## SUPPLEMENTARY INFORMATION





epoxidation followed by spontaneous rearrangement into trans-sobrerol.



**Figure S 2**. GC chromatogram of P450-BM3/2M catalyzed conversion of (-)-α-pinene (2 mM) into *trans*-sobrerol. The enzymatic reaction (top) is compared with commercial *trans*-sobrerol dissolved in ethyl acetate (*trans*-sobrerol reference, middle). As negative control (bottom), reaction buffer instead of P450-BM3/2M was prepared an incubated under the same conditions.



**Figure S 3.** Fragmentation pattern of *trans*-sobrerol (theoretical mass of 170 g/mol) from P450-BM3/2M catalyzed synthesis (Enzymatic *trans*-sobrerol) compared to the fragmentation pattern of synthetic *trans*-sobrerol (*trans*-sobrerol reference).



**Figure S 4**. GC-FID chromatogram of the of the lipase catalyzed conversion of *trans*-sobrerol into *trans*-sobreryl methacrylate (Enzymatic reaction) compared with synthetic *trans*-sobreryl methacrylate and commercially available Sobrerol. Conversion of 96% was detected



**Figure S 5**. Mass spectrum of *trans*-sobreryl methacrylate (theoretical mass of 238 g/mol) from the lipase catalyzed synthesis (Enzymatic *trans*-sobreryl methacrylate) compared to the mass spectrum of synthetic *trans*-sobreryl methacrylate.

Reaction time	60 °C (Conversion	70 °C (Conversion	80 °C (Conversion	90 °C (Conversion
[h]	SobMA[%])	SobMA[%])	SobMA[%])	SobMA[%])
0	n.d.	0	n.d.	4
24	32	71	86	75
48	68	77	97	86
72	70	83	98	91

**Table S 1.** Enzymatic transformation of Sob into SobMA at different temperatures. reactions were performed using 100 mg of Sob, 130 mg of lipase and 4 ml of VMA.

Table S 2. Enzymatic transformation of Sob into SobMA using different monomer ratios.

1		6 6	6 1	
	Sob:VMA 1:20	Sob:VMA 1:30	Sob:VMA 1:40	Sob:VMA 1:56
Reaction time	(Conversion	(Conversion	(Conversion	(Conversion
[h]	SobMA[%])	SobMA[%])	SobMA[%])	SobMA[%])
0	8	10	9	0
17	73	72	63	55
24	82	81	69	66
48	92	92	85	81
72	96	96	91	89

reactions were performed at 80 °C using 100 mg of Sob and 130 mg of lipase.



**Figure S 6**. <sup>1</sup>H NMR spectra showing the conversion of **SobMA** at different time points under RAFT polymerization conditions. Key signals are annotated.



**Figure S 7.** Polymerization kinetics of SobMA. (a) Time-dependent monomer conversions. (b)  $ln([M_0]/[M])$  vs time plots. SEC traces for free-radical (c), RAFT (d) and ATRP (e) polymerizations.



**Figure S 8.** Evolution of the molecular weights and dispersities of free-radical (a), RAFT (b), and ATRP (c) polymerizations.



**Figure S 9**. <sup>1</sup>H NMR spectra displaying the conversion of **SobMA** at different time points under free-radical polymerization conditions. Key signals are annotated.



**Figure S 10**. <sup>1</sup>H NMR spectra showing the conversion of **SobMA** at different time points under ATRP polymerization conditions. Key signals are annotated.



formation of  $P(SobMA_{R100}-b-MMA_{R100})$ . Samples taken before the MMA feed (6 h, maroon) and after the full polymerization time (24 h, black).



formation of  $P(SobMA_{R100}-b-BMA_{R100})$ . Samples taken before the BMA feed (6 h, maroon) and after the full polymerization time (24 h, black).



**Figure S 13.** Molecular weight distributions during the polymerization of  $P(SobMA_{R100}-b-MMA_{R100})$ ; before the methyl methacrylate feed (6 h) and after the full polymerization time (24 h).



**Figure S 14**. Molecular weight distributions during the polymerization of  $P(SobMA_{R100}-b-BMA_{R100})$ ; before the methyl methacrylate feed (6 h) and after the full polymerization time (24 h).



Figure S 15. <sup>1</sup>H NMR spectrum of P(SobMA<sub>R100</sub>-*b*-MMA<sub>R100</sub>) in CDCl<sub>3</sub>.



Figure S 16. <sup>1</sup>H NMR spectrum of P(SobMA<sub>R100</sub>-*b*-BMA<sub>R100</sub>) in CDCl<sub>3</sub>.



Figure S 17. <sup>1</sup>H NMR spectrum of P(SobMA<sub>R100</sub>-st-MMA<sub>R100</sub>) in CDCl<sub>3</sub>.



Figure S 18. <sup>1</sup>H NMR spectrum of P(SobMA<sub>R100</sub>-st-BMA<sub>R100</sub>) in CDCl<sub>3</sub>.



Figure S 19. FTIR spectra of poly(sobreryl methacrylates).



Figure S 20. <sup>1</sup>H NMR spectrum showing the precipitated PSobMA<sub>E</sub>.



Figure S 21. <sup>1</sup>H NMR spectrum of P(SobMA<sub>E25</sub>-st-BMA<sub>E75</sub>) in CDCl<sub>3.</sub>



Figure S 22. Molecular weight distribution of P(SobMA<sub>E25</sub>-st-BMA<sub>E75</sub>).



**Figure S 23.** Degradation thermograms (TGA) for sobreryl methacrylate homo (solid lines) and co-polymers (dashed lines) at a heating rate of 10 °C/min.



**Figure S 24**. DSC thermograms from the second heating of sobreryl methacrylate homo (solid lines) and co-polymers (dashed lines) at a heating rate of 10 °C/min.



Figure S 25. FTRaman spectroscopy of  $PSobMA_{R50}$  and its corresponding coating via thiol-ene chemistry.



**Figure S 26.** FTRaman spectroscopy of  $P(SobMA_{R25}-st-BMA_{R75})$  and its corresponding coating via thiol-ene chemistry



**Figure S 27**. Solubility test of the crosslinked  $PSobMA_{R50}$  and  $P(SobMA_{R25}-st-BMA_{R75})$  using TMTP. Polymers (c, d, g and h) and crosslinked films (a, b, e and f) were added into vials together with THF (a, c, e and g) and DCM (b, d, f and h) and shaked for 24h.



**Figure S 28.** FT-IR spectroscopy of PSobMA<sub>R50</sub>,  $P(SobMA_{E25}-st-BMA_{E75})$  and its corresponding coating via transetherification.



**Figure S 29.** Solubility test of the crosslinked  $PSobMA_{R50}$  and  $P(SobMA_{E25}$ -*st*-BMA<sub>E75</sub>) using HMMM. Crosslinked films (a, b) and polymers (c, d) and were added into vials together with THF and shaked for 24h.

Primer	Sequence
A74G fw	ATGAATCACGCTTTGATAAAAACTTAAGTCAAggcCTTAAATTTGTACG
A74G rev	TAACCCGTCTCCTGCAAAATCACGTACAAATTTAAGgccTTGACTTAAG
	<u> </u>
T 1000 C	
L188Q fw	TATGGTCCGTGCACTGGATGAAGCAATGAACAAG <u>cag</u> CAGCGAGC
I 1990 rou	
LIGOVIEV	

Table	<b>S 3</b> .	Listed	primers	used	for site	directed	mutagenesis.

 Table S 4. Site directed mutagenesis temperature program.

Program phase	Temperature	Time	Number of cycles
	(°C)	(min)	

Pre-phase	95	2	1
Initialize	95	0.33	
Annealing of primer	$T^*$	0.33	30
Elongation	72	2.13	
Final elongation	72	3 min	
End	4	$\infty$	1

\*Annealing temperatures: T = 58 °C for A74G mutation and T = 64 °C for L188Q mutation.





Figure S 30. SDS-PAGE of purification fractions of P450-BM3/2M. Purification fractions,

analyzed by SDS-PAGE (10% Mini PROTEAN TGX-gels, BioRad). S = supernatant, FT = flow through, W1 = wash buffer A, W2 = wash buffer B, E1-E5 = elution fractions 1 to 5.