

Supporting Information

## **Visualization of Slide-ring Effect: A Study on Movable Cross-Linking Point Using Mechanochromism**

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## Materials

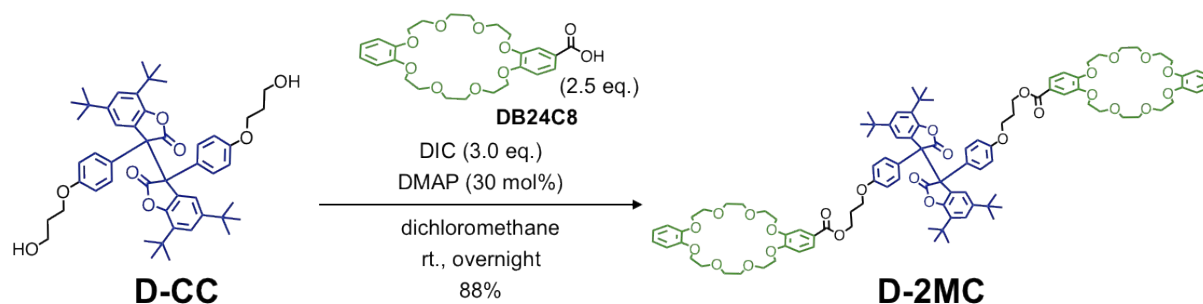
All solvents and reagents from Sigma-Aldrich, Wako Pure Chemical Industries, Tokyo Chemical Industry, and Kanto Chemical were used as received, unless otherwise noted. **D-CC**,<sup>[1]</sup> **DB24C8**,<sup>[2]</sup> **Axle**<sup>[3]</sup> were prepared according to previously published methods. Benzoyl peroxide (BPO) was reprecipitated with chloroform / methanol (1 / 3) at room temperature and dried in vacuo. Methyl acrylate (MA) monomer was passed through an ammonia column to remove the inhibitor. Dry dichloromethane was obtained by distillation over CaH<sub>2</sub> after being washed with water.

## Instruments

<sup>1</sup>H and <sup>13</sup>C NMR spectroscopic measurements were carried out using a Bruker AVANCE III HD500 spectrometer with tetramethylsilane (TMS) as an internal standard in chloroform-*d* (CDCl<sub>3</sub>), acetone-*d*<sub>6</sub>, DMSO-*d*<sub>6</sub>, or dichloromethane-*d*<sub>2</sub> (CD<sub>2</sub>Cl<sub>2</sub>). Tensile tests were carried out with dumbbell shaped samples (12 mm × 2 mm × 0.4 mm) at room temperature using a SHIMADZU EZ graph equipped with a 50 N load cell at elongation rate of 10 mm / min. The gel permeation chromatography (GPC) was performed at 30 °C in DMF (with 10 mM LiBr, 0.700 mL / min) using a JASCO PU-1580 system equipped with a set of a TOSOH TSK G2500H and a TOSOH TSK G4000H with a JASCO RI-1530 detector. The number average molecular weight (*M<sub>n</sub>*), weight average molecular weight (*M<sub>w</sub>*), and polydispersity index (*M<sub>w</sub>* / *M<sub>n</sub>*) of the polymers were calculated on the basis of a polystyrene calibration. IR spectra were recorded on a JASCO FT/IR-230 spectrometer. Melting points were measured on an RFS-10 (Round Science Inc.) instrument. FAB-MS spectra were obtained at the Center for Advanced Material Analysis, Tokyo Institute of Technology on request. In situ electron paramagnetic resonance (EPR) measurements were carried out on a JEOL JES-X320 X-band EPR spectrometer equipped with a tensile tester for long strip specimens (ca. 100 mm × 2 mm × 0.4 mm). The effective measuring range was 43.5 mm height. The specimens were stretched to 40, 80, 120, 160, 200, and 230% strains in sequence under a strain rate of 100 mm/min. The spectra were measured at each strain at room temperature using a microwave power of 0.4 mW and field modulation of 0.07 mT with time constant of 0.03 s and a sweep rate of 250 mT/s. The concentration of the radicals formed from the cleavage of DABBF was determined by comparing the area of the observed integral spectrum with a 0.001 mM solution of TEMPOL in benzene, which was placed in a 3 mm glass capillary, degassed, and measured under the same experimental conditions as the samples. The Mn<sup>2+</sup> signal was used as an auxiliary standard. The g value was determined as 2.003, which is a typical value for carbon centered radicals.



## Synthesis of D-2MC

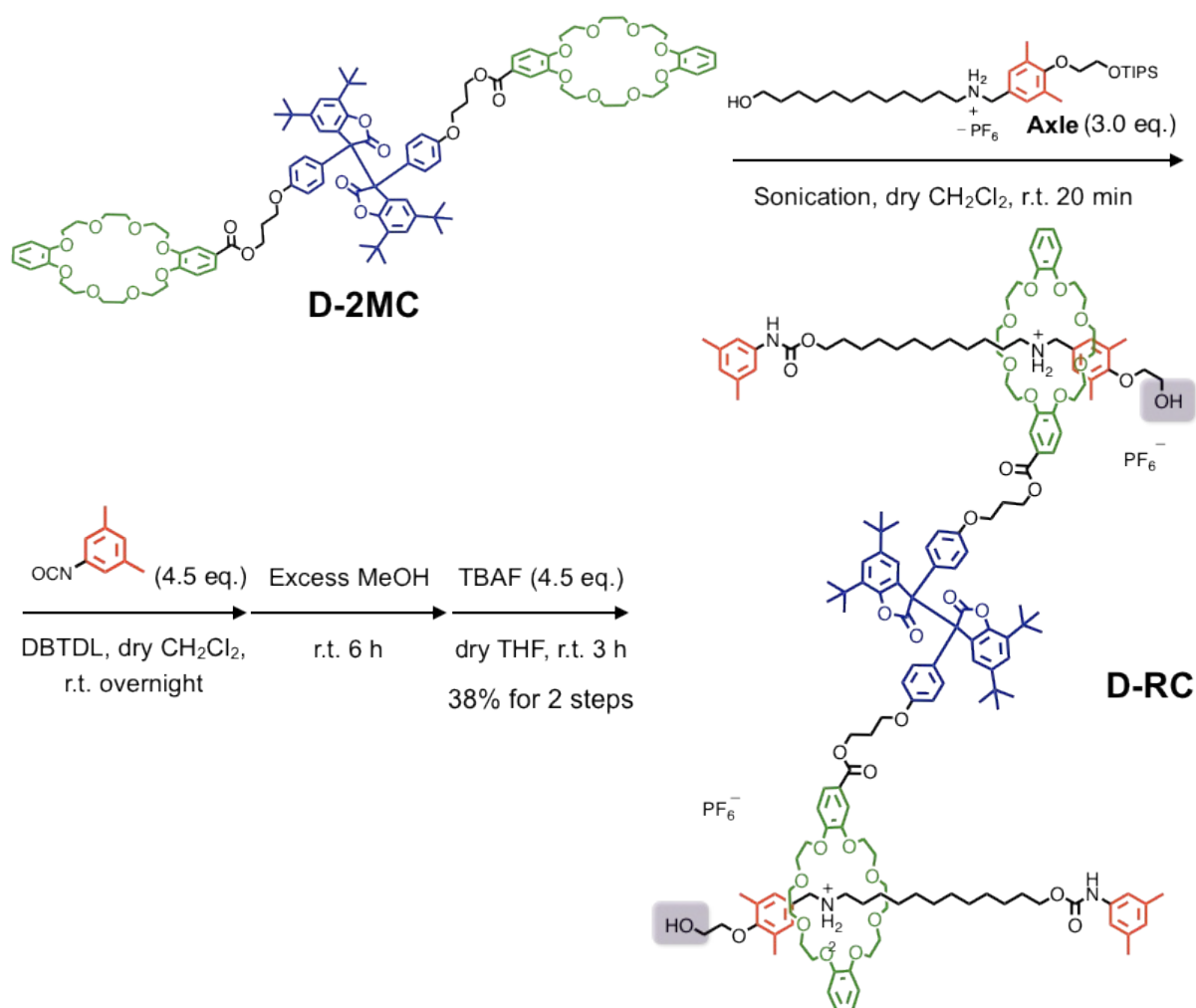


**Scheme S1.** Synthesis of **D-2MC**.

**D-CC** (5.3 g, 6.7 mmol) was mixed in dry dichloromethane (52 mL) with **DB24C8** (8.2 g, 17 mmol, 2.5 eq.) and 4-dimethylaminopyridine (DMAP, 0.38 g, 2.0 mmol, 30 mol%). After degassing followed by charging with argon 3 times, *N,N'*-diisopropylcarbodiimide (DIC, 3.1 mL, 2.5 g, 20 mmol, 3.0 eq.) was added dropwise to the mixture at 0 °C. The mixture was then allowed to warm up to room temperature and react overnight. The residue was washed with 0.1 M HCl aqueous solution once, and water 3 times. The organic layer was concentrated in vacuo. 30 mL dichloromethane was added and the suspension was filtered by a Buchner filter. The filter residue was washed with diethyl ether and the filtrate was concentrated. The residue was purified by column chromatography (silica gel, THF/dichloromethane = 15/85, v/v) to obtain a white solid **D-2MC** (10 g, 5.9 mmol, 88%).

m.p. 81.3–84.9 °C; FAB-MS (*m/z*): calcd for  $[M + Na]^+$ ,  $C_{100}H_{122}O_{26}Na$ , 1761.8122; found, 1761.7970;  $^1H$  NMR (500 MHz, 298 K,  $CD_2Cl_2$ ):  $\delta$  7.67 (d,  $J$  = 10 Hz, 1H), 7.56 (s, 1H), 7.10–7.42 (m, 4H), 6.82–6.98 (m, 7H), 4.48 (t,  $J$  = 6 Hz, 2H), 4.08–4.21 (m, 10H), 3.88–3.96 (m, 8H), 3.80–3.86 (m, 8H), 2.25 (m, 2H), 1.04–1.40 (m, 18H);  $^{13}C$  NMR (125 MHz, 298 K,  $CD_2Cl_2$ ):  $\delta$  166.02, 159.07, 153.03, 149.03, 149.00, 148.33, 133.38, 132.22, 124.21, 123.72, 122.93, 121.35, 121.32, 114.47, 114.20, 114.16, 113.13, 112.09, 71.21, 71.12, 71.05, 69.84, 69.67, 69.51, 69.42, 69.19, 69.11, 64.67, 61.57, 34.63, 34.26, 31.25, 29.40, 28.76, 22.01, 20.61; FT-IR (NaCl,  $cm^{-1}$ ): 2956, 2871, 1793, 1712, 1602, 1508, 1456, 1429, 1363, 1272, 1256, 1215, 1130, 1088, 1052, 995, 929, 821, 764.

## Synthesis of D-2MC



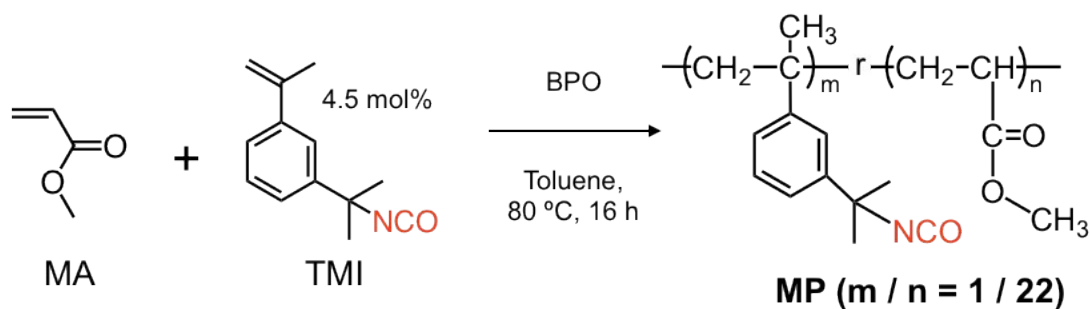
**Scheme S2. Synthesis of D-RC.**

In 13 mL dry dichloromethane, **D-2MC** (9.2 g, 5.3 mmol) was mixed with **Axle** (11 g, 16 mmol, 3.0 eq.). The mixture was sonicated for 20 min. To the solution were added 3,5-dimethylphenyl isocyanate (3.5 g, 3.4 mL, 24 mmol, 4.5 eq.) and 3 drops of dibutyltin dilaurate (DBTDL). The mixture was stirred overnight at room temperature under argon atmosphere and quenched by mixing with excess methanol for 6 hours. The solvent was removed in vacuum. To the residue was added 100 mL THF. Tetra-*n*-butylammonium fluoride (TBAF) in THF solution (1 mol/L, 24 mL, 4.5 eq.) was dropwise added to resulting solution at 0 °C. Then the mixture was allowed to warm to room temperature and react for 3 hours. The resultant mixture was poured into ethyl acetate and washed with ammonium chloride aqueous solution. After being dried upon  $\text{MgSO}_4$ , the solution was concentrated in vacuo and then purified by column

chromatography (silica gel, ethyl acetate  $\rightarrow$   $\text{CHCl}_3/\text{MeOH} = 100/3$ , v/v) to afford a white solid **D-RC** (6.2 g, 2.0 mmol, 38% for 2 steps).

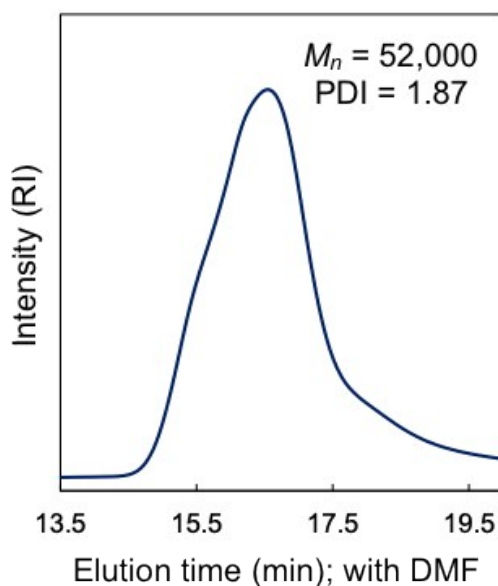
m.p. 104.5–106.7 °C; FAB-MS ( $m/z$ ): calcd for  $[\text{M} - \text{H}]^+$ ,  $\text{C}_{164}\text{H}_{223}\text{N}_4\text{O}_{34}$ , 2792.5844; found, 2792.5800;  $^1\text{H}$  NMR (500 MHz, 298 K,  $\text{CD}_2\text{Cl}_2$ ):  $\delta$  7.55 (dd,  $J = 3$  Hz, 1H), 7.37 (d,  $J = 2$  Hz, 1H), 7.32–6.68 (m, 14H), 6.61 (s, 1H), 6.56 (s, 1H), 4.43–4.34 (m, 4H), 4.16–3.95 (m, 12H), 3.88–3.68 (m, 10H), 3.66–3.60 (m, 8H), 3.58–3.47 (m, 4H), 3.17 (t,  $J = 15$  Hz, 2H), 2.21–2.13 (m, 8H), 1.97 (d,  $J = 2$  Hz, 6H), 1.56 (quint,  $J = 7$  Hz, 2H), 1.35–0.95 (m, 36H);  $^{13}\text{C}$  NMR (125 MHz, 298 K,  $\text{CD}_2\text{Cl}_2$ ):  $\delta$  159.05, 156.04, 153.59, 151.47, 148.97, 147.35, 147.33, 147.20, 138.71, 138.06, 133.42, 131.22, 130.01, 127.51, 124.87, 123.98, 123.41, 121.69, 116.24, 113.16, 113.03, 112.37, 111.50, 73.31, 70.95, 70.39, 70.34, 70.19, 68.38, 68.27, 68.08, 68.05, 65.11, 64.70, 62.05, 61.91, 51.91, 48.90, 34.26, 31.24, 29.48, 29.46, 29.38, 29.28, 29.23, 28.95, 28.69, 26.72, 26.44, 25.81, 21.09, 16.02; FT-IR (NaCl,  $\text{cm}^{-1}$ ): 2928, 2871, 1793, 1714, 1604, 1557, 1508, 1457, 1270, 1215, 1127, 1090, 1056, 953, 843, 764.

## Synthesis of MP

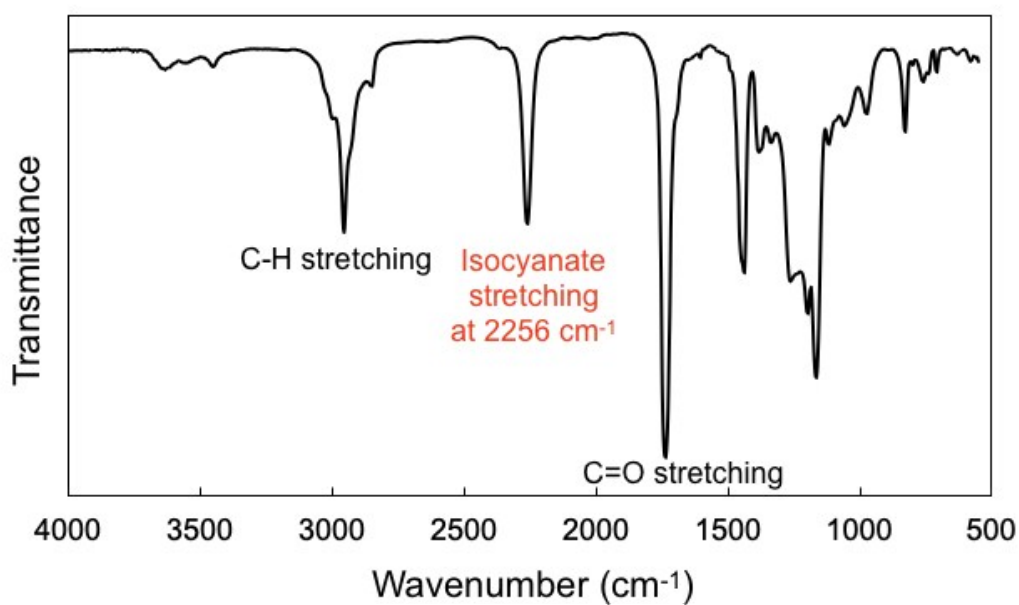


**Scheme S3.** Synthesis of MP.

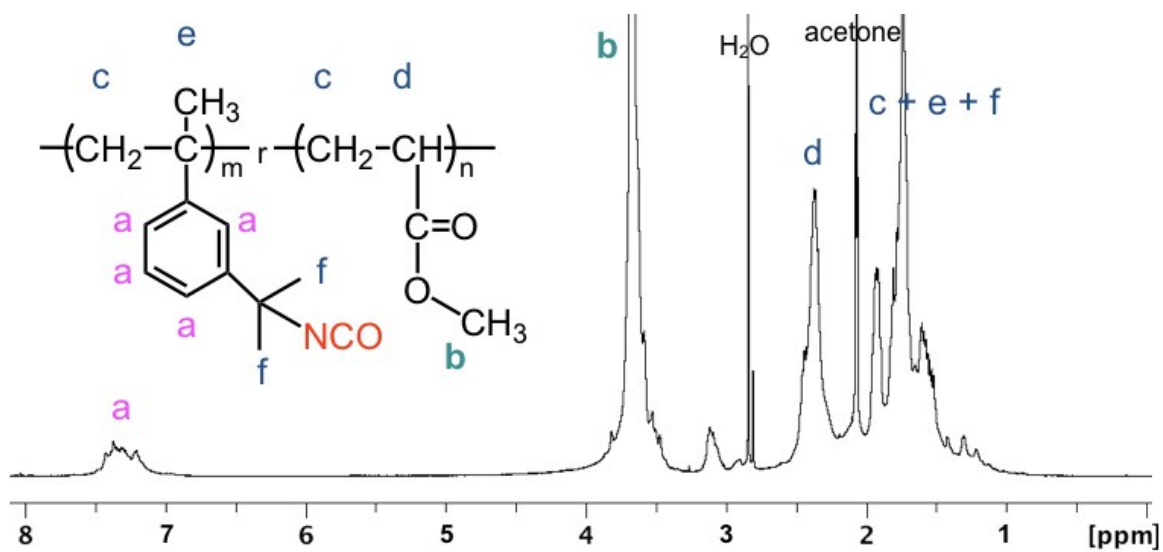
Methyl acrylate (MA, 5 mL, 0.06 mol), 3-isopropenyl- $\alpha,\alpha$ -dimethylbenzene isocyanate (TMI, 0.5 mL, 0.003 mol), benzoyl peroxide (BPO, 0.05 g, 0.2 mmol) were mixed in dry toluene (10 mL). The system was degassed with freeze-pump-thaw cycles and then heated at 80 °C for 16 h. The resulting solution was reprecipitated twice with dry  $\text{CHCl}_3$  and hexane, and dried in vacuum to afford a white solid (5.0 g, 95%).



**Figure S1.** GPC profile of MP.



**Figure S2.** IR spectrum of **MP** (NaCl).



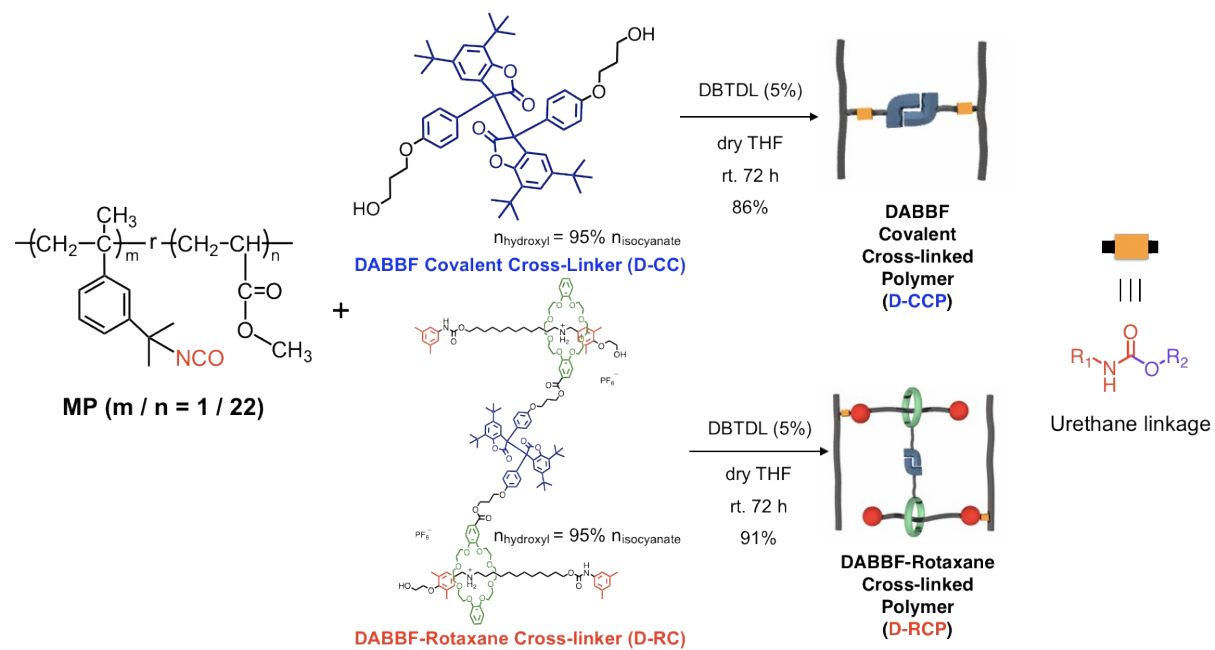
**Figure S3.** <sup>1</sup>H NMR spectrum of **MP** in acetone-*d*<sub>6</sub> (500 MHz, 25 °C).

**Table S1.** Analysis of  $^1\text{H}$  NMR spectrum of **MP**

Symbol	Origin	Integration value ( $^1\text{H}$ NMR)	Integration (a / b)	Proton dosage value (mmol)	Dosage ratio (a / b)
a	TMI and BPO	set as 1.00	$0.074 \pm 0.003$	12.0	0.073
b	MA	$13.6 \pm 0.4$		165	

From above **Table S1**, the radical polymerization proceeded quantitatively. Therefore, the composition of **MA** and **TMI** in **MP** was calculated based on dosage amount. That is, TMI, the isocyanate containing unit, accounted for 4.3% and MA, account for 95.7% of the polymer chain. The dosage ratio of cross-linking reaction was based on this result.

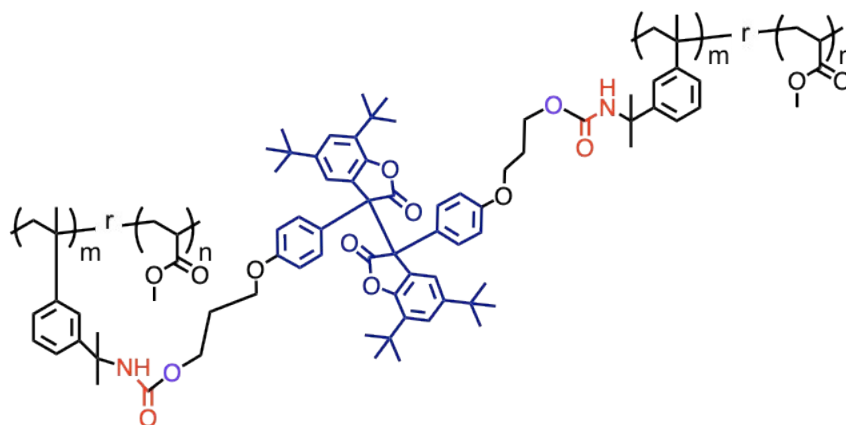
## Synthesis of D-RCP and D-CCP



**Scheme S4.** Cross-linking reactions to give **D-CCP** and **D-RCP**.

## Synthesis of **D-CCP**

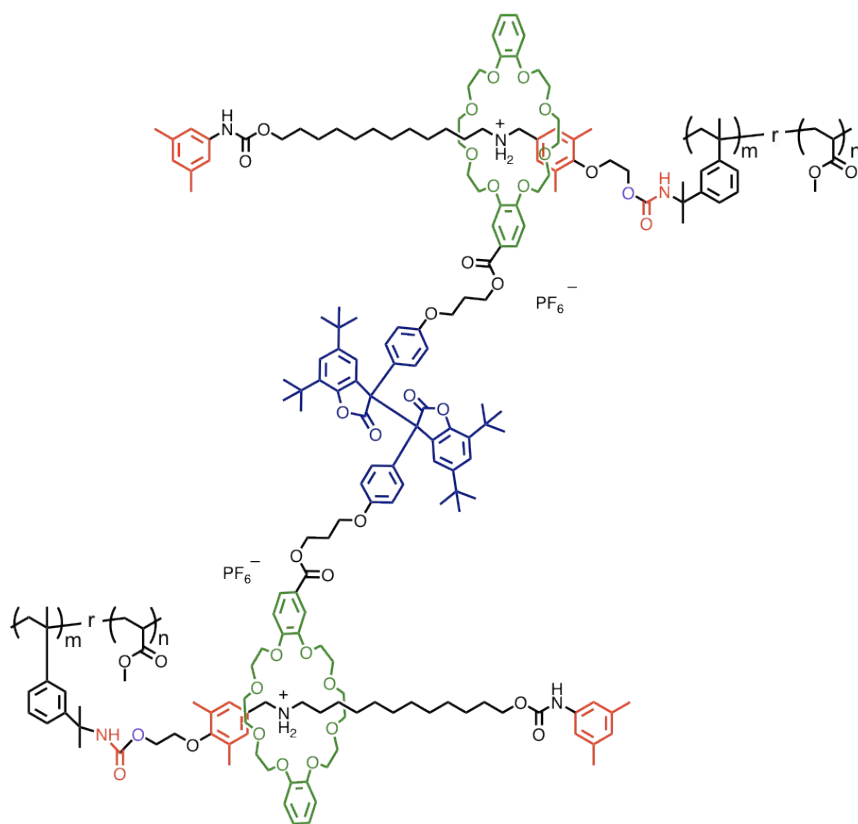
In a Teflon plate, polymer matrix **MP** (2.6 g, containing 1.2 mmol -NCO), **D-CC** (0.46 g, 0.58 mmol), and a catalytic amount of DBTDL were mixed in dry THF to give a homogeneous solution. The solution was allowed to stand for 3 days at room temperature in an argon-charged closed system. Then the solvent was allowed to evaporate naturally at room temperature. The residue after evaporation was taken from the mold and washed with ethyl acetate/methanol (1/1, v/v) solution at 0 °C to afford a white film of **D-CCP** (2.6 g, 86%).



**Figure S4.** Detailed structure of **D-CCP**.

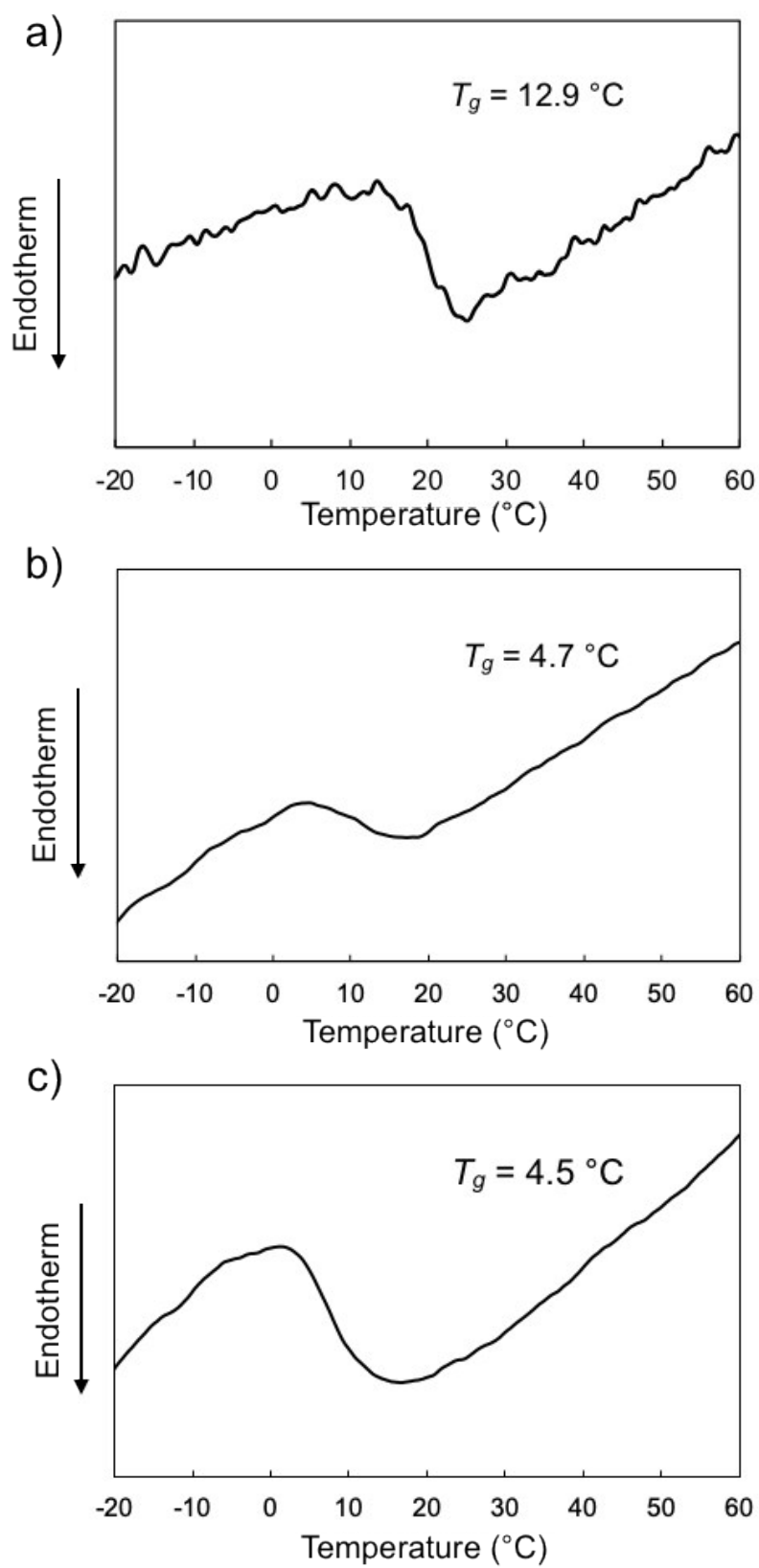
## Synthesis of **D-RCP**

In a Teflon plate, polymer matrix **MP** (1.7 g, containing 0.80 mmol -NCO), **D-RC** (1.2 g, 0.38 mmol), and a catalytic amount of DBTDL were mixed in dry THF to give a homogeneous solution. The solution was allowed to stand for 3 days at room temperature in an argon-charged closed system. Then the solvent was allowed to evaporate naturally at room temperature. The residue after evaporation was taken out from the mold and washed with acetate/methanol (1/1, v/v) mixed solution at 0 °C to afford a white film of **D-RCP** (2.6 g, 91%).



**Figure S5.** Detailed structure of **D-RCP**.

## Thermophysical properties of MP, D-CCP, and D-RCP



**Figure S6.** DSC charts of (a) **MP**, (b) **D-RCP** and (c) **D-CCP**.

## Results of swelling test and tensile test

Swelling test:

Swelling experiments were performed with 5.0–10 mg of cross-linked polymers in MeOH, THF, and CHCl<sub>3</sub> (ca. 5.0 mL) at 0 °C, to avoid the reorganization of the DABBF skeleton.<sup>[1]</sup> The swelling ratio was defined as the difference in weight of the swollen gel ( $W_{\text{swollen}}$ ) vs, the dried gel ( $W_{\text{dry}}$ ), according to equation (1).

$$\text{Swelling ratio [\%]} = \frac{W_{\text{swollen}} - W_{\text{dry}}}{W_{\text{dry}}} \times 100 \quad (1)$$

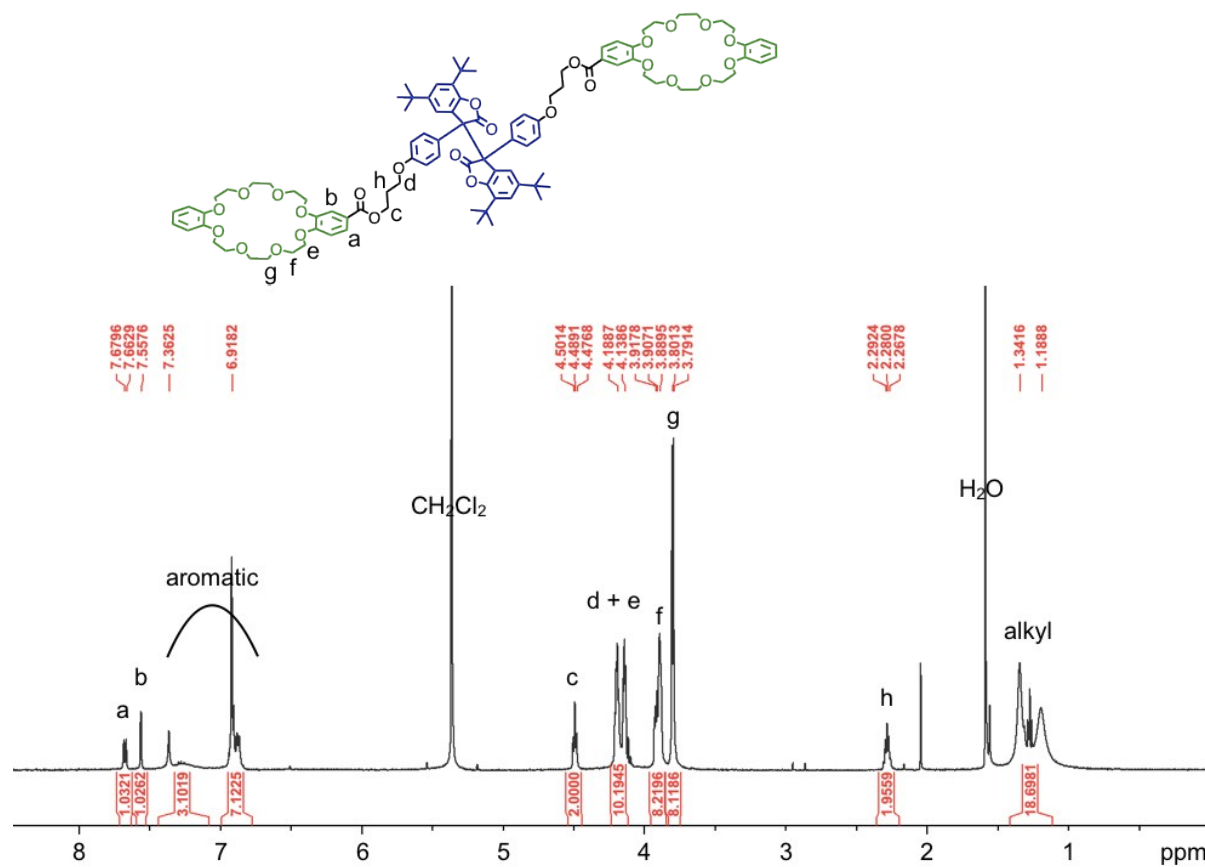
**Table S2.** Swelling ratio and tensile test results

Network polymer	Swelling ratio (%)			Tensile test <sup>a</sup>		
	MeOH	THF	CHCl <sub>3</sub>	Max strain (%)	Young's Modulus (MPa) <sup>b</sup>	Fracture energy (MJ / m <sup>3</sup> )
<b>D-CCP</b>	12 ± 3	418 ± 31	1281 ± 56	488 ± 32	8.4 ± 0.2	6.77 ± 0.45
<b>D-RCP</b>	13 ± 4	414 ± 22	1314 ± 78	568 ± 28	3.9 ± 0.3	4.41 ± 0.23

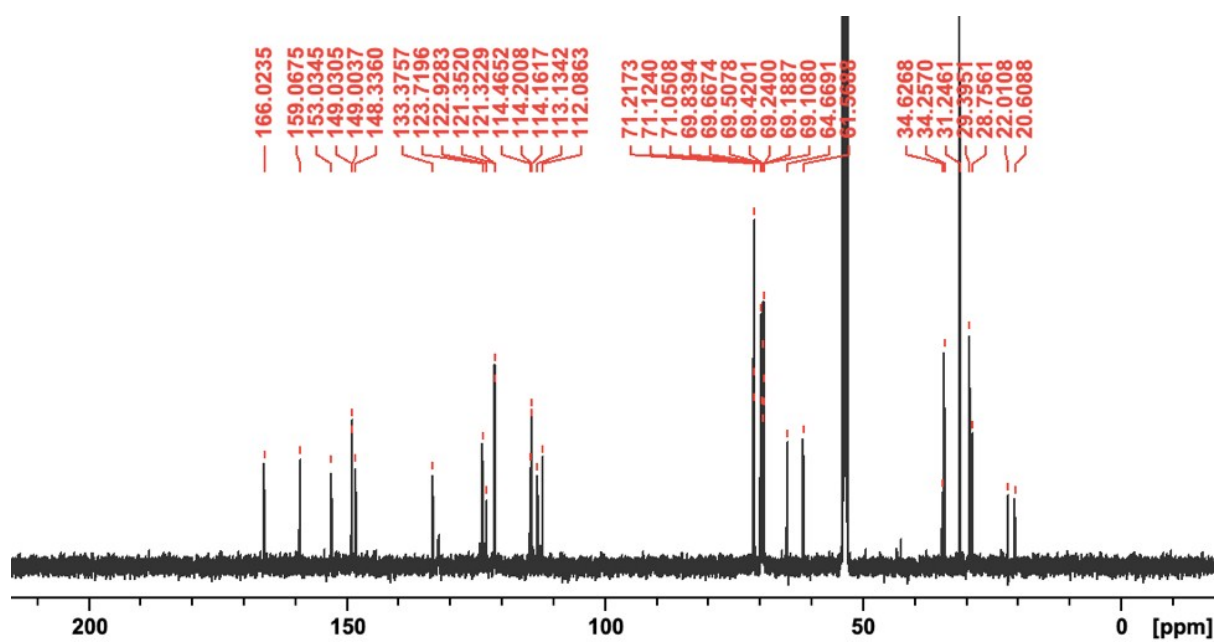
<sup>a</sup> As an elastomer. <sup>b</sup> Determined by the stress between 0 and 8% strain.

## Selected spectra information

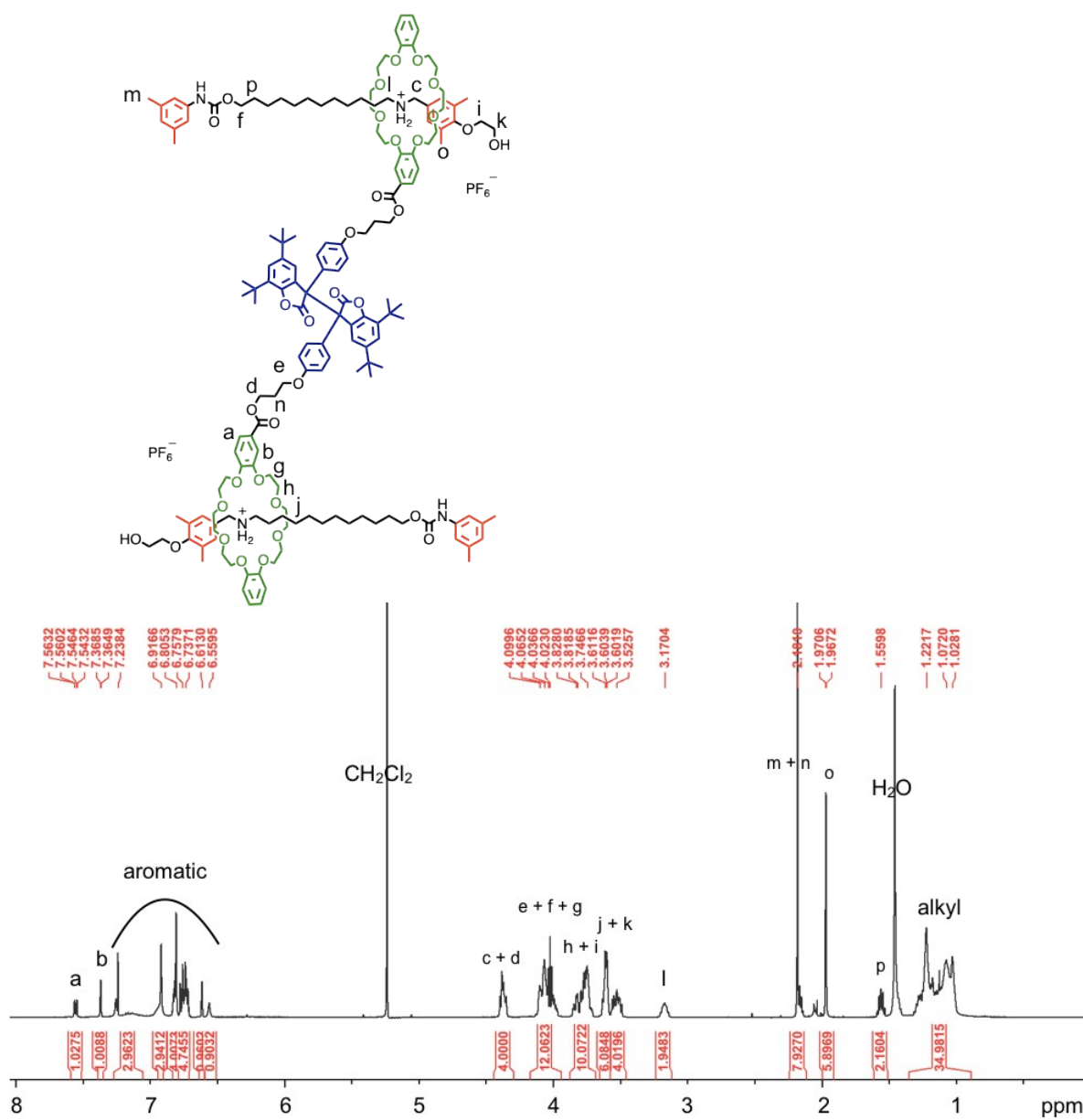
### NMR Spectra



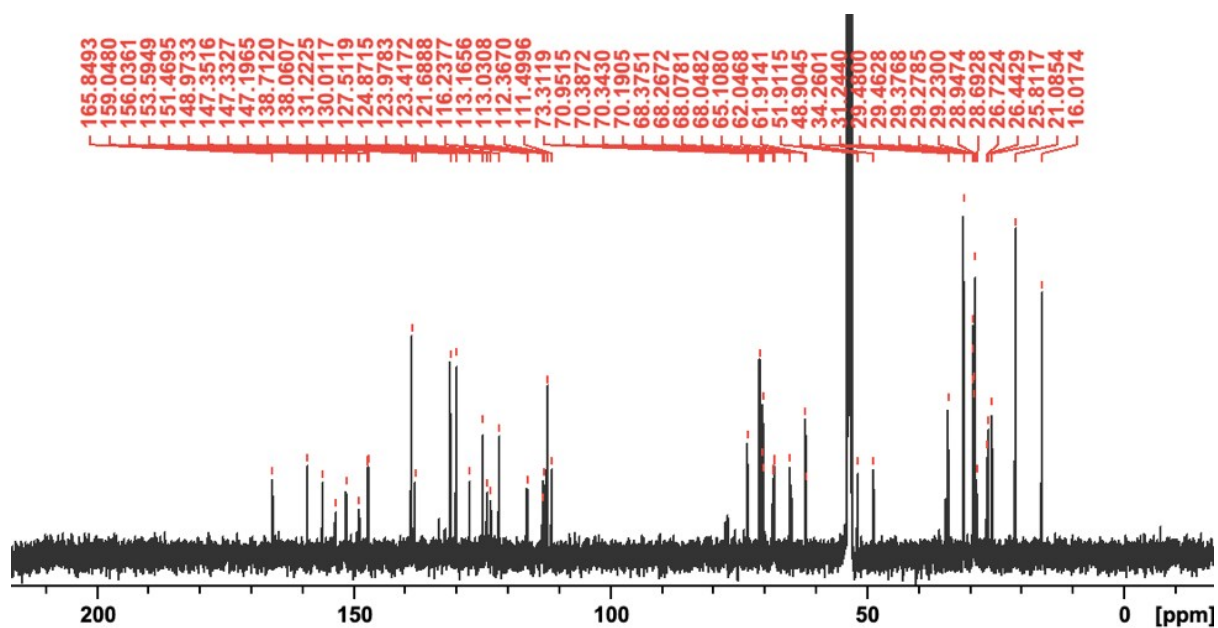
**Figure S7.** <sup>1</sup>H NMR spectrum of **D-2MC** in CD<sub>2</sub>Cl<sub>2</sub> (500 MHz, 25 °C).



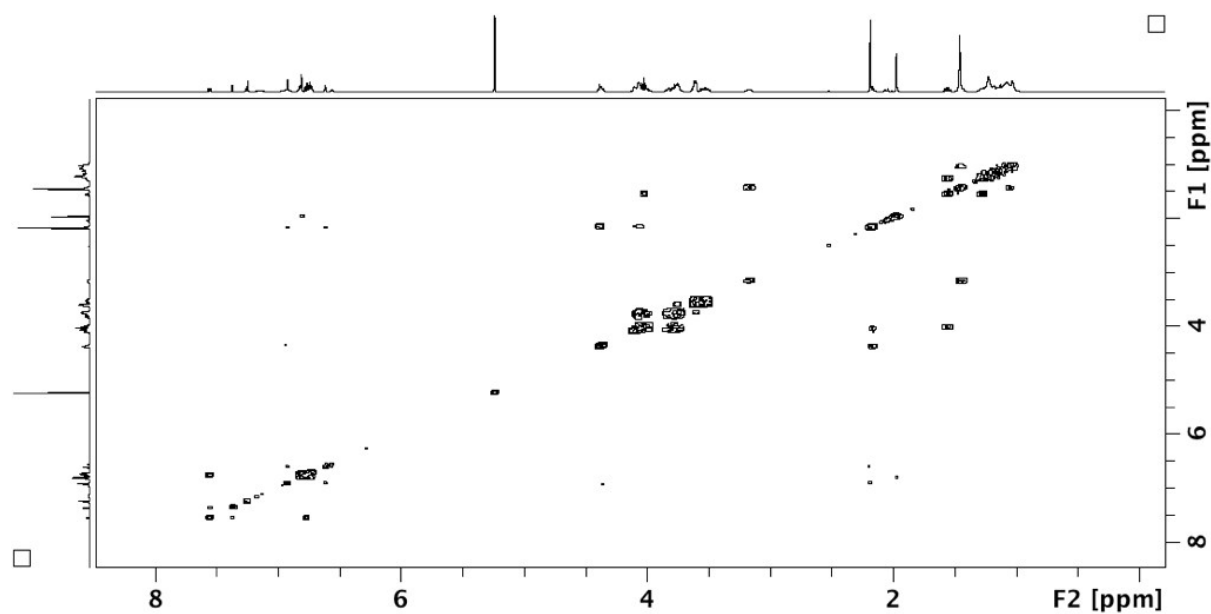
**Figure S8.**  $^{13}\text{C}$  NMR spectrum of **D-2MC** in  $\text{CD}_2\text{Cl}_2$  (125 MHz, 25 °C).



**Figure S9.**  $^1\text{H}$  NMR spectrum of **D-RC** in  $\text{CD}_2\text{Cl}_2$  (500 MHz, 25 °C).

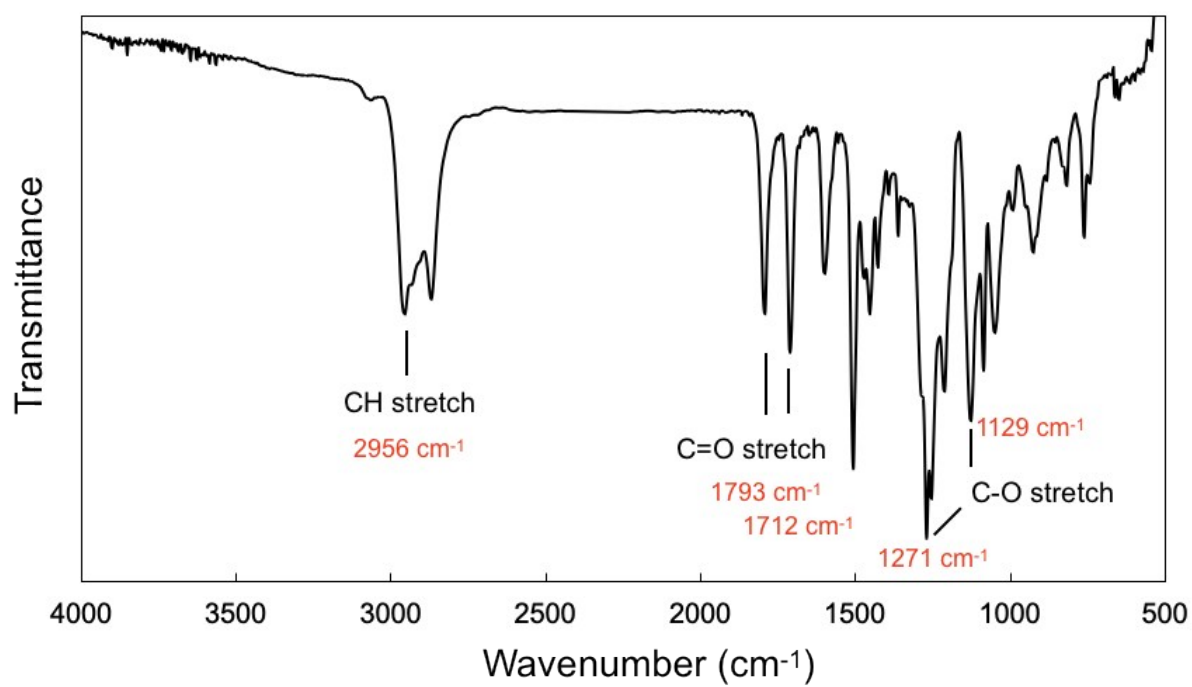


**Figure S10.**  $^{13}\text{C}$  NMR spectrum of **D-RC** in  $\text{CD}_2\text{Cl}_2$  (125 MHz, 25  $^\circ\text{C}$ ).

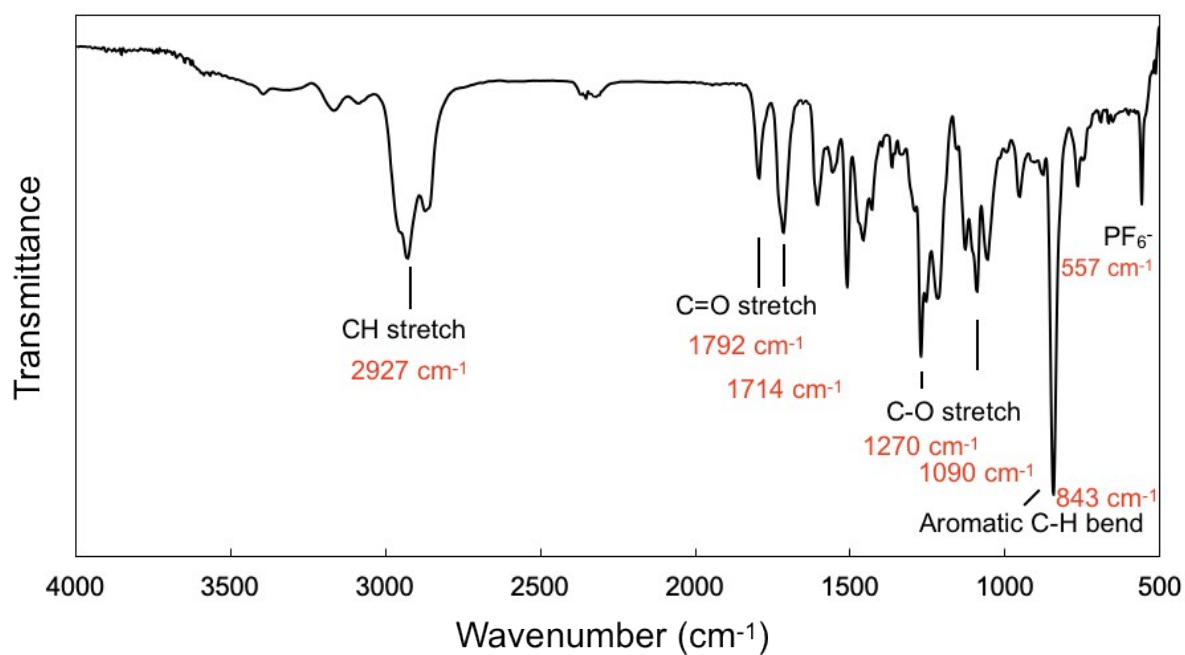


**Figure S11.** 2D H-H COSY NMR spectrum of **D-RC** in  $\text{CD}_2\text{Cl}_2$  (500 MHz, 25  $^\circ\text{C}$ ).

## IR spectra



**Figure S12.** IR spectrum of **D-2MC** (NaCl).



**Figure S13.** IR spectrum of **D-RC** (NaCl).

## References

- [1] K. Imato, A. Irie, T. Kosuge, T. Ohishi, M. Nishihara, A. Takahara, H. Otsuka, *Angew. Chem. Int. Ed.*, 2015, **54**, 6168–6172.
- [2] D. Feng, X. Li, X. Wang, X. Jiang, Z. Li, *Tetrahedron*, 2004, **60**, 6137–6144.
- [3] S. J. Rao, K. Nakazono, X. Liang, K. Nakajima, T. Takata, *Chem. Commun.*, 2019, **55**, 5231–5234.