Electronic Supplementary Material (ESI) for CrystEngComm. This journal is © The Royal Society of Chemistry 2019

Supporting information for: Mineralization of Magnetic Nano-Tape in Self-organized Nanospace Composed of Nucleopeptide and Peptide

Kazuki Murai,^{a*} Kaede Inagaki,^b Chisato Hiraoka,^b Sayaka Minoshima,^b Takatoshi Kinoshita,^c Kenji Nagata^b and Masahiro Higuchi^{b*}

 ^a Department of Chemistry and Materials, Faculty of Textile Science and Technology, Shinshu University, 3-15-1 Tokida, Ueda, Nagano 386-8567, Japan.
^b Department of Life Science and Applied Chemistry, Graduate School of Engineering, Nagoya Institute of Technology, Gokiso-cho, Showa-ku, Nagoya, 466-8555, Japan.
^c Nagoya Institute of Technology, Gokiso-cho, Showa-ku, Nagoya, 466-8555, Japan.
* Corresponding authors: murai kazuki@shinshu-u.ac.jp (Dr. K. Murai) and

higuchi.masahiro@nitech.ac.jp (Prof. Dr. M. Higuchi)

Contents

- 1. S1. Synthesis strategy of the nucleopeptide.
- 2. S2. ¹HNMR spectra of the spacer peptide.
- 3. S3. ¹HNMR spectrum of the peptide main chain of the nucleopeptide.
- 4. **S4.** ¹HNMR spectrum of the nucleopeptide.
- **5. S5.** Arrangement of the nucleotide chains in the nanosheet at each molar ratio, and growth mechanism of the nanosheets.
- 6. S6. Deconvolutions of UV-Vis spectra to absorption and scattering components.
- **7. S7.** TEM images of the nucleopeptide/spacer peptide aggregates obtained in the aqueous solution at pH 6.5.
- **8. S8.** Bright field- and dark field (DF)-TEM images and SAED pattern of the nucleopeptide/spacer peptide nanosheet.
- 9. S9. AFM image of the nucleopeptide/spacer peptide nanosheet.
- **10. S10.** EDX mapping data of the magnetite-nucleopeptide/spacer peptide hybrid nanosheet after mineralization.
- **11. S11.** Section analysis profile of the magnetite-nucleopeptide/spacer peptide hybrid nanosheet.

S1. Synthesis strategy of the nucleopeptide.



Figure S1. Synthesis strategy of the nucleopeptide.

S2. ¹HNMR spectra of the spacer peptide.



Intensity ratio of attributed protons.

Figure S2. ¹HNMR spectrum of the spacer peptide, Ac-(VE)₉-CONH₂.

S3. ¹HNMR spectrum of the peptide main chain of the nucleopeptide.



Figure S3. ¹HNMR spectrum of the peptide main chain of the nucleopeptide, Ac-VEVS-(VE)₇-CONH₂.

S4. ¹HNMR spectrum of the nucleopeptide.

		Intensity ratio	
Nucleopeptide		Experimental value	Calculated value
Ac-VEVS(g(GC) ₃)(VE) ₇ -CONH ₂	abegkq : lr	59.5 : 6	56 : 6
8 ^{H₀C−ℓ} −H−CH _d e H−CH _d	—сн—ё—н—сн—ё—_Я—_($ \begin{array}{c c} \mathbf{h} & \mathbf{f} & \mathbf{f} & \mathbf{c} \\ \mathbf{H} & \mathbf{c} \mathbf{H} & \mathbf{c} \mathbf{H} & \mathbf{c} \\ \mathbf{H} & \mathbf{c} \mathbf{H} & \mathbf{c} \mathbf{H} & \mathbf{c} \\ \mathbf{H} & \mathbf{c} \mathbf{H} & \mathbf{c} \mathbf{H} & \mathbf{c} \\ \mathbf{H}^{H_{1}} \mathbf{i} & \mathbf{e} & \mathbf{c} \mathbf{H} & \mathbf{c} \\ \mathbf{e} & \mathbf{c} \mathbf{H} & \mathbf{c} \mathbf{H} & \mathbf{c} \\ \mathbf{H}^{H_{2}} \mathbf{d} & \mathbf{c} \\ \mathbf{H}^{H_{2}} \mathbf{d} & \mathbf{c} \\ \mathbf{c} \mathbf{H}^{H_{2}} \mathbf{d} \\ \mathbf{c} \mathbf$	H2
		нј 	
	NH ₂	d	
	. f b	a b e g k q	
mst	cfh jopv	nu	
10 9 8	7 6 5 4	3 2 1 0 p	pm

Intensity ratio of attributed proton.

Figure S4. ¹HNMR spectrum of the nucleopeptide, Ac-VEVS(g(GC)₃)(VE)₇-CONH₂.

S5. Arrangement of the nucleotide chains in the nanosheet at each molar ratio, and growth mechanism of the nanosheets.



Figure S5. (a); Ideal arrangements nucleopeptide and spacer peptide, which take β -sheet conformation. (b); Schematic pictures of the 3D nucleopeptide/spacer peptide nanosheet, whose molar ratio is 1:5. (c); Schematic diagram for the growth process of the nucleopeptide/spacer peptide nanosheet, whose molar ratio is 1:6. We formed the assemblies on mica substrate (Area; 2.0 × 10⁻⁴ m²) and STEM grid (Area; 5.6 × 10⁻⁵ m²). We prepared the

assemble by incubating each substrate in 5.0 mL (mica system) or 1.5 mL (STEM grid system) of solutions (0.1 mM). The number of molecules of the peptide chains in the solution at the mica and STEM grid systems are 3.0×10^{17} and 9.0×10^{16} , respectively, and the molecular area of the peptide chains is 2.9 nm² per molecule. The total areas of the self-assembled peptide chains on mica substrate and STEM grid, assuming that the peptide chains lay on the surface as double layers, are 0.43 m² and 0.13 m², respectively. These values are large enough compared with the area of each substrate. Hence, the used amounts of the solution in this study do not affect assembly formation on mica substrate and STEM grid.



S6. Deconvolutions of UV-Vis spectra to absorption and scattering components.

Figure S6. UV-vis spectra of the self-assembly composed of the nucleopeptide and spacer peptide in aqueous solutions (a) were separated to absorption spectra based on the base paring (b) and scattering components (c) by peak deconvolution at various pH conditions (pH 6.5, pH 7.0, and pH 7.5), respectively. The molar ratio of the nucleopeptide and spacer peptide was fixed at 1:5.

S7. TEM images of the nucleopeptide/spacer peptide aggregates obtained in the aqueous solution at pH 6.5.



Figure S7. TEM images of the nucleopeptide/spacer peptide aggregates obtained in aqueous solution at the pH 6.5 after (a); 7 days and (b); 16 days incubation at room temperature, respectively. The molar ratio of the nucleopeptide and spacer peptide was fixed at 1:5. An aliquot sample solution that incubated each period was placed on STEM grid. After adsorption for 3 min, the excess solution was removed by absorption onto filter paper.

S8. Bright field- and dark field (DF)-TEM images and SAED pattern of the nucleopeptide/spacer peptide nanosheet.



Figure S8. Bright field- and dark field (DF)-TEM images and SAED pattern of the nucleopeptide/spacer peptide nanosheet formed on STEM grids after 10 days incubation at 15°C under the pH 7.0. Molar ratio of the nucleopeptide and spacer peptide was fixed at 1:5. DF-TEM images show the different domains attributed to each SAED spots (position 1-3).

S9. AFM image of the nucleopeptide/spacer peptide nanosheet.



Figure S9. AFM image of the nucleopeptide/spacer peptide nanosheet formed on mica for 7 days incubation under the pH 7.5 at 15° C. The molar ratio of the nucleopeptide and spacer peptide was fixed at 1:6.

S10. EDX mapping data of the magnetite-nucleopeptide/spacer peptide hybrid nanosheet after mineralization.



Figure S10. TEM and dark field STEM images, and EDX mapping of the magnetitenucleopeptide/spacer peptide hybrid nanosheet after mineralization. The nanosheet formed on STEM grid for 10 days incubation under the pH 7.5. The molar ratio of the nucleopeptide and spacer peptide was fixed at 1:6. Mineralization was carried out for 7 days at 15°C.

S11. Section analysis profile of the magnetite-nucleopeptide/spacer peptide hybrid

nanosheet.



Figure S11. Section analysis profile of modulus and adhesion images at same AFM observation spot in Figure 9(h) and (i).