SUPPLEMENTARY DOCUMENTATION

Liquid Biopsy using the Nanotube-CTC-Chip: Capture of Invasive CTCs with High Purity using Preferential Adherence in Breast Cancer Patients

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Table of Contents

Figure S_1 : (a) Fluorescence microscopy of blood droplet with MDA-MB-231-GFP cells at different depth of focus; (b) number of GFP observations versus number of cells spiked; (c) number of observed cells in each droplet. The observations indicated anywhere from 87-100% capture is possible. Slight errors in cell count at lower concentration is a result of spiking using a hemocytometer
Figure S_2 : Fluorescence microscopy of the entire droplet with MDA-MB-231-GFP cells4
Figure S ₃ : (a) RBC lysis protocol; (b) blood smear before and after lysis; (c) Adhered versus non-adhered cells
Figure S_4 : Method of preferential adherence of cancer cell lines on nanotube surface: (a) U-251, U-343 and LN-229 cells adhered to the nanotube surface; (b) Hela cells attached to the nanotube surface stained for CD59
Figure S_5 : Optical images and merge images of patient samples7
Figure S ₆ : Heterogeneous CTCs and WBCs on the same chip; optical, DAPI, EGFR, CK and merge images. A single CTC is seen at the bottom of each image suggesting this CTC was positive for DAPI, CK, EGFR suggesting multiple phenotypes on the same cell
Figure S ₇ : Epithelial, mesenchymal and EMT related CTCs along with WBCs (DAPI only)9
Table S1: Antibodies, manufacturer and dilutions 10



Figure S₁: (a) Fluorescence microscopy of blood droplet with MDA-MB-231-GFP cells at different depth of focus; (b) number of GFP observations versus number of cells spiked; (c) number of observed cells in each droplet. The observations indicated anywhere from 87-100% capture is possible. Slight errors in cell count at lower concentration is a result of spiking using a hemocytometer.



Figure S₂: Fluorescence microscopy of the entire droplet with MDA-MB-231-GFP cells.



Figure S_3 : (a) RBC lysis protocol; (b) blood smear before and after lysis; (c) Adhered versus non-adhered cells.



Figure S₄: Method of preferential adherence of cancer cell lines on nanotube surface: (a) U-251, U-343 and LN-229 cells adhered to the nanotube surface; (b) Hela cells attached to the nanotube surface stained for CD59.



Figure S₅: Optical images and merge images of patient samples.



Figure S₆: Heterogeneous CTCs and WBCs on the same chip; optical, DAPI, EGFR, CK and merge images. A single CTC is seen at the bottom of each image suggesting this CTC was positive for DAPI, CK, EGFR suggesting multiple phenotypes on the same cell.



Figure S7: Epithelial, mesenchymal and EMT related CTCs along with WBCs (DAPI only)

Antibody	Manufacturer	Host	Clone	Used dilution
				(µg/ml)
Cytokeratin	Thermofisher	Mouse	Zym5.2 (UCD/PR.10-11)	1:100
8/18	(#180213)			
EGFR	CellSignal (#4267S)	Rabbit	D38B1	1:50
Her2	Thermofisher	Mouse	e2-4001, 3B5	1:100
	(#MA5-14057)			
CD45	Thermofisher	Rat	YAML501.4	1:500
	(#MA5-17687)			
DAPI	CellSignal (#4083S)			0.1 μg/ml

Table S₁: Antibodies, manufacturer and dilutions