## 1 Stevia residue extract increases intestinal uric acid excretion via interacting with

## 2 intestinal urate transporters in hyperuricemic mice

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#### **36 Preparation of STVRE**

The Stevia leaves powder was extracted twice with 18 volumes of distilled water at 37 65°C for 1.5 hr and then filtrated. The filtrate was mixed with ferrous sulfate (0.9% 38 w/v, final concentration) and adjusted to pH 9.0-10.0 with calcium hydroxide. After 39 flocculation at 50°C for 30 min, the mixture was filtrated to obtain the Stevia residue. 40 The Stevia residue was then extracted three times with acidified 95% (v/v) acetone 41 (pH 1.5) at 45°C for 1 hr and filtrated. The filtrate was evaporated under reduced 42 pressure to remove acetone, and thereafter purified by extraction with ethyl acetate. 43 The partially purified extract was applied to an HZ841 macroporous resin column, 44 and then eluted it with 2 bed volume (BV) of distilled water, followed by 2 BV of 45 alkaline water (pH 9.5) at a flow rate of 1 BV/hr to remove impurities. Subsequently, 46 the adsorbed components were eluted with 3 BV of 17% (v/v) ethanol, followed by 47 1.5 BV of 50% (v/v) ethanol. The eluents were concentrated and lyophilized to obtain 48 SRE1 and SRE2, respectively. The resulting STVRE powder was stored at 49 -20°C prior 50 to use (1, 2).

Qualitative and quantitative analysis of the major bioactive compounds in SREs 51 The major bioactive compounds in SREs were identified by HPLC- ESI-QTOF-52 MS/MS (6530 HPLC-MS system; Aglient Technologies, Santa Clara, CA, USA) and 53 quantified by HPLC (1290 HPLC system; Aglient Technologies, Santa Clara, CA, 54 USA). The HPLC parameters were as follows: chromatographic column, Agilent 55 Eclipse Plus C18 column (50 mm × 2.1 mm i.d., 1.8 µm; Aglient Technologies, Santa 56 Clara, CA, USA); mobile phase, 0.1% formic acid in water (A) and methanol (B); 57 flow rate, 0.1 mL/min; elution procedure, 0–10 min, 10%–50% B; 10–12 min, 50%– 58

10% B; 12-15 min, 10% B; detection wavelength, 320 and 360 nm; column 59 temperature, 30°C; volume of sample injection, 1 µL. Authentic compounds 60 (3-O-caffeoylquinic acid, 4-O-caffeoylquinic acid, caffeic acid, quercetin, 61 3,4-O-dicaffeoylquinic acid, 3,5-O-dicaffeoylquinic acid, 4,5-O-di-caffeoylquinic 62 acid and quercetin-3-O-rhamnoside) were dissolved in methanol; the concentrations 63 were adjusted to 6.25, 12.5, 25, 50, 100, 200 and 400 µg/mL, respectively, and the 64 calibration curves were established under the same determination conditions. Mass 65 spectrometric detection was set as follows: Negative ion mass spectra of the column 66 eluate were recorded in the range of  $m/z \ 100-1500 \ (1, 2)$ . 67

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# 71 Supplementary Table 1. Retention times (Rt), MS and MS2 fragmentation ions of

72 the phenolic compounds in Stevia residue extract (STVRE).

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Compou nds	Rt (min )	Pseudomolec ular ion [M– H] <sup>–</sup> (m/z)	MS2 (m/z)	Tentative identification	mg/ml
1	3.8	353.0878	191.0571 (100), 179.0357 (72), 135.0451 (9)	3-O-Caffeoylquinic acid	$1.68 \pm 0.75$
2	4.2	353.0878	191.0567 (100), 179.0353 (76), 173.0431 (4), 135.0442 (13)	4-O-Caffeoylquinic acid	$3.79 \pm 0.41$
3	4.4	179.0350	135.0456 (100)	Caffeic acid	$8.54 \pm 0.82$
4	7.9	515.1195	353.0916 (100), 335.0823 (1), 191.0575 (12), 179.0361 (9), 135.0443 (2)	3,4-O-Dicaffeoylquinic acid	232.75 ± 2.17
5	8.0	515.1195	353.0914 (100), 335.0771 (1), 191.0564 (5), 179.036 (14), 173.0467 (24), 135.0440 (2	3,5-O-Dicaffeoylquinic acid	92.43 ± 0.97
6	9.1	515.1195	353.0921 (100), 191.0587 (4), 179.0365 (21), 173.0462 (19), 135.0459 (2)	4,5-O-Dicaffeoylquinic acid	265.94 ± 2.63
7	9.3	447.0933	301.0373 (100)	Quercetin-3-O-rhamnoside	$125.97 \pm 1.13$
8	11.0	301.0354	107.0139 (8), 121.0301 (11), 151.0042 (100), 178.9994 (72)	Quercetin	$3.05 \pm 0.63$
				Total chlorogenic acids	605.13 ± 5.66
				Total flavonoids	129.02 ± 20.61

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85 Supplementary Table 2. List of primer sequences uses in this study

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Gene Name	forward	reverse
ABCG2	TAAATGGAGCACCTCAACCT	GAGATGCCACGGATAAACTG
GLUT9	GAGATGCTCATTGTGGGACG	GTGCTACTTCGTCCTCGGT
GAPDH	GTGAAGGTCGGTGTGAACGG	GTGATGGCATGGACTGTGGTC

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### 88 Supplementary Table 3. Effect of STVRE on body weight of fructose-PO-YE

89 induced hyperuricemic mice (n = 8)

Treatments groups	Body Weight (g)	
GN	$52.23 \pm 2.35$	
GB	$49.86 \pm 4.56$	
GM	$50.56 \pm 1.93$	
GS1	$51.53 \pm 3.57$	
GS2	$50.08 \pm 2.98$	

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91 GN: normal group, GM: model group, GB: benzbromarone group (20 mg/kg bw),

92 GS1: stevia residue extract (200 mg/kg bw) and GS2: stevia residue extract (400 93 mg/kg bw)

94 #p<0.05, ##p<0.01, ###p<0.001 compare with GM, \*p<0.05, \*\*p<0.01, \*\*\*p<0.001</p>
95 compare with GN

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L. Zhao, X. Wang, M. Xu, Y. Lian, C. Wang, H. Yang, A. Mehmood, Dietary
 intervention with Stevia residue extracts alleviates impaired glucose regulation
 in mice. J Food Biochem, 2018, 42, e12651.

- L. Zhao, H. Yang, M. Xu, X. Wang, C. Wang, Y. Lian, *Stevia* residue extract
   ameliorates oxidative stress in d-galactose-induced aging mice via
   Akt/Nrf2/HO-1 pathway. J Funct Foods, 2019, 52, 587-595.