Photoinduced Interruption of Interannular Cooperativity for Delivery of Cationic Guests in Water

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SUPPORTING INFORMATION

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General Information

All chemicals were purchased from commercial sources and used as received unless stated otherwise. Basket 1 was synthesized according to the procedure recently reported by our group (see: Chem. Eur. J. 2019, 25, 273-279). NMR experiments were performed with Bruker 600 (5 mm Triple-resonance Inverse (TXI) cryoprobe with Z-Gradients), 700 (5 mm Triple-resonance Observe (TXO) cryoprobe with Z-Gradients) and 850 MHz (5mm Triple-resonance Inverse (TCI) cryoprobe with Z-Gradients) spectrometers. All NMR samples were contained in class B glass NMR tubes (Wilmad Lab Glass). ¹H NMR data were collected using excitation sculpting perfect echo pulse sequence for water suppression. For 2D homonuclear experiments, we typically used a spectral width of 2048×512 complex points in F2 and F1 dimensions. NOESY spectra shown in the main text were recorded using the Bruker zero-quantum suppression sequence noesygpphzs (see Angew. Chem. Int. Ed., 2003, 42, 3938–3941) with minor modifications to improve water suppression and reduce radiation damping at high magnetic fields equipped with cryoprobes. Zero-quantum suppression was achieved by the simultaneous application of a 20 ms adiabatic Chirp pulse with a B1 max of 1.545 kHz and a square gradient of about 0.7 G/cm at the beginning of the NOESY mixing time. Bipolar gradients with a strength of 0.1 G/cm were applied during the indirect acquisition time to suppress radiation damping. A 50 Hz B1 field was used for additional water suppression. ESI-TOF MS was recorded on a Waters Synapt G2 mass spectrometer, using a solution of 1 mg B-Glu and 0.169 mg 1,10-diaminodecane (1:1) in 1 mL water (pH=7, adjusted with ammonium hydroxide).

NMR Titrations



Figure S1. (Top) Water suppression ¹H NMR spectra (850 MHz, 298 K; 30 mM phosphate buffer at pH = 7.0 ± 0.1 , 10% D₂O) of 0.30 mM basket 2^{3-} obtained upon an incremental addition of 2.0, 10.0 and 100 mM solutions of 3^{2+} (30 mM phosphate buffer at pH = 7.0 ± 0.1). The number of molar equivalents of 3^{2+} are shown on the right.



7.7 7.6 7.5 7.4 3.5 3.4 3.3 3.2 3.1 3.0 2.9 2.8 2.7 2.6 2.5 2.4 2.3 2.2 2.1 2.0 1.9 1.8 1.7 1.6 1.5 1.4 1.3 1.2 1.1 1.0 0.9 [pom]



Figure S2. (Top) Water suppression ¹H NMR spectra (600 MHz, 298 K; 30 mM phosphate buffer at pH = 7.0 ± 0.1, 10% D₂O) of 0.50 mM basket 2^{3-} obtained upon an incremental addition of 2.0, 10.0 mM, 78.0 mM solutions of 4^{2+} (30 mM phosphate buffer at pH = 7.0 ± 0.1). The number of molar equivalents of 4^{2+} are shown on the right. (Bottom) Nonlinear least-square analysis of ¹H NMR binding data corresponding to the formation of $[4\subset 2]^-$ complex. The data were fit to 1:1 binding model (SigmaPlot) to give $K_1 = 52 \pm 5 \text{ M}^{-1}$; the residual distribution is shown below the binding isotherm.



Figure S3. (Top) Water suppression ¹H NMR spectra (600 MHz, 298 K; 30 mM phosphate buffer at pH = 7.0 ± 0.1, 10% D₂O) of 0.30 mM basket 2^{3-} obtained upon an incremental addition of 2.0 mM and 33.0 mM solutions of 5^{2+} (30 mM phosphate buffer at pH = 7.0 ± 0.1). The number of molar equivalents of 5^{2+} are shown on the right. (Right) Nonlinear least-square analysis of ¹H NMR binding data corresponding to the formation of [5⊂2]⁻ complex. The data were fit to 1:1 binding model (SigmaPlot) to give $K_1 = 408 \pm 11 \text{ M}^{-1}$; the residual distribution is shown below the binding isotherm.



Figure S4. Water suppression ¹H NMR spectra (600 MHz, 298 K; 30 mM phosphate buffer at pH = 7.0 ± 0.1 , 10% D₂O) of 0.27 mM basket 2^{3-} obtained upon an incremental addition of 2.0, 10.0 and 66.0 mM solutions of 6^{2+} (30 mM phosphate buffer at pH= 7.0 ± 0.1) to this solution. (Right) The number of molar equivalents of 6^{2+} .



Figure S5. A portion of TOF-ESI mass spectrum of basket 1^{6-} and diammonium decane 6^{2+} showing signals corresponding to doubly charged ternary $[6 \subset 1_2]^{2+}$ complexes. The ESI-TOF MS experiment was performed under the following conditions: ESI capillary voltage, 3 kV; sample cone voltage, 30 V; extraction cone voltage, 3.0 V; source temperature 80 °C; desolvation temperature, 80 °C; cone gas flow, 1 L per hour; desolvation gas flow, 700 L per hour (N₂); source gas control, 0 mL per minute; trap gas control, 2 mL per minute; sample flow rate, 5 µL per minute.



Figure S6. Water suppression ¹H NMR spectra (850 MHz, 298 K; 30 mM phosphate buffer at pH=7.0 \pm 0.1, 10% D₂O) of 0.50 mM basket **2**³⁻ and 0.5 mM **6**²⁺obtained upon an incremental addition of **7**³⁻ (3.1 mM solution in 30 mM phosphate buffer at pH=7.0 \pm 0.1). (Right) The molar equivalents of the corresponding components. This experiment was conducted in order to test if one basket could primarily utilize α -carboxylates (**7**³⁻) while another γ -carboxylates (**2**³⁻) for forming of ternary species. With the absence of a dramatic upfield shifting of the guest protons, we concluded that guest **6**²⁺ remained mostly free in solution with baskets **2**³⁻ and **7**³⁻ unable to form ternary complexes.



Figure S7. A steady irradiation (300 nm, Rayonet) of a solution of diammonium octane 4^{2+} (0.42 mM) and basket 1^{6-} (0.86 mM; 30 mM PBS at pH = 7.0 ± 0.1), constituting ternary $[4 \subset 1_2]^{10-}$, was monitored with ¹H NMR spectroscopy (600 MHz, 298 K). The inset shows a change in the normalized chemical shift of H_{1-4} resonances from 4^{2+} as a function of the irradiation time. ¹H NMR resonances labeled with black and red stars correspond to 1^{6-} and 2^{3-} , respectively.