

Supplementary Information

Formation of Supramolecular Single and Double Helix-like Structures by Designed Tripeptides

Rajat Subhra Giri and Bhubaneswar Mandal*

Department of Chemistry, Laboratory of Peptide and Amyloid Research, Indian Institute of
Technology Guwahati, Assam- 781039, India

Table of contents

1.	Crystal images	S3
2.	Table S1	S3
3.	Table S2	S4
4.	2D NMR and titration data (Table S3, S4, S5 and S6)	S5-S8
5.	Spectra of peptide 1 (Fig. S2-S9)	S9-S12
6.	Spectra of peptide 2 (Fig. S10-S17)	S13-S16
7.	References	S17

Crystal images:

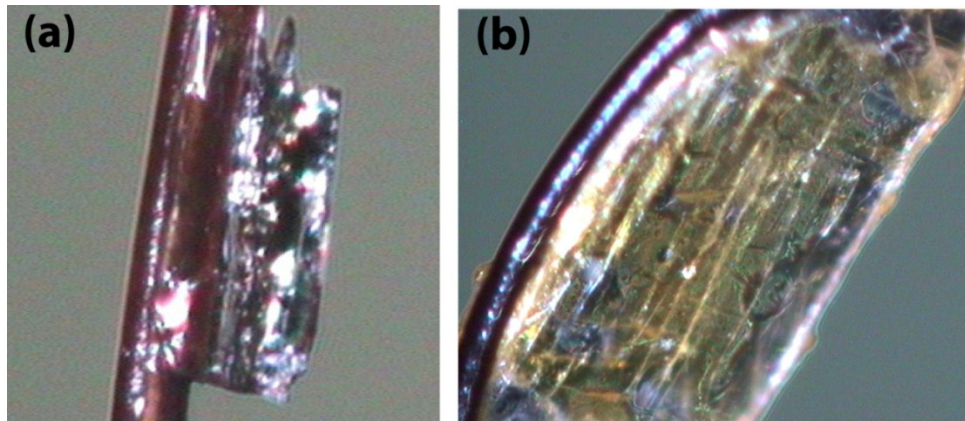


Fig. S1: Block shape crystal structure of (a) Boc-Gly-L-Phg-D-Phe-OMe (**1**) and Boc-Gly-L-Phg-D-Phg-OMe (**2**)

Table S1: Compares the torsional angles (deg) of L/L and D/L tripeptides

En try	Peptides		ϕ 1	ψ 1	ϕ 2 (L)	ψ 2 (L)	ϕ 3 (L)	ψ 3 (L)	Confor mations	Ref
1	Boc-Gly-L-Phg-L-Phe-OMe		126.2 (3)	-121.4 (3)	-127.6 (3)	124.4 (3)	-159.9 (3)	-179.0 (3)	Parallel β -sheet	1
					ϕ (L)	ψ (L)	ϕ (D)	ψ (D)		
2	β -helix peptide				-133	117	118	-130	β -helix	2,3
3	Boc-Gly-L-Phg-D-Phe-OMe (1)	Molecul e A	-59.8 (4)	150.1 (3)	-141.4 (3)	112.7 (3)	141.1 (3)	8.0 (5)	Anti-parallel β -sheet	This work
		Molecul e B	155.7 (3)	-143.2 (3)	-128.7 (3)	140.9 (3)	133.6 (3)	-109.7 (3)		
4	Boc-Gly-L-Phg-D-Phg-OMe (2)		172.9 (3)	-175.1 (3)	-138.0 (3)	141.3 (3)	142.3 (3)	18.1 (4)	Anti-parallel β -sheet	This work

Table S2: Hydrogen bonding distances (Å) and Bond angles (°) of peptide **1** and **2**

molecule	D–H $\cdots$ A	$d(\text{D}\cdots\text{H})$ /Å	$d(\text{H}\cdots\text{A})$ /Å	$d(\text{D}\cdots\text{A})$ /Å	$\angle\text{D–H}\cdots\text{A}/^\circ$	symmetry operation
Boc-Gly-L- Phg-D-Phe- OMe (1)	N1-H1 $\cdots$ O11	0.86	2.39	3.009(2)	130	$x, -1+y, z$
	N2-H2 $\cdots$ O10	0.86	2.09	2.906(2)	158	
	N3-H3 $\cdots$ O9	0.86	2.05	2.899(2)	168	$x, -1+y, z$
	N4-H4 $\cdots$ O6	0.86	2.56	3.390(2)	163	$x, 1+y, z$
	N5-H5 O4	0.86	2.03	2.870(2)	164	
	N6-H6 O3	0.86	2.01	2.850(2)	166	$x, 1+y, z$
	C8-H8 O9	0.98	2.53	3.400(2)	149	$x, -1+y, z$
	C17-H17B O8	0.97	2.57	3.179(2)	121	
	C20-H20 O5	0.93	2.58	3.336(2)	139	$-1+x, y, z$
	C31-H31A O4	0.97	2.43	3.278(2)	145	
	C33-H33 O3	0.98	2.39	3.213(2)	142	$x, 1+y, z$
	C37-H37 O3	0.93	2.59	3.432(3)	152	$-1+x, 1+y, z$
	C50-H50B O2	0.96	2.51	3.367(2)	148	$1+x, y, z$
Boc-Gly-L- Phg-D-Phg- OMe (2)	N1-H1 O4	0.86	2.18	3.021(1)	164	$1/2-x, -1/2+y, 1-z$
	N2-H2 O3	0.86	2.09	2.927(1)	164	$1/2-x, 1/2+y, 1-z$
	N3-H3 O2	0.86	2.12	2.968(1)	167	$1/2-x, -1/2+y, 1-z$
	C6-H6B O5	0.97	2.41	3.308(1)	155	$1/2+x, 1/2+y, z$
	C14-H14 O2	0.93	2.45	3.296(1)	151	$1/2-x, -1/2+y, 1-z$

2D NMR and titration data:

Table S3. Proton chemical shifts (ppm) for peptide **1**

Amino acid	HN	H α	H β	Others
Gly	5.24	3.81		Boc CH ₃ 1.43
Phg	7.41	5.50		OCH ₃ 3.70
Phe	6.55	4.90-4.85	2.97-2.95	

Table S4. Titration of peptide **1** in CDCl₃ with d₆-DMSO

Sl. No.	The volume of d ₆ -DMSO added (μL)	Chemical shifts δ (ppm)					
		NH of L-Phg	$\Delta\delta$ (NH) of L-Phg	NH of D-Phe	$\Delta\delta$ (NH) of D-Phe	NH of Gly	$\Delta\delta$ (NH) of Gly
1	0	7.410		6.550		5.238	
2	5	7.452		6.630		5.285	
3	10	7.485		6.783		5.341	
4	15	7.549		6.900		5.435	
5	20	7.649	0.239	7.036	0.486	5.674	0.436

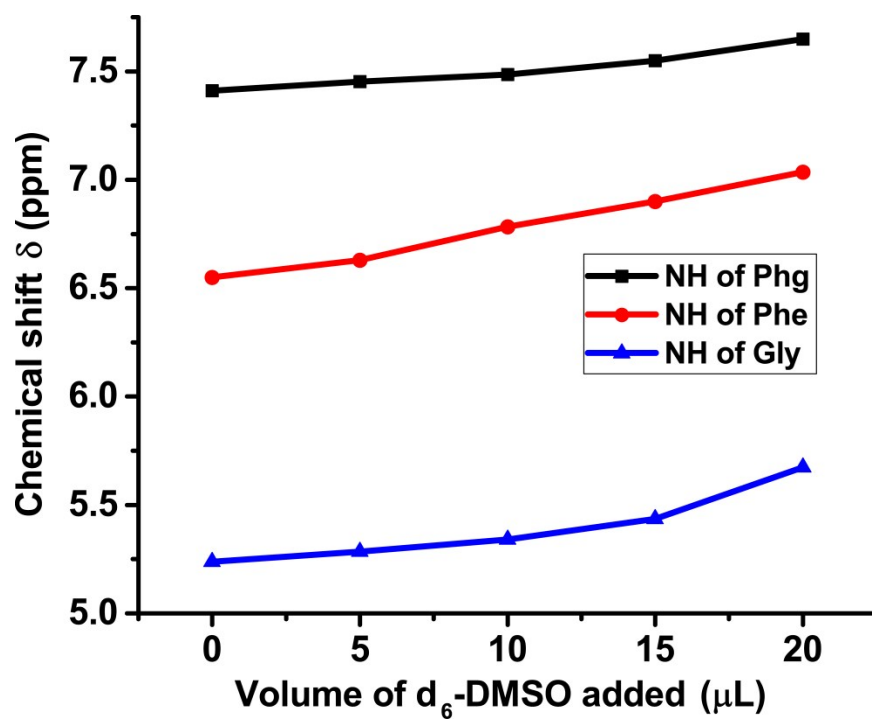
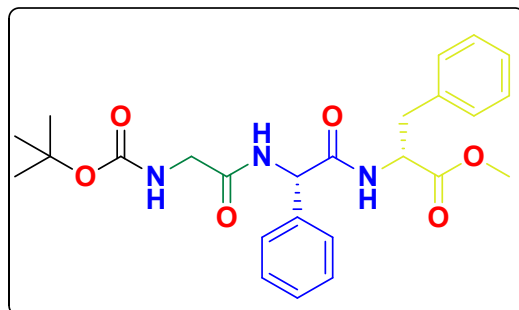


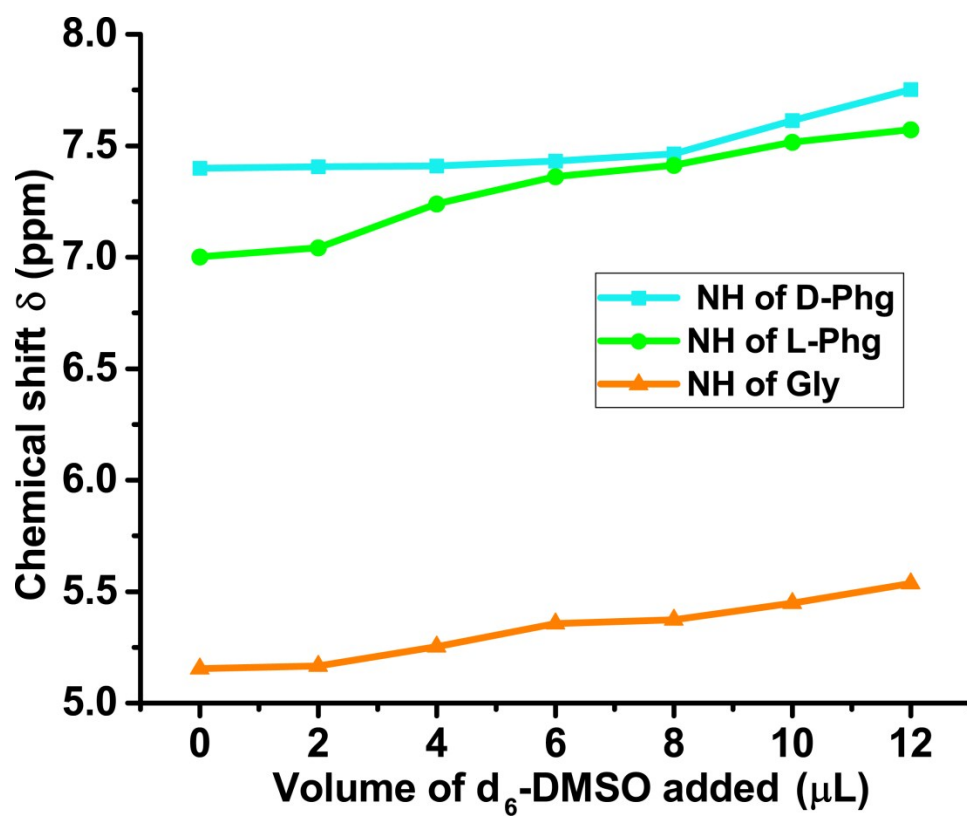
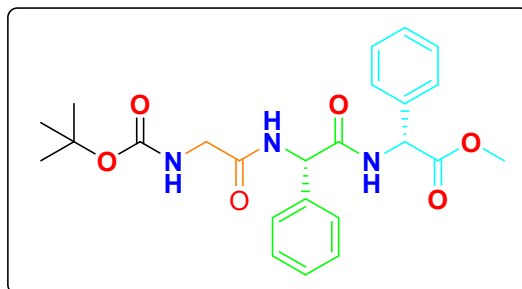
Table S5. Proton chemical shifts (ppm) for peptide **2**

Amino acid	HN	H α	Others
Gly	5.20	3.82	Boc CH ₃ 1.41
L-Phg	7.39	5.66	OCH ₃ 3.70
D-Phg	7.00	5.52-5.51	

Table S6. Titration of peptide **2** in CDCl₃ with d₆-DMSO

Sl. No.	The volume of d ₆ -DMSO added (μL)	Chemical shifts δ (ppm)					
		NH of D-Phg	$\Delta\delta$ (NH) of D-Phg	NH of L-Phg	$\Delta\delta$ (NH) of L-Phg	NH of Gly	$\Delta\delta$ (NH) of Gly
1	0	7.002		7.399		5.154	
2	2	7.0042		7.406		5.167	
3	4	7.239		7.409		5.253	
4	6	7.361		7.432		5.357	
5	8	7.412		7.463		5.374	

6	10	7.516		7.613		5.449	
7	12	7.573	0.571	7.752	0.353	5.536	0.382



Spectra of peptide 1:

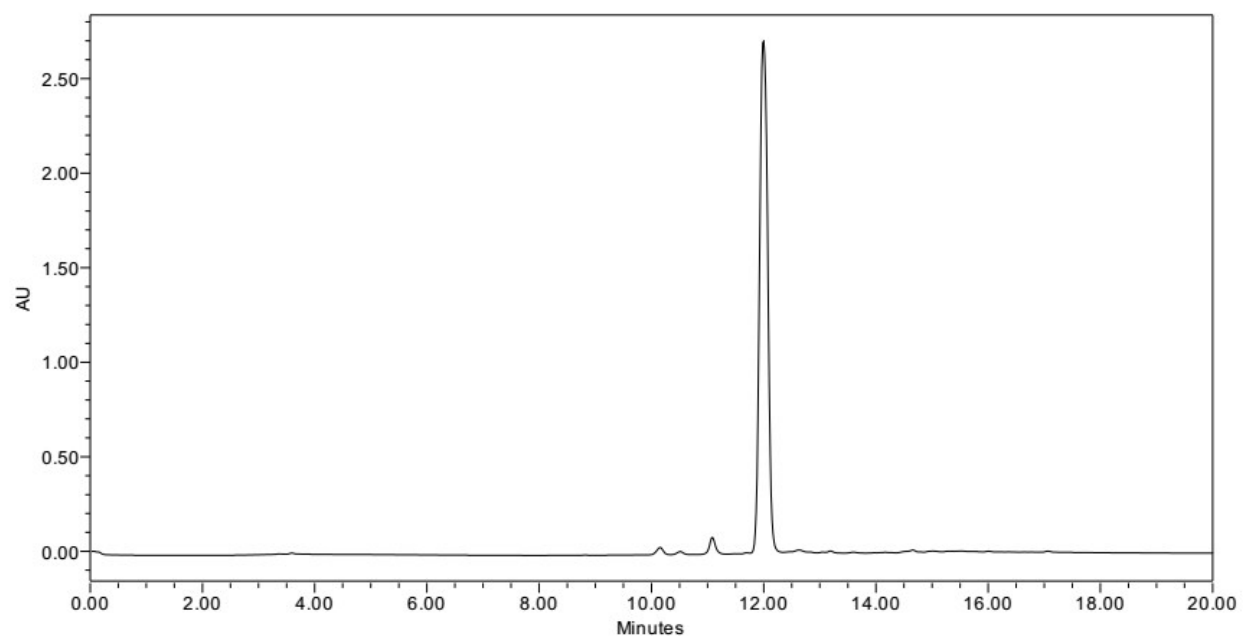


Fig. S2: HPLC profile picture of purified peptide **1**

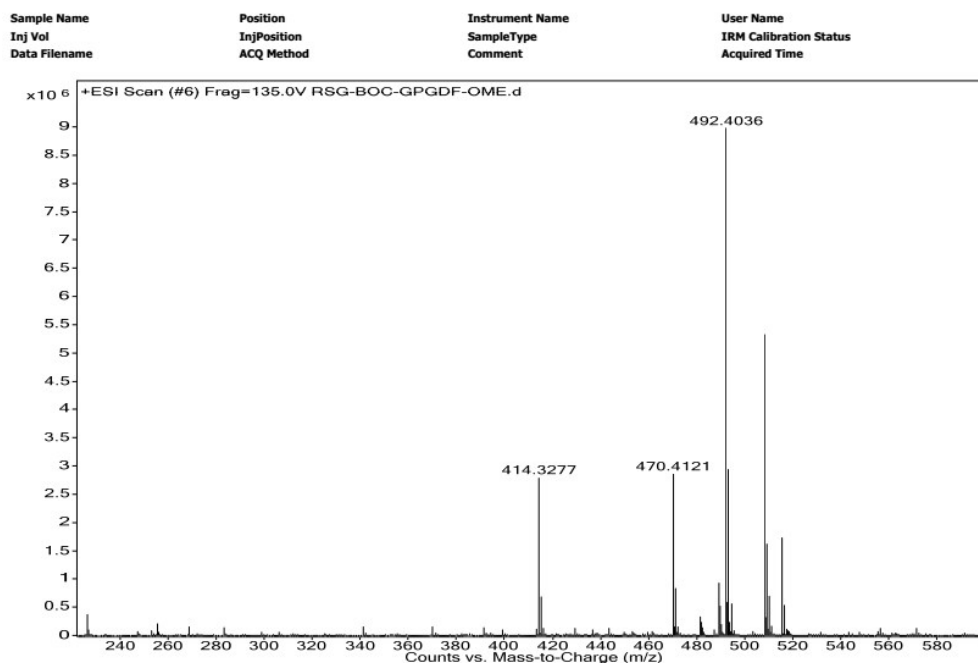


Fig. S3: MS spectra of peptide 1

RSG-GPGDF-0-1H
RSG-GPGDF-0-1H

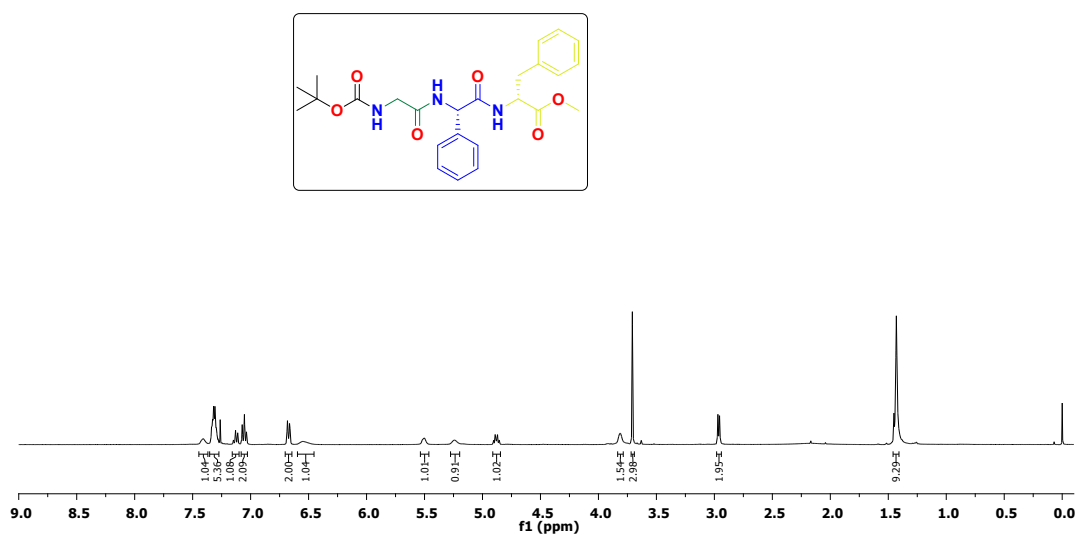


Fig. S4: ¹H NMR spectra of peptide 1

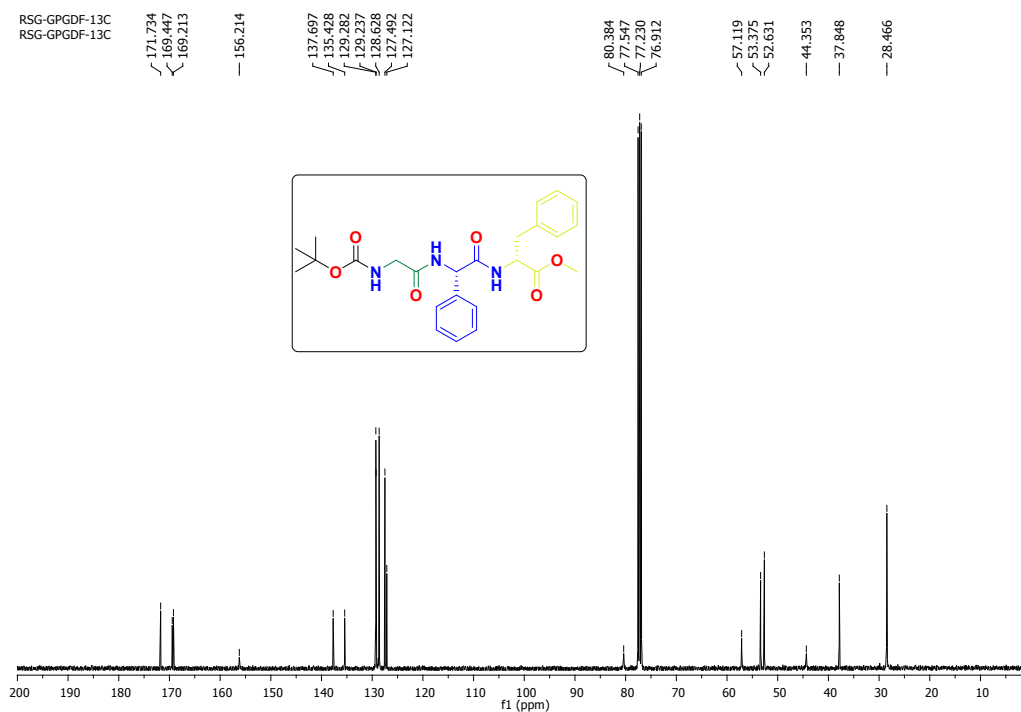


Fig. S5: ¹³C NMR spectra of peptide 1

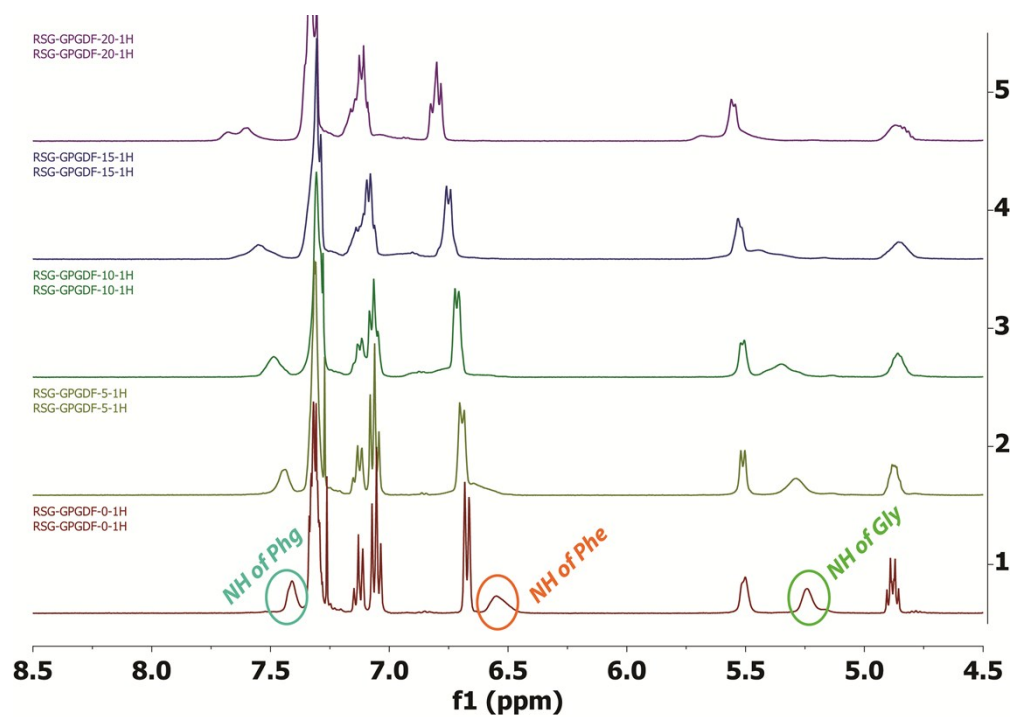


Fig. S6: Partial titration spectra of peptide 1 in CDCl₃ with d₆-DMSO

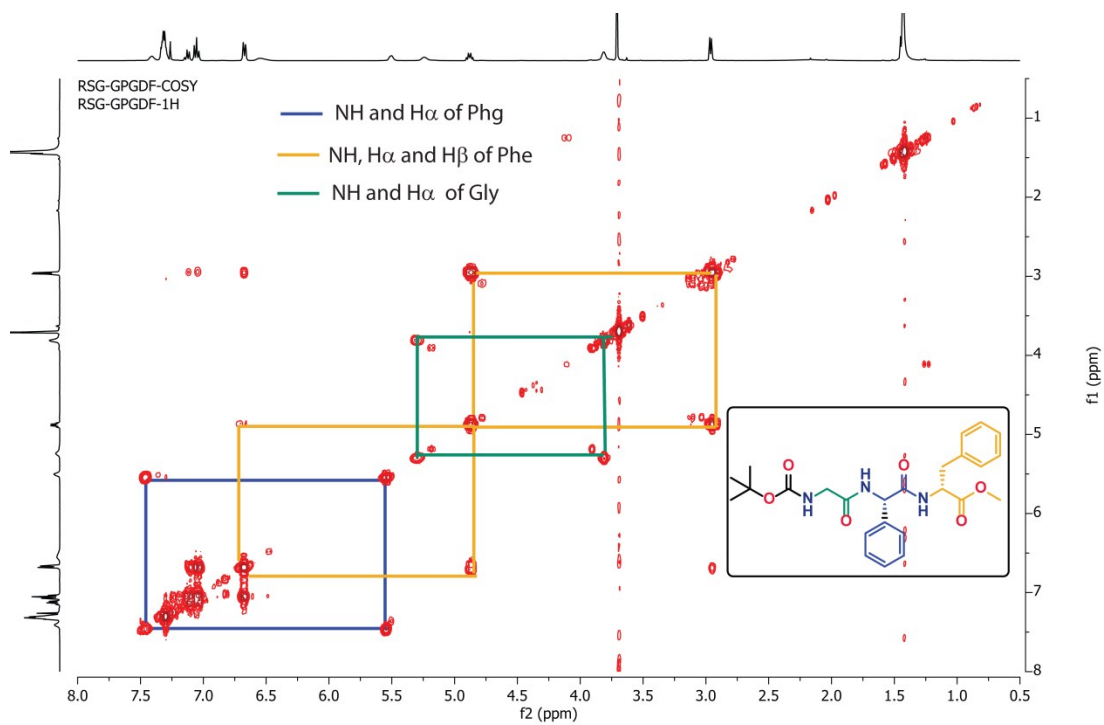


Fig. S7: COSY spectra of peptide 1

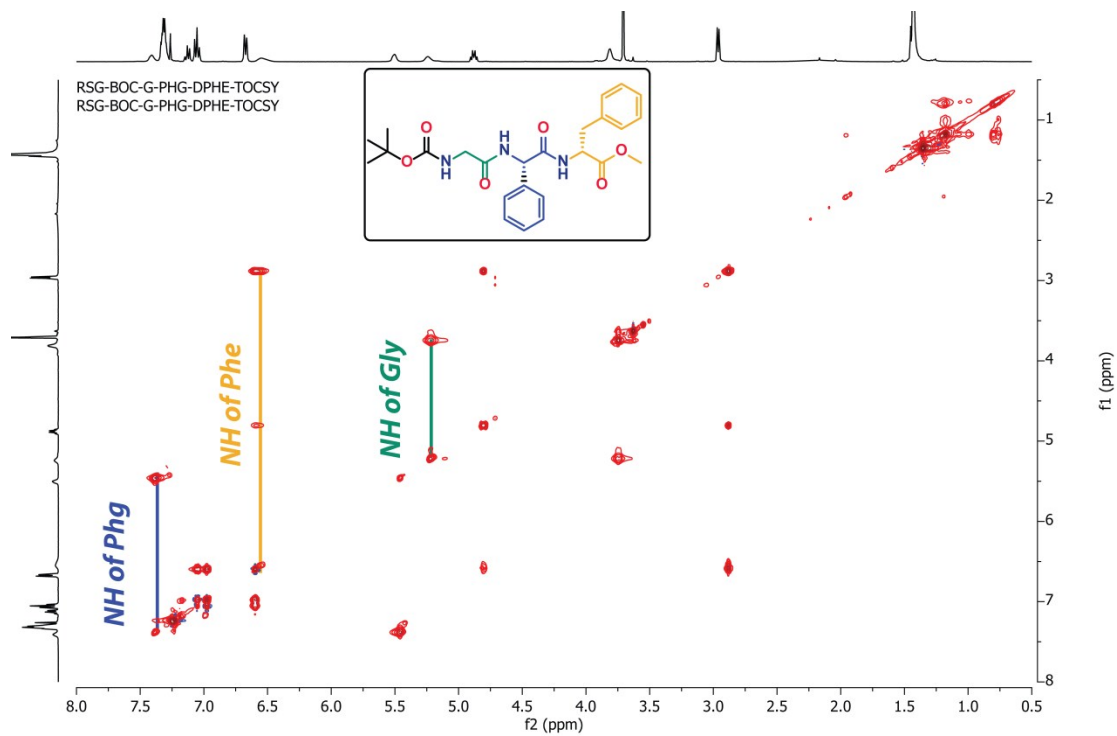


Fig. S8: TOCSY spectra of peptide 1

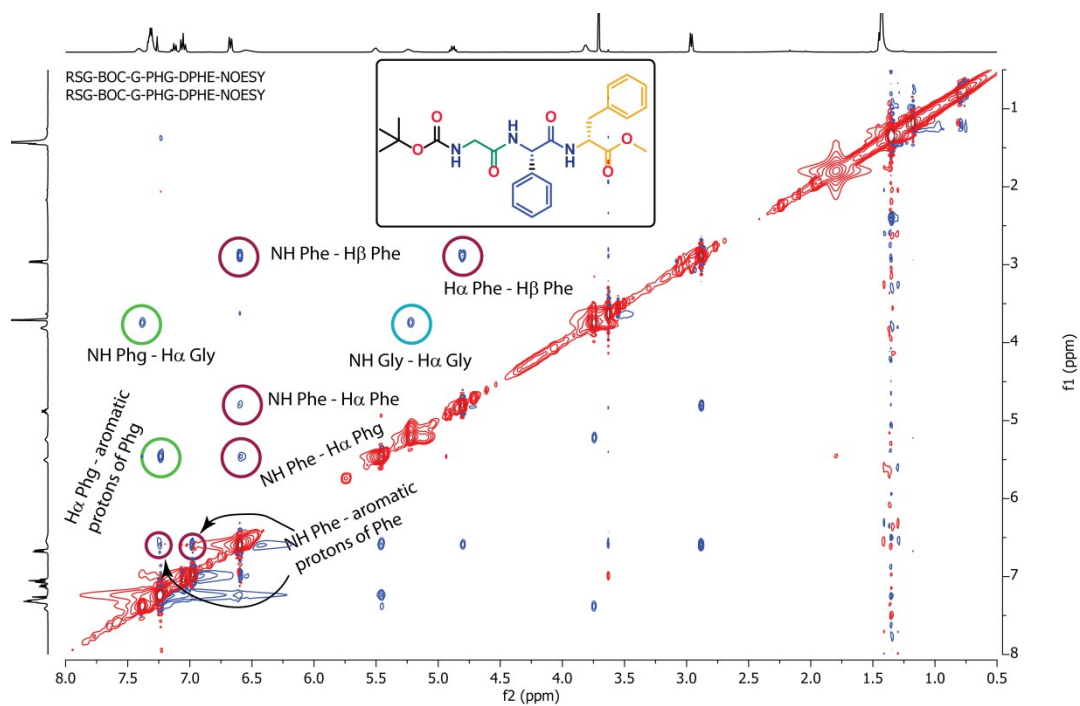


Fig. S9: NOESY spectra of peptide 1

Spectra of peptide 2:

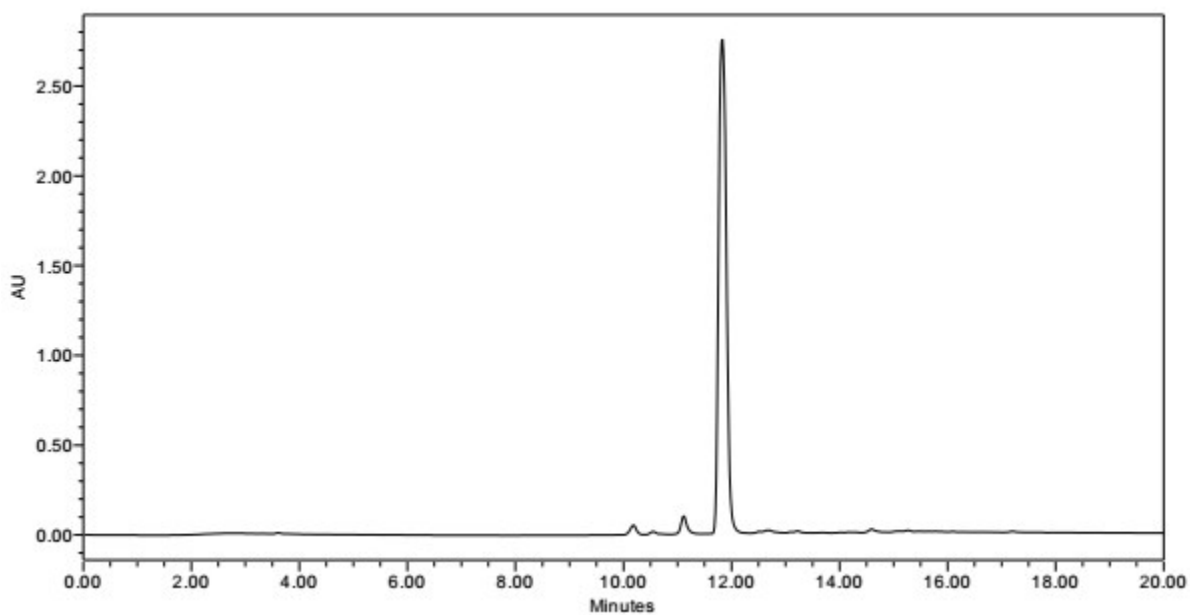


Fig. S10: HPLC profile picture of purified peptide 2

Sample Name	G-PHY-D-PHY	Position	P2-F4	Instrument Name	Instrument 1	User Name	
Inj Vol	20	InjPosition		SampleType	Sample	IRM Calibration Status	Success
Data Filename	G-PHY-D-PHY.d	ACQ Method	ESI ALS 50-500.m	Comment		Acquired Time	12/20/2018 12:35:01 PM

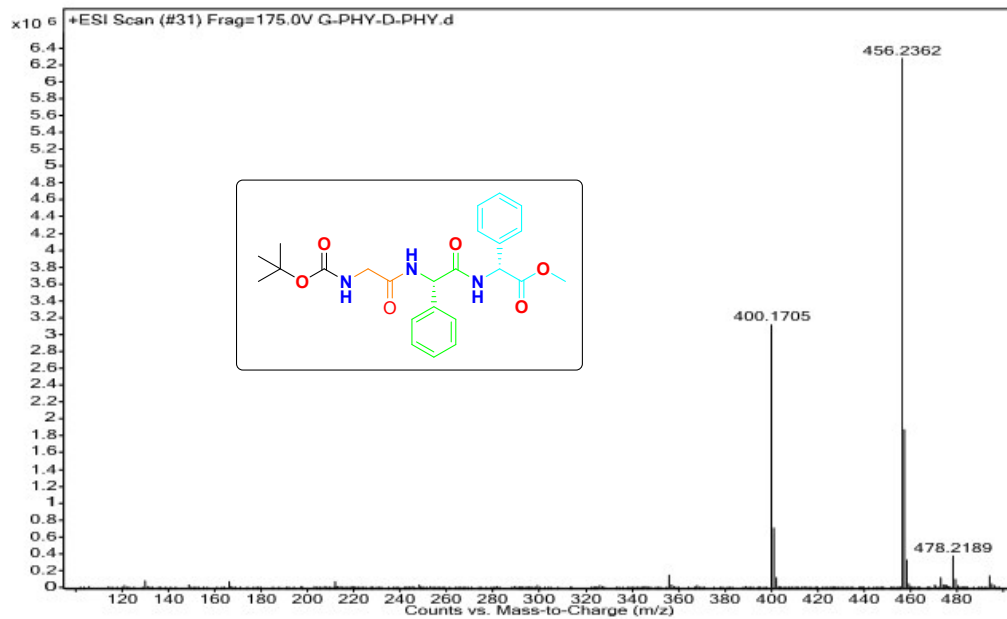


Fig. S11: MS spectra of peptide 2

RSG-gpgdpg-1H
RSG-gpgdpg-1H

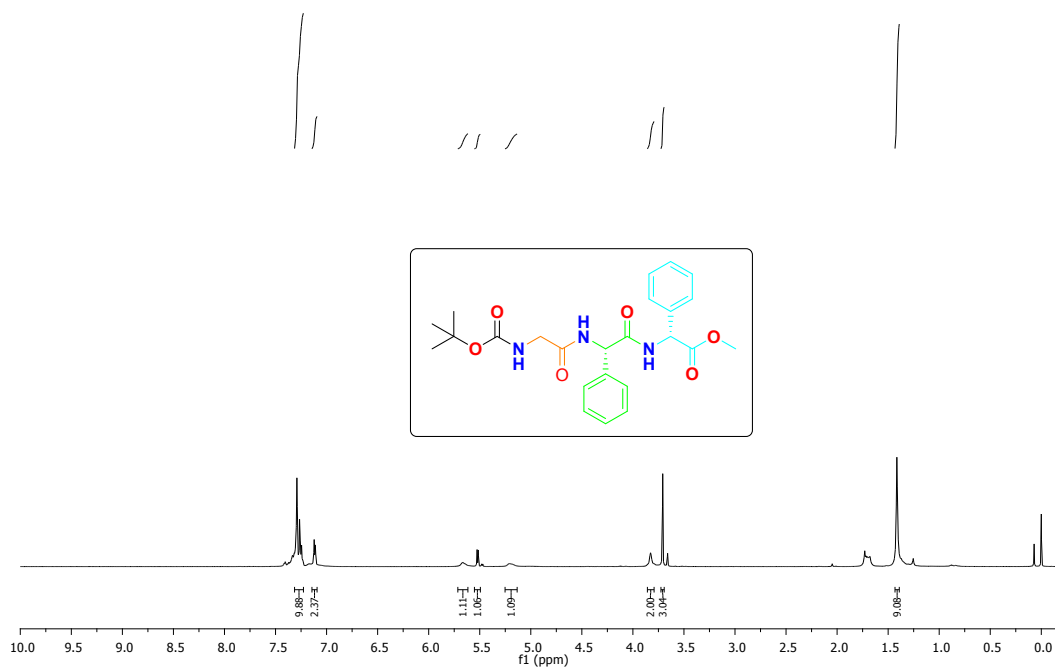


Fig. S12: ¹H NMR spectra of peptide 2

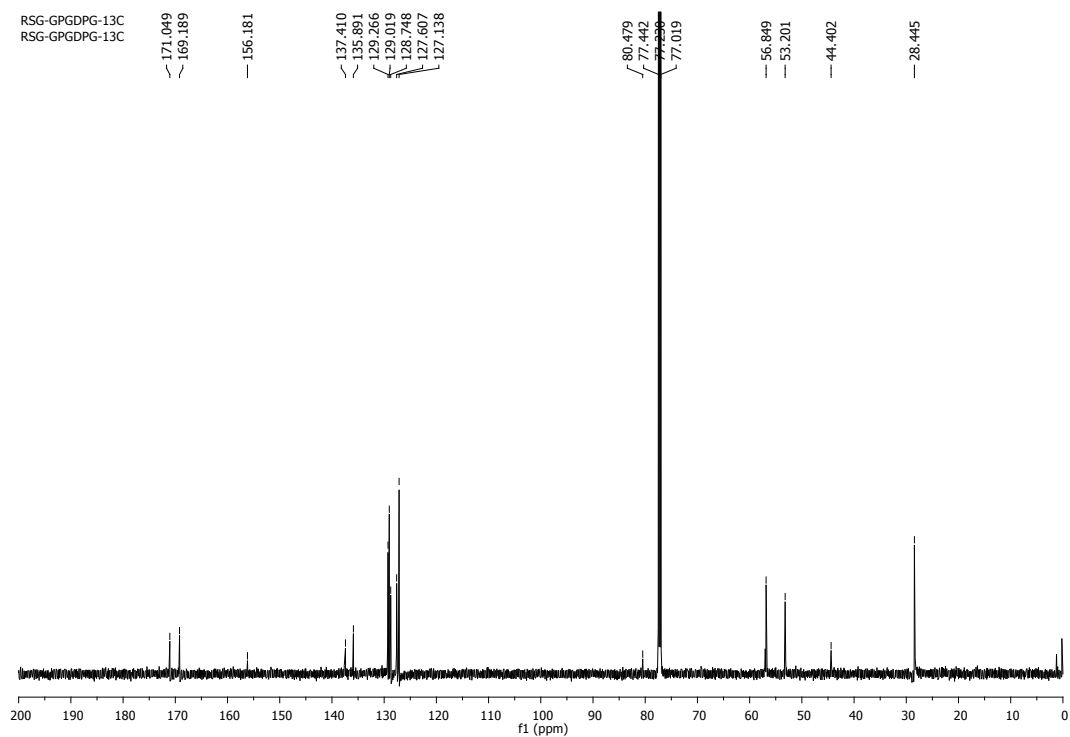


Fig. S13: ^{13}C NMR spectra of peptide 2

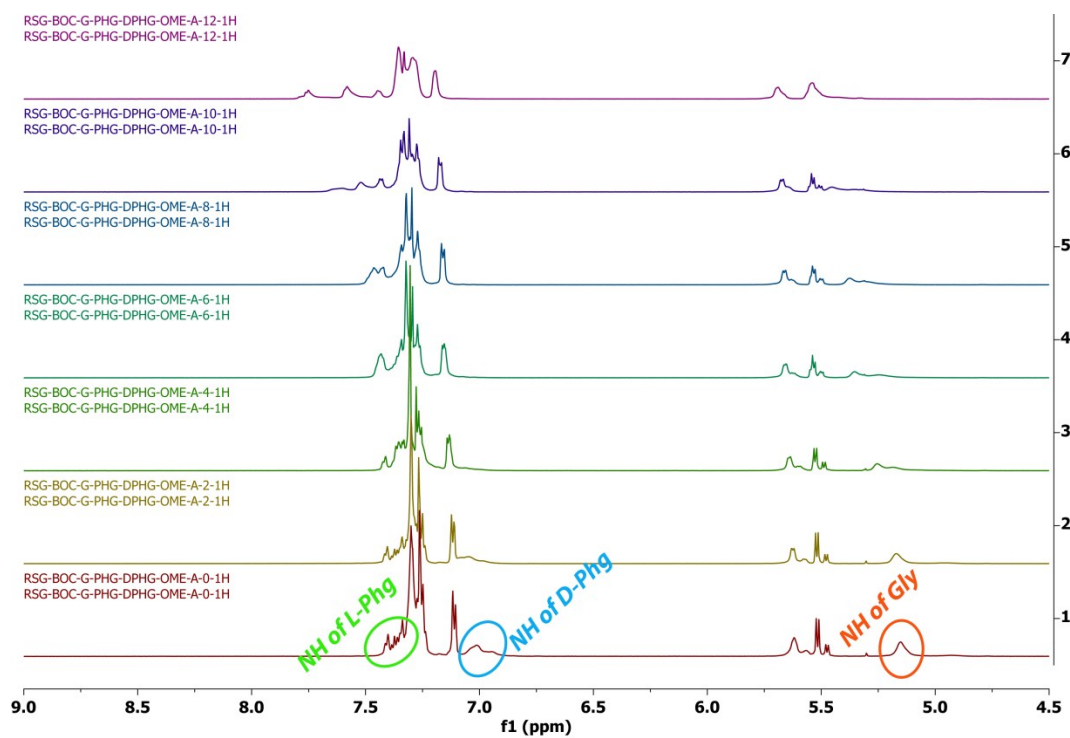


Fig. S14: Partial titration spectra of peptide 2 in CDCl_3 with d_6 -DMSO

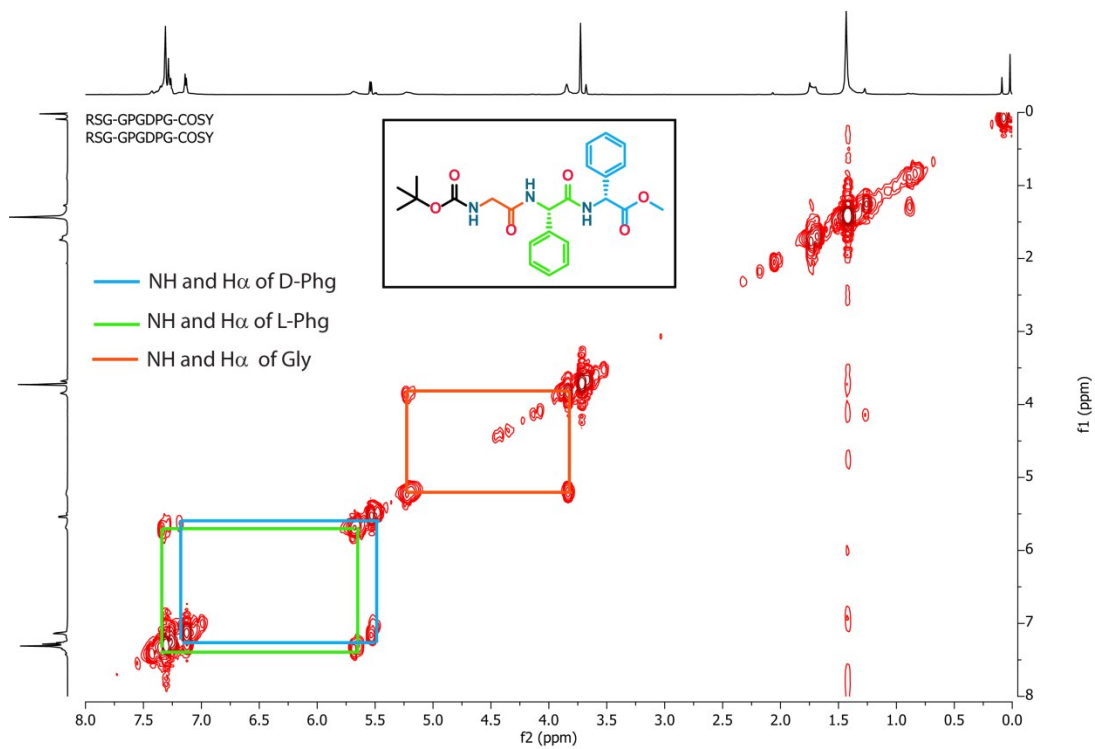


Fig. S15: COSY spectra of peptide 2

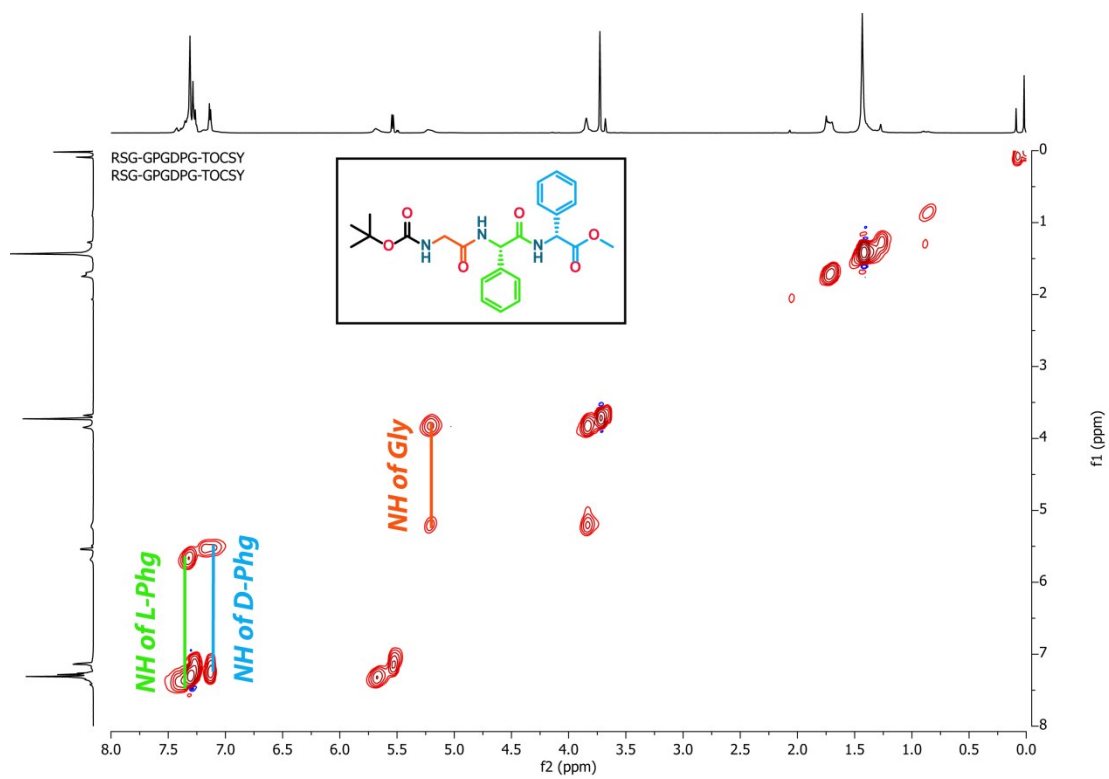


Fig. S16: TOCSY spectra of peptide 2

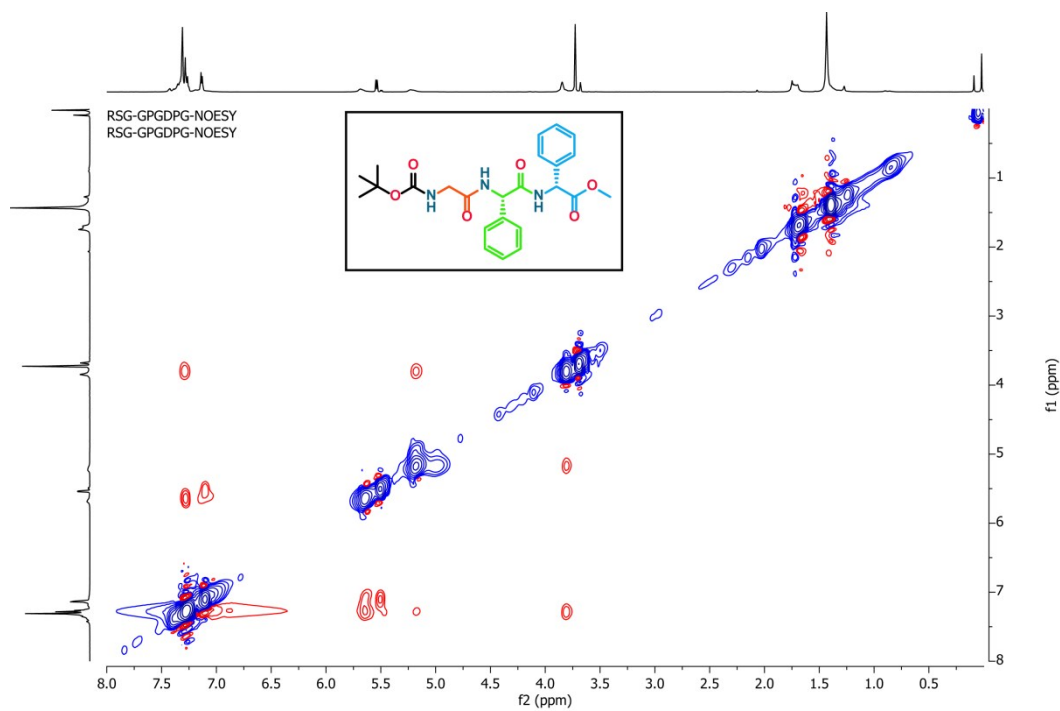


Fig. S17: NOESY spectra of peptide **2**

References:

1. R. S. Giri and B. Mandal, *CrystEngComm*, 2019, **21**, 236–243.
2. P. De Santis, S. Morosetti and R. Rizzo, *Macromolecules*, 1974, **7**, 52–58.
3. D. T. Bong, T. D. Clark, J. R. Granja and M. R. Ghadiri, *Angew. Chem. Int. Ed.*, 2001, **40**, 988–1011.