

Electronic Supplementary Information

Combination of ^{47}Ti and ^{48}Ti for the determination of highly polydisperse TiO_2 particle size distributions by spICP-MS

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Preparation of sample B1

Enrichment procedure for large TiO_2 particles ($\approx 200\text{-}500\text{ nm}$) contained in coconut syrup (A1) derived from centrifugation equations given in Gimbert *et al.* (2005).

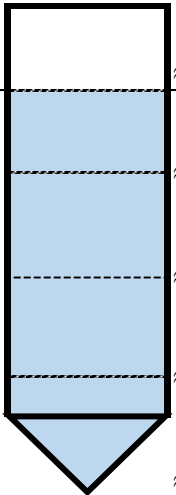
Sample A1 was first diluted 10 times with ultrapure water to reduce its viscosity and match roughly that of ultrapure water. The diluted solution (A1D) was then sonicated for 10 min (ultrasonic bath with 45 kHz frequency and 180 W maximum absorbed power) and centrifuged as follows:

Step 1: Centrifugation of (A1D) with $\approx 1.2\text{ }\mu\text{m}$ minimal cutoff (*i.e.* centrifugal force gradient in the sample (from tube bottom to liquid surface) ranging from 200 to 400 g for 2 min at 20°C) $\rightarrow$ recuperation of the supernatant (A1S1) containing TiO_2 particles mainly smaller than $0.6\text{ - }1.2\text{ }\mu\text{m}$

Step 2: Centrifugation of (A1S1) with $\approx 0.42\text{ }\mu\text{m}$ minimal cutoff (*i.e.* centrifugal force gradient in the sample (from tube bottom to liquid surface) ranging from 1400 to 2800 g for 2 min at 20°C) $\rightarrow$ discard supernatant containing TiO_2 particles smaller than $0.2\text{ - }0.4\text{ }\mu\text{m}$ and re-suspension of the pellet containing TiO_2 particles mainly between 0.2 and $1.2\text{ }\mu\text{m}$ by addition of ultrapure water and vigorous mixing (vortex) resulting in suspension (A1S2).

Step 3: repetition of Step 2 with suspension (A1S2) to retrieve suspension (A1S3)

Step 4: repetition of Step 2 with suspension (A1S3) to retrieve final suspension (A1S4) = sample B1

	Centrifuge Tube	2 min @ 4000 rpm (20°C)	2 min @ 1500 rpm (20°C)
Liquid surface		$\approx 2800\text{ g}$ - Cutoff $\approx 0.42\text{ }\mu\text{m}$	$\approx 400\text{ g}$ - Cutoff $\approx 1.2\text{ }\mu\text{m}$
75 % liquid height		$\approx 2800\text{ g}$ - Cutoff $\approx 0.38\text{ }\mu\text{m}$	$\approx 340\text{ g}$ - Cutoff $\approx 1.0\text{ }\mu\text{m}$
50 % liquid height		$\approx 2100\text{ g}$ - Cutoff $\approx 0.32\text{ }\mu\text{m}$	$\approx 300\text{ g}$ - Cutoff $\approx 0.86\text{ }\mu\text{m}$
25 % liquid height		$\approx 1800\text{ g}$ - Cutoff $\approx 0.24\text{ }\mu\text{m}$	$\approx 250\text{ g}$ - Cutoff $\approx 0.64\text{ }\mu\text{m}$
Tube bottom		$\approx 1400\text{ g}$	$\approx 200\text{ g}$

Additional information regarding sample preparation for spICP-MS analysis.

As sample A1 was already in a liquid form (syrup), it was simply diluted 10 times prior to sonication and centrifugation as described in the manuscript (section 2.2). Samples A2 and A3 both contained E171 additive in their coating, therefore the outer layer of these samples was simply separated from the almond or jellified core by swirling in ultrapure water for about 5 minutes prior to sonication and centrifugation of the resulting suspension as described in the manuscript (section 2.2).

Table S1: Typical spICP-MS operational parameters

Parameter	Value
Plasma Gas Flow (L min ⁻¹)	18.0
Auxiliary Gas Flow (L min ⁻¹)	1.20
Nebulizer Gas Flow ^a (L min ⁻¹)	1.00
RF Power (W)	1500
Sampling Depth (mm)	0.0 ^b
Sample Uptake ^c (mL min ⁻¹)	0.175
RPq (V)	0.45
Deflector Voltage (V)	-11
Quadrupole Rod Offset (V)	0.0
Torch Injector Inner Diameter (mm)	1.8
Nebulizer	Micromist 0.4 mL min ⁻¹
Spray Chamber	Cyclonic (glass) at room temperature
Sampler, skimmer, hyperskimmer cone	nickel
Uptake tubing	PVC 0.38 mm internal diameter
Drain tubing	Santoprene 1.30 mm internal diameter

^aNebulizer Gas Flow optimized daily

^bPosition relative to the initial torch sampling depth set with Perkin Elmer's torch alignment tool

^cSample Uptake measured gravimetrically daily

Validation of the 60 nm and 40 nm AuNPs suspensions used for TE determination and drift control

Since well characterized NIST AuNPs reference materials (RM) were not available at the time of this study, 60 nm and 40 nm AuNPs suspensions from BBI Solutions (Crumlin, UK) were used for TE determination and independent drift control (sizing and counting), respectively. Therefore, in order to make sure that BBI's 60 nm AuNPs suspension was reliable as a TE standard, we successfully took part in RIKILT's proficiency testing (Wageningen University & Research, The Netherlands) for size (z-score = 0.15) and particle concentration (z-score = 0.03) determination by spICP-MS in 2018. Additionally, we have also successfully determined the size of several other AuNPs suspensions including 20, 40, 80 and 100 nm AuNPs from BBI Solutions (Crumlin, UK) and 30 nm AuNPs from NIST (RM 8012).

Additional information regarding potential ^{48}Ca isobaric interference on ^{48}Ti

Samples A1, A2 and A3 supernatants (*i.e.* fractions $< 1.2\ \mu\text{m}$ subjected to spICP-MS analysis) were analyzed by conventional ICP-MS after acidic digestion for total Ti and Ca determination in order to determine the extent of ^{48}Ca isobaric interference on ^{48}Ti . The results presented in Table S2 showed a $^{48}\text{Ca}/^{48}\text{Ti}$ ratio below 0.1% for samples A1 and A2 and around 0.5% for sample A3 which contained about 10-times less TiO_2 compared to samples A1 and A2. These results demonstrated that ^{48}Ca isobaric interference on ^{48}Ti could be neglected for these samples even without using the ICP-MS collision cell. Additionally, the presence of Ca (and therefore 0.187% ^{48}Ca) or S (and therefore potentially $^{32}\text{S}^{16}\text{O}$) would essentially result in elevated ionic (“dissolved”) background. The “dissolved” ^{48}Ti concentration (included in the result summary from the Syngistix software) was carefully monitored and was systematically much lower than $0.05\ \mu\text{g L}^{-1}$ when the appropriate dilution factor was achieved (*i.e.* TiO_2 concentration about 1.5 to $10\ \mu\text{g L}^{-1}$).

Table S2: Ca and Ti contents in samples A1, A2 and A3 supernatants which were subjected to spICP-MS analysis

Sample	Total Ca ^a content ($\mu\text{g/g}$)	^{48}Ca content ^b ($\mu\text{g/g}$)	Total Ti ^c [TiO_2] ^d content ($\mu\text{g/g}$)	^{48}Ti content ^e ($\mu\text{g/g}$)	$^{48}\text{Ca}/^{48}\text{Ti}$ ratio (%)
A1	113	0.214	448 [747]	330	0.06%
A2	41.7	0.079	420 [700]	309	0.03%
A3	77.0	0.146	39.3 [65.5]	29.0	0.51%

^aTotal Ca content determined by ICP-MS using ^{44}Ca isotope and He collision gas

^bConsidering ^{48}Ca isotopic abundance of 0.187%

^dTotal Ti content determined by ICP-MS using ^{47}Ti isotope and He collision gas

^dConsidering Ti/ TiO_2 mass fraction of 59.9%

^eConsidering ^{48}Ti isotopic abundance of 73.72%

Table S3: Example of a typical spICP-MS analytical sequence

Sample	Analyte	Run time (s)	Verifications
ST0 Au (Blank)	¹⁹⁷ Au	60	¹⁹⁷ Au ionic calibration Linearity R² ≥ 0,999
ST1 Au (1 ppb)	¹⁹⁷ Au	60	
ST2 Au (5 ppb)	¹⁹⁷ Au	60	
ST3 Au (10 ppb)	¹⁹⁷ Au	60	
Blank AuNP	¹⁹⁷ Au	60	
60 nm AuNP STANDARD	¹⁹⁷ Au	60	
40 nm AuNP CONTROL #1	¹⁹⁷ Au	60	Size recovery : 95 – 105 %
ST0 Ti (Blank)	⁴⁸ Ti	60	⁴⁸ Ti ionic calibration Linearity R² ≥ 0,999
ST1 Ti (5 ppb)	⁴⁸ Ti	60	
ST2 Ti (10 ppb)	⁴⁸ Ti	60	
ST3 Ti (15 ppb)	⁴⁸ Ti	60	
ST0 Ti (Blank)	⁴⁷ Ti	60	⁴⁷ Ti ionic calibration Linearity R² ≥ 0,999
ST1 Ti (5 ppb)	⁴⁷ Ti	60	
ST2 Ti (10 ppb)	⁴⁷ Ti	60	
ST3 Ti (15 ppb)	⁴⁷ Ti	60	
Blank	⁴⁸ Ti	60	Same sample analyzed twice without delay (no rinsing sequence)
Blank	⁴⁷ Ti	60	
Sample 1 - C1.2µm D10000	⁴⁸ Ti	60	Same sample analyzed twice without delay (no rinsing sequence)
Sample 1 - C1.2µm D10000	⁴⁷ Ti	60	
40 nm AuNP CONTROL #2	¹⁹⁷ Au	60	Size recovery : 95 – 105 % Number recovery : 80 – 120 %

Additional data regarding sample preparation and spICP-MS analysis reliability and repeatability

Repeatability for both the sample preparation methodology and the spICP-MS analysis was demonstrated by repeating the entire protocol with different E171 food additives and final products. Reliability of the size measurement by spICP-MS was demonstrated through comparisons with scanning electron microscope (SEM) data. Table S4 presents the figures of repeatability for one E171 additive premix used as in-house control sample and Tables S4 and S5 present the figures of repeatability and reliability for two final products obtained through official sampling on the French market.

Relative standard deviations (RSDs) ≤ 5% were obtained for the mean and the median size measurements for the E171 additive premix (Table S4), the chewing gum coating (Table S5) and the chocolate candy coating (Table S6). These low RSDs demonstrated both intra-day and inter-day repeatability of the whole procedure for the E171 additive premix and final products with E171 containing coating.

Sizing accuracies (spICP-MS vs. SEM primary particle measurement) ranged between 93% and 98% for both the mean and the median size measurements of the chewing gum coating (Table S5) and the chocolate candy coating (Table S6). These good recoveries supported the reliability of the spICP-MS size measurements as well as the suitability of the sample preparation

methodology for final products with E171 containing coating. In addition, the similarity between spICP-MS and SEM results supports the idea that only few aggregates are present in the prepared solutions.

Table S4: Figures of repeatability obtained for in-house E171 additive premix used as spICP-MS control sample

In-house E171 additive premix used as spICP-MS control sample	Day #	Number of measurements	Mean size			D₅₀ – Median size		
			Mean (nm)	SD (nm)	RSD (%)	Mean (nm)	SD (nm)	RSD (%)
	1	1	173	N/A	N/A	162	N/A	N/A
	2	3	180	2,9	2%	168	2,0	1%
	3	2	174	2,8	2%	164	3,5	2%
	4	2	170	0,0	0%	157	0,7	0%
	5	3	167	3,1	2%	156	3,1	2%
	6	2	171	1,4	1%	161	3,5	2%
	7	5	174	3,0	2%	161	1,9	1%
	8	3	169	2,2	1%	156	5,9	4%
	9	3	170	6,0	4%	156	7,1	5%
	10	2	169	0,7	0%	157	0,7	0%
	11	2	166	1,4	1%	154	0,7	0%
Global		28	172	4,8	3%	159	5,1	3%

Table S5: Figures of repeatability and reliability (vs. SEM measurement) obtained by spICP-MS for a typical chewing gum tablet with white coating (analysis of the coating containing the E171 additive)

Chewing gum spICP-MS	Replicate preparation	Dilution level	Mean diameter (nm)	D50 - Median size (nm)	
	1 (Day 1)	1	144	139	
		2	137	132	
	2 (Day 1)	1	143	139	
		2	137	131	
	3 (Day 1)	1	142	136	
		2	140	132	
			Mean (nm)	140	135
			SD (nm)	2,8	3,6
			RSD (%)	2,0%	2,7%
SEM analysis for confirmation 300 primary particles measured			137	127	
% Sizing accuracy (spICP-MS vs. SEM)			98%	94%	

Table S6: Figures of repeatability and reliability vs. SEM measurement obtained by spICP-MS for a typical chocolate candy with peanut core and colored coating (analysis of the coating containing the E171 additive)

Chocolate candy spICP-MS	Replicate preparation	Dilution level	Mean diameter (nm)	D50 - Median size (nm)
	1 (Day 1)	1	125	110
		2	129	113
	2 (Day 2)	1	135	119
		2	132	116
	3 ^a (Day 3)	1	133	120
		Mean (nm)	131	116
		SD (nm)	3,7	4,4
		RSD (%)	2,8%	3,8%
SEM analysis for confirmation 300 primary particles measured			122	114
% Sizing accuracy (spICP-MS vs. SEM)			93%	98%

^aOnly one dilution level was analyzed on day 3

Determination of SiO₂ content and particle concentrations in silica suspensions by ICP-OES

Silica microsphere suspensions (300, 500 and 1000 nm nominal diameters) were thoroughly homogenized and diluted 10-fold in ultrapure water. Then, 500 µL aliquots were digested in duplicate with 100 µL HF (40%w/w) and 5000 µL HNO₃ (69%w/w). 50 µL of internal standard (1000 µg mL⁻¹ Sc) were added to the resulting digests and the volume was brought to 50 mL with ultrapure water for ICP-OES analysis (Varian 720-ES equipped with Agilent OneNeb HF compliant nebulizer; Sc 361.383 nm and Si 288.158 nm). Calibration was achieved using Si standards (0.5 – 1 – 5 – 10 – 20 µg mL⁻¹) supplemented with Sc internal standard. For further calculations, Si/SiO₂ mass fraction was considered to be 46.7% and SiO₂ density 2.0 g cm⁻³ (data provided by the manufacturer). Procedural blanks (*i.e.* 100 µL HF (40%w/w) + 5000 µL HNO₃ (69%w/w) + ultrapure water up to 50 mL) were analyzed and subtracted to the samples. Results are presented in Table S7.

SiO₂ particle concentrations are presented in Table S7 and were calculated according to the following equation:

$$SiO_2 \text{ particle conc.} = \frac{6 \times Cs}{10^{-18} \times Dp^3 \times \pi \times \rho}$$

where *SiO₂ particle conc.* is the target particle concentration (mL⁻¹), *Cs* is the SiO₂ concentration determined by ICP-OES (Table S7; mg mL⁻¹), *Dp* is the SiO₂ particle diameter provided by the manufacturer (nm) and *ρ* is the density of the SiO₂ particles provided by the manufacturer (2.0 g cm⁻³).

Table S7: SiO₂ and particle concentrations in silica suspensions – Uncertainties were calculated as the standard deviation of the duplicate values (n = 2 preparations).

SiO ₂ particle diameter from manufacturer (nm)	SiO ₂ conc. ICP-OES (mg mL ⁻¹)	SiO ₂ particle conc. (mL ⁻¹)
299	90 ± 9	(3.2 ± 0.3)10 ¹²
507	95 ± 8	(7.0 ± 0.5)10 ¹¹
1046	92 ± 9	(7.7 ± 0.8)10 ¹⁰

Sample B1 analysis with 10 millisecond dwell time was performed in order to determine whether the detector saturation phenomenon also occurred with this higher integration duration. Fig. S1 shows the TiO₂ PSDs obtained for sample B1 when acquiring ⁴⁷Ti and ⁴⁸Ti signals at 10 ms dwell time. Both ⁴⁷TiO₂ and ⁴⁸TiO₂ PSDs were very similar and all the following observations are comparable to those obtained with 100 µs dwell time. Indeed, very few particles (*i.e.* 23 out of 1195 particles) were detected below 100 nm and the frequency hardly reached a maximum of 7 for ⁴⁸Ti at 70 nm and 90 nm. From 100 nm up to 200 nm, both ⁴⁸TiO₂ and ⁴⁷TiO₂ frequencies overlapped quite well and increased steadily. Above 200 nm, ⁴⁷TiO₂ and ⁴⁸TiO₂ PSDs diverged significantly. ⁴⁸TiO₂ PSD increased steeply to reach a maximum frequency of 117 at 280 nm and collapsed rapidly ending around 400 nm while ⁴⁷TiO₂ PSD was shaped like a Gaussian curve (solid black line in Fig. S1) spreading from 100 nm up to 600 nm and centered on 352 nm with an average maximum frequency of 48 and a standard deviation of 94 nm (obtained with OriginPro 2015 software). The local maximum of the ⁴⁷TiO₂ PSD frequency is 55 at 350 nm. Combination of ⁴⁸TiO₂ (≤ 150 nm) and ⁴⁷TiO₂ (> 150 nm) PSD is presented as a solid blue line on Fig. S1 and once again is very similar to the one obtained with 100 µs dwell time. These observations demonstrated that the saturation of ⁴⁸Ti signal for larger TiO₂ particles was not limited to microsecond dwell time but might also appear using millisecond dwell times.

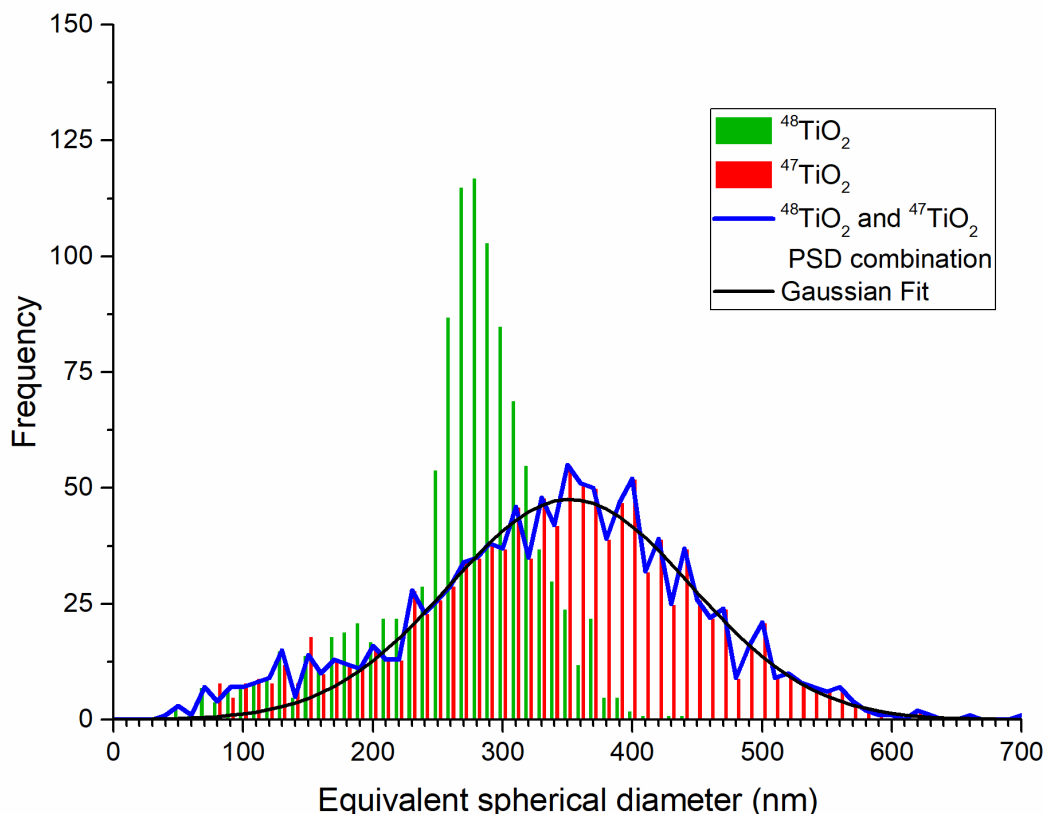


Fig. S1: TiO₂ PSD of sample B1 with 10 ms dwell time - ⁴⁷TiO₂ and ⁴⁸TiO₂ PSDs of sample B1, PSD combination (*i.e.* ⁴⁸TiO₂ data ≤ 150 nm and ⁴⁷TiO₂ data > 150 nm) as solid blue line and Gaussian fitting (OriginPro 2015 software) of the PSD combination as solid black line.

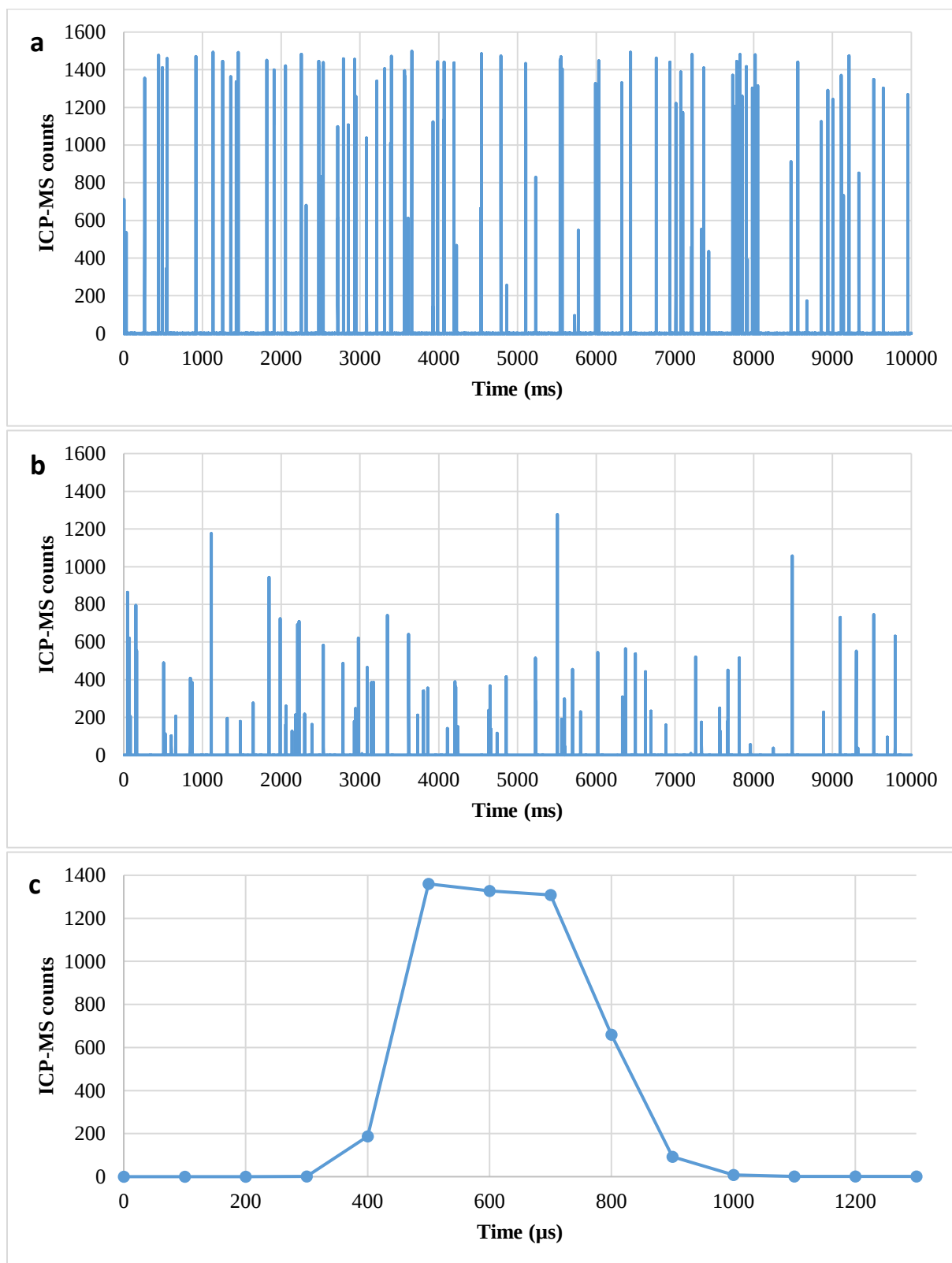
Table S8: Physico-chemical properties of Ti, TiO₂, Si and SiO₂ (ranges are provided when different values are reported in the literature especially depending on the crystallographic form *e.g.* TiO₂ rutile vs. anatase, SiO₂ amorphous vs. quartz)

Material	Molecular weight (g mol ⁻¹)	Usual density range (g cm ⁻³)	Melting point (°C)	Boiling point (°C)	Ionization Energy (eV)
Ti	47.87	4.51	1668	3287	6.8281 (Ti)
TiO ₂	79.87	3.79 - 4.23	1843 (1560 - 1912)	2972	7.8 - 9.5 (TiO ₂)
Si	28.09	2.33	1414	3265	8.1517 (Si)
SiO ₂	60.08	1.9 - 2.7	1713 - 1722	2950	11.8 - 12.6 (SiO ₂)

All data (except some ionization energy data for TiO₂ and SiO₂) are from Handbook of Chemistry and Physics 89th edition (edited by David R. Lide) and NIST online Chemistry WebBook n°69 (<https://doi.org/10.18434/T4D303> - last access 19/12/2018).

Additional ionization energy data from Scanlon *et al.* (2013), Maj *et al.* (1988) and Kostko *et al.* (2009).

Fig. S2: Illustration of spICP-MS signal saturation - Raw data (10 s section extract) for sample B1, comparison of raw signal (spikes) during ^{48}Ti (a) and ^{47}Ti (b) acquisition. Example of a TiO_2 particle clipped spike with detector saturation around 1400 counts during ^{48}Ti acquisition (c).



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