### **Electronic Supplementary Information**

# Dual-emitting Tb(III)&Yb(III) functionalized coordination polymer: "turn-on" sensor for N-methylformamide in urine and "turn-off" sensor for methylglyoxal in serum

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#### **Experimental Section**

#### Materials and methods

All chemicals employed were commercially available reagents of analytical grade and used without further purification. 1-(4-carboxylbenzyl)-3-(pyrzin-2-yl) (97%, Hcbpp) was purchased from Jinan Henghua Sci .& Tec .Co.,Ltd. Elemental analyses were carried out on a Perkin-Elmer 2400 automatic analyzer. FT-IR spectra data (4000-400 cm<sup>-1</sup>) were collected by a Nicolet impact 410 FT-IR spectrometer. The emission properties were recorded with Edinburgh FLS 920 fluorescence spectrometer equipped with a Peltier-cooled Hamamatsu R928 photomultiplier tube. An Edinburgh Xe900 450 W xenon arc lamp was used as an exciting light source. Thermal analysis was performed on a ZRY-2P thermogravimetric analysis from 30 to 700 °C with a heating rate of 10 °C min<sup>-1</sup> under a flow of air. Powder X-ray diffraction (PXRD) patterns were recorded in the 2 $\theta$  range of 5 – 50° using Cu K $\alpha$  radiation with a Shimadzu XRD-6000 X-ray diffractometer. Elemental analyses for Cu and Tb were conducted using a Perkin-Elmer Model Optima 3300 DV ICP spectrometer. Density functional theory (DFT) calculations were conducted by using the B3LYP/lanl2dz basis set implemented in the Gaussian 09 package. XPS experiments were carried out on a RBD upgraded PHI-5000C ESCA system (Perkin Elmer) with Mg Kα radiation (hv = 1253.6 eV). Lifetime studies were performed using photon-counting system with a microsecond pulse lamp as the excitation source. Data were analyzed through the nonlinear least squares procedure in combination with an iterative convolution method. The emission decays were analyzed by the sum of exponential functions. The decay curve is well fitted into a double exponential function:  $I = I_0 + A_1 \exp(-t/\tau_1) +$  $A_2 \exp(-t/\tau_2)$ , where I and  $I_0$  are the luminescent intensities at time t = t and t = 0, respectively, whereas  $\tau_1$  and  $\tau_2$  are defined as the luminescent lifetimes. The average

lifetime was calculated according to the following equation:  $\frac{\tau_1^2 A_1 \% + \tau_2^2 A_2 \%}{\tau_1 A_1 \% + \tau_2 A_2 \%}.$ 

#### X-ray data collection and structure determinations

X-ray single-crystal diffraction data of Cu-cbpp and Cu-Hcbpp were collected on a Rigaku SCX-mini diffractometer with graphite monochromatic Mo-K $\alpha$  radiation ( $\lambda$  = 0.71073 Å). The program CrystalClear was used for integration of the diffraction profiles. All the structures were solved by direct methods using the SHELXS program of the SHELXTL package and refined by full-matrix least-squares methods with SHELXL. Metal atoms were located from the E-maps and other non-hydrogen atoms excluded in conterions were located in successive difference Fourier syntheses and refined with anisotropic thermal parameters on F<sup>2</sup>. Further details of crystal data and structure refinement for Cu-cbpp and Cu-Hcbpp were summarized in Table S1.

Identification code	Cu-cbpp	Cu-Hcbpp
Empirical formula	$C_{30}H_{22}Cl_2Cu_2N_8O_4$	$C_{30}H_{24}Cl_4Cu_2N_8O_4$
CCDC	1920117	1920118
Formula mass	756.54	829.47
Crystal system	Monoclinic	Triclinic
Space group	$P 2_1/c$	Р
<i>a</i> (Å)	7.213(6)	7.305(9)
<i>b</i> (Å)	12.897(11)	8.862(11)
<i>c</i> (Å)	15.843(13)	24.838(3)
α (°)	90.00	92.59(2)
eta (°)	95.39(10)	97.87(2)
γ (°)	90.00	96.67(2)
V (Å <sup>3</sup> )	1467.3(2)	1578.8(3)
Ζ	2	2
$D_{\rm c}/({\rm g~cm^{-3}})$	1.712	1.745
$\mu$ (Mo K $\alpha$ )/mm <sup>-1</sup>	1.685	1.738
<i>F</i> (000)	764	836
$\theta$ range (°)	2.58 - 27.65	0.83 - 25.00
Limiting indices	$-8 \le h \le 9$	$-8 \le h \le 8$
	$-15 \le k \le 16$	$-9 \le k \le 10$
	$-18 \le l \le 20$	$-29 \le l \le 29$
Data/Restraints/Parameters	3365 / 0 / 208	5550 / 0 / 433
GOF on $F^2$	1.038	1.056
$R_1^{a}$	0.0322	0.0373
$wR_2^{b}$	0.0812	0.0913
$R_1$	0.0434	0.0580
$wR_2$	0.0865	0.1113

Table S1 Crystal data and structure refinement parameters

 ${}^{a}R_{1} = \sum ||F_{o}| - |F_{c}|| / \sum |F_{o};$ 

<sup>b</sup> wR<sub>2</sub> = [ $\sum [w (F_o^2 - F_c^2)^2] / \sum [w (F_o^2)^2]$ ]<sup>1/2</sup>.

Cu-cbpp			
Cu(1)-N(2)	2.006(18)	N(2)-Cu(1)-O(2)	162.18(7)
Cu(1)-O(2)	2.010(17)	N(2)-Cu(1)-O(1)	97.86(7)
Cu(1)-O(1)	2.046(17)	O(2)-Cu(1)-O(1)	64.60(7)
Cu(1)-N(3)	2.176(18)	N(2)-Cu(1)-N(3)	78.56(7)
Cu(1)-Cl(1)	2.228(7)	O(2)-Cu(1)-N(3)	101.04(7)
		O(1)-Cu(1)-N(3)	101.69(7)
		N(2)-Cu(1)-Cl(1)	96.58(6)
		O(2)-Cu(1)-Cl(1)	99.68(5)
		O(1)-Cu(1)-Cl(1)	143.01(6)
Cu-Hcbpp			
Cu(1)-N(1)	2.046(3)	N(3)-Cu(1)-Cl(1)	170.32(9)
Cu(1)-N(3)	2.065(3)	Cl(2)-Cu(1)-Cl(1)	92.18(4)
Cu(1)-Cl(2)	2.2371(10)	N(1)-Cu(1)-Cl(4)	99.03(9)
Cu(1)- $Cl(1)$	2.2716(10)	N(3)-Cu(1)-Cl(4)	89.51(9)
Cu(1)-Cl(4)	2.8791(12)	Cl(2)-Cu(1)-Cl(4)	96.06(4)
Cu(2)-N(5)	2.047(3)	Cl(1)-Cu(1)-Cl(4)	89.12(4)
Cu(2)-N(7)	2.076(3)	N(5)-Cu(2)-N(7)	79.81(12)
Cu(2)-Cl(4)	2.2526(11)	N(5)-Cu(2)-Cl(4)	175.14(9)
Cu(2)-Cl(3)	2.2586(10)	N(7)-Cu(2)-Cl(4)	97.83(9)
Cu(2)-Cl(1)	2.9330(12)	N(5)-Cu(2)-Cl(3)	90.86(9)
		N(7)-Cu(2)-Cl(3)	168.59(9)
		Cl(4)-Cu(2)-Cl(3)	91.94(4)
N(1)-Cu(1)-N(3)	79.84(12)	N(5)-Cu(2)-Cl(1)	87.38(9)
N(1)-Cu(1)-Cl(2)	164.64(9)	N(7)-Cu(2)-Cl(1)	83.75(8)
N(3)-Cu(1)-Cl(2)	97.49(9)	Cl(4)-Cu(2)-Cl(1)	88.14(4)
N(1)-Cu(1)-Cl(1)	90.92(9)	Cl(3)-Cu(2)-Cl(1)	102.52(4)

Table S2 Selected bond lengths (Å) and bond angles (°)



**Fig. S1** The structural unit of Cu-cbpp with labeling scheme and 50% thermal ellipsoids (hydrogen atoms are omitted for clarity)



**Fig. S2** The structural unit of Cu-Hcbpp with labeling scheme and 50% thermal ellipsoids (hydrogen atoms are omitted for clarity)



Fig. S3 PXRD patterns of Tb@Cu-Hcbpp in different pH value (from 3 to 11)



**Fig. S4** Electron distribution and energy diagram of HOMO-1 and LUMO+1 orbitals for Cu-cbpp and Cu-Hcbpp.



Fig. S5 Fluorescence spectra of Tb@Cu-Hcbpp for 3 days



Fig. S6 PXRD patterns of Tb@Cu-Hcbpp in different conditions



Fig. S7 UV-vis spectra of various organic solvents



Fig. S8 UV-vis spectra of Tb@Cu-Hcbpp in related circumstances



Fig. S9 Fluorescence response of Hcbpp in NMF



**Fig. S10** FT-IR spectra of  $Tb_{1-x}Yb_x@Cu-Hcbpp$  (x =0, 0.05, 0.10, 0.15, 0.20, 0.25, 0.30, 0.35 and 0.40)



Fig. S11  $Yb^{3+}$  ions emission of  $Tb_{0.85}Yb_{0.15}$  (Cu-Hcbpp with MGO, LA or PA.



Fig. S12 The molecular structure of MGO, PA and LA.



Fig. S13 FT-IR spectra of Cu-cbpp



Fig. S14 Thermal gravimetric curves of Cu-cbpp



Fig. S15 PXRD patterns of Cu-cbpp

Samples	Lifetime (µs)
Tb@Cu-Hcbpp	194.74
Tb@Cu-Hcbpp with NMF	399.61
Tb@Cu-Hcbpp with urea	196.19
Tb@Cu-Hcbpp with uric acid (UA)	195.66
Tb@Cu-Hcbpp with creatine	192.53
Tb@Cu-Hcbpp with creatinine (Cre)	195.36
Tb@Cu-Hcbpp with glucose (Glu)	193.17
Tb@Cu-Hcbpp with NaCl	194.23
Tb@Cu-Hcbpp with KCl	194.65
Tb@Cu-Hcbpp with NH <sub>4</sub> Cl	195.01
Tb@Cu-Hcbpp with Na <sub>3</sub> PO <sub>4</sub>	191.29

## Table S3 Lifetime of Tb@Cu-Hcbpp in different urine chemicals

No.	Fluorescence intensity (I <sub>386</sub> )	Fluorescence intensity $(I_{544})$	$I_{544}/I_{386}$
1	347.60 a.u.	380.37 a.u.	1.094275
2	345.47 a.u.	379.37 a.u.	1.098127
3	346.50 a.u.	379.95 a.u.	1.096536
4	345.41 a.u.	379.48 a.u.	1.098636
5	347.15 a.u.	380.23 a.u.	1.095290
Average Value			1.096573
Standard Deviation ( $\sigma$ )			0.000595
Slope (S) for NMF			0.08 µM <sup>-1</sup>
Detection limit $(3\sigma/S)$ for	or NMF		0.02 µM
Slope (S) for MGO			0.0072µM <sup>-1</sup>
Detection limit $(3\sigma/S)$ for	or MGO		0.25µM

 Table S4 Standard deviation calculation

Table S5	Selected	materials	for	sensing NMF
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No.	Complexes	Detection limit	References
1	Eu(III)@MOF-1	0.36 µM	Sens. Actuators, B 2018, 261, 153 <sup>1</sup>
2	[Eu <sub>0.1</sub> Tb <sub>1.9</sub> (FDA) <sub>3</sub> (DMF) <sub>2</sub> ]·2DMF		Inorg. Chem. Front. 2018, 5, 2971 <sup>2</sup>
3	Tb@Cu-Hcbpp	0.02 μΜ	This work

No.	Complexes	Detection limit	References
1	Methyl diaminobenzene-BODIPY		J. Am. Chem. Soc. 2013, 135,
	or MBo		12429. <sup>3</sup>
2	Zr-UiO-66-N <sub>3</sub>	118 µM	Anal. Chem. 2015, 87, 1757.4
3	PDN-1	77 nM	<i>Anal. Methods</i> 2015, 7, 2386. <sup>5</sup>
4	Pt/CeO <sub>2</sub> /GLO	2.14 nM	Anal. Methods 2016, 8, 2207.6
5	L		Dyes Pigm. 2017, 138, 23. <sup>7</sup>
6	V <sub>2</sub> O <sub>5</sub> modified electrode	0.24 μM	Biosens. Bioelectron. 2018, 103,
			143.8
7	CMFP	0.5 μΜ	Anal. Chem. 2019, 91, 5646.9
8	Tb <sub>0.85</sub> Yb <sub>0.15</sub> @Cu-Hcbpp	0.25 μM	This work

### Table S6 Selected materials for sensing MGO

Samples	Lifetime (µs)
Tb <sub>0.85</sub> Yb <sub>0.15</sub> @Cu-Hcbpp	477.59
Tb <sub>0.85</sub> Yb <sub>0.15</sub> @Cu-Hcbpp with MGO	223.13
Tb <sub>0.85</sub> Yb <sub>0.15</sub> @Cu-Hcbpp with PA	485.63
Tb <sub>0.85</sub> Yb <sub>0.15</sub> @Cu-Hcbpp with LA	472.38
Tb <sub>0.85</sub> Yb <sub>0.15</sub> @Cu-Hcbpp with AA	473.42
Tb <sub>0.85</sub> Yb <sub>0.15</sub> @Cu-Hcbpp with Cys	475.94
Tb <sub>0.85</sub> Yb <sub>0.15</sub> @Cu-Hcbpp with Ala	475.33
Tb <sub>0.85</sub> Yb <sub>0.15</sub> @Cu-Hcbpp with Glu	473.53
Tb <sub>0.85</sub> Yb <sub>0.15</sub> @Cu-Hcbpp with UA	475.98
Tb <sub>0.85</sub> Yb <sub>0.15</sub> @Cu-Hcbpp with NaCl	477.19
Tb <sub>0.85</sub> Yb <sub>0.15</sub> @Cu-Hcbpp with KCl	476.73

### **Table S7** Lifetime of $Tb_{0.85}Yb_{0.15}$ @Cu-Hcbpp in different serum chemicals

### Note and references

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