Electronic Supplementary Material (ESI) for MedChemComm. This journal is © The Royal Society of Chemistry 2019

SUPPORTING INFORMATION

Synthesis, Conformational Preferences, and Biological Activity of Conformational Analogues of the Microtubule-Stabilizing Agents, (–)-Zampanolide and (–)-Dactylolide

Jeffrey L. Henry,^a Matthew R. Wilson,^b Michael P. Mulligan, ^a Taylor R. Quinn ^a Dan L. Sackett^c and Richard E. Taylor* ^a

^aThe Warren Family Research Center for Drug Discovery and Development and the Department of Chemistry & Biochemistry, University of Notre Dame, Notre Dame, IN 46556-5670. Email: rtaylor@nd.edu ^bVertex Pharmaceuticals, 50 Northern Ave, Boston, MA 02210, USA. ^cEunice Kennedy Shriver National Institute of Child Health and Human Development, National Institutes of Health, Bethesda, MD 20892

General Experimental	2
Experimental Methods for new compounds	2
Cell Growth Inhibition Assay	13
References	13
Proton and 13C NMR for new compounds	14

General Information

Unless otherwise noted, all materials were used as received from a commercial supplier without further purification. All anhydrous reactions were performed using oven-dried or flame-dried glassware, which was then cooled under vacuum and purged with nitrogen gas. Tetrahydrofuran (THF), dichloromethane (CH_2Cl_2), toluene (PhMe), and diethyl ether (Et_2O) were filtered through activated alumina under nitrogen. Triethylamine (TEA) and N,Ndisopropylethylamine (*i*-Pr₂NEt) were distilled over CaH₂ and stored over KOH pellets. 4 Å molecular sieves were oven-dried overnight and then cooled under high vacuum prior to use. All reactions were monitored by Silicycle analytical thin layer chromatography (TLC) plates (AL SIL G/UV, glass back) and analyzed with 254 nm UV light and/or anisaldehyde - sulfuric acid or potassium permanganate treatment. Silica gel for column chromatography was purchased from Silicycle (Silica Gel 60, 230-400 mesh). Unless otherwise noted, all ¹H and ¹³C NMR spectra were recorded in CDCl₃ or DMSO d6 using either Bruker 400 spectrometers operating at 400.13 MHz for ¹H and 100.61 MHz for ¹³C, or Varian VNMRS 600 operating at 599.87 MHz for 1H and 150.84 MHz for ¹³C. Chemical shifts (δ) were reported in ppm relative to residual CHCl₃ as an internal reference (¹H: 7.26 ppm, ¹³C: 77.16 ppm). Coupling constants (J) were reported in Hertz (Hz). Peak multiplicity is indicated as follows: s (singlet), d (doublet), t (triplet), q (quartet), p (pentet), x (septet), h (heptet), b (broad), and m (multiplet). Mass spectra (FAB) were obtained at the Department of Chemistry and Biochemistry, University of Notre Dame using either a JEOL AX505HA or JEOL JMS-GCmate mass spectrometer.

Experimental Methods

OPMB Ō,,,

(S)-2-(((4-methoxybenzyl)oxy)methyl)oxirane: (4-methoxyphenyl)methanol (22 g, 20 mL, 1 eq, 0.16 mol) was slowly added to a vigorously stirred flask charged with 40% wt/wt NaOH (150 g NaOH in 230 mL of water) solution at 0 °C. 2-(chloromethyl)oxirane (60 g, 51 mL, 4 eq, 0.64 mol) was added by syringe pump over 30 min followed by tetrabutylammonium iodide (3.0 g, .05 Eq, 8.1 mmol). The solution was allowed to warm to 23 °C and stir for 3 days. The solution was diluted with water (250 mL) and Et₂O (250 mL). The aqueous layer was washed with Et₂O (100 mL x 3). The organics were dried with brine (100 mL), MgSO₄ and concentrated. The solution was purified via vacuum distillation (~.5 torr; 150° C) to afford 2-(((4-methoxybenzyl)oxy)methyl)oxirane (23 g, 74 %) as a clear oil.

In a round bottom flask charged with a stir bar and air, toluene (0.91 mL) was added to (*R*,*R*)-(–)-N,N'-Bis(3,5-di-tert-butylsalicylidene)-1,2-cyclohexanediaminocobalt(II) (193 mg, 0.0050 eq, 595 μ mol) followed by an addition of acetic acid (71.4 mg, 0.07 mL, 0.010 eq, 1.19 mmol). The solution was stirred vigorously at room temp for 1 h. The solvent was removed to afford a brown residue. At this point 2-(((4-methoxybenzyl)oxy)methyl)oxirane (23.1 g, 1 eq, 119 mmol) in 1.00 mL of THF was added to the residue and stirred at 0 °C until the brown residue was dissolved. Water (1.18 g, 1.18 mL, 0.55 eq, 65.4 mmol) was added dropwise to the solution and was allowed to slowly warm to 23° C and stirred for 16 hours. The resulting solution was concentrated *in vacuo*. The resulting residue was purified by vacuum distillation (~0.5 torr; 150° C) to afford (S)-2-(((4-methoxybenzyl)oxy)methyl)oxirane (9.70 g, 42 %) as a clear oil.

 $[α]_D^{23}$ = -3.5 (*c* 1.00, CHCl₃); ¹H NMR (400 MHz, CDCl₃): δ = 7.28 (d, J = 8.7 Hz, 2H), 6.88 (d, J = 8.7 Hz, 2H), 4.55 (d, J = 11.5 Hz, 1H), 4.49 (d, J = 11.5 Hz, 1H), 3.81 (s, 3H), 3.73 (dd, J = 11.4, 3.1 Hz, 1H), 3.41 (dd, J = 11.4, 5.9 Hz, 1H), 3.20-3.16 (m, 1H), 2.80 (dd, J = 5.0, 4.3 Hz, 1H), 2.61 (dd, J = 5.0, 2.7 Hz, 1H); ¹³C NMR (100 MHz, CDCl₃): δ = 159.4, 130.1, 129.6, 114.0, 73.1, 70.7, 55.4, 51.0, 44.5; HRMS-ESI (m/z): calcd. for C11H14NaO3 [M + Na]+: 217.0841, found 217.0853



(S)-1-((4-methoxybenzyl)oxy)pent-4-en-2-ol:

(S)-2-(((4-methoxybenzyl)oxy)methyl)oxirane (3.00 g, 1.00 eq, 15.0 mmol), copper(I) iodide (730 mg, 0.25 eq, 3.75 mmol), and THF (80.0 mL, M = 0.15) were added to a round bottom flask charged with a stir bar and Ar. The solution was cooled to -78 °C and vinylmagnesium bromide (23.0 mL, 1.50 eq, 23.0 mmol) was added in one portion, stirred for 5 min and was warmed to rt. The deep purple solution was quenched with 4:1 NH₄Cl:NH₄OH buffer (100 mL) and stirred overnight. The solution was diluted with water (200 mL) and separated. The aqueous layer was extracted with EtOAc (50 mL x 3). The organic layer was washed with brine (100 mL), dried MgSO₄ and then concentrated. The solution was purified via flash column chromatography (2:1 Hex:Et₂O) to afford alcohol **11** (3.30 g, 96%) as a yellow oil.

[α]_D²³ = +2.7 (*c* 1.00, CHCl₃); ¹H NMR (400 MHz, CDCl₃): δ = 7.26 (d, J = 8.7 Hz, 2H), 6.89 (d, J = 8.7 Hz, 2H), 5.82 (ddt, J = 17.1, 10.1, 7.1 Hz, 1H), 5.14-5.08 (m, 2H), 4.49 (s, 2H), 3.90-3.84 (m, 1H), 3.81 (s, 3H), 3.49 (dd, J = 9.5, 3.4 = Hz, 1H), 3.34 (dd, J = 9.5, 7.5 Hz, 1H), 2.32 (s, br, 1H), 2.25 (t, J = 6.8 Hz, 2H); ¹³C NMR (100 MHz, CDCl₃): δ = 159.4, 134.4, 130.2, 129.5, 117.8, 114.0, 73.7, 73.2, 69.8, 55.4, 38.0 HRMS-ESI (m/z): calcd. for C13H18NaO3 [M + Na]+: 245.1154, found 245.1148.



(S)-tert-butyl(2-(oxiran-2-yl)ethoxy)diphenylsilane: Epoxide **8** was prepared using known literature procedures.¹

 $[\alpha]_{D}^{23} = -6.3 (c \ 1.00, CHCl_3); {}^{1}H \ NMR (400 \ MHz, CDCl_3): \delta = 7.62-7.66 (m, 4H), 7.45-7.26 (m, 6H), 3.87-3.78 (m, 2H), 3.13-3.08 (m, 1H), 2.79 (dd, J = 5.0, 4.1 \ Hz, 1H), 2.52 (dd, J = 5.1, 2.8 \ Hz, 1H),$

1.80-1.75 (m, 2H), 1.06 (s, 9H); ¹³**C NMR** (100 MHz, $CDCI_3$): δ = 135.7, 135.7, 133.8, 133.8, 129.8, 127.8, 61.0, 50.3, 47.4, 35.8, 27.0, 19.3 **HRMS-ESI** (m/z): calcd. for C20H27O2Si [M + H]+: 327.1780, found 327.1775.



(R)-1-((tert-butyldiphenylsilyl)oxy)oct-7-en-3-ol: In a flame-dried 100 mL two-necked flask containing a reflux condenser and stir bar was added magnesium turnings (1.70 g, 71.1 mmol, 3.00 eq), 1-butenyl bromide (7.21 mL, 71.1 mmol, 3.00 eq), 1,2-dibromoethane (1-2 drops), and dry THF (50 mL). The resulting solution was heated gently to reflux with stirring which initiated the Grignard reaction (CAUTION: extremely exothermic). The refluxing solution became dark grey and was stirred until the magnesium turnings dissolved completely. In a separate 500 mL round bottom flask was added the known epoxide 8 (7.76 g, 23.7 mmol, 1.00 eq), dry THF (50 mL, M = 0.24), and CuI (1.13 g, 5.92 mmol, 0.25 eq). This flask was then cooled to -78 °C using a dry ice/acetone bath. Once the Grignard reaction had completed, the freshly prepared Grignard reagent was added in one portion via syringe to the cooled epoxide/THF/CuI reaction mixture giving a deep purple solution. After 5 min at -78 °C, the resulting dark purple reaction mixture was warmed to 0 °C and stirred at this temperature for 1 h. After 1 h, the reaction mixture was quenched carefully with saturated aqueous NH4Cl (80 mL) at 0 °C followed by 28% NH₄OH aqueous solution (20 mL). The reaction mixture was stirred for 15 min and then diluted with water (80 mL). The layers were separated, and the aqueous layer extracted with Et₂O (50 mL x 3). The combined organic layers were dried over MgSO₄, filtered, and concentrated in vacuo. Purification was accomplished by flash chromatography on silica gel (10:1 Hex:EtOAc) provided the corresponding alcohol (8.96 g, 23.40 mmol, 99%) as a colorless oil.

[α]_D²³ = +5.5 (*c* 1.00, CHCl₃); ¹H NMR (400 MHz, CDCl₃): δ = 7.69-7.67 (m, 4H), 7.47-7.38 (m, 6H), 5.82 (ddt, J = 17.0, 10.2, 6.7 Hz, 1H), 5.00 (d, J = 17.1 Hz, 1H), 4.95 (d, J = 10.2 Hz, 1H), 3.91-3.83 (m, 3H), 3.29 (d, J = 2.7 Hz, 1H), 2.11-2.05 (m, 2H), 1.79-1.61 (m, 2H), 1.59-1.50 (m, 2H), 1.49-1.40 (m, 2H), 1.05 (s, 9H); ¹³C NMR (100 MHz, CDCl₃): δ = 139.0, 135.7, 135.7, 133.1, 133.0, 130.0, 130.0, 127.9, 114.6, 71.9, 63.8, 38.4, 37.1, 33.9, 26.9, 25.0, 19.2 HRMS-ESI (m/z): calcd. for C24H34NaO2Si [M + Na]+: 383.2406, found 383.2419.



(*R*)-9-((tert-butyldiphenylsilyl)oxy)-7-hydroxynon-2-en-1-yl acetate: In a flame-dried twonecked flask containing a reflux condenser and stir bar was added alkene-ol (1.02 g, 2.66 mmol, 1.00 eq), (*Z*)-but-2-ene-1,4-diyl diacetate (4.57 g, 26.6 mmol, 10.0 eq) and DCM (30.0 mL, 0.10 M). The resulting solution was degassed with argon for 30 min. Grubbs II generation catalyst (113 mg, 0.13 mmol, 0.05 eq) was added in one portion, and the purple solution was heated to reflux. After 18 h, the resulting brown solution was allowed to cool to room temperature and concentrated in vacuo. Purification² by flash chromatography on silica gel (20:1 \rightarrow 10:1 \rightarrow 8:1 \rightarrow 5:1 \rightarrow 3:1 \rightarrow 2:1 Pentane: Et₂O) provided the corresponding allylic acetate (1.18 g, 2.61 mmol, 98%, 9:1 *E:Z*) as a brown oil.

[α]_D²³ = +7.9 (*c* 1.00, CHCl₃); ¹H NMR (400 MHz, CDCl₃): δ = 7.69-7.67 (m, 4H), 7.47-7.38 (m, 6H), 5.78 (td, J = 15.4 Hz, 1H), 5.58 (d, J = 15.4 Hz, 1H), 4.62 (d, J = 6.8 Hz, 0.2H), 4.51 (d, J = 6.5 Hz, 1.8H), 3.93-3.82 (m, 3H), 3.35 (s, br, 1H), 2.10-2.06 (m, 2H), 2.06 (s, 3H), 1.76-1.62 (m, 2H), 1.62-1.51 (m, 2H), 1.49-1.40 (m, 2H), 1.05 (s, 9H) ¹³C NMR (100 MHz, CDCl₃): δ = 171.0, 136.4, 135.7, 135.6, 133.1, 132.9, 130.0, 127.9, 124.1, 71.8, 65.4, 63.7, 38.4, 37.1, 32.3, 26.9, 24.9, 21.1, 19.1; HRMS-ESI (m/z): calcd. for C27H3O4Si [M + H]+: 455.2618, found 455.2512.



10

tert-butyldiphenyl(2-((2R,6S)-6-vinyltetrahydro-2H-pyran-2-yl)ethoxy)silane: In a flame-dried 25 mL cone shaped flask containing a stir bar was added freshly prepared $Pd_2(dba)_3 \cdot CHCl_3^3$ (0.05 g, 0.04 mmol, 0.02 eq), (*S,S*)-DACH-phenyl Trost ligand³ (0.091 g, 0.132 mmol, 0.06 eq), and dry DCM (15.0 mL, 0.15 M). The reaction mixture was stirred for 20 min allowing the initial purple homogenous solution to turn bright orange. In a separate flame-dried 100 mL round bottom flask containing a stir bar was added the allylic acetate (1.00 g, 2.20 mmol, 1.00 eq) and dry DCM (15.0 mL, 0.15 M) and the flask cooled to 0 °C. The catalyst solution was then added via cannula to the allylic acetate solution and the reaction mixture warmed to room temperature and stirred for 12 h. After 12 h, the reaction mixture was concentrated under reduced pressure giving an orange oil. Purification by flash chromatography on silica gel (10:1 Hex:EtOAc) provided *cis*-pyran **10** (0.82 g, 2.09 mmol, 95%, 10:1 dr) as a colorless oil.

[α]_D²³ = -16.4 (*c* 1.00, CHCl₃); ¹H NMR (400 MHz, CDCl₃): δ = 7.68-7.65 (m, 4H), 7.43-7.34 (m, 6H), 5.86 (ddd, J = 17.3, 10.6, 5.2 Hz, 1H), 5.21 (td, J = 17.4, 1.6 Hz, 1H), 5.08 (td, J = 10.6, 1.6 Hz, 1H), 3.90-3.79 (m, 2H), 3.73 (td, J = 10.0, 5.5 Hz, 1H), 3.65-3.59 (m, 1H), 1.88-1.50 (m, 6H), 1.34-1.16 (m, 2H), 1.04 (s, 9H); ¹³C NMR (100 MHz, CDCl₃): δ = 139.8, 135.7, 135.7, 134.2, 134.1, 129.6, 129.6, 127.7, 114.3, 78.2, 74.4, 60.4, 39.5, 31.6, 27.0, 23.8, 19.4, 0.14; HRMS-ESI (m/z): calcd. for C25H35O2Si+ [M + H]+: 395.2406, found 395.2401.



tert-butyl(2-((2R,6S)-6-((S,E)-5-((4-methoxybenzyl)oxy)-4-((triethylsilyl)oxy)pent-1-en-1-

yl)tetrahydro-2H-pyran-2-yl)ethoxy)diphenylsilane: To a flame-dried two-neck flask containing a stir bar and a reflux condenser, homoallylic alcohol **11** (0.56 g, 2.53 mmol, 2.00 eq), and vinyl pyran **10** (0.50 g, 1.27 mmol, 1.00 eq) was added dry DCM (12.5 mL, 0.11 M). This solution was deoxygenated with argon gas for 20 min and then Grubbs II generation catalyst (0.22 g, 0.25 mmol, 0.20 eq) was added resulting in a purple solution. The resulting reaction mixture was refluxed for 18 h. The dark brown solution was cooled to room temperature and concentrated under reduced pressure giving a brown oil. Purification of this crude oil by flash chromatography on silica gel (5:1 \rightarrow 3:1 Hex:EtOAc) provided an inseparable mixture of the homoallylic alcohol **11** and the cross metathesis product **7** as a yellow oil. The unreacted pyran starting material was re-isolated affording 60 mg (0.15 mmol, 12%) of the clear oil

The mixture of alcohols and DMF (5.0 mL, 0.30 M) were added to a flame-dried round bottom flask charged with a stir bar and argon. Imidazole (518 mg, 7.62 mmol, 6.00 eq) and TESCI (0.64 mL, 3.81 mmol, 3.00 eq) were added stirred for 18 h The brown solution was diluted with water (20 mL) and extracted with Et₂O (10 mL x 3). The organic layer was washed with brine (20 mL) and dried with MgSO₄. The solution was then concentrated under reduced pressure. The residue was purified via flash column chromatography (50:1 \rightarrow 20:1 \rightarrow 10:1 Hex:Et₂O) to afford the silyl ether (0.70 g, 0.99 mmol, 78% (90% brsm)) as a clear colorless oil over two steps.

[α]_D²³ = -11.7 (*c* 1.00, CHCl₃); ¹H NMR (400 MHz, CDCl₃): δ = 7.67-7.65 (m, 4H), 7.43-7.34 (m, 6H), 7.24 (d, J = 8.6 Hz, 2H), 6.86 (d, J = 8.6 Hz, 2H), 5.63 (td, J = 15.7, 6.7 Hz, 1H), 5.52 (dd, J = 15.7, 5.5 Hz, 1H), 4.43 (s, 2H), 3.87-3.80 (m, 2H), 3.80 (s, 3H), 3.77-3.69 (m, 2H), 3.60-3.55 (m, 1H), 3.34 (d, J = 5.3 Hz, 2H), 2.30 (td, J = 14.0, 6.7 Hz, 1H), 2.20 (td, J = 14.0, 6.7 Hz, 1H), 1.84-1.75 (m, 2H), 1.73-1.64 (m, 2H), 1.56-1.46 (m, 2H), 1.29-1.16 (m, 2H), 1.04 (s, 9H), 0.93 (t, J = 7.9 Hz, 9H), 0.58 (q, J = 7.9, 6H); ¹³C NMR (100 MHz, CDCl₃): δ = 159.2, 135.7, 135.7, 134.3, 134.2, 134.1, 130.7, 129.6, 129.4, 129.3, 127.7, 127.2, 113.8, 78.0, 74.5, 74.1, 73.1, 71.5, 60.5, 55.4, 39.5, 38.1, 31.7, 31.6, 27.0, 23.8, 19.4, 7.0, 5.1; HRMS-ESI (m/z): calcd. for C42H63O5Si2 [M + H]+: 703.4214, found 703.4209.



(*S,E*)-5-((2*S*,6*R*)-6-(2-((*tert*-butyldiphenylsilyl)oxy)ethyl)tetrahydro-2*H*-pyran-2-yl)-1-((4methoxybenzyl)oxy)pent-4-en-2-ol: The silyl ether (0.62 g, 0.88 mmol, 1.00 eq) and MeCN:H₂O (2.81 mL, 8:1) were added to a round bottom flask charged with a stir bar and argon. The flask was cooled to 0 °C and TFA (0.31 mL) was added and stirred for 30 min. The cloudy solution was carefully quenched with NaHCO₃ (sat) (until the pH =7-8 by litmus paper; ~10 mL) and EtOAc (20 mL). The aqueous layer was extracted with EtOAc (20 mL x 3). The organic layer was washed with brine (20 mL), dried MgSO₄, and concentrated in vacuo. The solution was purified via flash column chromatography (5:1 \rightarrow 3:1 Hex:EtOAc) to afford alcohol **7** (485 mg, 0.82 mmol, 93%) as a clear colorless oil.

[α]_D²³ = -12.3 (*c* 1.00, CHCl₃); ¹H NMR (400 MHz, CDCl₃): δ = 7.68-7.64 (m, 4H), 7.42-7.34 (m, 6H), 7.25 (d, J = 8.7 Hz, 2H), 6.88 (d, J = 8.7 Hz, 2H), 5.63 (td, J = 15.7, 6.7 Hz, 1H), 5.56 (dd, J = 15.7, 4.9Hz, 1H), 4.46 (s, 2H), 3.87-3.82 (m, 2H), 3.80 (s, 3H), 3.77-3.75 (m, 1H), 3.72 (td, J = 10.0, 5.7 Hz, 1H), 3.61-3.55 (m, 1H), 3.47 (dd, J = 9.5, 3.4 Hz, 1H), 3.33 (dd, J = 9.5, 7.4 Hz, 1H), 2.30-2.17 (m, 2H), 1.84-1.75 (m, 2H), 1.73-1.65 (m, 2H), 1.61-1.51 (m, 2H), 1.32-1.14 (m, 2H), 1.04 (s, 9H); ¹³C NMR (100 MHz, CDCl₃): δ = 159.4, 135.7, 135.7, 134.9, 134.2, 134.1, 130.2, 129.6, 129.6, 129.5, 129.5, 127.7, 126.2, 114.0, 77.8, 74.5, 73.7, 73.1, 70.0, 60.4, 55.4, 39.5, 36.7, 31.8, 31.5, 27.0, 23.7, 19.4; HRMS-ESI (m/z): calcd. for C36H49O5Si [M + H]+: 589.3349, found 589.3344.



(2E,4Z)-7-((tert-butyldimethylsilyl)oxy)-8-(diethoxyphosphoryl)-5-methylocta-2,4-dienoic acid: The acid prepared using the known procedure reported by Altmann and coworkers.¹

¹H NMR (400 MHz, CDCl₃): δ = 7.68 (dd, J = 15.2, 11.7 Hz, 1H), 6.11 (d, J = 11.7 Hz, 1H), 5.77 (d, 15.2, 1H), 4.18-4.08 (m, 5H), 2.69 (dd, J = 13.4, 8.4, 1H), 2.54 (dd, J = 13.4, 8.4, 1H), 2.09-1.99 (m, 2H), 1.93 (s, 3H), 1.33 (dt, J = 7.1, 2.8 Hz, 6H), 0.84 (s, 9H), 0.05 (s, 3H), -0.02 (s, 3H); ¹³C NMR (100 MHz, CDCl₃): δ =171.5, 148.0, 143.0, 126.5, 118.8, 67.4, 6.19 (dd, J = 6.5, 1.3 Hz), 41.7 (d, J = 3.6 Hz), 35.2 (d, J = 134.7 Hz), 27.1, 25.9, 25.7, 18.0, 16.6 (d, J = 6.1 Hz), -4.6, -4.7; HRMS-ESI (m/z): calcd. for C19H38O6PSi [M + H]+: 421.2175, found 421.2170.



(*S*,*E*)-5-((2*S*,6*R*)-6-(2-((*tert*-butyldiphenylsilyl)oxy)ethyl)tetrahydro-2*H*-pyran-2-yl)-1-((4-methoxybenzyl)oxy)pent-4-en-2-yl-(2*E*,4*Z*)-7-((*tert*-butyldimethylsilyl)oxy)-8-

(diethoxyphosphoryl)-5-methylocta-2,4-dienoate: A solution of acid 6 (1.24 g, 3.00 mmol, 1.30 eq) and NEt₃ (0.95 ml, 0.39 mmol, 3.00 eq) in toluene (11.0 mL, 0.10 M) was added to a flamedried round-bottom flask with a stir bar under Ar. 2,4,6-trichlorobenzoyl chloride (0.64 ml, 4.10 mmol, 1.80 eq) was subsequently added at room temperature, producing a cloudy pale yellow solution and was stirred for 1 h; then a solution of alcohol **7** (1.34 g, 2.30 mmol, 1.00 eq) and DMAP (278 mg, 2.30 mmol, 1.00 eq) in toluene (9.00 ml (the mixture was sonicated to produce a clear solution); plus 2 x 0.5 ml from rinsing)) was added, resulting in the immediate formation of an off-white suspension. After 1 h of stirring, NaHCO₃ (sat) (10 mL) and EtOAc (20 mL) were added and the phases were separated. The aqueous layer was extracted with EtOAc (20 mL x 3), and the combined organic extracts were washed with brine (20 mL), dried over MgSO₄ and concentrated under reduced pressure. Purification of the residue by flash chromatography (1:3 \rightarrow 1:1 EtOAc:Hex) afforded ester **12** (1.77 g, 1.80 mmol, 79%) as a clear yellow oil.

[α]_D²³ = -9.2 (*c* 1.00, CHCl₃); ¹H NMR (400 MHz, CDCl₃): δ = 7.67-7.64 (m, 4H), 7.59 (dd, J = 15.1, 11.7 Hz, 1H), 7.41-7.34 (m, 6H), 7.23 (d, J = 8.4 Hz, 2H), 6.85 (d, J = 8.0 Hz, 2H), 6.06 (d, J = 11.4 Hz, 1H), 5.80 (d, J = 15.1 Hz, 1H), 5.54 (m, 2H), 5.10-5.05 (m, 1H), 4.49 (d, J = 11.7, 1H), 4.41 (dd, J = 11.8, 2.9 Hz, 1H), 4.18-4.07 (m, 5H), 3.86-3.80 (m, 1H), 3.79 (s, 3H), 3.74-3.68 (m, 2H), 3.59-3.53 (m, 1H), 3.51 (d, J = 5.8 Hz, 2H), 2.64-2.53 (m, 2H), 2.41-2.39 (m, 2H), 2.01 (dd, J = 18.6, 6.2 Hz, 2H), 1.89 (s, 3H), 1.83-1.75 (m, 2H), 1.72-1.64 (m, 1H), 1.56-1.52 (m, 1H), 1.33 (dt, J = 7.1, 3.2, 6H), 1.28 (m, 2H), 1.03 (s, 9H), 0.83 (s, 9H), 0.05 (s, 3H), -0.03 (s, 3H); ¹³C NMR (100 MHz, CDCl₃): (There were more C13 peaks than expected due to the diastereotopic nature of the compound) δ = 167.0, 159.3, 146.3 146.3, 141.5, 141.5, 135.7, 135.7, 135.1, 135.1, 134.2, 134.1, 130.3, 129.6, 129.5, 129.4, 127.7, 126.8, 125.4, 125.4, 119.7, 119.7, 113.9, 77.8, 74.5, 72.9, 72.1 (d, J = 5.1 Hz), 70.2, 66.9, 61.7 (dd, J = 6.5, 2.1 Hz), 60.4, 55.4, 41.8 (d, J = 4.7 Hz), 39.5, 35.9, 34.5, 34.1, 31.7, 31.5, 27.0, 25.9, 25.2,23.7, 19.4, 17.9, 16.6, 16.6, -4.7; HRMS-ESI (m/z): calcd. for C55H84010PSi2 [M + H]+: 991.5341, found 991.5335.



(*S*,*E*)-5-((2*S*,6*R*)-6-(2-hydroxyethyl)tetrahydro-2*H*-pyran-2-yl)-1-((4-methoxybenzyl)oxy)pent-4-en-2-yl (2*E*,4*Z*)-8-(diethoxyphosphoryl)-7-hydroxy-5-methylocta-2,4-dienoate: Ester 12 (531 mg, 0.54 mmol, 1.00 eq) in THF (22.0 mL) was prepared in an oven-dried PTFE round-bottom flask equipped with a stir bar. 70% HF•py (5.30 mL) was carefully added at 0 °C. The mixture was warmed to room temperature after 5 min and continued stirring for 6 h. The solution was carefully pulled into a syringe and added to a vigorously stirred mixture of excess NaHCO₃ (sat) (200 mL) and EtOAc (200 mL); after ensuring the pH of the aqueous phase was neutral (pH = 7-8; litmus paper) the phases were separated. The aqueous phase was extracted with EtOAc (50 mL x 3). The combined organic extracts were washed with brine (200 mL), dried over MgSO₄, and concentrated under reduced pressure. The residue was purified by flash chromatography (1:0 -> 1:1 EtOAc:Acetone) to afford the diol (307 mg, 0.48 mmol, 92%) as a colorless, viscous oil.

[**α**]_D²³ = -14.6 (*c* 1.00, CHCl₃); ¹**H** NMR (400 MHz, CDCl₃): δ = 7.55-7.49 (m, 1H), 7.21 (d, J = 7.7 Hz, 2H), 6.84 (d, J = 8.6 Hz 2H), 6.10 (d, J = 11.7 Hz, 1H), 5.80 (d, J = 15.0 Hz, 1H), 5.55-5.46 (m, 2H), 5.08-5.03 (m, 1H), 4.47 (dd, J = 11.7, 1.7 Hz, 1H), 4.40 (dd, J = 11.7, 2.3 Hz, 1H), 4.17-4.14 (m, 1H), 4.13-4.05 (m, 4H), 3.77 (s, 3H), 3.73-3.71 (m, 2H), 3.63 (s, br, 1H), 3.56-3.53 (m, 1H), 3.51-3.46 (m, 2H), 3.01 (s, br, 1H), 2.62-2.56 (m, 1H), 2.46-2.39 (m, 1H), 2.38-2.28 (m, 2H), 1.93-1.88 (m, 5H), 1.80-1.78 (m, 1H), 1.75-1.69 (m, 1H), 1.65-1.62 (m, 1H), 1.53-1.45 (m, 3H), 1.30 (t, J = 7.1 Hz, 6H), 1.27-1.22 (m, 2H); ¹³C NMR (100 MHz, CDCl₃): (There were more C13 peaks than expected due to the diastereotopic nature of the compound) δ = 167.0, 167.0, 159.2, 145.6, 145.6, 140.5, 140.4, 134.7, 134.6, 130.2, 129.4, 126.7, 126.7, 125.8, 125.7, 120.1, 120.1, 113.8, 78.1, 78.0, 76.9, 72.8, 71.9, 70.2, 70.1, 65.3 (d, J = 4.6 Hz), 65.2 (d, J = 4.6 Hz), 62.1 (d, J = 6.3 Hz) 61.9 (d, J = 6.6 Hz), 61.3, 55.3, 41.3 (d, J = 4.7 Hz), 41.1 (d, J = 4.9 Hz), 38.2, 34.1, 34.0, 33.5, (d, J = 138.6 Hz), 33.4, (d, J = 138.6 Hz), 31.5, 31.3, 25.0, 24.9, 23.3, 16.5, (d, J = 2.4 Hz), 16.4, (d, J = 2.4 Hz); 16.4, (d, J



(*S*,*E*)-1-((4-methoxybenzyl)oxy)-5-((2*S*,6*R*)-6-(2-oxoethyl)tetrahydro-2*H*-pyran-2-yl)pent-4-en-2-yl (2*E*,4*Z*)-8-(diethoxyphosphoryl)-5-methyl-7-oxoocta-2,4-dienoate: The diol (261 mg, 0.41 mmol, 1.00 equiv) was charged in a flame-dried round bottom flask and dissolved in DCM (4.10 mL, M = 0.10). DMP (1.04 g, 0.41 mmol, 6.00 eq, addition in two equal portions, second addition after 30 min) was added at rt. After 2 h EtOAc (10 mL) and a mixture of saturated aqueous NaHCO₃ (10 mL) and saturated aqueous Na₂S₂O₃ (10 mL) were added then stirring was continued for 1 h. When two clear phases had formed, the phases were separated. The aqueous phase was extracted with EtOAc (5 mL x 3), and the combined organic extracts were dried over MgSO₄. Concentration of the solution under reduced pressure and purification of the residue by flash chromatography (EtOAc, 1% AcOH to deactivate the stationary phase) gave the β -ketophosphonate ester (216 mg, 0.34 mmol, 83%) as a pale-yellow oil.

[α]_D²³ = -7.4 (*c* 1.00, CHCl₃); ¹H NMR (500 MHz, CDCl₃): δ = 9.76 (s, 1H), 7.38 (dd, J = 14.9, 11.7 Hz, 1H), 7.21 (d, J = 8.4, 2H), 6.85 (d, J = 8.5 Hz, 2H), 6.19 (d, J = 11.5 Hz, 1H), 5.85 (d, J = 15.0 Hz, 1H), 5.54-5.50 (m, 2H), 5.10-5.04 (m, 1H), 4.49-4.39 (m, 2H), 4.17-4.13 (m, 4H), 3.87-3.85 (m, 1H), 3.78 (s, 4H), 3.62 (s, 2H), 3.49-3.48 (m, 2H), 3.13 (s, 1H), 3.08 (s, 1H), 2.59 (ddd, J = 16.3, 7.5, 2.1 Hz, 1H), 2.45 (ddd, J = 16.3, 5.0, 1.6 Hz, 1H), 2.41-2.32 (m, 2H), 1.93-1.82 (m, 4H), 1.61-1.53 (m, 3H), 1.34-1.22 (m, 8H); ¹³C NMR (125 MHz, CDCl₃): δ = 201.7, 198.3, 166.8, 159.3, 140.9, 139.8, 134.5, 130.2, 129.4, 127.5, 125.8, 121.1, 113.9, 78.1, 73.0, 72.9, 72.1, 70.1, 62.9

(d, J = 6.4 Hz), 55.3, 50.1, 47.6, 42.7 (d, J = 127 Hz), 34.0, 31.3, 31.2, 25.0, 23.3, 16.4 (d, J = 6.1 Hz); **HRMS-ESI** (m/z): calcd. for C33H47NaO10P [M + Na]+: 657.2805, found 657.2791.



(1S,2E,5S,8E,10Z,14E,17R)-5-(((4-methoxybenzyl)oxy)methyl)-11-methyl-6,21-

dioxabicyclo[15.3.1]henicosa-2,8,10,14-tetraene-7,13-dione: β -ketophosphonate ester (216 mg, 0.34 mmol, 1.00 eq) in THF/H₂O (40:1, 113 ml) was prepared in a round bottom flask with a stir bar. Freshly activated Ba(OH)₂•8H₂O (140 mg, 0.44 mmol, 1.30 eq) was added at 0 °C. The orange solution was stirred for 30 min and warmed to room temperature. The solution was stirred for 4 h until it was quenched by adding EtOAc (100mL) followed by washing the organic layer with NaHCO₃ (50 mL) and then with brine (50 mL). The clear organic phase was dried over MgSO₄ and concentrated under reduced pressure. The resulting yellow oil was purified by flash chromatography (3:1 Hex:EtOAc) to afford the PMB-protected ether (80.0 mg, 0.17 mmol, 49%) as a colorless oil.

[α]_D²³ = -99.6 (*c* 1.00, CHCl₃); ¹H NMR (400 MHz, CDCl₃): δ = 7.60 (dd, J = 15.0, 11.6 Hz, 1H), 7.24 (d, J = 8.7 Hz, 2H), 6.89-6.82 (m, 3H), 6.10 (d, J = 11.6 Hz, 1H), 5.96 (d, J = 16.2 Hz, 1H), 5.89 (d, J = 15.0 Hz, 1H), 5.58 (ddd, J = 15.1, 9.5, 5.4 Hz, 1H), 5.42 (dd, J = 15.1, 6.4 Hz, 1H), 5.27-5.21 (m, 1H), 4.52-4.45 (m, 2H), 3.91 (d, J = 14.0 Hz, 1H), 3.80 (s, 3H), 3.74 (dd, J = 10.2, 6.6 Hz, 1H), 3.58 (dd, J = 10.2, 5.8 Hz, 1H), 3.51 (dd, J = 10.2, 5.2 Hz, 1H), 3.38 (ddd, J = 8.8, 8.8, 2.3 Hz, 1H), 3.23 (d, J = 14.0 Hz, 1H), 2.42-2.37 (m, 1H), 2.34-2.20 (m, 3H), 1.85-1.81 (m, 4H), 1.53-1.48 (m, 2H), 1.26-1.20 (m, 3H); ¹³C NMR (100 MHz, CDCl₃): δ = 198.0, 166.9, 159.3, 146.6, 142.8, 139.4, 134.7, 131.4,130.2, 129.4, 126.0, 125.7, 121.3, 113.9, 78.1, 75.8, 73.0, 71.4, 70.9, 55.4, 44.9, 40.3, 35.4, 32.3, 31.6, 24.0, 23.5; HRMS-ESI (m/z): calcd. for C29H37O6 [M + H]+: 481.2590, found 481.2585.



(1S,2E,5S,8E,10Z,14E,17R)-5-(hydroxymethyl)-11-methyl-6,21-dioxabicyclo[15.3.1]henicosa-

2,8,10,14-tetraene-7,13-dione: PMB protected alcohol (118 mg, 0.25 mmol, 1.00 equiv) was dissolved in DCM:H₂O (5:1, 4.80 mL) buffer in a round bottom flask charged with a stir bar. DDQ (167 mg, 0.74 mmol, 3.00 equiv) was added at 0 °C, and the mixture was vigorously stirred for 4

h; the resulting reddish pink solution was quenched with NaHCO₃ (sat) (10 mL) and DCM (20 mL). The aqueous phase was extracted with DCM (10 mL x 3), and the combined organic extracts were washed with brine (20 mL), dried over MgSO₄ and concentrated under reduced pressure. Purification of the residue by flash chromatography (3:1 \rightarrow 1:1 Hex:EtOAc) delivered alcohol **3** (59.0 mg, 0.16 mmol, 67%) as a colorless amorphous solid.

 $[α]_D^{23} = -93.4$ (*c* 1.00, CHCl₃); ¹H NMR (400 MHz, CDCl₃): δ = 7.59 (dd, J = 15.0, 11.6 Hz, 1H), 6.86 (ddd, J = 16.1, 8.0, 8.0 Hz, 1H), 6.11 (d, J = 11.5 Hz, 1H), 5.98 (d, J = 16.2 Hz, 1H), 5.88 (d, J = 15.0 Hz, 1H), 5.62 (ddd, J = 15.1, 9.1, 5.6 Hz, 1H), 5.44 (dd, J = 15.3, 6.2 Hz, 1H), 5.12-5.07 (m, 1H), 3.85 (d, J = 14.1 Hz, 1H), 3.77-3.72 (m, 3H), 3.41-3.37 (m, 1H), 3.28 (d, J = 14.1 Hz, 1H), 2.38-2.24 (m, 5H), 1.88-1.82 (m, 4H), 1.53-1.48 (m, 2H), 1.26-1.17 (m, 3H); ¹³C NMR (100 MHz, CDCl₃): δ = 197.8, 167.4, 146.5, 143.2, 139.7, 134.7, 131.3, 125.7, 125.6, 120.9, 77.9, 75.8, 74.4, 65.0, 44.9, 40.2, 34.8, 32.2, 31.5, 24.1, 23.4; HRMS-ESI (m/z): calcd. for C21H29O5 [M + H]+: 361.2015, found 361.2010.



(15,2E,5S,8E,10Z,14E,17R)-11-methyl-7,13-dioxo-6,21-dioxabicyclo[15.3.1]henicosa-2,8,10,14tetraene-5-carbaldehyde: A solution of alcohol 3 (10.2 mg, 0.028 mmol, 1.00 eq) was dissolved in DCM (0.50 ml) in a flame-dried round bottom flask. DMP (48.0 mg, 0.11 mmol, 3.00 eq) added in 2 equal portions in 20 min intervals) at room temperature. Stirring was continued for 60 min then a mixture of NaHCO₃ (sat) (5 mL) and Na₂S₂O₃ (sat) (5 mL) was added and stirring was continued for 1 h until two clear phases were formed. The phases were separated and then the aqueous phase was extracted with DCM (5 mL x 3), the combined organic phases were dried over MgSO₄, concentrated under reduced pressure. The residue was purified using flash chromatography (3:1 Hex:EtOAc) to afford aldehyde **4** (6.80 mg, 0.019 mmol, 66%) as a colorless solid.

[α]_D²³ = -85.6 (*c* 1.00, CHCl₃); ¹H NMR (400 MHz, CDCl₃): δ = 9.57 (d, J = 0.9 Hz, 1H), 7.55 (dd, J = 15.1, 11.7 Hz, 1H), 6.91 (ddd, J = 15.9, 8.0, 7.3 Hz, 1H), 6.18 (d, J = 11.6 Hz, 1H), 6.10 (d, J = 15.9 Hz, 1H), 5.87 (d, J = 15.1 Hz, 1H), 5.66 (ddd, J = 15.4, 7.5, 7.5 Hz, 1H), 5.54 (dd, J = 15.4, 5.2 Hz, 1H), 4.98 (dd, J = 9.9, 3.2 Hz, 1H), 3.77 (dd, J = 11.0, 4.8 Hz, 1H), 3.67 (d, J = 15.5 Hz, 1H), 3.52-3.45 (m, 2H), 2.60 (d, J = 14.7 Hz, 1H), 2.47-2.28 (m, 3H), 1.91-1.82 (m, 4H), 1.56-1.49 (m, 2H), 1.31-1.17 (m, 3H); ¹³C NMR (100 MHz, CDCl₃): δ = 198.8, 196.9, 166.4, 146.1, 144.1, 140.7, 135.7, 131.2, 125.8, 123.7, 119.6, 78.0, 75.9, 44.8, 39.8, 32.6, 31.9, 31.4, 29.8, 25.1, 23.5; HRMS-ESI (m/z): calcd. for C21H27O5 [M + H]+: 359.1858, found 359.1853.

¹**H NMR** (600 MHz, (CD₃)₃SO) δ 9.50 (s, 1H), 7.35 (dd, J = 15.2, 11.6 Hz, 1H), 6.86 (dt, J = 15.2, 7.3 Hz, 1H), 6.26 (d, J = 11.5 Hz, 1H), 6.12 (d, J = 15.9 Hz, 1H), 5.90 (d, J = 15.2 Hz, 1H), 5.63 (dtd, J = 15.3, 7.6, 6.0, 1.4 Hz, 1H), 5.54 (dd, J = 15.5, 4.8 Hz, 1H), 4.99 (dd, J = 9.5, 3.7 Hz, 1H), 3.75 (dd, J = 10.4, 4.8 Hz, 1H), 3.68 (d, J = 16.6 Hz, 1H), 3.57 (d, J = 16.6 Hz, 1H), 3.48 (ddt, J = 11.3, 8.8, 2.7 Hz, 1H), 2.55 (dt, J = 15.0, 6.1, 4.8 Hz, 1H), 2.47 - 2.36 (m, 2H), 2.23 (dt, J = 14.9, 8.2 Hz, 1H), 1.87 (s, 3H), 1.81 - 1.69 (m, 2H), 1.51 (m, 4H).



(2Z,4E)-N-((S/R)-hydroxy((1S,2E,5S,8E,10Z,14E,17R)-11-methyl-7,13-dioxo-6,21-

dioxabicyclo[15.3.1]henicosa-2,8,10,14-tetraen-5-yl)methyl)hexa-2,4-dienamide: Amide 13 (14 mg, 0.1 mmol, 4.6 eq) was dissolved in THF (0.50 mL) and DIBAL-H (1M in DCM, 0.10 mL, 0.1 mmol, 3.8 eq) was added. Stirring was continued at room temperature for 45 min. After that time a solution of aldehyde 4 (9.6 mg, 0.03 mmol, 1.0 eq) in THF (0.5 mL, flask rinsed once with 0.5 mL of THF) was added. Stirring continued for a total of 3 h was quenched Rochelle salt (sat) (1 mL) was added as well as EtOAc (1 mL) and stirring was continued for 15 min then brine (1mL) was added followed by separation of phases and extraction with EtOAc (3 mL x 3). The combined organic phases were dried over MgSO₄, and the residue was purified using flash chromatography on deactivated stationary phase (3:1 \rightarrow 1:1 \rightarrow 1:2 Hex:EtOAc, 2% NEt₃) giving hemiaminal 5 (5.8 mg, 0.01 mmol, 49%) as a 1.5:1.0 mixture of both C20 epimers as a white solid.

¹**H NMR** (400 MHz, (CD₃)₃SO): δ = 8.36 (d, J = 9.1 Hz, 1H, isomer 1), 8.28 (d, J = 9.0 Hz, 1H, isomer 2), 7.51-7.41 (m, 2H, isomer 1+2), 6.81-6.72 (m, 1H, isomer 1+2), 6.42- 6.33 (m, 1H, isomer 1+2), 6.24-6.18 (m, 1H, isomer 1+2), 6.11 (d, J = 5.0 Hz, 1H, isomer 2), 6.02-5.92 (m, 3H), 5.88 (d, J = 15.0 Hz, 1H, isomer 2), 5.63 (d, J = 11.4 Hz, 1H, isomer 1+2), 5.59-5.52 (m, 1H, isomer 2), 5.51-5.42 (m, 1H, isomer 1), 5.40-5.31 (m, 2H), 4.93-4.88 (m, 1H, isomer 1), 4.86-4.82 (m, 1H, isomer 2), 3.90 (d, J = 14.6, 1H, isomer 1), 3.80 (d, J = 14.7 Hz, 1H, isomer 2), 3.68 (dd, J = 11.2, 7.0 Hz, 1H, isomer 1+2), 3.33-3.31 (m, 1H, isomer 2), 3.18 (d, J = 14.6 Hz, 1H, isomer 1), 2.45-2.42 (m, 1H, isomer 1+2), 2.34-2.12 (m, 4H),1.80-1.71 (m, 9H), 1.50-1.42 (m, 4H), 1.24-1.23 (m, 3H), 1.14-1.09 (m, 4H); ¹³**C NMR** (100 MHz, (CD₃)₃SO): δ = 197.2, 197.1, 165.9, 165.6, 165.3, 165.1, 146.2, 146.1, 143.1, 142.8, 140.9, 140.7, 139.7, 139.0, 137.5, 137.2, 134.5, 134.4, 130.7, 128.6, 125.4, 125.2, 125.1, 121.1, 120.5, 119.2, 119.0, 77.2, 77.0, 75.1, 75.1, 73.5, 73.4, 72.5, 72.4, 45.7, 44.7, 33.3, 33.2, 31.8, 31.8, 31.1, 31.1, 24.0, 23.9, 22.9, 22.8, 18.4, 18.4; **HRMS-ESI** (m/z): calcd. for C27H35NNaO6 [M + Na]+: 492.2362, found 492.2357.

Cell Growth Inhibition by Compounds

PC3, human prostate cancer, and A549, human lung cancer cell lines were obtained from the NCI anticancer drug screen. A2780(1A9) cells, human ovarian cancer, were the kind gift of Dr. Tito Fojo, Medical Oncology Branch, NCI, NIH. All cells were maintained in DMEM medium supplemented with 10% fetal bovine serum. Growth inhibition was determined using the Cell-Titer Assay Reagent (Promega), following exposure to serial dilutions of each compound for 3 days prior to assay and GI(50) determinations as described.⁴ Each compound was tested with an initial 10-dose dilution series, followed by narrower, replicate 6-dose series, each of which was a separate experiment. The results presented are mean GI(50) values based on multiple determinations, the SEM, and the number of replicates, *n*. Taxol was obtained through Millipore-Sigma and dactylolide and zampanolide were obtain via our previous total syntheses of the compounds.

ID	PC3			A549			1A9		
							(A2780/1A9)		
	GI(50)	SEM	n	GI(50)	SEM	n	GI(50)	SEM	n
Taxol	4.6	2	4	6	1.5	4	5.2	2	3
Zampanolide	3.2	0.4	3	5.2	2	4	2.1	1	3
Dactylolide	460	200	4	950	140	4	620	170	3
3	>30000		4	>30000		4	>30000		3
4	>30000		4	>30000		4	>30000		3
5	56	20	4	180	80	4	120	35	3

References:

- 1.) Zurwerra, D.; Gertsch, J.; Altmann, K-H. Org. Lett., 2010, 12, 2302–2305;
- 2.) The purification of the cross-metathesis product and (*Z*)-but-2-ene-1,4-diyl diacetate could be achieved by tedious column chromatography. However, when the reation was conducted on a large scale, the (*Z*)-but-2-ene-1,4-diyl could be removed via vacuum distillation to remove the coupling partner and then chromatographed on silica.
- 3.) The reaction could be conducted at 2 mol% palladium catalyst and 6 mol% ligand, but the reaction was particularly unreliable. Increasing the reaction to 10 mol% palladium catalyst and 30 mol% catalyst gave the most reproducible results.
- 4.) K. Präbst, H. Engelhardt, S. Ringgeler, H. Hübner, Methods Mol. Biol. 1601, 1-17 (2017).



































































