

Control of polymorphism in thiophene derivates by sublimation-aided nanostructuring

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ELECTRONIC SUPPORTING MATERIAL

Experimental

Thin deposits. ¹⁻⁴ Thin deposits of **1** were prepared by drop casting of 10 μ l of the solution of **1** (1 g/L) (Control samples) and **1** + **2** (**1**=1g/L, **2** =300 g/L) for ASB-SANS samples. The substrates consist of a 10x10 mm² piece of silicon covered by 200 nm of thermal oxides⁵.

X-Ray Diffraction. XRD were performed in specular geometry using a SmartLab-Rigaku diffractometer equipped with a rotating anode (Cu λ_{α} = 1.5405 Å), followed by a parabolic mirror to collimate the incident beam, and a series of variable slits (placed before and after the sample position) to reach an acceptance of 0.01°.

Grazing Incidence X-Ray Diffraction (GIXRD) measurements were performed at the beamline XRD1 of ELETTRA synchrotron facility (Trieste, Italy) by using a wavelength of 1 Å and an incident angle of 0.1°. GIXRD images were recorded for 40 seconds by a 2D camera (Pilatus detector), placed normal to the incident beam direction at 350 mm from the sample.

Index of the peaks were attributed considering the structure published in the references⁶.

Fluorescence microscopy. Fluorescence images were recorded with a Nikon i-80 microscope equipped with epi-fluorescence (FM) using FM filter Nikon Ex 420, DM 435, BA 475 and Ex 535, DM 570, BA 590. The FM images were recorded using a commercial CCD camera (Nikon CCD DS-2Mv). The illumination was performed by a 100 W Hg lamp at fixed power.

Confocal Fluorescence Imaging. Confocal fluorescence imaging was performed on an inverted Nikon Ti-E microscope (Nikon Co., Shinjuku, Japan) using an argon-ion CW laser as well as 405 nm pulsed/CW diode lasers (PicoQuant GmbH, Berlin, Germany). Images were collected using a Nikon Plan Apo VC 20X air objective with NA 0.8. Filters were set to register the fluorescence intensity in the 510-540 nm, 555-615 nm and 665-735 nm ranges. A Nikon A1 spectral module with a precisely corrected 32-PMT array detector was used for spectral imaging. Wavelength resolution was set to 6 nm per PMT array. Spectral images were obtained by exciting the sample at 488 nm in CW mode.

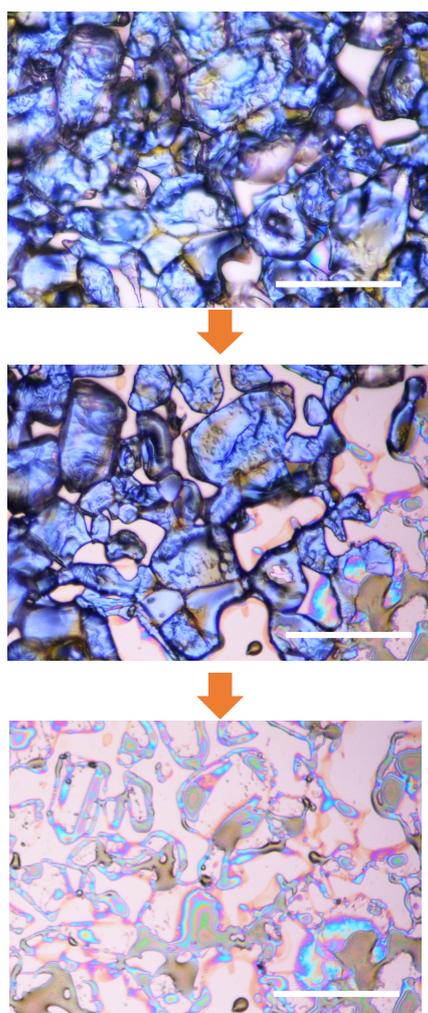


Figure S1. Time laps of the last step of the process: Sublimant compound sublimation

1. M. Cavallini and F. Biscarini, *Review of Scientific Instruments*, 2000, **71**, 4457-4460.
2. M. Cavallini, D. Gentili, P. Greco, F. Valle and F. Biscarini, *Nature Protocols*, 2012, **7**, 1668-1676.
3. P. Leclere, M. Surin, R. Lazzaroni, A. F. M. Kilbinger, O. Henze, P. Jonkheijm, F. Biscarini, M. Cavallini, W. J. Feast, E. W. Meijer and A. Schenning, *J. Mater. Chem.*, 2004, **14**, 1959-1963.
4. D. Gentili, P. Sonar, F. Liscio, T. Cramer, L. Ferlauto, F. Leonardi, S. Milita, A. Dodabalapur and M. Cavallini, *Nano Letters*, 2013, **13**, 3643-3647.
5. D. Gentili, F. Liscio, N. Demitri, B. Schafer, F. Borgatti, P. Torelli, B. Gobaut, G. Panaccione, G. Rossi, A. Degli Esposti, M. Gazzano, S. Milita, I. Bergenti, G. Ruani, I. Salitros, M. Ruben and M. Cavallini, *Dalton Transactions*, 2016, **45**, 134-143.
6. C. Cappuccino, L. Catalano, F. Marin, G. Dushaq, G. Raj, M. Rasras, R. Rezgui, M. Zambianchi, M. Melucci, P. Naumov and L. Maini, *Cryst. Growth Des.*, 2020, **in press**.