# **Supplementary Information**

# Bifunctional phase-transfer catalysts for fixation of CO<sub>2</sub> with epoxides under

## ambient pressure

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#### General

<sup>1</sup>H NMR, <sup>13</sup>C NMR, <sup>19</sup>F and <sup>31</sup>P spectra were recorded at room temperature using 400 MHz *Bruker spectrometer*. The data are reported as follows: chemical shift  $\delta$  in ppm (from internal tetramethylsilane on the  $\delta$  scale in case of <sup>1</sup>H and CDCl<sub>3</sub> triplet in case of <sup>13</sup>C), multiplicity (br = broad, s = singlet, d = doublet, t = triplet, q = quartet, m = multiplet), coupling constants (Hz) and integration. High resolution mass spectra were obtained by peak matching on BrukermaXis Spectrometer. Melting points are reported uncorrected and measured on Fukai-X-6 melting point apparatus. HPLC data were recorded on Agilent 1260 with UV detector. Analytical thin layer chromatography was performed on 0.25 mm silica gel plates with UV-254 fluorescent indicator. Flash column chromatography was performed using indicated solvent system on 200~300 mesh silica gel (SiO<sub>2</sub>). All air- and water-sensitive reactions were carried out under an inert atmosphere in glassware, which had been oven-dried as per standard procedure. Unless otherwise noted, all reagents were commercially obtained and used without further purification. Benzyl chloride and benzyl bromide were freshly distilled and used in following steps.

## 1. The synthesis of achiral phase-transfer catalysts (APTCs)

A. Synthesis of **APTCs** with a urea group



**APTC-10**:  $X = CH_2$ ,  $R = 3,5-(CH_3)_2$ ,  $Ar = 4-CF_3C_6H_4$ , 83% yield.

#### Typical procedure:

To 1.0 mmol of 2-(pyrrolidin-1-yl)ethan-1-amine (115 mg) in dry CH<sub>2</sub>Cl<sub>2</sub> (4.0 mL) was added by 1.05 mmol of 3,5-bistrifluorometylphenylisocyanate (270 mg) in dry  $CH_2Cl_2$  (1.0 mL) at room temperature (r.t.). The mixture was stirred for 8 h under inert atmosphere. When TLC indicates the complete consumption of starting materials, the reaction mixture was evaporated under reduced pressure to yield a light yellow glue which was purified by a chromatography (petroether/ethyl acetate = 2/1, v/v ) to give pure urea product as off-white flash column powder (351 mg, 95% yield). To the above-prepared urea in dry toluene (4.0 mL) was added by 1.05 mmol of BnBr (180 mg) in dry toluene (1.0 mL) at room temperature (r.t.). The mixture was heated to 80 °C by an oil-bath and stirred for 12 h under inert atmosphere. The reaction mixture was cooled to r.t. and the precipitate was collected and washed by  $dry Et_2O$  for three to five times. This precipitate was dried under a reduced pressure to give APTC-1 as a pale powder (490 mg, 91% yield in two steps) which was used directly without further purification.

APTC-4 and APTC-6~10 was obtained by using the same procedure as APTC-1, respectively.

**APTC-1**: pale powder, 91% yield; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$ : 9.45 (s, 1H), 8.06 (s, 2H), 7.90 (t, J = 5.76 Hz, 1H), 7.57-7.48 (m, 4H), 7.45 (s, 1H), 4.75 (s, 2H), 4.02-3.98 (m, 2H), 3.86-3.73 (m, 4H), 3.62 (t, *J* = 5.72 Hz, 2H), 2.33 (m, 2H), 2.19 (m, 2H), 1.80 (s, 2H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>) δ: 155.89, 141.16, 132.55, 131.98, 131.65, 131.19, 129.74, 126.90, 124.69, 121.98, 117.97, 115.13, 62.61, 61.43, 59.59, 58.41, 34.97, 21.30, 18.44; <sup>19</sup>F NMR (376 MHz, CDCl<sub>3</sub>) δ: -62.90; HRMS (ESI) *m/z* calcd. for C<sub>22</sub>H<sub>24</sub>F<sub>6</sub>N<sub>3</sub>O<sup>+</sup> [M-Br]<sup>+</sup>: 460.1818, found 460.1829.

**APTC-4**: white powder, 89% yield; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ: 9.52 (s, 1H), 8.03 (s, 2H), 7.61-7.44 (m, 6H), 7.28 (s, 1H), 5.08 (s, 2H), 4.13-4.00 (m, 7H), 3.88-3.85 (m, 2H), 3.54 (bs, 2H), 1.95 (bs, 1H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>) δ: 155.75, 140.92, 133.31, 132.10, 131.76, 131.41, 129.70, 125.37, 124.64, 121.93, 117.99, 115.38, 66.34, 60.38, 56.81, 56.32, 33.97; <sup>19</sup>F NMR (376 MHz, CDCl<sub>3</sub>) δ: -62.91; HRMS (ESI) *m/z* calcd. for C<sub>22</sub>H<sub>24</sub>F<sub>6</sub>N<sub>3</sub>O<sub>2</sub><sup>+</sup> [M-Br]+: 476.1767, found 476.1779.

**APTC-6**: white powder, 79% yield; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$ : 8.66 (bs, 1H), 7.53-7.39 (m, 8H), 7.00 (s, 2H), 4.75 (s, 2H), 3.87-3.46 (m, 8H), 2.24 (s, 3H), 2.11-2.06 (m, 4H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$ : 156.37, 136.79, 132.67, 131.76, 130.85, 129.55, 129.30, 127.36, 118.98, 62.49, 61.30, 59.44, 34.99, 21.27, 20.73; HRMS (ESI) *m/z* calcd. for C<sub>21</sub>H<sub>28</sub>N<sub>3</sub>O<sup>+</sup> [M-Br]<sup>+</sup>: 338.2227, found 338.2234.

**APTC-7**: white powder, 74% yield; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$ : 8.76 (s, 1H), 8.37 (s, 1H), 7.57-7.49 (m, 3H), 7.47-7.43 (m, 2H), 7.15 (s, 2H), 6.64 (s, 1H), 4.54 (s, 2H), 3.95-3.94 (m, 2H), 3.61 (m, 4H), 3.47 (m, 2H), 2.27 (s, 6H), 2.25 (s, 2H), 2.11 (m, 2 H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$ : 156.73, 138.43, 132.36, 131.14, 129.75, 126.92, 116.95, 62.00, 61.31, 60.01, 34.92, 21.36, 21.10; HRMS (ESI) *m/z* calcd. for C<sub>22</sub>H<sub>30</sub>N<sub>3</sub>O<sup>+</sup> [M-Br]<sup>+</sup>: 352.2383, found 352.2392.

**APTC-8**: white powder, 83% yield; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$ : 9.58 (s, 1H), 8.03 (s, 2H), 7.81 (d, J = 7.76 Hz, 2H), 7.67 (d, J = 7.84 Hz, 2H), 7.53 (bs, 1H), 7.42 (s, 1H), 5.01 (s, 2H), 4.05 (d, J = 5.12 Hz, 2H), 3.87 (br, 2H), 3.74-3.71 (m, 2H), 3.66 (br, 2H), 2.27 (s, 2H), 2.15 (s, 2H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$ : 155.83, 141.05, 133.29, 133.01, 131.86, 130.88, 129.35, 128.23, 126.49, 124.62, 121.91, 117.85, 115.25, 61.73, 61.45, 59.61, 35.11, 21.22; <sup>19</sup>F NMR (376 MHz, CDCl<sub>3</sub>)  $\delta$ : -62.99, -63.26; HRMS (ESI) *m/z* calcd. for C<sub>23</sub>H<sub>23</sub>F<sub>9</sub>N<sub>3</sub>O<sup>+</sup> [M-Br]<sup>+</sup>: 528.1692, found 528.1698.

**APTC-9**: white powder, 85% yield; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$ : 8.69 (s, 1H), 7.76 (d, J = 7.0 Hz, 2H), 7.58 (d, J = 7.04 Hz, 2H), 7.36 (d, J = 6.72 Hz, 3H), 7.01 (d, J = 7.36 Hz, 2H), 4.99 (s, 1H), 3.95 (s, 2H), 3.76 (s, 2H), 3.71 (s, 2H), 3.53 (s, 1H), 2.25 (s, 3H), 2.12 (br, 2H), 2.07 (br, 1H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$ : 156.34, 136.62, 133.39, 132.91, 132.58, 132.05, 131.29, 129.38, 126.30, 124.72, 122.01, 118.95, 61.59, 61.34, 59.51, 35.15, 21.18, 20.67; <sup>19</sup>F NMR (376 MHz, CDCl<sub>3</sub>)  $\delta$ : -63.08; HRMS (ESI) *m/z* calcd. for C<sub>22</sub>H<sub>27</sub>F<sub>3</sub>N<sub>3</sub>O<sup>+</sup> [M-Br]<sup>+</sup>: 406.2101, found 406.2107.

**APTC-10**: white powder, 83% yield; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$ : 8.47 (s, 1H), 7.74 (s, 2H), 7.67 (s, 2H), 7.05 (m, 2H), 6.63 (s, 1H), 4.87 (s, 2H), 3.90~3.52 (dd, J = 1.96, 6.56 Hz, 8H), 2.23 (s, 6H), 2.11 (br, 4H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$ : 156.21, 138.87, 138.52, 133.26, 131.22, 126.44, 124.43, 116.57, 61.53, 59.42, 49.51, 49.30, 34.89, 21.34, 21.37; <sup>19</sup>F NMR (376 MHz, CDCl<sub>3</sub>)  $\delta$ : -63.15; HRMS (ESI) *m/z* calcd. for C<sub>23</sub>H<sub>29</sub>F<sub>3</sub>N<sub>3</sub>O<sup>+</sup> [M-Br]<sup>+</sup>: 420.2257, found 420.2264.

B. Synthesis of APTCs with a squaramide group (APTC-3)

Step 1: To a solution of 3-((3,5-bis(trifluoromethyl)phenyl)amino)-4-methoxycyclobut-3-ene-1,2-dione (1.0 mmol) in MeOH (3 mL) was added a solution of 2-morpholinoethan-1-amine (1.0 mmol) in MeOH (2 mL) at r.t.. The mixture was stirred for 24 h. The reaction mixture was filtered, and the precipitate was washed with cold MeOH (2×1.0 mL) to afford pure squaramide.

Step 2: To 0.5 mmol of squaramide in anhydrous toluene (2.0 mL) was added by 0.6 mmol of BnBr in anhydrous toluene (1.0 mL) at room temperature (r.t.). The mixture was stirred for 12 h under inert atmosphere (checked by TLC) at 80 °C. The precipitate was collected and washed by anhydrous  $Et_2O$  (3×1.0 mL) to afford pure phase-transfer catalysts.



**APTC-3**: white powder, 76% yield; <sup>1</sup>H NMR (400 MHz, DMSO- $d_6$ )  $\delta$ : 10.84 (s, 1H), 8.27 (br, 1H), 8.12 (s, 2H), 7.69 (br, 1H), 7.64 (d, J = 5.08 Hz, 2H), 7.55 (m, 3H), 4.85, (s, 2H), 4.24 (s, 2H), 3.59 (s, 4H), 3.45 (s, 2H), 2.07 (s, 4H); <sup>13</sup>C NMR (100 MHz, DMSO- $d_6$ )  $\delta$ : 170.09, 163.94, 141.47, 133.18, 131.66, 130.86, 129.64, 128.80, 124.96, 122.25, 118.51, 115.51, 62.56, 61.43, 58.98, 38.53, 21.47; <sup>19</sup>F NMR (376 MHz, DMSO- $d_6$ )  $\delta$ : -61.73;

HRMS (ESI) m/z calcd. for  $C_{25}H_{24}F_6N_3O_2^+$  [M-Br]<sup>+</sup>: 512.1767, found 512.1777. Synthesis of **APTCs** with a thiourea group (**APTC-2** and **APTC-5**) APTC-2 and APTC-5 were synthesized according to the literature procedure.<sup>1</sup>



APTC-2: X = CH<sub>2</sub>, 47% overall yields in 4 steps; APTC-5: X = CH<sub>2</sub>O, 42% overall yields in 4 steps

**APTC-2**: light yellow powder, 47% yield for 4 steps; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$ : 7.49 (s, 1H), 7.38-7.24 (m, 5H), 7.21 (s, 2H), 4.30 (s, 2H), 3.85-3.82 (m, 2H), 3.37-3.35 (m, 6H), 2.18-2.14 (m, 4H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$ : 154.10, 150.74, 135.97, 132.80, 129.02, 128.66, 127.68, 124.79, 122.86, 122.08, 116.06, 55.26, 54.83, 39.09, 36.08, 23.35; <sup>19</sup>F NMR (376 MHz, CDCl<sub>3</sub>)  $\delta$ : -62.80; HRMS (ESI) *m/z* calcd. for C<sub>22</sub>H<sub>24</sub>F<sub>6</sub>N<sub>3</sub>S<sup>+</sup> [M-Br]<sup>+</sup>: 476.1590, found 476.1598.

**APTC-5**: light yellow powder, 42% yield for 4 steps; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$ : 7.49 (s, 1H), 7.32-7.27 (m, 5H), 7.20 (s, 2H), 4.93 (s, 2H), 4.06 (s, 2H), 3.68 (t, *J* = 4.68 Hz, 4H), 2.56 (t, *J* = 6.52 Hz, 2H), 2.49 (br, 3H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$ : 151.11, 133.39, 132.93, 128.83, 128.60, 127.81, 124.85, 122.83, 122.14, 115.78, 66.78, 56.33, 53.10, 38.85, 35.78; <sup>19</sup>F NMR (376 MHz, CDCl<sub>3</sub>)  $\delta$ : -64.22; HRMS (ESI) *m/z* calcd. for C<sub>22</sub>H<sub>24</sub>F<sub>6</sub>N<sub>3</sub>OS<sup>+</sup> [M-Br]<sup>+</sup>: 492.1539, found 492.1543.

C. Synthesis of APTC-1'(mono-NMe-APTC-1)



The free amino group  $(-NH_2)$  in 2-(pyrrolidin-1-yl)ethan-1-amine (330 mg) was protected by Boc<sub>2</sub>O in the mixed solvent of THF-H<sub>2</sub>O in the presence of NaHCO<sub>3</sub> to provide N-Boc-2-(pyrrolidin-1-yl)ethan-1-amine in quant. Yield which was directed reduced by 2.5 eq. LiAlH<sub>4</sub> in THF following by addition of 3,5-bistrifluorometylphenylisocyanate to give urea. To the above-prepared urea in dry toluene (4.0 mL) was added by 1.1 eq. of BnBr in dry toluene (1.0 mL) at room temperature (r.t.). The mixture was heated to 100 °C by an oilbath and stirred for 8 h under inert atmosphere. The reaction mixture was cooled to r.t. and the precipitate was collected and washed by dry Et<sub>2</sub>O for three to five times. This precipitate was dried under a reduced pressure to give **APTC-1'** as a white powder (76% overall yield for 4 steps) which was used directly without further purification.

**APTC-1**<sup>2</sup>: white powder, <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub> plus one drop of MeOD)  $\delta$ : 8.04 (s, 2H), 7.50-7.48 (m, 6H), 4.57 (s, 2H), 4.02 (t, *J* = 5.64 Hz, 2H), 3.88-3.84 (m, 2H), 3.74 (t, *J* = 5.76 Hz, 2H), 3.61-3.57 (m, 2H), 3.25 (s, 3H, N-Me), 2.34 (m, 2H), 2.07 (m, 2H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub> plus one drop of MeOD)  $\delta$ : 156.18, 141.05, 132.50, 131.64, 131.12, 129.67, 127.07, 124.86, 121.95, 119.87, 116.03, 61.90, 61.00, 43.44, 35.86, 21.38; <sup>19</sup>F NMR (376 MHz, CDCl<sub>3</sub>)  $\delta$ : -63.11.

2. The cycloaddition of CO<sub>2</sub> to epoxides by APTCs

*Typical procedure:* 

A mixture of 0.5 mmol of epoxide **1a** and 2.5 mol% of **APTC-1** (6.8 mg) was heated at 80 °C for 24 h under  $CO_2$  atmosphere (1atm, using a balloon). After cooled to room temperature, a small amount of  $CH_2Cl_2$  was added to the mixture and it was purified by a flash column chromatography (petroether/ethyl acetate = 10/1 to 3/1, v/v) to give cyclic carbonate **2a** as off-white solid.

Table S1. APTC-1 catalyzed cycloaddition of CO<sub>2</sub> to epoxides.



**2a**:<sup>2</sup> white powder, 87% yield; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$ : 7.32 (t, J = 8.36 Hz, 2H), 7.03 (t, J = 7.36 Hz, 1H), 6.92 (d, J = 7.96 Hz, 2H), 5.07-5.02 (m, 1H), 4.62 (t, J = 8.48 Hz, 1H), 4.54 (dd, J = 5.88, 2.52 Hz, 1H), 4.25 (dd, J = 6.76, 3.88 Hz, 1H), 4.14 (dd, J = 7.12, 3.52 Hz, 1H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$ : 157.78, 154.86, 129.71, 121.97, 114.61, 74.26, 66.87, 66.25; HRMS (ESI) *m/z* calcd. for C<sub>10</sub>H<sub>10</sub>O<sub>4</sub>Na<sup>+</sup> [M+Na]<sup>+</sup>: 217.0471, found 217.0473.

**2b**:<sup>2</sup> white powder, 91% yield; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$ : 7.41-7.33 (m, 5H), 4.86-4.81 (m, 1H), 4.61(q, J = 10.08 Hz, 2H), 4.50 (t, J = 8.36 Hz, 1H), 4.40(dd, J = 6.04, 2.28 Hz, 1H), 3.73(dd, J = 7.12, 3.84 Hz, 1H), 3.63(dd, J = 7.28, 3.68 Hz, 1H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$ : 155.03, 137.11, 128.61, 128.11, 127.79, 75.06, 73.69, 68.84, 66.32; HRMS (ESI) *m/z* calcd. for C<sub>11</sub>H<sub>12</sub>NaO<sub>4</sub><sup>+</sup> [M+Na]<sup>+</sup>: 231.0628, found 231.0631.

**2c**:<sup>2</sup> white powder, 90% yield; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$ : 8.05 (s, 1H), 8.03 (s, 1H), 7.62 (t, *J* = 7.40 Hz, 1H), 7.48 (t, *J* = 7.81 Hz, 2H), 5.10-5.06 (m, 1H), 4.67-4.59 (m, 2H), 4.53 (dd, *J* = 8.72, 3.88 Hz, 1H), 4.45 (dd, *J* = 5.64, 3.12 Hz, 1H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$ : 165.98, 154.53, 133.77, 129.80, 128.68, 73.94, 66.13, 63.66; HRMS (ESI) *m/z* calcd. for C<sub>11</sub>H<sub>10</sub>NaO<sub>5</sub><sup>+</sup> [M+Na]<sup>+</sup>: 245.0420, found 245.0418.

**2d**:<sup>2</sup> white powder, 90% yield; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$ : 7.05-6.89 (m, 4H), 5.04-5.00 (m, 1H), 4.61 (d, J = 7.00 Hz, 2H), 4.22 (q, J = 4.44 Hz, 2H), 3.85 (s, 3H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$ : 154.90, 150.36, 147.46, 123.41, 121.02, 116.59, 112.53, 74.58, 69.31, 66.37, 55.89; HRMS (ESI) *m/z* calcd. for C<sub>11</sub>H<sub>12</sub>NaO<sub>5</sub><sup>+</sup> [M+Na]<sup>+</sup>: 247.0577, found 247.0589.

**2e**:<sup>2</sup> white powder, 89% yield; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$ : 7.18 (d, J = 4.60 Hz, 2H), 6.95 (t, J = 7.36 Hz, 1H), 6.80 (d, J = 8.44 Hz, 1H), 5.08-5.04 (m, 1H), 4.66-4.57 (m, 2H), 4.27 (dd, J = 7.56, 3.12 Hz, 1H), 4.13 (dd, J = 7.96, 2.72 Hz, 1H), 2.24 (s, 1H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$ : 155.81, 154.92, 131.07, 127.07, 126.92, 121.64, 110.89, 74.34, 67.05, 66.27, 15.96; HRMS (ESI) *m/z* calcd. for C<sub>11</sub>H<sub>12</sub>NaO<sub>4</sub><sup>+</sup> [M+Na]<sup>+</sup>: 231.0628, found 231.0624. **2f**: <sup>2</sup> white powder, 81% yield; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$ : 8.14 (t, J = 5.24 Hz, 1H), 7.79 (t, J = 3.96 Hz, 1H), 7.52-7.47 (m, 3H), 7.35 (t, J = 7.60Hz, 1H), 6.77 (d, J = 7.60 Hz, 1H), 5.15 (m, 1H), 4.70-4.63 (m, 2H), 4.42 (dd, J = 7.64, 3.08 Hz, 1H), 4.26 (d, J = 10.72 Hz, 1H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$ : 155.15, 153.37, 134.47, 127.53, 126.77, 125.84, 125.49, 125.19, 121.61, 121.47, 104.83, 74.39, 67.21, 66.40; HRMS (ESI) *m/z* calcd. for C<sub>14</sub>H<sub>12</sub>NaO<sub>4</sub><sup>+</sup> [M+Na]<sup>+</sup>: 267.0628, found 267.0627.

**2**g:<sup>2</sup> white powder, 66% yield; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$ : 7.45-7.36 (m, 5H), 5.69 (t, *J* = 8.00 Hz, 1H), 4.81 (t, *J* = 8.40 Hz, 1H), 4.34 (dd, *J* = 7.84, 0.76 Hz, 1H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$ : 154.83, 135.81,129.76, 129.26, 125.89, 78.00, 71.18; HRMS (ESI) *m/z* calcd. for C<sub>9</sub>H<sub>8</sub>NaO<sub>3</sub><sup>+</sup> [M+Na]<sup>+</sup>: 187.0366, found 187.0372.

**2h**:<sup>2</sup> white powder, 65% yield; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$ : 7.38 (t, *J* = 7.08 Hz, 2H), 7.15 (t, *J* = 8.08 Hz, 2H), 5.69 (t, *J* = 7.96 Hz, 1H), 4.82 (t, *J* = 8.68 Hz, 1H), 4.34 (t, *J* = 8.60 Hz, 1H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$ : 164.64, 162.16, 154.63, 131.58, 128.07, 127.99, 116.49, 116.27, 71.11; <sup>19</sup>F NMR (376 MHz, CDCl<sub>3</sub>)  $\delta$ : -110.90; HRMS (ESI) *m/z* calcd. for C<sub>9</sub>H<sub>7</sub>FNaO<sub>3</sub><sup>+</sup> [M+Na]<sup>+</sup>: 205.0271, found 205.0276.

**2i**.<sup>2</sup> white powder, 70% yield; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$ : 7.43 (d, *J* = 8.20 Hz, 2H), 7.32 (d, *J* = 8.16 Hz, 2H), 5.68 (t, *J* = 7.92 Hz, 1H), 4.82 (t, *J* = 8.40 Hz, 1H), 4.32 (t, *J* = 8.12 Hz, 1H); HRMS (ESI) *m/z* calcd. for C<sub>9</sub>H<sub>7</sub>ClNaO<sub>3</sub>+ [M+Na]<sup>+</sup>: 220.9976, found 220.9980.

**2j**: <sup>2</sup> white powder, 70% yield; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$ : 7.46 (br, 3H), 7.40 (br, 2H), 5.69 (t, *J* = 7.76 Hz, 1H), 4.82 (t, *J* = 8.24 Hz, 1H), 4.36 (t, *J* = 7.84 Hz, 1H), 2.19 (s, 1H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$ : 154.77, 135.83, 129.74, 129.25, 125.85, 77.96, 71.14; HRMS (ESI) *m/z* calcd. for C<sub>9</sub>H<sub>8</sub>NaO<sub>3</sub><sup>+</sup> [M+Na]<sup>+</sup>: 187.0366, found 187.0368; HPLC: Chiralpak OD-H (<sup>*i*</sup>PrOH/*n*-Hexane) = 20:80, flow rate = 1.0 mL/min, 216 nm, *rac*-form: t<sub>r</sub> = 12.051 and 14.617 min; *enan*-form: *R*-major t<sub>r</sub> = 12.238 and *S*-minor t<sub>r</sub> = 14.870 min, 78.3% *ee*.

**2k**:<sup>2</sup> white powder, 71% yield; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$ : 7.45 (br, 3H), 7.39 (br, 2H), 5.69 (t, *J* = 7.68 Hz, 1H), 4.82 (t, *J* = 8.28 Hz, 1H), 4.36 (t, *J* = 7.96 Hz, 1H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$ : 154.82, 135.85, 129.73, 129.24, 125.88, 77.99, 71.16; HRMS (ESI) *m/z* calcd. for C<sub>9</sub>H<sub>8</sub>NaO<sub>3</sub><sup>+</sup> [M+Na]<sup>+</sup>: 187.0366, found 187.0368; HPLC: Chiralpak OD-H (<sup>*i*</sup>PrOH/*n*-Hexane) = 20:80, flow rate = 1.0 mL/min, 216 nm, *rac*-form: t<sub>r</sub> = 12.051 and 14.617 min; *enan*-form: *S*-major t<sub>r</sub> = 15.007 and *R*-minor t<sub>r</sub> = 14.876 min, 66.7% *ee*.

**21**:<sup>2</sup> white powder, 90% yield; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$ : 7.33 (t, J = 7.96 Hz, 2H), 7.04 (t, J = 7.36 Hz, 1H), 6.93 (d, J = 8.00 Hz, 2H), 5.07-5.02 (m, 1H), 4.64 (t, J = 8.44 Hz, 1H), 4.56 (dd, J = 5.92, 2.48 Hz, 1H), 4.26 (dd, J = 6.44, 4.12 Hz, 1H), 4.16 (dd, J = 7.08, 3.52 Hz, 1H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$ : 157.76, 154.70, 129.71, 122.01, 114.61, 74.13, 66.86, 66.25; HRMS (ESI) *m/z* calcd. for C<sub>10</sub>H<sub>10</sub>O<sub>4</sub>Na<sup>+</sup> [M+Na]<sup>+</sup>: 217.0471, found 217.0468; HPLC: Chiralpak OD-H (<sup>*i*</sup>PrOH/*n*-Hexane) = 20:80, flow rate = 1.0 mL/min, 216 nm, *rac*-form: t<sub>r</sub> = 24.394 and 31.924 min; *S*-form: t<sub>r</sub> = 31.441 min, >99% *ee*.

**2m**:<sup>2</sup> white powder, 85% yield; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$ : 7.40-7.28 (m, 5H), 4.84-4.79 (m, 1H), 4.60 (q, J = 9.52 Hz, 2H), 4.48 (t, J = 8.36 Hz, 1H), 4.39 (dd, J = 6.00, 2.24 Hz, 1H), 3.72 (dd, J = 7.32, 3.68 Hz, 1H), 3.62 (dd, J = 7.36, 3.64 Hz, 1H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$ : 155.07, 137.18, 128.59, 128.08, 127.76, 75.13, 73.65, 68.88, 66.30; HRMS (ESI) *m/z* calcd. for C<sub>11</sub>H<sub>12</sub>NaO<sub>4</sub><sup>+</sup> [M+Na]<sup>+</sup>: 231.0628, found 231.0619; HPLC: Chiralpak OD-H (*PrOH/n*-Hexane) = 10:90, flow rate = 1.0 mL/min, 254 nm, *rac*-form: t<sub>r</sub> = 35.168 and 49.587 min; *R*-form: t<sub>r</sub> = 34.856 min, >99% *ee*.

**2n**:<sup>2</sup> white powder, 89% yield; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$ : 7.40-7.28 (m, 5H), 4.85-4.79 (m, 1H), 4.60 (q, J = 9.36 Hz, 1H), 4.48 (t, J = 8.36 Hz, 1H), 4.38 (dd, J = 6.00, 2.24 Hz, 1H), 3.72 (dd, J = 7.36, 3.64 Hz, 1H), 3.61 (dd, J = 7.36, 3.64 Hz, 1H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$ : 155.08, 137.19, 128.59, 128.07, 127.76, 75.14, 73.64, 68.89, 66.30; HRMS (ESI) *m/z* calcd. for C<sub>11</sub>H<sub>12</sub>NaO<sub>4</sub><sup>+</sup> [M+Na]<sup>+</sup>: 231.0628, found 231.0624; HPLC: Chiralpak OD-H ('PrOH/*n*-Hexane) = 10:90, flow rate = 1.0 mL/min, 254 nm, t<sub>r</sub> = 35.168 and 49.587 min; *S*-form: t<sub>r</sub> = 49.568 min, >99% *ee*.

### 3. The synthesis of chiral phase-transfer catalysts (CPTC-1~15)

**CPTC-1**, **CPTC-2**, **CPTC-6**, **CPTC-7~14** are known compounds, and the synthesis of them can be found in our previous report.<sup>3</sup> **CPTC-3~5** were prepared as the procedure of **APTC-1**.



**CPTC-3**: white powder, 87% yield for two steps;  $[\alpha]^{20}_D - 25^\circ$  (c = 0.20, CH<sub>2</sub>Cl<sub>2</sub>); <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$ : 9.57 (s, 1H), 8.10 (s, 2H), 8.00-7.93 (m, 1H), 7.60-7.41(m, 5H), 4.82 (d, *J* = 13.44 Hz, 1H), 4.63 (m, 1H), 4.45 (d, *J* = 13.48 Hz, 1H), 4.24 (m, 1H), 3.83 (dd, *J* = 11.44, 2.44 Hz, 1H), 3.73-3.61 (m, 3H), 3.30 (d, *J* = 13.56 Hz, 1H), 2.49 (m, 1H), 2.34 (m, 1H), 2.17 (m, 2H), 1.95-1.88 (m, 1H), 1.84 (s, 2H), 1.09 (d, *J* = 6.76 Hz, 6H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$ : 155.88, 141.26, 132.27, 131.97, 131.65, 131.31, 129.81, 126.94, 124.73, 122.02, 118.04, 115.09, 63.42, 61.92, 61.69, 61.58, 49.64, 32.87, 21.61, 20.42, 19.62, 17.37; <sup>19</sup>F NMR (376 MHz, CDCl<sub>3</sub>)  $\delta$ : -62.86; HRMS (ESI) *m/z* calcd. for C<sub>25</sub>H<sub>34</sub>BrF<sub>6</sub>N<sub>4</sub>O<sup>+</sup> [M+NH<sub>4</sub>]<sup>+</sup>: 599.1815, found 599.1832.

**CPTC-4**: white powder, 79% yield for two steps;  $[\alpha]^{20}_D$  -14° (c = 0.25, CH<sub>2</sub>Cl<sub>2</sub>); <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$ : 8.60 (s, 1H), 7.81 (d, *J* = 9.6 Hz, 1H), 7.55-7.50 (m, 4H), 7.22 (s, 2H), 6.64 (s, 1H), 4.87 (d, *J* = 13.52 Hz, 1H), 4.62 (t, *J* = 12.76 Hz, 1H), 4.43 (d, *J* = 13.44 Hz, 1H), 4.30-4.23 (m, 1H), 3.76 (t, *J* = 12.76 Hz, 1H), 3.62 (m, 2H), 3.27 (d, *J* = 13.72 Hz, 1H), 2.45 (s, 1H), 2.27 (s, 6H), 2.19 (s, 1H), 2.13 (m, 2H), 1.89 (t, *J* = 5.52 Hz, 1H), 1.85 (s, 1H), 1.7 (d, *J* = 6.56 Hz, 6H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$ : 156.26, 138.32, 132.45, 131.06, 129.65, 127.29, 124.05, 116.51, 63.41, 61.75, 49.57, 32.87, 21.63, 21.38, 20.50, 19.68, 17.52; HRMS (ESI) *m/z* calcd. for C<sub>25</sub>H<sub>36</sub>BrN<sub>3</sub>NaO<sup>+</sup> [M+Na]<sup>+</sup>: 499.1934, found 499.1966.

**CPTC-5**: white powder, 82% yield for two steps;  $[\alpha]^{20}_D$  -32° (c = 0.20, CH<sub>2</sub>Cl<sub>2</sub>); <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$ : 8.78 (s, 1H), 7.68-7.65 (m, 5H), 7.19 (s, 2H), 6.65 (s, 1H), 5.03 (d, *J* = 13.04 Hz, 1H), 4.82 (d, *J* = 13.2 Hz, 1H), 4.51 (m, 1H), 4.22 (m, 1H), 3.91-3.85 (m, 2H), 3.56 (d, *J* = 7.28 Hz, 1H), 3.50-3.46 (m, 1H), 3.34 (d, *J* = 6.90 Hz, 1H), 2.36 (m, 1H), 2.26 (s, 6H), 2.20-2.14 (m, 1H), 2.08 (s, 2H), 1.92 (s, 2H), 1.09-1.06 (m, 6H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$ : 156.31, 139.22, 136.48, 133.27, 126.46, 124.27,116.48, 63.41, 61.96, 61.21, 60.90, 49.82, 32.87, 21.40, 21.22, 20.62, 19.66, 17.65; <sup>19</sup>F NMR (376 MHz, CDCl<sub>3</sub>)  $\delta$ : -63.12; HRMS (ESI) *m/z* calcd. for C<sub>26</sub>H<sub>35</sub>BrF<sub>3</sub>KN<sub>3</sub>O<sup>+</sup> [M+K]<sup>+</sup>: 580.1547, found 580.1549.

4. The kinetic resolution of epoxides by CO<sub>2</sub> in the presence of CPTCs

Typical procedure:

A mixture of 0.25 mmol of epoxide **1g** and 1.0 mol% of **CPTC-2** (1.5 mg) was heated at 60 °C for 24 h under  $CO_2$  atmosphere (1atm, using a balloon). After cooled to room temperature, a small amount of  $CH_2Cl_2$  was added to the mixture and it was purified by a flash column chromatography (petroether/ethyl acetate = 10/1 to 3/1, v/v) to give cyclic carbonate and epoxide respectively. The *ee* of products were determined by HPLC using a chiral column (Daicel Chiralpak AD-H or OD-H), and the absolute configurations of products were assigned according to literature report.<sup>4</sup>

### **Notes and References**

- 1. J. Novacek, M. Waser, Eur. J. Org. Chem., 2014, 802.
- (a) W. Clegg, R. W. Harrington, M. North, R. Pasquale, *Chem.-Eur. J.*, 2010, 16, 6828; (b) L. Wu, H. Yang, H. Wang, J. Lu, *RSC Adv.*, 2015, 5, 23189; (c) H. Büttner, J. Steinbauer, T. Werner, *ChemSusChem*, 2015, 8, 2655.
- 3. J.-C. Zhu, D.-X. Cui, Y.-D. Li, J.-X. He, W.-P. Chen, P.-A. Wang, Org. Biomol. Chem., 2018, 16, 3012.
- (a) Y. Toda, Y. Komiyama, A. Kikuchi, H. Suga, ACS Catal., 2016, 6, 6906; (b) S.-Y. Liu, N. Suematsu, K. Maruoka, S. Shirakawa, Green Chem., 2016, 18, 4611; (c) J. Qin, V. A. Larionov, K. Harms, E. Meggers, ChemSusChem, 2019, 12, 320; (d) T. Ema, M. Yokoyama, S. Watanabe, S. Sasaki, H. Ota, K. Takaishi, Org. Lett., 2017, 19, 4070; (e) K. Takaishi, T. Okuyama, S. Kadosaki, M. Uchiyama, T. Ema, Org. Lett. 2019, 21, 1397.



NMR spectra copies of all new compounds







S12























































10.0 9.5 9.0 8.5 8.0
























## HRMS copies of all new compounds



S43



















Display Report							
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HPLC copies of cyclic carbonates from APTC-1 catalyzed fixation of CO<sub>2</sub> with chiral epoxides



HPLC: Chiralpak OD-H (<sup>*i*</sup>PrOH/*n*-Hexane) = 20:80, flow rate = 1.0 mL/min, 216 nm.



HPLC: Chiralpak OD-H (<sup>*i*</sup>PrOH/*n*-Hexane) = 20:80, flow rate = 1.0 mL/min, 216 nm.



HPLC: Chiralpak OD-H (<sup>*i*</sup>PrOH/*n*-Hexane) = 10:90, flow rate = 1.0 mL/min, 254 nm.



HPLC copies of producs from the kinetic resolution of rac-epoxides by CO<sub>2</sub> with CPTCs



For epoxide 1g and 1g':

HPLC: Chiralpak AD-H (<sup>*i*</sup>PrOH/*n*-Hexane) = 0.2:99.8, flow rate = 0.3 mL/min, 216 nm.



For cyclic carbonate **2g** and **2j**:

HPLC: Chiralpak OD-H (<sup>i</sup>PrOH/*n*-Hexane) = 20:80, flow rate = 1.0 mL/min, 216 nm.





For epoxide **1g** and **1g'**: HPLC: Chiralpak AD-H (*i*PrOH/*n*-Hexane) = 1:99, flow rate = 0.3 mL/min, 216 nm.



For cyclic carbonate 2g and 2j:

HPLC: Chiralpak OD-H (<sup>*i*</sup>PrOH/*n*-Hexane) = 20:80, flow rate = 1.0 mL/min, 216 nm.






For epoxide: HPLC: Chiralpak AD-H (<sup>*i*</sup>PrOH/*n*-Hexane) = 1:99, flow rate = 0.3 mL/min, 216 nm. For cyclic carbonate HPLC: Chiralpak OD-H (<sup>*i*</sup>PrOH/*n*-Hexane) = 20:80, flow rate = 1.0 mL/min, 216 nm.



For cyclic carbonate HPLC: Chiralpak OD-H (PrOH/n-Hexane) = 20:80, flow rate = 1.0 mL/min, 216 nm.





For cyclic carbonate HPLC: Chiralpak OD-H (PrOH/n-Hexane) = 20:80, flow rate = 1.0 mL/min, 216 nm.



For epoxide: HPLC: Chiralpak AD-H (<sup>*i*</sup>PrOH/*n*-Hexane) = 1:99, flow rate = 0.3 mL/min, 216 nm. For cyclic carbonate HPLC: Chiralpak OD-H (<sup>*i*</sup>PrOH/*n*-Hexane) = 20:80, flow rate = 1.0 mL/min, 216 nm.







For cyclic carbonate HPLC: Chiralpak OD-H (*i*PrOH/*n*-Hexane) = 20:80, flow rate = 1.0 mL/min, 216 nm.



For cyclic carbonate HPLC: Chiralpak OD-H (<sup>*i*</sup>PrOH/*n*-Hexane) = 5:95, flow rate = 1.0 mL/min, 216 nm.





For epoxide: HPLC: Chiralpak OD-H (<sup>*i*</sup>PrOH/*n*-Hexane) = 10:90, flow rate = 1.0 mL/min, 254 nm. For cyclic carbonate HPLC: Chiralpak OD-H (<sup>*i*</sup>PrOH/*n*-Hexane) = 20:80, flow rate = 1.0 mL/min, 216 nm.



For epoxide: HPLC: Chiralpak OD-H (<sup>*i*</sup>PrOH/*n*-Hexane) = 10:90, flow rate = 1.0 mL/min, 254 nm. For cyclic carbonate HPLC: Chiralpak OD-H (<sup>*i*</sup>PrOH/*n*-Hexane) = 20:80, flow rate = 1.0 mL/min, 216 nm.





For cyclic carbonate HPLC: Chiralpak OD-H (PrOH/n-Hexane) = 20:80, flow rate = 1.0 mL/min, 216 nm



For epoxide: HPLC: Chiralpak OD-H (<sup>*i*</sup>PrOH/*n*-Hexane) = 10:90, flow rate = 1.0 mL/min, 254 nm. For cyclic carbonate HPLC: Chiralpak OD-H (<sup>*i*</sup>PrOH/*n*-Hexane) = 10:90, flow rate = 1.0 mL/min, 216 nm.







For epoxide: HPLC: Chiralpak OD-H (<sup>*i*</sup>PrOH/*n*-Hexane) = 10:90, flow rate = 1.0 mL/min, 254 nm. For cyclic carbonate HPLC: Chiralpak OD-H (<sup>*i*</sup>PrOH/*n*-Hexane) = 10:90, flow rate = 1.0 mL/min, 216 nm.



For epoxide: HPLC: Chiralpak OD-H (<sup>*i*</sup>PrOH/*n*-Hexane) = 10:90, flow rate = 1.0 mL/min, 254 nm. For cyclic carbonate HPLC: Chiralpak OD-H (<sup>*i*</sup>PrOH/*n*-Hexane) = 10:90, flow rate = 1.0 mL/min, 216 nm.



