Heteronuclear d-d and d-f Ru(II)/M complexes [M = Gd(III), Yb(III), Nd(III), Zn(II) or Mn(II)] of ligands combining phenanthroline and aminocarboxylate binding sites: combined relaxivity, cell imaging and photophysical studies.

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Experimental Supporting Information



Fig. S1. ¹H NMR spectrum (400 MHz, d6-acetone) of **Ru**•**E**. H_b = Proton on bipyridine ligand; H_p = proton on phenanthroline ligand; H_{py} = proton on pyridine ring.



Fig. S2. High-resolution ES mass spectrum of Ru•E



Fig. S3. ¹H NMR spectrum (500 MHz, d6-DMSO) of **Ru**•L. H_b = Proton on bipyridine ligand; H_p = proton on phenanthroline ligand; H_{py} = proton on pyridine ring.



Fig. S4. High-resolution ES mass spectrum of Ru-L



Fig. S5. ¹H NMR spectrum (400 MHz, d₆-acetone) of **Ru**•**E**₂ at 298 K. H_b = proton on bipyridine ligand, H_p = proton on phenanthroline ligand, H_{py} = proton on pyridine ring.



Fig. S6 High-resolution ES mass spectrum of Ru•E2



Fig. S7. ¹H NMR spectrum (400 MHz, D₂O) of **Ru**•L₂ at 298 K. H_b = proton on bipyridine ligand, H_p = proton on phenanthroline ligand, H_{py} = proton on pyridine ring.



Fig. S8. High-resolution ES mass spectrum of Ru•L2



Fig. S9. Part of the high-resolution ES mass spectrum of **Ru**•**Gd** showing (top) observed and (below) calculated signals for $[C_{49}H_{37}N_9O_8RuGd + H]^{2+}$



Fig. S10. Part of the high-resolution ES mass spectrum of $\mathbf{Ru} \cdot \mathbf{Gd}_2$ showing (top) observed and (bottom) calculated signals for $[C_{66}H_{50}N_{12}O_{16}RuGd_2+2H]^{2+}$



Fig. S11. Part of the high resolution ES mass spectrum of **Ru**•Nd showing the signal for $[C_{49}H_{37}N_9O_8RuNd + H]^{2+}$.



Fig. S12. Part of the high resolution ES mass spectrum of $\mathbf{Ru} \cdot \mathbf{Yb}$ showing the signal for $[C_{49}H_{37}N_9O_8RuYb + H]^{2+}$.



Fig. S13. Part of the high resolution ES mass spectrum of $\mathbf{Ru} \cdot \mathbf{Nd}_2$ showing the signal for $[C_{66}H_{50}N_{12}O_{16}RuNd_2 + 2H]^{2+}$.



Fig. S14. Part of the high resolution ES mass spectrum of $\mathbf{Ru} \cdot \mathbf{Yb}_2$ showing the signal for $[C_{66}H_{50}N_{12}O_{16}RuYb_2 + 2H]^{2+}$.



Fig. S15. ¹H NMR spectrum (400 MHz, D₂O) of **Ru**•**Zn** at 298 K. (H_b = proton on bipyridine ligand, H_p = proton on phenanthroline ligand, H_{py} = proton on pyridine ring).



Fig. S16. Part of the high resolution ES mass spectrum of **Ru**•**Zn** showing the signal for $[C_{49}H_{37}N_9O_8RuZn+2H]^{2+}$.



Fig. S17. Part of the high resolution ES mass spectrum of **Ru**•**Mn** showing the signal for $[C_{49}H_{37}N_9O_8RuMn+2H]^{2+}$.



Fig. S18. ¹H NMR spectrum (400 MHz, D₂O) of **Ru**•**Zn**₂ at 298 K. (H_b = proton on bipyridine ligand, H_p = proton on phenanthroline ligand, H_{py} = proton on pyridine ring).



Fig. S19. Part of the high resolution ES mass spectrum of $\mathbf{Ru} \cdot \mathbf{Zn}_2$ showing the signal for $[C_{66}H_{50}N_{12}O_{16}RuZn_2]^{2-}$.



Fig. S20. Part of the high resolution ES mass spectrum of $\mathbf{Ru} \cdot \mathbf{Mn}_2$ showing the signal for $[C_{66}H_{50}N_{12}O_{16}RuMn_2]^{2-}$.



Fig. S21. UV/Vis absorption spectra in MeOH, normalised at 435 nm, of **Ru**•**E** (green) and **Ru**•**E**₂ (red), emphasising the difference in the ¹MLCT absorption region (500 – 550 nm). For extinction coefficients see main text.



Fig. S22. Excitation spectra recorded in D₂O at 298K of (a) **Ru**•Nd and (b) **Ru**•Nd₂, monitoring excitation at 1060 nm in both cases, showing overlap of the main feature of the excitation spectra with the Ru-based ¹MLCT absorption manifold.



Fig. S23. Decay-associated spectra obtained by global analysis of the transient absorption spectral data of **Ru**•**Zn** (left) and **Ru**•**Mn**, in water at RT.



Fig. S24. Evolution-associated spectra, illustrating the appearance of the experimental TA spectra at different time delays, for **Ru**•**Zn** (left) and **Ru**•**Mn** (right), in water at RT.



Fig. S25. Plot of relaxation rate of water protons (s⁻¹) *vs.* concentration of Gd(III)-containing complex (mM) for **Ru•Gd** (red dots), **Ru•Gd**₂ (blue dots) and Magnevist (green dots) (400 MHz, 298 K, D₂O).



Fig. S26. Clonogenic toxicity assay of HeLa cells incubated with **Ru**•**Gd** (red line) or **Ru**•**Gd**₂ (blue line) at concentrations of 50 μ M and 200 μ M for 4h. Incubations were carried out in full DMED at 37 °C under an atmosphere of CO₂/air (5/95, *v*/*v*) for 7 – 10 days until visible cell colonies had formed. Error bars represent the standard deviation of six data points (three repeats of each of duplicate datasets).



Fig. S27. Plot of concentration (mM) *vs.* relaxation rate of water protons (s⁻¹) to determine the concentration-normalised longitudinal relaxivity values for **Ru**•**Mn** (orange) and **Ru**•**Mn**₂ (purple). The data for **Ru**•**Gd** (red), and **Ru**•**Gd**₂ (blue) and the commercial MRI contrast agent Magnevist® (green) are also included for comparison (400 MHz, 298 K, D₂O).