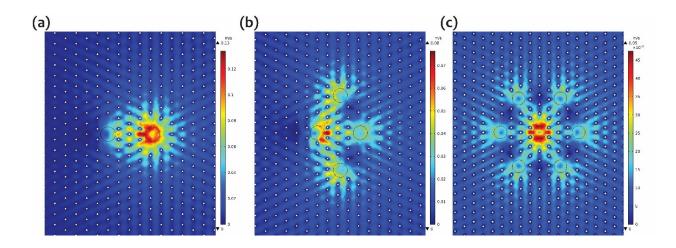
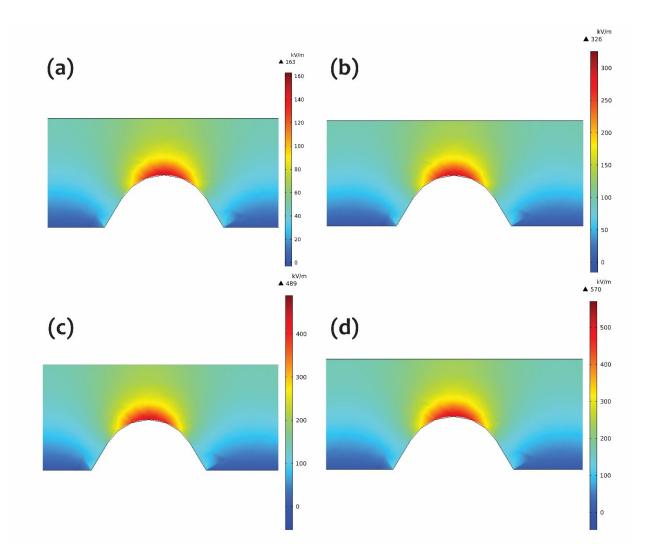
## **Supplementary Figures**

## Microelectrofluidic Probe for Sequential Cell Separation and Patterning

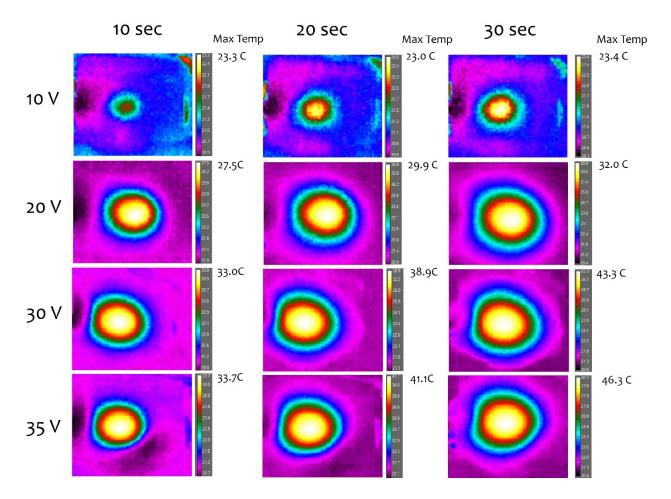
Ayoola T. Brimmo<sup>1, 2</sup>, Anoop Menachery<sup>1</sup>, Mohammad A. Qasaimeh<sup>1,2</sup> <sup>1</sup>Division of Engineering, New York University Abu Dhabi <sup>2</sup>Department of Mechanical and Aerospace Engineering, New York University, New York



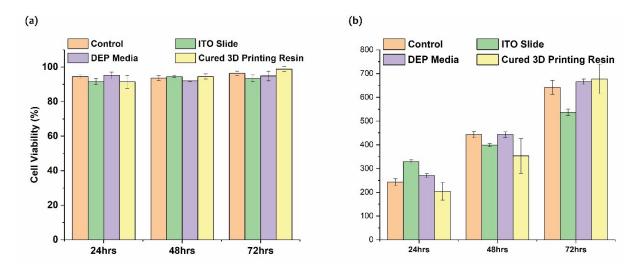
**Fig. S1. Velocity profile at capture plane with 20 µl/min inlet flow rate.** (a) Dipole Microfluidic configuration. (b) Quadrupole Microfluidic configuration. (c) Heptapole Microfluidic configuration.



**Fig. S2. Electric field strength as a function applied voltage.** (a) 10 Vpk-pk. (b) 20 Vpk-pk. (c) 30V pk-pk. (d) 40 Vpk-pk.



**Fig. S3. Temperature measurements as a function of applied voltage and exposure time.** Transient Infrared images as a function of applied voltage showed that the maximum temperature cells were exposed to at 35 V is 46.3 C



**Fig. S4. Impact of materials on cell viability and growth density.** (a) Cell viability as a function of time for different materials used in the study. (b) Cell density as a function of time for different materials used in the study. Error bars denote standard error.

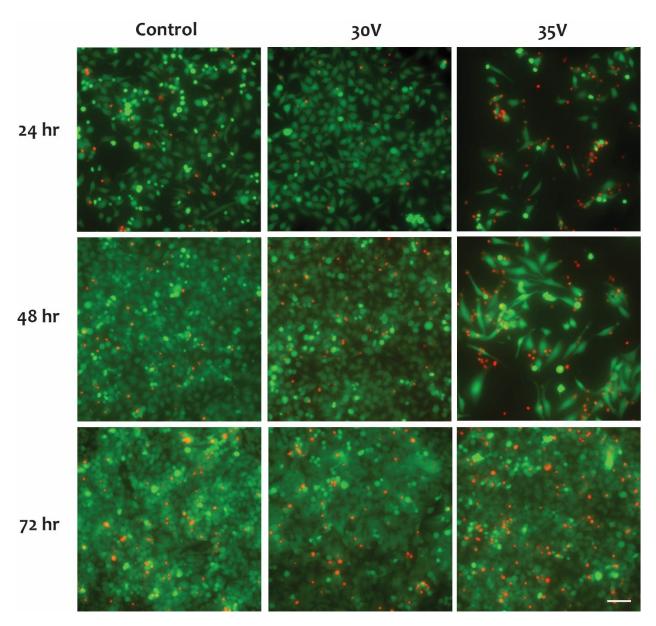
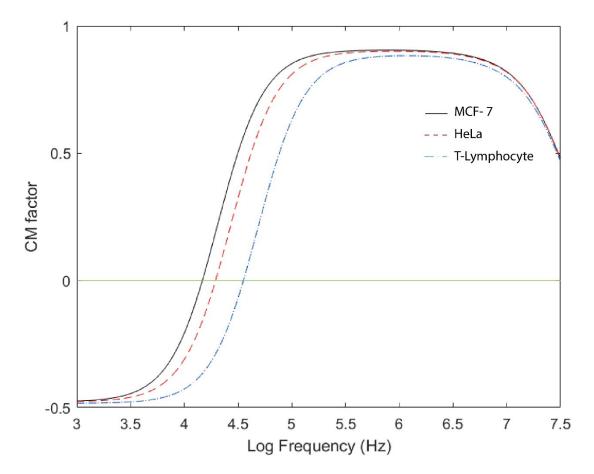


Fig. S5. Fluorescent image for live/dead viability study. Green denotes live cells and red denotes dead cells. Scale bar is  $100 \ \mu m$ .



**Fig. S6. Calculated CM function of MCF 7, HeLa, and T-lymphocytes.** Frequency dependent CM-function calculated using the two-shelled model