Electronic Supplementary Information

Highly Efficient and Practical Aerobic Oxidation of Alcohols with by Inorganic-ligand Supported Copper Catalysis

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Table of contents

I.	General information	3
II.	Preparation of inorganic-ligand supported copper-catalyst	3
III.	FT-IR spectra of catalyst 1	4
IV.	XRD spectra of catalyst 1	4
V.	General procedure for catalytic oxidation of alcohols	5
VI.	Optimization of reaction conditions	5
VII.	Recycling experiments of catalyst for oxidative reaction	11
VIII.	ESI-Ms spectra of catalyst 1 intermediate	13
IX.	References	14
X.	NMR data of products	14
XI.	NMR spectra	20

I. General information

The catalysts was prepared according to published literature methods.^[1,2] All reagents were purchased from Sigma-Aldrich and Adamas-beta, which were used without further purification. ¹H and ¹³C Nuclear Magnetic Resonance (NMR) spectra were recorded on Bruker AVANCE III 500 MHz (500 MHz for proton, 125MHz for carbon) spectrometer and Bruker AVANCE III 400MHz (400MHz for proton, 100MHz for carbon) with tetra-methylsilane as the internal reference using CDCl₃ or DMSO-d₆ as solvent in all cases, and chemical shifts were reported in parts per million (ppm, δ). FT-IR spectra were recorded on a Thermo Fisher Nicolet 6700. ESI-MS was performed on Analysis Center of Shanghai Institute of Technology. GC mass spectra were recorded on Shimadzu GCMS-QP2010 with RTX-5MS column (0.25 mm× 30 m). Column chromatography was performed using 300-400 mesh silica gel.

II. Preparation of inorganic-ligand supported copper-catalyst

 $(NH_4)_4[CuMo_6O_{18}(OH)_6]\cdot 5H_2O$ was synthesized according to a published procedure^[1,2] with modification: $(NH_4)_6Mo_7O_{24}\cdot 4H_2O$ (5.3g) was dissolved in water (80 ml) and then heated to 100°C. $CuSO_4\cdot 5H_2O$ (0.99g) was dissolved in water (20ml), which was slowly added into the above solution under stirring. The mixture solution was kept stirring for 1h after $CuSO_4$ being completely added. On cooling, the mixture solution was filtered to remove any insoluble precipitate produced, and the filtrate was allowed to stand at room temperature for crystalization without disturbance. Light blue-green crystalline aim product (3.1g) was deposited and collected within a week. (Figure. S1) IR: 1631.65 (δ OH m), 1400.56 (δ NH, s), 931.62(v Mo=O, vs), 897.66(v Mo=O, vs), 640.18 (v Mo-O-Mo, vs), 577.68 (v M-O-Mo, w) cm⁻¹.



Figure. S1 Appearance of $(NH_4)_4[CuMo_6O_{18}(OH)_6]$ ·5H₂O

III. FT-IR spectra of catalysts 1



Figure S2. FT-IR spectra of catalyst 1

IV. XRD spectra of catalyst 1



Figure S3. XRD spectra of catalyst 1

V. General procedure for catalytic oxidation of alcohol

Aerobic oxidation reactions of alcohols were carried out in 2 ml aqueous solution containing 1 mmol alcohols and 10 mol% NaCl, together with 1.0 mol% cat. **1.** in test tubes. Meanwhile, oxygen gas balloonswere utilized to fulfill the reaction tubes with oxygen, and the reactions were kept at 60 °C for 6-24 hours. The reaction yield was determined *via* gas chromatography-mass spectrometry (GC-MS). When the reaction finished, a small amount of ethyl acetate was added into the reaction mixture and the solution was quickly filtered. The filtered solid was washed with ether, dried and redispersed for recyclability tests. Finally, the solvent was removed in vacuum, and the corresponding aldehydes were purified by washing through silica gel column. (EA:PE=1:6 or 1:10)

VI. Optimization of reaction conditions

$\begin{array}{c} O \\ O \\ O \\ O \end{array} + O_2(1 \text{ atm}) \\ (\text{balloon}) \end{array} \xrightarrow{\begin{array}{c} \textbf{Cat.1} (1.0 \text{ mol\%}) \\ \text{Solvent (2.0 ml)} \\ 60 \text{ °C, t} \end{array}} \xrightarrow{\begin{array}{c} O \\ O \\ O \end{array} \xrightarrow{\begin{array}{c} O \\ O \\ O \end{array}} \xrightarrow{\begin{array}{c} O \\ O \\ O \end{array}}$				
Entry	Solvent	Time (h)	Sel. (%) ⁺	Yield (%)
1	H ₂ O	24	99	72
2	CH₃CN	16	99	66
3	DCM	12	99	60
4	Dioxane	15	93	32
5	DMF	20	90	30
6	DMSO	20	86	<10
7	H_2O/CH_3CN	12	99	90
8	H ₂ O/DCM	10	99	81
9	$H_2O/Dioxane$	14	95	52
10	H ₂ O/DMF	16	99	46
11	H ₂ O/DMSO	18	99	17

Table S1. The effect of solvent.^[a]

[a] Reaction conditions: Cat. 1(1.0 mol%), alcohol (2.0 mmol), O_2 (1 atm), and solvent (2 mL) at 60°C, Volume ratio: v/v=1/1, unless otherwise noted. [b], [c] Selectivity and yields were calculated by GC and confirmed by GC-MS.

Table S2. The effect of water. [a]

	$\begin{array}{c} O \\ O \\ O \\ O \end{array} \begin{array}{c} O \\ O \\ O \\ O \end{array} \begin{array}{c} O \\ O $	$\begin{array}{c} 1 (1.0 \text{ mol}\%) \\ 1_3 \text{CN} = X (2 \text{ ml}) \\ 0 \text{ °C, 12h} \end{array}$	°0
Entry	H_2O/CH_3CN (v : v)	Sel.(%) ^[b]	Yield (%) ^[c]
1	0.5:1	97	83
2	1:1	>99	90
3	2:1	99	87
4	3:1	99	87
5	4:1	95	85
6	5:1	92	83

[a] Reaction conditions: Cat. 1(1.0 mol%), alcohol (2.0 mmol), O_2 (1 atm), and solvent (2 mL) at 60°C, Volume ratio: v/v=1/1, unless otherwise noted. [b], [c] Selectivity and yields were calculated by GC and confirmed by GC-MS.



Figure S4. The effect of solvent

O OF	$\begin{array}{c} \text{Cat. I (1.0 mol)} \\ \text{NaCl (10 mol)} \\ \text{H}_2\text{O/CH}_3\text{CN} = 1 \\ \text{(balloon)} \\ \end{array}$	$\frac{\sqrt{6}}{1:1}$ 0 +	ОН
7a		7	7b
 Time (h)	Recovery of	piperinic aldehyde	piperinic acid (%) ^d
2	97	-	-
4	95	3	-
6	95	4	-
8	94	4	-
10	93	5	-
12	92	5	-
14	90	7	-
16	89	7	-
18	88	8	-
20	86	10	-
22	85	12	-
24	82	13	_

Table S3. Monitoring the reaction of piperitol with NaCl and 18-Crown-6^[a]

[a] The reaction was carried out on a 10 mmol scale of piperitol in 40 mL of H_2O/CH_3CN (1:1). The reaction mixtures were calculated by GC and confirmed by GC-MS once per hour. [b],[c],[d] Yields were calculated from GC-Ms.



Figure S5. Monitoring the reaction of piperitol with NaCl and 18-Crown-6

Table S4. Monitoring the reaction of piperitol with NaCl [a]

piperitol	$\begin{array}{c} & Cat.1 (1.0) \\ & O_2 (1 \text{ atm}) \\ & H_2O/CH_3(0) \\ & (balloon) \end{array}$	$ \begin{array}{c} mol\%) \\ \hline mol\%) \\ \hline CN = 1:1 \\ t (h) \end{array} \qquad $	OH perinic acid
Time (h)	piperitol (%) ^b	piperinic aldehyde ^c	piperinic acid ^d
2	92	8	-
4	85	14.5	-
6	78.5	21	-
8	70.5	28.5	-
10	62	33	-
12	58.5	39.5	-
14	50.5	48.5	-
16	46	53	-
18	42.5	57	-
20	37.5	62.5	-
22	33.5	65.5	-
24	31	69	-

^a The reaction was carried out on a 10 mmol scale of piperitol in 40 mL of H_2O/CH_3CN (1:1). Every one hour, the reaction mixtures were calculated by GC and confirmed by GC-MS. ^{b,c,d} Yields were calculated from GC-MS.



Figure S6. Monitoring the reaction of piperitol with NaCl

	n-CoH47OH	+ O ₂ (1 atm) —	Cat.1 additives(10 mol%) ➤ n-C ₇ H₁₅CH	O + <i>n</i> -C7H15CO2H
	Octanal alcol	hol (balloon)	I ₂ O/CH ₃ CN =1/1(2 ml) 60 °C, 12h Octanaldeh	yde Octanal acid
Entry	Additive	Sel. (%) ^b	Yield of aldehyde (%)	Yield of acid (%) ^d
1	Na ₂ CO ₃	72	12	53
2	Na ₂ SO ₄	85	15	68
3	NaSO ₃	99	95	0
4	NaClO ₄	99	4	93
5	NaHSO ₄	92	33	60
6	NaF	76	20	51
7	NaCl	99	27	75
8	NaBr	92	21	73
9	Nal	91	16	69
10	K_2SO_3	95	14	70
11	MgSO₃	15	<5	<5
12	CaSO ₃	18	5	11
13 ^e	NaSO ₃	72	34	13
14 ^f	NaSO ₃	92	85	5
15 ^g	NaSO ₃	96	88	-
16 ^h	NaSO ₃	-	<5	-

Table S5. Optimization of reaction conditions [a]

^[a] Reaction conditions: Cat. **1** (1.0 mol%), alcohols (1.0 mmol), O₂ (1 atm), additive (10 mol%) and CH₃CN / H₂O=1/1 (2 mL) at 60°C. Unless otherwise noted. ^{[b], [c], [d]} Selectivity were determined by GC and confirmed by GC-MS. ^[c] Yields were calculated from ¹H NMR spectra. ^[e] At 25 °C. ^[f] At 70 °C. ^[g] Reactions were carried out under atmospheric air, 24 h. ^[h] Reactions were carried out under nitrogen atmosphere.



Figure S7. Screening of pH on the catalytic oxidation of piperonyl alcohol. Reaction conditions: Cat. 1 (1.0 mol%), Piperonyl alcohol (1.0 mmol), O_2 (1 atm), NaCl (10 mol%) and MeCN/H₂O=1/1 (2 mL) at

60°C. Unless otherwise noted, Selectivity were determined by GC and confirmed by GC-MS. Yields were calculated from ¹H NMR spectra.

A: Control experiments



B: Kinetic Isotopic Effect experiments







Potentiol Vs. Fc/Fc⁺(V)

Figure S8. Experimental studies providing insight into the mechanism of the copper-catalyzed oxidation of alcohols. These include (A) control experiments, (B) kinetic isotope effects, (C) cyclic voltammograms of catalyst **1**/NaCl (1:10).



Schem S1. Gram-scale oxidation of benzyl alcohol to benzaldehyde



Figure S9. Recycling experiments of the Catalyst 1

VII. Recycling experiments of catalyst for oxidative reaction

The copper catalyst was precipitated after the oxidative reaction experiments, and then recovered for reuse. The recovered catalyst was characterized by FT-IR and XRD (Figure. S9 and S10.). The infrared image contains a blue line of fresh catalyst and a red line of the catalyst recycled five times.



Figure S10. FT-IR spectra of the Catalyst 1 before and after reaction



Figure S11. XRD of recycled Catalyst 1

VIII. ESI-Ms spectra of cat. 1 intermediate









IX. References

- [1]. Nomiya, K.; Takahashi, T.; Shirai, T.; Miwa, M. Polyhedron. 1987, 6, 213.
- [2]. Blazevic, A.; Al-Sayed, E.; Roller, A.; Giester, G.; Rompel, A. Chemistry. 2015, 21, 4762.

X. NMR data of products



Benzaldehyde (2): ¹H NMR (500 MHz, CDCl₃) δ 10.20 (s, 1H), 8.06 (dd, *J* = 8.2, 1.3 Hz, 2H), 7.81 (ddd, *J* = 7.4, 4.1, 1.3 Hz, 1H), 7.71 (dd, *J* = 10.7, 4.5 Hz, 2H); ¹³C NMR (125 MHz, CDCl₃) δ 192.4 (s), 136.4 (s), 134.5 (s), 129.8 (s), 129.0 (s).



p-Tolualdehyde (3): ¹H NMR (500 MHz, CDCl₃) δ 9.93 (d, *J* = 2.9 Hz, 1H), 7.79 – 7.71 (m, 2H), 7.30 (d, *J* = 7.5 Hz, 2H), 2.40 (d, *J* = 2.7 Hz, 3H); ¹³C NMR (125 MHz, CDCl₃) δ 191.9 (s), 145.5 (s), 134.2 (s), 129.8 (d, *J* = 15.0 Hz), 21.8 (s).



4-Methoxylbenzaldehyde (4): ¹H NMR (500 MHz, CDCl₃) δ 9.81 (d, *J* = 2.5 Hz, 1H), 7.84 – 7.69 (m, 2H), 7.01 – 6.86 (m, 2H), 3.81 (dd, *J* = 4.3, 3.0 Hz, 3H); ¹³C NMR (125 MHz, CDCl₃) δ 190.9 (s), 164.6 (s), 132.0 (s), 129.9 (s), 114.3 (s), 55.6 (s).



4-Isopropylbenzaldehyde (5): ¹H NMR (500 MHz, CDCl₃) δ 9.91 (s, 1H), 7.75 (d, *J* = 8.2 Hz, 2H), 7.32 (d, *J* = 8.1 Hz, 2H), 2.91 (dt, *J* = 13.8, 6.9 Hz, 1H), 1.21 (d, *J* = 7.1 Hz, 6H); ¹³C NMR (125 MHz, CDCl₃) δ 192.1 (s), 156.3 (s), 134.6 (s), 130.4 (s), 130.0 (s), 127.2 (s), 125.6 (s), 34.5 (s), 23.7 (s).



4-Fluorobenzaldehyde (6): ¹H NMR (500 MHz, CDCl₃) δ 9.97 (s, 1H), 7.91 (dd, J = 8.5, 5.5 Hz, 2H), 7.21 (t, J = 8.5 Hz, 2H); ¹³C NMR (125 MHz, CDCl₃) δ 190.5 (s), 167.5 (s), 133.0 (s), 132.2 (s), 116.4 (s).



4-Chlorobenzaldehyde (7): ¹H NMR (500 MHz, CDCl₃) δ 10.01 (s, 1H), 7.85 (d, *J* = 8.3 Hz, 2H), 7.54 (d, *J* = 8.3 Hz, 2H); ¹³C NMR (125 MHz, CDCl₃) δ 190.9 (s), 141.0 (s), 134.8 (s), 130.9 (s), 129.5 (s).



4-Bromobenzaldehyde (8): ¹H NMR (500 MHz, CDCl₃) δ 10.14 (s, 1H), 7.91 (d, *J* = 8.4 Hz, 2H), 7.85 (d, *J* = 8.3 Hz, 2H); ¹³C NMR (125 MHz, CDCl₃) δ 191.1 (s), 135.1 (s), 132.5 (s), 131.0 (s), 129.8 (s).



4-Nitrobenzaldehyde (9): ¹H NMR (500 MHz, CDCl₃) δ 10.33 (s, 1H), 8.55 (d, *J* = 8.6 Hz, 2H), 8.25 (d, *J* = 8.6 Hz, 2H); ¹³C NMR (125 MHz, CDCl₃) δ 190.4 (s), 151.1 (s), 140.1 (s), 130.5 (s), 124.3 (s).



2-Fluorobenzaldehyde (10): ¹H NMR (500 MHz, CDCl₃) δ 10.55 (s, 1H), 8.05 (dd, *J* = 10.4, 4.2 Hz, 1H), 7.84 – 7.71 (m, 1H), 7.45 (dd, *J* = 9.6, 5.5 Hz, 1H), 7.40 - 7.29 (m, 1H); ¹³C NMR (125 MHz, CDCl₃) δ 187.3 (s), 165.8 (s), 163.7 (s), 136.4 (s), 128.7 (s), 124.7 (s), 116.6 (s).



2-Bromoaldehyde (11): ¹H NMR (500 MHz, CDCl₃) δ 10.39 (s, 1H), 7.94 (dd, *J* = 7.1, 2.1 Hz, 1H), 7.70 – 7.64 (m, 1H), 7.51 – 7.44 (m, 2H); ¹³C NMR (125 MHz, CDCl₃) δ 192.0 (s), 135.4 (s), 133.9 (s), 133.5 (s), 129.9 (s), 128.0 (s), 100.0 (s).



Salicylaldehyde (12): ¹H NMR (500 MHz, CDCl₃) δ 11.04 (s, 1H), 9.90 (s, 1H), 7.66 – 7.47 (m, 2H), 7.13 – 6.95 (m, 2H); ¹³C NMR (125 MHz, CDCl₃) δ 196.6 (s), 161.6 (s), 137.0 (s), 133.7 (s), 120.7 (s), 119.8 (s), 117.6 (s).



5-Methylsalicylaldehyde (13): ¹H NMR (500 MHz, CDCl₃) δ 10.85 (s, 1H), 9.87 (s, 1H), 7.36 (d, *J* = 6.3 Hz, 2H), 6.91 (d, *J* = 9.1 Hz, 1H), 2.35 (s, 3H); ¹³C NMR (125 MHz, CDCl₃) δ 196.6 (s), 159.6 (s), 138.1 (s), 133.4 (s), 129.2 (s), 120.4 (s), 117.4 (s), 20.2 (s).



2,4,6-Trimethylbenzaldehyde (14): ¹H NMR (500 MHz, CDCl₃) δ 10.57 (s, 1H), 6.91 (s, 2H), 2.59 (s, 6H), 2.33 (s, 3H); ¹³C NMR (125 MHz, CDCl₃) δ 193.0 (s), 143.8 (s), 141.5 (s), 130.5 (s), 130.0 (s), 21.5 (s), 20.5 (s).



Terephthalaldehyde (15): ¹H NMR (500 MHz, CDCl₃) δ 10.30 (s, 2H), 8.22 (s, 4H); ¹³C NMR (125 MHz, CDCl₃) δ 191.6 (s), 140.0 (s), 130.2 (s).



4-Methylsulphonylbenzaldehyde (16): ¹H NMR (400 MHz, DMSO) δ 9.94 (d, *J* = 5.3 Hz, 1H), 7.81 – 7.76 (m, 2H), 7.40 – 7.35 (m, 2H), 2.37 (d, *J* = 5.2 Hz, 3H); ¹³C NMR (100 MHz, DMSO) δ 193.2 (s), 145.9 (s), 1 34.7 (s), 130.3 (d, *J* = 11.5 Hz), 22.0 (s).



3,5-Bibromo-2-aminobenzaldehyde (17): ¹H NMR (500 MHz, DMSO) δ 9.81 (s, 1H), 7.90 (d, *J* = 2.3 Hz, 2H), 3.34 (s, 2H); ¹³C NMR (125 MHz, DMSO) δ 193.8 (s), 146.7 (s), 139.9 (s), 137.9 (s), 120.4 (s), 110.3 (s), 105.7 (s).



Piperonal (18): ¹H NMR (500 MHz, DMSO) δ 9.87 – 9.67 (m, 1H), 7.55 (d, *J* = 4.0 Hz, 1H), 7.25 (d, *J* = 83.4 Hz, 1H), 7.15 (d, *J* = 5.6 Hz, 1H), 6.24 – 6.03 (m, 2H); ¹³C NMR (125 MHz, DMSO) δ 191.4 (s), 158.0 (s), 153.2 (s), 129.0 (s), 125.7 (s), 109.1 (s), 106.8 (s), 102.8 (s).



Acetophenone (19): ¹H NMR (400 MHz, DMSO) δ 7.95 (d, *J* = 7.2 Hz, 2H), 7.75 – 7.58 (m, 1H), 7.58 – 7.45 (m, 2H), 2.57 (d, *J* = 0.8 Hz, 3H); ¹³C NMR (100 MHz, DMSO) δ 198.5 (s), 137.5 (s), 133.8 (s), 128.8 (s), 27.3 (s).



4-Methylaldehyde (20): ¹H NMR (500 MHz, CDCl₃) δ 7.88 (d, *J* = 8.0 Hz, 2H), 7.28 (d, *J* = 7.7 Hz, 2H), 2.60 (s, 3H), 2.43 (s, 3H); ¹³C NMR (125 MHz, CDCl₃) δ 198.0 (s), 171.0 (s), 143.9 (s), 134.7 (s), 129.3 (s), 128.5 (s), 26.6 (s), 21.7 (s).



4-Fluoroacetophenone (21): ¹H NMR (500 MHz, CDCl₃) δ 7.97 (ddd, *J* = 6.7, 5.4, 2.1 Hz, 2H), 7.11 (t, *J* = 8.6 Hz, 2H), 2.57 (s, 3H); ¹³C NMR (125 MHz, CDCl₃) δ 196.5 (s), 166.7 (s), 133.6 (s), 130.9 (s), 115.5 (s), 26.5 (s).



4-Chloroacetophenone (22): ¹H NMR (500 MHz, DMSO) δ 7.96 (d, *J* = 8.5 Hz, 2H), 7.57 (d, *J* = 8.5 Hz, 2H), 2.58 (s, 3H); ¹³C NMR (125 MHz, DMSO) δ 197.4 (s), 138.6 (s), 135.9 (s), 130.5 (s), 129.2 (s), 27.15 (s).



4-Nitroacetophenone (23): ¹H NMR (500 MHz, CDCl₃) δ 8.34 (d, *J* = 8.6 Hz, 2H), 8.13 (d, *J* = 8.6 Hz, 2H), 2.70 (s, 3H); ¹³C NMR (126 MHz, CDCl₃) δ 196.4 (s), 150.4 (s), 141.4 (s), 129.3 (s), 123.9 (s), 27.0 (s).



1-Naphthaldehyde (24): ¹H NMR (500 MHz, CDCl₃) δ 10.41 (s, 1H), 9.28 (d, *J* = 8.6 Hz, 1H), 8.10 (d, *J* = 8.2 Hz, 1H), 7.99 (d, *J* = 7.0 Hz, 1H), 7.93 (d, *J* = 8.2 Hz, 1H), 7.71 (t, *J* = 7.6 Hz, 1H), 7.68 -7.54 (m, 2H); ¹³C NMR (125 MHz, CDCl₃) δ 193.6 (s), 139.0 (s), 136.7 (s), 135.3 (s), 133.8 (s), 131.4 (s), 130.6 (s), 129.1 (s), 128.5 (s), 126.0 (s), 124.9 (s).



2-Naphthaldehyde (25): ¹H NMR (500 MHz, CDCl₃) δ 10.20 (s, 1H), 8.38 (s, 1H), 8.04 (d, *J* = 8.1 Hz, 1H), 8.01 – 7.92 (m, 3H), 7.70 – 7.60 (m, 2H); ¹³C NMR (126 MHz, CDCl₃) δ 193.6 (s), 136.7 (s), 135.3 (s), 133.8 (s), 131.4 (s), 130.6(s), 129.1 (s), 128.5 (s), 127.0 (s), 124.9 (s).



Acetonaphthone (26): ¹H NMR (400 MHz, CDCl₃) δ 8.82 (d, *J* = 8.7 Hz, 1H), 7.91 (ddd, *J* = 21.4, 19.0, 8.2 Hz, 3H), 7.55 (dddd, *J* = 28.0, 15.4, 7.4, 4.4 Hz, 3H), 2.73 (s, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 201.8 (s), 135.4 (s), 134.0 (s), 133.1 (s), 130.2 (s), 128.8 (s), 128.5 (s), 128.1 (s), 125.5 (s), 125.1 (s), 124.3 (s), 30.0 (s).



Benzophenone (27): ¹H NMR (500 MHz, CDCl₃) δ 7.83 (d, *J* = 7.3 Hz, 4H), 7.62 (t, *J* = 7.4 Hz, 2H), 7.51 (t, *J* = 7.6 Hz, 4H); ¹³C NMR (126 MHz, CDCl₃) δ 194.6 (s), 134.9 (s), 133.0 (s), 129.9 (s), 129.0 (s).



Benzil (28): ¹H NMR (400 MHz, CDCl₃) δ 7.99 – 7.94 (m, 4H), 7.64 (t, J = 7.4 Hz, 2H), 7.50 (t, J = 7.7 Hz, 4H). ¹³C NMR (101 MHz, CDCl₃) δ 194.69 (s), 135.01 (s), 133.06 (s), 130.0 (s), 129.1 (s).



Cinnamaldehyde (29): ¹H NMR (500 MHz, CDCl₃) δ 9.71 (d, *J* = 7.7 Hz, 1H), 7.63 – 7.51 (m, 2H), 7.51 – 7.33 (m, 4H), 6.72 (dd, *J* = 16.0, 7.7 Hz, 1H); ¹³C NMR (125 MHz, CDCl₃) δ 193.7 (s), 152.8 (s), 134.0 (s), 131.3 (s), 129.1 (s), 128.6 (d, *J* = 7.6 Hz).



4-Methylcinnamaldehyde (30): ¹H NMR (500 MHz, DMSO) δ 9.65 (d, *J* = 7.8 Hz, 1H), 7.68 (dd, *J* = 27.5, 11.9 Hz, 3H), 7.29 (d, *J* = 7.8 Hz, 2H), 6.82 (dd, *J* = 15.9, 7.8 Hz, 1H), 2.35 (s, 3H); ¹³C NMR (125 MHz, DMSO) δ 194.9 (s), 153.8 (s), 141.9 (s), 131.9 (s), 130.2 (s), 129.3 (s), 128.1 (s), 21.6 (s).

4-Methoxylcinnamaldehyde (31): ¹H NMR (500 MHz, DMSO) δ 9.61 (d, *J* = 7.8 Hz, 1H), 7.72 (d, *J* = 8.6 Hz, 2H), 7.67 (d, *J* = 15.8 Hz, 1H), 7.03 (d, *J* = 8.6 Hz, 2H), 6.75 (dd, *J* = 15.8, 7.8 Hz, 1H), 3.82 (s, 3H); ¹³C NMR (125 MHz, DMSO) δ 194.6 (s), 162.2 (s), 153.7 (s), 131.2 (s), 127.2 (s), 125.8 (s), 115.0 (s), 55.9 (s).



4-Chlorocinnamaldehyde (32): ¹H NMR (500 MHz, DMSO) δ 9.68 (d, *J* = 7.7 Hz, 1H), 7.77 (dd, *J* = 23.9, 12.2 Hz, 3H), 7.55 (d, *J* = 8.4 Hz, 2H), 6.90 (dd, *J* = 15.9, 7.7 Hz, 1H); ¹³C NMR (125 MHz, DMSO) δ 194.9 (s), 152.1 (s), 136.2 (s), 133.5 (s), 130.9 (s), 129.6 (d, *J* = 5.4 Hz).



2-Pyridinecarboxaldehyde (33): ¹H NMR (500 MHz, CDCl₃) δ 9.54 (dd, *J* = 3.9, 0.7 Hz, 1H), 7.20 (d, *J* = 7.9 Hz, 1H), 7.03 (td, *J* = 7.8, 1.6 Hz, 1H), 6.38 (ddd, *J* = 7.4, 4.8, 1.1 Hz, 1H); ¹³C NMR (125 MHz, DMSO) δ 194.1 (s), 152.8 (s), 150.6 (s), 138.0 (s), 128.8 (s), 122.0 (s).



2-Thenaldehyde (34): ¹H NMR (500 MHz, CDCl₃) δ 9.89 (s, 1H), 7.75 (d, *J* = 3.3 Hz, 1H), 7.72 (d, *J* = 4.8 Hz, 1H), 7.22 - 7.12 (m, 1H); ¹³C NMR (125 MHz, CDCl₃) δ 183.1 (s), 144.0 (s), 136.6 (s), 135.2 (s), 128.4 (s).



Furfural (35): ¹H NMR (500 MHz, CDCl₃) δ 9.67 (s, 1H), 7.71 (s, 1H), 7.27 (d, *J* = 3.5 Hz, 1H), 6.62 (d, *J* = 1.9 Hz, 1H); ¹³C NMR (125 MHz, CDCl₃) δ 177.9 (s), 153.0 (s), 148.1 (s), 121.1 (s), 112.6 (s).



2-Acetylthiophene (36): ¹H NMR (500 MHz, CDCl₃) δ 7.55 (s, 1H), 7.49 (s, 1H), 6.97 (s, 1H), 2.40 (s, 3H); ¹³C NMR (125 MHz, CDCl₃) δ 190.6 (s), 144.4 (s), 133.8 (s), 132.6 (s), 128.2 (s), 26.7 (s).



2-Acetylfuran (37): ¹H NMR (500 MHz, CDCl₃) δ 7.60 (s, 1H), 7.20 (d, *J* = 3.5 Hz, 1H), 6.59 – 6.50 (m, 1H), 2.49 (s, 3H); ¹³C NMR (125 MHz, CDCl₃) δ 152.9 (s), 146.5 (s), 117.3 (s), 112.3 (s), 26.0 (s).



2-acetylpyridine (38): ¹H NMR (500 MHz, CDCl₃) δ 8.54 (dd, *J* = 3.9, 0.7 Hz, 1H), 7.89 (d, *J* = 7.9 Hz, 1H), 7.70 (td, *J* = 7.8, 1.6 Hz, 1H), 7.34 (ddd, *J* = 7.4, 4.8, 1.1 Hz, 1H), 2.58 (s, 3H); ¹³C NMR (125 MHz, CDCl₃) δ 199.8 (s), 153.4 (s), 148.9 (s), 136.7 (s), 126.0 (s), 121.4 (s), 25.6 (s).

XI. NMR spectra





 ^{13}C NMR spectra of $\boldsymbol{3}$ (125 MHz, CDCl_3)









 ^{13}C NMR spectra of **6** (125 MHz, CDCl_3)





 ^{13}C NMR spectra of $\boldsymbol{8}$ (125 MHz, CDCl_3)





 ^{13}C NMR spectra of 10 (125 MHz, CDCl_3)



¹³C NMR spectra of **11** (125 MHz, CDCl₃)







 ^{13}C NMR spectra of 14 (125 MHz, CDCl_3)



 ^{13}C NMR spectra of 15 (125 MHz, CDCl_3)



¹³C NMR spectra of **16** (125 MHz, DMSO)



¹³C NMR spectra of **17** (125 MHz, DMSO)



¹³C NMR spectra of **18** (125 MHz, DMSO)



¹³C NMR spectra of **19** (125 MHz, DMSO)



 ^{13}C NMR spectra of 20 (125 MHz, CDCl_3)





¹³C NMR spectra of **22** (125 MHz, DMSO)



 ^{13}C NMR spectra of 23 (125 MHz, CDCl_3)



 ^{13}C NMR spectra of 24 (125 MHz, CDCl_3)



 ^{13}C NMR spectra of 25 (125 MHz, CDCl_3)



 $^{\rm 13}{\rm C}$ NMR spectra of ${\bf 26}$ (125 MHz, CDCl_3)



¹³C NMR spectra of **27** (125 MHz, CDCl₃)



¹³C NMR spectra of **28** (125 MHz, CDCl₃)



 ^{13}C NMR spectra of 29 (125 MHz, CDCl_3)



¹³C NMR spectra of **30** (125 MHz, DMSO)



¹³C NMR spectra of **31** (125 MHz, DMSO)



¹³C NMR spectra of **32** (125 MHz, DMSO)





 ^{13}C NMR spectra of 34 (125 MHz, $\text{CDCl}_3)$



¹³C NMR spectra of **35** (125 MHz, CDCl₃)



 ^{13}C NMR spectra of 36 (125 MHz, CDCl_3)





 $^{\rm 13}{\rm C}$ NMR spectra of ${\bf 38}$ (125 MHz, CDCl_3)