

## Supplementary Materials for

# Efficient Microbial Synthesis of Key Steroidal Intermediates from Bio-Renewable Phytosterols by Genetically Modified *Mycobacterium fortuitum* Strains

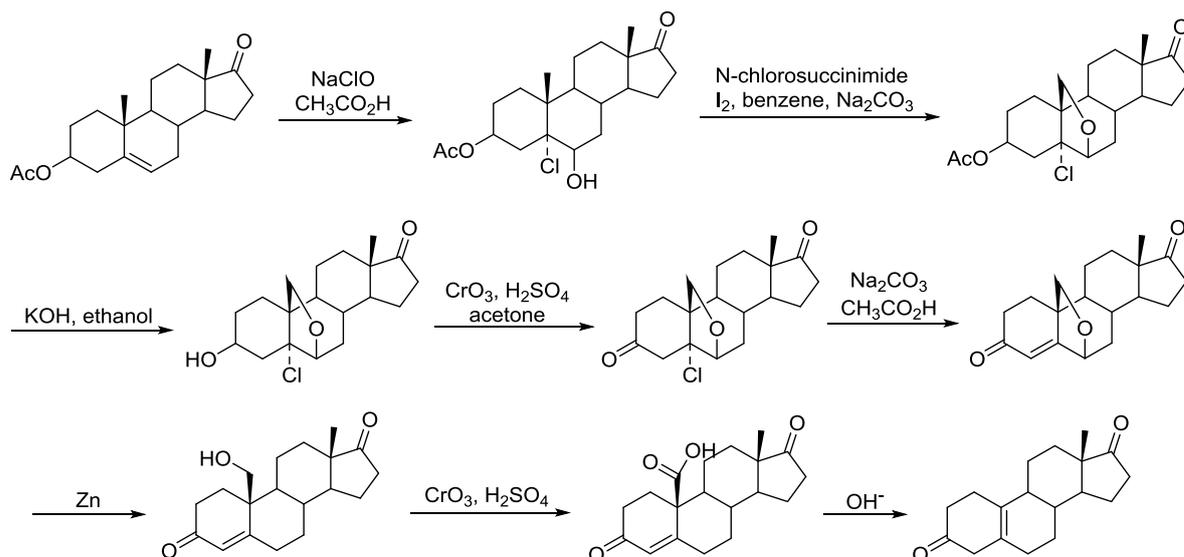
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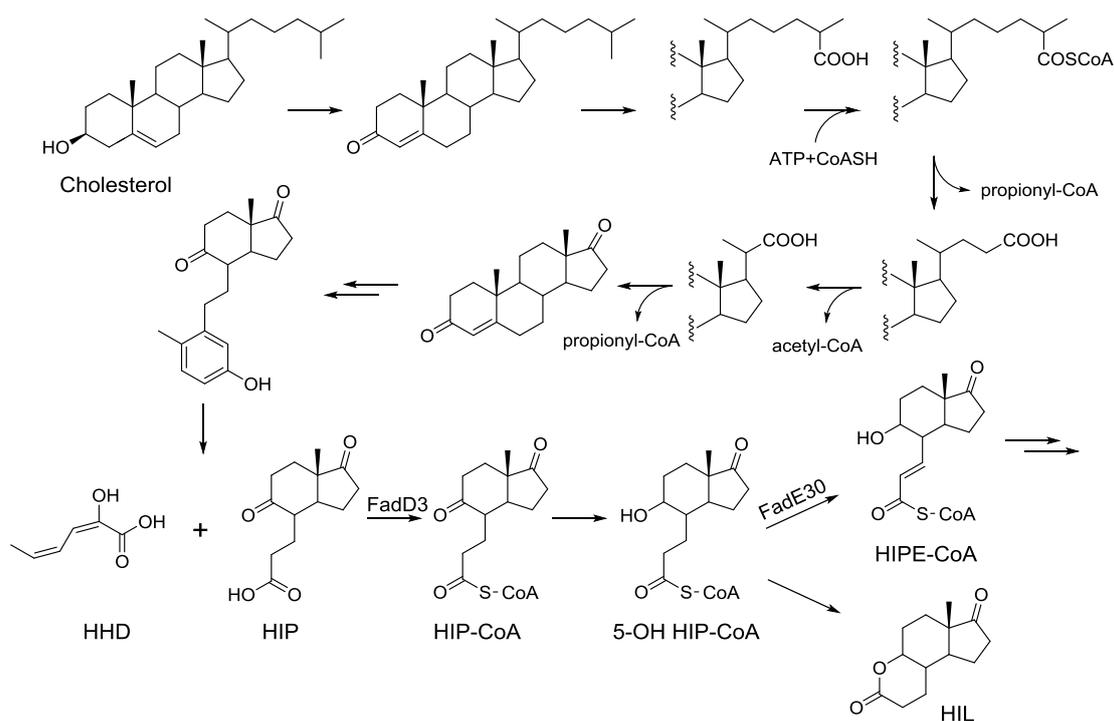
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**Scheme S1** The elimination of the C-19 methyl group by chemical synthesis.<sup>1-3</sup>



**Scheme S2** Overview of the cholesterol degradation pathway.<sup>4-7</sup> The side chain and AB-ring of cholesterol are degraded to produce HIP and 2-hydroxyhexa-2,4-dienoic acid (HHD). The thioesterification of HIP, which is catalyzed by an acyl-CoA synthase FadD3, initiates the catabolism of CD-ring. FadE30, an acyl-CoA dehydrogenase, catalyzes the further degradation of metabolite 5-OH HIP-CoA.

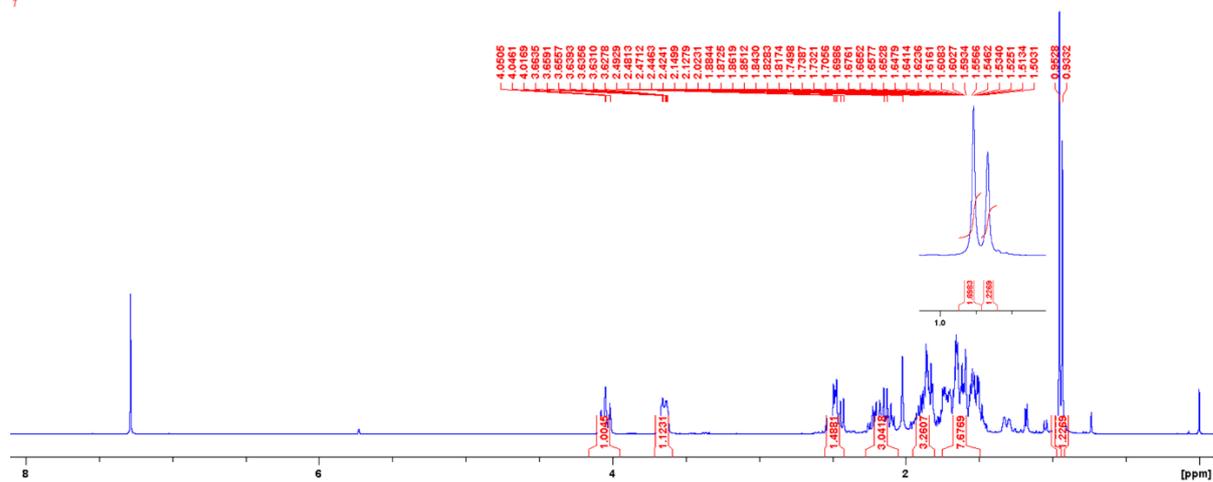


Fig. S1 <sup>1</sup>H NMR spectrum of HIK.

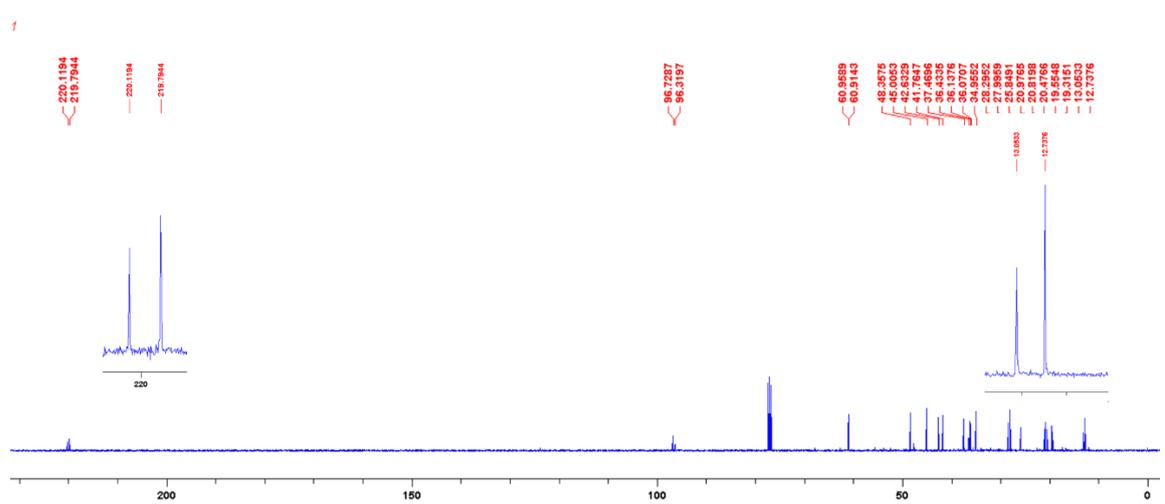
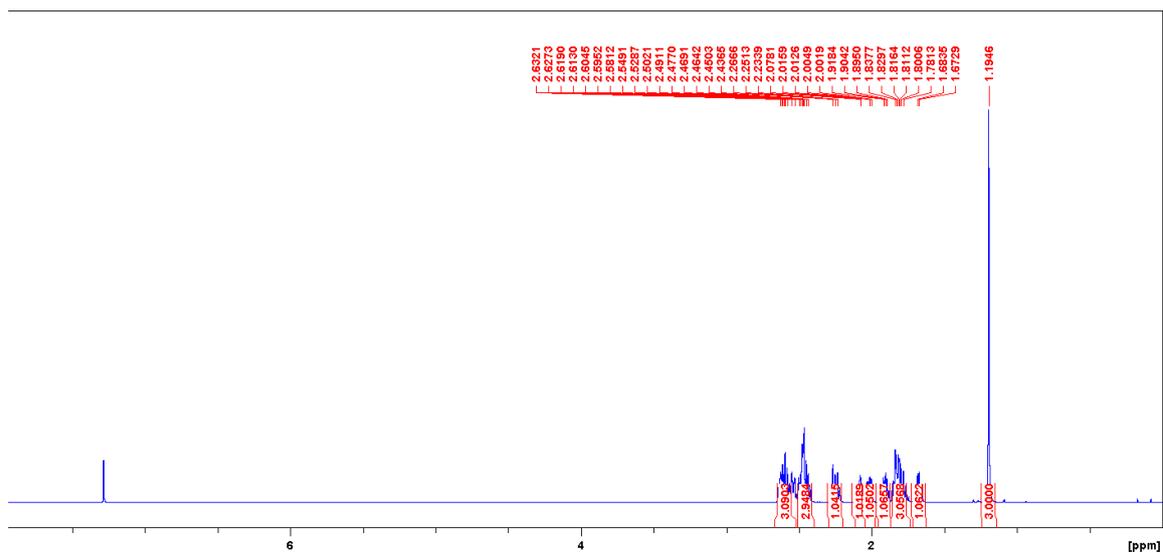
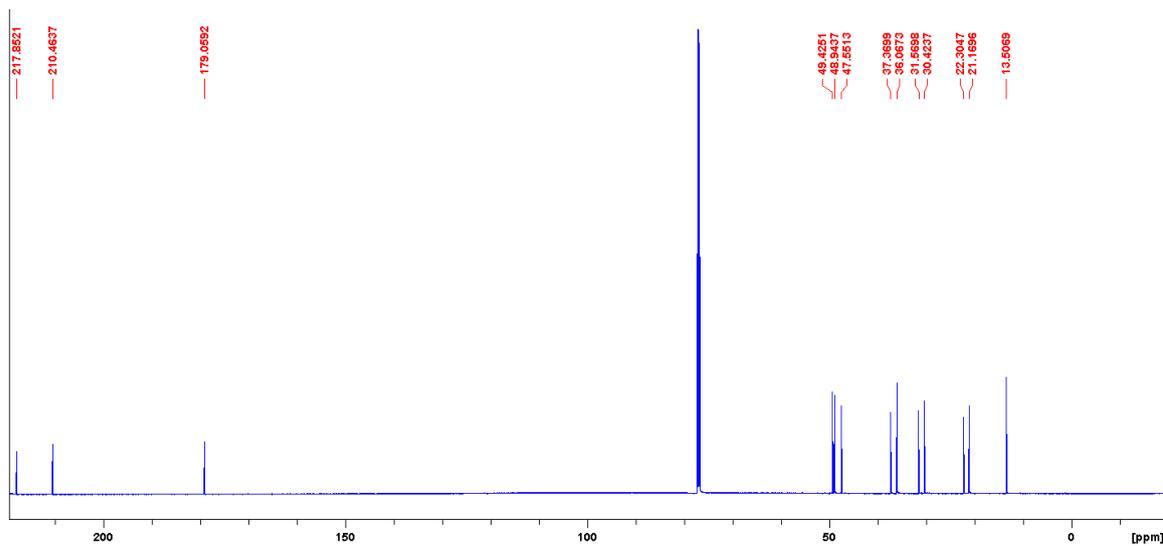


Fig. S2 <sup>13</sup>C NMR spectrum of HIK.



**Fig. S3** <sup>1</sup>H NMR spectrum of HIP.



**Fig. S4** <sup>13</sup>C NMR spectrum of HIP.

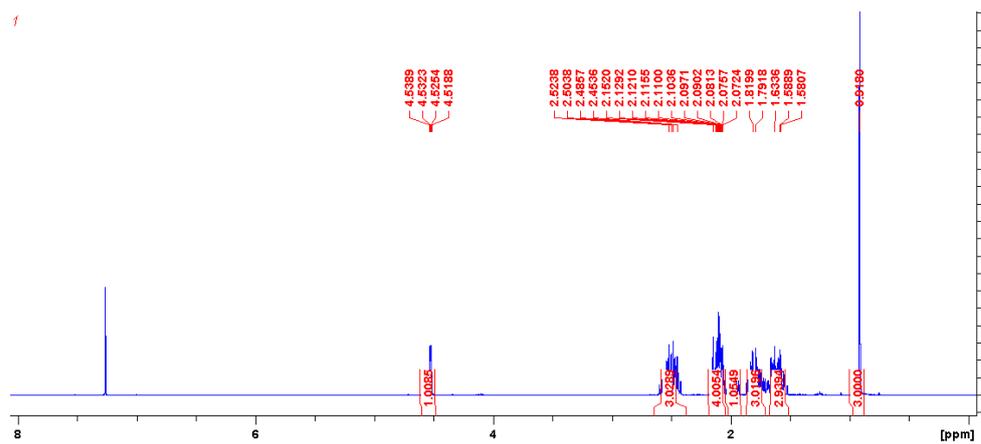


Fig. S5  $^1\text{H}$  NMR spectrum of HIL.

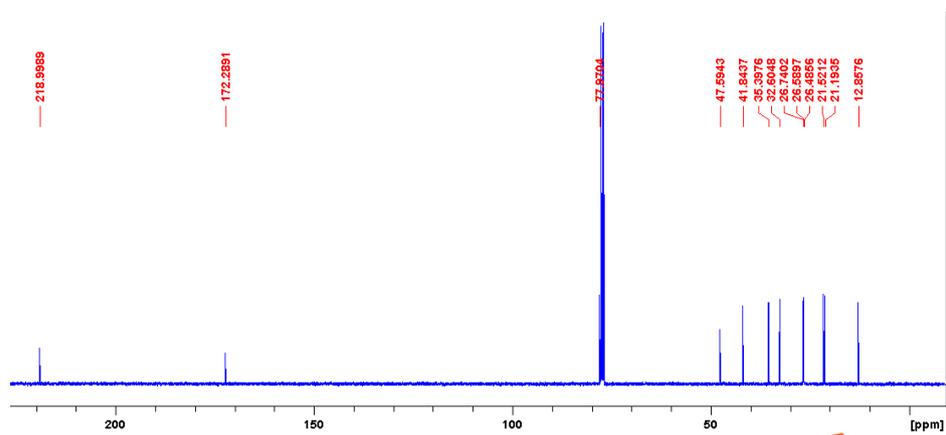
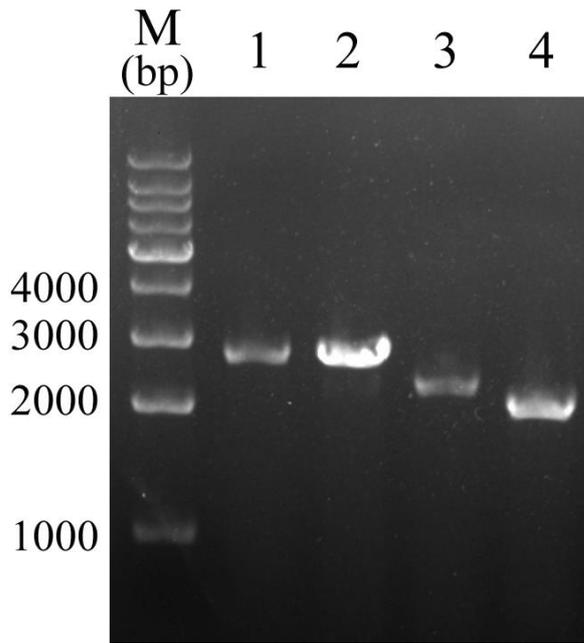
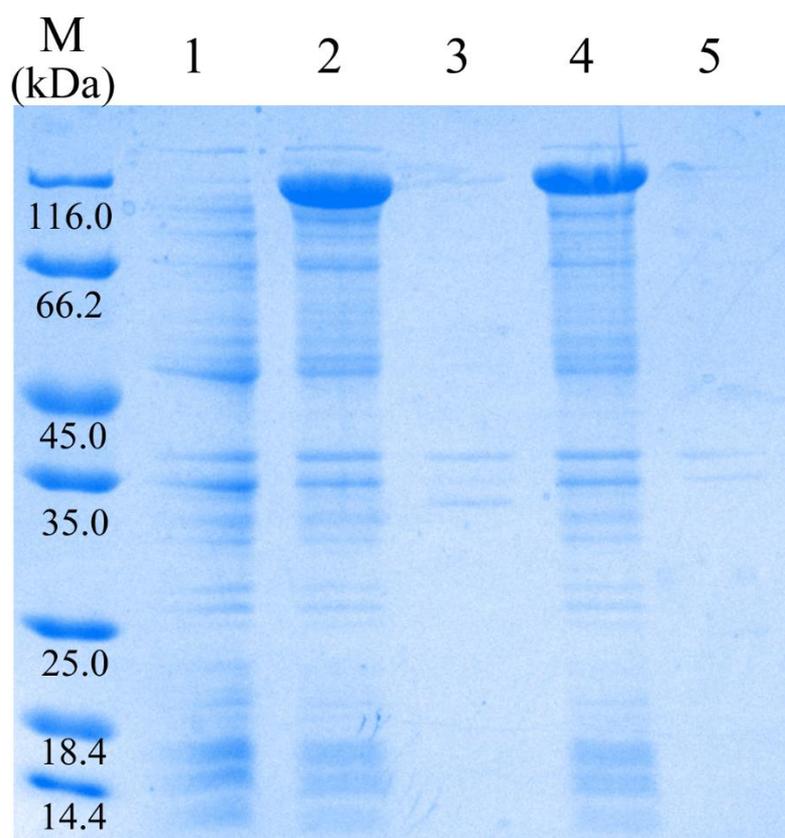


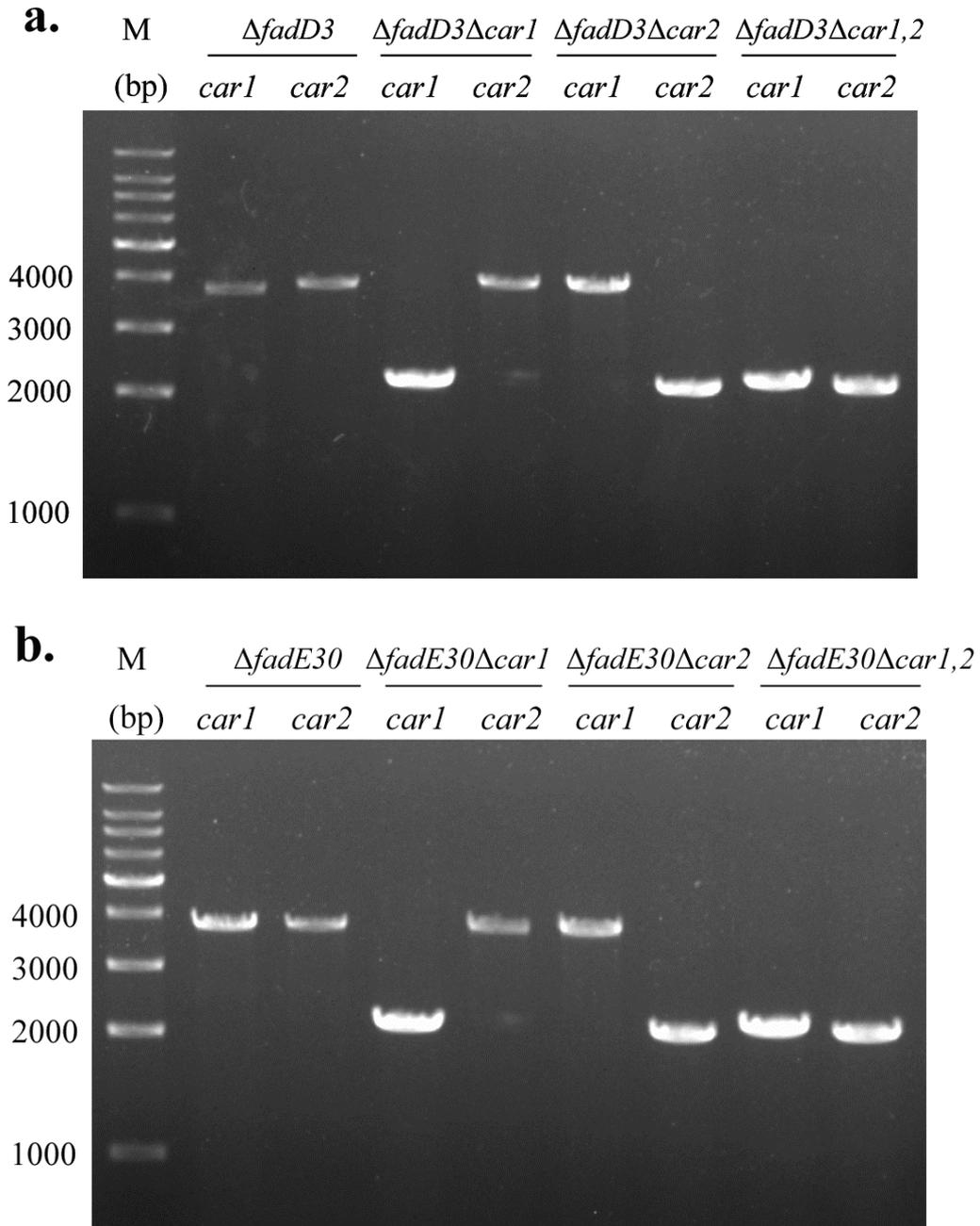
Fig. S6  $^{13}\text{C}$  NMR spectrum of HIL.



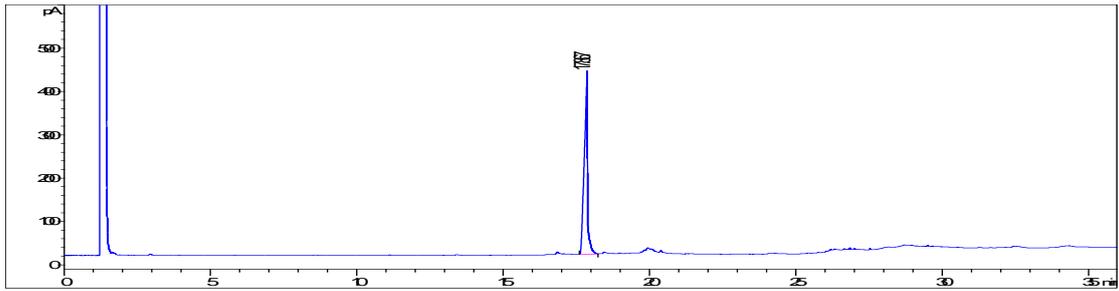
**Fig. S7** Identification of *fadD3* and *fadE30* gene deleted mutants of ATCC 6841 by PCR. Lanes: (M) DNA markers; (1) The PCR products of *fadD3* using ATCC 6841 as the control; (2) The PCR products of *fadE30* using ATCC 6841 as the control; (3) The PCR products of *fadD3* with a shortened size (~ 2300 bp) represented a successful deletion; (4) The PCR products of *fadE30* with a shortened size (~ 1900 bp) represented a successful deletion.



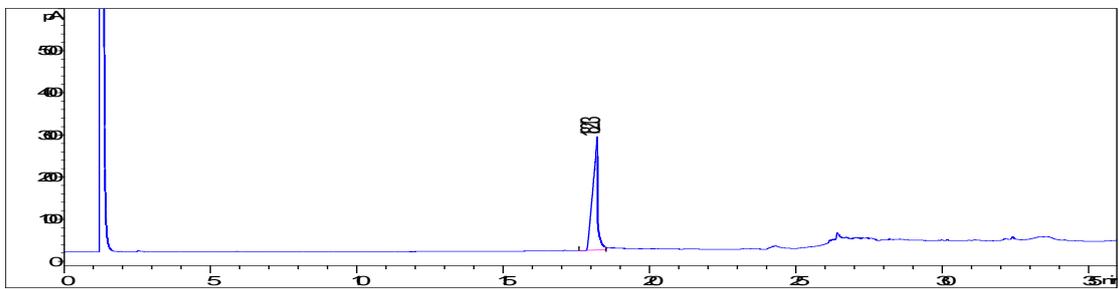
**Fig. S8** SDS-PAGE analysis of heterologous expression of CAR1 and CAR2 from ATCC 6841 in *E.coli* BL21(DE3) host cells. Lanes: (1) total cell extracts of *E.coli* BL21(DE3) harboring blank pET28a(+) as the control; (2) supernatant fraction of CAR1<sub>*E.coli*</sub>; (3) precipitant fraction of CAR1<sub>*E.coli*</sub>; (4) supernatant fraction of CAR2<sub>*E.coli*</sub>; (5) precipitant fraction of CAR2<sub>*E.coli*</sub>; (M) protein markers.



**Fig. S9** Identification of *car1* and *car2* gene deleted strains by PCR. The PCR products of *car1* and *car2* with a shortened size (2100bp and 2000 bp) represented a successful deletion. (a) Identification of *car1* and *car2* deleted strains with  $\Delta$ fadD3 background. (b) Identification of *car1* and *car2* deleted strains with  $\Delta$ fadE30 background.



**Fig. S10** The biotransformation of phytosterols by strain  $\Delta fadD3\Delta car1,2$ . The product HIP (17.9 min) was detected from the extract of fermentation supernatant by GC.



**Fig. S11** The biotransformation of phytosterols by strain  $\Delta fadE30\Delta car1,2$ . The product HIL (18.2 min) was detected from the extract of fermentation supernatant by GC.

**Table S1** Strains used in this study.

Name	Description	Source
ATCC 6841	Wild type strain	ATCC
$\Delta fadD3$	<i>fadD3</i> -deleted strain of 6841	This study
$\Delta fadD3\Delta car1$	<i>car1</i> -deleted strain with $\Delta fadD3$ background	This study
$\Delta fadD3\Delta car2$	<i>car2</i> -deleted strain with $\Delta fadD3$ background	This study
$\Delta fadD3\Delta car1,2$	<i>car1</i> and <i>car2</i> -deleted strain with $\Delta fadD3$ background	This study
$\Delta fadE30$	<i>fadE30</i> -deleted strain of 6841	This study
$\Delta fadE30\Delta car1$	<i>car1</i> -deleted strain with $\Delta fadE30$ background	This study
$\Delta fadE30\Delta car2$	<i>car2</i> -deleted strain with $\Delta fadE30$ background	This study
$\Delta fadE30\Delta car1,2$	<i>car1</i> and <i>car2</i> -deleted strain with $\Delta fadE30$ background	This study
pET28a <sub><i>E.coli</i></sub>	<i>E.coli</i> BL21(DE3) harboring pET28a(+)	Novagen
CAR1 <sub><i>E.coli</i></sub>	<i>E.coli</i> BL21(DE3) harboring pET28a(+)- <i>car1</i>	This study
CAR2 <sub><i>E.coli</i></sub>	<i>E.coli</i> BL21(DE3) harboring pET28a(+)- <i>car2</i>	This study

**Table S2** Primers and plasmids used in this study.

Name	Description	Source/reference
<b>Primers</b>		
<i>fadE30</i> <sub>del</sub> -U-F	<u>TGTTGCCATTGCTGCAGGCACCGGGTACAGAGTCG</u>	This study
<i>fadE30</i> <sub>del</sub> -U-R	TGATTCCACGCCCGACGCTCTTC	This study
<i>fadE30</i> <sub>del</sub> -D-F	<u>CGTCGGGCGTGGAATCACCGCTGATCCGCCAAAAG</u>	This study
<i>fadE30</i> <sub>del</sub> -D-R	<u>GTACCGCGGCCGCTTAATTAAGCCGAACGCCTCGTCTGA</u>	This study
<i>fadD3</i> <sub>del</sub> -U-F	<u>ACGTTGTTGCCATTGCTGCAGCGGTGACGTCGAGGATCTTG</u>	This study
<i>fadD3</i> <sub>del</sub> -U-R	<u>GGTAGCCGAGCATGACATTGGGTTGATGCACAGATAGCGGTCG</u>	This study
<i>fadD3</i> <sub>del</sub> -D-F	<u>CGACCGCTATCTGTGCATCAACCCAATGTCATGCTCGGCTACC</u>	This study
<i>fadD3</i> <sub>del</sub> -D-R	<u>GTACCGCGGCCGCTTAATTAACTCGTTGCTCGACGTGCTCATG</u>	This study
<i>car1</i> <sub>del</sub> -U-F	<u>TGTTGCCATTGCTGCAGGCATCTCGCACCATCAGC</u>	This study

Name	Description	Source/reference
<i>car1</i> <sub>del</sub> -U-R	<u>AGGTC</u> ACTTGGTCGAGCCAGCGCCGCCTGATTCTC	This study
<i>car1</i> <sub>del</sub> -D-F	CAGCGCCGCCTGATTCTCGCTCGACCAAGTGACCTG	This study
<i>car1</i> <sub>del</sub> -D-R	<u>CGCGGCCGCTTAATTA</u> ACGATCGGCTTGCTCTAGG	This study
<i>car2</i> <sub>del</sub> -U-F	<u>TGTTGCCATTGCTGC</u> AGGATCAGACTCACAGCACATTG	This study
<i>car2</i> <sub>del</sub> -U-R	GATCTTCGTGGTGAGCGCGGCGAGCGAGGCATACAG	This study
<i>car2</i> <sub>del</sub> -D-F	<u>CTGTATGCCTCGCTCGCC</u> GCGCTCACCACGAAGATC	This study
<i>car2</i> <sub>del</sub> -D-R	<u>CGCGGCCGCTTAATTA</u> AAGGTTCCCCTGAGCAAATC	This study
<i>car1</i> -F	<u>CGCGGCAGCCATATG</u> ACCACCGAAACGC	This study
<i>car1</i> -R	GAGCTCGAATTCGGATCCTTACAGCAATCCGAGCAG	This study
<i>car2</i> -F	<u>CGCGGCAGCCATATG</u> TCGTTTGATACTCGC	This study
<i>car2</i> -R	GAGCTCGAATTCGGATCCTAGAGCAGGCCGAGCTG	This study

Name	Description	Source/reference
<b>Plasmids</b>		
p2NIL	Gene manipulation vector, Kan <sup>R</sup>	Parish and Stoker <sup>8</sup>
pGOAL19	<i>Hyg</i> <i>Pag85-lacZ</i> <i>P<sub>hsp60</sub>-sacB</i> , <i>PacI</i> cassette vector, Amp <sup>R</sup>	Parish and Stoker <sup>8</sup>
pET28a(+)	<i>E. coli</i> expression vector, Kan <sup>R</sup>	Novagen
pKH <sub>del</sub> - <i>fadE30</i>	p2NIL carrying the alleles of <i>fadE30</i> and the selection cassette from pGOAL19, Kan <sup>R</sup> , Hyg <sup>R</sup>	This study
pKH <sub>del</sub> - <i>fadD3</i>	p2NIL carrying the alleles of <i>fadD3</i> and the selection cassette from pGOAL19, Kan <sup>R</sup> , Hyg <sup>R</sup>	This study
pKH <sub>del</sub> - <i>car1</i>	p2NIL carrying the alleles of <i>car1</i> and the selection cassette from pGOAL19, Kan <sup>R</sup> , Hyg <sup>R</sup>	This study
pKH <sub>del</sub> - <i>car2</i>	p2NIL carrying the alleles of <i>car2</i> and the selection cassette from pGOAL19, Kan <sup>R</sup> , Hyg <sup>R</sup>	This study
pET28a(+)- <i>car1</i>	pET-28a(+) harboring the <i>car1</i> gene, Kan <sup>R</sup>	This study
pET28a(+)- <i>car2</i>	pET-28a(+) harboring the <i>car2</i> gene, Kan <sup>R</sup>	This study

Notes: Kan<sup>R</sup> kanamycin-resistant, Amp<sup>R</sup> ampicillin-resistant, Hyg<sup>R</sup> hygromycin-resistant, the restriction enzyme cutting sites were in bold, and the homologous sequence were underlined.

## References

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