Supporting Information

to

Steric Influence of Salicylaldehyde-based Schiff Base Ligands in the Formation

of trans-[Re(PR₃)₂(Schiff Base)]⁺ Complexes

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Section 1. SCXRD refinement details for compounds 2, 3•CH₃OH, 5, 7•2.333CH₃OH, and 8-11 (Table S1); ORTEP representations for compounds 2, 5, 8, 9, 10 (Figures S1-S5)

Section 2.¹H-NMR spectra for compounds 2-11 (Figures S6-S21)

Section 3. Cyclic voltammograms for compounds 2-7 (Figures S22-S28)

Supplemental Information Section 1. SCXRD refinement details for compounds 2, 3•CH₃OH, 5, 7•2.333CH₃OH, and 8-11

Compounds 2, 3•CH₃OH, 5, and 11 were solved by iterative dual space recycling as implemented in SHELXT v. 2014/5.1 Compounds 7•2.333CH₃OH, 8, 9, and 10 were solved by direct methods as implemented in SHELXS.² All structures were refined by full-matrix least squares refinement against F² using SHELXL v. 2017/1.³ Full occupancy non-hydrogen atoms were refined anisotropically to convergence. Hydrogen atoms were placed in calculated positions and constrained to ride on the carrier atoms; a ridingrotating model was used for methyl group hydrogen atoms. $[PF_6]^-$ ions in 2 and 5 and two ethyl groups from a P(Et)₃ ligand in 5 were found to be disordered over two positions; the relative occupancies were constrained to add to 1 and refined to values of 87.5%, 69.3%, 53.2%, and 54.3% for the major parts of all four groups, respectively. A total of five distance restraints were used to keep the disordered groups in 5 near reasonable values. Due to systematic errors in the intensities for 11 caused by satellite crystals, rigid group restraints were used to keep the anisotropic thermal ellipsoids for the terminal Re=O oxygen atom within reasonable values.⁴ For compounds **3**•CH₃OH and **7**•2.333CH₃OH, contributions to scattering from highly disordered solvent molecules were determined and removed using PLATON SQUEEZE.⁵ The structure of **3**•CH₃OH was found to contain 70 electrons per unit cell distributed across 256 Å³ of void space, in good agreement with the expected values for four methanol molecules (72 electrons). For 7•2.333CH₃OH, 336 electrons were found distributed through a total void volume of 1902 Å³, equivalent to 18.667 methanol molecules per unit cell (7 methanol molecules per 3 molecules of the metal complex).

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For Compound 2, the thermal ellipsoid of the Cl⁻ ion is significantly greater than the average values for other non-hydrogen atoms in the structure; greater thermal motion is chemically reasonable for this moiety as it is situated within the disordered solvent void. Compounds 8 and 11 contain significant difference map peaks in chemically unreasonable positions, which are likely caused by extra intensity from diffraction by unremovable satellite crystals. This is supported by the fact that $|F_{obs}| > |F_{calc}|$ for the most disagreeable reflections. For compound 11, which crystallized in the form of highly intergrown clumps, this problem is especially severe; nearly complete powder diffraction rings are observed on almost every frame of the data. Nevertheless, the data that was able to be obtained for 11 could be reliably indexed to a single unit cell and refined anisotropically as a structure fully consistent with the NMR data. Since both morphological inspection and the appearance of the diffraction data suggested the crystal of 11 is likely a single crystal core heavily coated in smaller, randomly oriented satellites, improving the model with de-twinning was not feasible.

References

- 1. Sheldrick, G. M., Acta Crystallogr. A **2015**, *71*, 3-8.
- 2. Sheldrick, G. M. SHELXS, v.2013-1, 2013.
- 3. Sheldrick, G. M., Acta Crystallogr. C 2015, 71, 3-8.
- 4. Thorn, A.; Dittrich, B.; Sheldrick, G. M., Acta Crystallogr. A **2012**, 68, 448-451.
- 5. Spek, A. L., Acta Crystallogr. C 2015, 71, 9-18.

	2	3 •CH₃OH	5	7●2.333CH₃O H	8	9	10	11
CCDC	1910610	1910611	1910612	1910613	1910614	1910615	1910616	1912253
formula	$C_{28}H_{44}F_6N_2O_2$ P_3Re	$\begin{array}{c} C_{44}H_{44}ClN_2O\\ {}_2P_2Re \end{array}$	$C_{30}H_{48}F_6N_2O_2$ P_3Re	$C_{46}H_{48}F_6N_2O_2$ P_3Re	$C_{24}H_{33}F_6N_2O_3$ P_2Re	$\begin{array}{c} C_{28}H_{33}F_6N_2\\ O_3P_2Re \end{array}$	$\begin{array}{c} C_{32}H_{33}F_{6}N_{2}\\ O_{3}P_{2}Re \end{array}$	$C_{36}H_{33}F_6N_2O_3P_2Re$
fw	833.76	980.48	861	1053.97	759.66	807.70	855.74	903.78
cryst system	Monoclinic	Triclinic	Monoclinic	Monoclinic	Monoclinic	Triclinic	Monoclinic	Triclinic
space group	$P2_1/c$	P-1	$P2_1/c$	C2/c	$P2_1/n$	P-1	$P2_1/n$	P-1
a (Å)	10.462(1)	10.5691(6)	23.173(2)	42.450(4)	8.7506(3)	10.7455(9)	9.327(1)	11.694(1)
<i>b</i> (Å)	15.724(2)	12.4434(8)	15.547(1)	10.1296(9)	10.9321(4)	12.292(1)	19.497(2)	13.536(2)
<i>c</i> (Å)	19.766(3)	17.083(1)	21.324(2)	30.348(3)	29.4316(11)	13.353(1)	17.659(2)	13.657(2)
a (deg)	90	100.042(1)	90	90	90	65.584(1)	90	68.440(3)
β (deg)	98.111(3)	91.437(1)	117.133(1)	130.284(1)	90.9445(12)	70.251(1)	94.463(2)	70.666(4)
γ (deg)	90	106.144(1)	90	90	90	83.092(1)	90	64.490(4)
$V(Å^3)$	3219.2(7)	2118.6(2)	6836.9(9)	9955(2)	2815.1(2)	1511.1(2)	3201.5(6)	1775.6(4)
Ζ	4	2	8	8	4	2	4	2
ρ (g/cm ³)	1.720	1.537	1.675	1.406	1.792	1.775	1.775	1.690
T (K)	100	223	100	100	100	100	100	100
μ (mm ⁻¹)	3.988	3.052	3.758	2.596	4.499	4.197	3.967	3.582
λ souce (Å)	0.71073	0.71073	0.71073	0.71073	0.71073	0.71073	0.71073	0.71073
$R(F)^a$	0.0289	0.0326	0.0263	0.0196	0.0307	0.0180	0.0208	0.0812
$Rw(F)^2$	0.0459	0.0667	0.0473	0.0426	0.0643	0.0391	0.0421	0.1896
GoF	1.005	1.017	0.944	1.025	1.390	1.036	1.350	1.193

Table S1. X-ray crystal data, data collection parameters, and refinement parameters for 2, 3•*CH*₃*OH, 5, 7*•*2.333CH*₃*OH, and 8-11.*



Figure S1. X-ray crystal structure of **2** with 50% probability thermal ellipsoids. Only the cation is shown.



Figure S2. X-ray crystal structure of **5** with 50% probability thermal ellipsoids. Only the cation is shown.



Figure S3. X-ray crystal structure of 8 with 50% probability thermal ellipsoids. Only the cation is shown.



Figure S4. X-ray crystal structure of **9** with 50% probability thermal ellipsoids. Only the cation is shown.



Figure S5. X-ray crystal structure of **10** with 50% probability thermal ellipsoids. Only the cation is shown.



Figure S6. Full view of ¹H NMR of **2** *trans*-[Re(PEt₃)₂(sal₂en)][PF₆] in CD₃CN (600 MHz, calibrated to residual CHD₂CN at 1.94 ppm).



Figure S7. Narrow view of ¹H NMR of **2** *trans*-[Re(PEt₃)₂(sal₂en)][PF₆] in CD₃CN (600 MHz, calibrated to residual CHD₂CN at 1.94 ppm).

¹H NMR [600 MHz, CD₃CN, r.t., δ (ppm)]: 60.06 (s, 2H, -HC=N-); 27.68 (d, *J* = 8.3 Hz, 2H, Ar**H**); 17.08 (t, *J* = 7.5 Hz, 2H, Ar**H**); 1.91 (t, *J* = 7.4 Hz, 18H, PCH₂C**H**₃); 1.76 (q, J = 7.3 Hz, 12H, PC**H**₂CH₃); -7.15 (t, *J* = 8.3 Hz, 2H, Ar**H**); -19.10 (d, *J* = 7.4 Hz, 2H, Ar**H**); -26.35 (s, 4H, -NC**H**₂C**H**₂N-).



Figure S8. Full view of ¹H NMR of **3** *trans*-[Re(PEtPh₂)₂(sal₂en)]Cl in CD₃CN (500 MHz, calibrated to residual CHD₂CN at 1.94 ppm).





Figure S9. Narrow view of ¹H NMR of **3** *trans*-[Re(PEtPh₂)₂(sal₂en)]Cl in CD₃CN (500 MHz, calibrated to residual CHD₂CN at 1.94 ppm).

¹H NMR [500 MHz, CD₃CN, r.t., δ (ppm)]: 61.16 (s, 2H, -HC=N-); 27.00 (d, *J* = 8.2 Hz, 2H, ArH); 16.45 (t, *J* = 7.3 Hz, 2H, ArH); 8.11 (t, *J* = 7.4 Hz, 4H, PPh para); 7.13 (t, *J* = 6.7 Hz, 6H, PCH₂CH₃); 6.61 (d, *J* = 7.7, 8H, PPh ortho); 6.56 (br s, 8H, PPh meta); 2.10 (q, *J* = 7.2 Hz, 4H, PCH₂CH₃); -7.84 (t, *J* = 8.0 Hz, 2H, ArH); -19.70 (d, *J* = 7.8 Hz, 2H, ArH); -20.28 (s, 4H, -NCH₂CH₂N-).

1H trans-[Re(PPh3)2(sal2en)][PF6]



Figure S10. Full view of ¹H NMR of **4** *trans*-[Re(PPh₃)₂(sal₂en)][PF₆] in CD₃CN (600 MHz, calibrated to residual CHD₂CN at 1.94 ppm).

1H trans-[Re(PPh3)2(sal2en)][PF6]



Figure S11. Narrow view of ¹H NMR of 4 *trans*-[Re(PPh₃)₂(sal₂en)][PF₆] in CD₃CN (600 MHz, calibrated to residual CHD₂CN at 1.94 ppm).

¹H NMR [600 MHz, CD₃CN, r.t., δ (ppm)]: 59.37 (s, 2H, -HC=N-); 26.54 (d, *J* = 7.8 Hz, 2H, Ar**H**); 15.75 (t, *J* = 7.5 Hz, 2H, Ar**H**); 8.20 (t, *J* = 7.6 Hz, 6H, P**Ph** meta); 7.24 (t, *J* = 7.1 12H, P**Ph** meta); 6.44 (br s, 12H, P**Ph** ortho); -9.30 (t, *J* = 8.1 Hz, 2H, Ar**H**); -19.51 (s, 4H, -NC**H**₂C**H**₂N-); -20.94 (d, *J* = 7.5 Hz, 2H, Ar**H**).



Figure S12. Full view of ¹H NMR of **5** *trans*-[Re(PEt₃)₂(sal₂ibn)][PF₆] in CD₃CN (600 MHz, calibrated to residual CHD₂CN at 1.94 ppm).

1H trans-[Re(PEt3)2(sal2ibn)]+



Figure S13. Narrow view of ¹H NMR of 5 *trans*-[Re(PEt₃)₂(sal₂ibn)][PF₆] in CD₃CN (600 MHz, calibrated to residual CHD₂CN at 1.94 ppm).

¹H NMR [600 MHz, CD₃CN, r.t., δ (ppm)]: 84.49 (s, 1H, -HC=N-); 56.59 (s, 1H, -HC=N-); 28.75 (d, *J* = 8.2 Hz, 1H, ArH); 27.75 (d, *J* = 8.3 Hz, 1H, ArH); 18.03 (t, *J* = 7.1 Hz, 1H, ArH); 17.25 (t, *J* = 7.0 Hz, 1H, ArH); 1.65 (s, 18H, P(CH₂CH₃)₃); 0.64-0.54 (br m, 6H, P(CH₂CH₃)₃); 0.34-0.25 (br m, 6H, P(CH₂CH₃)₃); -1.65 (s, 6H, -C(CH₃)₂CH₂-); -6.66 (t, *J* = 8.3 Hz, 1H, ArH); -6.95 (t, *J* = 8.3 Hz, 1H, ArH); -18.96 (d, *J* = 7.1 Hz, 1H, ArH); -19.75 (d, *J* = 7.1 Hz, 1H, ArH); -31.06 (s, 2H, -C(CH₃)₂CH₂-).



Figure S14. Full view of ¹H NMR of **6** *trans*-[Re(PEt₂Ph)₂(sal₂ibn)][PF₆] in CD₃CN (600 MHz, calibrated to residual CHD₂CN at 1.94 ppm).



Figure S15. Narrow view of ¹H NMR of **6** *trans*-[Re(PEt₂Ph)₂(sal₂ibn)][PF₆] in CD₃CN (600 MHz, calibrated to residual CHD₂CN at 1.94 ppm).

¹H NMR [600 MHz, CD₃CN, r.t., δ (ppm)]: 83.34 (s, 1H, -CH=N-); 55.92 (s, 1H, -CH=N-); 27.94 (d, J = 8.2 Hz, 1H, ArH); 27.64 (d, J = 8.5 Hz, 1H, ArH); 17.48 (t, J = 6.6 Hz, 1H, ArH); 17.15 (t, J = 7.5 Hz, 1H, ArH); 13.62 (d, J = 7.5 Hz, 4H, PPh *ortho*); 9.20 (t, J = 7.4 Hz, 2H, PPh *para*); 8.42 (t, J = 7.6 Hz, 4H, PPh *meta*); 3.41 (q, J = 6.7 Hz, 8H, PCH₂CH₃); 1.12 (t, J = 6.6 Hz, 12H, PCH₂CH₃); -2.53 (s, 6H, -C(CH₃)₂CH₂-); -6.41 (t, J = 8.2 Hz, 1H, ArH); -7.57 (t, J = 8.2 Hz, 1H, ArH); -18.51 (d, J = 7.2 Hz, 1H, ArH); -20.29 (d, J = 7.1 Hz, 1H, ArH); -28.65 (s, 2H, -C(CH₃)₂CH₂-).



Figure S16. Full view of ¹H NMR of **7** *trans*-[Re(PEtPh₂)₂(sal₂ibn)][PF₆] in CD₃CN (600 MHz, calibrated to residual CHD₂CN at 1.94 ppm).



Figure S17. Narrow view of ¹H NMR of **7** *trans*-[Re(PEtPh₂)₂(sal₂ibn)][PF₆] in CD₃CN (600 MHz, calibrated to residual CHD₂CN at 1.94 ppm).

¹H NMR [600 MHz, CD₃CN, r.t., δ (ppm)]: 81.68 (s, 1H, -CH=N-); 57.17 (s, 1H, -CH=N-); 27.78 (d, J = 8.2 Hz, 1H, ArH); 27.50 (d, J = 8.2 Hz, 1H, ArH); 16.91 (t, J = 7.6 Hz, 1H, ArH); 16.71 (t, J = 7.4 Hz, 1H, ArH); 8.43 (t, J = 7.6 Hz, 2H, PPh para); 8.36 (t, J = 7.3 Hz, 2H, PPh para); 7.93 (br s, 4H, PPh ortho); 7.84 (br s, 4H, PPh ortho); 7.18 (t, J = 7.6 Hz, 4H, PPh meta); 7.01 (t, J = 7.4 Hz, 4H, PPh meta); 5.54 (m, 6H, PCH₂CH₃); 0.72 (br s, 2H, PCH₂CH₃); 0.31 (br s, 2H, PCH₂CH₃); -2.39 (s, 6H, -C(CH₃)₂CH₂-); -6.59 (t, J = 8.2 Hz, 1H, ArH); -8.46 (t, J = 8.2 Hz, 1H, ArH); -18.85 (d, J = 7.3 Hz, 1H, ArH); -20.92 (d, J = 7.3 Hz, 1H, ArH); -26.39 (s, 2H, -C(CH₃)₂CH₂-).



Figure S18. ¹H NMR of **8** *cis*-[ReO(PEt₃)(sal₂ibn)][PF₆] in CD₃CN (500 MHz, calibrated to residual CHD₂CN at 1.94 ppm).

¹H NMR [600 MHz, CD₃CN, r.t., δ (ppm)]: 9.04 (d, *J* = 9.7 Hz, 1H, -HC=N-); 8.60 (s, 1H, -HC=N-); 7.75-7.79 (m, ArH, 3H); 7.52-7.55 (m, ArH, 1H); 7.35 (m, 1H, ArH); 7.24 (m, 1H, ArH); 7.09 (m, ArH, 1H); 6.98 (d, *J* = 8.5 Hz, 1H, ArH); 4.86 (d, *J* = 11.1 Hz, 1H, -NCH₂-C(CH₃)); 4.30 (d, *J* = 11.0 Hz, 1H, -NCH₂-C(CH₃)); 1.77-1.85 (m, 3H, PCH₂CH₃); 1.75 (s, 3H, -CH₃); 1.41-1.46 (m, 1H, PCH₂CH₃); 1.06-1.11 (m, 9H, PCH₂CH₃) 1.02 (s, 3H, -CH₃).





Figure S19. ¹H NMR of 9 *cis*-[ReO(PEt₂Ph)(sal₂ibn)][PF₆] in CD₃CN (500 MHz,

calibrated to residual CHD₂CN at 1.94 ppm).

¹H NMR [600 MHz, CD₃CN, r.t., δ (ppm)]: 8.94 (d, 1H, -HC=N-); 8.05 (s, 1H, -HC=N-); 7.82 (m, J = 1.4, 7.7 Hz, 1H, ArH); 7.72, 7.70 (dd, J = 1.8, 7.9 Hz, 1H, ArH); 7.50, 7.49 (d, J = 8.4 Hz, 1H, ArH); 7.47-7.45 (m, 1H, PPh *para*); 7.37-7.35 (m, 4H, PPh); 7.43, 7.42 (dd, J = 1.7, 8.0 Hz, 1H, ArH); 7.20 (m, J = 1.7, 6.3 Hz, 1H, ArH); 7.12 (m, J = 1.1, 7.5 Hz, 1H, ArH); 7.05 (m, J = 1.0, 7.5 Hz, 1H, ArH); 6.31, 6.29 (d, J = 8.5 Hz, 1H, ArH); 4.75, 4.73 (d, J = 11 Hz, 1H, -NCH₂-C(CH₃)); 4.33, 4.31 (d, J = 11 Hz, 1H, -NCH₂-C(CH₃)-); 2.20-2.08 (m, 4H, PCH₂CH₃); 1.76 (s, 3H, -CH₃); 1.15-1.02 (d of d of t, J = 7.5 Hz, 6H, PCH₂CH₃); 1.01 (s, 3H, -CH₃). 1H cis-[ReO(PEtPh2)(sal2ibn)][PF6]



Figure S20. ¹H NMR of **10** *cis*-[ReO(PEtPh₂)(sal₂ibn)][PF₆] in CD₃CN (500 MHz, calibrated to residual CHD₂CN at 1.94 ppm).

¹H NMR [500 MHz, CD₃CN, r.t., δ (ppm)]: 8.95 (d, *J* = 10 Hz, 1H, -HC=N-); 8.20 (s, 1H, -HC=N-); 7.87-6.99 (m, ArH, 8H); 7.53-7.36 (m, PPh, 10H); 4.69 (d, *J* = 11 Hz, 1H, -NCH₂-C(CH₃)); 4.33 (d, *J* = 11 Hz, 1H, -NCH₂-C(CH₃)); 2.68-2.58 (m, 1H, PCH₂CH₃); 2.25-2.15 (m, 1H, PCH₂CH₃); 1.73 (s, 3H, -CH₃); 1.15-1.02 (d of t, *J* = 7.5 Hz, 3H, PCH₂CH₃); 0.96 (s, 3H, -CH₃).



Figure S21. ¹H NMR of **11** *cis*-[ReO(PPh₃)(sal₂ibn)][PF₆] in CD₃CN (600 MHz, calibrated to residual CHD₂CN at 1.94 ppm).

¹H NMR [600 MHz, CD₃CN, r.t., δ (ppm)]: 8.96 (d, *J* = 10 Hz, 1H); 8.22 (-HC=N-); 7.84-7.74, 7.34-6.44 (m, ArH); 7.52-7.39 (m, PPh, 15H); 4.67 (d, *J* = 11 Hz, 1H, -NCH₂-C(CH₃)); 4.34 (d, *J* = 11 Hz, 1H, -NCH₂-C(CH₃)); 1.73 (s, 3H, -CH₃); 1.00 (s, 3H, -CH₃).



Figure S22. Repetitive cyclic voltammogram (CV) for *trans*- $[Re(PEt_3)_2(sal_2en)][PF_6]$ **2** in the region -1.3 to +0.6 V.



Figure S23. Repetitive CV for *trans*-[Re(PEtPh₂)₂(sal₂en)][PF₆] **3** in the region -1.2 to +0.6 V.



Figure S24. Repetitive CV for *trans*- $[Re(PPh_3)_2(sal_2en)][PF_6]$ **4** in the region +0.1 to +0.7 V.



Figure S25. Repetitive CV for *trans*-[Re(PPh₃)₂(sal₂en)][PF₆] **4** -1.05 to -0.65 V.



Figure S26. Repetitive CV for *trans*-[Re(PEt₃)₂(sal₂ibn)][PF₆] **5** in the region -1.3 to -0.8 V.



Figure S27. Repetitive CV for *trans*-[Re(PEt₃)₂(sal₂ibn)][PF₆] **5** in the region +0.1 to +0.6 V.



Figure S28. Repetitive CV for *trans*-[Re(PEtPh₂)₂(sal₂ibn)][PF₆] **6** in the region -1.2 to +0.6 V.