Electronic Supporting Information (ESI)

Fully bio-derived CO₂ polymers for non-isocyanate based polyurethane synthesis

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1. Characterization of PBDs

1.1 Summarized properties of PBD1 - PBD4

	м		double bond units [%] ^b				double bond units [%] ^c		double bonds	т	Ŧ
polymer	[kg/mol]ª	Ъ	1,	,4-	1,:	2-	1,4-	1,2-	[mmol/g] ^d	(°C)°	°C) ^f
			cis	trans	cyclic	vinyl	cis/-trans	cyclic/-vinyl			
PBD1	1.14	2.3 3	40	0.8	30.1	29.1	40.7	59.3	11.7	- 56	249
PBD2	2.63	2.5 8	44.4	36.6	1.1	18.9	81.4	18.6	16.0	- 94	347
PBD3	11.0	1.1 2	42.1	44.0	0.4	13.5	88.1	11.9	16.7	- 94	351
PBD4	39.4	1.1 2	45.8	43.9	0.2	10.2	91.6	8.40	14.2	- 99	365

Table S1. Detailed properties of PBD1 - PBD4.

^aMeasured by GPC (THF, RT) using polystyrene standards. ^bDetermined by ¹H NMR analysis using

 $X_{double \ bond \ type}(\%) = \frac{A_{double \ bond \ type}}{A_{total}} \cdot 100$ normalized proton areas (A): NMR analysis using normalized carbon areas (A): $X_{1,2-cylic/-vinyl}(\%) = \frac{A_{1,2-cyclic/-vinyl, = CH_2}}{A_{total -} A_{styrene}} \cdot 100.$ and $X_{1,2-cylic/-vinyl}(\%) = \frac{A_{1,2-cyclic/-vinyl, = CH_2}}{A_{total -} A_{styrene}} \cdot 100.$

Supporting Information Section 3). Determined by DSC; data are taken from the second heating. Decomposition temperature (T_d) determined by TGA at 1% weight loss.

1.2 Spectroscopic analysis



Figure S1. ¹H NMR spectrum of PBD1 in CDCl₃ (Table S1).



¹H NMR (CDCl₃, 400 MHz): *δ* (ppm) = 7.50-6.85 (m, 5H, 1-CH),6.1-5.69 (m, 1H, 17-CH), 5.69-5.49 (m, 1H, 9-CH), 5.49-5.23 (m, 2H, 4-CH, 5-CH), 5.23-4.50 (m, 2H, 10-CH₂, 18-CH₂), 3.00-2.57 (m, 2H, 2x2-CH), 2.57-0.48 (m, 16H, 2x3-CH, 2x6-CH, 2x7-CH, 8-CH, 2x11-CH, 12-CH, 13-CH, 2x14-CH, 15-CH, 2x16-CH).





¹³**C NMR** (CDCl₃, 125 MHz): δ (ppm) = 145.0-141.8 (1x1-C (styrene end group), 9-C, 17-C), 132.5-123.2 (5x1-C (styrene end group), 4-C, 5-C), 115.7-111.5 (10-C, 18-C), 46.7-6.05 (2-C, 3-C, 6-C, 7-C, 8-C, 11-C, 12-C, 13-C, 14-C, 15-C, 16-C).



re S3. IR spectrum of PBD1 (Table S1).

1.2.2 PBD2

Please note the peak assignments in Figure S1 and S2. The signal for 1, 4-cis and 1,4-trans double bonds splits into the signals at δ (ppm) = 5.42 (m, 2H (trans)) and 5.38 (m, 2H, (cis)).



Figure S5. ¹³C NMR spectrum of PBD2 in CDCl₃ (Table S1).



re S6. IR spectrum of PBD2 (Table S1).

1.2.3 PBD3



Figure S8. ¹³C NMR spectrum of PBD3 in CDCl₃ (Table S1).



Figure S9. IR spectrum of PBD3 (Table S1).

1.2.4 PBD4







Figure S12. IR spectrum of PBD4 (Table S1).



1.3 Thermal analysis

Figure S13. Thermogravimetric analysis of PBD1 (Table S1).



Figure S14. Differential Scanning Calorimetry trace of PBD1 (Table S1).

1.3.2 PBD2



Figure S15. Thermogravimetric analysis of PBD2 (Table S1).



Figure S16. Differential Scanning Calorimetry trace of PBD2 (Table S1).



1.3.3 PBD3

Figure S17. Thermogravimetric analysis of PBD3 (Table S1).



Figure S18. Differential Scanning Calorimetry trace of PBD3 (Table S1).





Figure S19. Thermogravimetric analysis of PBD4 (Table S1).



Figure S20. Differential Scanning Calorimetry trace of PBD4 (Table S1).

2. Characterization of PE-PBDs

2.1 Summarized properties of PE-PBD1 - PE-PBD4

Table S2. Detailed properties of PE-PBD1 - PE-PBD4.

		Đª		epoxic	le units [%				
polymer	M _n [kg/mol]ª			1, 4	_ b	1,2- <i>^b</i>	EP-number ^a	T _g (°C)⁰	T _d (°C) ^f
	[Kg/mol]		total	trans	cis	cyclic/-vinyl	[mmon/9]		
PE-PBD1	1.10	2.43	13.9 ^b	6.50	5.95	1.47	1.62	- 45	259
			10.9°						
PE-PBD2	2.64	2.56	o.20° 7.20°	3.85	4.02	0.39	1.33	- 87	322
PE-PBD3	11.5	4.40	11.8 ^b	57 4	5.05	0.07	1.96	- 83	362
		1.16	11.2°	5.74	5.95				
PE-PBD4	39.7	1.13	10.7 ^b	5.13	5.32	0.20	1.51	- 88	375

^aMeasured by GPC (THF, RT) using polystyrene standards. ^bDetermined by ¹H NMR analysis using normalised proton areas for

$$X_{1,2-cyclic/-vinyl\,epoxide} \text{ or } X_{1,4-cis\,epoxide}(\%) = \frac{A_{epoxide\,type}}{A_{total} - A_{methylene\,group, styrene}} \cdot 100$$
epoxide and double bonds:

$$X_{1,4-trans\,epoxide}(\%) = \frac{A_{epoxide\,type} - A_{methylene\,group, styrene}}{A_{total} - A_{methylene\,group, styrene}} \cdot 100$$
Chetermined by ¹³C igated NMR analysis using normalized

using

 $X_{epoxide}(\%) = \frac{A_{epoxide}}{A_{total-}A_{styrene}} \cdot 100$. "Measured by epoxide titration (see carbon areas of epoxides and double bonds: Supporting Information Section 3). "Determined by DSC; data are taken from the second heating. "Decomposition temperature (T_d) determined by TGA at 1% weight loss.

2.2 Spectroscopic analysis

2.2.1 PE-PBD1



¹H NMR (CDCl₃, 400 MHz): δ (ppm) = 7.50-6.85 (m, 5H, 1-CH),6.0-5.69 (m, 1H, 17-CH), 5.69-5.49 (m, 1H, 9-CH), 5.49-5.22 (m, 2H, 4-CH, 5-CH), 5.23-4.50 (m, 2H, 10-CH₂, 18-CH₂), 3.25-3.00 (m, 2H, 9'-CH, 17'-CH), 3.00-2.80 (m, cis epoxide, 2H, 4'-CH, 5'-CH), 2.80-2.57 (m, trans epoxide, 4H, 2x2-CH, 4'-CH, 5'-CH), 2.57-0.48 (m, 36H, 2x3-CH, 2x3'-CH, 2x6-CH, 2x6'-CH, 2x7'-CH, 8-CH, 8'-CH, 2x10'-CH, 2x11-CH, 2x11'-CH, 12-CH, 12'-CH, 13-CH, 13'-CH, 2x14-CH, 2x14'-CH, 15-CH, 15'-CH, 2x16-CH, 2x16'-CH, 2x18'-CH).

Signals from side reactions e. g. epoxide ring opening (-OH formation) and cross-linking (C-O-C formation) typically between δ (ppm) = 3 - 6 ppm were not observed for all samples.



Figure S22. ¹³C NMR spectrum of PE-PBD1 in CDCl₃ (Table S2).



¹³**C NMR** (CDCl₃, 125 MHz): δ (ppm) = 145.0-141.8 (6x1-C (styrene), 9-C, 17-C), 132.5-123.2 (4-H, 5-C), 115.7-111.5 (10-C, 18-C), 66.0-63.5 (9'-C, 17'-C), 59.5-51.0 (4'-C, 5'-C), 46.7-6.05 (10'-C, 18'-C, 2-C, 3-C, 3'-C, 6-C, 6'-C, 7-C, 7'-C, 8-C, 8'-C, 11-C, 11'-C, 12-C, 12'-C, 13-C, 13'-C, 14-C, 14'-C, 15-C, 15'-C, 16-C, 16'-C).



Figure S23. IR spectrum of PE-PBD1 (Table S2).

Signals from side reactions e. g. epoxide ring opening (-OH formation) and cross-linking (C-O-C formation) typically between $3600 - 3200 \text{ cm}^{-1}$ and $1300 - 1000 \text{ cm}^{-1}$ were not observed for all samples.

2.2.2 PE-PBD2

Please note the peak assignments in Figure S21 and S22. The signal for 1, 4-cis and 1, 4-trans double bonds splits into the signals at δ (ppm) = 5.45 (m, 2H (trans)) and 5.42 (m, 2H, (cis)).







ep. Ultra PM 4 2.0 Figure S26. IR spectrum of PE-PBD2 (Table S2).

2.2.3 PE-PBD3

Please note the peak assignments in Figure S21 and S22.The signal for 1, 4-cis and 1, 4-trans double bonds splits into the signals at δ (ppm) = 5.36 (m, 2H (trans)) and 5.32 (m, 2H, (cis)).



Figure S28. ^{13}C NMR spectrum of PE-PBD3 in CDCl_3 (Table S2). S22



ep. UltraN4 5000 2 128.0

Figure S29. IR spectrum of PE-PBD3 (Table S2).

2.2.4 PE-PBD4

Please note the peak assignments in Figure S21 and S22.The signal for 1, 4-cis and 1, 4-trans double bonds splits into the signals at δ (ppm) = 5.40 (m, 2H (trans)) and 5.36 (m, 2H, (cis)).



S24



ep. LBR 300 1 128.0

Figure S32. IR spectrum of PE-PBD4 (Table S2).

2.3Thermal analysis

2.3.1 PE-PBD1







Figure S34. Differential Scanning Calorimetry trace of PE-PBD1 (Table S2).

2.3.2 PE-PBD2



Figure S35. Thermogravimetric analysis of PE-PBD2 (Table S2).



Figure S36. Differential Scanning Calorimetry trace of PE-PBD2 (Table S2).

2.3.3 PE-PBD3



Figure S37. Thermogravimetric analysis of PE-PBD3 (Table S2).



Figure S38. Differential Scanning Calorimetry trace of PE-PBD3 (Table S2).

2.3.4 PE-PBD4



Figure S39. Thermogravimetric analysis of PE-PBD4 (Table S2).



Figure S40. Differential Scanning Calorimetry trace of PE-PBD4 (Table S2).

3. Determination of PBDs double bond contents

EP-numbers of several partly epoxidized PBDs were determined by the titration technique described in the Experimental Section. The relative amounts of converted double bonds were calculated by ¹H NMR analysis. Both results were applied to the graph shown in figure S41.



Figure S41. EP-numbers of partly epoxidized PBDs and relative amount of epoxidized double bonds applied to a graph and linearly fitted.

Linear fitting and extrapolation leads to the EP-values expectable at 100 % epoxide conversion. This is equated with the total amount of double bonds in the polymers, respectively. The results are summarized in table S3. All samples differ from the theoretical value of 18.5 mmol/g.

Table S3. Summary of data derived by linear fit and extrapolation.

PBD	slope, a [<u><i>mol</i>/100</u> <u></u> <i>mol</i> %	coefficient of determination, R ²	EP-value at 100% epoxide conversion [mol/100g]
PBD1	0.0117	0.9990	1.17
PBD2	0.0160	0.9998	1.60
PBD3	0.0167	0.9967	1.67
PBD4	0.0142	0.9962	1.42

4. Characterization of PC-PBDs

4.1 Spectroscopic analysis



Figure S42. ¹H NMR spectrum of PC-PBD1 in CDCl₃ (Table 2).



¹H NMR (CDCl₃, 400 MHz): *δ* (ppm) = 7.39-6.85 (m, 5H, 1-CH),6.25-5.68(m, 1H, 17-CH), 5.68-5.49 (m, 1H, 9-CH), 5.49-5.23 (m, 2H, 4-CH, 5-CH), 5.23-4.73 (m, 2H, 10-CH₂, 18-CH₂), 4.73-3.85 (m, 8H, 4"-H, 5"-H, 9"-H, 10"-H, 17"-H, 18"-H), 3.25-3.00 (m, 2H, 9'-CH, 17'-CH), 3.00-2.80 (m, cis-epoxide, 2H, 4'-CH, 5'-CH), 2.80-2.57 (m, trans-epoxide, 4H, 2x2-CH, 4'-CH, 5'-CH), 2.57-0.48 (m, 52H, 2x3-CH, 2x3'-CH, 2x3"-CH, 2x6-CH, 2x6'-CH, 2x6"-CH, 2x7-CH, 2x7'-CH, 2x7"-CH 8-CH, 8'-CH, 8"-CH, 2x10'-CH, 2x11-CH, 2x11'-CH, 2x11"-CH 12-CH, 12'-CH, 12"-CH, 13-CH, 13'-CH, 13"-CH, 2x14-CH, 2x14'-CH, 2x14"-CH, 15-CH, 15'-CH), 15"-CH, 2x16-CH, 2x16'-CH, 2x16'-CH, 2x16"-CH).





¹³**C NMR** (CDCl₃, 125 MHz): δ (ppm) = 145.0-141.8 (6x1-C (styrene), 9-C, 17-C), 132.5-123.2 (4-H, 5-C), 115.7-111.5 (10-C, 18-C), 83.0-77.7 (4"-C, 5"-C, 9"-C, 10"-C, 17"-C, 18"-C), 66.0-63.5 (9'-C, 17'-C), 59.5-51.0 (4'-C, 5'-C), 46.7-6.05 (2-C, 3-C, 3'-C, 3"-C, 6-C, 6'-C, 6"-C, 7-C, 7'-C, 7"-C, 8-C, 8'-C, 8"-C, 11-C, 11'-C, 11"-C, 12-C, 12'-C, 12"-C, 13-C, 13'-C, 13"-C, 14-C, 14'-C, 14"-C, 15'-C, 15'-C, 15"-C, 16'-C, 16'-C).



carb. Ultra AL 4.0 Figure S44. IR spectrum of PC-PBD1 (Table 2).

4.1.2 PC-PBD2



Figure S46. ^{13}C NMR spectrum of PC-PBD2 in CDCl3 (Table 2). S34



carb. Ultra PM4 2.0

Figure S47. IR spectrum of PC-PBD2 (Table 2).

4.1.3 PC-PBD3

Please note the peak assignments in Figure S42 and S43.





carb. Ultra N4 5000 2.0

Figure S50. IR spectrum of PC-PBD3 (Table 2).

4.1.4 PC-PBD4

Please note the peak assignments in Figure S42 and S43.



Figure S52.¹³C NMR spectrum of PC-PBD4 in CDCl₃ (Table 2).



carb. LBR 300.0 Figure S53. IR spectrum of PC-PBD4 (Table 2).

4.2 Thermal analysis

4.2.1 PC-PBD1



Figure S54. Thermogravimetric analysis of **PC-PBD1** (Table 2).



Figure S55. Differential Scanning Calorimetry trace of PC-PBD1 (Table 2).

4.2.2 PC-PBD2



Figure S56. Thermogravimetric analysis of PC-PBD2 (Table 2).



Figure S57. Differential Scanning Calorimetry trace of PC-PBD2 (Table 2).

4.2.3 PC-PBD3



Figure S58. Thermogravimetric analysis of PC-PBD3 (Table 2).



Figure S59. Differential Scanning Calorimetry trace of PC-PBD3 (Table 2).

4.2.4 PC-PBD4



Figure S60. Thermogravimetric analysis of PC-PBD4 (Table 2).



Figure S61. Differential Scanning Calorimetry trace of PC-PBD4 (Table 2).

5. Selectivities towards the conversion of 1,2-cyclic/vinyl, 1,4-*cis* and 1,4*trans* epoxides depending on reaction conditions demonstrated by epoxide residues in the partyl carboxylated **PE-PBDs**



Figure S62. Epoxide ratios in partly carboxylated **PE-PBD1** after the application of different reaction conditions, solvents and reaction times.



ure S63. Epoxide ratios in partly carboxylated **PE-PBD2** after the application of different reaction conditions, solvents and reaction times.



■ 1,4-trans epoxides ■ 1,4-cis epoxides ■ 1,2-cyclic/vinyl epoxides

Figure S64. Epoxide ratios in partly carboxylated **PE-PBD3** after the application of different reaction conditions, solvents and reaction times.



Figure S65. Epoxide ratios in partly carboxylated **PE-PBD4** after the application of different reaction conditions, solvents and reaction times.

6. GPC analysis



Figure S66. GPC analysis of the sequential transformation a) from PBD1 to PC-PBD1, b) PBD2 to PC-PBD2, c) PBD3 to PC-PBD3 and d) PBD4 to PC-PBD4.

7. IR Characterization NIPU reactions



Figure S67. IR-spectra of PC-PBD1 cured with PDA (1/1) at 70 °C and 130°C for 16h.



Figure S68. IR-spectra of PC-PBD1 cured with ODA (1/1) at 70 °C and 130°C for 16h.



Figure S69. IR-spectra of PC-PBD2 cured with PDA (1/1) at 70 °C and 130°C for 16h.



Figure S70. IR-spectra of PC-PBD2 cured with ODA (1/1) at 70 °C and 130°C for 16h.