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Supporting Information

Cercosporin-Bioinspired Selective Photooxidation Reactions under Mild Conditions

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1. Materials and Methods

Cercosporin was biosynthesized by a new cercosporin producing strain in our laboratory.¹ Hypocrellin A and hypocrellin B were commercially available and used without further purification. All other commercially available reagents and solvents were used without further purification. Thin-layer chromatography was performed using silica gel plates F254. Visualization was accomplished with short wavelength UV light (254 nm) and UVA light (366 nm) sources. ¹H and ¹³C NMR spectra were recorded on Bruker AV400 (400 MHz) spectrometer in CDCl₃ and DMSO-d₆ solutions with internal solvent signals (for ¹H and ¹³C) as reference (7.26 and 77.2, 2.50 and 39.5 for CDCl₃ and DMSO- d_6 , respectively). ¹H NMR data are reported as follows: chemical shift (ppm), multiplicity (s = singlet, br. s. = broad singlet, d = doublet, t = triplet, q = quartet, quint = quintet, sext = sextet, hept = heptet, dd = doubletdoublet of doublets, dd = doublet of doublets of doublets, td = triplet of doublets, gd = quartet of doublets, m = multiplet), coupling constants (Hz), and numbers of protons. Data for ¹³C NMR are reported in terms of chemical shift and no special nomenclature is used for equivalent carbons. High resolution mass spectra (HRMS) were recorded on Waters Xevo G2 Q-TOF instrument. Gas chromatography (GC) and gas chromatography coupled to low-resolution mass spectrometry (GC-MS) analysis were performed through a capillary column (length: 30 m; diam.: 0.25 mm; film: 0.25 µM) using He gas as carrier. GC (TRACE 1300) was equipped with an FID detector. GC-MS (TSQ 8000) was performed on triple quadruple detector. UV-Vis and fluorescence measurements were performed with Shimadzu UV-3600plus spectrophotometer and F-2700 spectrofluorometer. The lifetime was measured on Edinburgh FLS920 fluorescence spectrometer. IR spectra was measured on Thermo Scientific Nicolet iS5 FTIR. Cyclic voltammetry (CV) was performed using an CHI600E electrochemical workstation: Au wire ($\phi = 1.6$ mm) sealed in a Teflon jacket as working electrode, Pt wire as the counter electrode, Ag/AgCl (KCl, 3 M) electrode as the reference electrode, and ferrocenium/ferrocene (Fc⁺/Fc) as the internal standard. Scan rate: 50 mV s⁻¹ (in the range -1 to +2.2 V). Bu₄NPF₆ (0.1 M in MeCN) was used as the supporting electrolyte. Household CFLs (compact fluorescent lamps) were used as the light source. The emission spectrum of each light source was measured with Hitachi F-2700 spectrofluorometer. The intensity of irradiation was measured by a FZ-A radiometer (Photoelectric Instrument Factory of Beijing Normal University) equipped with a 400-1000 nm sensor. The sample was placed at an approximate distance of 2 cm to the lamp. The light intensity was measured to be 21.3 mW/cm² for 23w CFL and

13.7 mW/cm² for 15w CFL, respectively. Additionally, the light intensity was also measured to be 6.51 mW/cm² for 5w blue LED and 2.39 mW/cm² for 5w green LED.

2. Biosynthesis of Cercosporin

Cercosporin was biosynthesized from microbial fermentation with cheap glucose as starting material by a new cercosporin producing strain in our laboratory.¹ Briefly, the neighbor-joining phylogenetic tree was constructed based on the 18S rDNA nucleotide sequences, showing that this new strain belongs to the genus *Cercospora*, thus designated as *Cercospora* sp. JNU001 (deposited to China Center for Type Culture Collection (CCTCCNo. M2017842). Next, cercosporin was produced by liquid fermentation using S7 medium (1 g/L Gluose, 6 g/L Fructose, 3g/L Sucrose, 1 g/L sodium acetate, 1 g/L soy peptone, 5 mg/L phenylalanine, 100 mg/L Sodium benzoate, 136 mg/L KH₂PO₄, 1 mg/L biotin, 6.5 mg/L Ca(NO₃)₂, 1 mg/L pyridoxal, 1 mg/L calcium pantothenate, 1 mg/L thiamine, 5 mg/L MnCl₂, 2 mg/L FeCl₂, 1 g/L Cu(NO₃)₂, 3.6 mg/L MgSO₄ and 2.5 mg/L ZnSO₄). A single colony of *Cercospora* sp. JNU001 was inoculated into 50 mL PDA medium and cultured for 7 days at 25 °C. Then, they were inoculated into a 3 L Erlenmeyer flask containing 1L S-7 medium (pH=8.5) medium and shaked with 150 rpm at 25°C under continuous light for 10 days. Cercosporin was purified by Sephadex LH-20 column after extraction by dichloromethane and identified by HPLC, LC-MS and NMR. A maximum of cercosporin production was obtained (128.2 mg/L), providing adequate amounts for further analysis.

3. Photostability Experiment of Cercosporin

The stability experiment of cercosporin under visible light Irradiation was performed. Upon irradiation with 23 W CFL for 48h, new signal related to the photolysis of cercosporin was not detected by HPLC. This result indicated the excellent stability of cercosporin as the photocatalyst.



Fig. S1 High-performance liquid chromatogram analysis of photostability of cercosporin without irradiation (a) and with photoirradiation under 23 W CFL for 48h (b).



4. Spectra of the Light Source and Photochemical Reaction System

Fig. S2 Spectra of light source. (a) blue LED, (b) green LED, (c) CFL. (d) Photograph of the photochemical reaction set up.

5. Optimization of Reaction Conditions

5.1 Optimization of the reaction conditions for the selective oxidation of benzylic C-H bonds



Entry	Photocatalyst	Additive	Solvent	Light source	Yield $(\%)^b$
	(X mol%)				
1	CP (2)	_	DMSO	23 W CFL	Trace
2	CP (2)	_	Toluene	23 W CFL	Trace
3	CP (2)	_	DMF	23 W CFL	Trace
4	CP (2)	_	DCE	23 W CFL	Trace
5	CP (2)	_	CH ₃ CN	23 W CFL	5

Table S1	Optimization	of reaction	conditions ^a
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6 ^c	CP (2)		CH ₃ OH	23 W CFL	20
7	CP (2)	KBr (0.2 equiv)	CH ₃ OH	23 W CFL	78
8	CP (2)	LiBr (0.2 equiv)	CH ₃ OH	23 W CFL	26
9	CP (2)	NaBr (0.2 equiv)	CH ₃ OH	23 W CFL	25
10	CP (2)	CsBr (0.2 equiv)	CH ₃ OH	23 W CFL	20
11	CP (2)	KF (0.2 equiv)	CH ₃ OH	23 W CFL	30
12	CP (2)	KCl (0.2 equiv)	CH ₃ OH	23 W CFL	24
13	CP (2)	KI (0.2 equiv)	CH ₃ OH	23 W CFL	35
14	CP (2)	KO ^t Bu (0.2 equiv)	CH ₃ OH	23 W CFL	29
15	CP (3)	KBr (0.2 equiv)	CH ₃ OH	23 W CFL	78
16	CP (1)	KBr (0.2 equiv)	CH ₃ OH	23 W CFL	72
17	CP (2)	KBr (0.1 equiv)	CH ₃ OH	23 W CFL	75
18	CP (2)	KBr (0.3 equiv)	CH ₃ OH	23 W CFL	75
19	CP (2)	KBr (0.4 equiv)	CH ₃ OH	23 W CFL	78
20	CP (2)	KBr (0.2 equiv)	CH ₃ OH	15 W CFL	75
21	CP (2)	KBr (0.2 equiv)	CH ₃ OH	5 W blue LED	70
22	CP (2)	KBr (0.2 equiv)	CH ₃ OH	5 W green LED	62
23	HA (2)	KBr (0.2 equiv)	CH ₃ OH	23 W CFL	30
24	HB (2)	KBr (0.2 equiv)	CH ₃ OH	23 W CFL	15
25		KBr (0.2 equiv)	CH ₃ OH	23 W CFL	trace
26	CP (2)	KBr (0.2 equiv)	CH ₃ OH	_	trace
27 ^d	CP (2)	KBr (0.2 equiv)	CH ₃ OH	23 W CFL	trace

^{*a*}Reaction conditions: **1a** (0.25 mmol), photocatalyst in solvent (2.0 mL) at room temperature in O₂ under irradiation for 30h. ^{*b*}Yields of isolated product. ^{*c*}Reaction time was 40h. ^{*d*}Reaction was conducted in nitrogen atmosphere.

5.2 Optimization of the reaction conditions for the selective oxidation of amines





Entry	Photocatalyst	Solvent	Light source	yield	(%) ^b
	(X mol%)			4a	5a
1	CP (1)	CHCl ₃	15 W CFL	60	23
2	CP (1)	THF	15 W CFL	59	31
3	CP (1)	CH ₃ CN	15 W CFL	49	29
4	CP (1)	CH ₃ OH	15 W CFL	95	0
5	CP (1)	DMF	15 W CFL	trace	trace
6	CP (1)	DMSO	15 W CFL	trace	trace
7	CP (1)	CH ₃ OH	5 W blue LED	8	35
8	CP (1)	CH ₃ OH	5 W green LED	0	20
9	CP (2)	CH ₃ OH	15 W CFL	95	0
10	CP (0.5)	CH ₃ OH	15 W CFL	0	79
11	HA (1)	CH ₃ OH	15 W CFL	80	5
12	HB (1)	CH ₃ OH	15 W CFL	62	23
13	_	CH ₃ OH	15 W CFL	0	0
14	CP (1)	CH ₃ OH	_	0	0
15 ^c	CP (1)	CH ₃ OH	15 W CFL	0	trace

^{*a*}All reactions were carried out on a scale of 0.25 mmol of **3a** in 2 mL of solvent under 15W CFL for 8h. ^{*b*} Yields of isolated product. ^{*c*} Reaction was conducted in nitrogen atmosphere.

5.3 Optimization of the reaction conditions for the selective oxidation of sulfides

S-S-S-S-S-S-S-S-S-S-S-S-S-S-S-S-S-S-S-	$\frac{\text{Cercosporin}}{\text{solvent, O}_2, \text{ rt, light source}}$		
6a		7a	8a

Table S3 Optimization of reaction conditions ^{<i>a</i>}	imization of reaction conditions ^{<i>a</i>}
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Entry	Photocatalyst	Solvent	Light source	yield	(%) ^b
	(X mol%)			7a	8a
1	CP (1)	CHCl ₃	15 W CFL	79	0
2	CP (1)	THF	15 W CFL	20	0
3	CP (1)	CH ₃ OH	15 W CFL	95	0
4	CP (1)	DMF	15 W CFL	18	0

5	CP (1)	DMSO	15 W CFL	trace	0
6	CP (1)	CH ₃ CN	15 W CFL	81	0
7	CP (1)	CH ₃ OH	5 W blue LED	35	8
8	CP (1)	CH ₃ OH	5 W green LED	20	0
9	CP (2)	CH ₃ OH	15 W CFL	95	0
10	CP (0.5)	CH ₃ OH	15 W CFL	79	0
11	_	CH ₃ OH	15 W CFL	0	0
12	CP (1)	CH ₃ OH		0	0
13 ^c	CP (1)	CH ₃ OH	15 W CFL	trace	0

^{*a*}All reactions were carried out on a scale of 0.25 mmol of **6a** in 2 mL of solvent under 15W CFL for 6h. ^{*b*} Yields of isolated product. ^{*c*} Reaction was conducted in nitrogen atmosphere.

6. General Procedures

6.1 General procedure for the selective oxidation benzylic C-H bonds with cercosporin

In a dried schlenk tube, benzylic derivatives 1 (0.25 mmol), potassium bromide (0.2 equiv) and cercosporin (2 mol%) was added in 2.0 mL methanol. Next, a balloon was purged with oxygen and fixed on the top of the schlenk tube. The reaction mixture was stirred and irradiated by 23 W white CFL at room temperature under an atmospheric pressure oxygen atmosphere. When the reaction was finished, the reaction mixture was washed with brine. The aqueous phase was re-extracted with ethyl acetate. The combined organic extracts were dried over Na_2SO_4 , concentrated in vacuum, and the resulting residue was purified by silica gel column chromatography to afford the desired product 2.

Acetophenone (2a)²

The representative procedure was followed using ethylbenzene (**1a**) (0.25 mmol) as substrate. Isolation by column chromatography (PE/EtOAc: 10/1) yielded **2a** (23 mg, 78%) as a colorless oil. ¹H NMR (400 MHz, CDCl₃): δ ppm 7.96 (d, 2H, *J* = 8 Hz, ArH), 7.58-7.55 (m, 1H, ArH), 7.48-7.44 (m, 2H, ArH), 2.58 (s, 3H, CH₃). ¹³C NMR (101 MHz, CDCl₃) δ ppm 198.04, 137.16, 133.05, 128.54, 128.27, 26.54. 1-(4-Bromophenyl)ethan-1-one (**2b**)³



The representative procedure was followed using 1-bromo-4-ethylbenzene (**1b**) (0.25 mmol) as substrate. Isolation by column chromatography (PE/EtOAc: 10/1) yielded **2b** (32 mg, 65%) as a colorless oil. ¹H NMR (400 MHz, CDCl₃): δ ppm 7.82 (d, 2H, *J* = 8 Hz, ArH), 7.61 (d, 2H, *J* = 8 Hz, ArH), 2.59 (s, 3H, CH₃). ¹³C NMR (101 MHz, CDCl₃) δ ppm 196.89, 135.86, 131.87, 129.81, 128.25, 26.48.

 $1-(4-Methoxyphenyl)ethan-1-one (2c)^4$



The representative procedure was followed using 1-ethyl-4-methoxybenzene (1c) (0.25 mmol) as substrate. Isolation by column chromatography (PE/EtOAc: 10/1) yielded 2c (28 mg, 75%) as colorless oil. ¹H NMR (400 MHz, CDCl₃): δ ppm 7.94 (d, 2H, *J* = 8 Hz, ArH), 6.94 (d, 2H, *J* = 12 Hz, ArH), 3.87 (s, 3H, OCH₃), 2.56 (s, 3H, CH₃). ¹³C NMR (101 MHz, CDCl₃) δ ppm 196.65, 163.48, 130.54, 130.36, 113.67, 55.42, 26.26.

2,3-Dihydro-1H-inden-1-one (2d)⁵



The representative procedure was followed using 2,3-dihydro-1*H*-indene (**1d**) (0.25 mmol) as substrate. Isolation by column chromatography (PE/EtOAc: 10/1) yielded **2d** (24 mg, 73%) as colorless oil. ¹H NMR (400 MHz, CDCl₃): δ ppm 7.77 (d, 1H, *J* = 8 Hz, ArH), 7.61-7.57 (m, 1H, ArH), 7.49 (d, 1H, *J* = 4 Hz, ArH), 7.39-7.36 (m, 1H, ArH), 3.17-3.14 (m, 2H, CH₂), 2.72-2.69 (m, 2H, CH₂). ¹³C NMR (101 MHz, CDCl₃) δ ppm 206.95, 155.12, 137.07, 134.56, 127.24, 126.68, 123.65, 36.20, 25.79. 3,4-Dihydronaphthalen-1(2*H*)-one (**2e**)⁶



The representative procedure was followed using 1,2,3,4-tetrahydronaphthalene (1e) (0.25 mmol) as

substrate. Isolation by column chromatography (PE/EtOAc: 10/1) yielded **2e** (29 mg, 81%) as colorless oil. ¹H NMR (400 MHz, CDCl₃): δ ppm 8.04 (d, 1H, *J* = 8 Hz, ArH), 7.49-7.45 (m, 1H, ArH), 7.33-7.29 (m, 1H, ArH), 7.25 (d, 1H, *J* = 8 Hz, ArH), 2.99-2.96 (m, 2H, CH₂), 2.68-2.65 (m, 2H, CH₂), 2.18-2.11 (m, 2H, CH₂). ¹³C NMR (101 MHz, CDCl₃) δ ppm 198.25, 144.46, 133.35, 132.63, 128.75, 127.13, 126.59, 39.15, 29.70, 23.29.

Isobenzofuran-1(3H)-one (2f)³



The representative procedure was followed using 1,3-dihydroisobenzofuran (**1f**) (0.25 mmol) as substrate. Isolation by column chromatography (PE/EtOAc: 10/1) yielded **2f** (23 mg, 71%) as colorless oil. ¹H NMR (400 MHz, CDCl₃): δ ppm 7.93 (d, 1H, *J* = 8 Hz, ArH), 7.71-7.68 (m, 1H, ArH), 7.56-7.50 (m, 2H, ArH), 5.33 (s, 2H, CH₂). ¹³C NMR (101 MHz, CDCl₃) δ ppm 171.06, 146.59, 134.01, 128.95, 125.57, 125.48, 122.22, 69.69.

Isochroman-1-one $(2g)^7$



The representative procedure was followed using isochromane (**1g**) (0.25 mmol) as substrate. Isolation by column chromatography (PE/EtOAc: 10/1) yielded **2g** (27 mg, 75%) as colorless oil. ¹H NMR (400 MHz, CDCl₃): δ ppm 8.10 (d, 1H, *J* = 8 Hz, ArH), 7.56-7.52 (m, 1H, ArH), 7.41-7.38 (m, 1H, ArH), 7.27-7.26 (m, 1H, ArH), 4.55-4.52 (m, 2H, CH₂), 3.08-3.05 (m, 2H, CH₂). ¹³C NMR (101 MHz, CDCl₃) δ ppm 165.11, 39.64, 133.67, 130.16, 127.57, 127.53, 127.32, 125.21, 67.33, 27.75, 27.72, 27.56. Benzophenone (**2h**)⁸



The representative procedure was followed using diphenylmethane (**1h**) (0.25 mmol) as substrate. Isolation by column chromatography (PE/EtOAc: 10/1) yielded **2h** (34 mg, 75%) as colorless oil. ¹H NMR (400 MHz, CDCl₃): δ ppm 7.82-7.80 (m, 2H, ArH), 7.60-7.57 (m, 1H, ArH), 7.50-7.47 (m, 2H, ArH). ¹³C NMR (101 MHz, CDCl₃) δ ppm 196.67, 137.64, 132.39, 130.03, 128.28.

Phenyl (p-tolyl)methanone (2i)9



The representative procedure was followed using 1-benzyl-4-methylbenzene (**1i**) (0.25 mmol) as substrate. Isolation by column chromatography (PE/EtOAc: 10/1) yielded **2i** (38 mg, 78%) as colorless oil. ¹H NMR (400 MHz, CDCl₃): δ ppm 7.78 (d, 2H, *J* = 8 Hz, ArH), 7.72 (d, 2H, *J* = 8 Hz, ArH), 7.59-7.55 (m, 1H, ArH), 7.49-7.45 (m, 2H, ArH), 7.28 (d, 2H, *J* = 8 Hz, ArH), 2.44 (s, 3H, CH₃). ¹³C NMR (101 MHz, CDCl₃) δ ppm 196.47, 143.22, 137.98, 134.91, 132.15, 130.30, 129.92, 128.98, 128.21, 21.65. Phenyl (*o*-tolyl)methanone (**2j**)¹⁰



The representative procedure was followed using 1-benzyl-2-methylbenzene (**1j**) (0.25 mmol) as substrate. Isolation by column chromatography (PE/EtOAc: 10/1) yielded **2j** (37 mg, 77%) as colorless oil. ¹H NMR (400 MHz, CDCl₃): δ ppm 7.81-7.79 (m, 2H, ArH), 7.59-7.56 (m, 1H, ArH), 7.47-7.43 (m, 2H, ArH), 7.41-7.37 (m, 1H, ArH), 7.32-7.22 (m, 3H, ArH), 2.33 (s, 3H, CH₃). ¹³C NMR (101 MHz, CDCl₃) δ ppm 197.60, 137.61, 136.73, 135.70, 132.08, 129.96, 129.19, 129.09, 127.47, 127.42, 124.16, 18.93.

o-Tolyl(p-tolyl)methanone (2k)¹¹



The representative procedure was followed using 1-methyl-2-(4-methylbenzyl)benzene (**1k**) (0.25 mmol) as substrate. Isolation by column chromatography (PE/EtOAc: 10/1) yielded **2k** (42 mg, 80%) as colorless oil. ¹H NMR (400 MHz, CDCl₃): δ ppm 7.70 (d, 2H, *J* = 8 Hz, ArH), 7.39-7.36 (m, 1H, ArH), 7.30-7.22 (m, 5H, ArH), 2.42 (s, 3H, CH₃), 2.31 (s, 3H, CH₃). ¹³C NMR (101 MHz, CDCl₃) δ ppm 198.32, 144.07, 139.03, 136.46, 135.21, 130.91, 130.30, 130.01, 129.20, 128.26, 125.19, 21.71, 19.90. Di*-p*-tolylmethanone (**2l**)¹²



The representative procedure was followed using di-*p*-tolylmethane (**11**) (0.25 mmol) as substrate. Isolation by column chromatography (PE/EtOAc: 10/1) yielded **21** (39 mg, 76%) as colorless oil. ¹H NMR (400 MHz, CDCl₃): δ ppm 7.70 (d, 2H, *J* = 8 Hz, ArH), 7.29 (d, 2H, *J* = 8 Hz, ArH), 2.44 (s, 3H, CH₃). ¹³C NMR (101 MHz, CDCl₃) δ ppm 196.23, 142.91, 135.25, 130.18, 128.90, 21.62.

(4-(*tert*-Butyl) phenyl) (phenyl) methanone (**2m**)¹³



The representative procedure was followed using 1-benzyl-4-(tert-butyl) benzene (**1m**) (0.25 mmol) as substrate. Isolation by column chromatography (PE/EtOAc: 10/1) yielded **2m** (47 mg, 80%) as colorless oil. ¹H NMR (400 MHz, CDCl₃): δ ppm 7.81-7.75 (m, 4H, ArH), 7.60-7.56 (m, 1H, ArH), 7.51-7.49 (m, 4H, ArH), 1.37 (s, 9H, 3CH₃). ¹³C NMR (101 MHz, CDCl₃) δ ppm 196. 42, 156.18, 137.97, 134.85, 132.16, 130.14, 129.96, 128.21, 125.25, 35.12, 31.17.

(4-(*tert*-Butyl) phenyl) (o-tolyl)methanone (2n)¹⁴



The representative procedure was followed using 1-(4-(*tert*-butyl) benzyl)-2-methylbenzene (**1n**) (0.25 mmol) as substrate. Isolation by column chromatography (PE/EtOAc: 10/1) yielded **2n** (49 mg, 79%) as colorless oil. ¹H NMR (400 MHz, CDCl₃): δ ppm 7.75 (d, 2H, *J* = 8 Hz, ArH), 7.46 (d, 2H, *J* = 8 Hz, ArH), 7.39-7.36 (m, 1H, ArH), 7.31-7.21 (m, 3H, ArH), 2.33 (s, 3H, CH₃), 1.35 (s, 9H, 3CH₃). ¹³C NMR (101 MHz, CDCl₃) δ ppm 198.21, 156.96, 139.01, 136.54, 135.10, 130.91, 130.17, 129.99, 128.31, 125.43, 125.12, 35.17, 31.13, 19.90.

(4-(*tert*-Butyl) phenyl) (*p*-tolyl) methanone (**20**)¹¹



The representative procedure was followed using 1-(tert-butyl)-4-(4-methylbenzyl) benzene (10) (0.25

mmol) as substrate. Isolation by column chromatography (PE/EtOAc: 10/1) yielded **2o** (47 mg, 76%) as colorless oil. ¹H NMR (400 MHz, CDCl₃): δ ppm 7.76-7.71 (m, 4H, ArH), 7.49 (d, 2H, *J* = 4 Hz, ArH), 7.27 (d, 2H, *J* = 8 Hz, ArH), 2.44 (s, 3H, CH₃), 1.36 (s, 9H, 3CH₃). ¹³C NMR (101 MHz, CDCl₃) δ ppm 196.15, 155.87, 142.90, 135.24, 135.19, 130.21, 130.00, 128.89, 125.16, 35.08, 31.17, 21.62.

Bis (4-(*tert*-butyl)phenyl) methanone (2p)¹⁵



The representative procedure was followed using bis(4-(*tert*-butyl) phenyl) methane (**1p**) (0.25 mmol) as substrate. Isolation by column chromatography (PE/EtOAc: 10/1) yielded **2p** (60 mg, 82%) as colorless oil. ¹H NMR (400 MHz, CDCl₃): δ ppm 7.76 (d, 2H, *J* = 8 Hz, ArH), 7.49 (d, 2H, *J* = 8 Hz, ArH), 1.37 (s, 9H, 3CH₃). ¹³C NMR (101 MHz, CDCl₃) δ ppm 196.13, 155.87, 135.18, 130.04, 125.16, 35.09, 31.18.

(4-Fluorophenyl) (phenyl) methanone $(2q)^{16}$

The representative procedure was followed using 1-benzyl-4-fluorobenzene (**1q**) (0.25 mmol) as substrate. Isolation by column chromatography (PE/EtOAc: 10/1) yielded **2q** (31 mg, 62%) as colorless oil. ¹H NMR (400 MHz, CDCl₃): δ ppm 7.87-7.83 (m, 2H, ArH), 7.78-7.76 (m, 2H, ArH), 7.61-7.58 (m, 1H, ArH), 7.51-7.47 (m, 2H, ArH), 7.18-7.14 (m, 2H, ArH). ¹³C NMR (101 MHz, CDCl₃) δ ppm 195.16, 166.64, 164.11, 137.53, 133.85, 133.82, 132.68, 132.59, 132.43, 129.84, 128.34, 115.54, 115.32. (4-Fluorophenyl) (*p*-tolyl) methanone (**2r**)¹⁷



The representative procedure was followed using 1-fluoro-4-(4-methylbenzyl)benzene (1r) (0.25 mmol)

as substrate. Isolation by column chromatography (PE/EtOAc: 10/1) yielded **2r** (32 mg, 60%) as colorless oil. ¹H NMR (400 MHz, CDCl₃): δ ppm 7.84-7.81 (m, 2H, ArH), 7.69 (d, 2H, *J* = 8 Hz, ArH), 7.30-7.26 (m, 2H, ArH), 7.17-7.13 (m, 2H, ArH), 2.44 (s, 3H, CH₃). ¹³C NMR (101 MHz, CDCl₃) δ ppm 195.01, 166.50, 163.98, 143.31, 134.80, 134.17, 134.14, 132.54, 132.45, 130.12, 129.04, 115.46, 115.24, 21.63.

(4-(*tert*-Butyl)phenyl) (4-fluorophenyl) methanone (2s)¹⁸



The representative procedure was followed using 1-(*tert*-butyl)-4-(4-fluorobenzyl) benzene (**1s**) (0.25 mmol) as substrate. Isolation by column chromatography (PE/EtOAc: 10/1) yielded **2s** (44 mg, 69%) as colorless oil. ¹H NMR (400 MHz, CDCl₃): δ ppm 7.86-7.83 (m, 2H, ArH), 7.43 (d, 2H, *J* = 8 Hz, ArH), 7.50 (d, 2H, *J* = 8 Hz, ArH), 7.17-7.13 (m, 2H, ArH), 1.37 (s, 9H, 3CH₃). ¹³C NMR (101 MHz, CDCl₃) δ ppm 194.94, 166.51, 163.99, 156.26, 134.74, 134.15, 134.12, 132.59, 132.50, 129.96, 125.32, 115.45, 115.23, 115.18, 114.97, 35.12, 31.14.

Bis (4-fluorophenyl) methanone $(2t)^{19}$



The representative procedure was followed using bis (4-fluorophenyl) methane (1t) (0.25 mmol) as substrate. Isolation by column chromatography (PE/EtOAc: 10/1) yielded **2t** (26 mg, 48%) as colorless oil. ¹H NMR (400 MHz, CDCl₃): δ ppm 7.83-7.80 (m, 2H, ArH), 7.19-7.15 (m, 2H, ArH). ¹³C NMR (101 MHz, CDCl₃) δ ppm 193.70, 166.65, 164.12, 133.74, 133.71, 132.50, 132.41, 115.64, 115.42. (3-Chlorophenyl) (phenyl) methanone (**2u**)²⁰



The representative procedure was followed using 1-benzyl-3-chlorobenzene (1u) (0.25 mmol) as substrate. Isolation by column chromatography (PE/EtOAc: 10/1) yielded 2u (35 mg, 65%) as colorless

oil. ¹H NMR (400 MHz, CDCl₃): δ ppm 7.79 (d, 2H, *J* = 8 Hz, ArH), 7.67 (d, 1H, *J* = 8 Hz, ArH), 7.61-7.55 (m, 3H, ArH), 7.52-7.48 (m, 2H, ArH), 7.42-7.40 (m, 1H, ArH). ¹³C NMR (101 MHz, CDCl₃) δ ppm 195.17, 139.30, 136.98, 134.58, 132.80, 132.32, 129.99, 129.88, 129.61, 128.45, 128.08. Bis (4-chlorophenyl) methanone (**2**v)²¹

The representative procedure was followed using bis (4-chlorophenyl) methane (1v) (0.25 mmol) as substrate. Isolation by column chromatography (PE/EtOAc: 10/1) yielded 2v (36 mg, 58%) as colorless oil. ¹H NMR (400 MHz, CDCl₃): δ ppm 7.73 (d, 2H, *J* = 8 Hz, ArH), 7.47 (d, 2H, *J* = 8 Hz, ArH). ¹³C NMR (101 MHz, CDCl₃) δ ppm 194.13, 139.14, 135.54, 131.29, 128.75. (4-(*tert*-Butyl)phenyl) (3-chlorophenyl) methanone (2w)



The representative procedure was followed using 1-(4-(tert-butyl)benzyl)-3-chlorobenzene (**1w**) (0.25 mmol) as substrate. Isolation by column chromatography (PE/EtOAc: 10/1) yielded **2w** (48 mg, 72%) as colorless oil. ¹H NMR (400 MHz, CDCl₃): δ ppm 7.77-7.74 (m, 3H, ArH), 7.67-7.66 (m, 1H, ArH), 7.56-7.50 (m, 3H, ArH), 7.43-7.39 (m, 1H, ArH), 1.37 (s, 9H, 3CH₃). ¹³C NMR (100 MHz, CDCl₃): δ ppm 194.9, 156.7, 139.6, 134.5, 134.2, 132.1, 130.1, 129.8, 129.6, 128.0, 125.5, 35.2, 31.1. HRMS (ESI-Q-TOF) exact mass calcd for C₁₇H₁₈ClO [M+H]⁺ 273.1046, found 273.1023. IR: 3064.8, 2962.5, 2904.1, 2868.1, 1658.0, 1604.5, 1566.1, 1463.3, 1314.3, 1276.0, 945.5, 849.7, 761.5, 709.2 cm⁻¹.

6.2 General procedure for the selective oxidation of amines with cercosporin

In a dried schlenk tube, amines **3** (0.25 mmol) and cercosporin (1 mol%) was added in 2.0 mL methanol. Next, a balloon was purged with oxygen and fixed on the top of the schlenk tube. The reaction mixture was stirred and irradiated by 15 W white CFL at room temperature under an atmospheric pressure oxygen atmosphere. The reaction progress was monitored by GC analysis. Photooxidation yields of **4** were calculated from GC measurements using internal standards.

6.3 General procedure for the selective oxidation of sulfides with cercosporin

In a dried schlenk tube, sulfides 6 (0.25 mmol) and cercosporin (1 mol%) was added in 2.0 mL

methanol. Next, a balloon was purged with oxygen and fixed on the top of the schlenk tube. The reaction mixture was stirred and irradiated by 15 W white CFL at room temperature under an atmospheric pressure oxygen atmosphere. When the reaction was finished, the reaction mixture was washed with brine. The aqueous phase was re-extracted with ethyl acetate. The combined organic extracts were dried over Na₂SO₄, concentrated in vacuum, and the resulting residue was purified by silica gel column chromatography to afford the desired product **7**.

(Benzylsulfinyl)benzene (7a)²²

The representative procedure was followed using benzyl(phenyl)sulfane (**6a**) (0.25 mmol) as substrate. Isolation by column chromatography (PE/EtOAc: 5/1) yielded **7a** (51 mg, 95%) as a white viscous solid. ¹H NMR (400 MHz, CDCl₃): δ ppm 7.47-7.36 (m, 5H, ArH), 7.29-7.21 (m, 3H, ArH), 6.98-6.96 (m, 2H, ArH), 4.00-3.97 (m, 2H, CH₂). ¹³C NMR (101 MHz, CDCl₃) δ ppm 142.82, 131.15, 130.35, 129.16, 128.85, 128.44, 128.23, 124.43, 63.61.

(Sulfinylbis(methylene))dibenzene (7b)²³

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The representative procedure was followed using dibenzylsulfane (**6b**) (0.25 mmol) as substrate. Isolation by column chromatography (PE/EtOAc: 5/1) yielded **7b** (51 mg, 90%) as a white viscous solid. ¹H NMR (400 MHz, CDCl₃): δ ppm 7.40-7.34 (m, 3H, ArH), 7.30-7.28 (m, 2H, ArH), 3.94-3.86 (m, 2H, CH₂). ¹³C NMR (101 MHz, CDCl₃) δ ppm 130.15, 128.96, 128.36, 57.34.

(Methylsulfinyl)benzene (7c)²⁴

7c

The representative procedure was followed using methyl(phenyl)sulfane (**6c**) (0.25 mmol) as substrate. Isolation by column chromatography (PE/EtOAc: 5/1) yielded **7c** (32 mg, 94%) as a white viscous solid. ¹H NMR (400 MHz, CDCl₃): δ ppm 7.67-7.64 (m, 2H, ArH), 7.56-7.48 (m, 3H, ArH), 2.73 (s, 3H, CH₃). ¹³C NMR (101 MHz, CDCl₃) δ ppm 145.69, 131.00, 129.32, 123.45, 43.92.

1-Chloro-4-(methylsulfinyl)benzene (7d)²⁵



The representative procedure was followed using (4-chlorophenyl)(methyl)sulfane (**6d**) (0.25 mmol) as substrate. Isolation by column chromatography (PE/EtOAc: 5/1) yielded **7d** (38 mg, 90%) as a white viscous solid. ¹H NMR (400 MHz, CDCl₃): δ ppm 7.60 (d, 2H, *J* = 8 Hz, ArH), 7.51 (d, 2H, *J* = 8 Hz, ArH), 2.73 (s, 3H, CH₃). ¹³C NMR (101 MHz, CDCl₃) δ ppm 144.26, 137.24, 129.64, 124.97, 44.03. 1-Bromo-4-(methylsulfinyl)benzene (**7e**)²⁶



The representative procedure was followed using (4-bromophenyl) (methyl)sulfane (**6e**) (0.25 mmol) as substrate. Isolation by column chromatography (PE/EtOAc: 5/1) yielded **7e** (47 mg, 87%) as a white viscous solid. ¹H NMR (400 MHz, CDCl₃): δ ppm 7.67 (d, 2H, *J* = 8 Ha, ArH), 7.53 (d, 2H, *J* = 8 Hz, ArH), 2.72 (s, 3H, CH₃). ¹³C NMR (101 MHz, CDCl₃) δ ppm 144.94, 132.56, 125.44, 125.14, 43.99. 1-(Methylsulfinyl)-4-nitrobenzene (**7f**)²⁷



The representative procedure was followed using methyl (4-nitrophenyl) sulfane (**6f**) (0.25 mmol) as substrate. Isolation by column chromatography (PE/EtOAc: 5/1) yielded **7f** (37 mg, 80%) as a white viscous solid. ¹H NMR (400 MHz, CDCl₃): δ ppm 8.40 (d, 2H, *J* = 8 Hz, ArH), 7.85 (d, 2H, *J* = 8 Hz, ArH), 2.80 (s, 3H, CH₃). ¹³C NMR (101 MHz, CDCl₃) δ ppm 153.29, 149.49, 124.68, 124.45, 43.85. Sulfinyldibenzene (**7g**)²⁸



The representative procedure was followed using diphenylsulfane (6g) (0.25 mmol) as substrate.

Isolation by column chromatography (PE/EtOAc: 5/1) yielded **7g** (45 mg, 90%) as a white viscous solid. ¹H NMR (400 MHz, CDCl₃): δ ppm 7.66-7.64 (m, 4H, ArH), 7.49-7.42 (m, 6H, ArH). ¹³C NMR (75 MHz, CDCl₃) δ ppm 145.64, 131.08, 129.35, 124.80.

1-Methoxy-4-(*p*-tolylsulfinyl)benzene (7h)²⁹



The representative procedure was followed using (4-methoxyphenyl) (*p*-tolyl)sulfane (**6h**) (0.25 mmol) as substrate. Isolation by column chromatography (PE/EtOAc: 5/1) yielded **7h** (53 mg, 87%) as a white viscous solid. ¹H NMR (400 MHz, CDCl₃): δ ppm 7.55 (d, 2H, *J* = 8 Hz, ArH), 7.49 (d, 2H, *J* = 8 Hz, ArH), 7.25 (d, 2H, *J* = 8 Hz, ArH), 6.95 (d, 2H, *J* = 8 Hz, ArH), 3.81 (s, 3H, OCH₃), 2.36 (s, 3H, CH₃). ¹³C NMR (101 MHz, CDCl₃) δ ppm 161.91, 142.68, 141.24, 137.00, 129.90, 127.05, 124.73, 114.77, 55.47, 21.35.

1-Chloro-4-(p-tolylsulfinyl)benzene (7i)³⁰



The representative procedure was followed using (4-chlorophenyl) (*p*-tolyl)sulfane (**6i**) (0.25 mmol) as substrate. Isolation by column chromatography (PE/EtOAc: 5/1) yielded **7i** (59 mg, 95%) as a white viscous solid. ¹H NMR (400 MHz, CDCl₃): δ ppm 7.56 (d, 2H, *J* = 8 Hz, ArH), 7.51 (d, 2H, *J* = 8 Hz, ArH), 7.42 (d, 2H, *J* = 8 Hz, ArH), 7.27 (d, 2H, *J* = 8 Hz, ArH), 2.37 (s, 3H, CH₃). ¹³C NMR (101 MHz, CDCl₃) δ ppm 144.44, 142.14, 142.02, 137.07, 130.16, 129.52, 126.01, 124.92, 21.41.

1,3-Dimethyl-5-(p-tolylsulfinyl)benzene (7j)³¹



The representative procedure was followed using (3,5-dimethyl phenyl) (*p*-tolyl)sulfane (**6j**) (0.25 mmol) as substrate. Isolation by column chromatography (PE/EtOAc: 5/1) yielded **7j** (54 mg, 90%) as a white viscous solid. ¹H NMR (400 MHz, CDCl₃): δ ppm 7.52 (d, 2H, *J* = 8 Hz, ArH), 7.26-7.23 (m, 4H, ArH), 7.03 (s, 1H, ArH), 2.36 (s, 3H, CH₃), 2.32 (s, 6H, 2CH₃). ¹³C NMR (101 MHz, CDCl₃) δ ppm 145.45,

142.65, 141.40, 139.19, 132.72, 129.96, 124.94, 122.17, 21.38, 21.28.

1-(tert-Butyl)-4-(phenylsulfinyl)benzene (7k)³²





The representative procedure was followed using (4-(*tert*-butyl) phenyl) (phenyl)sulfane (**6k**) (0.25 mmol) as substrate. Isolation by column chromatography (PE/EtOAc: 5/1) yielded **7k** (55 mg, 86%) as a white viscous solid. ¹H NMR (400 MHz, CDCl₃): δ ppm 7.66-7.64 (m, 2H, ArH), 7.56 (d, 2H, *J* = 8 Hz, ArH), 7.48-7.41 (m, 5H, ArH), 1.30 (s, 9H, 3CH₃). ¹³C NMR (101 MHz, CDCl₃) δ ppm 154.64, 145.70, 142.41, 130.84, 129.22, 126.37, 124.80, 124.68, 34.91, 31.16.

4,4'-Sulfinylbis(bromobenzene) (71)33



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The representative procedure was followed using bis(4-bromophenyl) sulfane (**6**I) (0.25 mmol) as substrate. Isolation by column chromatography (PE/EtOAc: 5/1) yielded **7**I (83 mg, 93%) as a white viscous solid. ¹H NMR (400 MHz, CDCl₃): δ ppm 7.61 (d, 2H, *J* = 8 Hz, ArH), 7.50 (d, 2H, *J* = 8 Hz, ArH). ¹³C NMR (101 MHz, CDCl₃) δ ppm 144.40, 132.69, 126.16, 125.92.

1-Bromo-4-(phenylsulfinyl)benzene (7m)³²



7m

The representative procedure was followed using (4-bromophenyl) (phenyl) sulfane (**6m**) (0.25 mmol) as substrate. Isolation by column chromatography (PE/EtOAc: 5/1) yielded **7m** (63 mg, 92%) as a white viscous solid. ¹H NMR (400 MHz, CDCl₃): δ ppm 7.64-7.62 (m, 2H, ArH), 7.59 (d, 2H, J = 8 Hz, ArH), 7.51 (d, 2H, J = 8 Hz, ArH), 7.48-7.46 (m, 3H, ArH). ¹³C NMR (101 MHz, CDCl₃) δ ppm 145.23, 144.90, 132.46, 131.32, 129.45, 126.18, 125.46, 124.63.

7. Mechanism Research

7.1 Time profile of the photooxidation of benzylic C-H bond with and without visible light



In a dried schlenk tube, 4-ethylbenzene (1a) (0.25 mmol), potassium bromide (0.2 equiv) and cercosporin (2 mol%) was added in 2.0 mL methanol. Next, a balloon was purged with oxygen and fixed on the top of the schlenk tube. Then, the reaction mixtures were placed under 23 W CFL at given time intervals. The yield was detected by GC using diphenyl as internal standard.

Light ON/OFF	Time (h)	Yield of 2a
0	0	0
2 hour on	2	20
1 hour off	3	22
2 hour on	5	45
1 hour off	6	46
2 hour on	8	68
1 hour off	9	70
2 hour on	11	83







7.2 Time profile of the photooxidation of amine with and without visible light



In a dried schlenk tube, phenylmethanamine (**3a**) (0.2 mmol) and cercosporin (1 mol%) was added in 2.0 mL CH₃OH. Next, a balloon was purged with oxygen and fixed on the top of the schlenk tube. Then, the reaction mixtures were placed under 15 W CFL at given time intervals. The yield was detected by GC using diphenyl as internal standard.

Light ON/OFF	Time (h)	Yield of 4a
0	0	0
2 hour on	2	26
1 hour off	3	29
2 hour on	5	52
1 hour off	6	55
2 hour on	8	72
1 hour off	9	75
2 hour on	11	95

Table S5. On/off experiment for the photooxidation of amine



Fig. S4 Time profile of the photooxidation of amine with and without visible light.

7.3 Time profile of the photooxidation of sulfide with and without visible light



In a dried schlenk tube, diphenylsulfane (6g) (0.25 mmol) and cercosporin (1 mol%) was added in 2.0 mL CH₃OH. Next, a balloon was purged with oxygen and fixed on the top of the schlenk tube. Then,

the reaction mixtures were placed under 15 W CFL at given time intervals. The yield was detected by GC using diphenyl as internal standard.



Table S6. On/off experiment for the photooxidation of sulfide.

Fig. S5 Time profile of the photooxidation of sulfide with and without visible light.

7.4 Alkane radical capturing experiment in the photooxidation of benzylic C-H bond



Fig. S6 Alkane radical capturing experiment in the photooxidation of benzylic C-H bond

8. References

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9. ¹H, ¹³C and IR Spectra.























S34





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S48



S49



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S73











S78























S89







S92









S96

