## **Electronic Supplementary Information**

## A "turn-on" sensor based on MnO<sub>2</sub> coated UCNPs for detection of alkaline phosphatase and ascorbic acid

Mei-yu Liang, Bing Zhao, Yan Xiong, Wen-xin Chen, Jian-zhong Huo, Fei Zhang, Lu Wang, Yan Li\*

Key Laboratory of Inorganic-Organic Hybrid Functional Material Chemistry (Tianjin Normal University), Ministry of Education, Tianjin Key Laboratory of Structure and Performance for Functional Molecule, College of Chemistry, Tianjin Normal University, 393 Binshui West Road, Tianjin 300387, PR China

\* Corresponding authors. E-mail address: nkliyan398@gmail.com (Y. Li).



Fig. S1 XRD pattern of NaYF<sub>4</sub>:Yb<sup>3+</sup>, Tm<sup>3+</sup> UCNPs which reflections were coincident with the diffraction peaks of the  $\beta$ -phase.

phase	ICP-MS tested result		The molar ratios of	
	Y	Yb Tm	Y: Yb: Tm	
β	19.72%	8.04% 0.11%	77.54%: 16.27%: 0.22%	

**Table S1** ICP-MS quantitative analysis of molar ratios of Y: Yb: Tm in the sample.



Fig. S2 FT-IR spectra of  $NaYF_4$ : Yb<sup>3+</sup>, Tm<sup>3+</sup> UCNPs (curve a) and pure PEI (curve b).



Fig. S3 Energy dispersive X-ray spectroscope (EDS) spectrum of  $MnO_2$  coated UCNPs.



**Fig. S4** (A) Luminescence decay curves of UCNPs (black line) and MnO<sub>2</sub> coated UCNPs (red line). (B) Luminescence decay curves of UCNPs and MnO<sub>2</sub> coated UCNPs and corresponding mono-exponential fitting based on  $y=y_0+A_1*exp[-(x-x_0)/\tau_1]$  (red line and blue line).



**Fig. S5** (A) The restored luminescence intensity of the sensing system against the different concertrations of AAP in the presence of ALP (100 mU mL<sup>-1</sup>). (B) The restored luminescence intensity of the sensing system against the incubation time of ALP (100 mU mL<sup>-1</sup>) and AAP (6 mM). (C) The restored luminescence intensity of the sensing system against the varying pH of ALP (100 mU mL<sup>-1</sup>) and AAP (6 mM). (D) The luminescence reaction time of the sensing system.

Assay method		Detection limit	Ref.
Specific determination of ascorbic acid with high-performance liquid chromatography		1 pM	1
Determination of ascorbic acid by capillary electrophoresis with electrochemical detection		2×10-7 g mL-1	2
Bimodal electrochemiluminescence of G-CNQDs for ascorbic acid detection		110 pM	3
Colorimetric detection of ascorbic acid using Pt/CeO <sub>2</sub> nanocomposites		80 nM	4
A MnO <sub>2</sub> nanosheet-based ratiometric fluorescent nanosensor for detection of ascorbic acid		10 nM	5
Graphene oxide nanosheets functionalized with poly(styrene sulfonate) for colorimetric assay for ascorbic of acid		0.15 μΜ	6
An "on-off-on" fluorescent nanoprobe for recognition of ascorbic acid based on carbon quantum dot		1.35 μM	7
Near-infrared quantum dots-gold nanoclusters nanohybrid for ratiometric fluorescent detection of ascorbic acid		1.5 µM	8
Based on MnO <sub>2</sub> coated UCNPs for detection of alkaline phosphatase and ascorbic acid		0.29 μΜ	This work

## **Table S2** Comparison of different detection methods of AA with the linear range and the detection limit

## References

- 1 E. Kishida, Y. Nishimoto and S. Kojo, Anal. Chem., 1992, 64, 1505-1507.
- 2 Y. Peng, Y. Zhang and J. Ye, J. Agr. Food Chem., 2008, 56, 1838-1844.
- 3 H. Wang, G. Pu, S. Devaramani, Y. Wang, Z. Yang, L. Li, X. Ma and X. Lu, *Anal. Chem.*, 2018, **90**, 4871-4877.
- 4 X. Liu, X. Wang, C. Qi, Q. Han, W. Xiao, S. Cai, C. Wang and R. Yang, *Appl. Surf. Sci.*, 2019, **479**, 532-539.
- 5 Y. Lyu, Z. Tao, X. Lin, P. Qian, Y. Li, S. Wang and Y. Liu, *Anal. Bioanal. Chem.*, 2019, **411**, 4093-4101.
- 6 J. Chen, J. Ge, L. Zhang, Z. Li, J. Li, Y. Sun and L. Qu, *Microchim. Acta*, 2016, **183**, 1847-1853.
- 7 X. Gong, Y. Liu, Z. Yang, S. Shuang, Z. Zhang and C. Dong, *Anal. Chim. Acta*, 2017, **968**, 85-96.
- 8 P. Zhao, K. He, Y. Han, Z. Zhang, M. Yu, H. Wang, Y. Huang, Z. Nie and S. Yao, Anal. Chem., 2015, 87, 9998-10005.