# **Supplementary Information**

# Selective Recognition of DNA Defects By Cyclometalated Ir(III) complexes

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#### **Materials and Methods**

## Synthesis and characterization of [Ir(phpy)2(L)]PF<sub>6</sub>.

A mixture of  $[(phpy)_2Ir(\mu-Cl)]_2$  (0.12 g, 0.1 mmol) and Imiphen(L1) or Furphen(L2) or Faqphen(L3) (0.07 g, 0.25 mmoL) in MeOH/CH<sub>2</sub>Cl<sub>2</sub> (50 mL, 1:1) was heated to reflux with stirring under N<sub>2</sub> for 12h. The reaction mixture was then cooled to room temperature, followed by addition of NH<sub>4</sub>PF<sub>6</sub> (0.04 g, 2.5 mmol). The solution was stirred for 30 min. After removal of solvent under reduced pressure, the crude product was purified by column chromatography on silica gel using CH<sub>2</sub>Cl<sub>2</sub>/CH<sub>3</sub>CN (5:1, V:V) as eluent to afford the desired [Ir(phpy)<sub>2</sub>(N^N)]PF<sub>6</sub> as orange solid.

**Complex Ir-1:** <sup>1</sup>H NMR (400 MHz,  $d_6$ -DMSO),  $\delta$ (ppm) : 9.4(m,1H), 9.3(s,1H), 8.32 (m,1H), 7.9(m,1H), 7.7(m, 2H), 7.5 (s,1H), 7.3(s,1H), 7.1 (m,2H), 6.9(s,1H), 6.4(m,1H). ESI-MS (m/z) : 788.16 (M-PF<sub>6</sub>)<sup>+</sup>

**Complex Ir-2:** <sup>1</sup>H NMR (400 MHz, *d*<sub>6</sub>-DMSO), δ (ppm): 9.4(d, 1H), 9.0 (dd, 1H), 8.32 (dd, 1H), 8.2 (dd, 1H), 8.1 (d,1H), 7.9(m, 2H), 7.7(m, 2H), 7.1 (m, 1H), 6.9(m, 2H), 6.3(m,1H). ESI-MS (*m/z*): 839.08 (M+H<sub>2</sub>O+Cl-PF<sub>6</sub>)

**Complex Ir-3:** <sup>1</sup>H NMR (400 MHz, *d*<sub>6</sub>-DMSO), δ (ppm): 9.24 (dd, 1H), 8.8 (dd, 1H), 8.4 (d, 2H), 8.3 (t,2H), 8.14(dd, 1H), 7.9(m, 5H), 7.6 (d, 1H), 7.1(m, 3H), 6.3(d, 1H) ESI-MS (*m/z*): 927.18 (M-PF<sub>6</sub>)<sup>+</sup>

The molecular mass of the Ir(III) complexes was recorded with Agilent mass spectrometer equipped with an electron spray source. <sup>1</sup>H NMR for the ligands and its corresponding complexes was performed using Bruker 400MHz spectrometer. Electronic absorption spectra of all the three complexes in acetonitrile were carried out using Shimazdu UV-1700 model spectrophotometer. Steady state emission spectra of the complexes were recorded on a Hitachi F-7000 spectrofluorimeter at 25°C. Steady-state luminescence spectra of the Ir(III) complexes were recorded using Hitachi F7000 spectrofluorimeter equipped with thermostat. The binding constants for the Ir(III) with different oligonucleotides were calculated using Scatchard plot.<sup>1</sup> The room temperature luminescence lifetimes of the Ir(III) complexes with and without oligonucleotides were measured with Time Correlated Single Photon Counting Fluorescence Spectrophotometer (TCSPC-FS) (Edinburgh model-FLS980), UK. The lifetimes of the complexes were calculated using the software provided along with the TCSPC instrument.

### **Fluorescence experiments:**

#### With Oligonucleotides:

Duplex DNA were prepared by annealing the respective primer and complementary sequence in Tris buffer (10mM)/50mM slowly after heating to 90°C for five minutes followed by slow cooling to room temperature over a period of 2-3 h. The sample were incubated at 4°C for overnight before experiments. Fluorescence experiments were carried out by incubating at iridium (III) complexes (1 $\mu$ M) and varying the duplex concentration (0-2 $\mu$ M) in Tris buffer (pH-7.4). The complex were excited between 370-400nm and the emission intensity was integrated from 500-700nm.

# Reference

1. C. V. Kumar and E. H. Asuncion, J. Am. Chem.Soc., 1993, 115, 8547-8553.



Figure S1. ESI-MS of complex Ir-1







Fig. S4. <sup>1</sup>H NMR spectra of (a) Ir-1 (b) Ir-2 and (c) Ir-3



**Figure S5.** (a) UV-Visible in acetonitrile and (b) Emission spectra of Ir(III) complexes in dichloromethane

(a)

(b)



**(b)** 



FigureS6.ExcitedstatelifetimedecayplotsofIr(III)complexes(a)and(b)withcontrol and DNAdefects



Figure S7. Emission spectra of Ir-3 with match and mismatch oligonucleotide