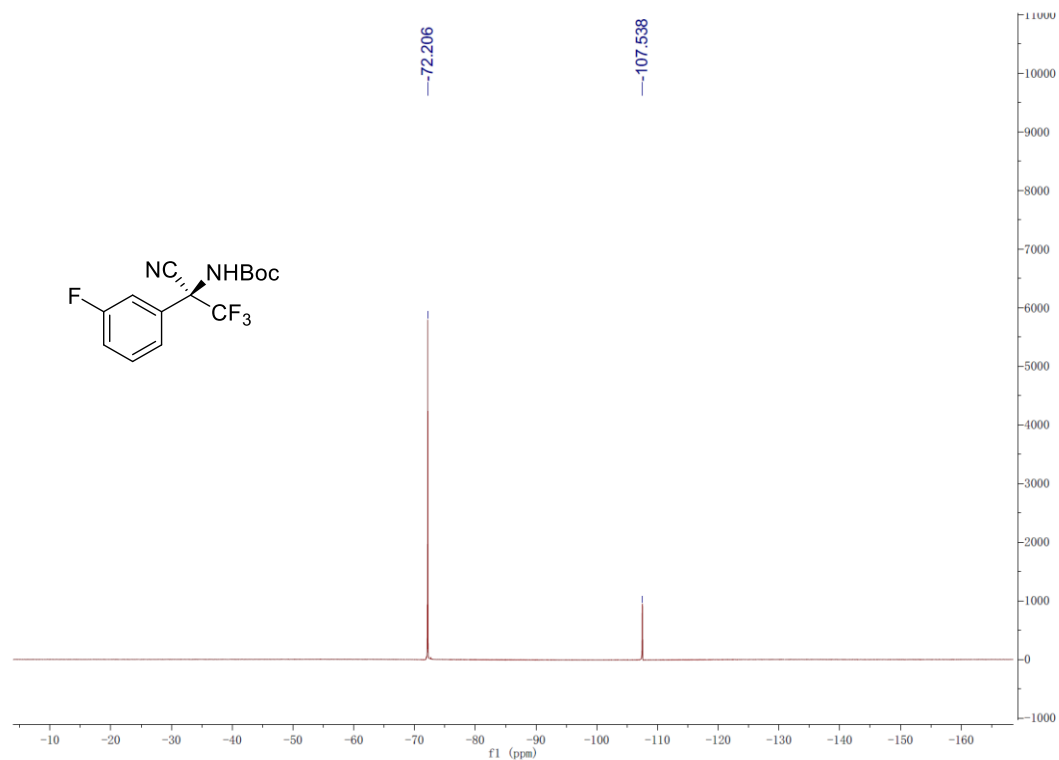
Peak#	Ret. Time	Area	Height	Area %	Height %
1	11.402	75836	3422	50.406	58.644
2	16.148	74616	2413	49.594	41.356
Total		150452	5835	100.000	100.000



Detector A Ch1 254nm

Peak#	Ret. Time	Area	Height	Area %	Height %
1	11.424	122320	5407	7.341	10.374
2	16.162	1543906	46719	92.659	89.626
Total		1666226	52127	100.000	100.000

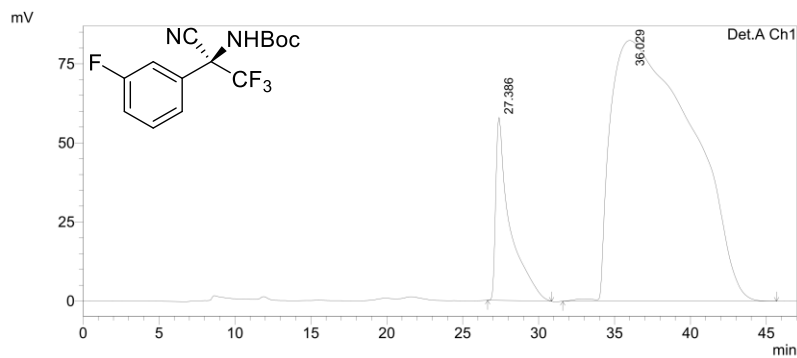




1 Det.A Ch1/254nm

PeakTable

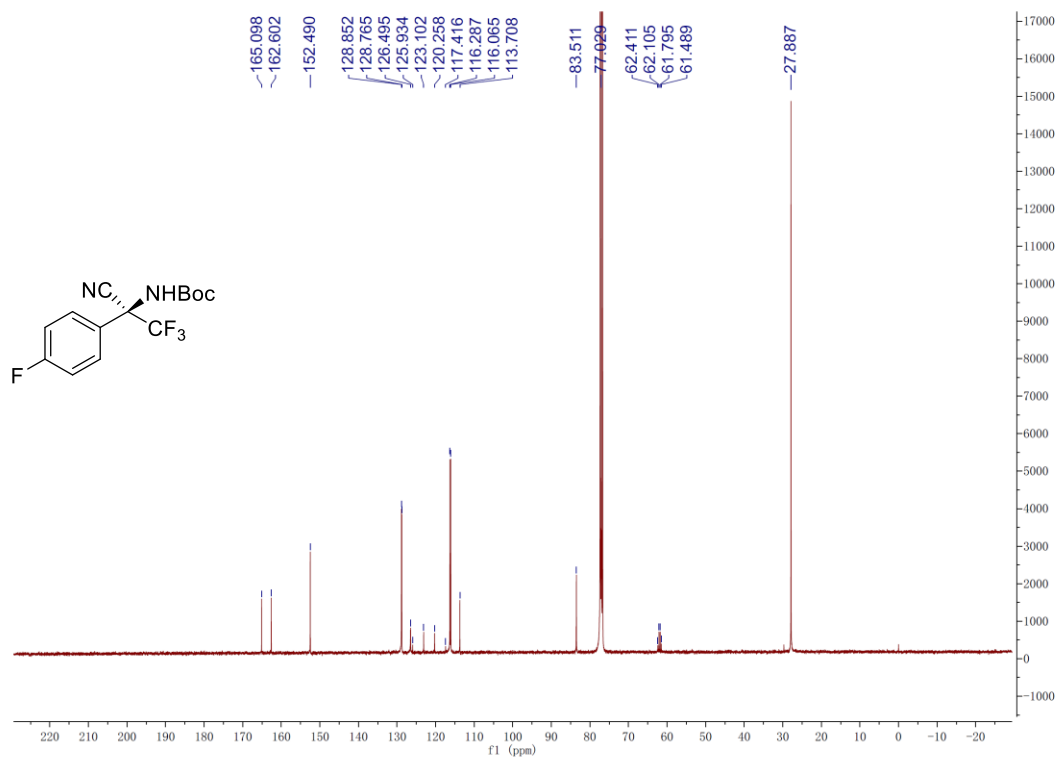
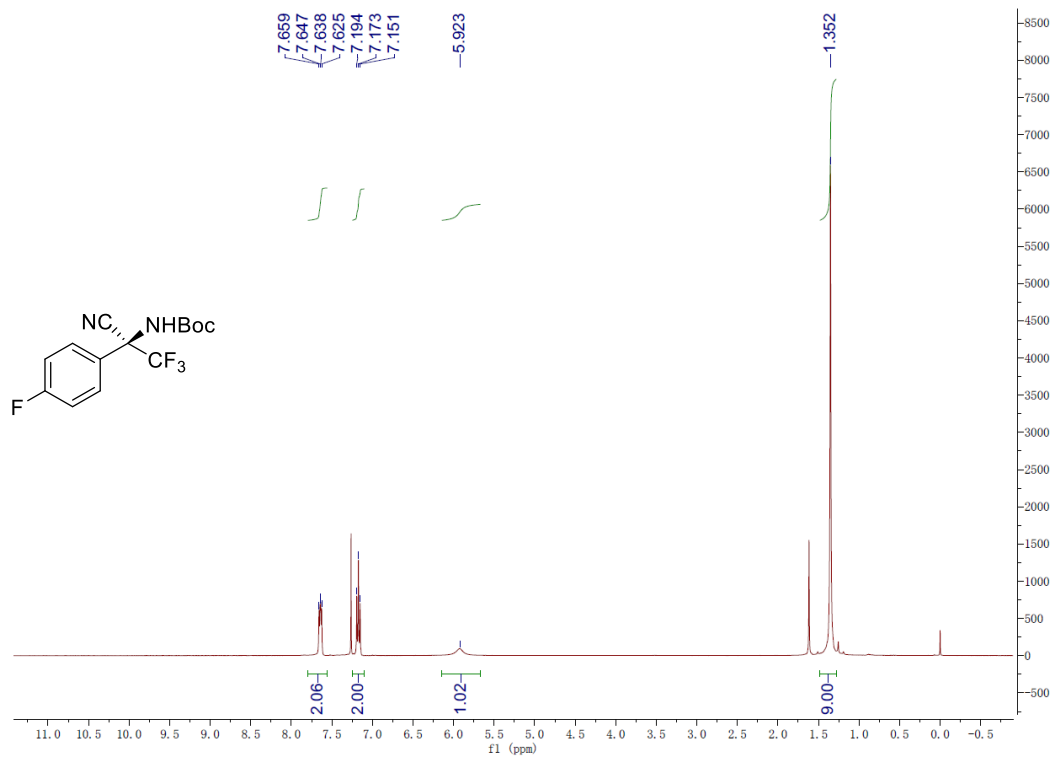
Peak#	Ret. Time	Area	Height	Area %	Height %
1	28.169	5256870	57601	49.964	68.169
2	39.435	5264340	26896	50.036	31.831
Total		10521210	84497	100.000	100.000

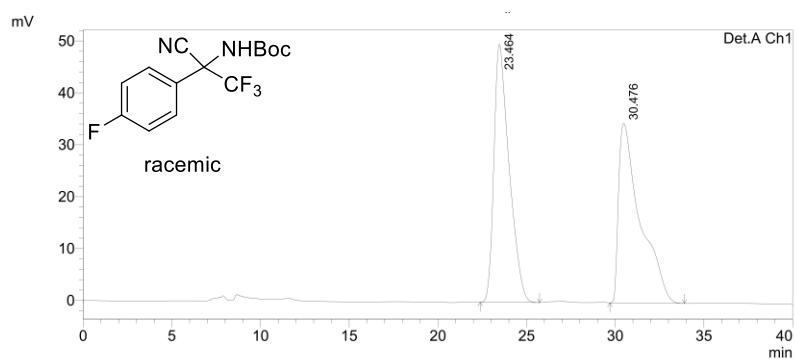
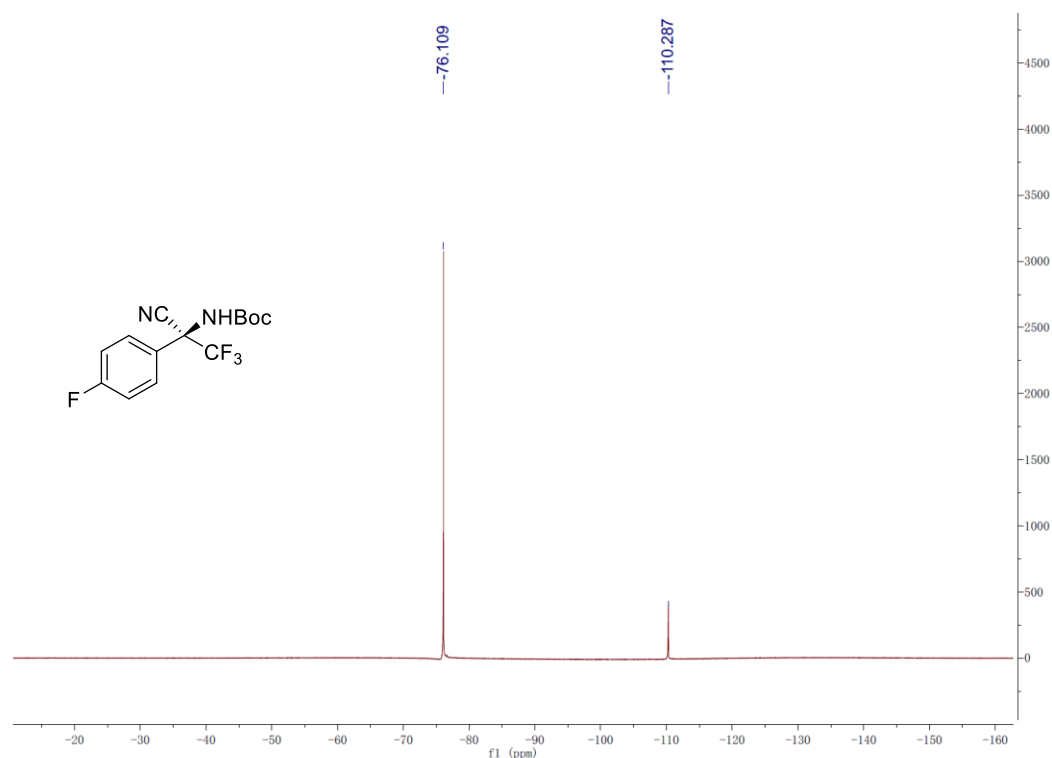


1 Det.A Ch1/254nm

PeakTable

Peak#	Ret. Time	Area	Height	Area %	Height %
1	27.386	3804323	57773	11.035	41.218
2	36.029	30671670	82392	88.965	58.782
Total		34475993	140165	100.000	100.000

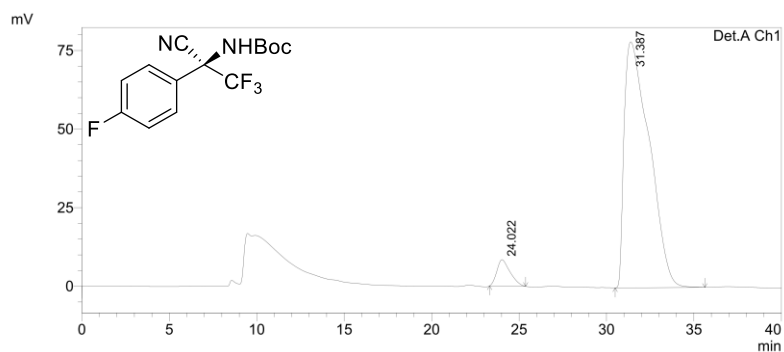




1 Det.A Ch1/254nm

PeakTable

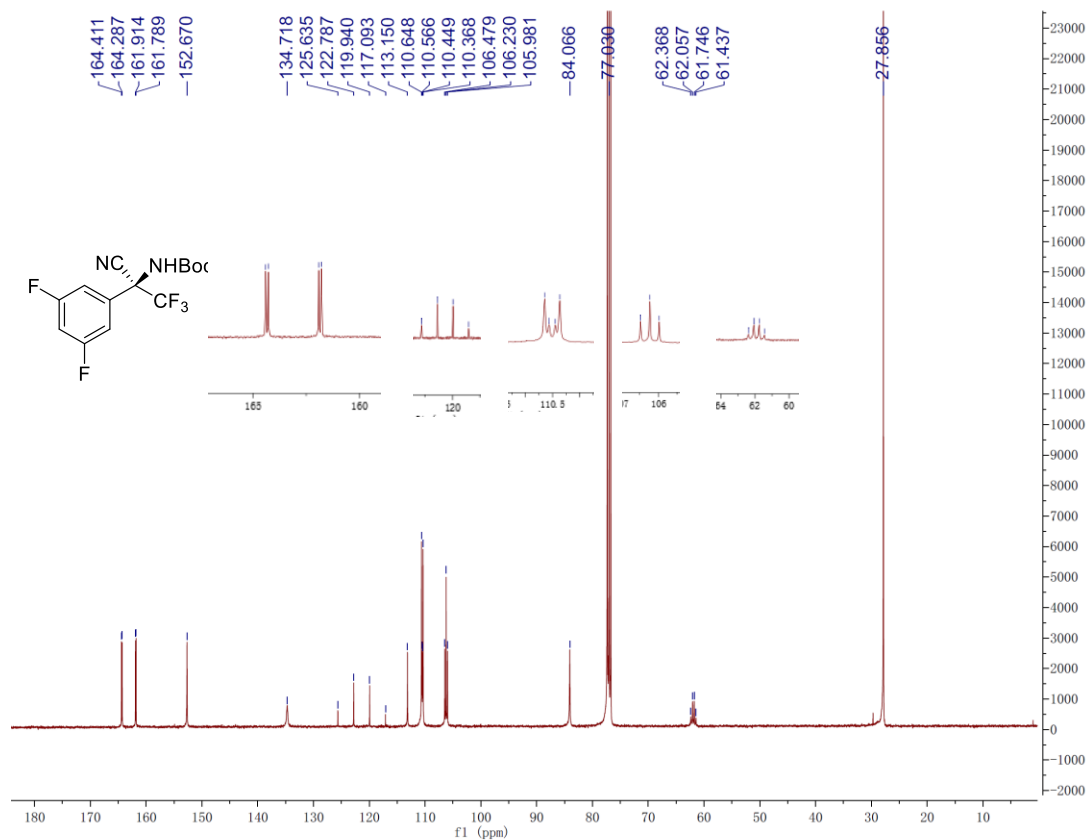
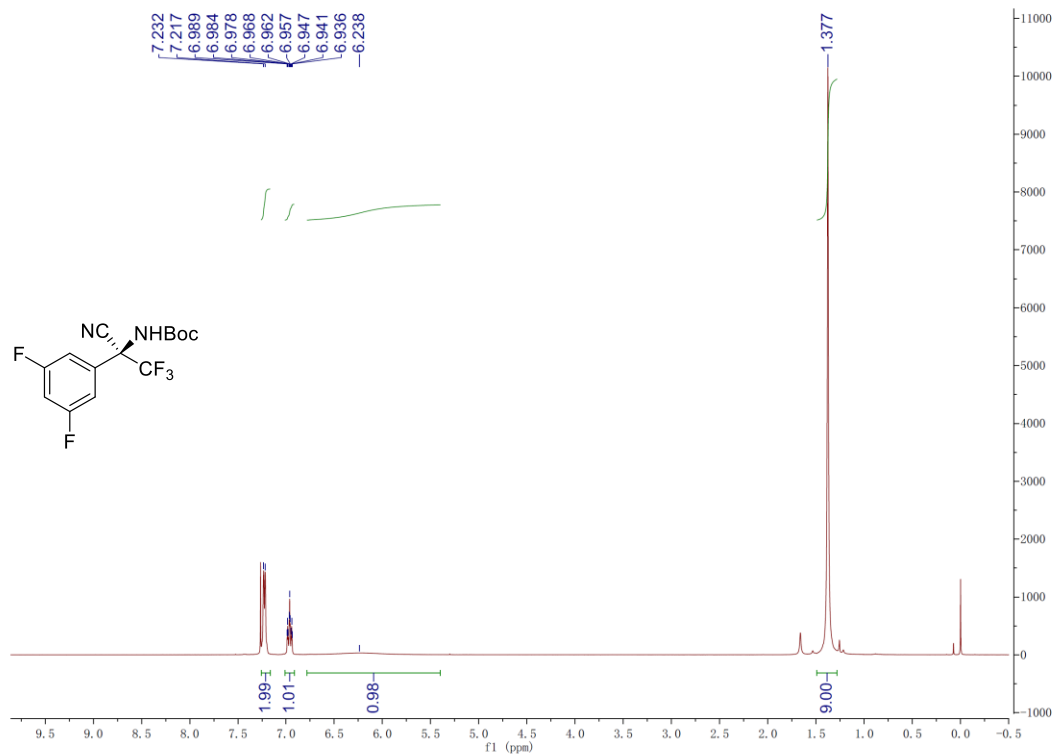
Peak#	Ret. Time	Area	Height	Area %	Height %
1	23.464	2969201	49678	50.259	58.953
2	30.476	2938619	34589	49.741	41.047
Total		5907821	84267	100.000	100.000

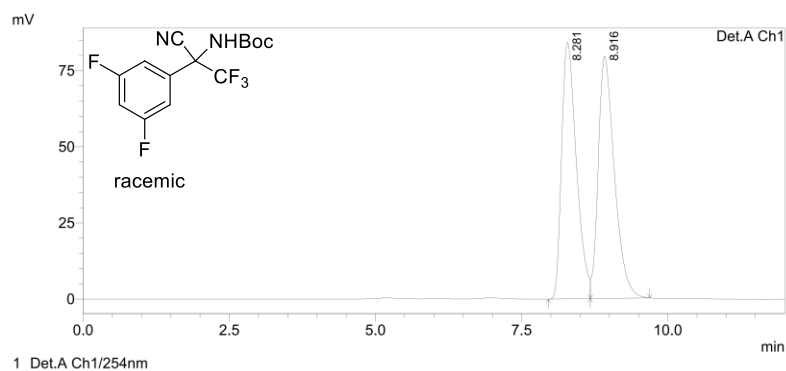
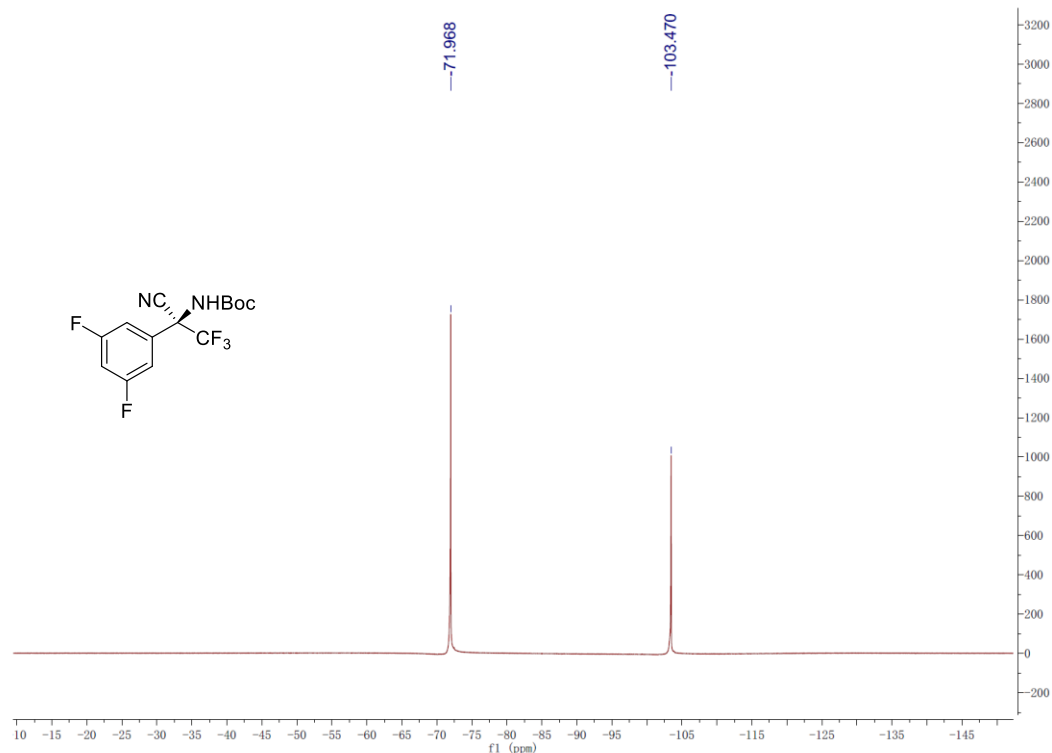


1 Det.A Ch1/254nm

PeakTable

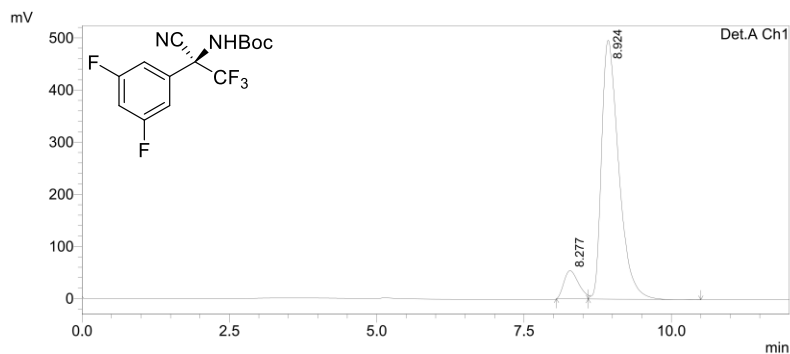
Peak#	Ret. Time	Area	Height	Area %	Height %
1	24.022	440700	8385	5.335	9.678
2	31.387	7820426	78252	94.665	90.322
Total		8261127	86636	100.000	100.000





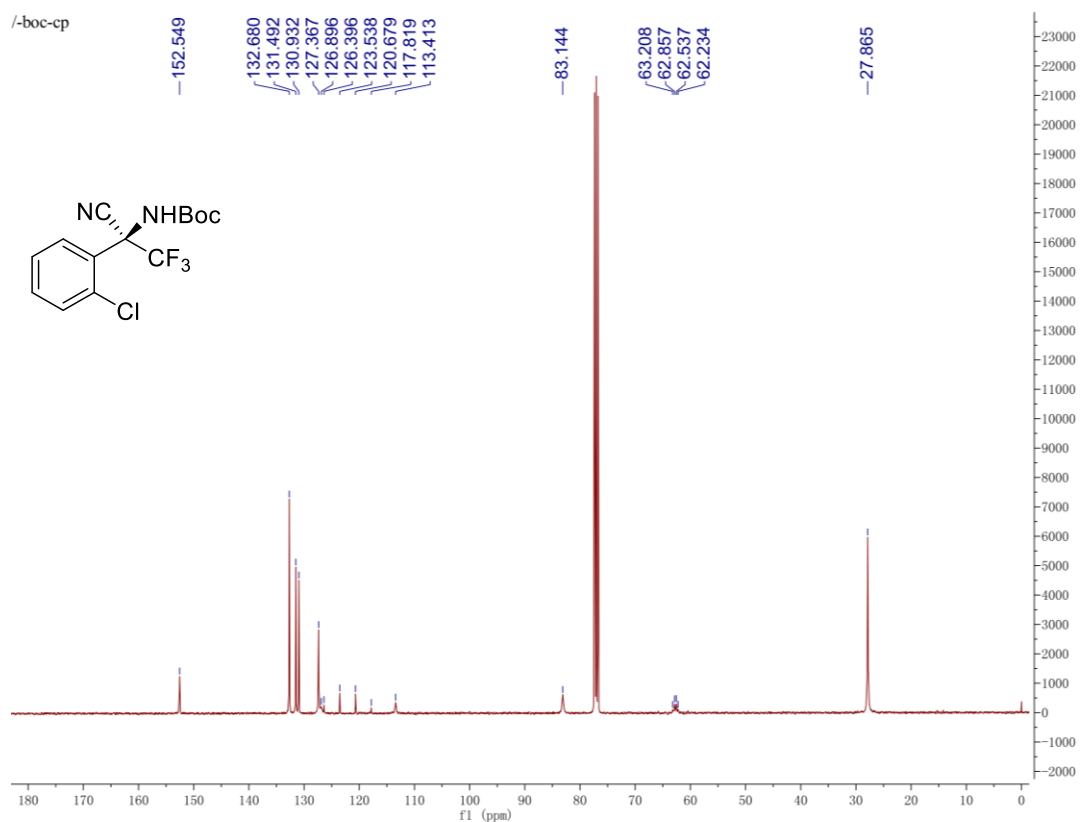
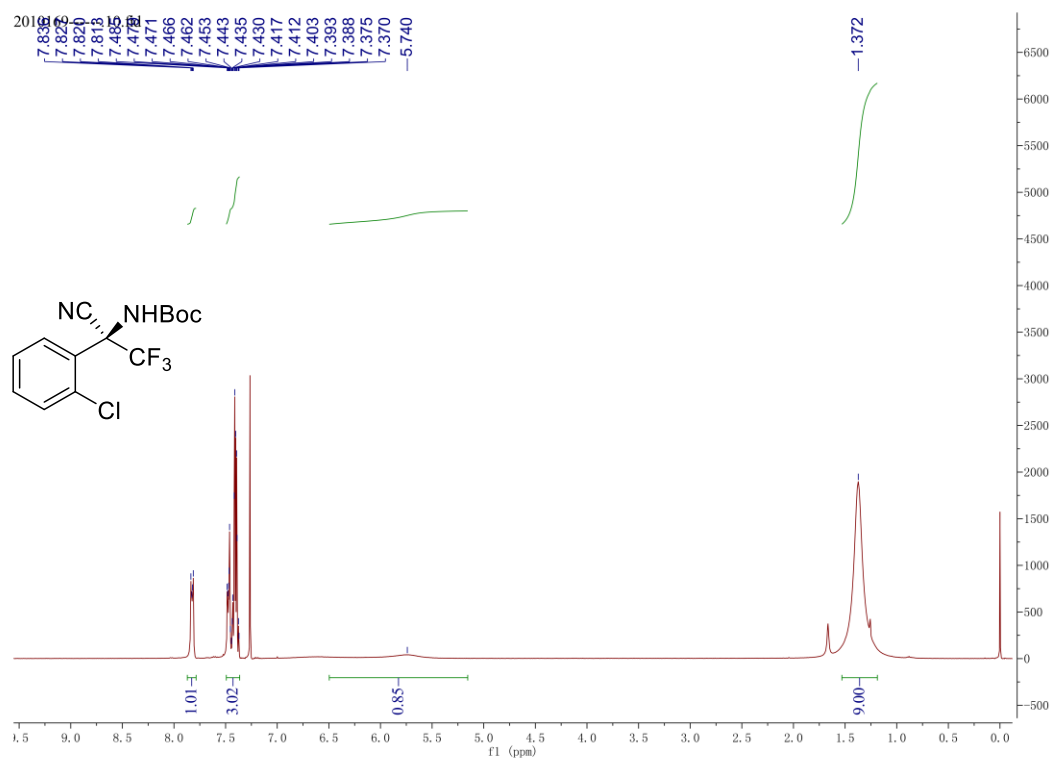
1 Det.A Ch1/254nm

PeakTable					
Peak#	Ret. Time	Area	Height	Area %	Height %
1	8.281	1479171	84252	49.110	51.472
2	8.916	1532811	79433	50.890	48.528
Total		3011982	163685	100.000	100.000

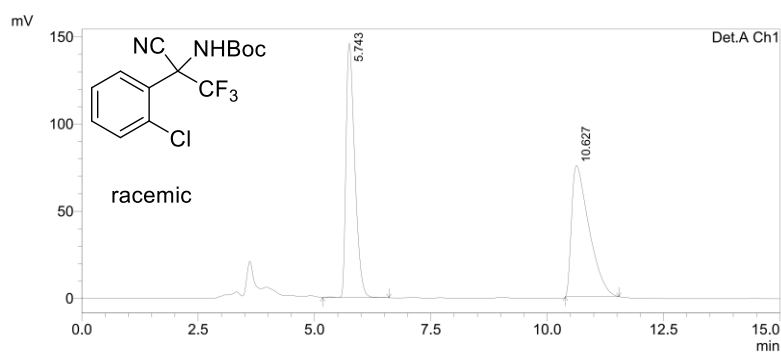
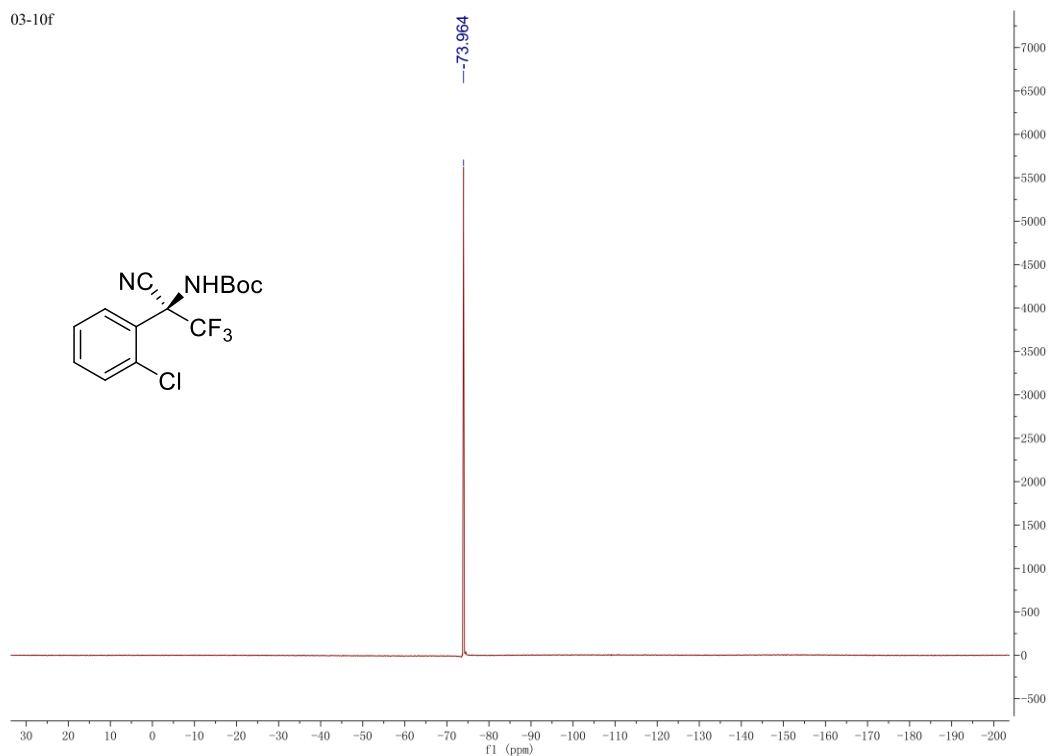


1 Det.A Ch1/254nm

PeakTable					
Peak#	Ret. Time	Area	Height	Area %	Height %
1	8.277	933576	54205	8.477	9.845
2	8.924	10078868	496386	91.523	90.155
Total		11012444	550591	100.000	100.000

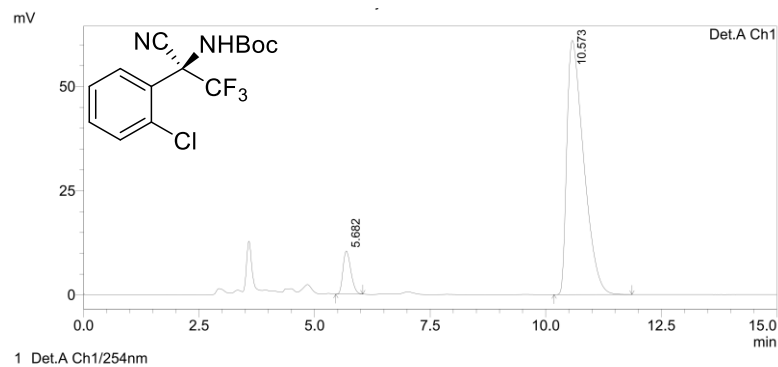


03-10f



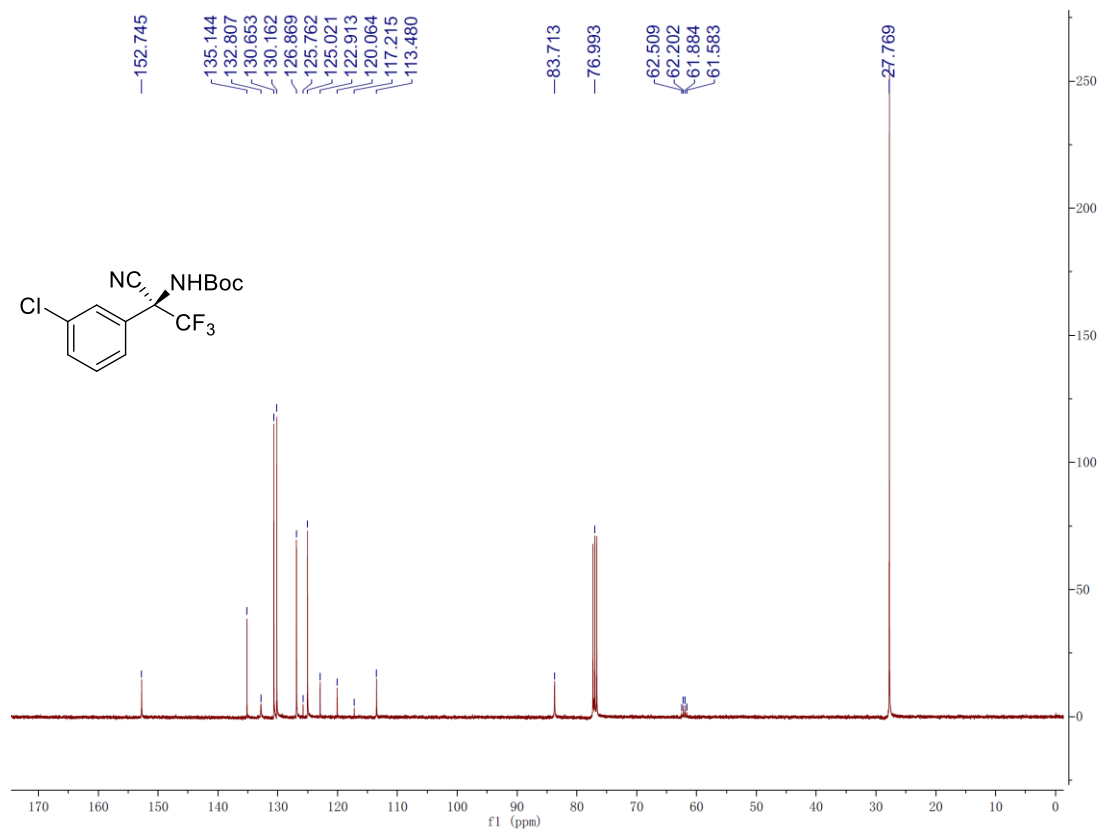
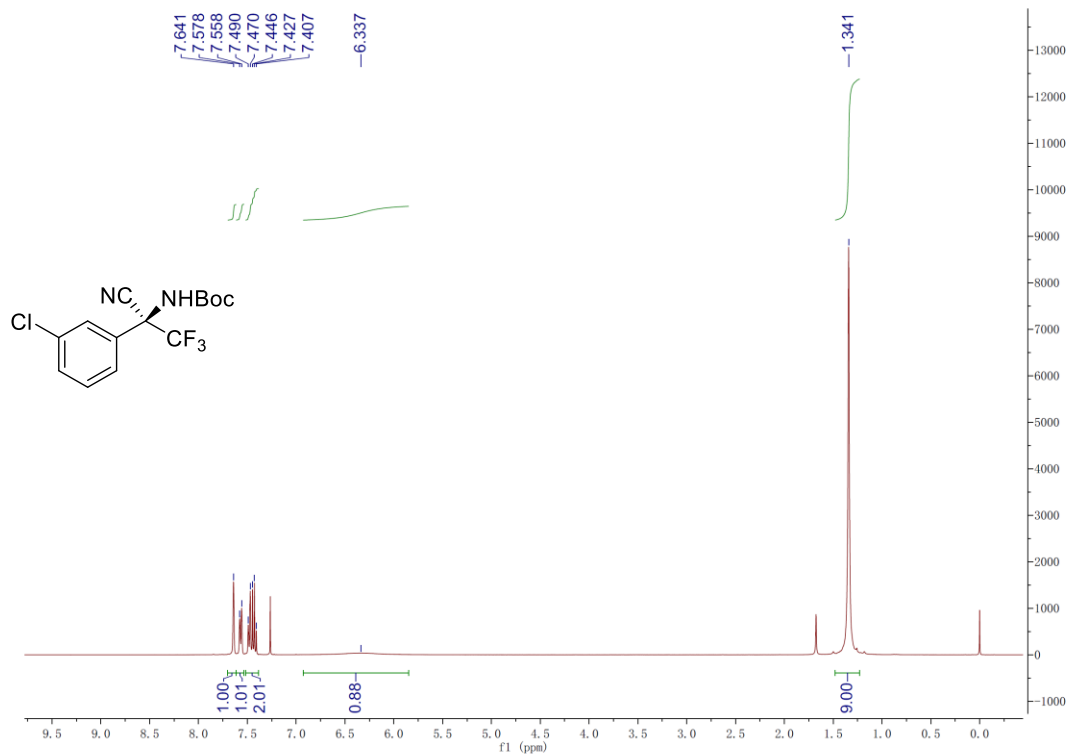
PeakTable

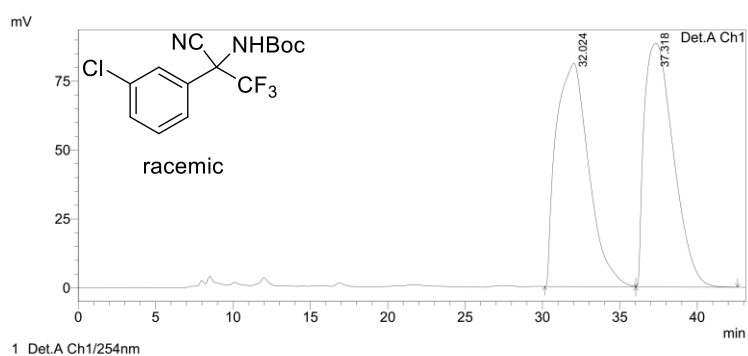
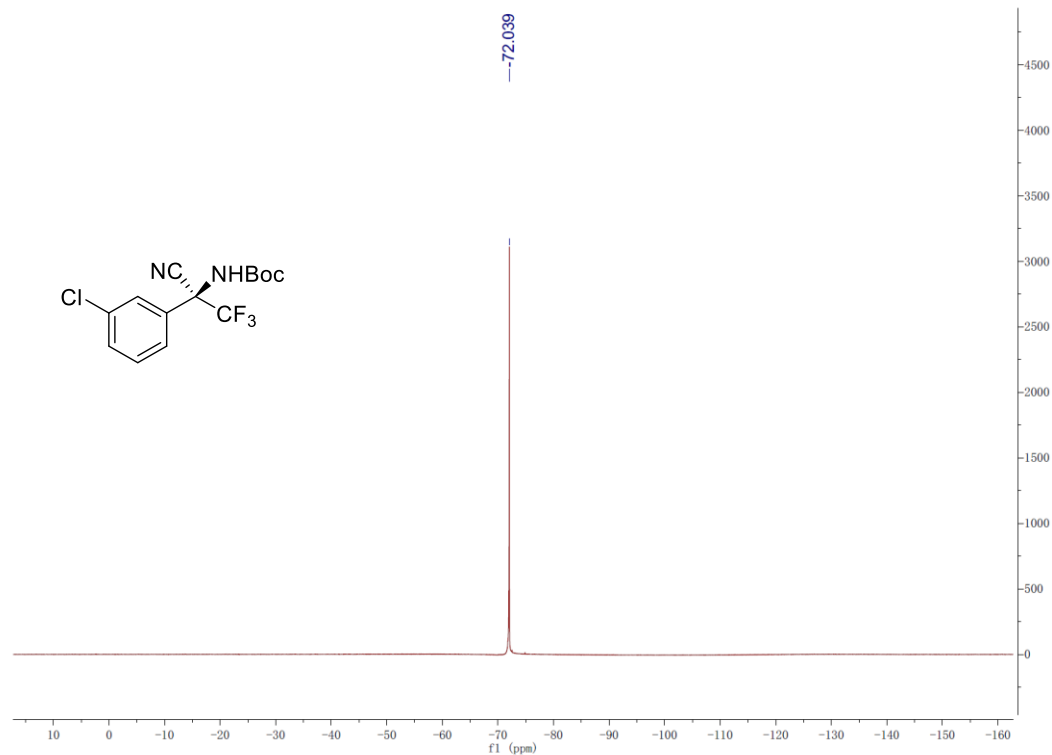
Peak#	Ret. Time	Area	Height	Area %	Height %
1	5.743	1912930	145744	49.325	65.936
2	10.627	1965297	75295	50.675	34.064
Total		3878227	221039	100.000	100.000



PeakTable

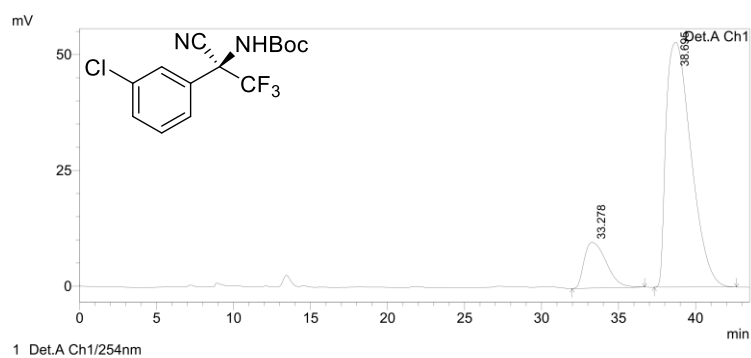
Peak#	Ret. Time	Area	Height	Area %	Height %
1	5.682	122731	10354	7.367	14.513
2	10.573	1543324	60988	92.633	85.487
Total		1666055	71342	100.000	100.000





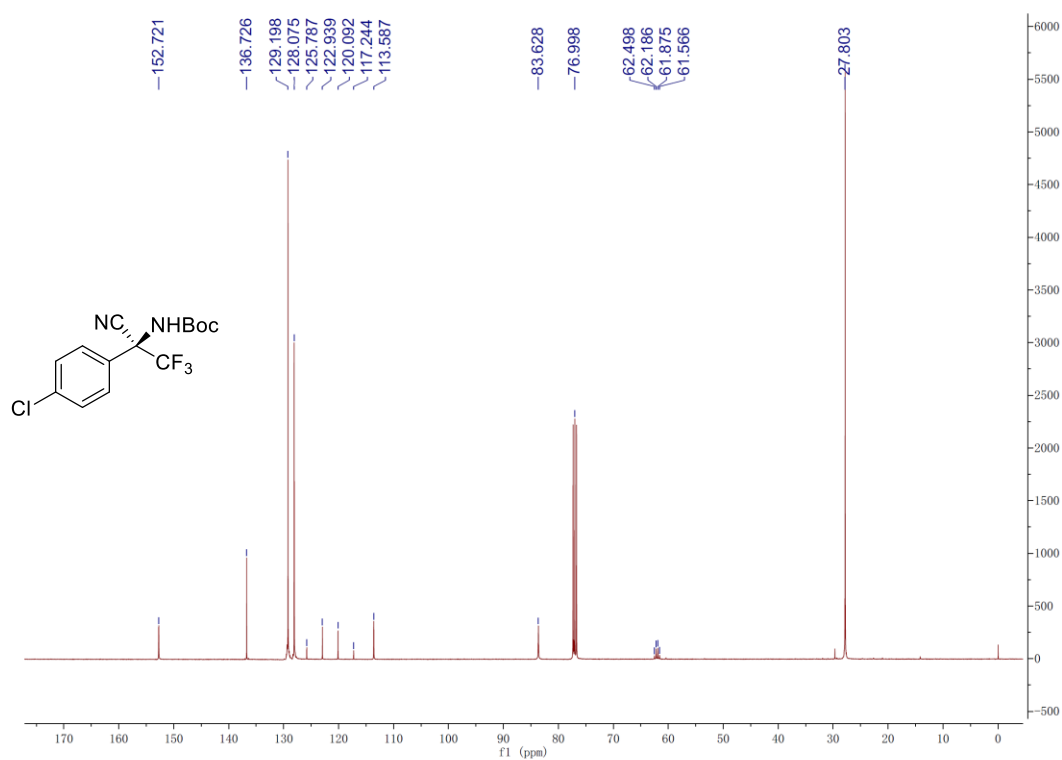
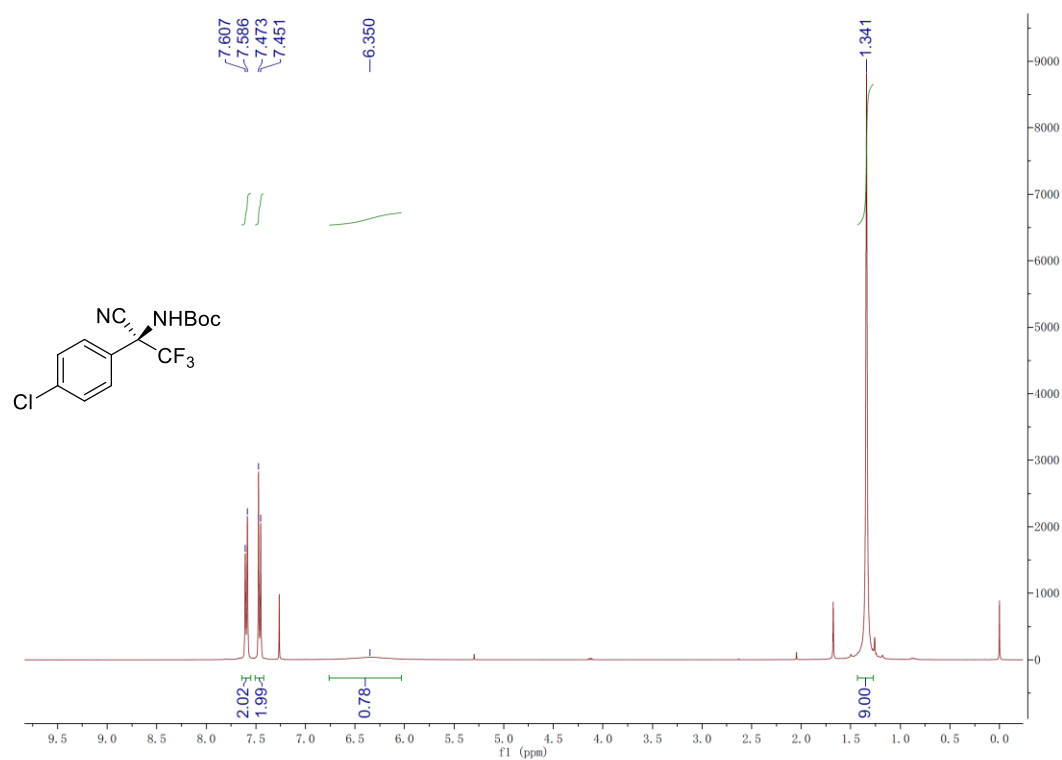
PeakTable

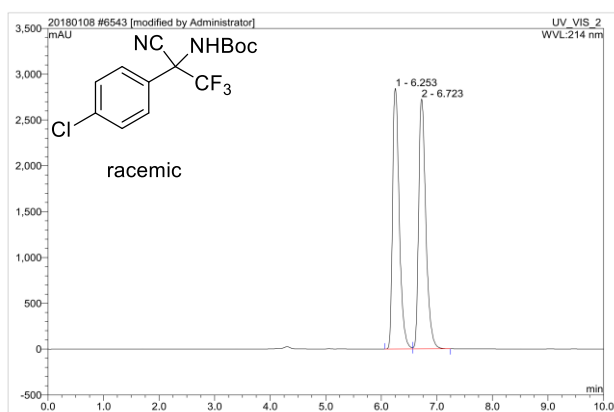
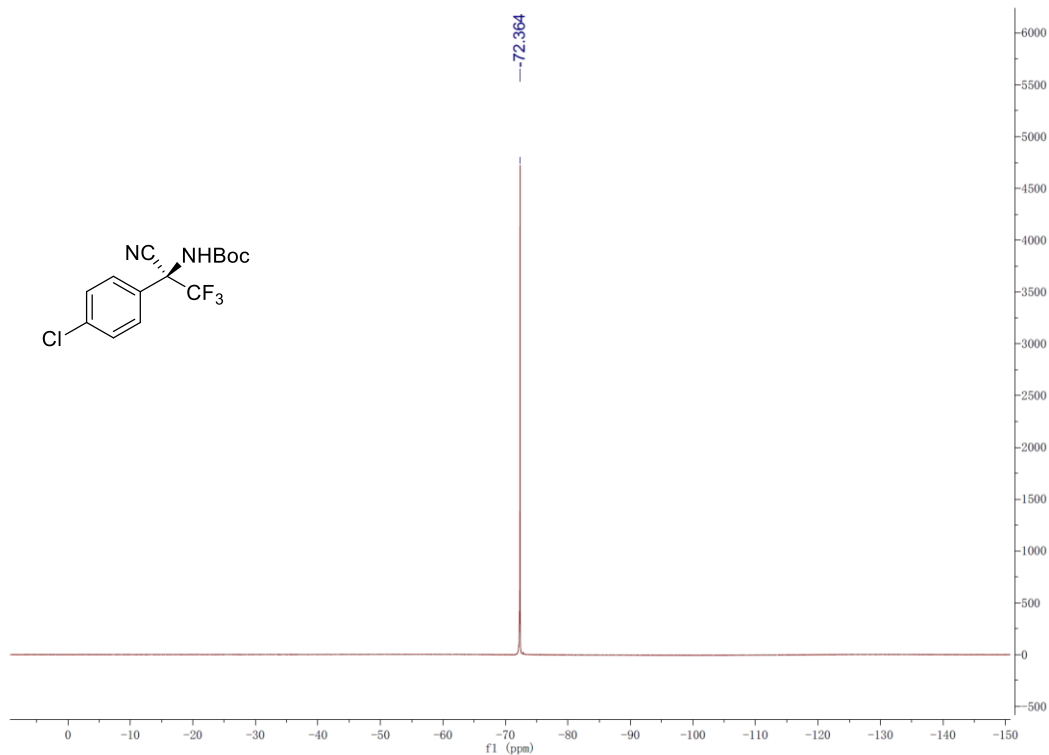
Peak#	Ret. Time	Area	Height	Area %	Height %
1	32.024	11848979	81108	50.542	47.845
2	37.318	11594683	88414	49.458	52.155
Total		23443662	169522	100.000	100.000



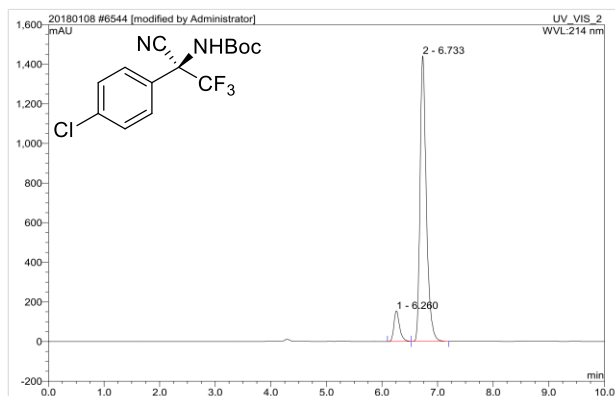
PeakTable

Peak#	Ret. Time	Area	Height	Area %	Height %
1	33.278	984057	9872	14.175	15.738
2	38.695	5958203	52854	85.825	84.262
Total		6942259	62726	100.000	100.000

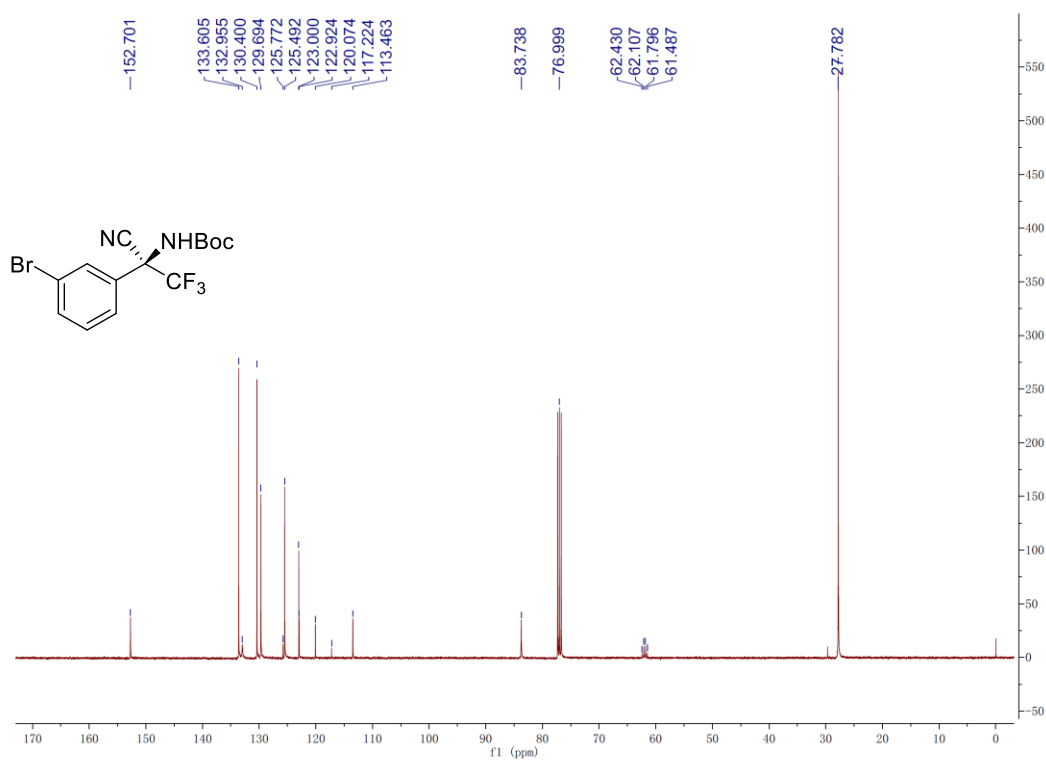
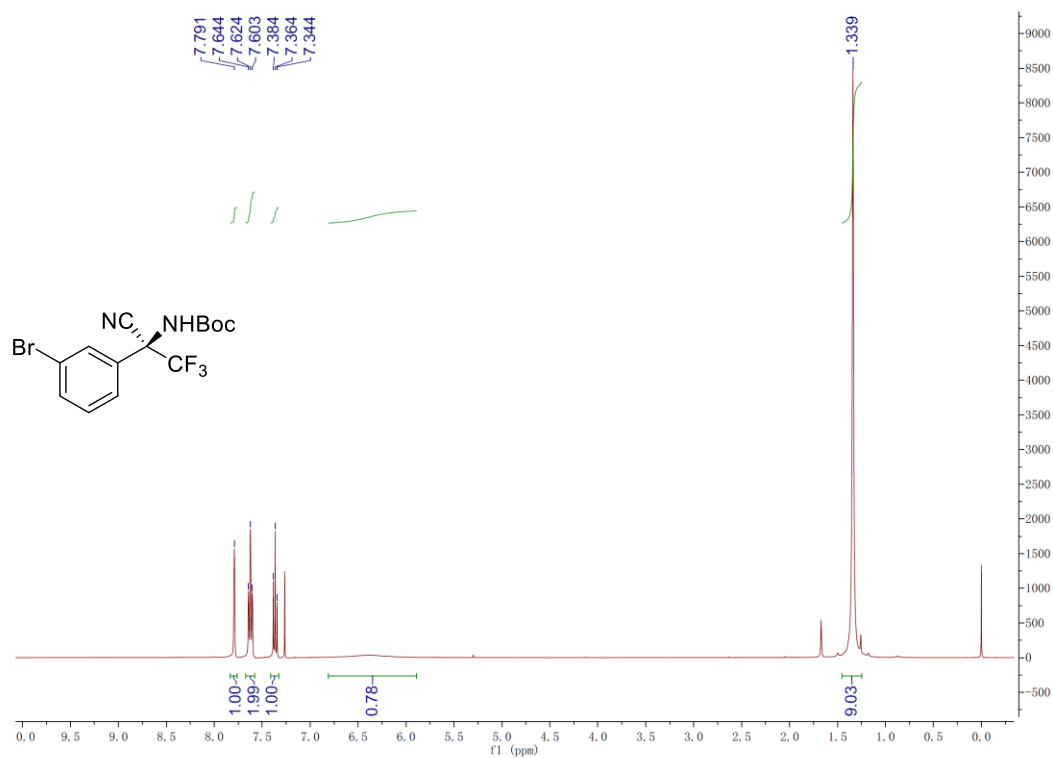


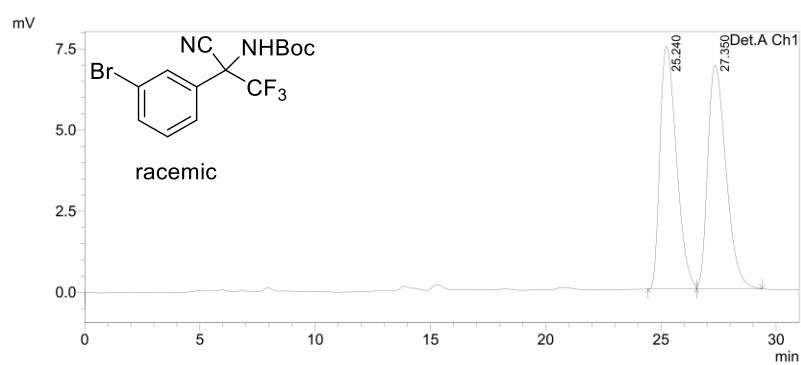
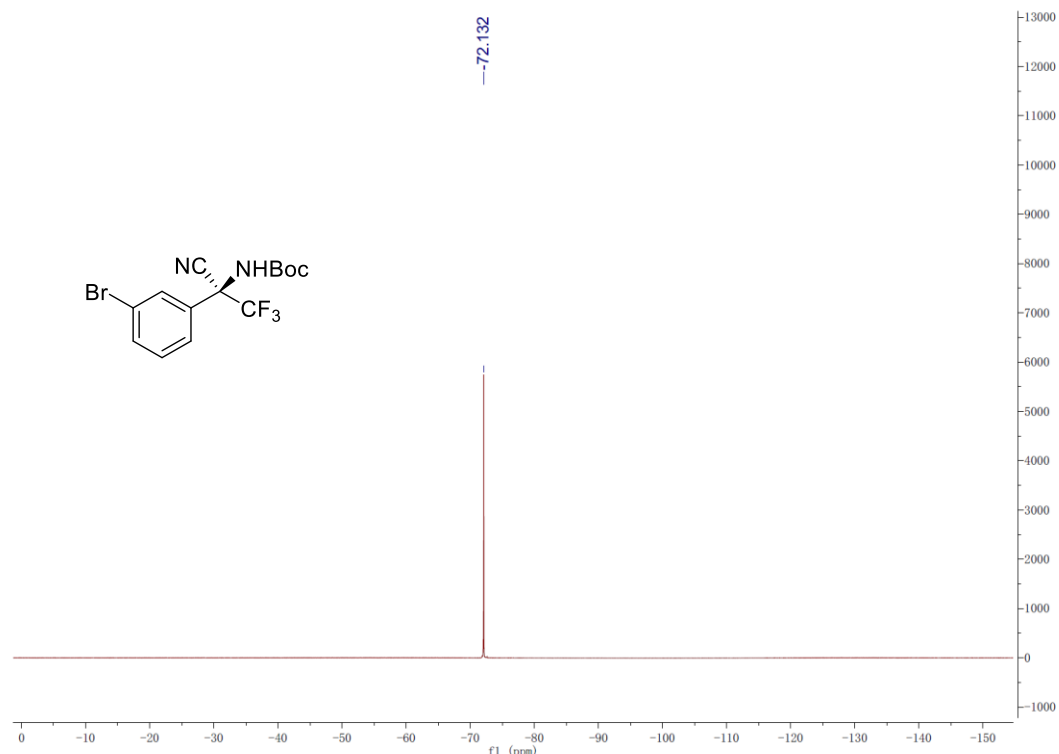


No.	Ret.Time min	Peak Name	Height mAU	Area mAU*min	Rel.Area %	Amount	Type
1	6.25	n.a.	2844.704	397.448	49.31	n.a.	BM
2	6.72	n.a.	2726.574	408.540	50.69	n.a.	MB
Total:			5571.278	805.988	100.00	0.000	



No.	Ret.Time min	Peak Name	Height mAU	Area mAU*min	Rel.Area %	Amount	Type
1	6.26	n.a.	154.471	18.074	8.71	n.a.	BMb*
2	6.73	n.a.	1441.387	189.329	91.29	n.a.	bMB*
Total:			1595.858	207.403	100.00	0.000	

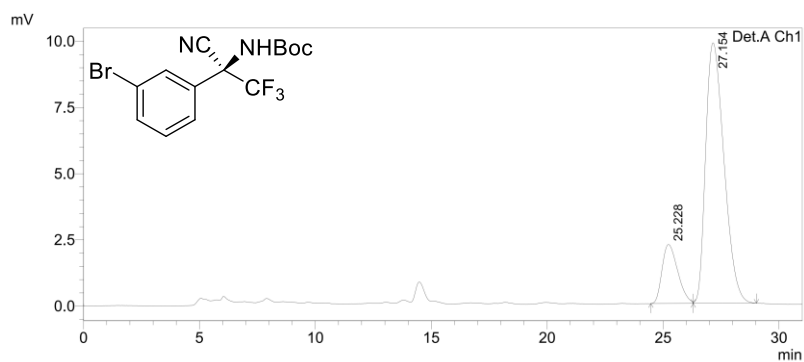




1 Det.A Ch1/254nm

PeakTable

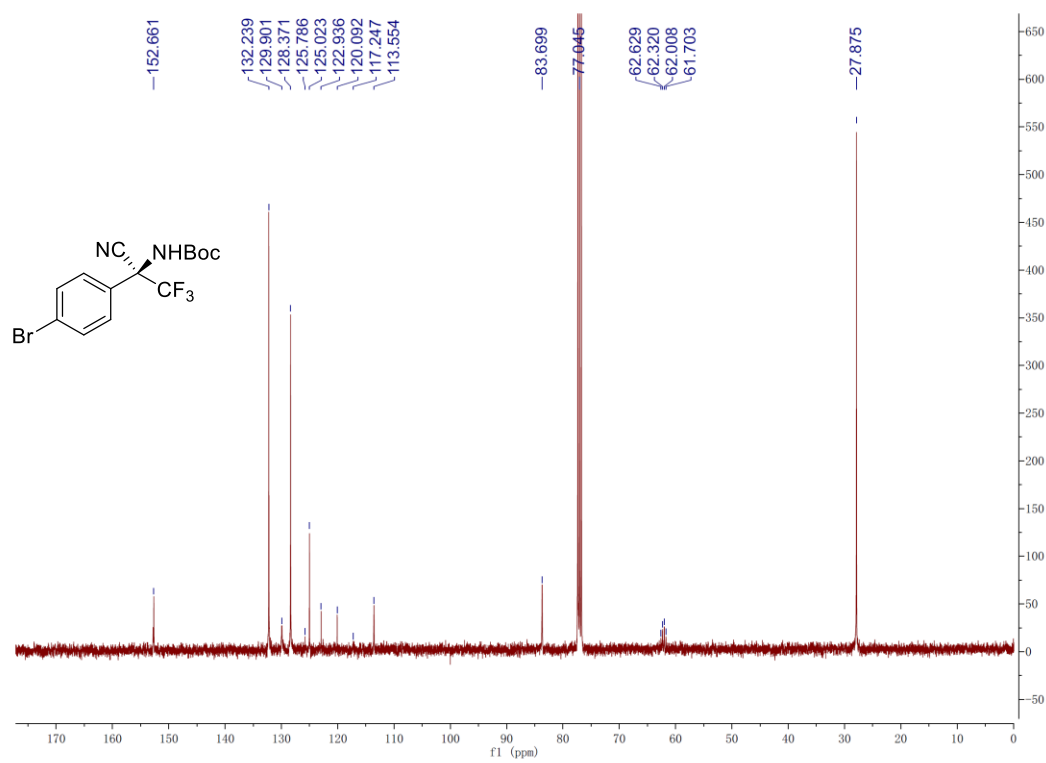
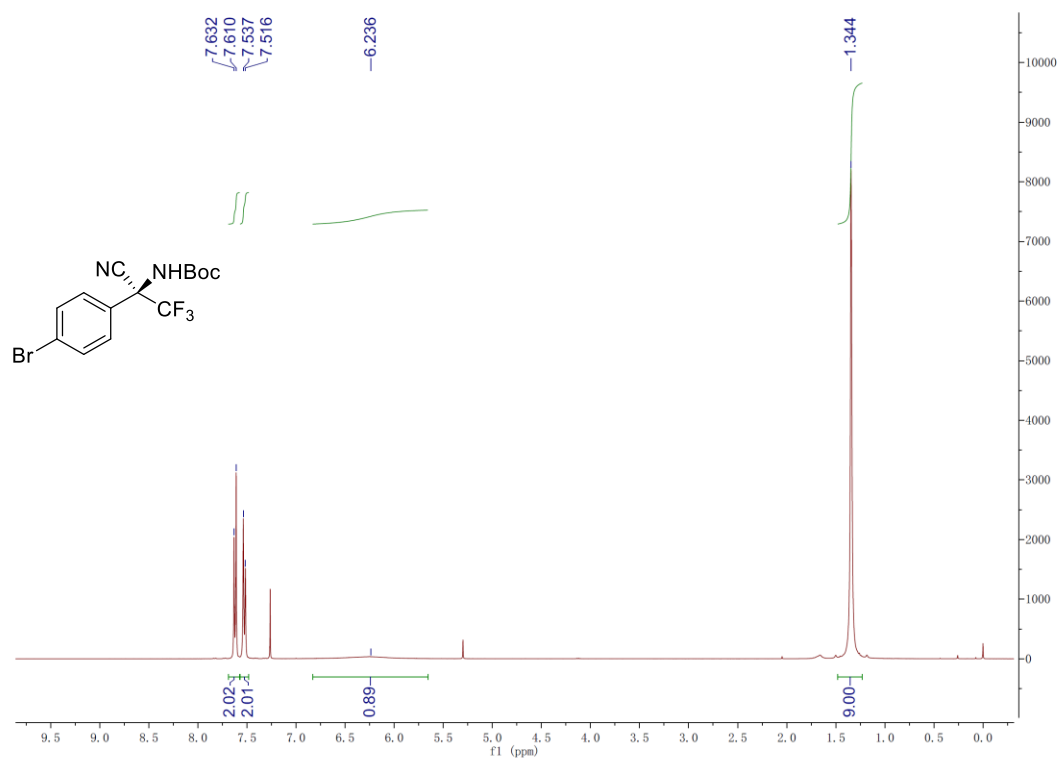
Peak#	Ret. Time	Area	Height	Area %	Height %
1	25.240	371787	7482	49.453	52.048
2	27.350	380018	6893	50.547	47.952
Total		751805	14376	100.000	100.000

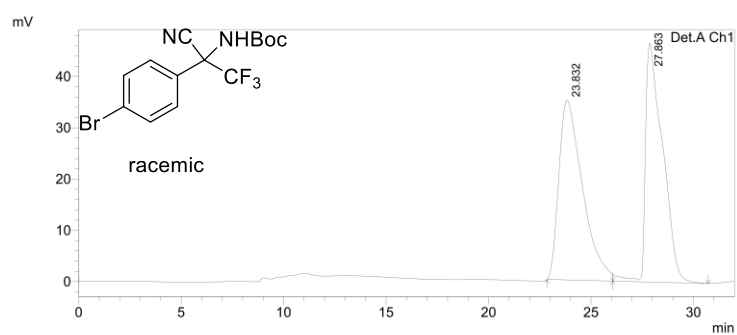
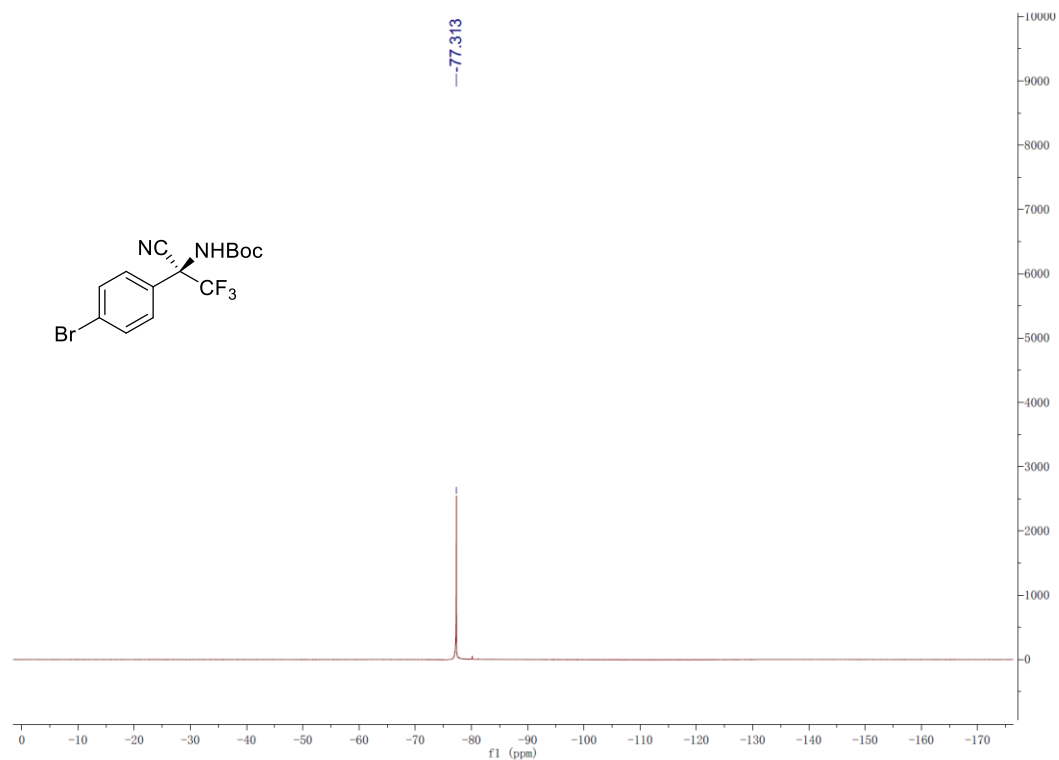


1 Det.A Ch1/254nm

PeakTable

Peak#	Ret. Time	Area	Height	Area %	Height %
1	25.228	104230	2226	16.113	18.450
2	27.154	542641	9839	83.887	81.550
Total		646870	12064	100.000	100.000

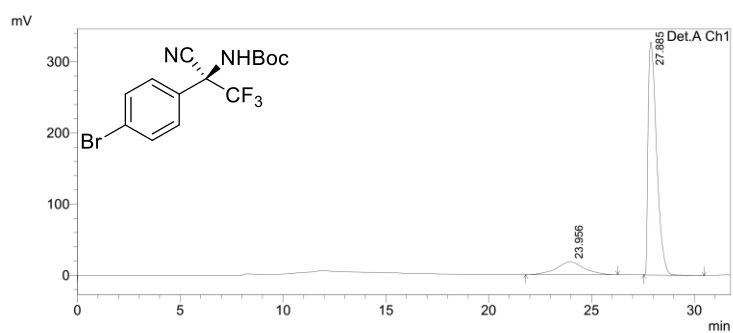




1 Det.A Ch1/254nm

PeakTable

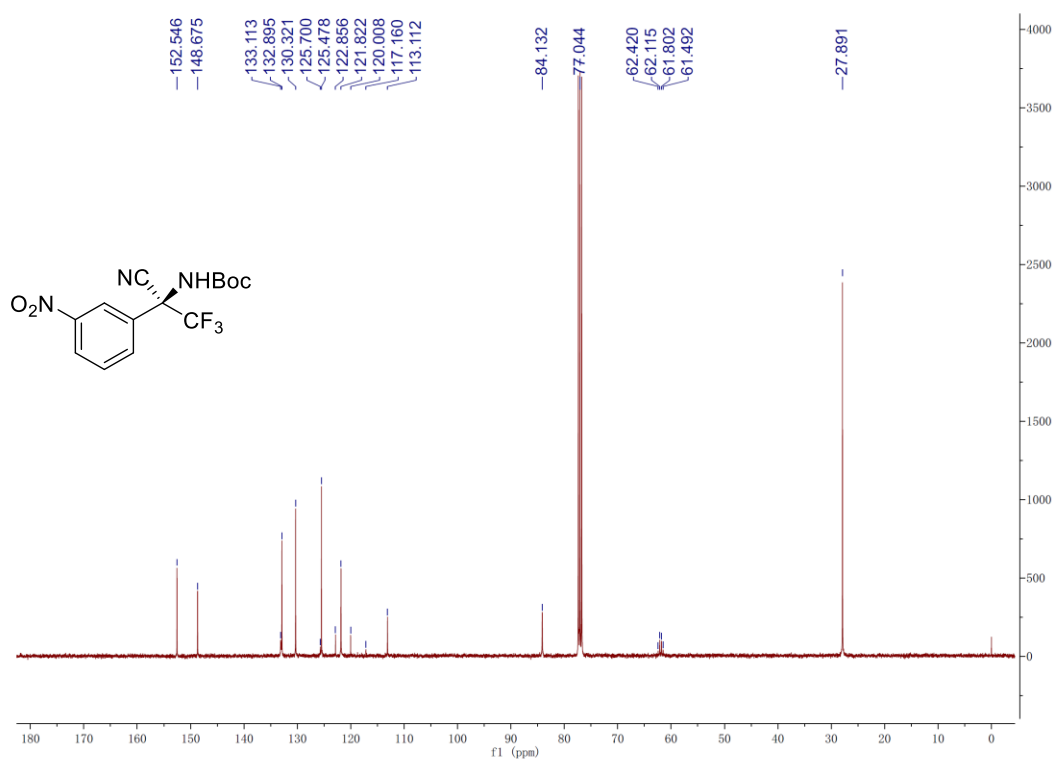
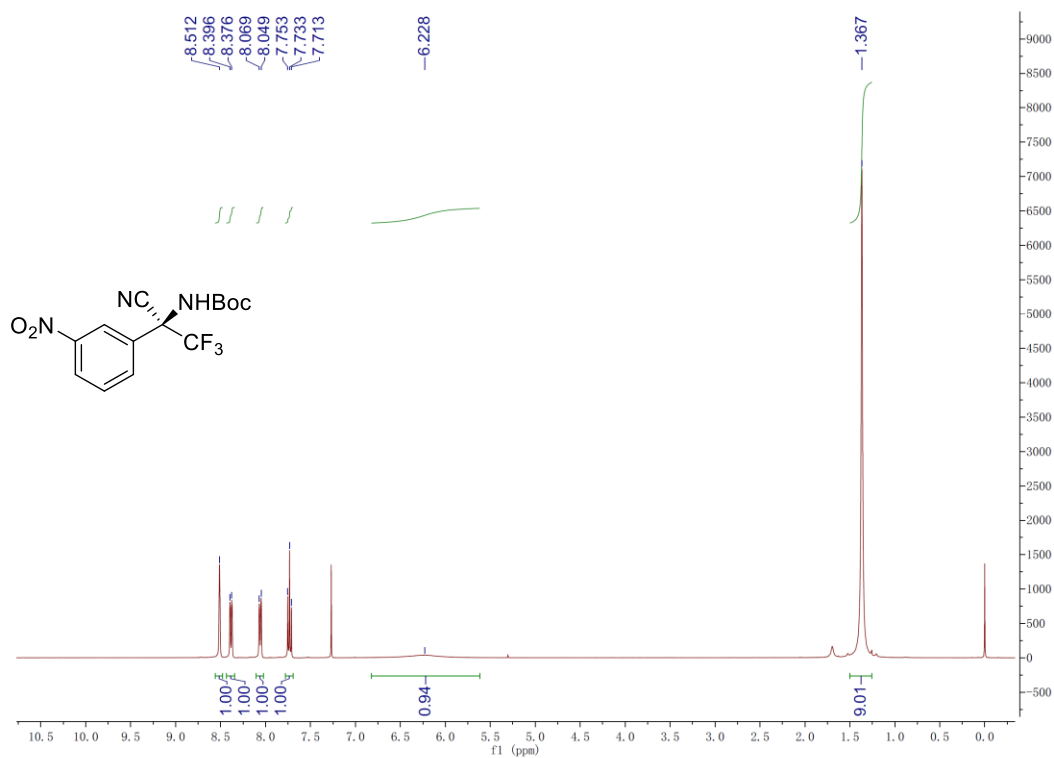
Peak#	Ret. Time	Area	Height	Area %	Height %
1	23.832	2810148	35043	50.614	42.923
2	27.863	2741990	46599	49.386	57.077
Total		5552138	81641	100.000	100.000

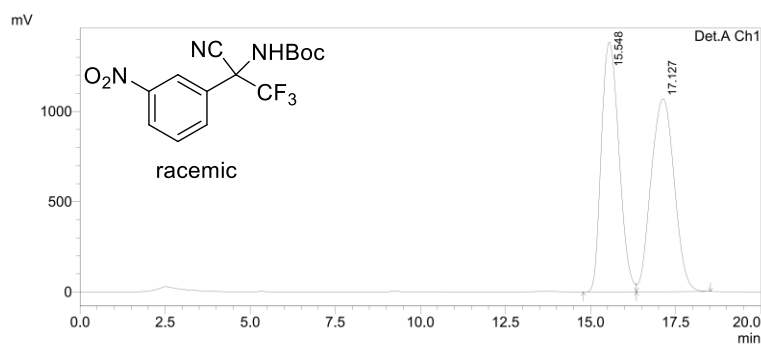
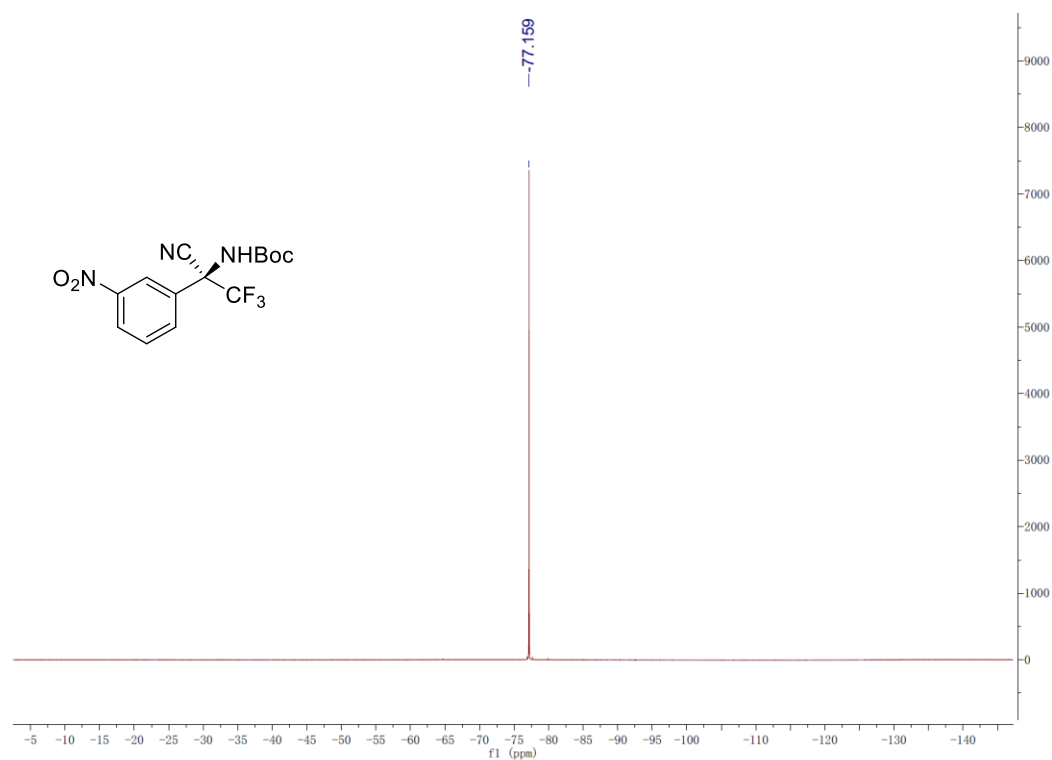


1 Det.A Ch1/254nm

PeakTable

Peak#	Ret. Time	Area	Height	Area %	Height %
1	23.956	1682653	18113	15.091	5.239
2	27.885	9467668	327596	84.909	94.761
Total		11150321	345709	100.000	100.000

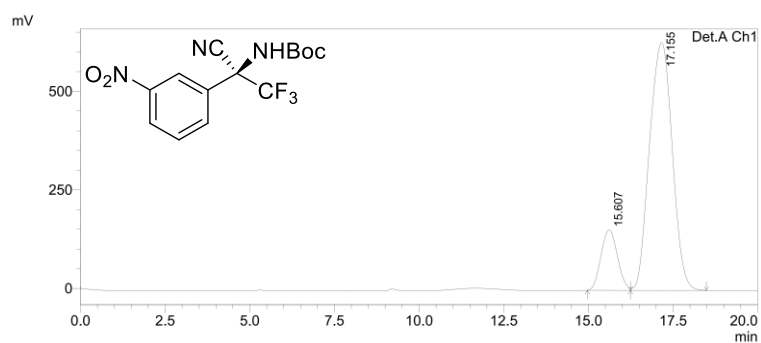




1 Det.A Ch1/254nm

PeakTable

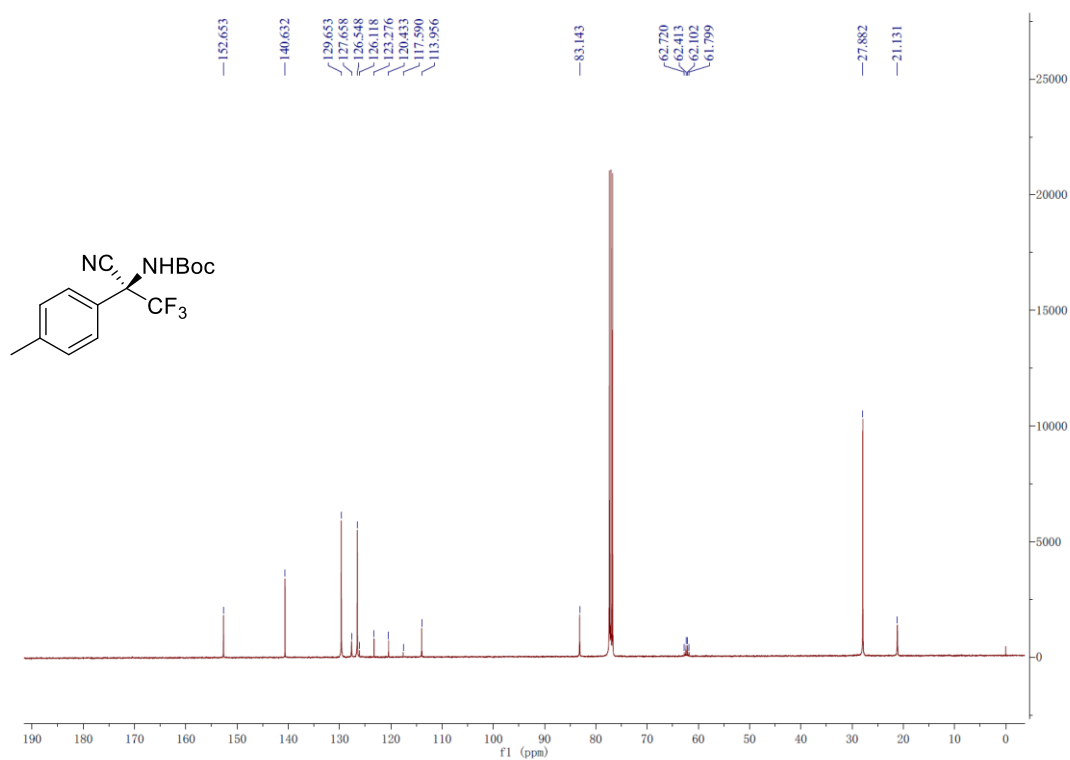
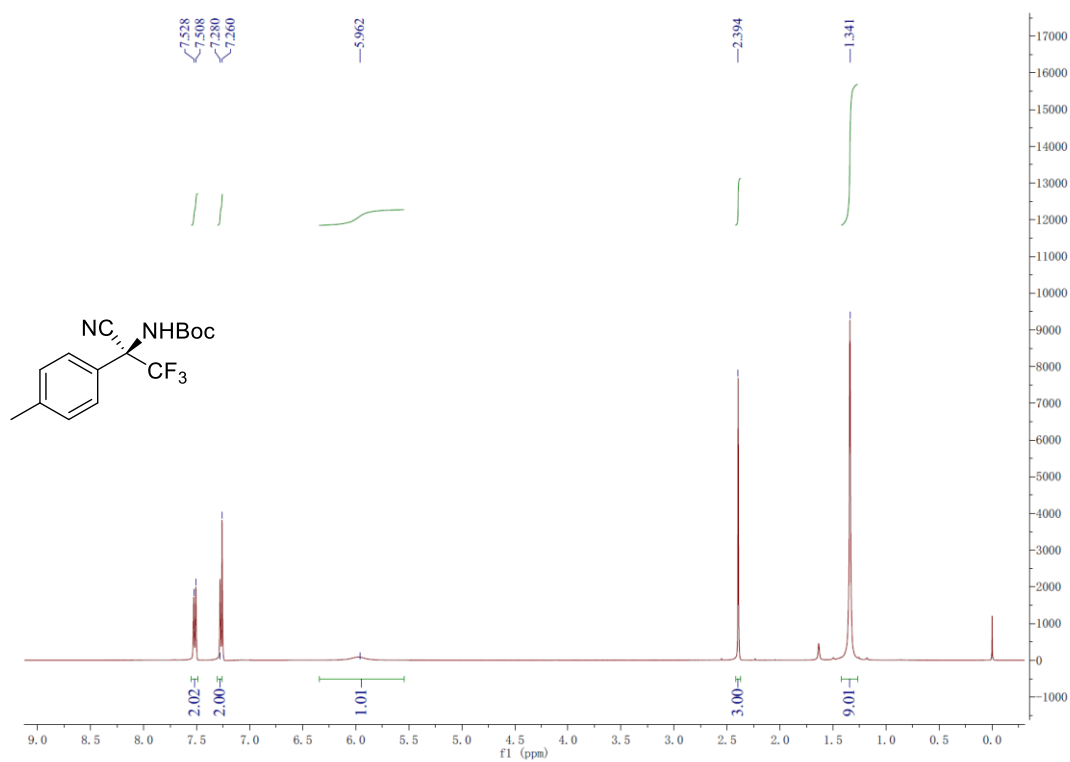
Peak#	Ret. Time	Area	Height	Area %	Height %
1	15.548	49836195	1384486	49.541	56.422
2	17.127	50760596	1069339	50.459	43.578
Total		100596791	2453825	100.000	100.000

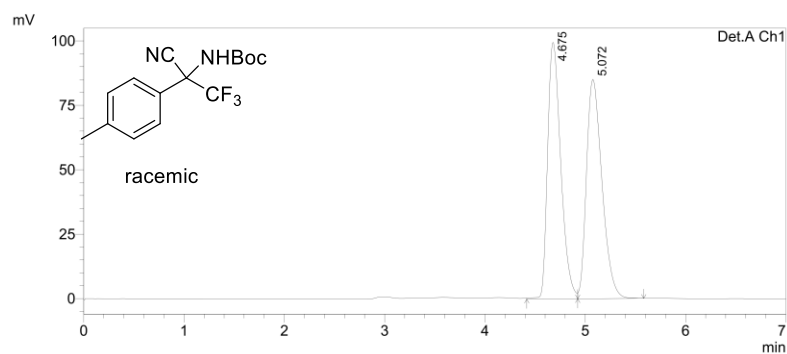
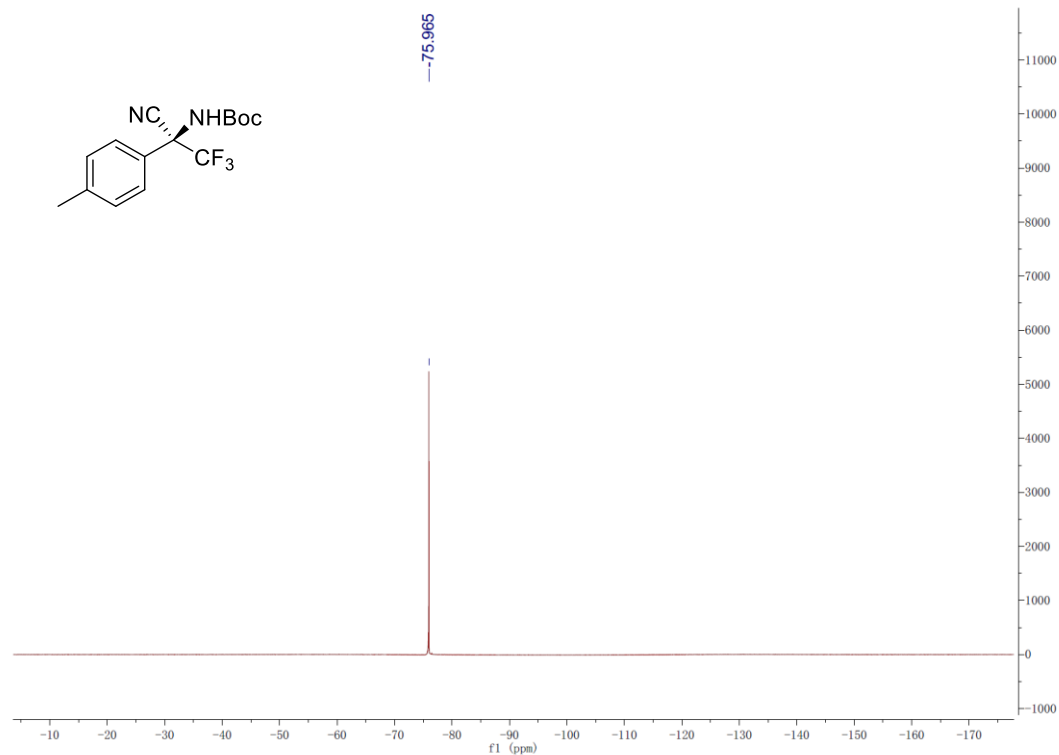


1 Det.A Ch1/254nm

PeakTable

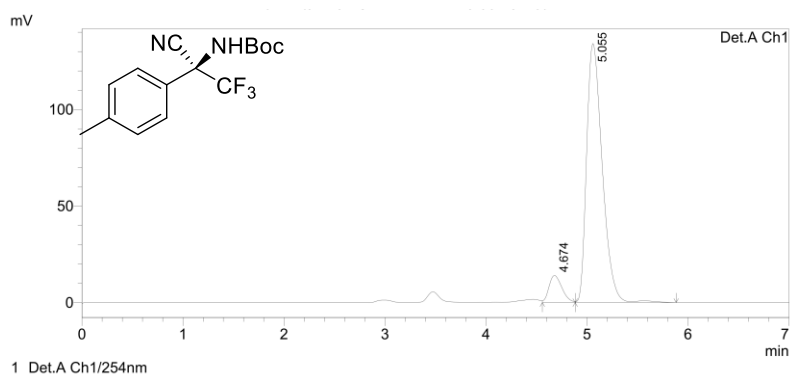
Peak#	Ret. Time	Area	Height	Area %	Height %
1	15.607	5159651	153472	14.932	19.620
2	17.155	29394668	628742	85.068	80.380
Total		34554319	782214	100.000	100.000





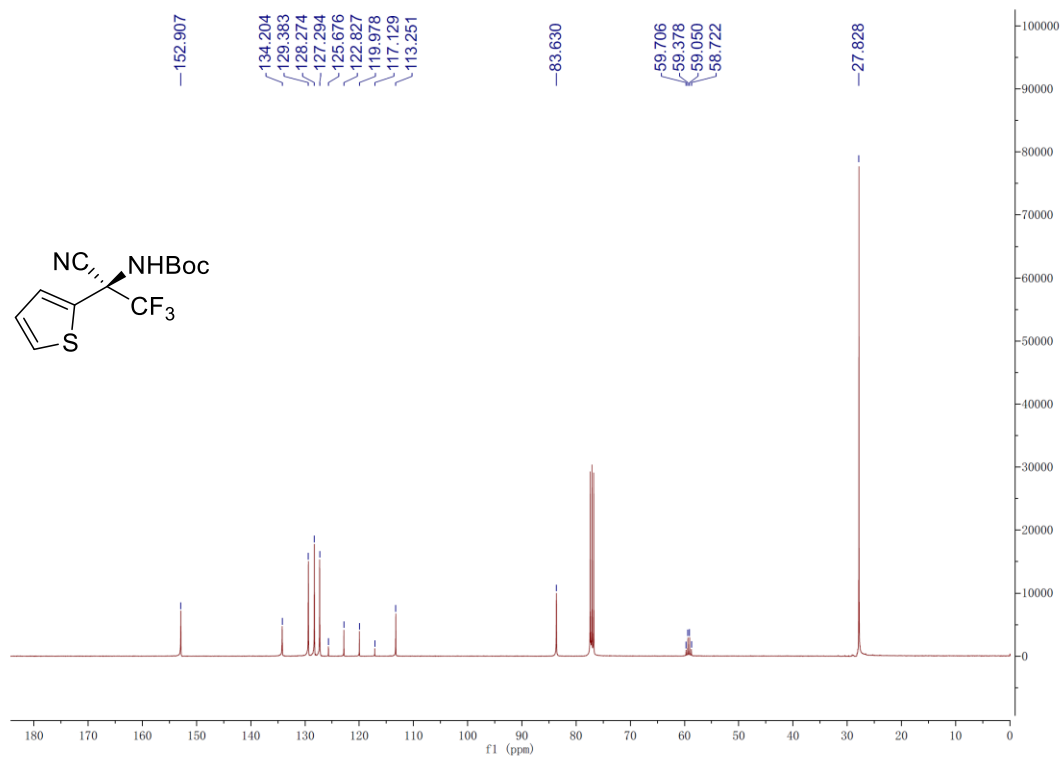
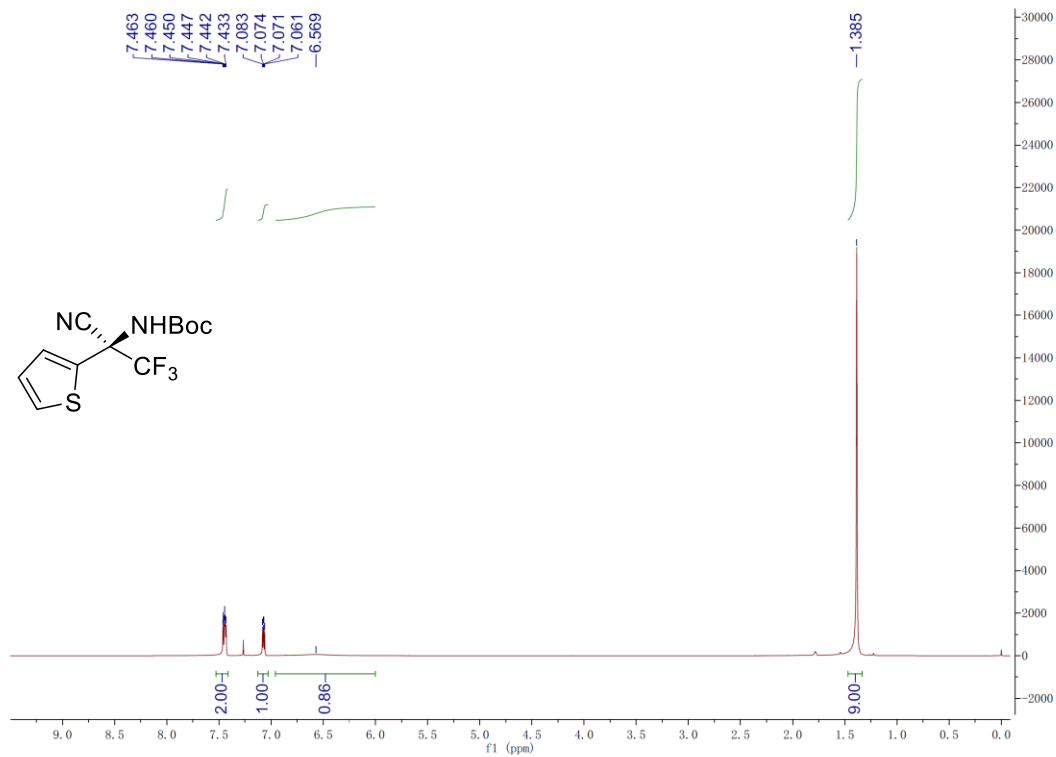
PeakTable

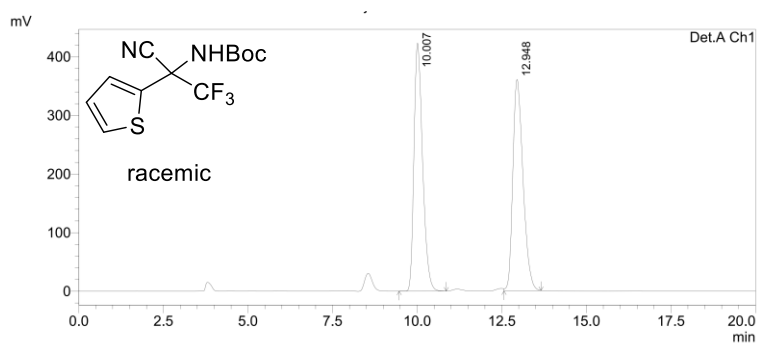
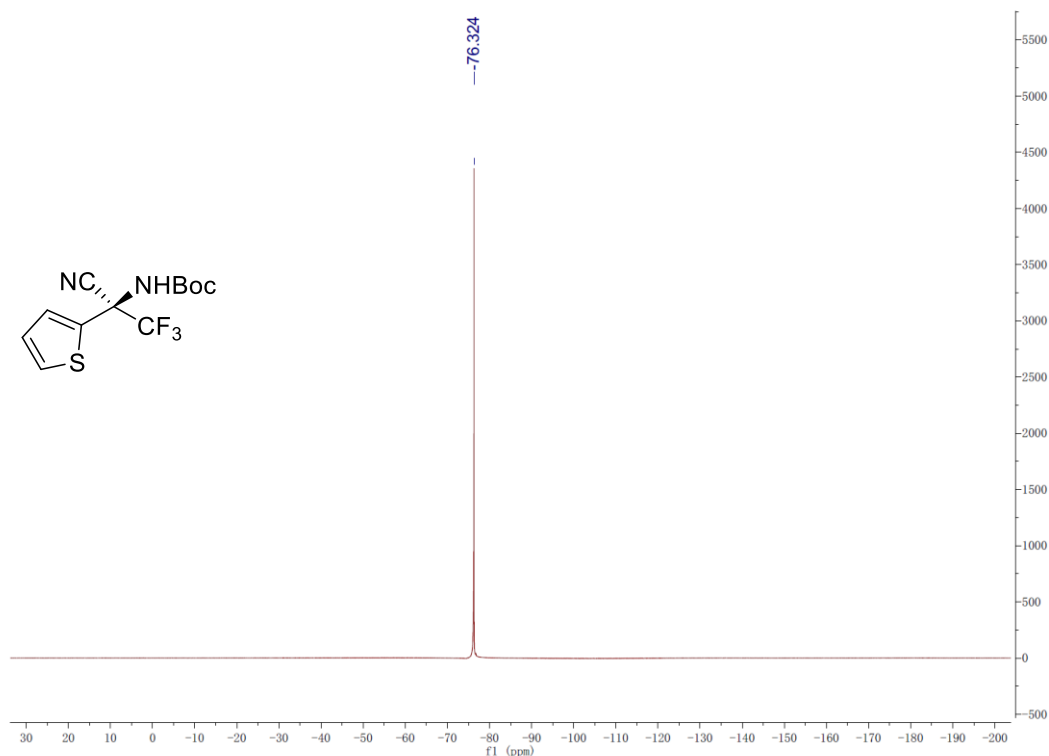
Peak#	Ret. Time	Area	Height	Area %	Height %
1	4.675	907044	99442	49.763	53.879
2	5.072	915695	85122	50.237	46.121
Total		1822740	184564	100.000	100.000



PeakTable

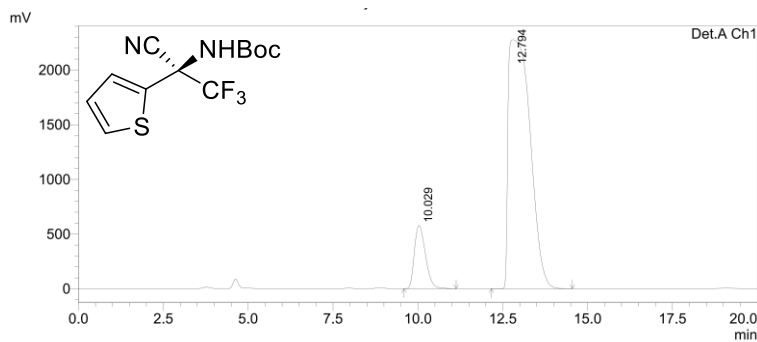
Peak#	Ret. Time	Area	Height	Area %	Height %
1	4.674	125300	14016	8.006	9.463
2	5.055	1439864	134099	91.994	90.537
Total		1565164	148115	100.000	100.000





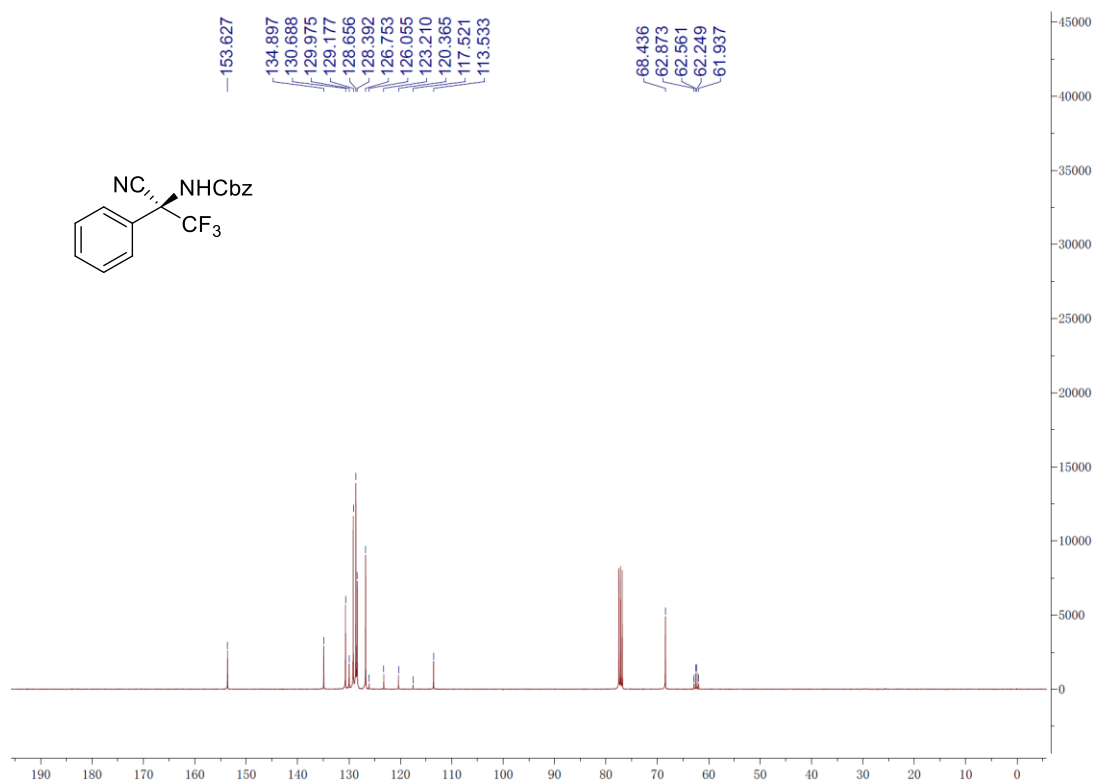
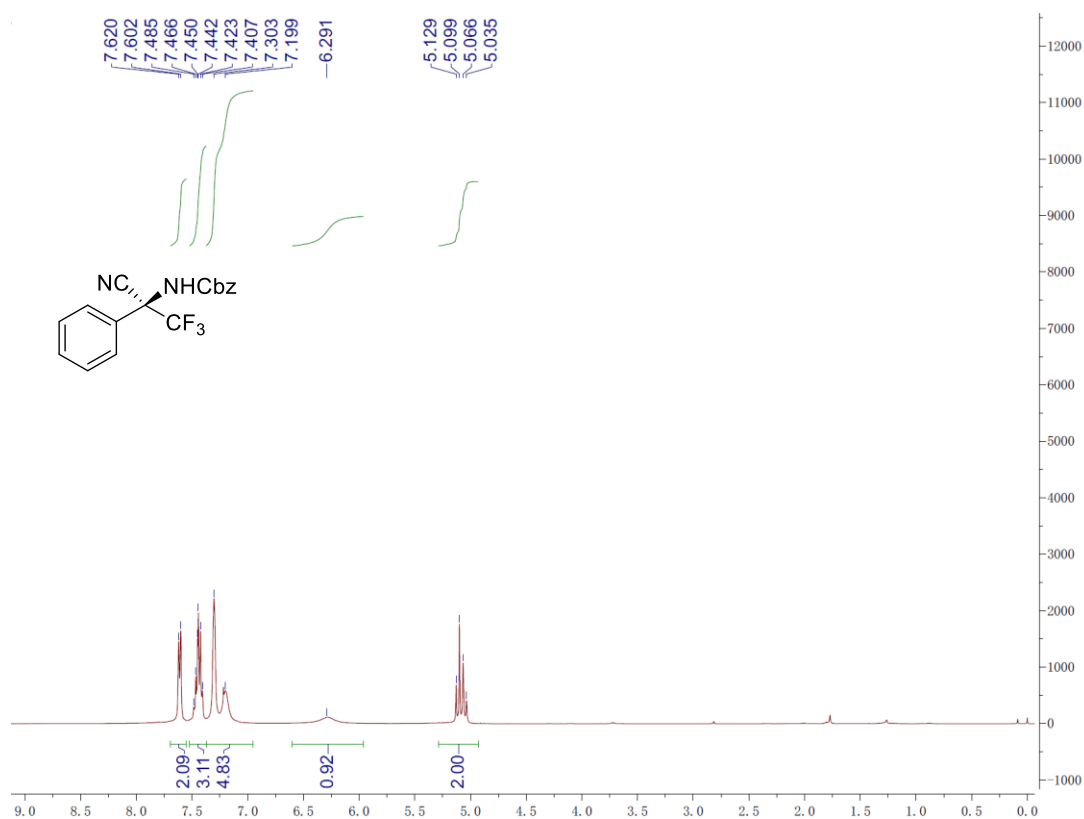
1 Det.A Ch1/220nm

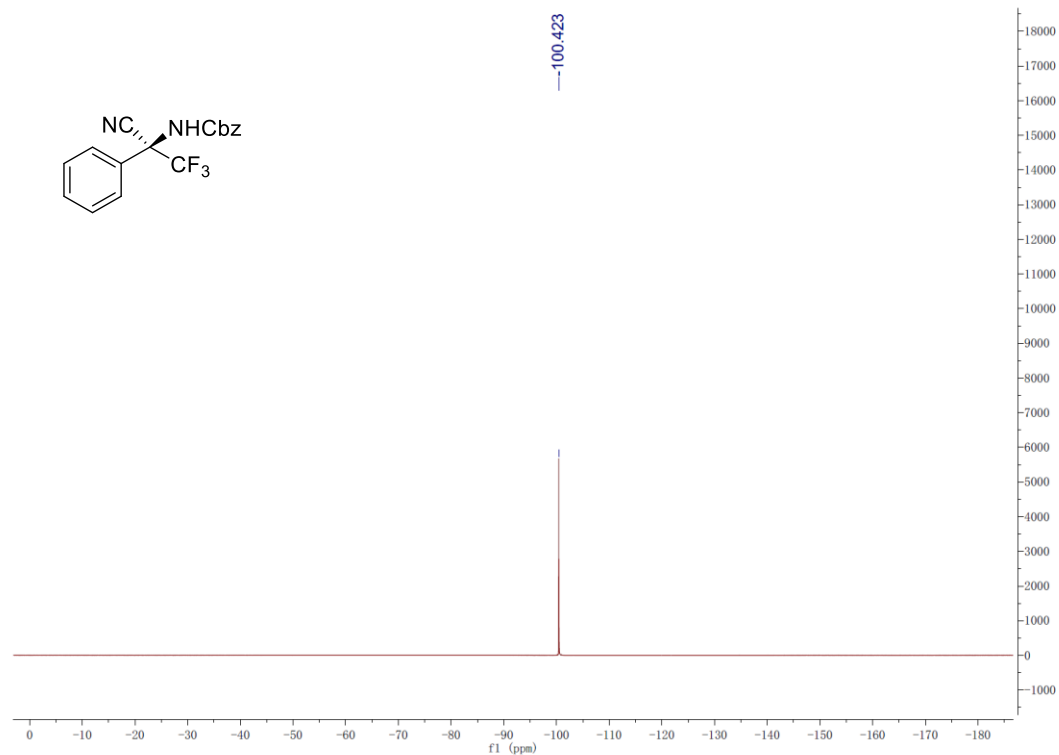
PeakTable					
Peak#	Ret. Time	Area	Height	Area %	Height %
1	10.007	7486355	423183	49.789	54.000
2	12.948	7549700	360488	50.211	46.000
Total		15036055	783671	100.000	100.000



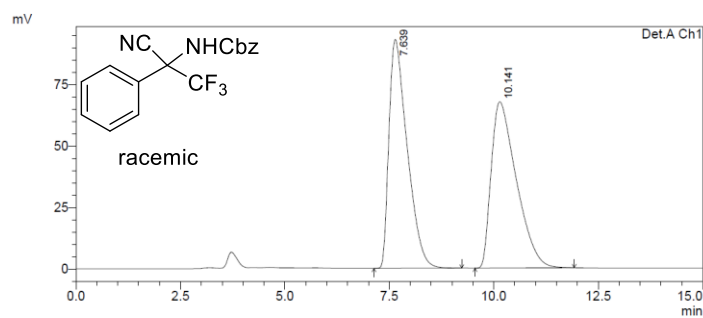
1 Det.A Ch1/220nm

PeakTable					
Peak#	Ret. Time	Area	Height	Area %	Height %
1	10.029	13124581	577375	11.385	20.230
2	12.794	102153772	2276748	88.615	79.770
Total		115278352	2854123	100.000	100.000





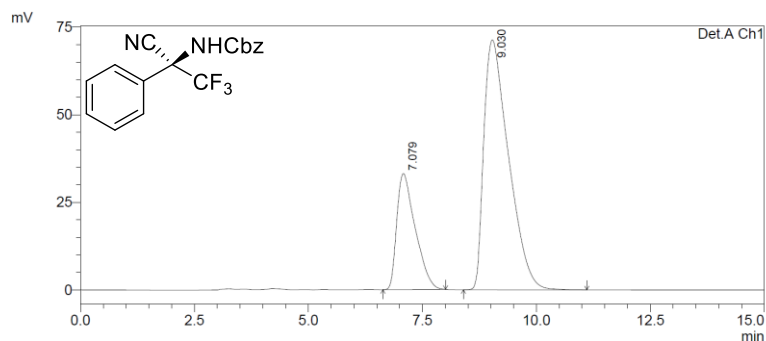
ee



1 Det.A Ch1/254nm

PeakTable

Peak#	Ret. Time	Area	Height	Area %	Height %
1	7.639	2828525	93104	49.928	57.917
2	10.141	2836710	67650	50.072	42.083
Total		5665235	160754	100.000	100.000

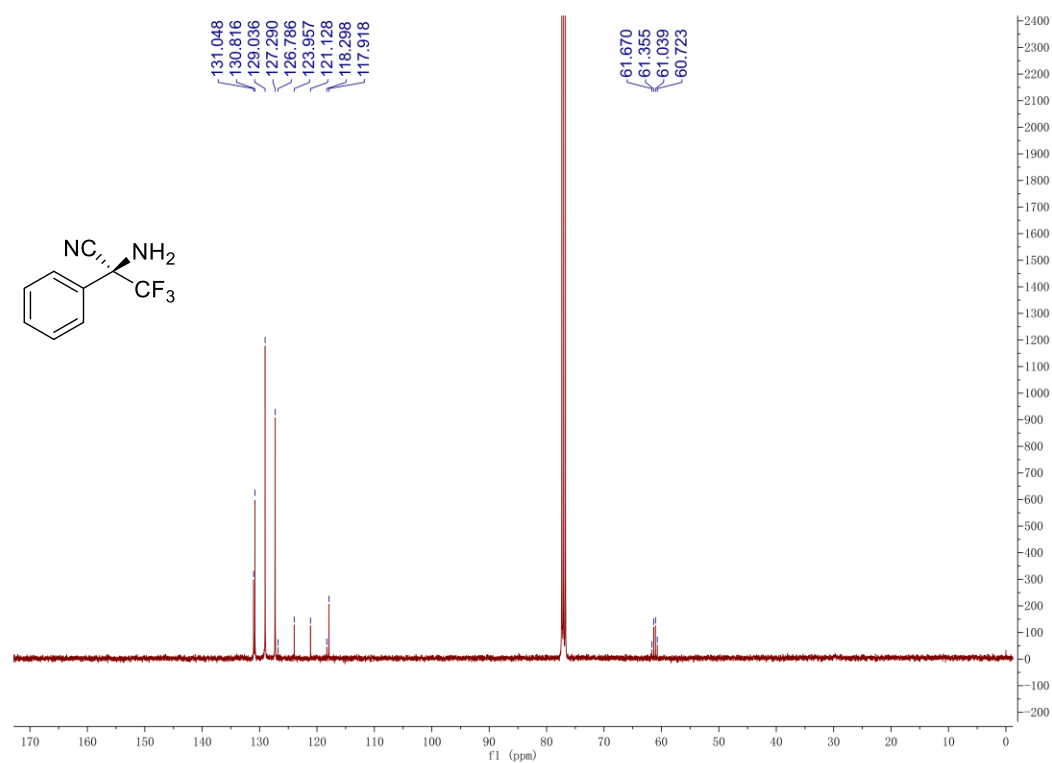
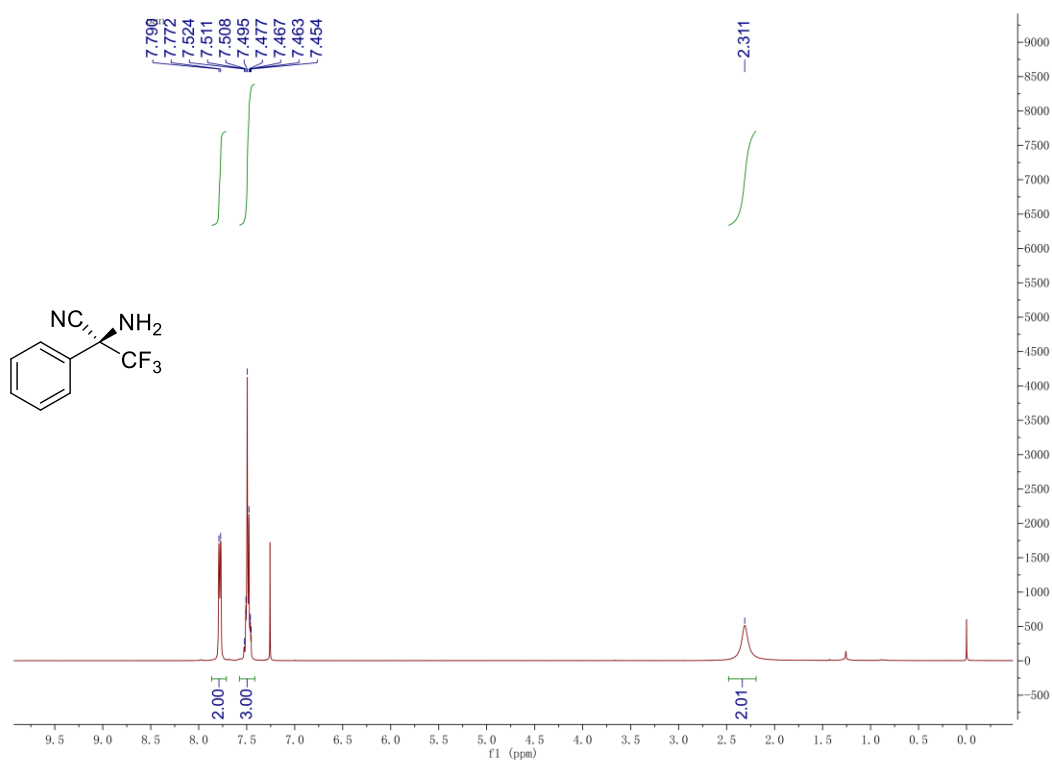


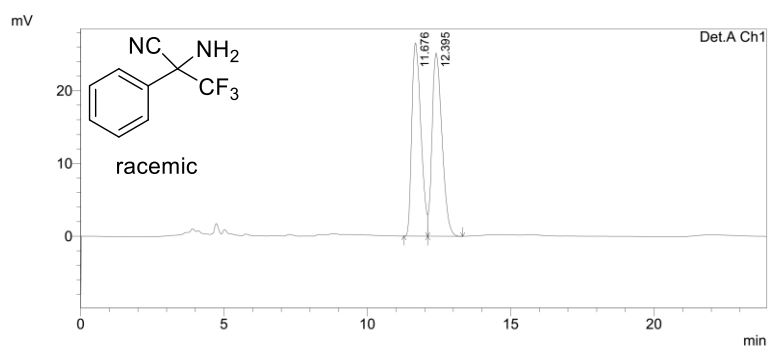
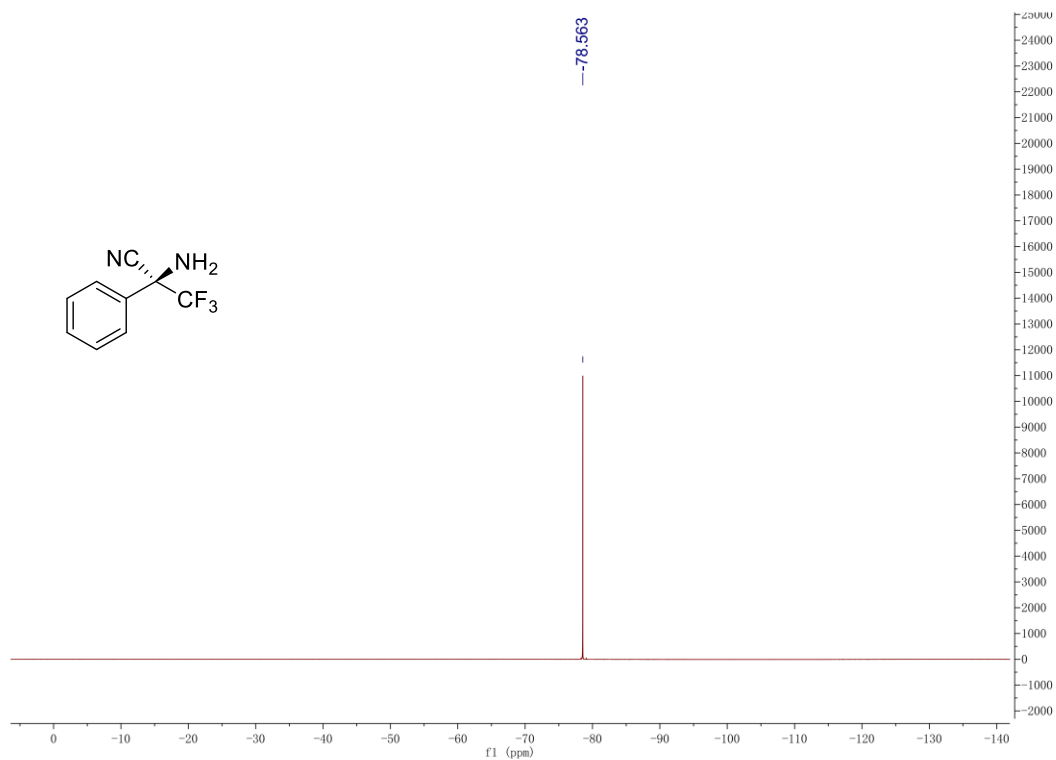
1 Det.A Ch1/254nm

PeakTable

Peak#	Ret. Time	Area	Height	Area %	Height %
1	7.079	932218	33136	25.695	31.711
2	9.030	2695840	71358	74.305	68.289
Total		3628058	104495	100.000	100.000

Copies of NMR and HPLC spectra for 5a-5f

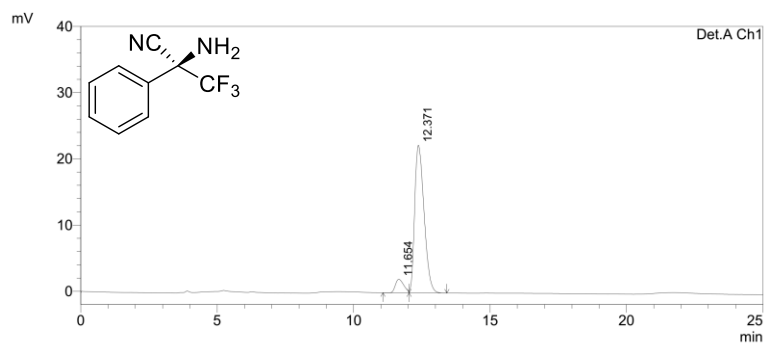




1 Det.A Ch1/220nm

PeakTable

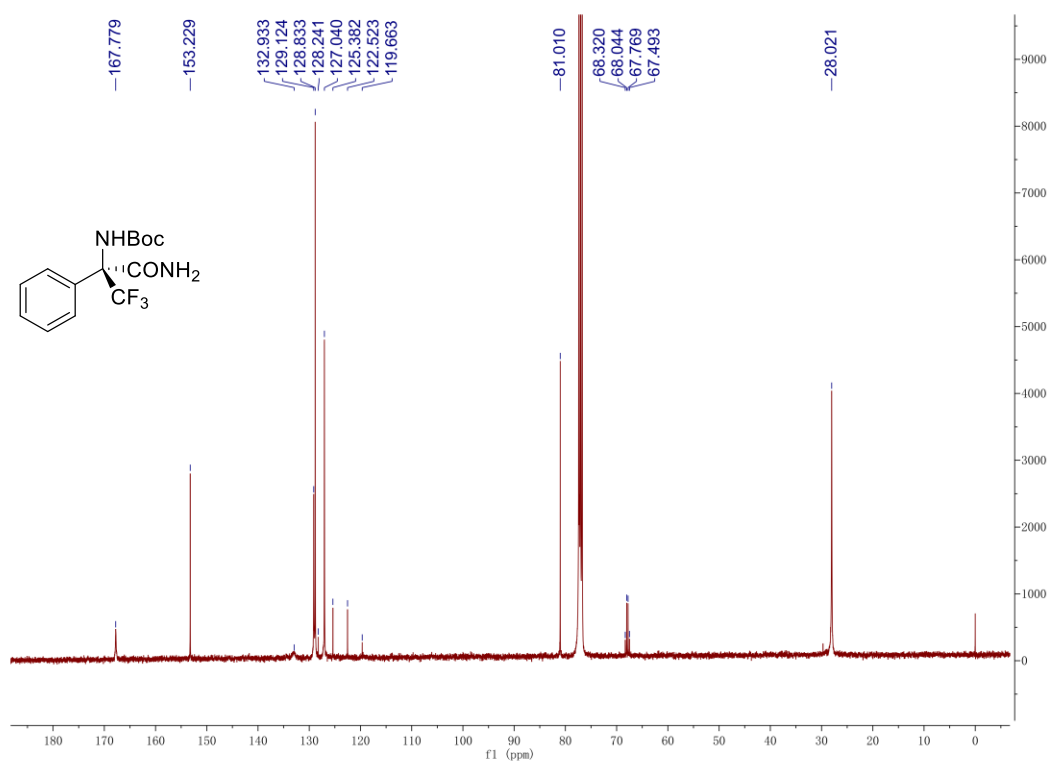
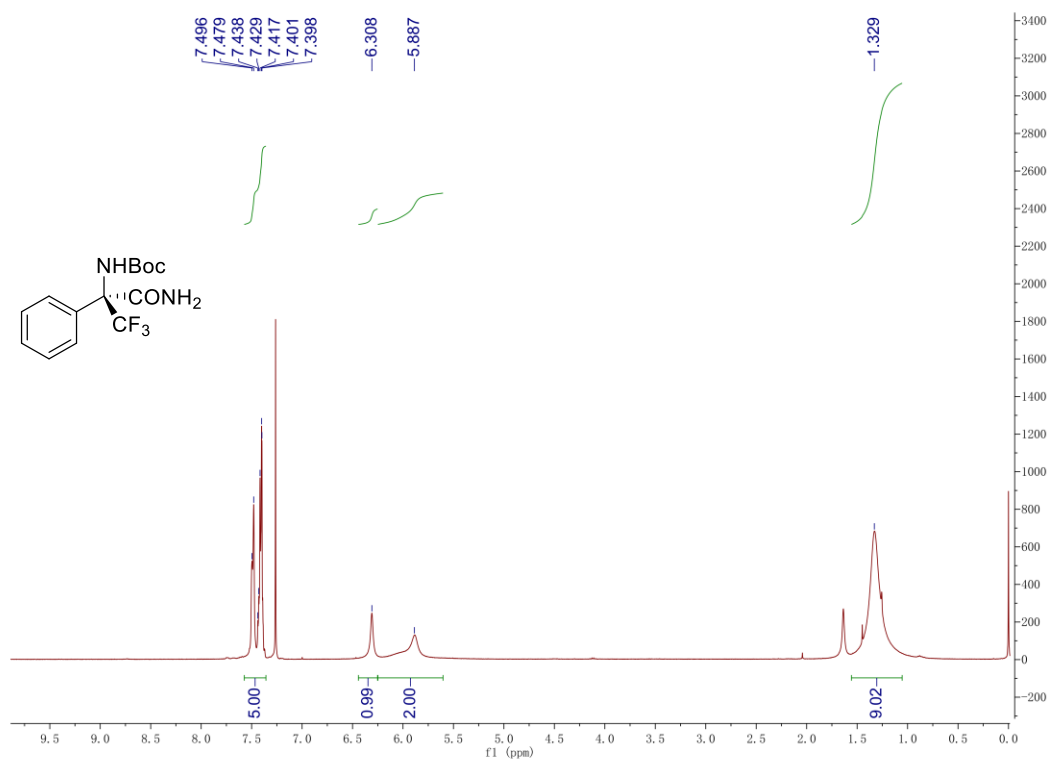
Peak#	Ret. Time	Area	Height	Area %	Height %
1	11.676	590196	26563	49.251	51.346
2	12.395	608146	25170	50.749	48.654
Total		1198342	51732	100.000	100.000

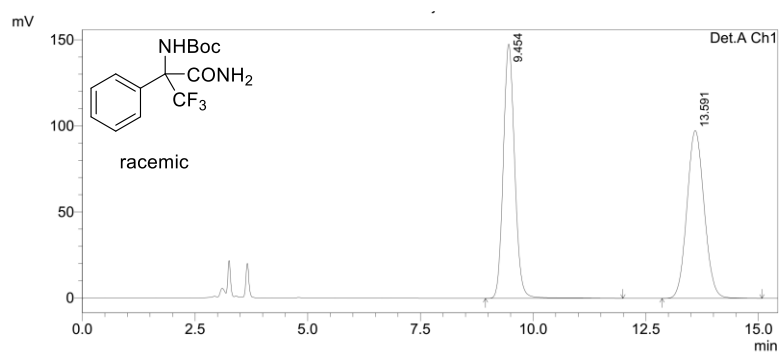
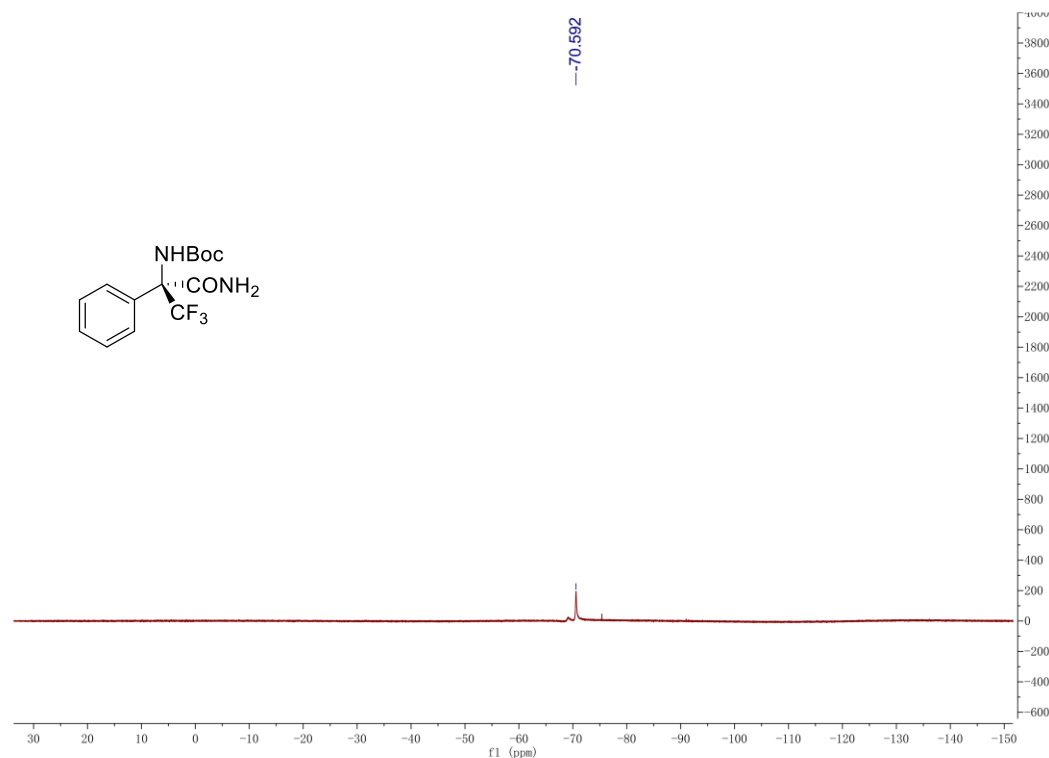


1 Det.A Ch1/220nm

PeakTable

Peak#	Ret. Time	Area	Height	Area %	Height %
1	11.654	43659	2040	7.556	8.376
2	12.371	534179	22314	92.444	91.624
Total		577838	24354	100.000	100.000

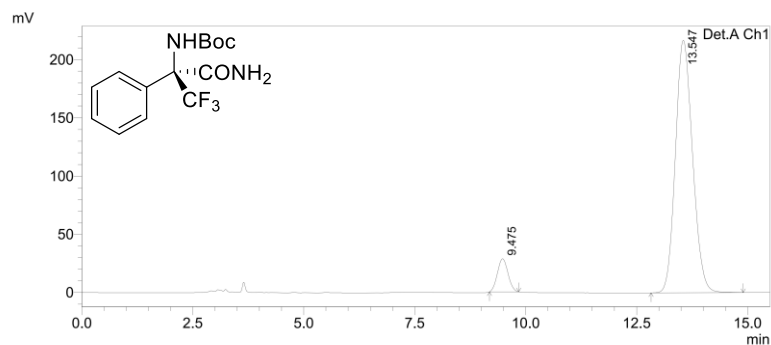




1 Det.A Ch1/220nm

PeakTable

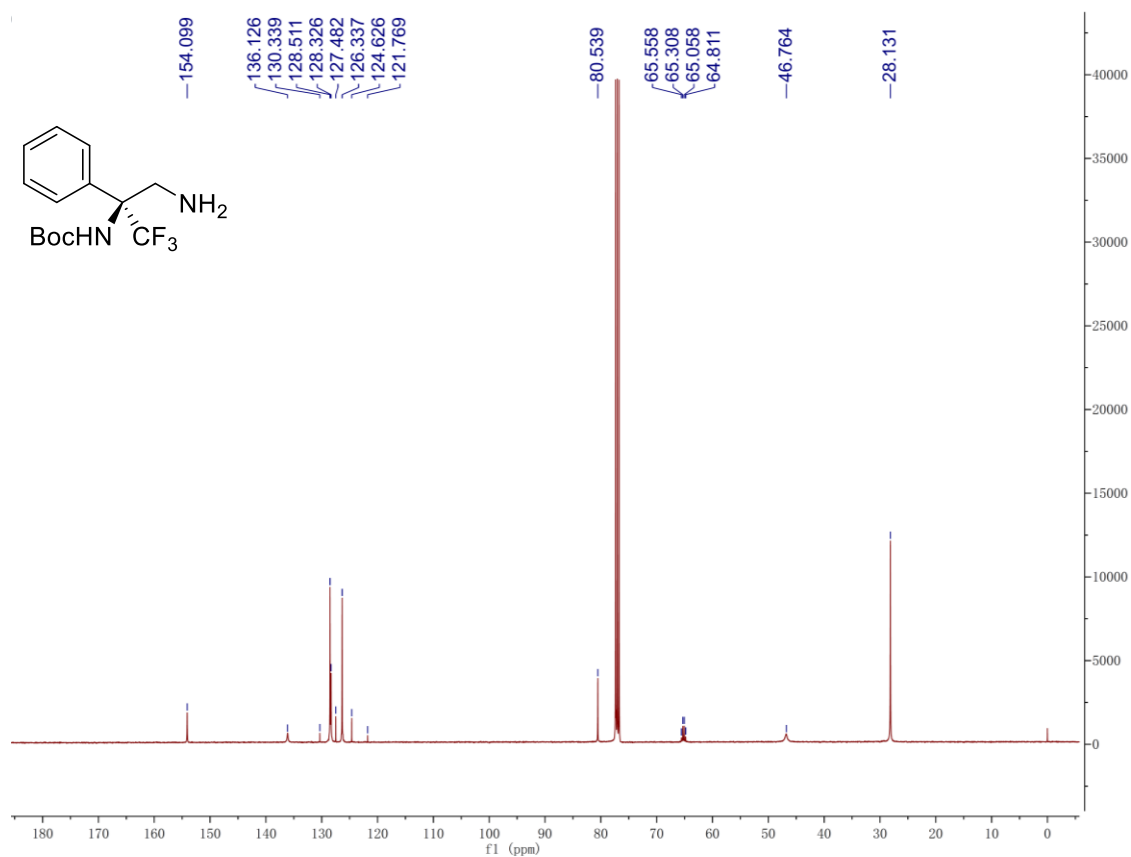
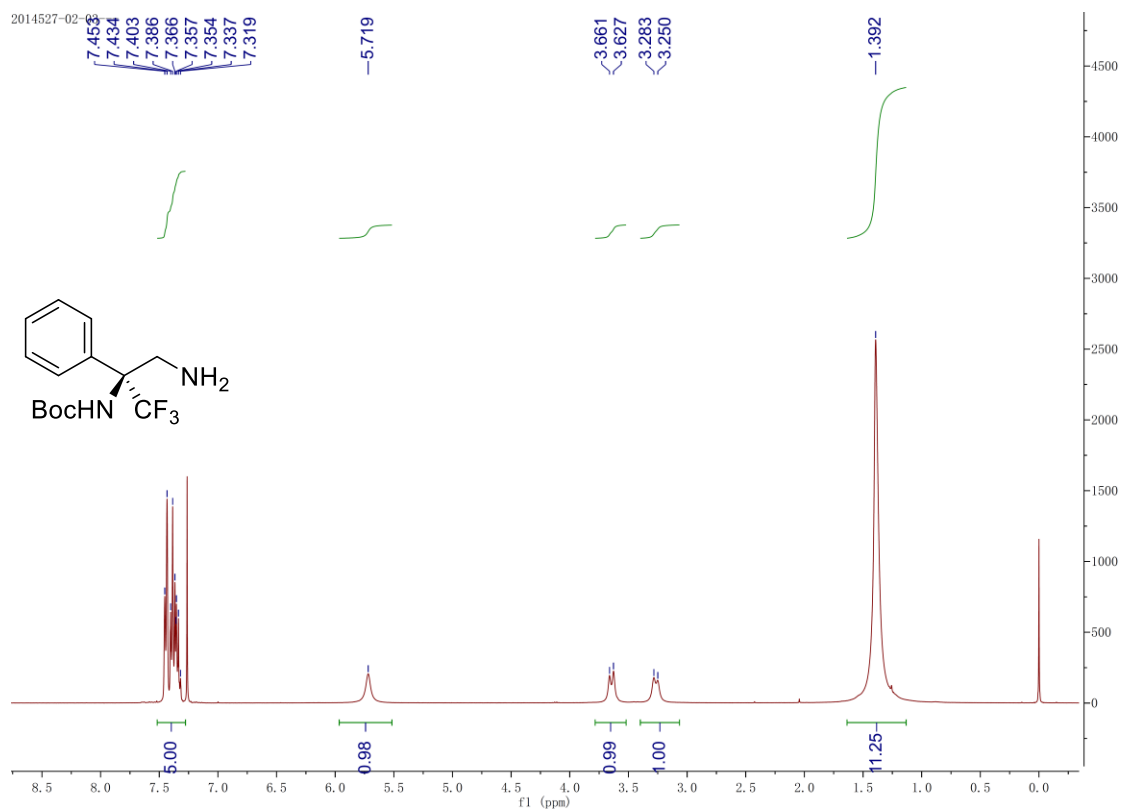
Peak#	Ret. Time	Area	Height	Area %	Height %
1	9.454	2597590	147805	50.048	60.250
2	13.591	2592658	97516	49.952	39.750
Total		5190249	245320	100.000	100.000



1 Det.A Ch1/220nm

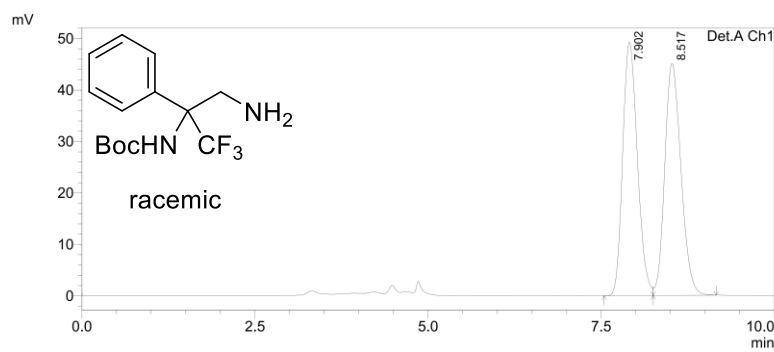
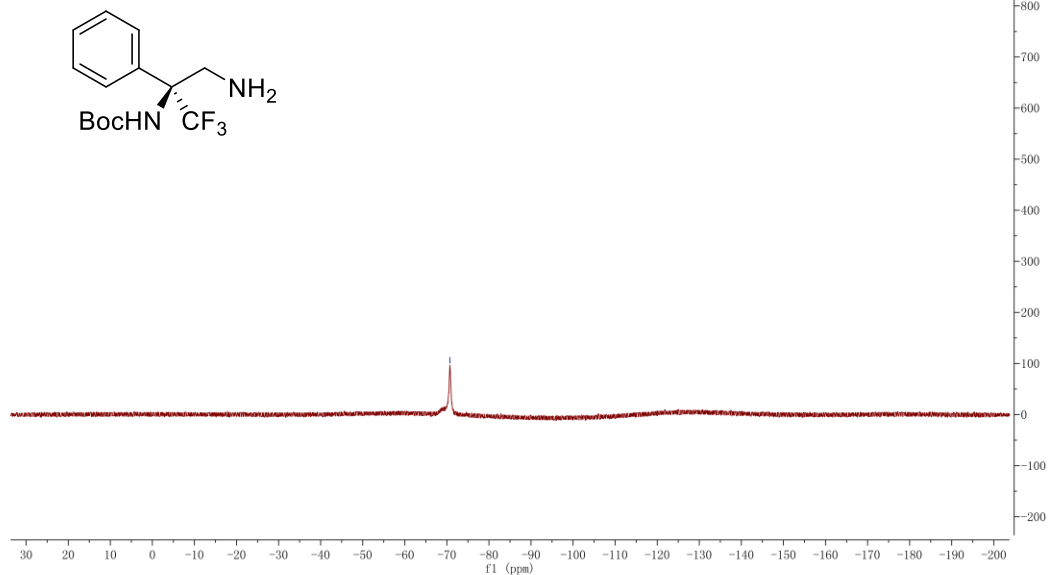
PeakTable

Peak#	Ret. Time	Area	Height	Area %	Height %
1	9.475	470381	28371	7.458	11.568
2	13.547	5836663	216892	92.542	88.432
Total		6307044	245263	100.000	100.000



2017226-CH2NH2_FLUORINE_01
2017226-CH2NH2

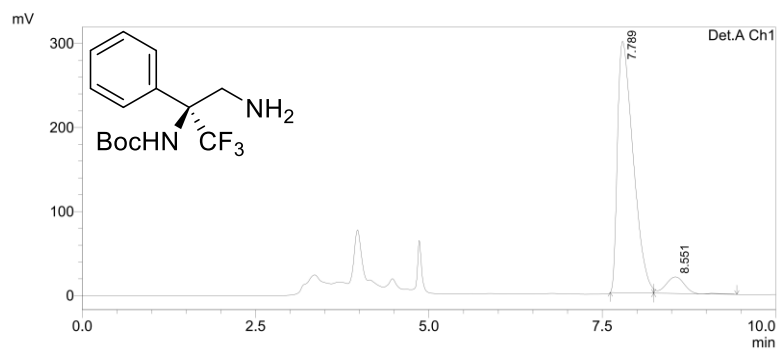
—70.749



1 Det.A Ch1/254nm

PeakTable

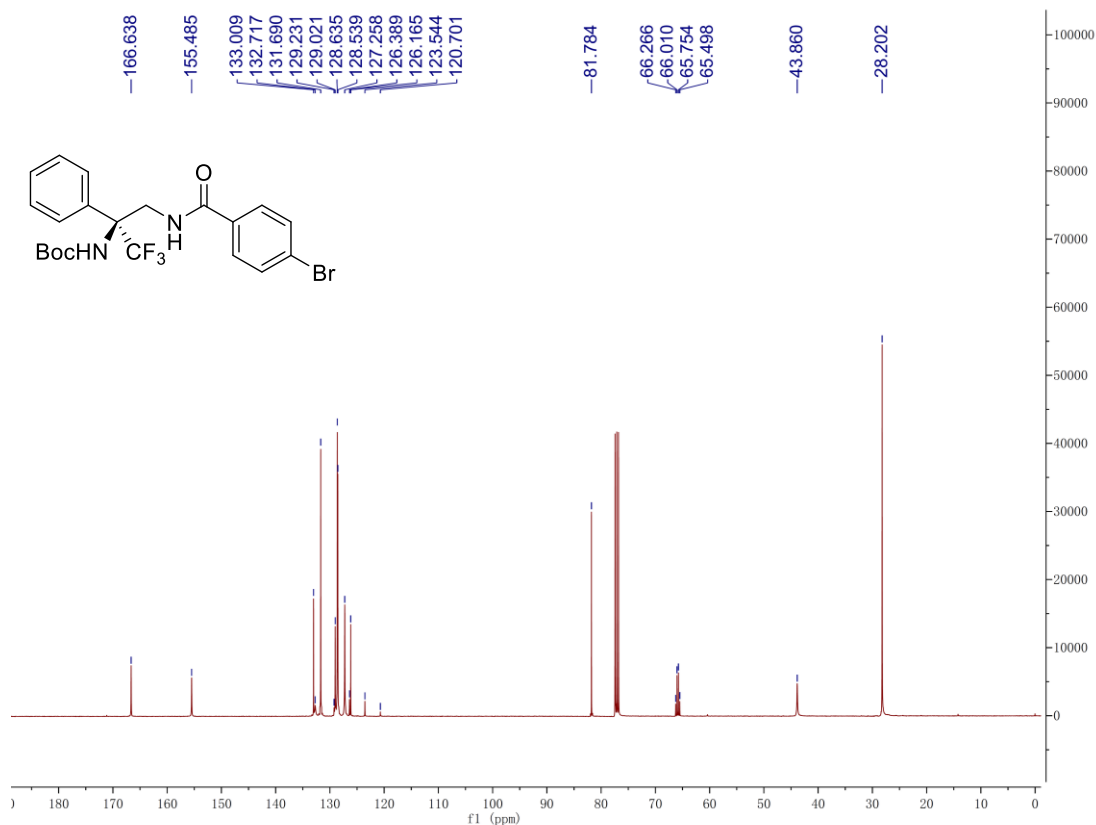
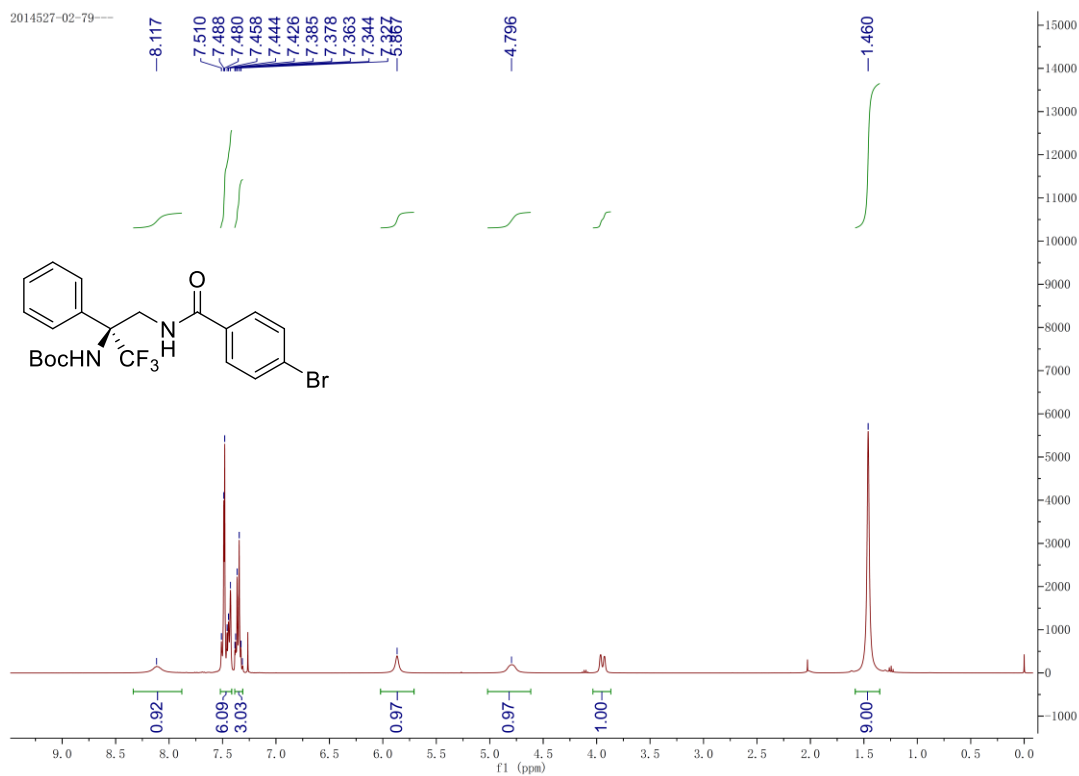
Peak#	Ret. Time	Area	Height	Area %	Height %
1	7.902	726903	49250	49.712	52.241
2	8.517	735324	45025	50.288	47.759
Total		1462227	94275	100.000	100.000

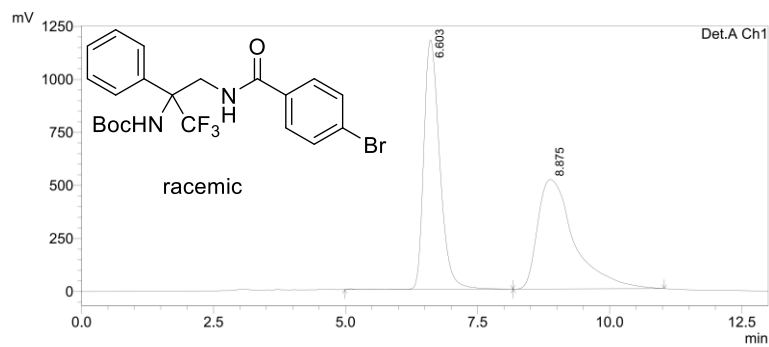
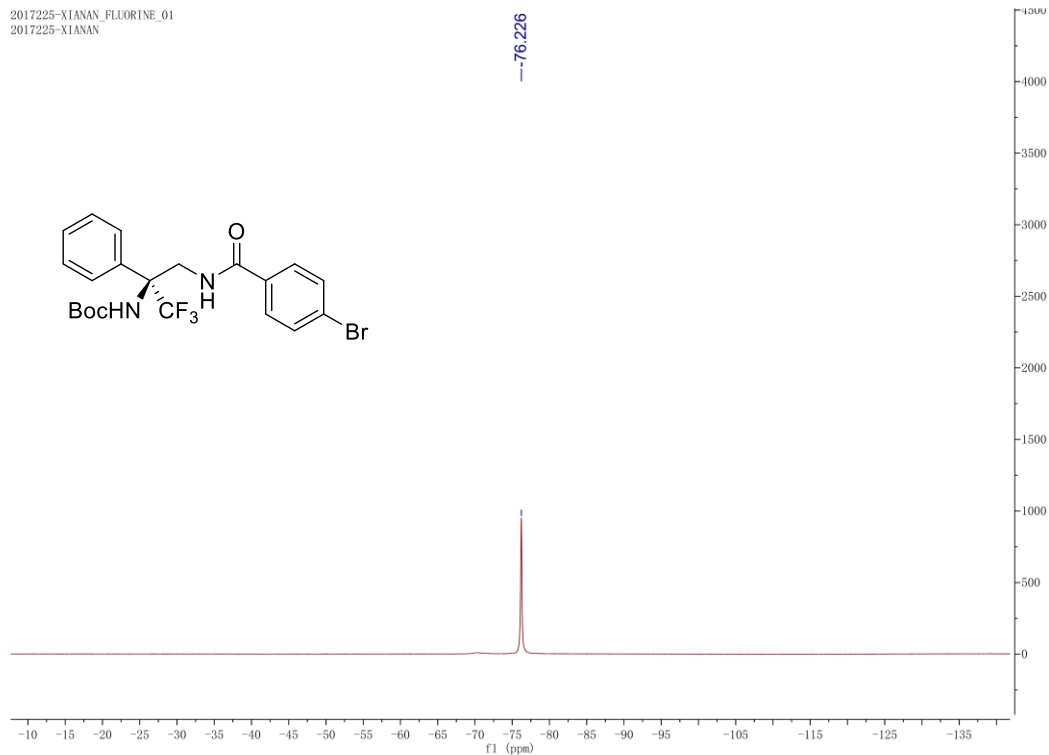


1 Det.A Ch1/254nm

PeakTable

Peak#	Ret. Time	Area	Height	Area %	Height %
1	7.789	4703628	299095	92.558	93.827
2	8.551	378214	19679	7.442	6.173
Total		5081842	318774	100.000	100.000

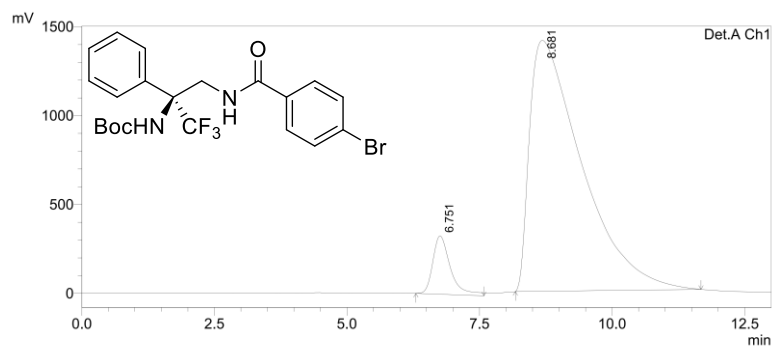




1 Det.A Ch1/254nm

PeakTable

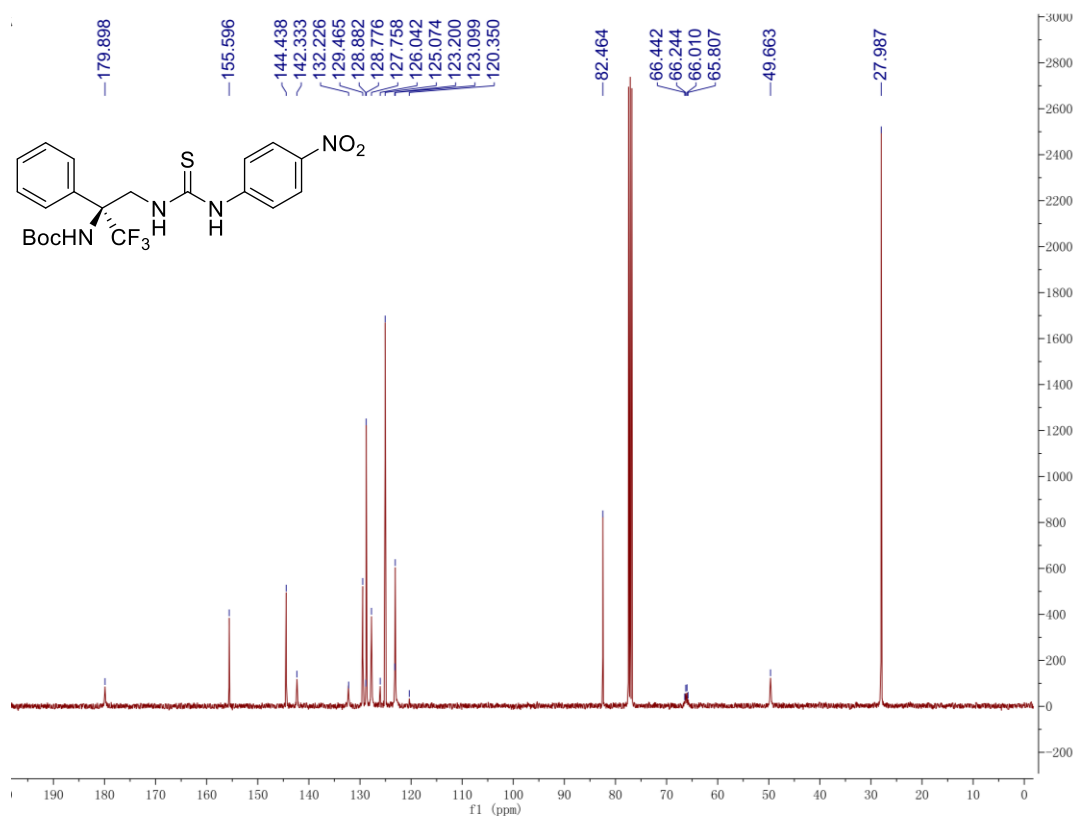
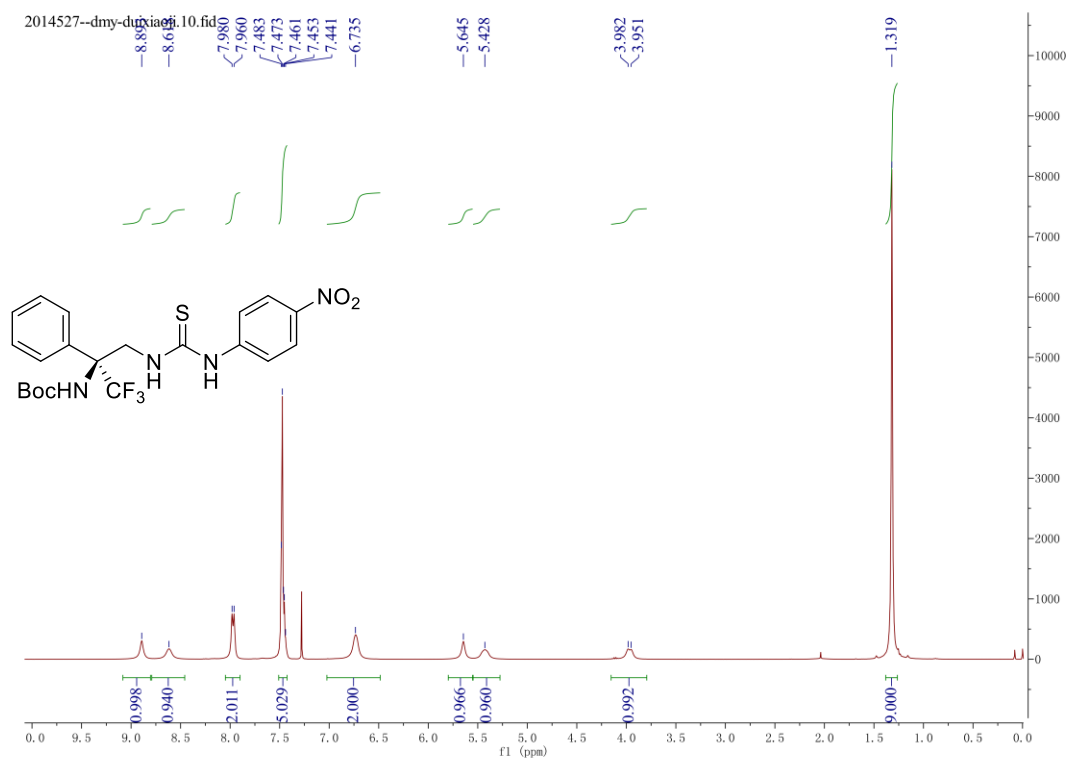
Peak#	Ret. Time	Area	Height	Area %	Height %
1	6.603	24556553	1176033	49.646	69.510
2	8.875	24906577	515866	50.354	30.490
Total		49463130	1691899	100.000	100.000

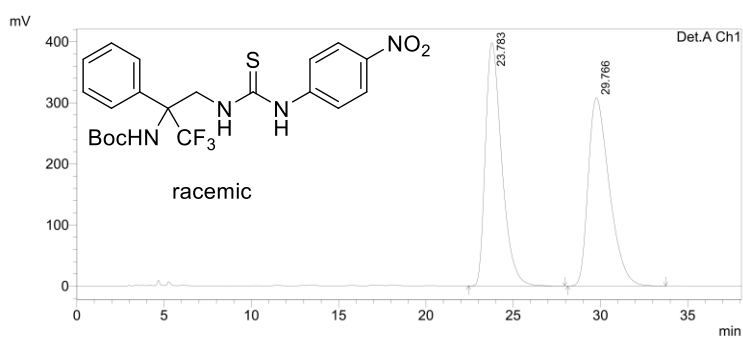
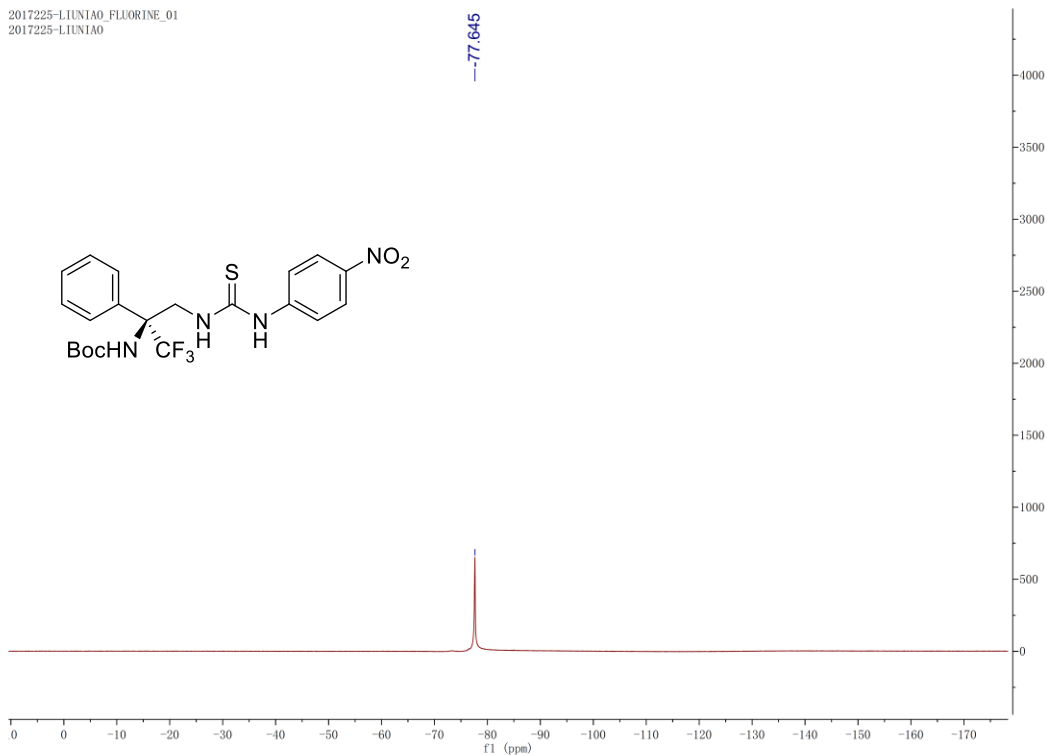


1 Det.A Ch1/254nm

PeakTable

Peak#	Ret. Time	Area	Height	Area %	Height %
1	6.751	7568486	328220	7.255	18.874
2	8.681	96756331	1410833	92.745	81.126
Total		104324817	1739053	100.000	100.000

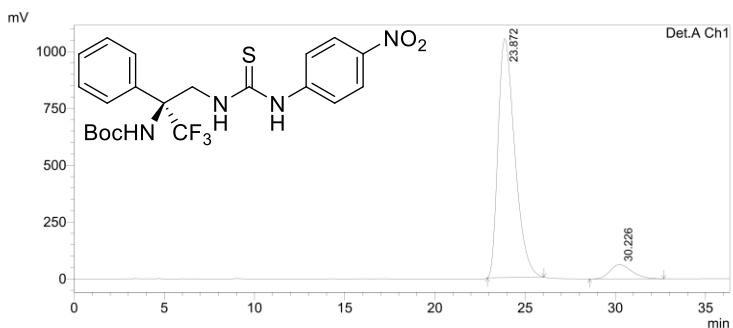




1 Det.A Ch1/254nm

PeakTable

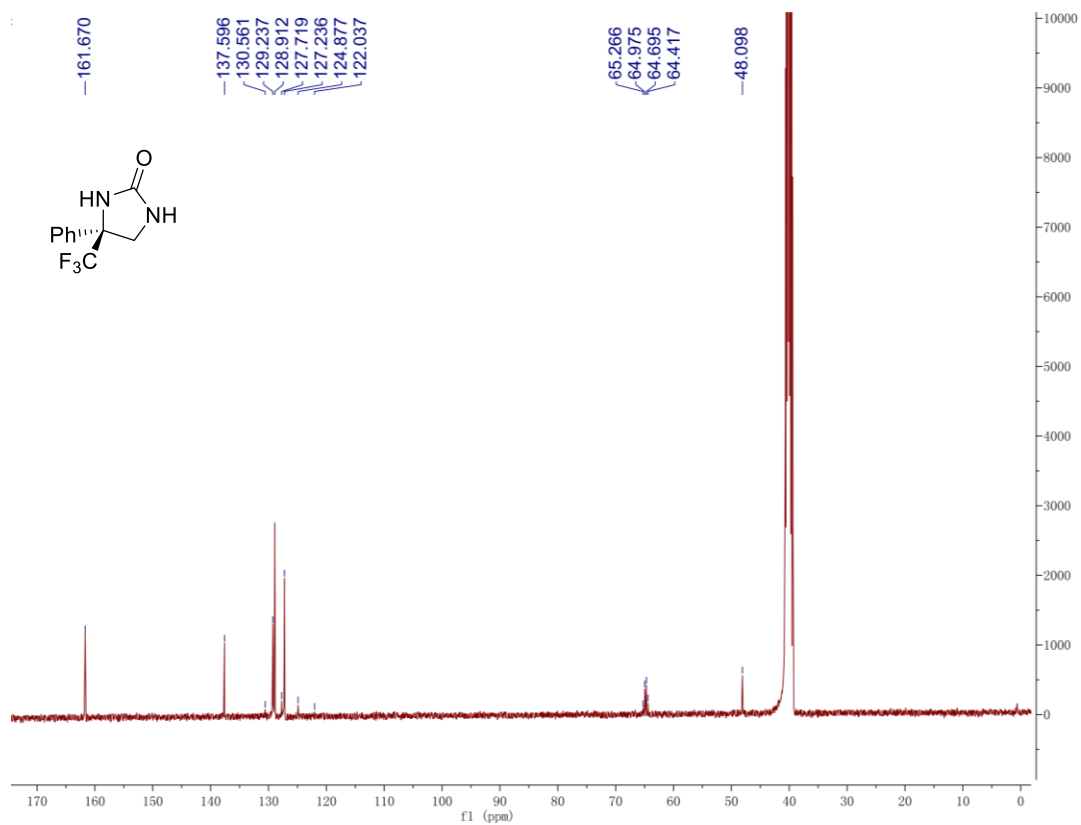
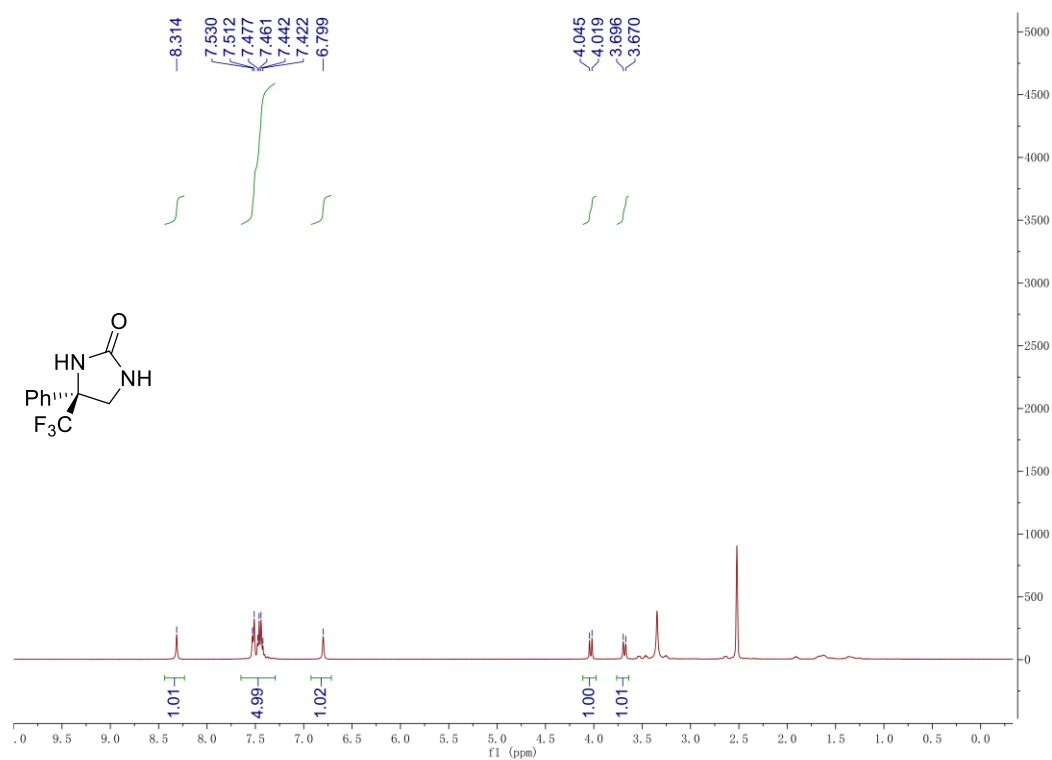
Detector A Ch1 254nm					
Peak#	Ret. Time	Area	Height	Area %	Height %
1	23.783	25979287	398267	50.068	56.416
2	29.766	25909195	307674	49.932	43.584
Total		51888482	705941	100.000	100.000

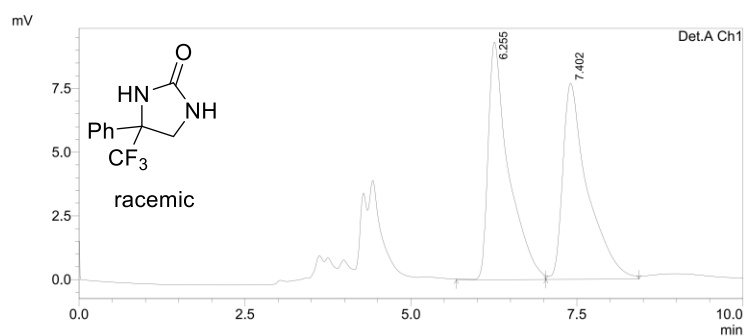
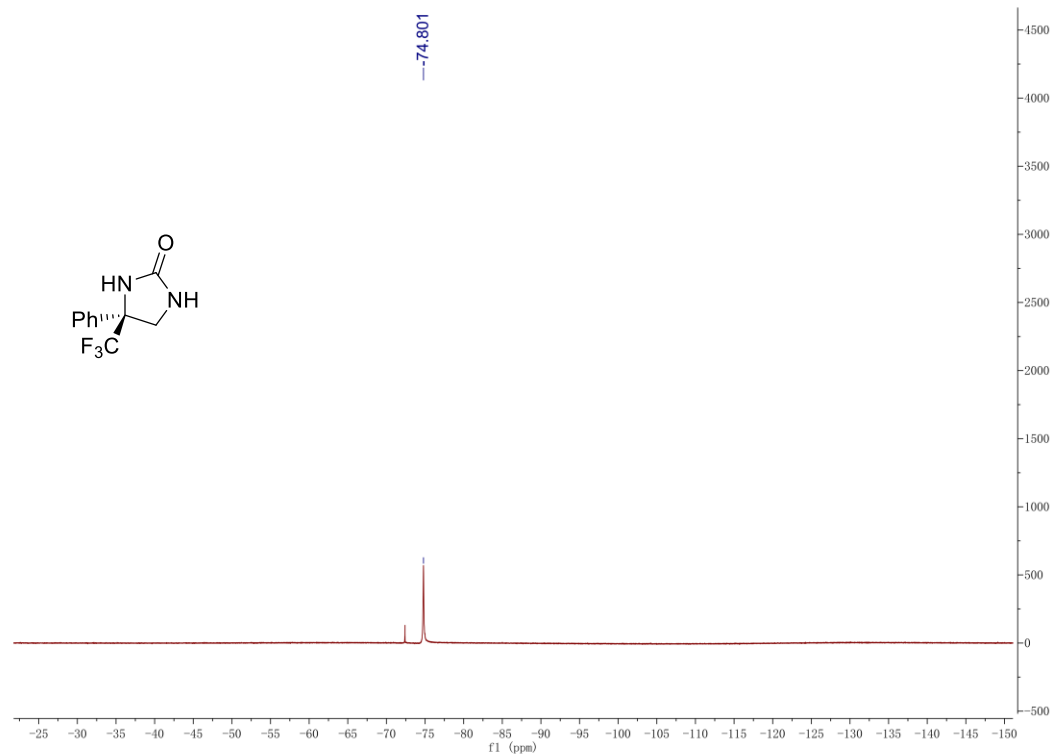


1 Det.A Ch1/254nm

PeakTable

Detector A Ch1 254nm					
Peak#	Ret. Time	Area	Height	Area %	Height %
1	23.872	67918212	1051086	92.854	94.390
2	30.226	5227166	62467	7.146	5.610
Total		73145378	1113553	100.000	100.000

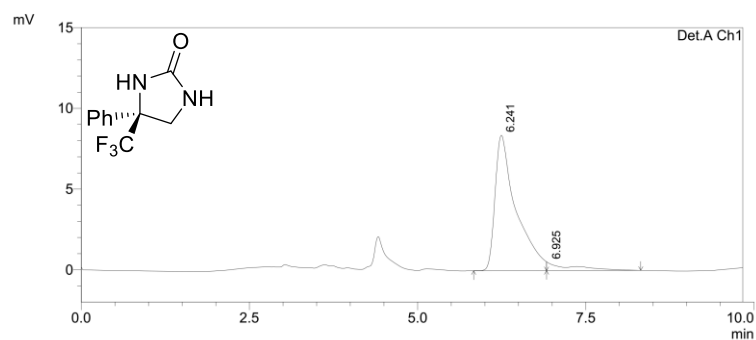




1 Det.A Ch1/220nm

PeakTable

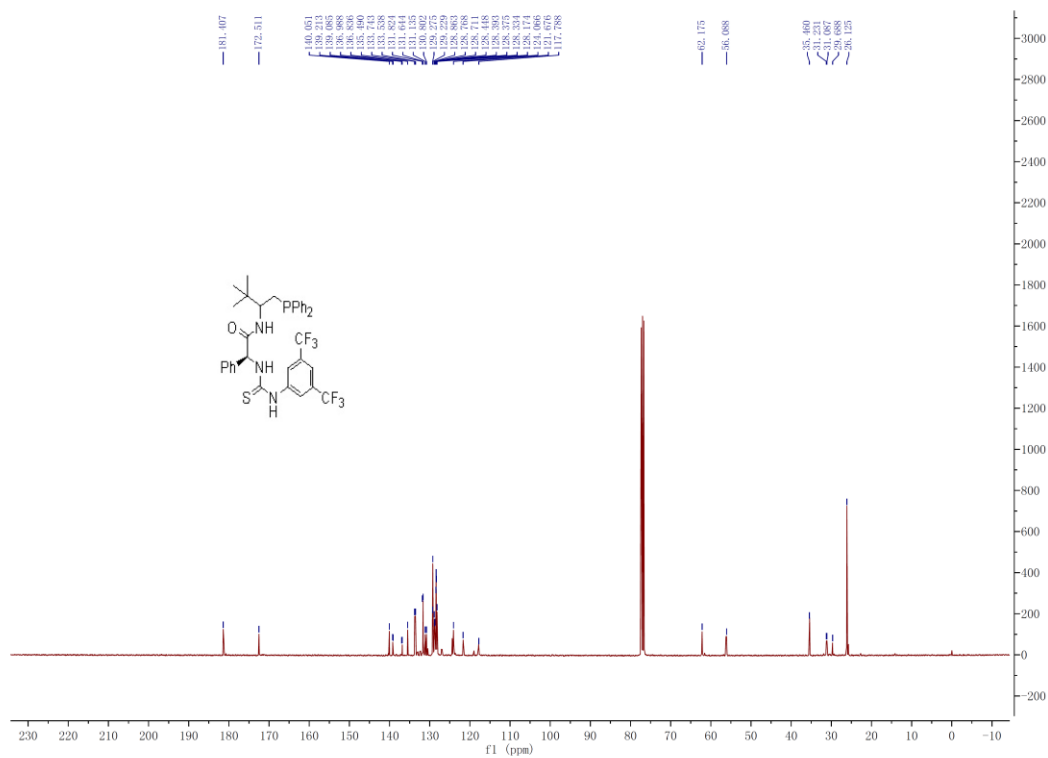
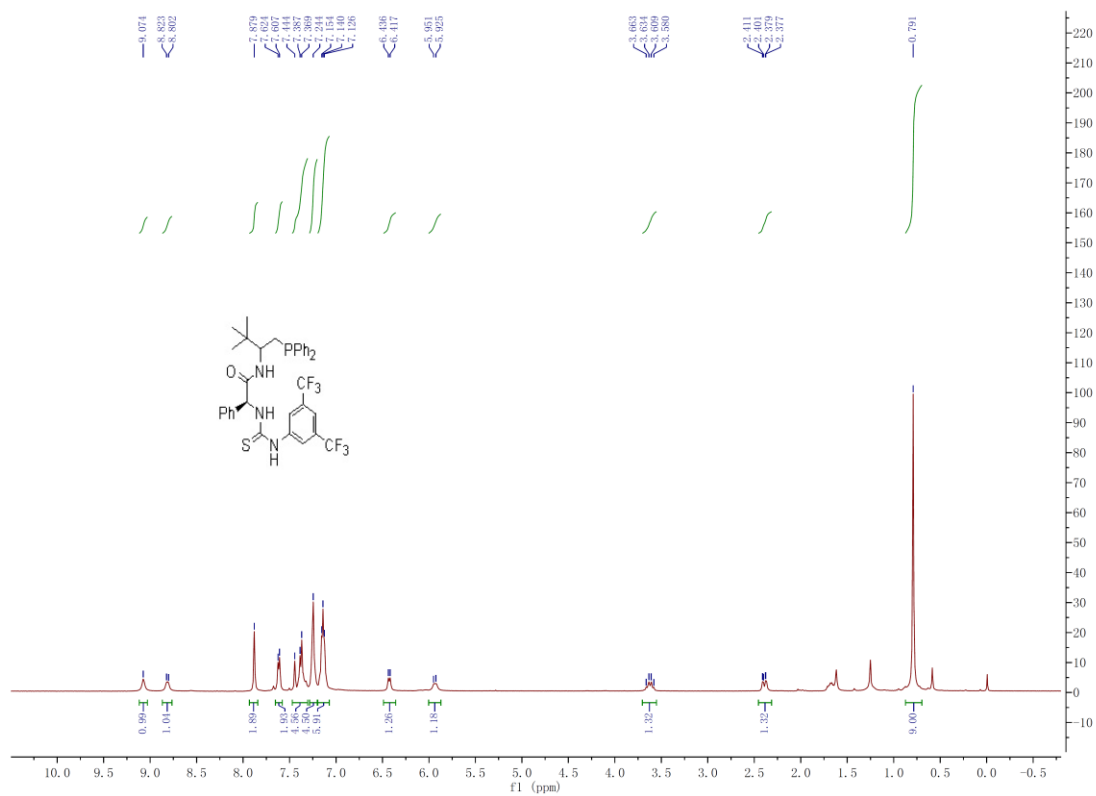
Peak#	Ret. Time	Area	Height	Area %	Height %
1	6.255	200620	9329	49.998	54.791
2	7.402	200635	7698	50.002	45.209
Total		401256	17027	100.000	100.000

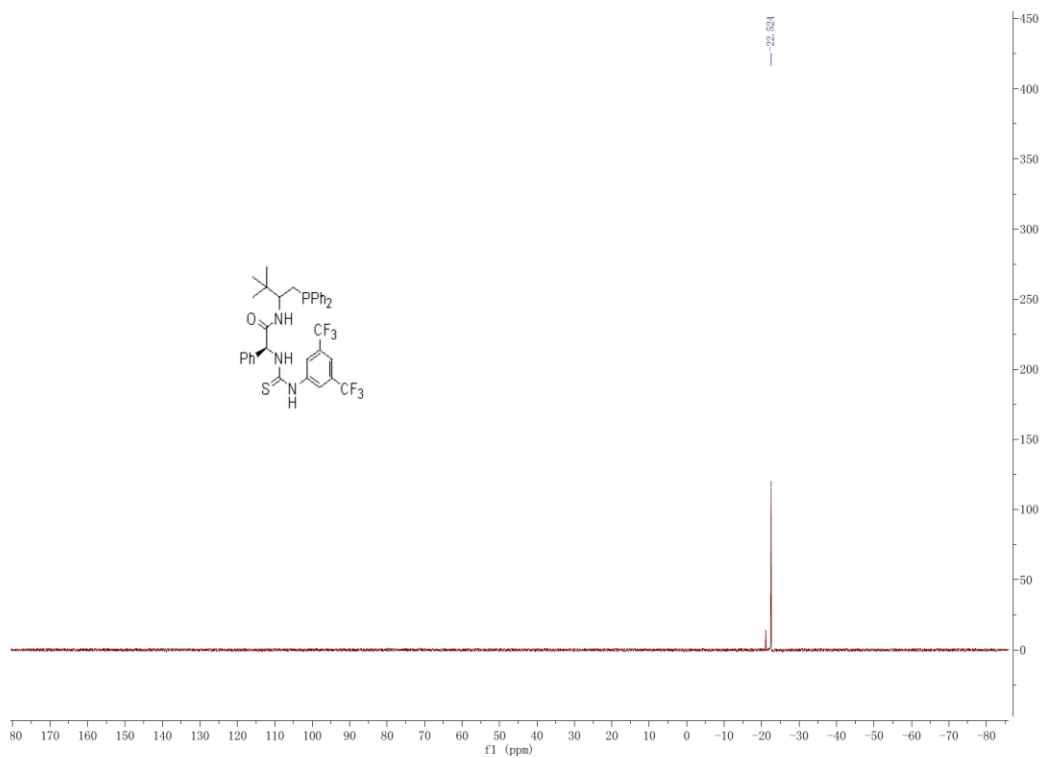
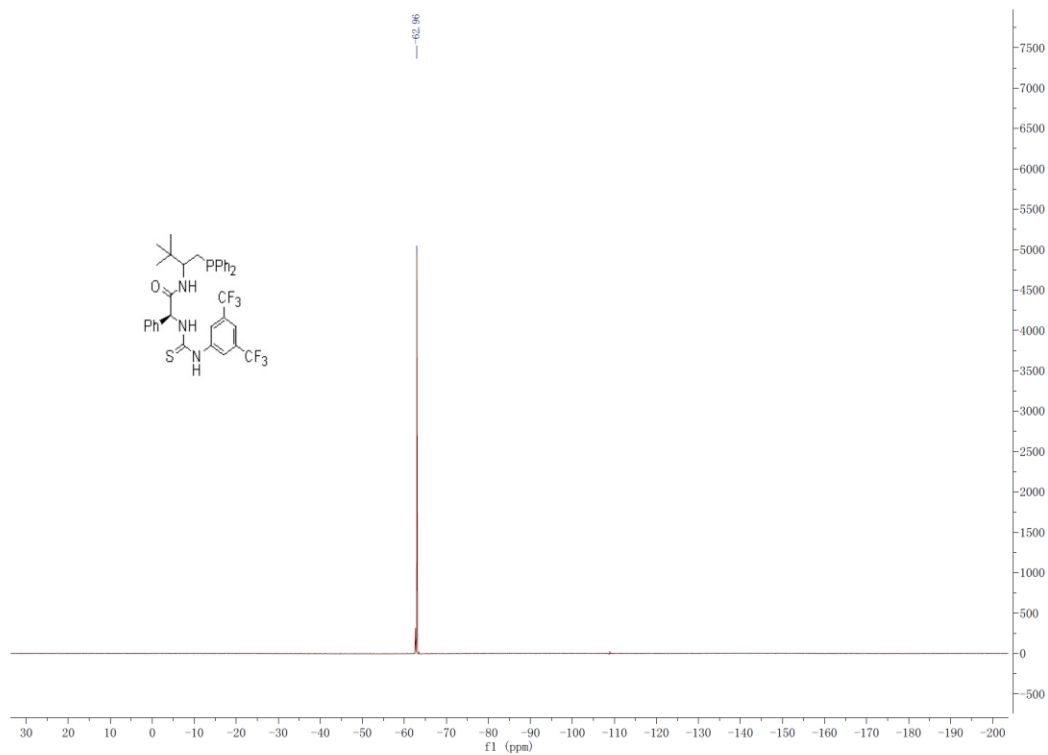


1 Det.A Ch1/220nm

PeakTable

Peak#	Ret. Time	Area	Height	Area %	Height %
1	6.241	181967	8389	94.160	94.166
2	6.925	11286	520	5.840	5.834
Total		193254	8908	100.000	100.000





VI. Crystal structure data for 3'i

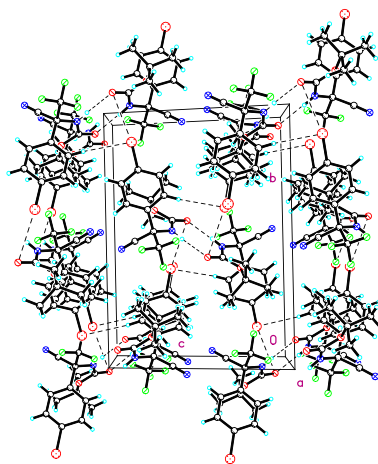
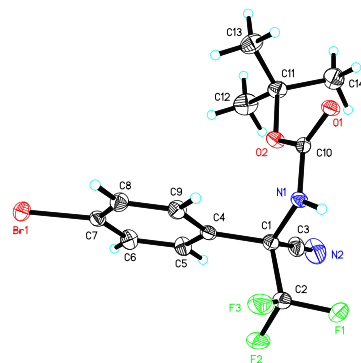


Table 1. Crystal data and structure refinement for mo_d8v19305_0m.

Identification code	mo_d8v19305_0m	
Empirical formula	C14 H14 Br F3 N2 O2	
Formula weight	379.18	
Temperature	193(2) K	
Wavelength	0.71073 Å	
Crystal system	Monoclinic	
Space group	P 21/n	
Unit cell dimensions	a = 9.2263(4) Å	$\alpha = 90^\circ$.
	b = 15.7424(7) Å	$\beta = 108.325(2)^\circ$.
	c = 11.8768(6) Å	$\gamma = 90^\circ$.
Volume	1637.56(13) Å ³	
Z	4	
Density (calculated)	1.538 Mg/m ³	
Absorption coefficient	2.547 mm ⁻¹	
F(000)	760	
Crystal size	0.170 x 0.150 x 0.120 mm ³	
Theta range for data collection	2.588 to 25.998°.	
Index ranges	-11 ≤ h ≤ 11, -19 ≤ k ≤ 19, -12 ≤ l ≤ 14	
Reflections collected	16250	
Independent reflections	3207 [R(int) = 0.0356]	
Completeness to theta = 25.242°	99.6 %	
Absorption correction	Semi-empirical from equivalents	
Max. and min. transmission	0.7456 and 0.4692	
Refinement method	Full-matrix least-squares on F ²	
Data / restraints / parameters	3207 / 0 / 207	
Goodness-of-fit on F ²	1.040	
Final R indices [I > 2σ(I)]	R1 = 0.0291, wR2 = 0.0706	
R indices (all data)	R1 = 0.0389, wR2 = 0.0753	
Extinction coefficient	0.0023(7)	
Largest diff. peak and hole	0.521 and -0.344 e.Å ⁻³	

Table 2. Atomic coordinates ($\times 10^4$) and equivalent isotropic displacement parameters ($\text{\AA}^2 \times 10^3$) for mo_d8v19305_0m. U(eq) is defined as one third of the trace of the orthogonalized U^{ij} tensor.

	x	y	z	U(eq)
Br(1)	11226(1)	8777(1)	8432(1)	38(1)
F(1)	8340(2)	3917(1)	6905(2)	60(1)
F(2)	10260(2)	4619(1)	8014(2)	59(1)
F(3)	9302(2)	4932(1)	6168(2)	53(1)
O(1)	4050(2)	5749(1)	5563(1)	30(1)
O(2)	5422(2)	6197(1)	7403(1)	31(1)
N(1)	6500(2)	5346(1)	6412(2)	26(1)
N(2)	7320(3)	4646(2)	9273(2)	63(1)
C(1)	7867(2)	5323(1)	7424(2)	29(1)
C(2)	8958(3)	4691(2)	7115(3)	43(1)
C(3)	7550(3)	4955(2)	8494(2)	41(1)
C(4)	8654(2)	6194(1)	7707(2)	26(1)
C(5)	9432(3)	6436(2)	8854(2)	34(1)
C(6)	10186(3)	7211(2)	9081(2)	36(1)
C(7)	10140(2)	7733(1)	8146(2)	29(1)
C(8)	9347(3)	7511(1)	6996(2)	34(1)
C(9)	8608(2)	6736(2)	6781(2)	32(1)
C(10)	5216(2)	5770(1)	6405(2)	25(1)
C(11)	4143(3)	6661(2)	7639(2)	34(1)
C(12)	4901(3)	7011(2)	8865(2)	54(1)
C(13)	3612(3)	7369(2)	6743(2)	46(1)
C(14)	2889(3)	6033(2)	7628(3)	52(1)

Table 3. Bond lengths [Å] and angles [°] for mo_d8v19305_0m.

Br(1)-C(7)	1.899(2)
F(1)-C(2)	1.335(3)
F(2)-C(2)	1.338(3)
F(3)-C(2)	1.317(3)
O(1)-C(10)	1.218(2)
O(2)-C(10)	1.324(3)
O(2)-C(11)	1.488(3)
N(1)-C(10)	1.357(3)
N(1)-C(1)	1.444(3)
N(1)-H(1)	0.84(2)
N(2)-C(3)	1.123(3)
C(1)-C(3)	1.507(3)
C(1)-C(4)	1.538(3)
C(1)-C(2)	1.539(3)
C(4)-C(5)	1.379(3)
C(4)-C(9)	1.382(3)
C(5)-C(6)	1.388(3)
C(5)-H(5)	0.9500
C(6)-C(7)	1.371(3)
C(6)-H(6)	0.9500
C(7)-C(8)	1.376(3)
C(8)-C(9)	1.382(3)
C(8)-H(8)	0.9500
C(9)-H(9)	0.9500
C(11)-C(12)	1.508(4)
C(11)-C(13)	1.511(4)
C(11)-C(14)	1.519(3)
C(12)-H(12A)	0.9800
C(12)-H(12B)	0.9800
C(12)-H(12C)	0.9800
C(13)-H(13A)	0.9800
C(13)-H(13B)	0.9800
C(13)-H(13C)	0.9800
C(14)-H(14A)	0.9800
C(14)-H(14B)	0.9800
C(14)-H(14C)	0.9800

C(10)-O(2)-C(11)	121.31(17)
C(10)-N(1)-C(1)	123.38(19)
C(10)-N(1)-H(1)	115.5(17)
C(1)-N(1)-H(1)	119.8(17)
N(1)-C(1)-C(3)	111.12(18)
N(1)-C(1)-C(4)	112.88(18)
C(3)-C(1)-C(4)	111.26(19)
N(1)-C(1)-C(2)	106.44(19)
C(3)-C(1)-C(2)	105.5(2)
C(4)-C(1)-C(2)	109.24(17)
F(3)-C(2)-F(1)	107.9(2)
F(3)-C(2)-F(2)	107.9(2)
F(1)-C(2)-F(2)	107.2(2)
F(3)-C(2)-C(1)	111.6(2)
F(1)-C(2)-C(1)	111.2(2)
F(2)-C(2)-C(1)	110.7(2)
N(2)-C(3)-C(1)	177.0(3)
C(5)-C(4)-C(9)	119.5(2)
C(5)-C(4)-C(1)	121.7(2)
C(9)-C(4)-C(1)	118.8(2)
C(4)-C(5)-C(6)	120.5(2)
C(4)-C(5)-H(5)	119.7
C(6)-C(5)-H(5)	119.7
C(7)-C(6)-C(5)	118.9(2)
C(7)-C(6)-H(6)	120.6
C(5)-C(6)-H(6)	120.6
C(6)-C(7)-C(8)	121.6(2)
C(6)-C(7)-Br(1)	119.58(17)
C(8)-C(7)-Br(1)	118.84(17)
C(7)-C(8)-C(9)	119.0(2)
C(7)-C(8)-H(8)	120.5
C(9)-C(8)-H(8)	120.5
C(8)-C(9)-C(4)	120.5(2)
C(8)-C(9)-H(9)	119.7
C(4)-C(9)-H(9)	119.7
O(1)-C(10)-O(2)	126.13(19)
O(1)-C(10)-N(1)	122.5(2)
O(2)-C(10)-N(1)	111.40(18)

O(2)-C(11)-C(12)	101.98(18)
O(2)-C(11)-C(13)	109.32(18)
C(12)-C(11)-C(13)	111.1(2)
O(2)-C(11)-C(14)	108.98(18)
C(12)-C(11)-C(14)	111.5(2)
C(13)-C(11)-C(14)	113.3(2)
C(11)-C(12)-H(12A)	109.5
C(11)-C(12)-H(12B)	109.5
H(12A)-C(12)-H(12B)	109.5
C(11)-C(12)-H(12C)	109.5
H(12A)-C(12)-H(12C)	109.5
H(12B)-C(12)-H(12C)	109.5
C(11)-C(13)-H(13A)	109.5
C(11)-C(13)-H(13B)	109.5
H(13A)-C(13)-H(13B)	109.5
C(11)-C(13)-H(13C)	109.5
H(13A)-C(13)-H(13C)	109.5
H(13B)-C(13)-H(13C)	109.5
C(11)-C(14)-H(14A)	109.5
C(11)-C(14)-H(14B)	109.5
H(14A)-C(14)-H(14B)	109.5
C(11)-C(14)-H(14C)	109.5
H(14A)-C(14)-H(14C)	109.5
H(14B)-C(14)-H(14C)	109.5

Symmetry transformations used to generate equivalent atoms:

Table 4. Anisotropic displacement parameters ($\text{\AA}^2 \times 10^3$) for mo_d8v19305_0m. The anisotropic displacement factor exponent takes the form: $-2\pi^2 [h^2 a^{*2} U^{11} + \dots + 2 h k a^* b^* U^{12}]$

	U^{11}	U^{22}	U^{33}	U^{23}	U^{13}	U^{12}
Br(1)	40(1)	28(1)	46(1)	-9(1)	10(1)	-3(1)
F(1)	44(1)	28(1)	92(1)	-9(1)	2(1)	5(1)
F(2)	32(1)	49(1)	78(1)	-5(1)	-9(1)	13(1)
F(3)	42(1)	56(1)	64(1)	-12(1)	22(1)	9(1)
O(1)	22(1)	33(1)	31(1)	-6(1)	2(1)	4(1)
O(2)	26(1)	40(1)	28(1)	-8(1)	8(1)	2(1)
N(1)	22(1)	29(1)	26(1)	-6(1)	5(1)	1(1)
N(2)	60(2)	70(2)	50(2)	19(1)	4(1)	-14(1)
C(1)	25(1)	29(1)	30(1)	1(1)	2(1)	0(1)
C(2)	29(1)	34(1)	59(2)	-5(1)	1(1)	4(1)
C(3)	35(1)	39(1)	38(2)	9(1)	-3(1)	-7(1)
C(4)	20(1)	29(1)	29(1)	-1(1)	6(1)	2(1)
C(5)	34(1)	38(1)	27(1)	2(1)	7(1)	-4(1)
C(6)	38(1)	41(1)	26(1)	-8(1)	7(1)	-5(1)
C(7)	24(1)	27(1)	36(1)	-7(1)	9(1)	1(1)
C(8)	35(1)	32(1)	31(1)	1(1)	7(1)	0(1)
C(9)	32(1)	33(1)	26(1)	-2(1)	1(1)	-2(1)
C(10)	25(1)	23(1)	28(1)	-1(1)	9(1)	-1(1)
C(11)	35(1)	38(1)	35(1)	-6(1)	18(1)	6(1)
C(12)	66(2)	61(2)	37(2)	-12(1)	20(1)	11(2)
C(13)	57(2)	38(1)	44(2)	-4(1)	20(1)	11(1)
C(14)	44(2)	51(2)	73(2)	-4(1)	38(2)	0(1)

Table 5. Hydrogen coordinates ($\times 10^4$) and isotropic displacement parameters ($\text{\AA}^2 \times 10^{-3}$) for mo_d8v19305_0m.

	x	y	z	U(eq)
H(1)	6400(30)	5006(16)	5850(20)	32(7)
H(5)	9451	6069	9494	41
H(6)	10726	7376	9871	43
H(8)	9309	7885	6360	40
H(9)	8063	6575	5990	39
H(12A)	5252	6541	9425	81
H(12B)	4167	7358	9107	81
H(12C)	5776	7363	8861	81
H(13A)	4492	7709	6721	68
H(13B)	2885	7730	6970	68
H(13C)	3113	7125	5957	68
H(14A)	2408	5827	6817	78
H(14B)	2122	6314	7911	78
H(14C)	3329	5553	8148	78

Table 6. Torsion angles [°] for mo_d8v19305_0m.

C(10)-N(1)-C(1)-C(3)	58.8(3)
C(10)-N(1)-C(1)-C(4)	-67.0(3)
C(10)-N(1)-C(1)-C(2)	173.1(2)
N(1)-C(1)-C(2)-F(3)	60.5(2)
C(3)-C(1)-C(2)-F(3)	178.61(19)
C(4)-C(1)-C(2)-F(3)	-61.7(3)
N(1)-C(1)-C(2)-F(1)	-60.1(3)
C(3)-C(1)-C(2)-F(1)	58.1(3)
C(4)-C(1)-C(2)-F(1)	177.8(2)
N(1)-C(1)-C(2)-F(2)	-179.23(19)
C(3)-C(1)-C(2)-F(2)	-61.1(2)
C(4)-C(1)-C(2)-F(2)	58.6(3)
N(1)-C(1)-C(4)-C(5)	147.1(2)
C(3)-C(1)-C(4)-C(5)	21.4(3)
C(2)-C(1)-C(4)-C(5)	-94.7(3)
N(1)-C(1)-C(4)-C(9)	-34.5(3)
C(3)-C(1)-C(4)-C(9)	-160.19(19)
C(2)-C(1)-C(4)-C(9)	83.7(2)
C(9)-C(4)-C(5)-C(6)	-1.1(3)
C(1)-C(4)-C(5)-C(6)	177.3(2)
C(4)-C(5)-C(6)-C(7)	0.3(3)
C(5)-C(6)-C(7)-C(8)	1.0(3)
C(5)-C(6)-C(7)-Br(1)	-177.73(17)
C(6)-C(7)-C(8)-C(9)	-1.3(3)
Br(1)-C(7)-C(8)-C(9)	177.40(17)
C(7)-C(8)-C(9)-C(4)	0.4(3)
C(5)-C(4)-C(9)-C(8)	0.8(3)
C(1)-C(4)-C(9)-C(8)	-177.7(2)
C(11)-O(2)-C(10)-O(1)	5.1(3)
C(11)-O(2)-C(10)-N(1)	-175.52(17)
C(1)-N(1)-C(10)-O(1)	-176.48(19)
C(1)-N(1)-C(10)-O(2)	4.1(3)
C(10)-O(2)-C(11)-C(12)	178.2(2)
C(10)-O(2)-C(11)-C(13)	-64.1(3)
C(10)-O(2)-C(11)-C(14)	60.2(3)

Symmetry transformations used to generate equivalent atoms:

Table 7. Hydrogen bonds for mo_d8v19305_0m [$\text{\AA}$ and $^\circ$].

D-H...A	d(D-H)	d(H...A)	d(D...A)	$\angle(\text{DHA})$
C(14)-H(14A)...O(1)	0.98	2.44	3.002(3)	116.1
C(13)-H(13C)...O(1)	0.98	2.43	2.997(3)	116.3
C(13)-H(13B)...Br(1)#1	0.98	3.12	4.070(3)	163.2
N(1)-H(1)...O(1)#2	0.84(2)	1.99(3)	2.826(2)	174(2)

Symmetry transformations used to generate equivalent atoms:

#1 $x-1, y, z$ #2 $-x+1, -y+1, -z+1$