An Efficient Way for the N-Formylation of Amines by Inorganic-ligand Supported Iron Catalysis

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I. General information

The catalyst was prepared according to published literature methods.^[1] All reagents were purchased from Sigma-Aldrich and Adamas-beta, which were used without further purification. FT-IR spectra were recorded on a Thermo Fisher Nicolet 6700. XRD were explored on D/max 2200 PC of Janpan. GC analyses were performed on Shimadzu GC-2014 with a flame ionization detector equipped with an Rtx-1 capillary column (internal diameter = 0.25 mm, length = 30 m) or a Stabil wax capillary column (internal diameter = 0.25 mm, length = 30 m). GC mass spectra were recorded on Shimadzu GCMS-QP2010 with RTX-5MS column (0.25 mm× 30 m). ¹H and ¹³C Nuclear Magnetic Resonance (NMR) spectra were recorded on Bruker AVANCE III 500 MHz (500 MHz for proton, 125MHz for carbon) spectrometer with tetramethylsilane as the internal reference using CDCl₃ and DMSO as solvent in all cases, and chemical shifts were reported in parts per million (ppm, δ). Column chromatography was performed using 200-300 mesh silica gel.

II. Preparation and Characterizations of catalyst

 $[NH_4]_3[FeMo_6O_{18}(OH)_6]$ was prepared according to the published literature methods and what we previously reported^[1,2]. Firstly, $(NH_4)_6Mo_7O_{24} \times 4H_2O$ (5.0 g, 4 .0 mmol) was dissolved in water (80 ml) under stirring in an oil bath at 100 °C. Then, an aqueous solution of Fe₂(SO₄)₃ (2.3 g, 5.75 mmol) dissolved in 20 ml of water was added drop by drop to the above solution. The pH of the solution needs to be controlled at around 4.0 to 6.5 in this process strictly. After the dropwise addition is completed, the mixed solution is further stirred at a constant temperature of 100 °C for 2 hour. Following by, the solution is filtered immediately. The obtained solution was cooling at room temperature for 12 hours and precipitated the white crystals ^[1]. After recrystallized, filtered and vacuum dried, the white crystals (5.1 g) was deposited and collected. IR: 3174.36 (vasNH, m), 1635.68 (δ OH m), 1400.75 (δ NH, s), 947.41 (v Mo=O, vs), 891.89 (v Mo=O, vs), 650.93 (v Mo-O-Mo, vs), 572.87 (v M-O-Mo, w) cm⁻¹.



Figure S1. [NH₄]₃[FeMo₆O₁₈(OH)₆]

III. FT-IR spectra of catalyst 1



Figure S2. FT-IR spectra of catalyst 1

IV. XRD spectra of catalyst 1



Figure S3. XRD spectra of catalyst 1

V. ESI-MS spectra of catalyst 1



Figure S4. ESI-MS of (NH₄)₃[FeMo₆O₁₈(OH)₆].



Figure S5. Zoom the area of ESI-MS of $(NH_4)_3$ [FeMo₆O₁₈(OH)₆], (m/z = 1010-1500, $\{NH_4H[FeMo_6O_{24}H_6]\}^{1-} = 1043.34$ g/mol).

VI. General procedure for N-Formylation of Amines with Formic acid

The Cat.1 (1.0 mol%), amines (1.0 mmol), formic acid (2.0 ml), Na₂SO₃ (0.05 eq.) stirring at a reaction tube at 80 °C for 2 h (diamines for 12 h). Afterwards, a small amount of ethyl acetate was added into the reaction mixture and the solution was quickly filtered. The filtered solid was washed, dried and then recycled. Reaction mixture was analyzed by GC-MS analysis. Finally, the solvent was removed in vacuo, and the corresponding Formamides was purified by washing through base-washed

silica gel column. (Petrol: EtOAc = 1:2).

VII. Recycling experiments of catalyst 1

The POM catalyst was filtered and dried after the direct coupling experiment. The recovered catalyst was characterized by FTIR and X-RD. The infrared image contains a new catalyst and the sixth catalyst.



Figure S6. The catalyst recovery



Figure S7. Recycling experiments for the Cat. 1



Figure S8. FT-IR spectra of the Cat. 1 before and after reaction



Figure S9. XRD of Cat. 1 before and after reaction

VIII. Optimization of reaction conditions

NH ₂	0 + II _	Cat. 1 (1.0 mol%) Additive (0.05 eq.)	N O	
	НОН	80 °C, 2 h		
Entry	Catalyst [mol%]	Additive [eq.]	Yield ^[j] [%]	
1	1 (1.0)	NaBr	24	
2	1 (1.0)	NaI	45	
3	1 (1.0)	Na ₂ CO ₃	77	
4	1 (1.0)	KCl	32	
5	1 (1.0)	K_2SO_4	53	
6	1 (1.0)	KBr	45	
7	1 (1.0)	KI	64	
8	1 (1.0)	NaHCO ₃	72	
9	1 (1.0)	Na ₂ SO ₃	>99	
10 ^b	1 (1.0)	Na_2SO_3	86	
11 ^c	1 (1.0)	Na_2SO_3	89	
12 ^d	1 (0.1)	Na_2SO_3	63	
13 ^e	1 (0.5)	Na ₂ SO ₃	76	
14 ^f	1 (1.5)	Na ₂ SO ₃	93	

Table S1: Effect of the amount of catalyst, additive on N-Formylation of Amines

^a Reaction conditions: Cat. **1** (1.0 mol%), benzylamine (1.0 mmol), formic acid (2.0 ml), 80 °C, 2 h. ^b Na₂SO₃ (0.01 eq.). ^cNa₂SO₃ (0.1 eq.). ^d Cat. **1** (0.1 mol%). ^e Cat. **1** (0.5 mol%). ^fCat. **1** (1.5 mol%). ^g Substrate conversion and yield were determined by GC-MS (internal standard is toluene) analysis.

	NH ₂	Cat. 1 (1.0 mc O Na ₂ SO ₃ (0.05	bl%) 5 eq.)	^N∕∽o
	Ŧ	H OH T, h, Solver	nt	н
Entry	T [°C]	Solvent [2.0 mL]	Time [h]	Yield ^[b] [%]
1	50	none	4	43
2	60	none	4	66
3	70	none	4	78
4	75	none	4	83
5	80	none	4	92
6	85	none	4	90
7	90	none	4	74
8	80	none	2	>99
9	80	none	6	94
10 ^b	80	diethyl ether	2	75
11°	80	MeCN	2	68
12 ^d	80	Toluene	2	81
13 ^e	80	DMF	2	83

^aReaction conditions: Cat. **1** (1.0 mol%), benzylamine (1.0 mmol), formic acid (2.0 ml), Na₂SO₃ (0.05 eq.) stirring at a reaction tube at 80 °C for 2 h. ^{b-e} Cat. **1** (1.0 mol%), benzylamine (1.0 mmol), formic acid (2.0 mmol), Na₂SO₃ (0.05 eq.) stirring at a reaction tube at 80 °C for 2 h. ^fSubstrate conversion and yield were determined by GC-MS (internal standard is toluene) analysis.

Table S3: Optimization of N-formylation on diamine

	NH ₂ O	Cat. 1 (1.0 mol% Na ₂ SO ₃ (0.05 e	o) q.)	N O
	NH ₂ H OH	T, h		N O H
Entry	T [°C]	HCOOH (ml)	T [h]	Yield ^[b] [%]
1	60	1.0	12	38
2	70	1.0	12	45
3	75	1.0	12	51
4	80	1.0	12	63
5	85	1.0	12	60
6	90	1.0	12	56
7	80	1.0	12	67
8	80	1.5	12	78
9	80	1.8	12	80
10	80	2.0	12	82
11	80	2.5	12	60
12	80	3.0	12	75
13	80	2.0	6	59
15	80	2.0	18	78

^a Reaction conditions: Cat. **1** (1.0 mol%), phenylenediamine (1.0 mmol), formic acid (2.0 mL), Na₂SO₃ (0.05 eq.) stirring at a reaction tube at 80 °C for 12 h. ^b Yields were determined by ¹H-NMR, values in parentheses are the isolated yield.

IX. References

- K, Nomiya; T, Takahashi; T, Shirai; M, Miwa. Andersontype heteropolyanions of molybdenum(VI) and tungsten(VI) *Polyhedron.* 1987, 6, 213-218.
- [2] H, Yu.; Q, Zhao.; Z, Wei.; Z, Wu.; Q, L.; S, Han.; Y, Wei. Ironcatalyzed oxidative functionalization of C(sp³)–H bonds under bromide-synergized mild conditions. *Chem. Commun.* 2019, 55, 7840.
- C, C, Chong; R, Kinjo. Hydrophosphination of CO₂ and Subsequent Formate Transfer in the 1,3,2-Diazaphospholene-Catalyzed N-Formylation of Amines. *Angew. Chem. Int. Ed.* 2015, 54, 12116.
- [4] B, Kang; S, H, Hong. Cheminform. 2015, 357, 834.
- [5] S, Chakraborty; U, Gellrich; Y, Diskin-Posner. Manganese-Catalyzed N-Formylation of Amines by Methanol Liberating H₂: A Catalytic and Mechanistic Study. *Angew. Chem. Int. Ed. Engl.* 2017, *56*, 4229.

 [6] O, Jacquet; C, D, N, Gomes; M, Ephritikhine; T, Cantat. Recycling of Carbon and Silicon Wastes: Room Temperature Formylation of N-H Bonds Using Carbon Dioxide and Polymethylhydrosiloxane. J. Am. Chem. Soc. 2012, 134, 2934-2937.

X. NMR data of products

N O H

N-benzylformamide(2)^[3-6]: ¹H NMR (500 MHz, CDCl₃) δ 8.10 (s, 1H), 7.37 – 7.27 (m, 3H), 7.24 (t, *J* = 7.3 Hz, 2H), 6.99 (s, 1H), 4.37 (s, 2H). ³C NMR (125 MHz, CDCl₃) δ 160.81 (s), 137.74 (s), 129.35 (s), 128.08 (s), 42.37 (s).



N-(4-methylbenzyl)formamide(3) ^[3-6]: ¹H NMR (500 MHz, CDCl₃) δ 8.24 (s, 1H), 7.18 (dd, *J* = 15.2, 7.5 Hz, 4H), 6.09 (s, 1H), 4.44 (s, 2H), 2.35 (s, 3H). ¹³C NMR (125 MHz, CDCl₃) δ 161.06 (s), 137.43 (s), 134.59 (s), 129.45 (s), 127.81 (s), 126.96 (s), 41.95 (s), 21.10 (s).



N-(2-methylbenzyl)formamide(4) ^[3-6]: ¹H NMR (500 MHz, CDCl₃) δ 8.17 (s, 1H), 7.35 (s, 4H), 6.02 (s, 1H), 5.23 (s, 2H), 1.53 (s, 3H). ¹³C NMR (125 MHz, CDCl₃) δ 160.38 (s), 142.51 (s), 128.79 (s), 127.60 (s), 126.17 (s), 47.65 (s), 21.74 (s).



N-(4-methoxybenzyl)formamide(5) ^[3-6]: ¹H NMR (500 MHz, DMSO) δ 8.45 (s, 1H), 8.22 (s, 1H), 7.27 (d, J = 8.4 Hz, 2H), 6.92 (d, J = 8.5 Hz, 2H), 4.34 (s, 2H), 3.75 (s, 3H). ¹³C NMR (125 MHz, DMSO) δ 161.51 (s), 158.88 (s), 131.30 (s), 129.10 (s), 114.10 (s), 55.24 (s), 40.82(s).



N-(4-isopropylbenzyl)formamide(6) ^[3-6]: ¹H NMR (500 MHz, CDCl₃) δ 8.17 (s, 1H), 7.20 (d, J = 5.8 Hz, 4H), 6.52 (s, 1H), 4.40 (s, 2H), 2.90 (s, 1H), 1.23 (d, J = 6.8 Hz, 6H). ¹³C NMR (125 MHz, CDCl₃) δ 161.40 (s), 148.39 (s), 134.95 (s), 127.86 (s), 127.33 (d, J = 129.7 Hz), 41.91 (s), 33.82 (s), 23.85 (s).



N-(benzo[d][1,3]dioxol-5-ylmethyl)formamide(7) ^[3-6]: ¹H NMR (500 MHz, DMSO) δ 8.44 (s, 1H), 8.12 (s, 1H), 6.85 (d, *J* = 8.2 Hz, 2H), 6.75 (d, *J* = 7.9 Hz, 1H), 5.98 (s, 2H), 4.22 (d, *J* = 6.0 Hz, 2H). ¹³C NMR (125 MHz, CDCl₃) δ 161.38 (s), 147.96 (s), 147.09 (s), 131.39 (s), 121.12 (s), 108.39 (d, *J* = 7.1 Hz), 101.13 (s), 42.03 (s).



N-benzhydrylformamide(8) ^[3-6]: ¹H NMR (500 MHz, CDCl₃) δ 8.29 (s, 1H), 7.37 – 7.34 (m, 4H), 7.31 (d, J = 7.1 Hz, 2H), 7.26 (d, J = 7.1 Hz, 4H), 6.39 (s, 1H), 6.35 (s, 1H). ¹³C NMR (125 MHz, CDCl₃) δ 160.22 (s), 140.93 (s), 128.97 (s), 128.76 (s), 128.03 (s), 127.66 (s), 127.32 (d, J = 18.7 Hz),



N-(4-fluorobenzyl)formamide(9) ^[3-6]: ¹H NMR (500 MHz, CDCl₃) δ 8.44 (s, 1H), 8.12 (s, 1H), 6.85 (d, J = 8.2 Hz, 2H), 6.75 (d, J = 7.9 Hz, 1H), 5.98 (s, 2H), 4.22 (d, J = 6.0 Hz, 2H). ¹³C NMR (125 MHz, CDCl₃) δ 161.04 (s), 129.49 (d, J = 8.1 Hz), 115.71 (s), 115.54 (s), 41.46 (s).



N-(4-chlorobenzyl)formamide(10) ^[3-6]: ¹H NMR (500 MHz, CDCl₃) δ 8.29 (s, 1H), 7.33 (d, J = 8.4 Hz, 2H), 7.25 (d, J = 8.3 Hz, 2H), 5.96 (s, 1H), 4.47 (d, J = 6.0 Hz, 2H). ¹³C NMR (125 MHz, CDCl₃) δ 160.96 (s), 136.14 (s), 133.58 (s), 129.14 (s), 128.93 (s), 128.31 (s), 41.50 (s).



N-(4-bromobenzyl)formamide(11) ^[3-6]: ¹H NMR (500 MHz, CDCl₃) δ 8.44 (s, 1H), 8.12 (s, 1H), 6.85 (d, J = 8.2 Hz, 2H), 6.75 (d, J = 7.9 Hz, 1H), 5.98 (s, 2H), 4.22 (d, J = 6.0 Hz, 2H).¹³C NMR (125 MHz, CDCl₃) δ 161.03 (s), 136.65 (s), 131.90 (s), 129.49 (s), 121.63 (s), 41.56 (s).



N-(3,5-dichlorobenzyl)formamide(12) ^[3-6]: ¹H NMR (500 MHz, CDCl₃) δ 8.24 (s, 1H), 7.39 (s, 1H), 7.33 (d, J = 7.7 Hz, 1H), 7.28 (s, 1H), 7.24 (s, 1H), 6.33 (s, 1H), 4.52 (s, 2H). ¹³C NMR (125 MHz, CDCl₃) δ 161.21 (s), 134.20 (s), 133.69 (s), 131.00 (s), 129.39 (s), 127.46 (s), 39.59 (s).



N-(thiophen-2-ylmethyl)formamide(13) ^[3-6]: ¹H NMR (500 MHz, DMSO) δ 8.52 (s, 1H), 8.02 (s, 1H), 7.30 (d, J = 6.0 Hz, 1H), 6.92 – 6.85 (m, 2H), 4.40 (d, J = 6.1 Hz, 2H). ¹³C NMR (125 MHz, DMSO) δ 161.40 (s), 142.30 (s), 127.18 (s), 126.05 (s), 125.55 (s), 36.20 (s).



N-(furan-2-ylmethyl)formamide(14) ^[3-6]: ¹H NMR (500 MHz, DMSO) δ 8.17 (s, 1H), 7.32 (s, 1H), 6.78 (s, 1H), 6.26 (d, *J* = 34.9 Hz, 2H), 4.44 (d, *J* = 5.1 Hz, 2H). ¹³C NMR (125 MHz, DMSO) δ 161.38 (s), 152.26 (s), 142.63 (s), 110.89 (s), 107.40 (s), 34.43 (s).



N-(pyridin-2-ylmethyl)formamide(15) ^[3-6]: ¹H NMR (500 MHz, CDCl₃) δ 9.00 (s, 1H), 8.60 (s, 1H), 8.21 (s, 1H), 7.68 (s, 1H), 7.37 (s, 1H), 7.21 (s, 1H), 4.82 (d, *J* = 5.6 Hz, 2H). ¹³C NMR (125 MHz, CDCl₃) δ 164.65 (s), 156.97 (s), 149.22 (s), 137.35 (s), 126.28 (s), 122.34 (s), 44.69 (s).



N-(naphthalen-1-ylmethyl)formamide(16) ^[3-6]: ¹H NMR (500 MHz, CDCl₃) δ 8.20 (s, 1H), 7.99 (s, 1H), 7.90 – 7.77 (m, 3H), 7.55 (s, 1H), 7.43 (s, 1H), 6.09 (s, 1H), 4.90 (s, 2H). ¹³C NMR (120 MHz, CDCl₃) δ 160.95 (s), 133.88 (s), 132.82 (s), 131.25 (s), 128.84 (s), 126.78 (d, *J* = 8.1 Hz), 126.12 (s), 125.41 (s), 123.36 (s), 40.30 (s).



N-(cyclohexylmethyl)formamide(17) ^[3-6]: ¹H NMR (500 MHz, CDCl₃) δ 7.94 (s, 1H), 6.81 (s, 1H), 3.67 (s, 1H), 1.77 (d, J = 12.3 Hz, 2H), 1.59 (d, J = 3.6 Hz, 2H), 1.21 (t, J = 12.5 Hz, 3H), 1.13 – 0.98 (m, 3H). ¹³C NMR (125 MHz, CDCl₃) δ 160.15 (s), 32.68 (d, J = 5.4 Hz), 31.64 (s), 24.62 (s).



N-phenylformamide(18) ^[3-6]: ¹H NMR (500 MHz, CDCl₃) δ 8.78 (s, 1H), 8.32 (s, 1H), 7.32 (dt, J = 16.0, 8.0 Hz, 4H), 7.19 (d, J = 7.5 Hz, 1H). ¹³C NMR (125 MHz, CDCl₃) δ 160.88 (s), 130.41 (s), 129.74 (s), 129.12 (s), 128.39 (s), 119.67 (s).



N-propylformamide(19) ^[3-6]: ¹H NMR (500 MHz, DMSO) δ 8.85 (s, 1H), 8.00 (s, 1H), 2.98 (s, 2H), 1.28 (s, 2H), 1.18 (s, 2H), 0.75 (s, 3H). ¹³C NMR (125 MHz, DMSO) δ 161.46 (s), 39.33 (s), 22.69 (s), 11.69 (s).

H N /O

N-butylformamide(20) ^[3-6]: ¹H NMR (500 MHz, DMSO) δ 8.85 (s, 1H), 8.00 (s, 1H), 2.98 (s, 2H), 1.28 (s, 2H), 1.18 (s, 2H), 0.75 (s, 3H). ¹³C NMR (125 MHz, DMSO) δ 161.30 (s), 37.18 (s), 31.52 (s), 19.91 (s), 13.79 (s).



pyrrolidine-1-carbaldehyde(21) ^[3-6]: ¹H NMR (500 MHz, DMSO) δ 8.06 (s, 1H), 3.33 (s, 2H), 3.11 (s, 2H), 1.69 (s, 4H).¹³C NMR (125 MHz, DMSO) δ 161.13 (s), 45.84 (s), 43.11 (s), 24.87 (s), 24.19 (s).



piperidine-1-carbaldehyde(22) ^[3-6]: ¹H NMR (500 MHz, CDCl₃) δ 8.02 (s, 1H), 3.42 (dd, *J* = 74.1, 14.0 Hz, 4H), 1.84 - 1.33 (m, 6H). ¹³C NMR (125 MHz, CDCl₃) δ 161.09 (s), 47.02 (s), 40.80 (s), 26.58 (s), 25.09 (s), 24.68 (s).



N,N-dimethylformamide(23) ^[3-6]: ¹H NMR (500 MHz, DMSO) δ 7.88 (s, 1H), 2.80 (s, 3H), 2.64 (s, 3H). ¹³C NMR (125 MHz, DMSO) δ 162.46 (s), 35.78 (s), 30.69 (s).



N,N'-(1,2-phenylene)diformamide(24): ¹H NMR (500 MHz, DMSO) δ 8.28 (s, 1H), 8.16 (s, 1H), 7.61 (s, 2H), 7.20 (d, J = 4.8 Hz, 2H), 6.45 (s, 2H). ¹³C NMR (125 MHz, DMSO) δ 163.56 (s), 122.39 (s), 115.74 (s).



N,N'-(1,3-phenylene)diformamide(25): ¹H NMR (500 MHz, DMSO) δ 10.04 (s, 2H), 8.27 (s, 1H), 8.10 (s, 1H), 7.98 (s, 1H), 7.31 (s, 1H), 7.18 (d, *J* = 7.7 Hz, 2H). ¹³C NMR (125 MHz, DMSO) δ 160.21 (s), 138.82 (s), 130.18 (s), 129.60 (s), 115.31 (s), 110.90 (s), 109.54 (s).



N,N'-(4-methyl-1,3-phenylene)diformamide(26): ¹³C NMR (125 MHz, DMSO) δ 160.21 (s), 138.82 (s), 130.18 (s), 129.60 (s), 115.31 (s), 110.90 (s), 109.54 (s). ¹³C NMR (125 MHz, DMSO) δ 160.22 (s), 159.79 (s), 136.73 (s), 136.21 (s), 131.51 (s), 130.91 (s), 124.80 (s), 116.09 (s), 17.79 (s).



N,N'-(methylenebis(4,1-phenylene))diformamide(27): ¹H NMR (500 MHz, DMSO) δ 10.11 (s, 2H), 8.24 (d, J = 1.4 Hz, 2H), 7.49 (d, J = 8.3 Hz, 4H), 7.15 (d, J = 8.1 Hz, 4H), 3.84 (s, 2H). ¹³C NMR (125 MHz, DMSO) δ 159.86 (s), 137.33 – 137.13 (m), 136.91 (d, J = 49.1 Hz), 129.99 (s), 129.46 (s), 119.75 (s), 118.29 (s).



N,N'-(((sulfonylbis(4,1-phenylene))bis(sulfanediyl))bis(4,1-phenylene))diformamide(28): ¹H NMR (500 MHz, DMSO) δ 10.47 (s, 2H), 8.34 (s, 2H), 7.77 (d, *J* = 8.5 Hz, 4H), 7.73 (d, *J* = 8.4 Hz, 4H), 7.51 (d, *J* = 8.4 Hz, 4H), 7.19 (d, *J* = 8.5 Hz, 4H).¹³C NMR (125 MHz, DMSO) δ 160.46 (s), 146.73 (s), 140.17 (s), 138.24 (s), 136.29 (s), 128.48 (s), 127.00 (s), 121.01 (s).



N,N'-(butane-1,4-diyl)bis(4-((4-formamidophenyl)thio)benzamide)(29): 1H NMR (500 MHz, DMSO) δ 10.41 (s, 2H), 8.43 (s, 2H), 8.33 (s, 2H), 7.76 (d, J = 8.2 Hz, 4H), 7.69 (d, J = 8.4 Hz, 4H), 7.46 (d, J = 8.3 Hz, 4H), 7.18 (d, J = 8.2 Hz, 4H), 3.26 (d, J = 4.4 Hz, 4H), 1.54 (s, 4H). ¹³C NMR (125 MHz, DMSO) δ 165.93 (s), 160.35 (s), 141.54 (s), 139.36 (s), 134.95 (s), 128.52 (s), 127.58 (s), 120.82 (s), 118.90 (s), 27.17 (s).



N,N'-(hexane-1,6-diyl)bis(4-((4-formamidophenyl)thio)benzamide)(30): ¹H NMR (500 MHz, DMSO) δ 10.41 (s, 2H), 8.39 (s, 2H), 8.32 (s, 2H), 7.75 (d, J = 8.2 Hz, 5H), 7.68 (s, 4H), 7.44 (d, J = 6.5 Hz, 5H), 7.18 (d, J = 8.2 Hz, 5H), 3.35 (s, 4H), 1.48 (s, 4H), 1.25 (s, 4H). ¹³C NMR (125 MHz, DMSO) δ 165.86 (s), 160.35 (s), 141.49 (s), 139.36 (s), 134.96 (s), 128.51 (s), 127.56 (s), 120.80 (s), 118.88 (s), 29.49 (d, J = 17.0 Hz), 29.24 (s), 26.95 (s).

IX. NMR spectra



¹H NMR spectra of **2** (500 MHz, CDCl₃)



¹C NMR spectra of **2** (125 MHz, CDCl₃)



¹C NMR spectra of **3** (125 MHz, CDCl₃)



¹C NMR spectra of 4 (125 MHz, CDCl₃)



¹C NMR spectra of **5** (125 MHz, DMSO)







¹C NMR spectra of 6 (125 MHz, CDCl₃)



¹H NMR spectra of 7 (500 MHz, DMSO)



¹C NMR spectra of 7 (125 MHz, DMSO)







¹C NMR spectra of 8 (125 MHz, CDCl₃)







¹C NMR spectra of 9 (125 MHz, CDCl₃)



¹H NMR spectra of **10** (500 MHz, CDCl₃)



¹C NMR spectra of **10** (125 MHz, CDCl₃)







¹C NMR spectra of **11** (125 MHz, CDCl₃)



¹H NMR spectra of **12** (500 MHz, CDCl₃)



¹C NMR spectra of **12** (125 MHz, CDCl₃)







¹C NMR spectra of **13** (125 MHz, DMSO)



¹C NMR spectra of **14** (125 MHz, DMSO)



¹C NMR spectra of **15** (125 MHz, CDCl₃)







¹C NMR spectra of **16** (125 MHz, CDCl₃)







 ^{1}C NMR spectra of 17 (125 MHz, CDCl₃)







¹C NMR spectra of **18** (125 MHz, CDCl₃)



¹H NMR spectra of **19** (500 MHz, DMSO)



¹C NMR spectra of **19** (125 MHz, DMSO)



¹H NMR spectra of **20** (500 MHz, DMSO ₃)



¹C NMR spectra of **20** (125 MHz, DMSO)



¹C NMR spectra of **21** (125 MHz, DMSO)



¹H NMR spectra of **22** (500 MHz, CDCl₃)



¹C NMR spectra of **22** (125 MHz, CDCl₃)







¹C NMR spectra of 23 (125 MHz, DMSO)



¹H NMR spectra of **24** (500 MHz, DMSO)



¹C NMR spectra of **24** (125 MHz, DMSO)







¹C NMR spectra of **25** (125 MHz, DMSO)







¹C NMR spectra of **26** (125 MHz, DMSO)



¹C NMR spectra of **27** (125 MHz, DMSO)







¹C NMR spectra of 28 (125 MHz, DMSO)



¹H NMR spectra of **29** (500 MHz, DMSO)



¹C NMR spectra of **29** (125 MHz, DMSO)







¹C NMR spectra of **30** (125 MHz, DMSO)