

Supplementary Information

Microfluidic screening system based on boron-doped diamond electrodes and dielectrophoretic sorting for directed evolution of NAD(P)-dependent oxidoreductases

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Supplementary Table S1 The oligonucleotide sequences used in this study

Primer	Sequence
T7-F	5'-TAATACGACTCACTATAGGG-3'
T7-R	5'-GCTAGTTATTGCTCAGCGG-3'
T7-F2	5'-TAATACGACTCAACTATAGGGAGACCACAAC-3'
T7-R2	5'-GCTAGTTATTGCTCAGCGGTGGC-3'
Insert-F	5'-ACCGCGAACAGATTGGAGGTGCGGAAAAAGTGTCGTTTGA-3'
Insert-R	5'-AGCAGCCGGATCCTCGAGTCACTCGAGGGCGGCCGGAGCGT-3'
Vector-F	5'-ACCTCCAATCTGTTCGCGGTG-3'
Vector-R	5'-TGA CT CGAGGATCCGGCTGCT-3'

Supplementary Table S2 The data corresponding to Figure 3 for peak currents of plugs at various concentrations (0, 1, 10, 20 and 50 μ M) of NADH.

	0 μ M	1 μ M	10 μ M	20 μ M	50 μ M
Mean	4.13	7.17	22.9	51.5	98.3
SD	2.08	3.38	4.75	3.71	12.0
Number of samples	1036	652	606	490	383

Supplementary Table S3 The data corresponding to Figure 4 for peak currents of plugs containing microbeads on which the wild-type ADH (WT) or randomly mutated ADH (Mn^{2+} concentration: 0.1, 0.3 or 0.5 mM) was immobilized.

	WT	0.1 mM	0.3 mM	0.5 mM
Mean	68.0	48.4	56.8	52.9
SD	16.0	10.8	9.9	36.8
Number of samples	208	182	327	141

Supplementary Table S4 The data corresponding to Figure 6 for relative peak currents of plugs containing microbeads on which the wild-type IDH (WT) or randomly mutated IDH library before (Mut) and after the first or second round of screening (1st and 2nd) was immobilized.

	WT	Mut (R0)	1st (R1)	2nd (R2)
Mean	1.0	0.92	0.98	1.0
SD	0.038	0.019	0.038	0.019
Number of samples	493	534	837	439

a

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10      20      30      40      50      60      70      80      90
CATATGAAAGCAGCCGTGGTAGAACAGTTTAAAGAGCCGCTGAAATCAAAGAAGTGGAGAAACCGACGATCTCGTATGGCGAAGTTTAA
M K A A V V E Q F K E P L K I K E V E K P T I S Y G E V L

100     110     120     130     140     150     160     170     180
GTGCGCATCAAAGCATGCGGTGTATGCCATACCGATCTTACCGCAGCTCATGGGGATTGGCCGGTAAACCAAACCTGCCGTTAATCCCA
V R I K A C G V C H T D L H A A H G D W P V K P K L P L I P

190     200     210     220     230     240     250     260     270
GGACACGAAGGTGTGGGCATCGTGGAGAAGTTGGTCTGGGGTTACGCACCTGAAAGTTGGCGATCGCGTCGGTATTCTTGGCTGTAT
G H E G V G I V E E V G P G V T H L K V G D R V G I P W L Y

280     290     300     310     320     330     340     350     360
TCAGCATGTGGCCATTGCGACTATTGCCTGTCTGGTCAGGAAACGTTATGCGAACACCAGAAGAATGCAGGCTATTCCGTGGATGGTGGT
S A C G H C D Y C L S G Q E T L G E H Q K N A G Y S V D G G

370     380     390     400     410     420     430     440     450
TATGCCGAGTACTGTGCTGCTGCGGCAGACTACGTGGTTAAGATTCCGGACAACCTTGTGTTGAAGAAGCTGCCCGGATTTTCTGTGCT
Y A E Y C R A A A D Y V V K I P D N L S F E E A A P I F C A

460     470     480     490     500     510     520     530     540
GGCGTCACAACCTACAAAGCGCTGAAAGTTACCGGTGCTAAACCAGGCGAATGGGTTGCGATTTATGGCATCGGTGGCTTAGGTCATGTT
G V T T Y K A L K V T G A K P G E W V A I Y G I G G L G H V

550     560     570     580     590     600     610     620     630
GCGGTGCAATACGCGAAAGCCATGGGCCTGAATGTCGTGGCCGTGGACATTGGTGACGAGAAGCTGGAACCTGCCAAAGAACTCGGTGCA
A V Q Y A K A M G L N V V A V D I G D E K L E L A K E L G A

640     650     660     670     680     690     700     710     720
GATCTGGTGGTCAATCCGCTGAAGGAAGATGCCGCGAAATTCATGAAAGAGAAAGTCCGAGGGGTGCATGCGGCTGTCGTAACCTGCGGTA
D L V V N P L K E D A A K F M K E K V G G V H A A V V T A V

730     740     750     760     770     780     790     800     810
AGTAAGCCGGCCTTTTCAGAGCGCGTACAACAGCATTCCGGCGTGGAGGCGCGTGTGTGCTGGTTGGGCTTCCGCCTGAAGAGATGCCATT
S K P A F Q S A Y N S I R R G G A C V L V G L P P E E M P I

820     830     840     850     860     870     880     890     900
CCGATTTTCGACACTGTGCTGAACGGGATCAAATTATCGGAAGCATTGTAGGCACCCGCAAGATCTGCAGGAAGCGCTCCAATTTGCC
P I F D T V L N G I K I I G S I V G T R K D L Q E A L Q F A

910     920     930     940     950     960     970     980     990
GCGGAAGGCAAAGTCAAGACCATCATTGAAGTTCAACCCCTTGAGAAGATCAACGAAGTCTTTGATCGCATGTTGAAAGGCCAGATTAAT
A E G K V K T I I E V Q P L E K I N E V F D R M L K G Q I N

1000    1010    1020    1030    1040    1050    1060    1070    1080
GGCCGTGTCGTAATGACATTGGAGGATAAAATGATCGCGGCCGTTTACCCATACGATGTTCCCTGACTATGCGGGCTATCCCTATGACGTC
G R V V L T L E D K G S A A V Y P Y D V P D Y A G Y P Y D V

1090    1100    1110    1120    1130    1140
CCGGAATGACAGGGTCCCTATCCCTATGACGTTCCAGATTACGCTCCGGCCGCCCTCGAG
P D Y A G S Y P Y D V P D Y A P A A L E
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Supplementary Fig. S1 The DNA and amino acid sequences of dehydrogenases used in this study. (a) Alcohol dehydrogenase (ADH) from *Geobacillus stearothermophilus*. (b) Isocitrate dehydrogenase (IDH) from *Streptococcus mutans*. The codon usage is optimized for expression in *Escherichia coli*. NdeI, BamHI and XhoI recognition sites are indicated in the boxes. Underline indicates 3xHA-tag.

b

10 20 30 40 50 60 70 80 90
CATATGCGGAAAAAGTGTGTTTGAAGAGGGTAACTTCAAGTGCCGGATAAACCGGTTATACCGTACATTGAAGGTGATGGAGTTGGT
M A E K V S F E E G K L Q V P D K P V I P Y I E G D G V G

100 110 120 130 140 150 160 170 180
CAGGACATTTGGAAGAATGCGCAAATCGTCTTCGACAAAGCCATTGCCAAGGTGATGGGGGTACAAAACAGGTGATCTGGCGTGAAGTT
Q D I W K N A Q I V F D K A I A K V Y G G H K Q V I W R E V

190 200 210 220 230 240 250 260 270
CTTGCAAGGCAAAAAGGCCTATAACGAAACGGGCAACTGGCTGCCAATGAGACGCTGGAATCATAAAACCCACTTACTGGCGATAAAA
L A G K K A Y N E T G N W L P N E T L E I I K T H L L A I K

280 290 300 310 320 330 340 350 360
GGGCCACTGGAACCTCCGGTTGGTGGGGCATTGTTCACTGAACGTAGCGTTACGTCAAGAACTGGACCTGTTTGCCTGCGTTCCACCA
G P L E T P V G G G I R S L N V A L R Q E L D L F A C V R P

370 380 390 400 410 420 430 440 450
GTACGCTACTTCAAGGGTGTACCCTCTCTGAAACATCCGAAAAACCGGCTATTACGATTTTCCGGGAAAACACCGAGGACATTTAT
V R Y F K G V P S P L K H P E K T A I T I F R E N T E D I Y

460 470 480 490 500 510 520 530 540
GCTGGCATTGAATGGAATGCGGGTACTGCGGAAGTGCAGAAAGTATCAACTTTTTGCAGGATGATATGCAGGTAAAGAAGATCCGCTTT
A G I E W N A G T A E V Q K V I N F L Q D D M Q V K K I R F

550 560 570 580 590 600 610 620 630
CCGAAAAGTCCAGCATTGGCATTAAACCGATAAGCATCGAAGGGTCCGCAACGCCTATTAGAGCCGCAATTGAGTATGCACTGGCCAAT
P K S S S I G I K P I S I E G S Q R L I R A A I E Y A L A N

640 650 660 670 680 690 700 710 720
AACCTGACCAAAGTGACCCTGGTCCATAAAGGAAACATCCAGAAGTTCACCGAAGGAGGCTTTCCGCAATGGGGGTATGAGTTGGCTAAA
N L T K V T L V H K G N I Q K F T E G G F R K W G Y E L A K

730 740 750 760 770 780 790 800 810
CGCGAGTATGCTGCCAACTTGCAGTGGCCAGTTAGTGGTTGACGACATTATTGCGGATAACTTCTCCAGCAGATTCTGCTGAAACCT
R E Y A A E L A S G Q L V V D D I I A D N F L Q Q I L L K P

820 830 840 850 860 870 880 890 900
GAGCGTTTTGATGTGGTGGCACTGACCAACCTTAACGGCGATTACGCCAGTGTGCGTTAGCTGCACAAGTTGGCGGTATCGGCATTTCA
E R F D V V A L T N L N G D Y A S D A L A A Q V G G I G I S

910 920 930 940 950 960 970 980 990
CCAGGTGCGAATATCAATTACCAGACTGGCCATGCCATTTTCGAAGCGACACACGGAACCGCACCCGATATCGCAGGTCAAGATCTCGCG
P G A N I N Y Q T G H A I F E A T H G T A P D I A G Q D L A

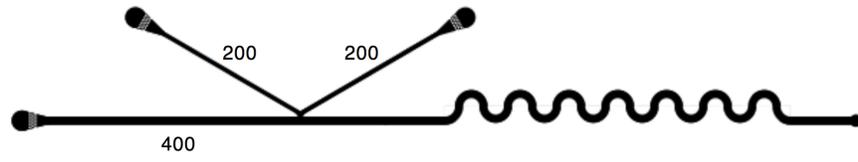
1000 1010 1020 1030 1040 1050 1060 1070 1080
AATCCTAGCTCCGTCTTGTGTCGGGTTGTATGCTCTTTGACTATATCGGCTGGTCTAAAGTTAGCGATCTGATCATGAAAGCTGTGCGAG
N P S S V L L S G C M L F D Y I G W S K V S D L I M K A V E

1090 1100 1110 1120 1130 1140 1150 1160 1170
AAAGCAATTGCGAATGGCCAGGTGACAATCGATTTTGCCAAAGAACTAGGCGTGAAGCGTTGACCACACGTCAGTTTAGCGAAGTCTCTG
K A I A N G Q V T I D F A K E L G V E A L T T R Q F S E V L

1180 1190 1200 1210 1220 1230 1240 1250 1260
CTGACGTAATGATCCGCGGCGTTTACCCATACGATGTTCTGACTATGCGGGCTATCCCTATGACGTCCCGGACTATGCAGGGTCC
L T Y L G S A A V Y P Y D V P D Y A G Y P Y D V P D Y A G S

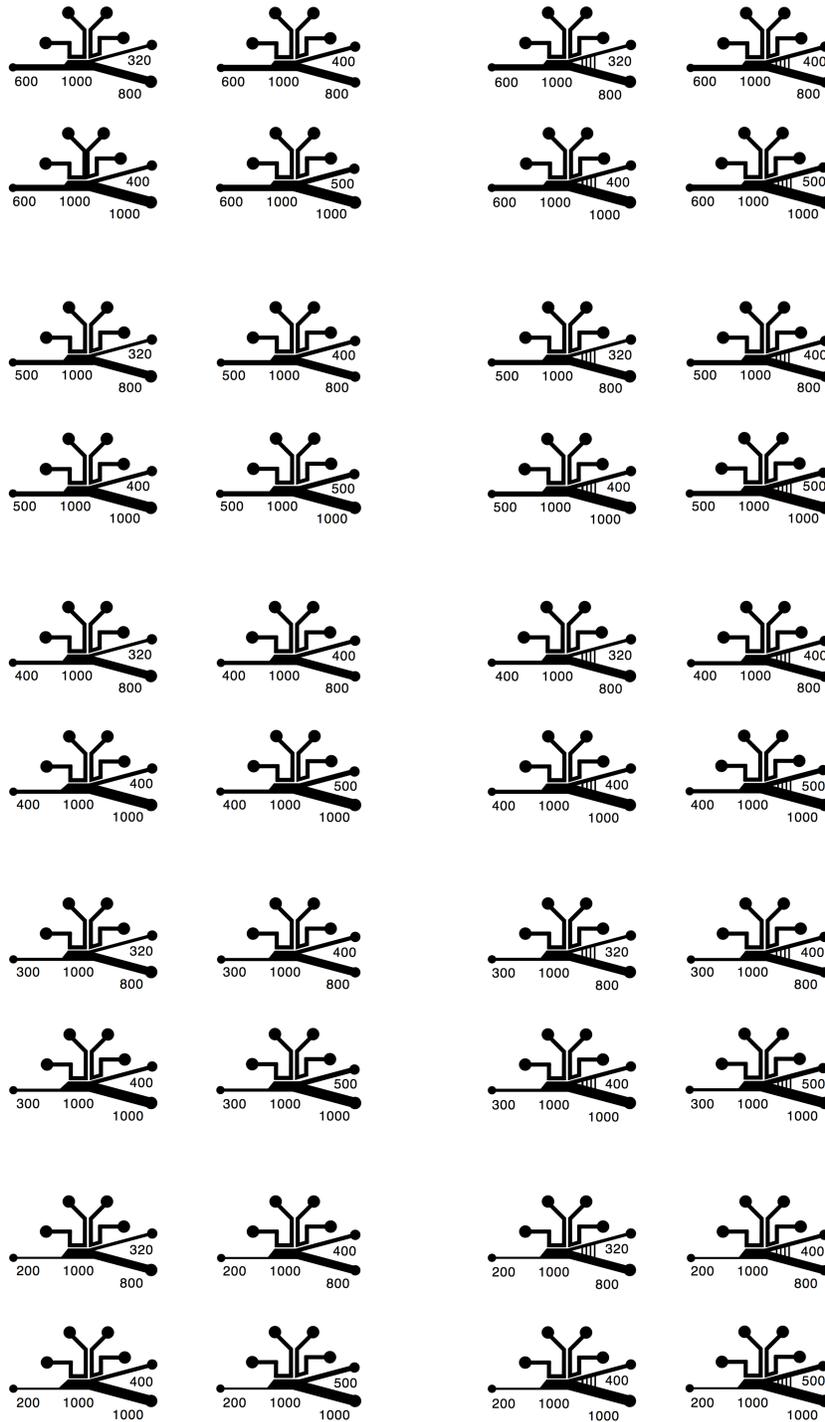
1270 1280 1290 1300
TATCCCTATGACGTTCCAGATTACGCTCCGGCCGCCCTCGAG
Y P Y D V P D Y A P A A L E

Supplementary Fig. S1 (continued)



Supplementary Fig. S2 Masks for the fabrication of the encapsulating device.

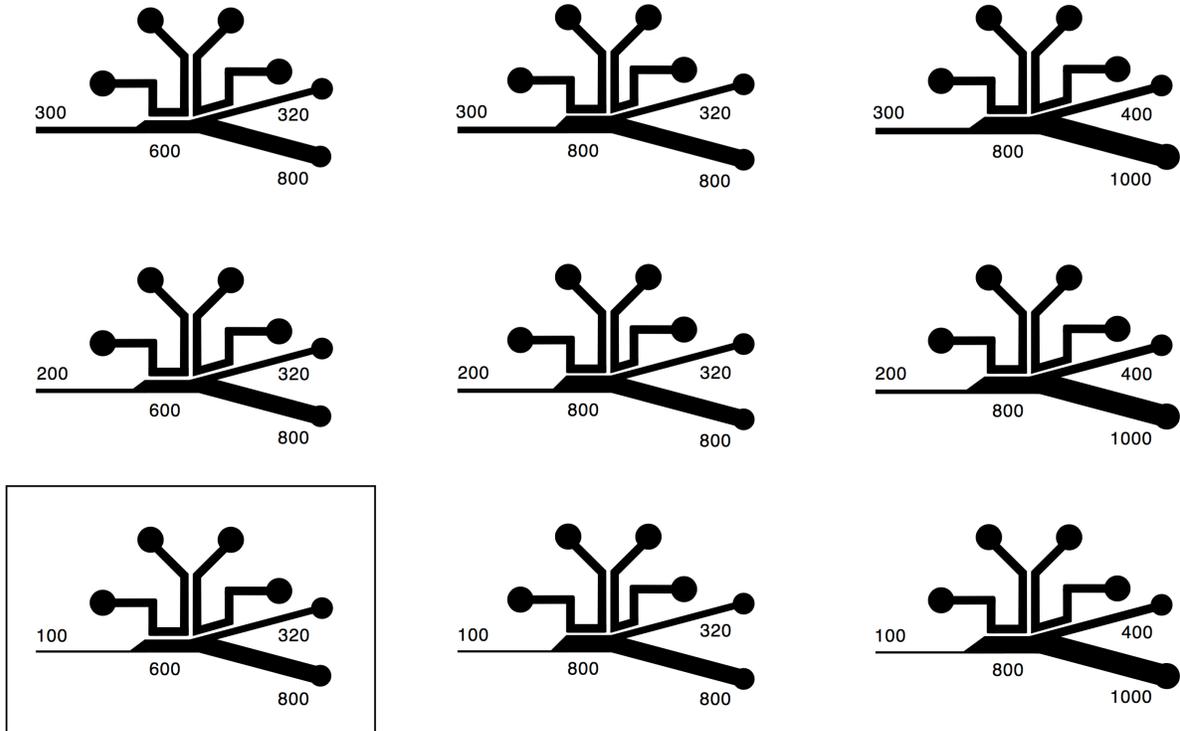
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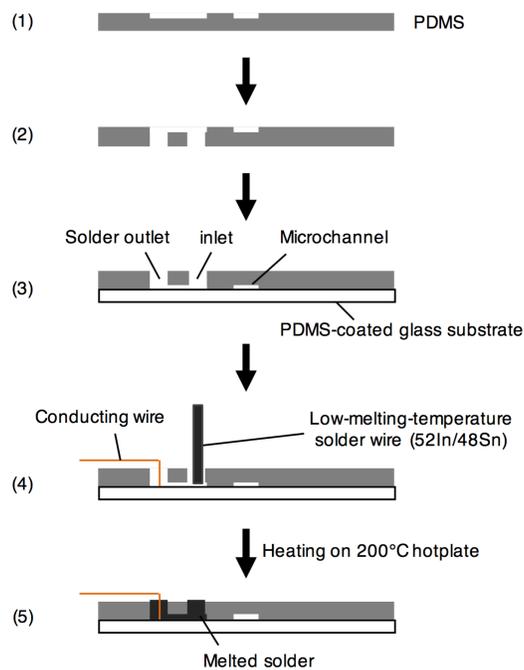
Supplementary Fig. S3 Masks for the fabrication of the droplets-sorting device.

(a) Forty patterns of devices with different channel widths [inlet, center, outlet (collect) and outlet (waste)] and with (right) or without (left) a shunt structure. (b) Nine patterns of devices. Box, optimized device.

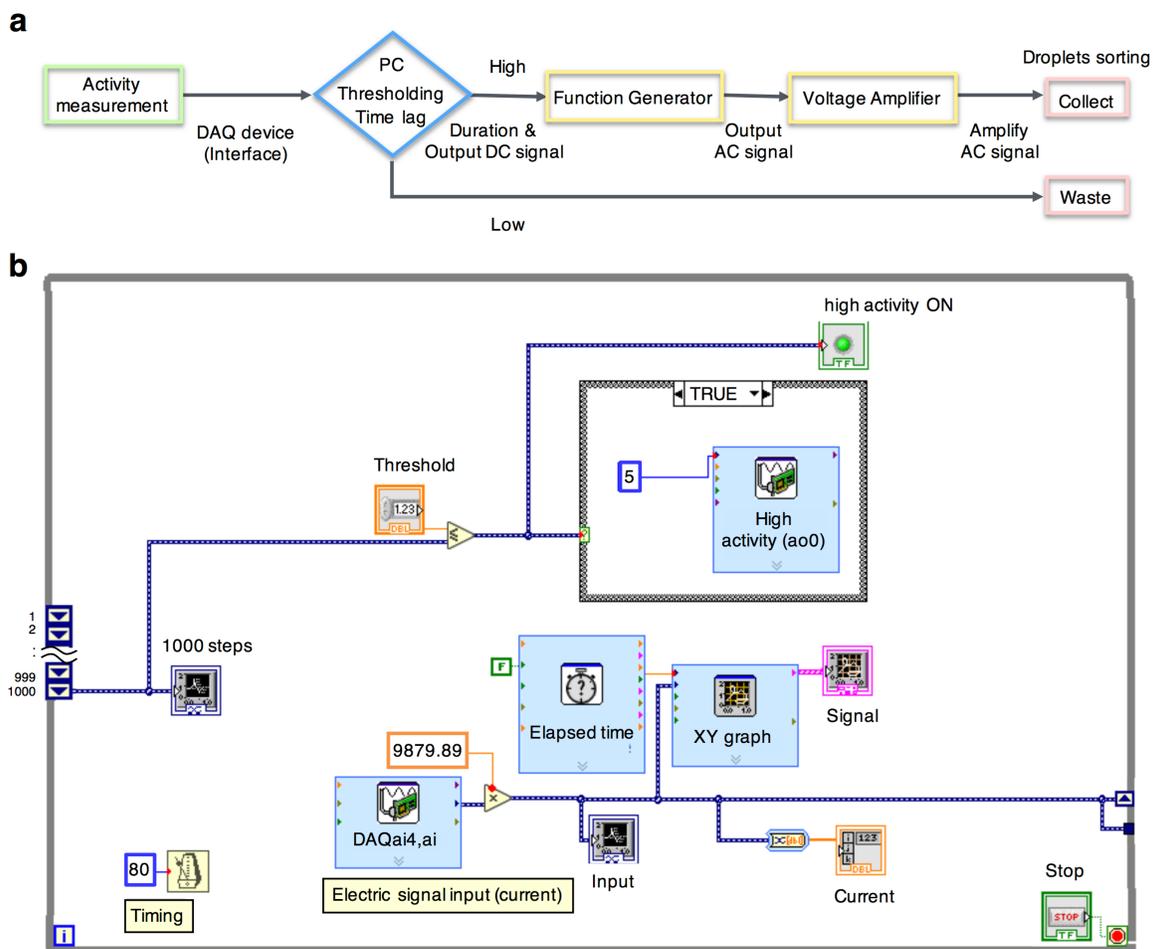
b



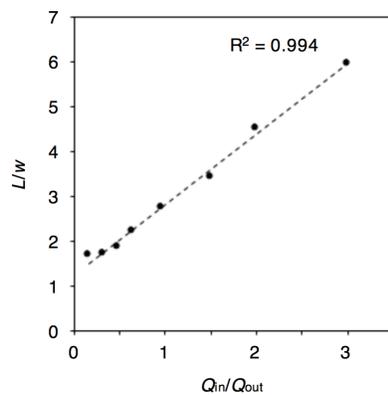
Supplementary Fig. S3 (continued)



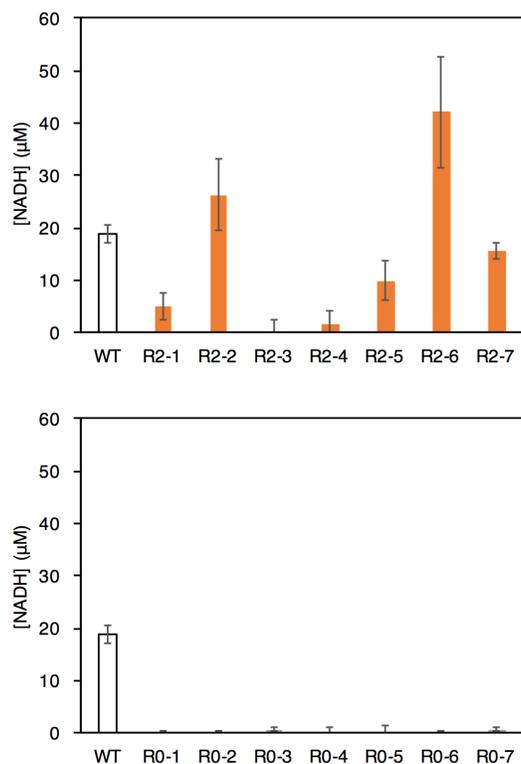
Supplementary Fig. S4 Fabrication method of the droplet-sorting device. (1) PDMS structured by using standard soft lithography. (2) Punched holes as the low-melting-temperature solder wire inlets and outlets. (3) The structured PDMS was sealed by oxygen plasma to a flat surface to enclose the channels. (4) The conducting wire was set at the outlet holes and the low-melting-temperature solder wire at the inlet hole on the 200°C hot plate. (5) After the solder wire melted, it was removed from the hot plate to harden.



Supplementary Fig. S5 Fabrication of data acquisition and control system. (a) A flow chart of our detection and sorting system. First, the continuous analog signals detected by the NAD(P)H-measuring device were converted to digital signals *via* a DAQ device. Then, each signal was compared with a threshold value and divided into high and low signals to the function generator by the LabVIEW program (b) on a PC. When only high signals were sent, the function generator output AC signal using burst oscillation. Then, signals were amplified by voltage amplifier. The amplified signals to the dielectrophoretic electrode controlled switching and duration. Finally, on the droplet-sorting device, droplets containing enzymes with higher activity were collected *via* the "collect channel." (b) A block diagram of the LabVIEW program.



Supplementary Fig. S6 Relationship between the plug size and flow rates in the encapsulating device. Dimensionless droplet length (L/w) was plotted as a function of ratios of the rates of flow of the dispersed and continuous phases (Q_{in}/Q_{out}) for the reference geometry ($h = 65 \mu\text{m}$, $w = 400 \mu\text{m}$).



Supplementary Fig. S7 Activity measurement of IDH clones by observing NADH absorbance.

Each seven clones were randomly picked from the library after two rounds of screening (top) and the initial library (bottom), respectively. These clones were translated by the PURE system and the activities of the products were measured by absorbance of NADH at 340 nm. Mean \pm SD. N = 3. WT; wild-type. Mutants; R2-1 (V358A), R2-2 (I304V), R2-3 (P300L), R2-4 (R197C, Y420C), R2-5 (K12E, A36Q), R2-6 (K17R), R2-7 (M1T, S286G), R0-1 (frameshift), R0-2 (Q162H, V275M, K360N), R0-3 (A2E, Q13R, N104D, K131R), R0-4 (E24G, E153G, G157D), R0-5 (L251F, D254G, K268R, E270G, D343G, T368A, A431P), R0-6 (nonsense), R0-7 (frameshift).