

Supporting Information for

**FgaPT2, a biocatalytic tool for alkyl-diversification of indole natural products**

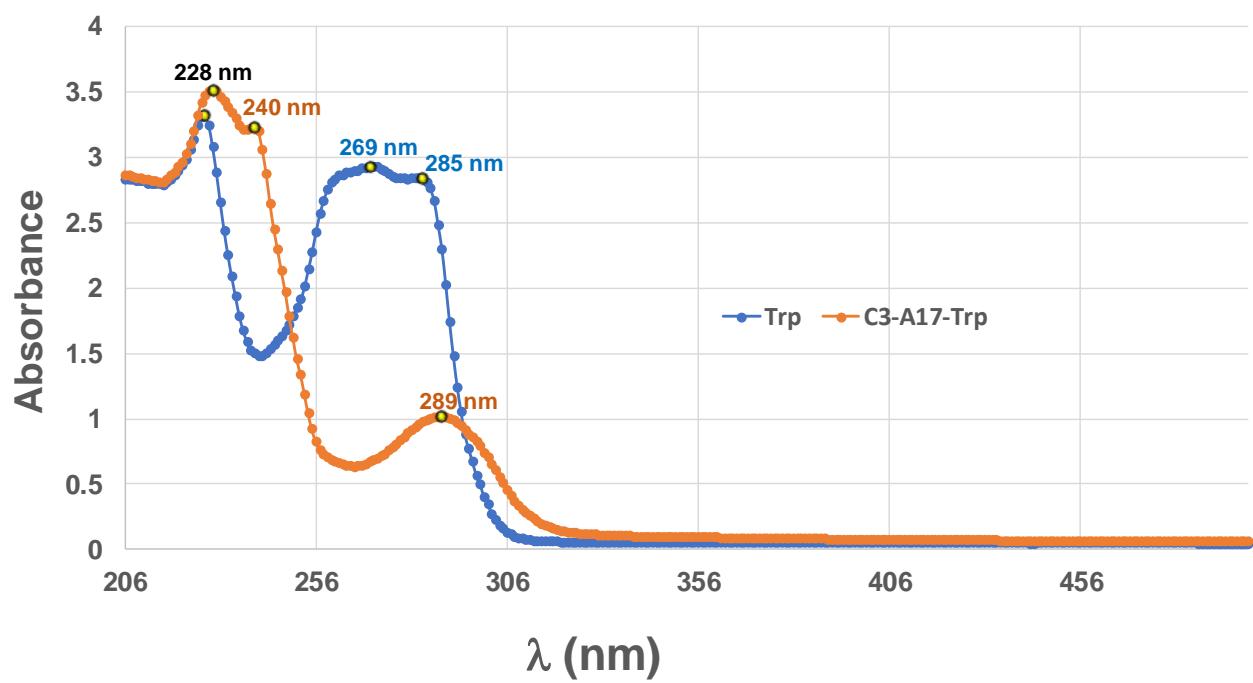
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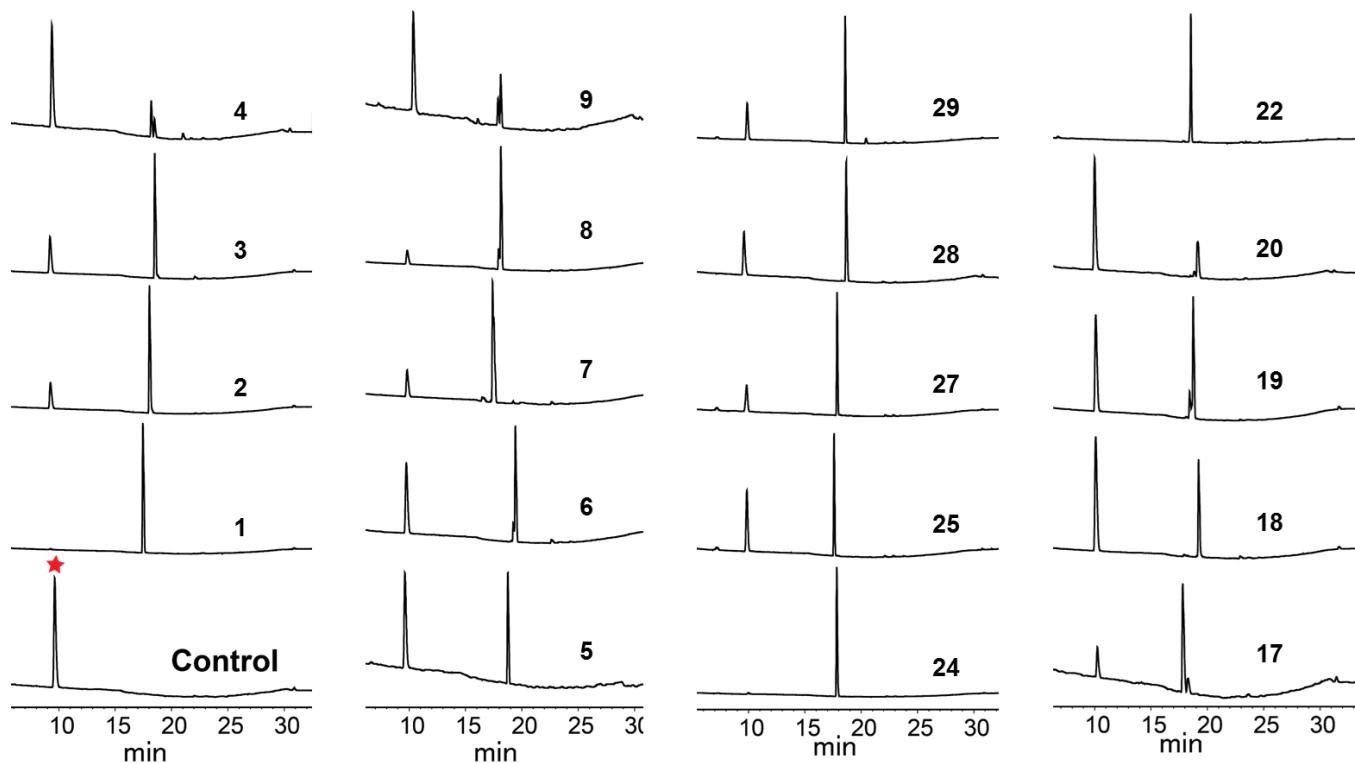
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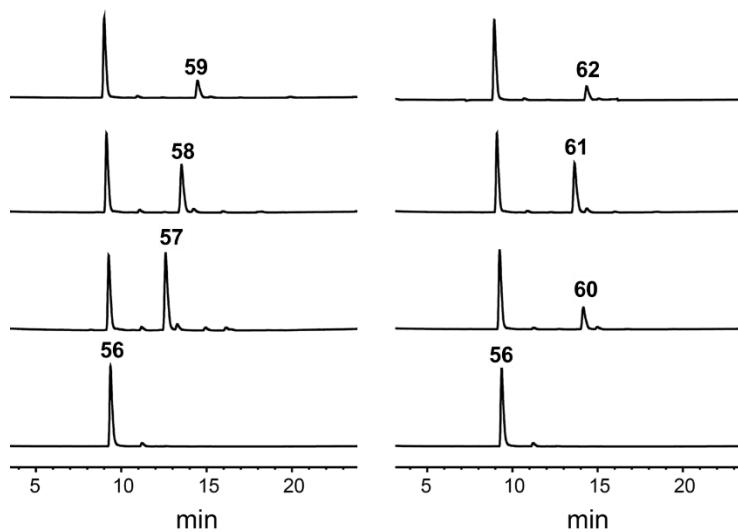
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**Figure S1:** Comparison of UV wavelength scan of L-Trp (blue) and **17** (orange).



**Figure S2:** RP-HPLC chromatograms of FgaPT2 catalyzed reactions with L-Trp (labelled with red asterisk) and alkyl-PP analogues that led to the formation of corresponding alkyl tryptophan analogues. The alkyl-PP analogue used for the reaction is labelled on the chromatogram.



**Figure S3:** RP-HPLC chromatograms of FgaPT2 catalyzed reactions with 7-hydroxy-K252c (**56**) and alkyl-PP analogues that led to the formation of corresponding alkyl-7-hydroxy-K252c analogues (**57-62**).

Alkyl-PP analogue	Concentration of Alkyl-PP (mM)	$k_{cat}$ (min <sup>-1</sup> )	$K_M$ (mM)	$k_{cat} / K_M$ (mM <sup>-1</sup> min <sup>-1</sup> )
1	2	65 ± 3	0.17 ± 0.04	380 ± 90
2	5	14.3 ± 0.2	0.28 ± 0.02	51 ± 4
3	5	20.80 ± 0.4	0.26 ± 0.02	78 ± 6
4	5	0.38 ± 0.01	0.18 ± 0.02	2.1 ± 0.2
6	10	5.2 ± 0.5	0.14 ± 0.07	36 ± 16
7	2	0.84 ± 0.05	0.19 ± 0.06	4 ± 1
8	5	3.0 ± 0.1	0.16 ± 0.03	18 ± 4
9	5	1.44 ± 0.08	0.33 ± 0.08	4 ± 1
17	5	33 ± 2	0.24 ± 0.06	137 ± 35
20	10	2.23 ± 0.04	0.31 ± 0.02	7.2 ± 0.5
22	5	5.1 ± 0.2	0.17 ± 0.02	29 ± 4
24	5	5.48 ± 0.09	0.22 ± 0.02	25 ± 2
25	5	0.88 ± 0.02	0.20 ± 0.02	4.3 ± 0.5
27	5	0.31 ± 0.01	0.12 ± 0.02	2.5 ± 0.4
28	5	0.123 ± 0.004	0.25 ± 0.03	0.48 ± 0.06
29	5	0.242 ± 0.006	0.25 ± 0.03	1.0 ± 0.1

**Table S1:** Kinetic parameters for FgaPT2 with varied concentration of L-Trp and constant concentration with representative alkyl-PP analogues in 25 mM Tris, 5 mM MgCl<sub>2</sub>, 50 mM KCl, pH 7.5, 35 °C.

**Table S2:** Summary of HRMS data of synthetic alkyl-PP analogues.

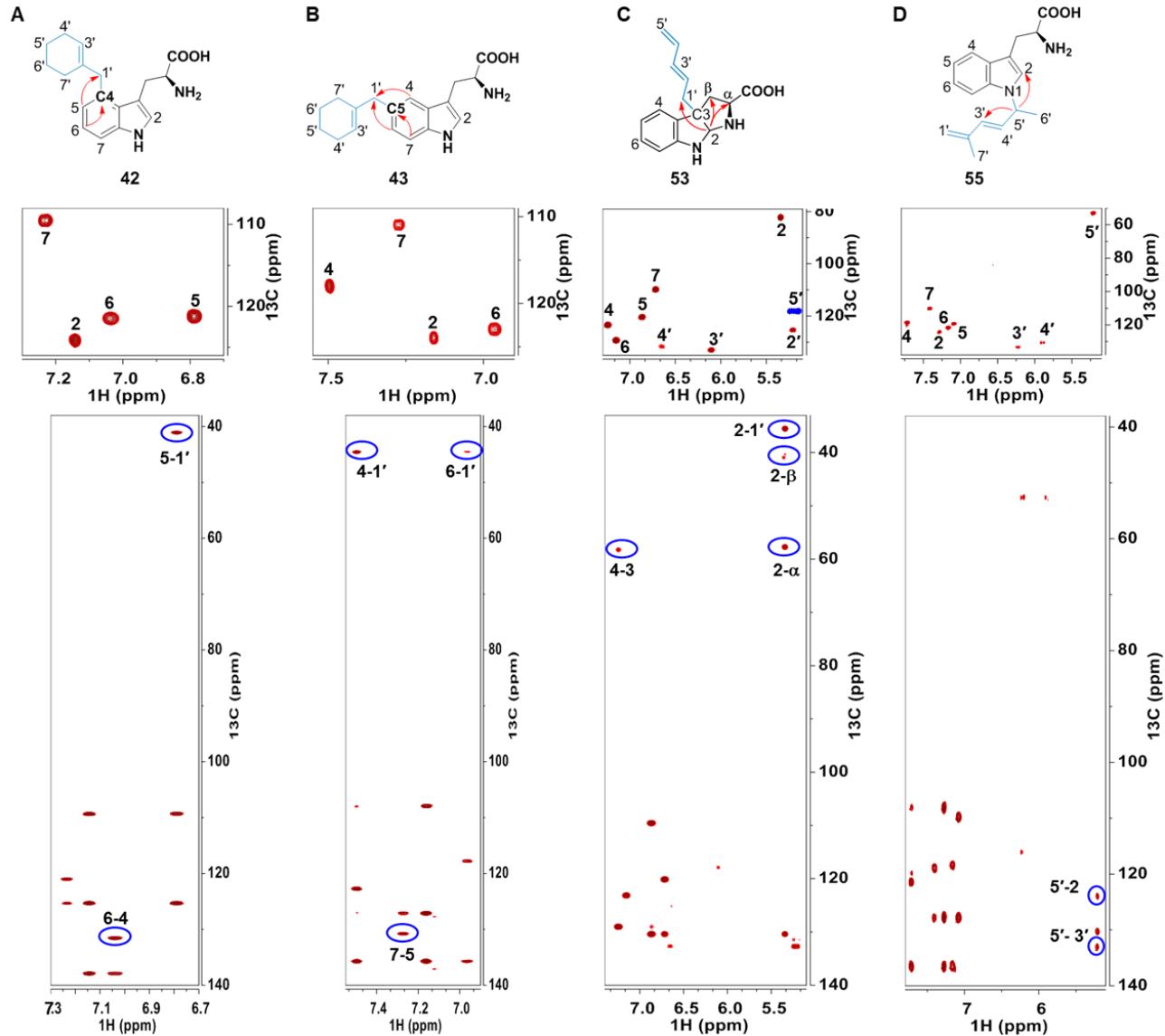
Compound	Chemical Formula	Calculated Mass (Da)	Observed (Da)
<b>5</b>	C <sub>7</sub> H <sub>13</sub> O <sub>7</sub> P <sub>2</sub> [M-H] <sup>-</sup>	271.0142	271.0148
<b>9</b>	C <sub>4</sub> H <sub>8</sub> BrO <sub>7</sub> P <sub>2</sub> [M-H] <sup>-</sup>	308.8934	308.8932
<b>22</b>	C <sub>7</sub> H <sub>9</sub> O <sub>7</sub> P <sub>2</sub> [M-H] <sup>-</sup>	266.9824	266.9828
<b>23</b>	C <sub>8</sub> H <sub>11</sub> O <sub>8</sub> P <sub>2</sub> [M-H] <sup>-</sup>	296.9929	296.9939
<b>24</b>	C <sub>8</sub> H <sub>11</sub> O <sub>8</sub> P <sub>2</sub> [M-H] <sup>-</sup>	296.9929	296.9927
<b>25</b>	C <sub>8</sub> H <sub>11</sub> O <sub>8</sub> P <sub>2</sub> [M-H] <sup>-</sup>	296.9929	296.9926
<b>26</b>	C <sub>8</sub> H <sub>11</sub> O <sub>7</sub> P <sub>2</sub> [M-H] <sup>-</sup>	280.9980	280.9978
<b>27</b>	C <sub>7</sub> H <sub>8</sub> FO <sub>7</sub> P <sub>2</sub> [M-H] <sup>-</sup>	284.9729	284.9728
<b>28</b>	C <sub>7</sub> H <sub>8</sub> ClO <sub>7</sub> P <sub>2</sub> [M-H] <sup>-</sup>	300.9439	300.9428
<b>29</b>	C <sub>7</sub> H <sub>8</sub> BrO <sub>7</sub> P <sub>2</sub> [M-H] <sup>-</sup>	344.8929	344.8927
<b>30</b>	C <sub>7</sub> H <sub>8</sub> NO <sub>9</sub> P <sub>2</sub> [M-H] <sup>-</sup>	311.9674	311.9677
<b>31</b>	C <sub>9</sub> H <sub>13</sub> O <sub>9</sub> P <sub>2</sub> [M-H] <sup>-</sup>	327.0035	327.0048
<b>32</b>	C <sub>9</sub> H <sub>13</sub> O <sub>9</sub> P <sub>2</sub> [M-H] <sup>-</sup>	327.0035	327.0027
<b>33</b>	C <sub>7</sub> H <sub>7</sub> F <sub>2</sub> O <sub>7</sub> P <sub>2</sub> [M-H] <sup>-</sup>	302.9635	302.9629
<b>34</b>	C <sub>10</sub> H <sub>15</sub> O <sub>10</sub> P <sub>2</sub> [M-H] <sup>-</sup>	357.0140	357.0135

**Table S3:** Summary of HRMS data of alkyl-Trp analogues from FgaPT2 catalyzed alkylation reaction with L-Trp and synthetic alkyl-PP analogues. The subscript ‘2’ represent the minor product based on HPLC retention time.

Alkyl -PP used for the reaction	Enzyme Product	Chemical Formula	Calculated Mass (Da)	Observed Mass (Da)
1	35	C <sub>16</sub> H <sub>21</sub> N <sub>2</sub> O <sub>2</sub> [M+H] <sup>+</sup>	273.1603	273.1602
2	36	C <sub>17</sub> H <sub>23</sub> N <sub>2</sub> O <sub>2</sub> [M+H] <sup>+</sup>	287.1760	287.1754
3	37	C <sub>18</sub> H <sub>25</sub> N <sub>2</sub> O <sub>2</sub> [M+H] <sup>+</sup>	301.1916	301.1916
4	42	C <sub>18</sub> H <sub>23</sub> N <sub>2</sub> O <sub>2</sub> [M+H] <sup>+</sup>	299.1760	299.1756
4	43	C <sub>18</sub> H <sub>23</sub> N <sub>2</sub> O <sub>2</sub> [M+H] <sup>+</sup>	299.1760	299.1761
5	38	C <sub>18</sub> H <sub>23</sub> N <sub>2</sub> O <sub>2</sub> [M+H] <sup>+</sup>	299.1760	299.1768
6	44	C <sub>19</sub> H <sub>25</sub> N <sub>2</sub> O <sub>2</sub> [M+H] <sup>+</sup>	313.1916	313.1913
6	6-Trp <sub>2</sub>	C <sub>19</sub> H <sub>25</sub> N <sub>2</sub> O <sub>2</sub> [M+H] <sup>+</sup>	313.1916	313.1911
7	40	C <sub>15</sub> H <sub>19</sub> N <sub>2</sub> O <sub>2</sub> [M+H] <sup>+</sup>	259.1447	259.1449
7	41	C <sub>15</sub> H <sub>19</sub> N <sub>2</sub> O <sub>2</sub> [M+H] <sup>+</sup>	259.1447	259.1444
8	45	C <sub>15</sub> H <sub>18</sub> CIN <sub>2</sub> O <sub>2</sub> [M+H] <sup>+</sup>	293.1057	293.1061
8	8-Trp <sub>2</sub>	C <sub>15</sub> H <sub>18</sub> CIN <sub>2</sub> O <sub>2</sub> [M+H] <sup>+</sup>	293.1057	293.1062
9	46	C <sub>15</sub> H <sub>18</sub> BrN <sub>2</sub> O <sub>2</sub> [M+H] <sup>+</sup>	337.0552	337.0552
9	9-Trp <sub>2</sub>	C <sub>15</sub> H <sub>18</sub> BrN <sub>2</sub> O <sub>2</sub> [M+H] <sup>+</sup>	337.0552	337.0547
10	10-Trp	C <sub>14</sub> H <sub>17</sub> N <sub>2</sub> O <sub>2</sub> [M+H] <sup>+</sup>	245.1290	245.1294
13	13-Trp	C <sub>20</sub> H <sub>21</sub> N <sub>2</sub> O <sub>2</sub> [M+H] <sup>+</sup>	321.1603	321.1601
14	14-Trp	C <sub>21</sub> H <sub>23</sub> N <sub>2</sub> O <sub>3</sub> [M+H] <sup>+</sup>	351.1709	351.1711
15	15-Trp	C <sub>21</sub> H <sub>23</sub> N <sub>2</sub> O <sub>2</sub> [M+H] <sup>+</sup>	335.1760	335.1757
16	16-Trp	C <sub>26</sub> H <sub>25</sub> N <sub>2</sub> O <sub>2</sub> [M+H] <sup>+</sup>	397.1916	397.1912
17	53	C <sub>16</sub> H <sub>19</sub> N <sub>2</sub> O <sub>2</sub> [M+H] <sup>+</sup>	271.1447	271.1442
17	17-Trp <sub>2</sub>	C <sub>16</sub> H <sub>19</sub> N <sub>2</sub> O <sub>2</sub> [M+H] <sup>+</sup>	271.1447	271.1444
18	47	C <sub>18</sub> H <sub>23</sub> N <sub>2</sub> O <sub>2</sub> [M+H] <sup>+</sup>	299.1760	299.1768
19	54	C <sub>17</sub> H <sub>21</sub> N <sub>2</sub> O <sub>2</sub> [M+H] <sup>+</sup>	285.1603	285.1602
19	19-Trp <sub>2</sub>	C <sub>17</sub> H <sub>21</sub> N <sub>2</sub> O <sub>2</sub> [M+H] <sup>+</sup>	285.1603	285.1605
20	55	C <sub>18</sub> H <sub>23</sub> N <sub>2</sub> O <sub>2</sub> [M+H] <sup>+</sup>	299.1715	299.1751
20	20-Trp <sub>2</sub>	C <sub>18</sub> H <sub>23</sub> N <sub>2</sub> O <sub>2</sub> [M+H] <sup>+</sup>	299.1715	299.1749
21	21-Trp	C <sub>22</sub> H <sub>23</sub> N <sub>2</sub> O <sub>2</sub> [M+H] <sup>+</sup>	347.1760	347.1767
22	48	C <sub>18</sub> H <sub>19</sub> N <sub>2</sub> O <sub>2</sub> [M+H] <sup>+</sup>	295.1447	295.1430
24	39	C <sub>19</sub> H <sub>21</sub> N <sub>2</sub> O <sub>3</sub> [M+H] <sup>+</sup>	325.1552	325.1547
25	49	C <sub>19</sub> H <sub>21</sub> N <sub>2</sub> O <sub>3</sub> [M+H] <sup>+</sup>	325.1552	325.1540
27	50	C <sub>18</sub> H <sub>18</sub> FN <sub>2</sub> O <sub>2</sub> [M+H] <sup>+</sup>	313.1352	313.1338
28	51	C <sub>18</sub> H <sub>18</sub> CIN <sub>2</sub> O <sub>2</sub> [M+H] <sup>+</sup>	329.1057	329.1039
29	52	C <sub>18</sub> H <sub>18</sub> BrN <sub>2</sub> O <sub>2</sub> [M+H] <sup>+</sup>	373.0552	373.0531

**Table S4:** Summary of HRMS data of alkyl-7-hydroxy-K252c analogues (**57-62**) from FgaPT2 catalyzed alkylation reaction with 7-hydroxy-K252c (**56**) and synthetic alkyl-PP analogues.

Alkyl -PP used for the reaction	Compound	Chemical Formula	Calculated Mass (Da)	Observed Mass (Da)
-	<b>56</b>	C <sub>20</sub> H <sub>12</sub> N <sub>3</sub> O <sub>2</sub> [M-H] <sup>-</sup>	326.0930	326.0934
<b>1</b>	<b>57</b>	C <sub>25</sub> H <sub>20</sub> N <sub>3</sub> O <sub>2</sub> [M-H] <sup>-</sup>	394.1556	394.1553
<b>2</b>	<b>58</b>	C <sub>26</sub> H <sub>22</sub> N <sub>3</sub> O <sub>2</sub> [M-H] <sup>-</sup>	408.1712	408.1712
<b>3</b>	<b>59</b>	C <sub>27</sub> H <sub>24</sub> N <sub>3</sub> O <sub>2</sub> [M-H] <sup>-</sup>	422.1869	422.1882
<b>4</b>	<b>60</b>	C <sub>27</sub> H <sub>22</sub> N <sub>3</sub> O <sub>2</sub> [M-H] <sup>-</sup>	420.1712	420.1727
<b>5</b>	<b>61</b>	C <sub>27</sub> H <sub>22</sub> N <sub>3</sub> O <sub>2</sub> [M-H] <sup>-</sup>	420.1712	420.1709
<b>6</b>	<b>62</b>	C <sub>28</sub> H <sub>24</sub> N <sub>3</sub> O <sub>2</sub> [M-H] <sup>-</sup>	434.1869	434.1878



**Figure S4:** Representative 2D  $^1\text{H}, ^{13}\text{C}$  HSQC (top) and HMBC (bottom) demonstrating the C4 (**A**), C5 (**B**), C3 (**C**) and N1 (**D**) regio-chemical attachment of the alkyl groups on L-Trp. The blue circled HMBC peaks show a correlation between atoms with red arrows on the structure. Black and blue colored labels denote signals from the indole ring and alkyl-group, respectively.

### The NMR analysis of C4-alkyl-Trp analogues

Six allylic analogues (**1**, **2**, **3**, **4**, **5**, and **7**) and an o-methoxy benzyl analogue (**24**) in FgaPT2 catalyzed L-Trp alkylation reactions produced normal C4 alkylated products. The reaction involving **1**, **2**, **3**, **5**, and **24** produced a single C4 product, whereas **4** and **7** produced the C4-isomer as the major product. Based on <sup>1</sup>H and 2D NMR analysis of the alkyl-group, all seven C4-alkyl-Trp analogues were confirmed to be normal alkylated. The <sup>1</sup>H NMR of C4-alkyl-Trp analogues when compared to that of tryptophan, showed the disappearance of the downfield shifted aromatic doublet signal corresponding to the H4 (7.58 ppm) of tryptophan. Among the four observed aromatic signals of these alkyl-tryptophan analogues, one was a singlet corresponding to H2 of the indole moiety, two doublets (H5 and H7) and one triplet (H6). Assignment of the aromatic signals was further supported by the COSY correlation peaks (H5-H6 and H6-H7) and carbon chemical shift values derived from <sup>1</sup>H-<sup>13</sup>C-HSQC, that confirmed the substitution of the alkyl group to take place on the C4 position of the aromatic ring of tryptophan (top panel of **Figure S4A**, a representative <sup>1</sup>H-<sup>13</sup>C-HSQC spectrum of **42**). In order to unambiguously confirm the findings, representative 2D <sup>1</sup>H-<sup>13</sup>C-HMBC spectra were collected. The sequential connectivity between H5 of the indole moiety to the C1' of the alkyl group (bottom panel of **Figure S4A**, a representative <sup>1</sup>H-<sup>13</sup>C-HMBC spectrum of **42**, **Table S5**) and/or H1' of the alkyl group to the C5 and C3a of the tryptophan aromatic moiety (**Table S5**) unambiguously confirmed the attachment of alkyl group at C4 position of tryptophan. Further, the NMR data of **35** and **40** matched the previously reported values,<sup>30, 41</sup> that formed the basis for comparison with the rest of the C4-isomers. Additional confirmation of the products was obtained from HRMS data, that displayed mass corresponding to the alkylated tryptophan product (**Table S3**). These results demonstrate the ability of FgaPT2 to catalyze C4-normal alkylation of tryptophan using unnatural alkyl donors including, o-methoxybenzyl and other allylic (**1-5**, and **7**) alkyl donors.

### The NMR analysis of C5-alkyl-Trp analogues

Eleven alkyl-PP analogues (**4**, **6**, **7**, **8**, **9**, **18**, **22**, **25**, **27**, **28**, and **29**) in a FgaPT2 catalyzed alkylation reaction with tryptophan produced normal C5 alkylated products. These include single product of FgaPT2 reactions with **18**, **22**, **25**, **27**, **28**, **29**; major product of reactions involving **6**, **8**, and **9**; and minor product of reactions with **4** and **7**. Detailed <sup>1</sup>H and 2D NMR analysis of the alkyl-group of all 11 of the C5-alkyl-Trp analogues were confirmed to be normal alkylated (**Table S5**). Comparison of <sup>1</sup>H NMR and <sup>1</sup>H-<sup>13</sup>C-HSQC spectra of the C5-alkyl-Trp analogues with that of tryptophan revealed that the upfield shifted triplet signal corresponding to H5 of the indole moiety (<sup>1</sup>H, <sup>13</sup>C = 7.05 ppm, 118.1 ppm) has disappeared. Further, the presence of two singlets (H2 and H4) and two doublets (H6 and H7) and COSY correlation between H6-H7, along with their signature carbon chemical shift values derived from the combined analysis of COSY, and <sup>1</sup>H-<sup>13</sup>C-HSQC spectra (top panel of **Figure S4B**, a representative <sup>1</sup>H-<sup>13</sup>C-HSQC

spectrum of **43**, **Table S5**); implied the alkyl addition to take place at the C5 position of the aromatic ring of tryptophan. To confirm the C5 regio-specific attachment of the alkyl group and to identify the aromatic signals of the benzylic alkyl substituents,  $^1\text{H}$ - $^{13}\text{C}$ -HMBC spectra were collected for all new benzylic alkyl tryptophan analogues (involving **25**, **27**, **28** and **29**) and two representative allylic alkyl tryptophan analogues (involving **4** and **9**) (**Table S5**). The HMBC spectra displayed the correlation from H4 and H6 of tryptophan to the C1' of the alkyl-group (bottom panel of **Figure S4B**, a representative  $^1\text{H}$ - $^{13}\text{C}$ -HMBC spectrum of **43**, **Table S5**); and/or from H1' of the alkyl group to C4 and C6 of the tryptophan aromatic moiety (see Supporting Information, **Table S5**), confirming the C5-alkylation of tryptophan. In addition, the NMR data of **48** matched the previously reported values.<sup>41, 42</sup> Additional confirmation of alkyl group attachment to tryptophan was obtained via HRMS analysis (**Table S3**). These results support the ability of FgaPT2 to catalyze normal alkylation at the C5 position of unsubstituted L-tryptophan using diverse benzylic and allylic alkyl donors.

#### The NMR analysis of C3-alkyl-Trp analogue (**53**)

The UV spectra of the major product of the FgaPT2 catalyzed a reaction with **17** and tryptophan displayed maximum absorption at 228, 240, and 289 nm (**Figure S1**), suggesting the presence of an indoline-like structure. This was further confirmed by HRMS and NMR analysis (**Tables S3 & S5**). The  $^1\text{H}$  and 2D NMR analysis of the alkyl group suggested the alkylation to happen in a normal manner. In addition, the coupling pattern and COSY correlation of the four aromatic protons suggested the six-membered aromatic ring of the indole moiety is similar to that of tryptophan, implying the alkyl substitution is not directly on the six-membered ring of tryptophan. Further, the combined analysis of  $^1\text{H}$ - $^{13}\text{C}$ -HSQC and  $^1\text{H}$ - $^{13}\text{C}$ -HMBC spectra revealed the upfield shifted signals of H2/C2 (5.34 ppm/82.2 ppm) and C3 (58.2 ppm) of tryptophan (**Figure S4C**), clearly indicating the broken aromaticity of the pyrrole ring. The unambiguous confirmation of cyclization, that yielded a hexahydro-pyrroloindoline moiety was derived from  $^1\text{H}$ - $^{13}\text{C}$ -HMBC analysis, that displayed the connectivity between H1' of the alkyl group to C2, C $\beta$  and C3a position; and C2 to C $\alpha$  of the alkyl-pyrroloindoline moiety (bottom panel of **Figure S4C**, **Table S5**). These analyses confirmed FgaPT2 to catalyze normal C3-alkylation followed by cyclization of tryptophan in the presence of **17**.

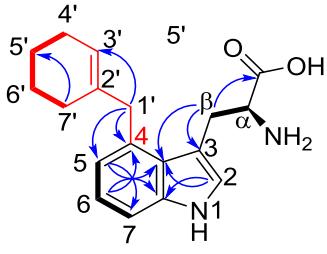
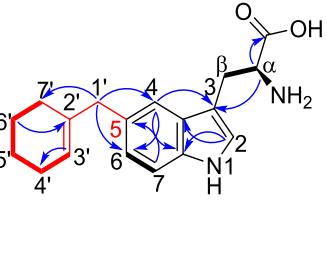
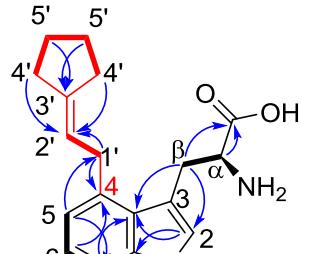
#### The NMR analysis of N1-alkyl-Trp analogues (**54** and **55**)

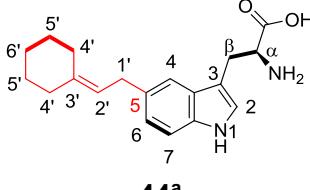
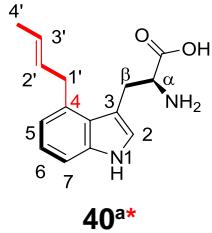
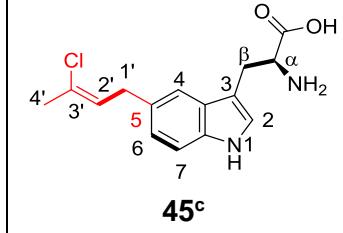
The NMR analysis of the major product of FgaPT2 catalyzed reaction using alkyl-PP analogues, **19** and **20**, confirmed them to be N1-alkylated. The detailed analysis of **54** suggested a normal alkylation based on the pattern of signals of the alkyl moiety, whereas, **55** indicated it to be reverse alkylated via the C5' (reverse $\text{C}5'$ ) of alkyl-PP analogue, **20** (**Figure 5**). More specifically, the chemical shifts, coupling pattern

and COSY correlation of all aromatic protons (H2, H4, H5, H6, and H7) of the **54** and **55** analogues appeared similar to that of tryptophan, implying no substitution to take place directly on the carbon atom of the aromatic ring of tryptophan. This is supported by the  $^1\text{H}$  and  $^{13}\text{C}$  chemical shift assignment (top panel of **Figure S4D**, a representative  $^1\text{H}$ - $^{13}\text{C}$ -HSQC spectrum of **55**, **Table S5**) and COSY correlations (H4-H5, H5-H6, H6-H7) of the **54** and **55**. The H1' chemical shift value of the alkyl group in **54** at 5.05 ppm supported the N-alkylation of tryptophan with attachment via C1'. The pattern of signals of alkyl moiety in **55** suggested C5' of **20** was attached to N1 of the indole ring with C5'/H5' chemical shift values at 53 ppm/5.21 ppm respectively (top panel of **Figure S4D**, **Table S5**). The confirmation of the N1-alkylation and alkyl group chemical shift assignment of **55** was derived from the  $^1\text{H}$ - $^{13}\text{C}$  HMBC spectra, that displayed a correlation between H5' of the alkyl group and C2 of the tryptophan aromatic moiety (bottom panel of **Figure S4D** for **55**, **Table S5**). This is the first demonstration of FgaPT2 to carry out the reverse alkylation (although via C5' group of **20**) at N1 of tryptophan. Therefore, while the FgaPT2-catalyzed N1-normal alkylation has been previously demonstrated with tryptophan derivatives using DMAAPP,<sup>34</sup> this study revealed the possibility of N1-reverse<sub>C5'</sub> alkylation of native tryptophan by an unnatural alkyl-donor, **20**.

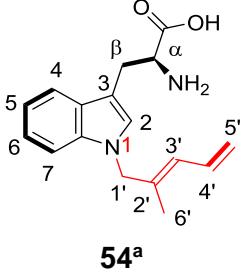
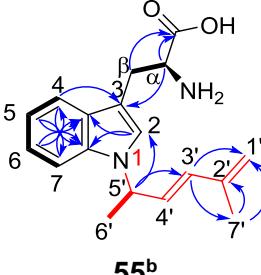
**Table S5.** Summary of NMR chemical shift assignment of the alkyl-L-Trp analogues (500 MHz). Where  $\delta_H$  and  $\delta_C$  values are in ppm and coupling constant,  $J$  is in Hz. a= DMSO-d<sub>6</sub>, b= MeOH-d<sub>4</sub>

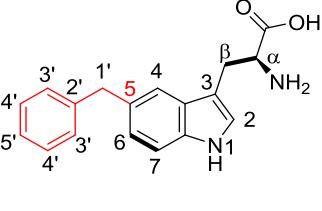
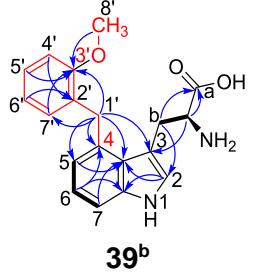
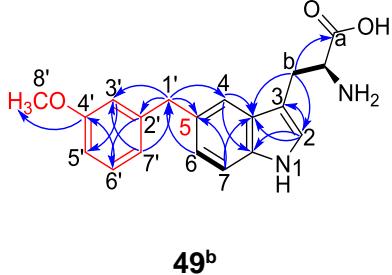
Structure / COSY HMBC Correlation / Compound						
Position	$\delta_H$ , multi ( $J$ )	$\delta_C$	$\delta_H$ , multi ( $J$ )	$\delta_C$	$\delta_H$ , multi ( $J$ )	$\delta_C$
COOH	-	n.f.	-	n.f.	-	n.f.
NH	10.99, s	-	10.88, d (2.5)	-	10.89, d (2.5)	-
2	7.18, s	125.3	7.18, d (2.5)	124.8	7.18, d (2.5)	124.6
3	-	n.f.	-	n.f.	-	n.f.
3a	-	n.f.	-	n.f.	-	n.f.
4	-	n.f.	-	n.f.	-	n.f.
5	6.73, d (7.7)	119.4	6.71, d (7.6)	118.9	6.72, d (7.6)	118.7
6	6.97, t (7.7)	121.7	6.95, t (7.6)	121.5	6.95 t, (7.6)	121.5
7	7.19, d (7.7)	110.1	7.17, d (7.6)	109.9	7.17, d (7.6)	109.9
7a	-	n.f.	-	n.f.	-	n.f.
$\beta$	3.51, dd (15.6, 4.4); 3.11, dd (15.6, 10.5)	n.f.	3.57, dd (15.6, 3.3); 2.94, dd (15.6, 10.7)	n.f.	3.58, dd (15.6, 3.2); 2.95, dd (15.6, 10.6)	29.7
$\alpha$	3.41	n.f.	3.39, dd (10.6, 3.2)	56.1	3.39, dd (10.6, 3.2)	55.9
1'	3.7-3.66, m	31.9	3.68, dd (16.0, 7.8) 3.74, dd (16.0, 6.0)	31.5	3.71, dd (16.0, 7.8) 3.76, dd (16.0, 7.1)	31.1
2'	5.29, dd (7.8, 6.0)	124.1	5.32 dd (7.8, 6.0)	122.7	5.29, t (7.1)	122.1
3'	-	n.f.	-	n.f.	-	n.f.
4'	1.70, s	26.1	1.71, s	16.6	2.08-2.02, m	29.2
5'	1.70, s	18.4	2.02, q (7.4)	32.4	0.98, t (7.5)	13.4
6'	-	-	0.98, t (7.4)	13.1	2.18-2.12,m	23.5
7'	-	-	-	-	0.97, t (7.5)	13.4

Structure / COSY HMBC Correlation / Compound			
	<b>42<sup>b</sup></b>	<b>43<sup>b</sup></b>	<b>38<sup>b</sup></b> (80% C4 <sup>N</sup> isomer + 20% unknown isomer)
Position	$\delta_{\text{H}}$ , multi (J)	$\delta_{\text{C}}$	$\delta_{\text{H}}$ , multi (J)
COOH	-	173.0	-
NH	exc.	-	exc
2	7.14, s	124.1	7.16, s
3	-	109.3	-
3a	-	125.3	-
4	-	131.5	7.50, d (1.5)
5	6.79, d (7.6)	121.2	-
6	7.04, t (7.6)	121.5	6.97, dd (8.4, 1.5)
7	7.23, d (7.6)	109.5	7.27, d (8.4)
7a	-	137.8	-
$\beta$	3.76, dd (15.6, 3.5) 3.02, dd (15.6,11.2)	29.3	3.53, dd (15.2, 3.9); 3.10, dd (15.2, 9.9)
$\alpha$	3.83, dd (11.2, 3.5)	56.2	3.86, dd (9.9, 3.9)
1'	3.67, s	41.5	3.33, s
2'	-	138.1	-
3'	5.08, s	121.8	5.54-5.43, m
4'	1.96-1.9, m	25.1	2.08-2.00, m
5'	1.60-1.54, m	22.6	1.64-1.52, m
6'	1.69-1.63, m	23.1	1.64-1.52, m
7'	2.13-2.06, m	28.8	1.93-1.86, m
			27.8

Structure / COSY HMBC Correlation / Compound					
Position	$\delta_H$ , multi ( $J$ )	$\delta_C$	$\delta_H$ , multi ( $J$ ) /	$\delta_C$	$\delta_H$ , multi ( $J$ )
COOH	-	n.f.	-	n.f.	-
NH	10.73, br.s	-	10.88, br. s	-	exc
2	7.14, d (1.9)	124.5	7.16, d (2.1)	124.7	7.30, s
3	-	n.f.	-	n.f.	-
3a	-	n.f.	-	n.f.	-
4	7.31, s	117.6	-/	n.f.	7.58, s
5	-	n.f.	6.73, d (7.1)	119.7	-
6	6.88, dd (8.4, 1.2)	122.3	6.97, t (7.6)	121.5	7.47, d (8.4)
7	7.24, d (8.4)	111.8	7.18, dd (7.6, 7.1)	110.4	7.18, d (8.4)
7a	-	n.f.	-	n.f.	-
$\beta$	3.33-3.25, m 2.85, dd (15.2, 9.7)	27.9	3.57,dd (15.5, 3.6) 2.88,dd (15.5, 11.1)	29.7	3.47, dd (15.2, 4.8) 3.24, dd (15.2, 8.5)
$\alpha$	3.39, dd, (9.7, 3.9)	55.3	3.36-3.34, m	56.1	4.04, dd (8.5, 4.8)
1'	3.36, d (7.5)	33.5	3.74, dd (15.5, 6.4) 3.62, dd (15.5, 6.1 )	36.2	3.63, d (7.4)
2'	5.27, t (7.5)	121.7	5.68-5.56, m	131.7	5.92-5.81, m
3'	-	n.f.	5.52-5.42, m	125.5	-
4'	2.25, t (5.2) 2.07, t (5.2)	28.7, 37.1	1.62, d (6.1)	18.2	2.18-2.13, m
5'	1.60-1.44, m	28	-	-	-
6'	1.60-1.44, m	28	-	-	-

Structure / COSY HMBC Correlation / Compound					
Position	$\delta_H$ , multi (J)	$\delta_C$	$\delta_H$ , multi (J)	$\delta_C$	$\delta_H$ , multi (J)
COOH	-	173.0		169.2	-
NH	exc	-	exc	-	exc
2	7.18, s	124.2	5.34, s	82.2	7.17, s
3	-	107.9	-	58.2	-
3a	-	127.1	-	130.5	-
4	7.53, s	122.4	7.24, d (7.6)	123.5	7.51, s
5	-	129.9	6.86, t (7.6)	120.3	-
6	6.99, d (8.3)	117.4	7.15, t (7.6)	129.2	6.98, d (8.3)
7	7.29, d (8.3)	111.3	6.71, d (7.6)	109.7	7.28, d (8.3)
7a	-	135.8	-	148.7	-
$\beta$	3.51, dd (15.3, 4.0) 3.12, dd (15.3, 9.5)	27.3	2.76, dd (13.3, 6.2) 2.44, dd (13.3, 11.8)	40.7	3.51, dd (15.3, 4.0) 3.10, dd (15.3, 9.6)
$\alpha$	3.86, dd, (9.5, 4.0)	55.5	4.00, dd (11.8, 6.2)	57.9	3.85, dd (9.6, 4.0)
1'	3.57, d (7.0)	37.9	2.78-2.68, m	34.9	3.52-3.49, m
2'	5.92, t (7.0)	129.1	5.24-5.16, m	125.3	5.71, dt (14.8, 7.00)
3'	-	121.4	6.10, t (11.0)	132.7	6.31, dd (14.8, 11.0)
4'	2.32, s	27.7	6.65, ddd (16.7, 11.0, 10.6)	131.6	5.79, d (11.0)
5'	-	-	5.22, d (16.7) 5.15, d (10.6)	118.0	-
6'	-	-	-	-	1.75, (s); 1.73, (s)

Structure / COSY HMBC Correlation/ Compound			
Position	$\delta_H$ , multi (J)	$\delta_H$ , multi (J)	$\delta_C$
COOH	-	-	172.9
NH	-	-	-
2	7.19, s	7.27, s	124.2
3	-	-	108.1
3a	-	-	127.8
4	7.58, d (7.9)	7.71, d (7.8)	118.6
5	7.02, dd (7.9, 7.6)	7.08, dd (7.8, 7.6)	119.1
6	7.12, dd (8.2, 7.6)	7.16, dd (8.2, 7.6)	121.6
7	7.39, d (8.2)	7.40, d (8.2)	109.8
7a	-	-	136.5
$\beta$	3.34-3.24, m 2.93, dd (15.1, 8.8)	3.52, dd (15.2, 4.1) 3.15, dd (15.2, 9.3)	27.4
$\alpha$	3.41-3.36, m	3.85, dd (9.3, 4.1)	55.6
1'	5.05-4.90, m	4.96, s 4.93, s	116.3
2'	-	-	141.3
3'	6.30, d (15.8)	6.22, d (15.8)	133.1
4'	5.80, dt (15.8, 6.1)	5.89, dd (15.8, 6.01)	130.3
5'	4.89-4.77, m	5.21, p (7.0, 6.0)	52.9
6'	1.76, s	1.68, d (7.0)	19.6
7'	-	1.81, s	17.6

Structure / COSY / HMBC Correlation / Compound						
<b>Position</b>	<b><math>\delta_H</math>, multi (<math>J</math>)</b>	<b><math>\delta_C</math></b>	<b><math>\delta_H</math>, multi (<math>J</math>)</b>	<b><math>\delta_C</math></b>	<b><math>\delta_H</math>, multi (<math>J</math>)</b>	<b><math>\delta_C</math></b>
COOH	-	n.f.	-	170.2	-	170.8
NH	10.77, s	-	exc	-	exc	-
2	7.15, s	124.9	7.14, s	124.9	7.18, s	124.6
3	-	n.f.	-	107.4	-	106.5
3a	-	n.f.	-	124.8	-	127.0
4	7.41, s	118.5	-	131.5	7.48, d (1,6)	117.5
5	-	n.f.	6.77, d (7.2)	121.6	-	132.0
6	6.92, dd (8.3, 1.6)	122.8	7.08, dd (8.2, 7.2)	121.9	7.00, dd (8.4, 1.6)	123.4
7	7.24, d, (8.3)	111.9	7.31, d (8.2)	110.1	7.30, d (8.4)	111.5
7a	-	n.f.	-	138.0	-	135.7
$\beta$	3.28, dd(15.2, 3.7) 2.86, dd (15.2, 9.6)	n.f.	3.52, dd (15.4, 3.9); 2.98, dd (15.4, 11.4)	28.4	3.50, dd, (15.3, 4.7) 3.27, dd (15.3, 8.4)	26.5
$\alpha$	3.38, dd (9.6, 3.7)	n.f.	3.73, dd (11.4, 3.9)	53.7	4.19, dd (8.4, 4.7)	53.5
1'	3.98, s	42.3	4.42, d (16.9); 4.28, d (16.9)	33.2	4.03, s	41.7
2'	-		-	129.5	-	144.1
3'	7.29 – 7.17, m	128.9	-	157.1	6.76, t (2.1)	114.2
4'	7.29 – 7.17, m	128.9	6.96, d, (8.2)	109.9	-	159.8
5'	7.18 – 7.1, m	125.9	7.18, dd (8.2,7.4)	127.2	6.71, dd (8.2, 2.1)	110.7
6'	-	-	6.74, t (7.4)	120.0	7.15, t (7.9)	129.1
7'	-	-	6.64, d (7.4)	129.1	6.80, dd (7.9, 2.1)	121.1
8'	-	-	3.85, s	54.5	3.73 (s, 3H)	54.3

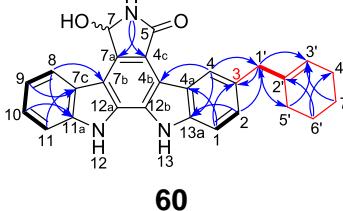
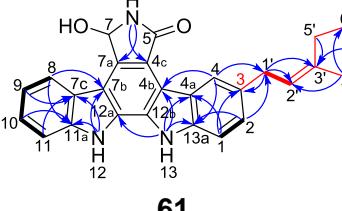
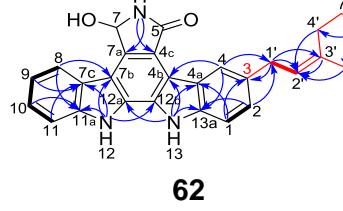
Structure / COSY HMBC Correlation / Compound	50 <sup>b</sup>		51 <sup>b</sup>		52 <sup>b</sup>	
Position	$\delta_{\text{H}}$ , multi (J)	$\delta_{\text{C}}$	$\delta_{\text{H}}$ , multi (J)	$\delta_{\text{C}}$	$\delta_{\text{H}}$ , multi (J)	$\delta_{\text{C}}$
COOH	-	170.3	-	170.5	-	170.4
NH	exc	-	exc	-	exc	-
2	7.18, s	124.4	7.19, s	124.7	7.19, s	124.7
3	-	106.2	-	106.5	-	106.4
3a	-	127.1	-	127.1	-	127.1
4	7.46, d (1.4)	117.4	7.47, s	117.5	7.46, d (1.4)	117.5
5	-	131.9	-	131.5	-	131.5
6	6.98, dd (8.3, 1.4)	122.9	6.98, d (8.3)	123.0	6.98, dd (8.3, 1.4)	123.0
7	7.31, d (8.3)	111.2	7.31, d (8.3)	111.5	7.31, d (8.3)	111.5
7a	-	135.7	-	136.2	-	135.7
$\beta$	3.48, dd (15.2, 4.9) 3.30, dd (15.2, 8.1)	26.1	3.48, dd (15.3, 4.8) 3.28, dd (15.3, 8.1)	26.5	3.48, dd (15.3, 4.8) 3.28, dd (15.3, 8.1)	26.5
$\alpha$	4.23, dd, (8.1, 4.9)	53.2	4.19, dd (8.1, 4.8)	53.5	4.22, dd (8.1, 4.9)	53.4
1'	4.04, s	40.8	4.04, s	41.2	4.03, s	41.0
2'	-	138.5	-	141.4	-	141.8
3'	7.21, dd (8.6, 5.5)	129.9	7.19, d (8.3)	130.1	7.14, d (8.3)	130.4
4'	6.96, dd (9.1, 8.6)	114.3	7.23, d (8.3)	128.1	7.38, d (8.3)	131.1
5'	-	161.3 d. (240.3)	-	131.1	-	119.4

\*NMR assignment of these compounds has been reported previously; — = COSY correlations; ↗ = HMBC correlations.

**Table S6.** Summary of NMR chemical shift assignment of the alkyl-7-hydroxy-K252c analogues in Acetone-d<sub>6</sub> (500 MHz). Where δ<sub>H</sub> and δ<sub>C</sub> values are in ppm and coupling constant, J is in Hz.

Structure / COSY / HMBC Correlation / Compound	57	58	59			
Position	δH, multi (J)	δC	δH, multi (J)	δC	δH, multi (J)	δC
1	7.53, d (8.1)	111.2	7.53, d (8.0)	110.9	7.54, d (8.3)	110.9
2	7.26, dd (8.1, 1.8)	126.7	7.26, dd (8.0, 1.8)	126.3	7.28, dd (8.3, 1.8)	126.1
3	-	132.7	-	132.7	-	132.8
4	9.18, d (1.8)	125.4	9.18, d (1.8)	125.2	9.20, d (1.8)	124.9
4a	-	123.6	-	123.6	-	123.7
4b	-	116.3	-	116.3	-	116.31
4c	-	n.f.	-	135.1	-	n.f.
5	-	n.f.	-	n.f.	-	n.f.
6	7.72, s	-	7.72, s	-	7.72, s	-
7	6.57, d (10.6)	79.6	6.56, d (5.4)	79.4	6.59, d (8.6)	79.3
7-OH	5.24, d (10.6)	-	5.24, d (5.4)	-	5.24, d (8.6)	-
7a	-	n.f.	-	118.6	-	118.6
7b	-	n.f.	-	115.7	-	115.8
7c	-	122.9	-	122.9	-	122.9
8	8.49, d (7.9)	123.7	8.50, d (7.8)	123.4	8.49, d (7.9)	123.4
9	7.30-7.26, m	120.4	7.30-7.26, m	119.9	7.31-7.27, m	120.0
10	7.46-7.42, m	125.7	7.45-7.41, m	125.3	7.46-7.42, m	125.3
11	7.67, d (7.9)	111.8	7.66, d (8.0)	111.5	7.66, d (8.1)	111.5
11a	-	140.1	-	140.1	-	140.1
12	11.08, s	-	11.14, s	-	11.03, s	-
12a	-	n.f.	-	n.f.	-	n.f.
13	10.85, s	-	10.91, s	-	10.81, s	-
13a	-	138.5	-	138.6	-	138.6
1'	3.54, d (7.3)	34.9	3.55, d (7.2)	34.5	3.57, d (7.4)	33.9
2'	5.46, t (7.3)	125.4	5.48, t (7.2)	123.5	5.43, t (7.4)	122.9
3'	-	130.9	-	136.6	-	142.3
4'	1.82, s	17.3	1.82, s	15.7	2.11, q (7.6)	29.2
5'	1.76, s	25.6	2.07, q (7.4)	32.5	2.28, q (7.6)	23.1
6'	-	-	1.04, t (7.4)	12.6	1.09, t (7.6)	12.6
7'	-	-	-	-	1.04, t (7.6)	12.5

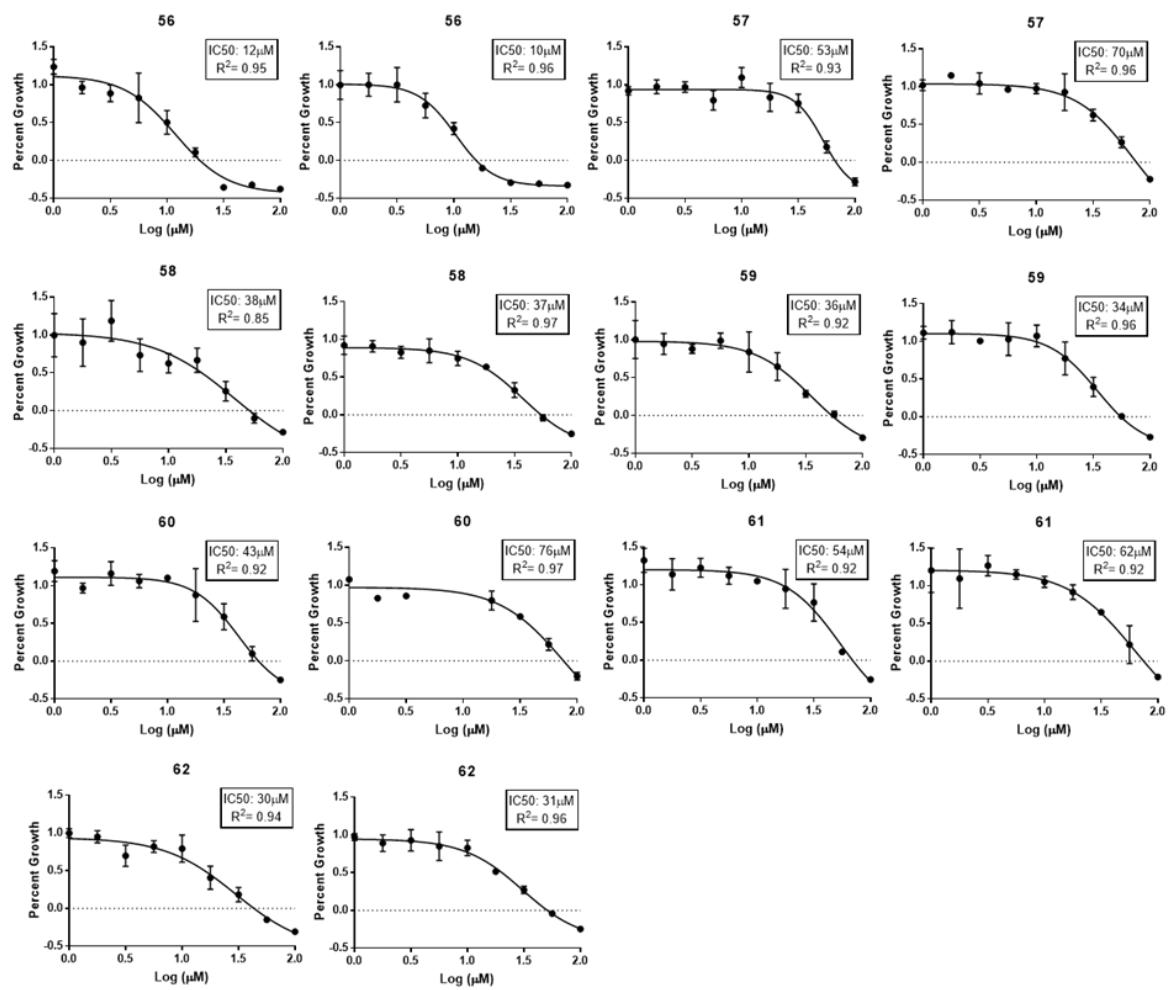
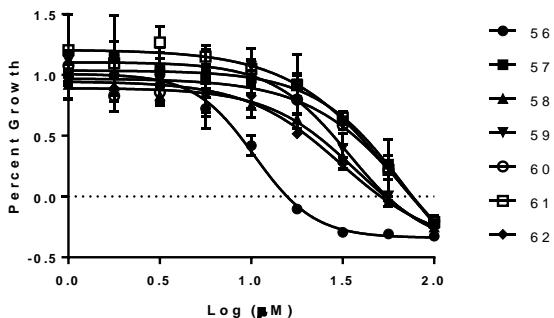
\*NMR assignment of these compounds has been reported previously; — = COSY correlations; ↗ = HMBC correlations.

Structure / COSY HMBC Correlation / Compound						
Position	$\delta$ H, multi (J)	$\delta$ C	$\delta$ H, multi (J)	$\delta$ C	$\delta$ H, multi (J)	$\delta$ C
1	7.53 (d, (8.2))	110.8	7.55, d (8.3)	111.0	7.53, d (8.2)	110.9
2	7.26 d, (8.2)	126.6	7.26, d (8.3)	125.9	7.26, d (8.2)	126.4
3	-	131.1	-	132.1	-	132.7
4	9.16 (s)	125.9	9.16, s	124.9	9.17, s	125.1
4a	-	123.5	-	123.5	-	123.6
4b	-	116.3	-	115.9	-	116.2
4c	-	135.2	-	134.3	-	135.1
5	-	n.f.	-	n.f.	-	n.f.
6	7.75 (s)	-	7.66, s	-	7.73, s	-
7	6.57 (d, (8.1))	79.4	6.55, d (8.3)	79.4	6.56, d (9.0)	79.3
7-OH	5.24 (d, (8.1))	-	5.28, d (8.3)	-	5.24, d (9.0)	-
7a	-	118.5	-	118.1	-	118.5
7b	-	115.7	-	115.3	-	115.6
7c	-	122.9	-	122.9	-	122.9
8	8.49 (d, (7.9))	123.4	8.46, d (7.9)	123.3	8.48, d (7.9)	123.3
9	7.29 (dd, (7.9, 7.6))	119.8	7.25, dd (7.9, 7.7)	119.7	7.28, t (7.9)	119.9
10	7.44 (dd, (8.1, 7.6))	125.2	7.42, dd (8.3, 7.7)	125.1	7.43, t (7.6)	125.3
11	7.66 (d, (8.1))	111.5	7.67, d (8.3)	111.6	7.66, d (8.1)	111.4
11a	-	140.1	-	140.4	-	140.1
12	11.02 (s)	-	12.24, s	-	11.16, s	-
12a	-	n.f.	-	127.3	-	126.9
12b	-	n.f.	-	n.f.	-	128.6
13	10.81 (s)	-	11.96, s	-	10.92, s	-
13a	-	n.f.	-	138.8	-	138.5
1'	3.43 (s)	44.9	3.51, d (7.3)	36.3	3.53, d (7.5)	33.6
2'	-	138.2	5.57, tp (7.3, 2.4)	120.4	5.39, t (7.5)	121.5
3'	5.52 (s)	122.2	-	142.7	-	139.1
4'	2.05 – 2.00 (m)	25.4	2.42, td (7.3, 2.4)	28.9	2.15, t (5.5)	37.3
5'	2.00 – 1.91 (m)	27.9	2.29, td (7.3, 2.4)	33.7	2.37, t (5.2)	28.6
6'	1.81 – 1.38 (m)	22.8	1.74, p (7.3, 6.9)	26.2	1.73 – 1.51, m	27.7
7'	1.81 – 1.38 (m)	22.8	1.64, p (7.3, 6.9)	26.4	1.73 – 1.51, m	28.6
8'	-	-	-	-	1.73 – 1.51, m	26.8

n.f. = Not found; exec.=exchangeable

**Table S7:** Values used to generate Figure 3 obtained from two experimental replicates.

Alkyl-PP analogue	% Conversion
1	94 ± 8
2	77 ± 2
3	66 ± 8
4	40 ± 16
5	42 ± 4
6	52 ± 5
7	70 ± 16
8	78 ± 10
9	41 ± 8
10	1 ± 0
13	1 ± 1
14	0.5 ± 0.1
15	0.9 ± 0.1
16	1 ± 0.8
17	84 ± 9
18	38 ± 11
19	54 ± 9
20	22.4 ± 0.6
21	0.7 ± 0.0
22	100.0 ± 0.0
24	94 ± 8
25	46 ± 15
27	70 ± 13
28	60 ± 2
29	61 ± 4

**A.****B.**

**Figure S5:** Anticancer activity of alkyl-7-hydroxy-K252c analogues. Individual cytotoxicity curves (duplicate) for the 7-hydroxy-K252c parent compound (**56**) and analogues (**57-62**) in HCT-116 cells. **(B)** Cytotoxicity curves for the 7-hydroxy-K252c parent compound and analogues (**56-62**) in HCT-116 cells. Error bars represent standard deviation within one independent experiment.

**Supporting Information for**

**FgaPT2, a biocatalytic tool for alkyl-diversification of indole natural products**

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Department of Chemistry and Biochemistry, University of Oklahoma, Stephenson Life Sciences Research Center, 101 Stephenson Parkway, Norman, Oklahoma 73019, United States.

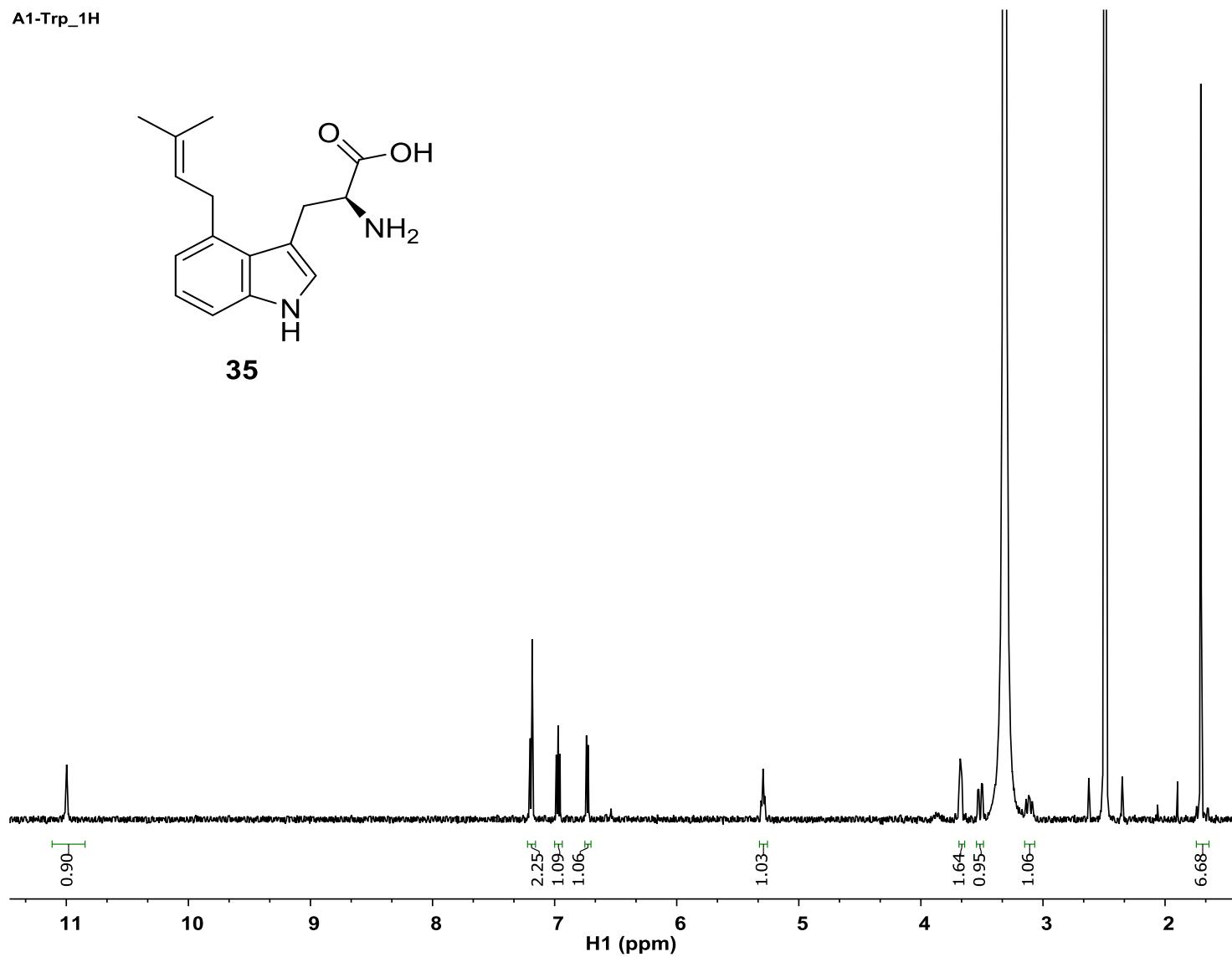
E-mail: [shanteri.singh@ou.edu](mailto:shanteri.singh@ou.edu)

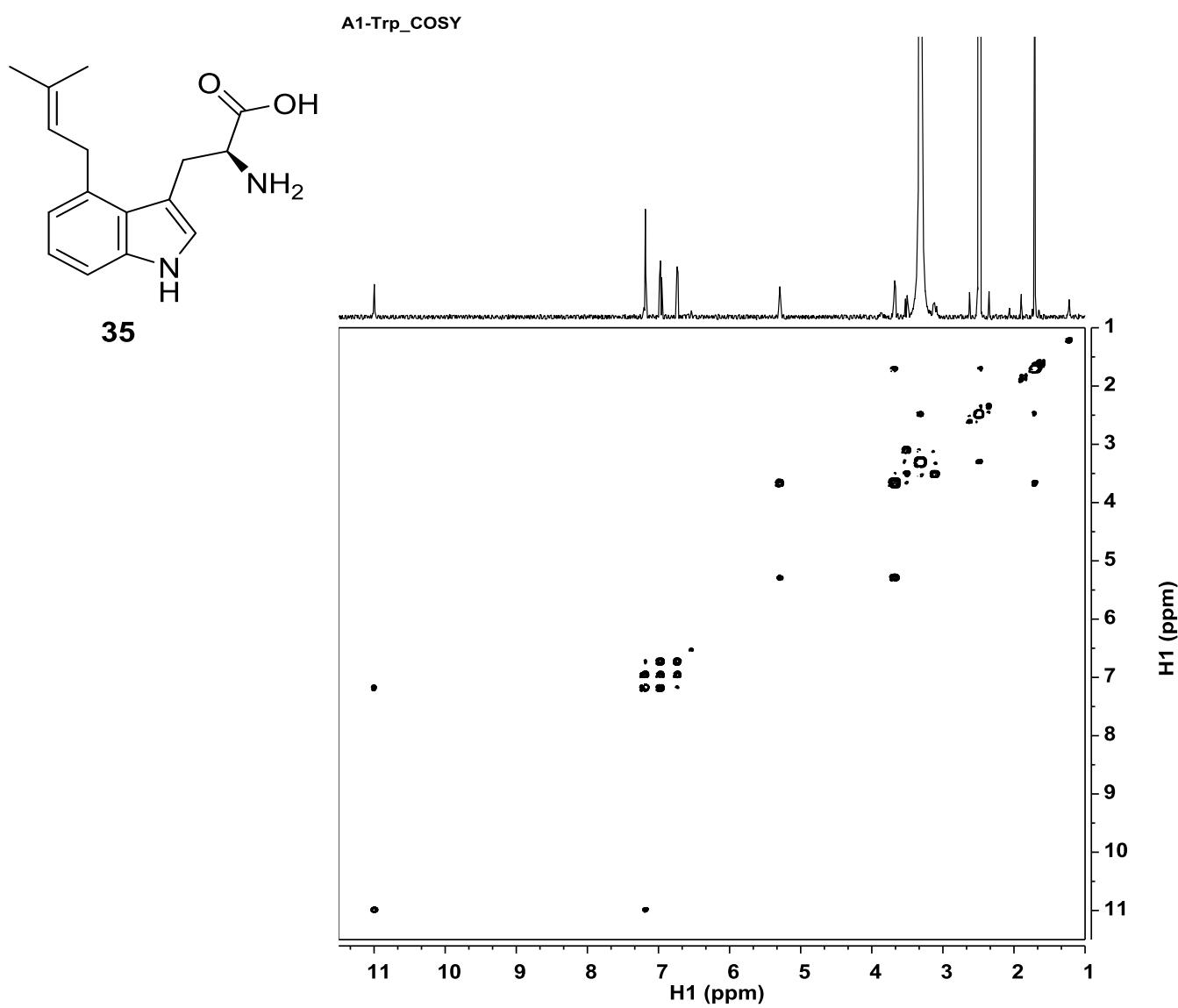
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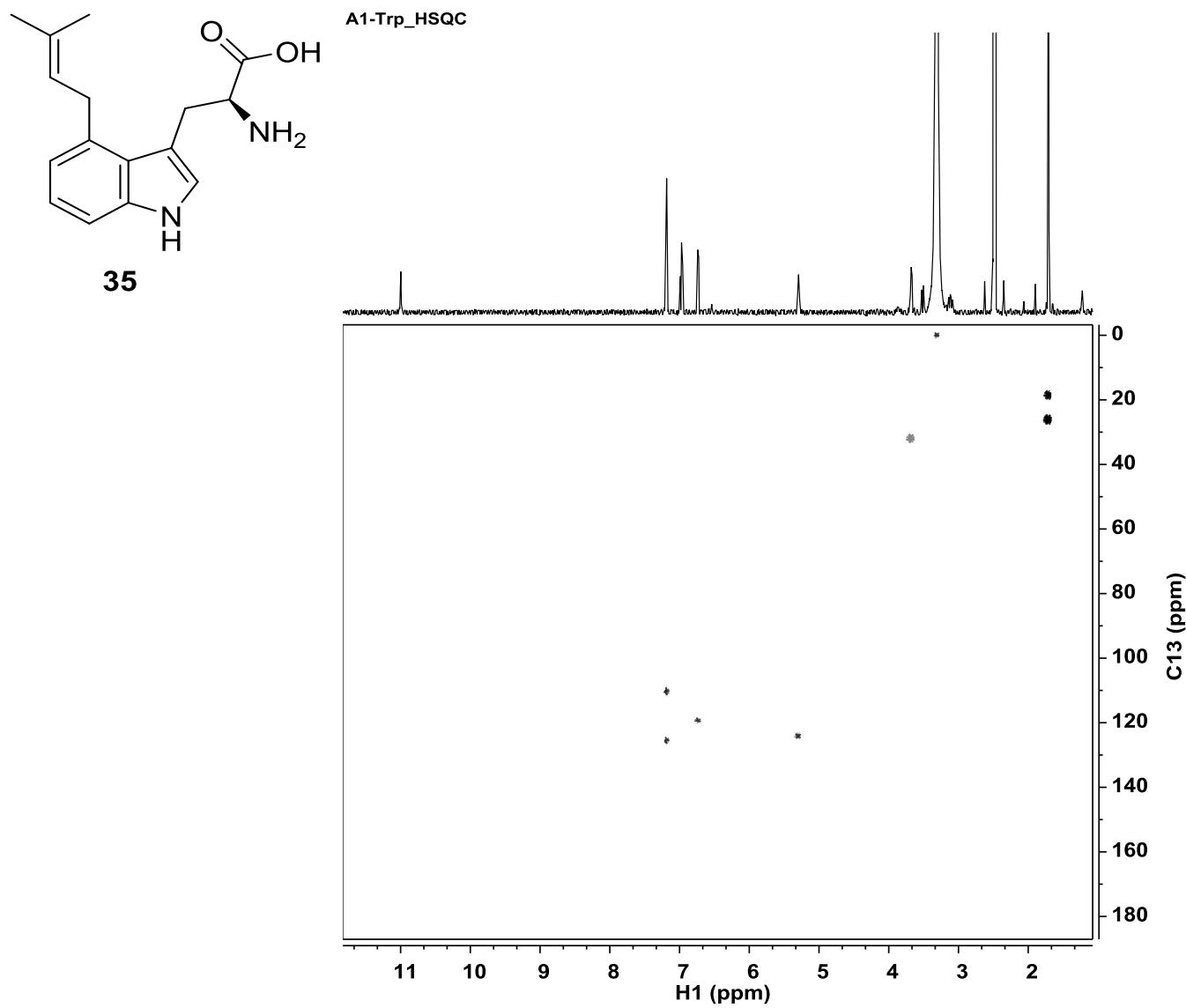
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3. NMR and HRMS Data of <b>37</b>	S11-14
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9. NMR and HRMS Data of <b>40</b>	S35-38
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23. HRMS Data of <b>13-Trp</b>	S70

24. HRMS Data of <b>14-Trp</b>	S71
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42. NMR and HRMS data of <b>9</b>	S138-140
43. NMR and HRMS data of <b>22</b>	S141-143
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A1-Trp\_1H

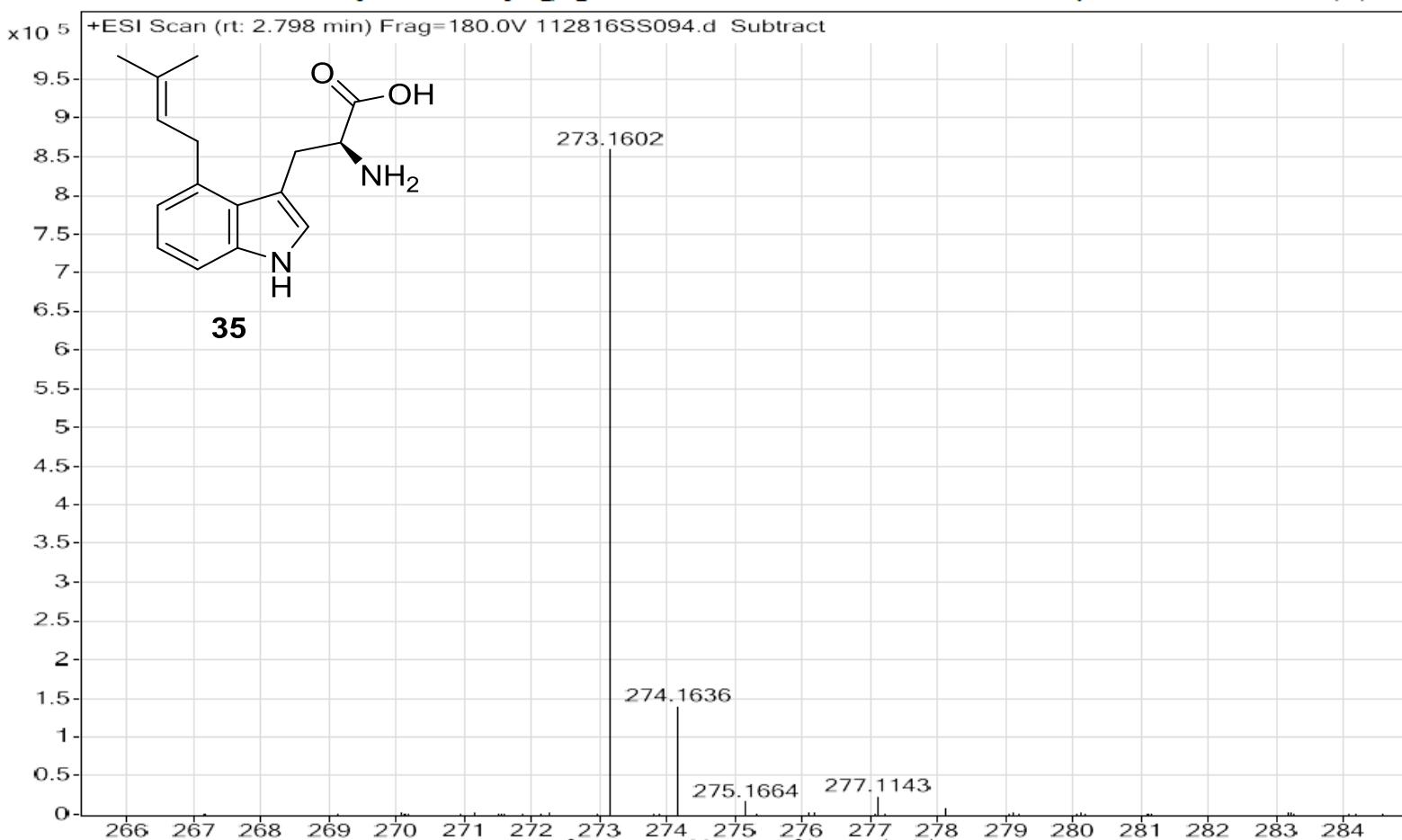






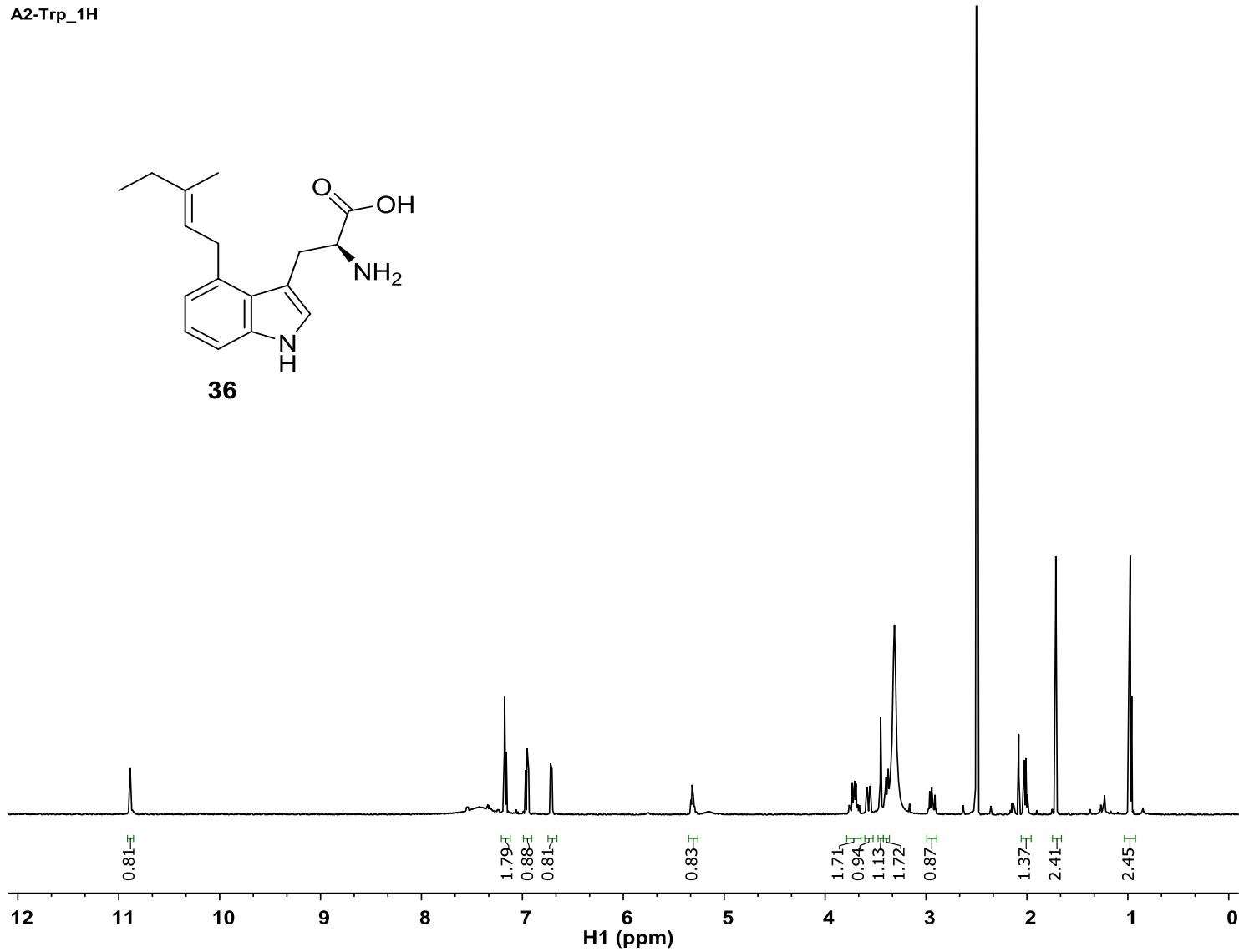
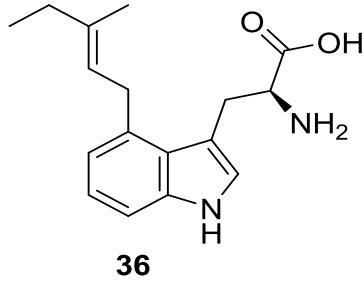
2D  $^1\text{H}$ - $^{13}\text{C}$  HSQC NMR spectrum (500 MHz) of **35** in  $\text{DMSO-d}_6$

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Inj Vol	5	InjPosition		SampleType	Sample	IRM Calibration Status	Success
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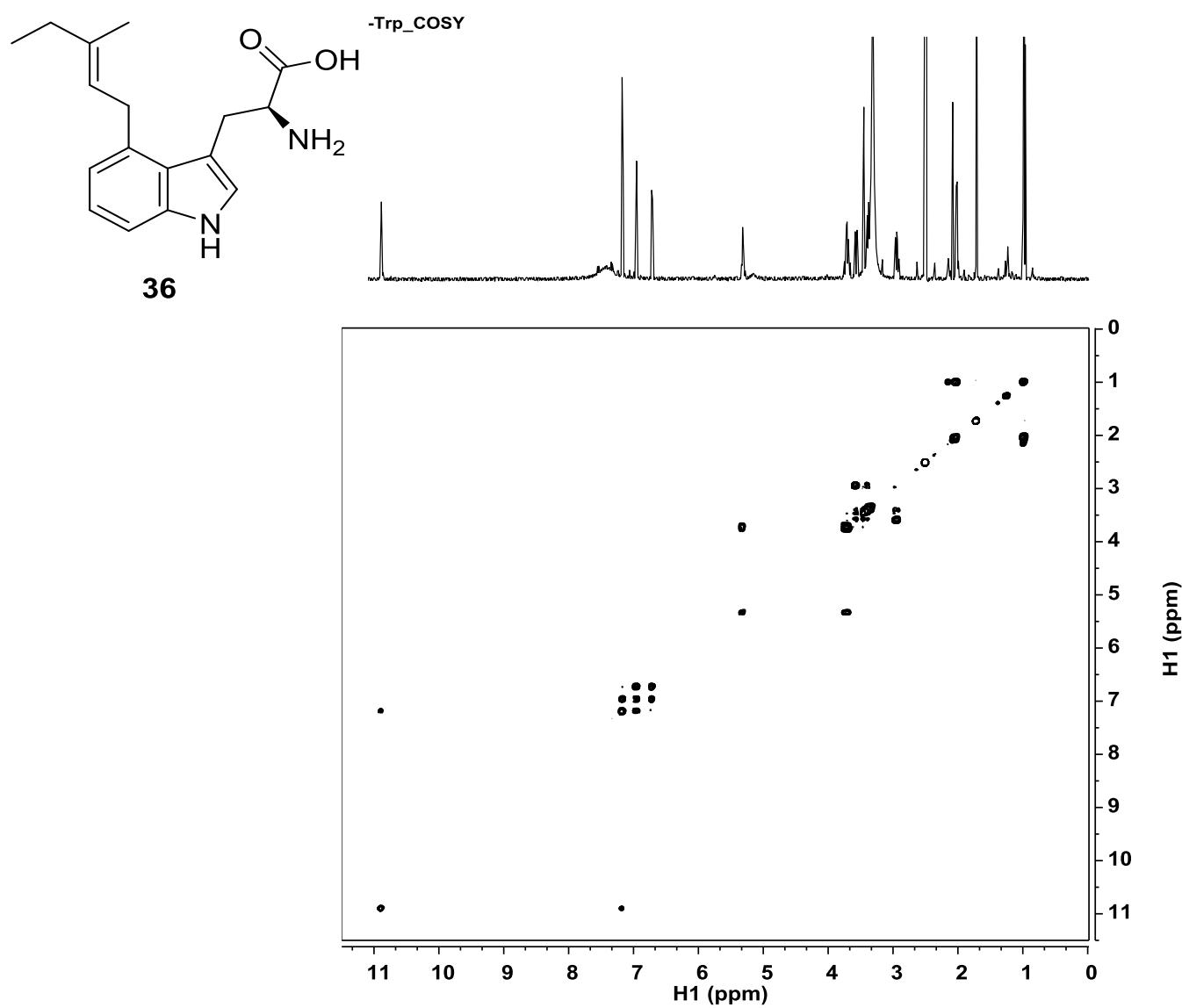


HRMS spectrum of 35

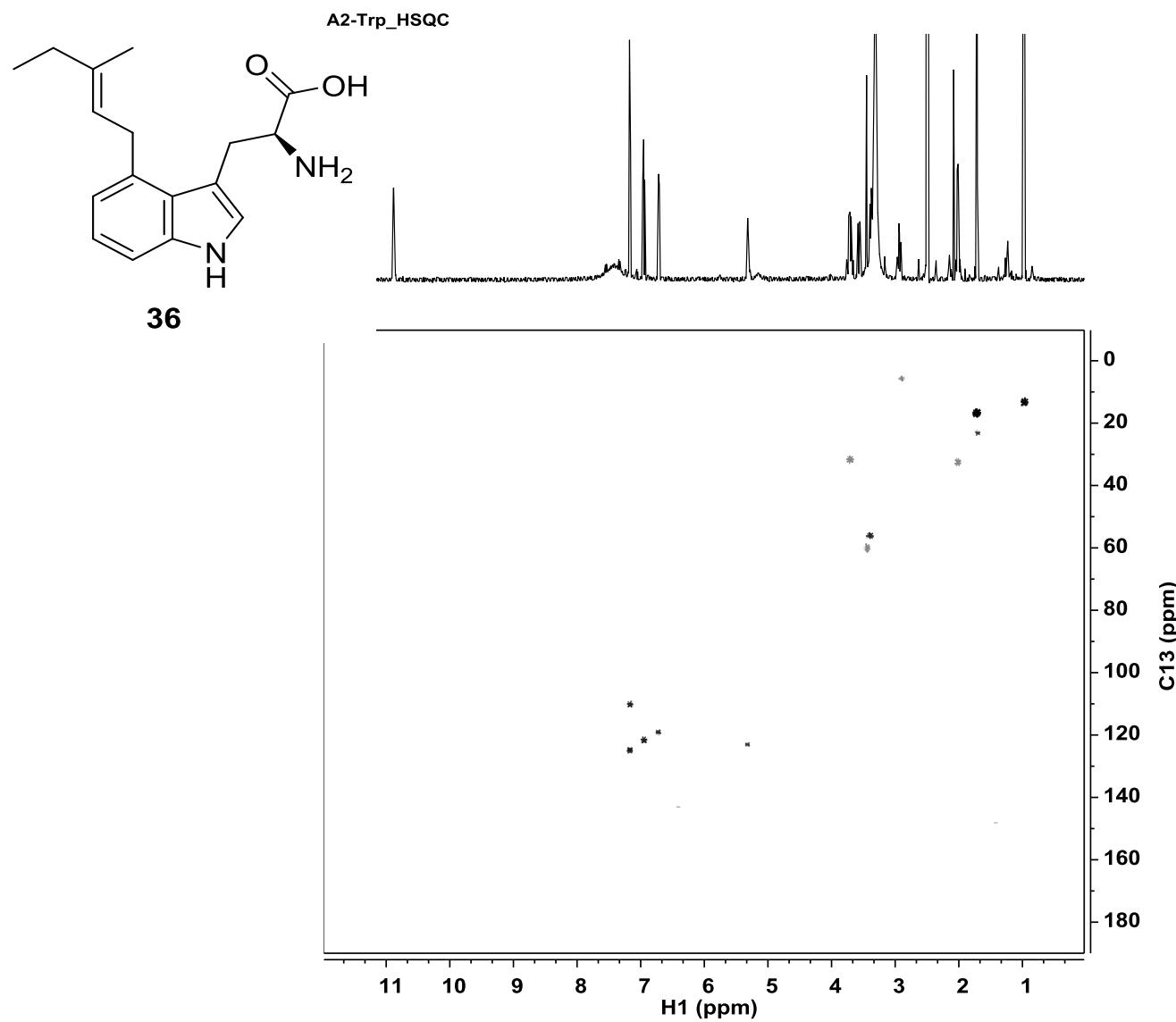
A2-Trp\_1H



<sup>1</sup>H NMR Spectrum (500 MHz) of compound **36** in DMSO-d<sub>6</sub>

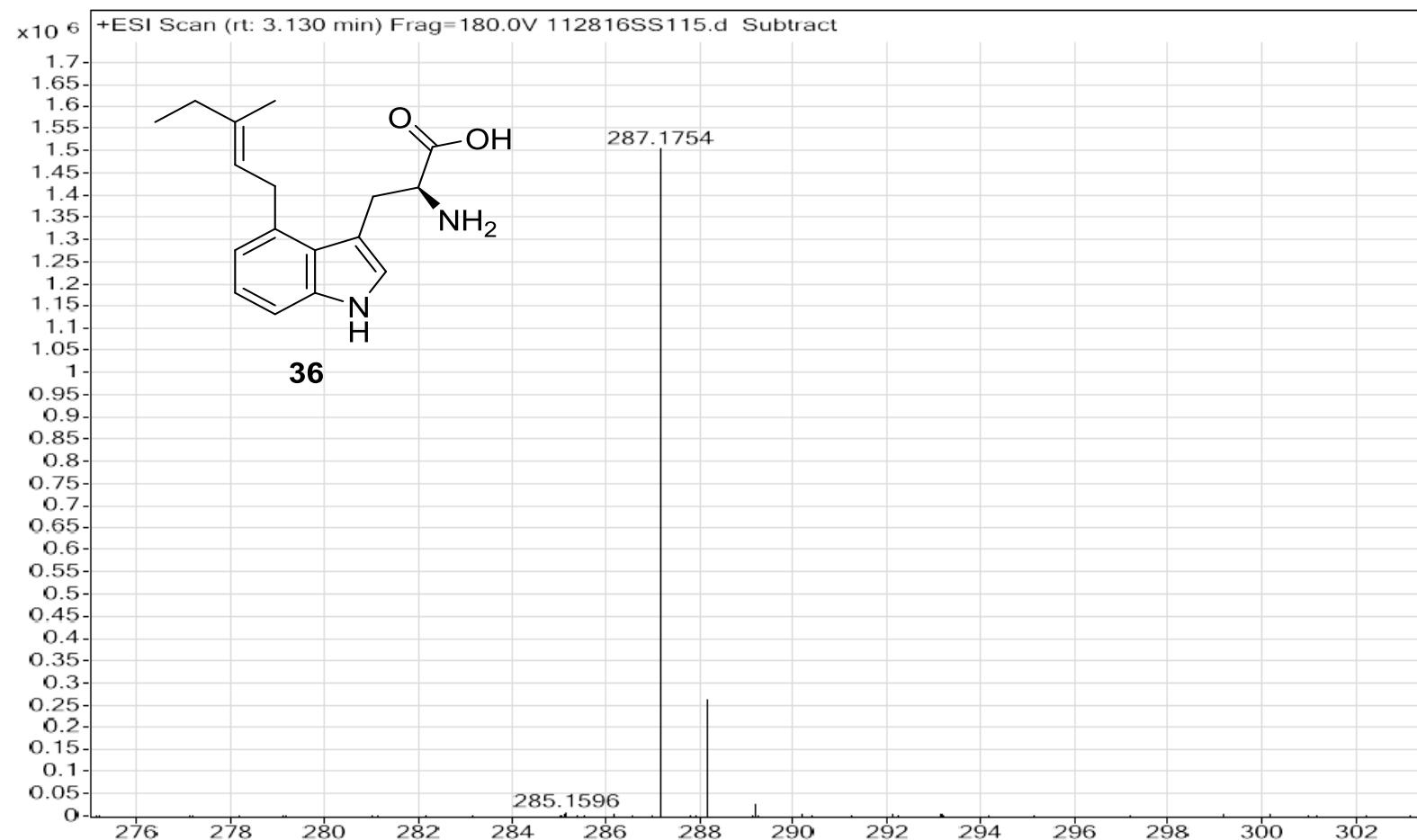


2D  $^1\text{H}$ - $^1\text{H}$  COSY NMR spectrum (500 MHz) of **36** in DMSO-d<sub>6</sub>



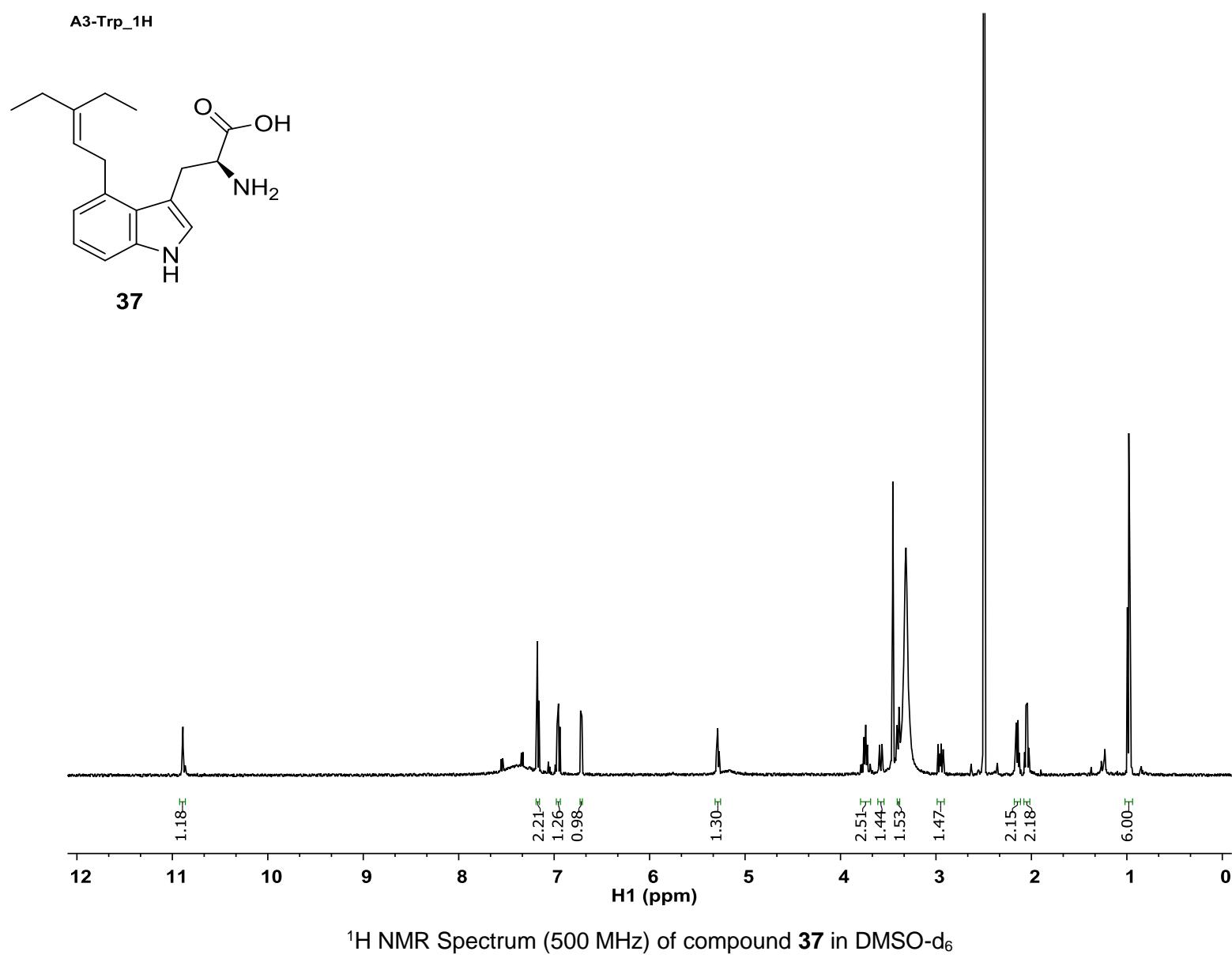
2D  $^1\text{H}$ - $^{13}\text{C}$  HSQC NMR spectrum (500 MHz) of **36** in  $\text{DMSO-d}_6$

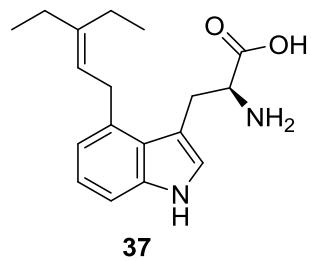
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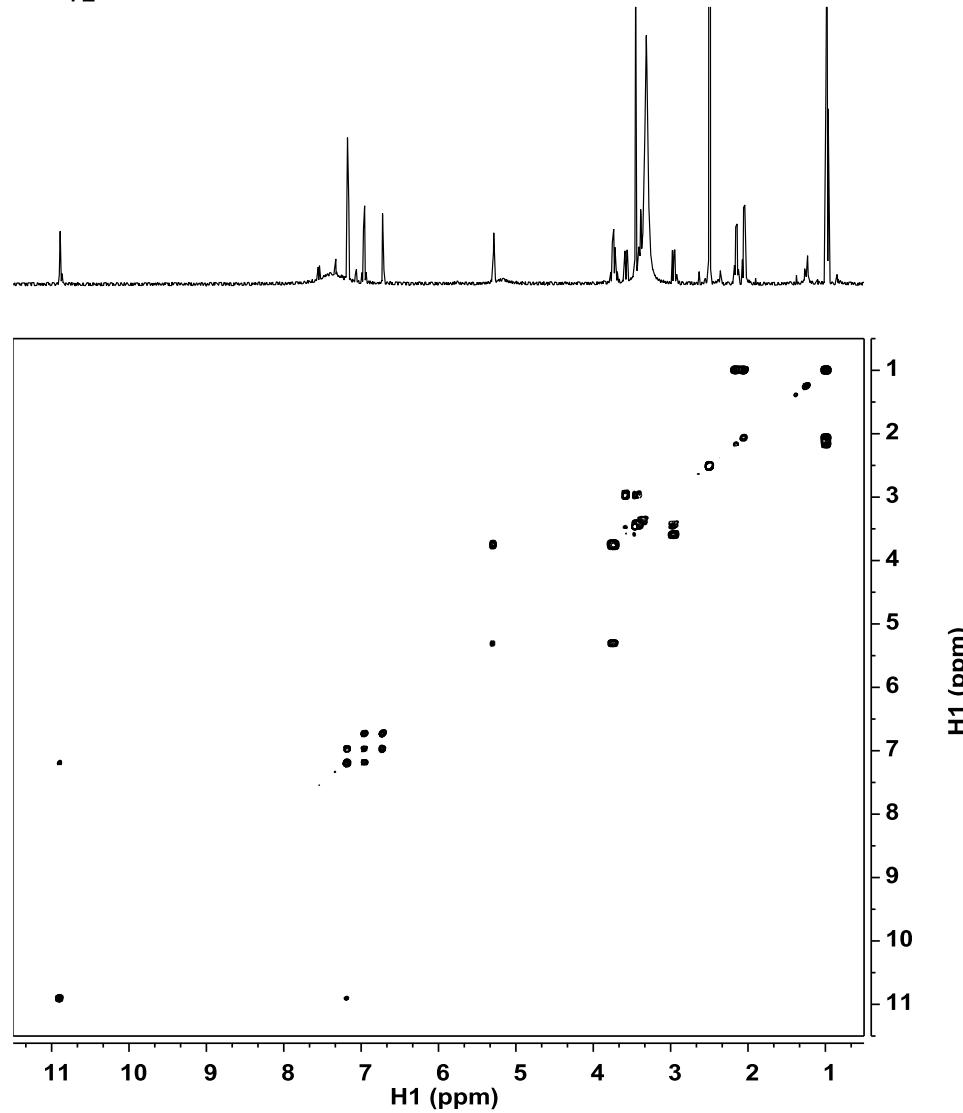
HRMS spectrum of **36**

A3-Trp\_1H

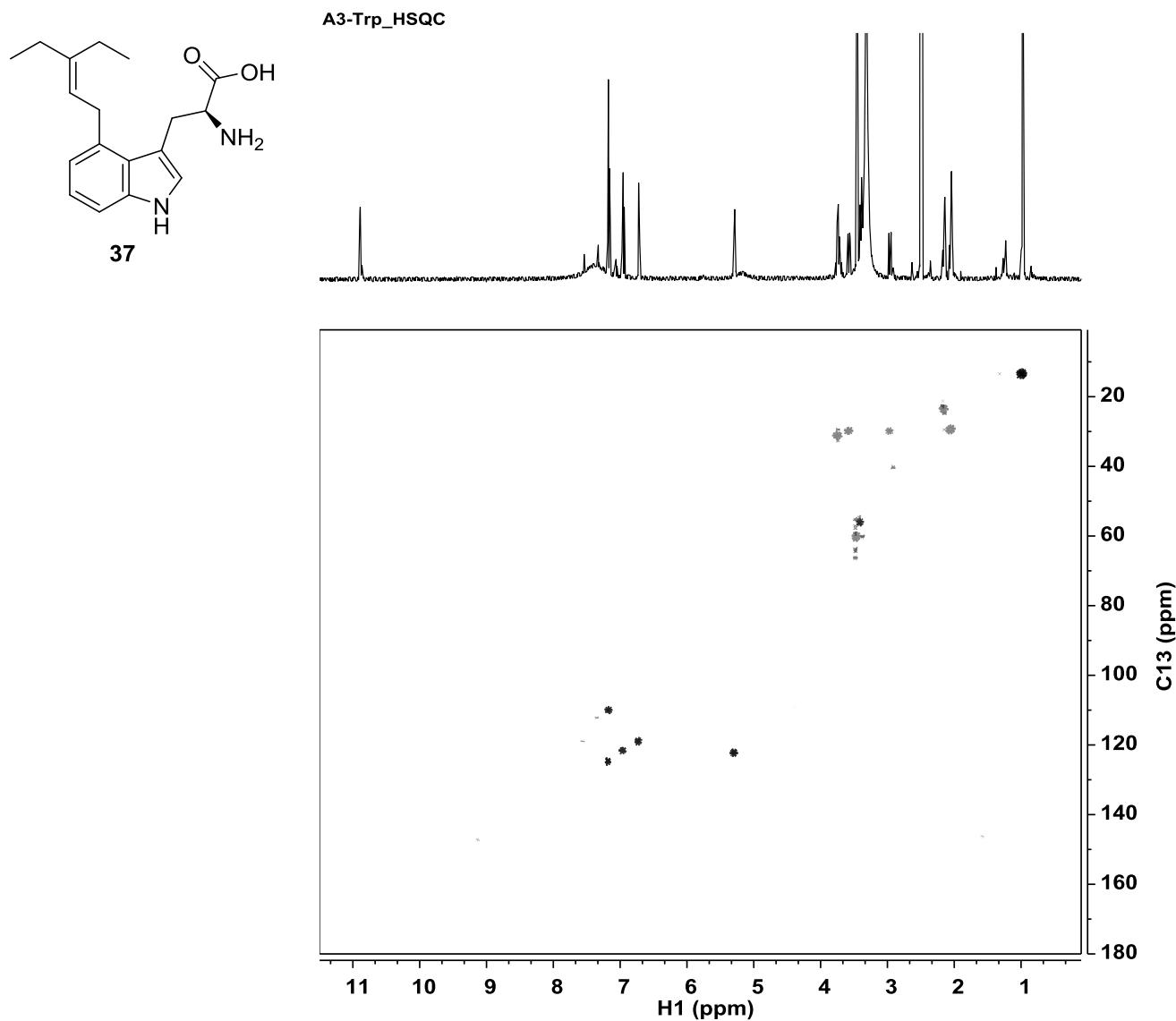




A3-Trp\_COSY

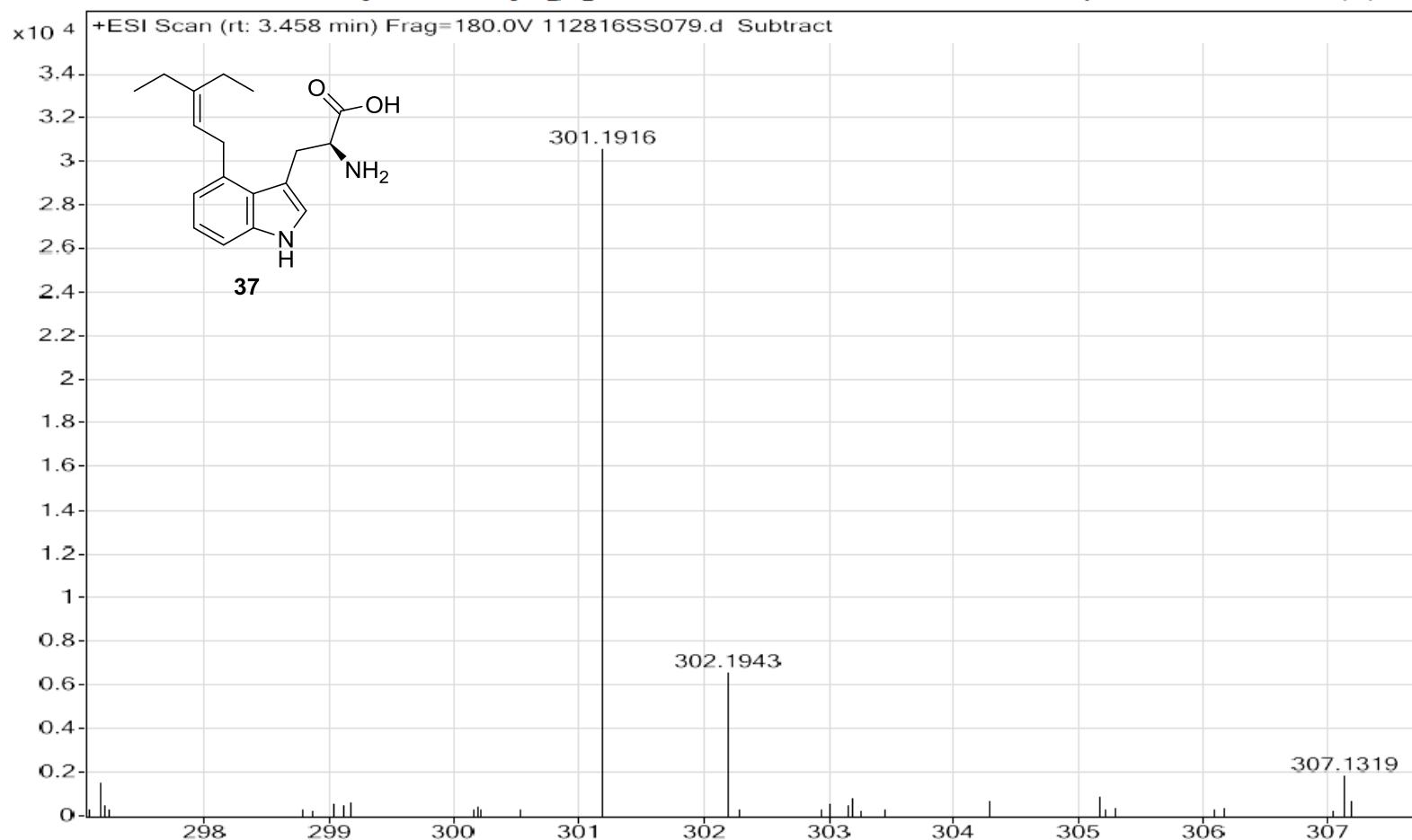


2D  $^1\text{H}$ - $^1\text{H}$  COSY NMR spectrum (500 MHz) of 37 in  $\text{DMSO-d}_6$



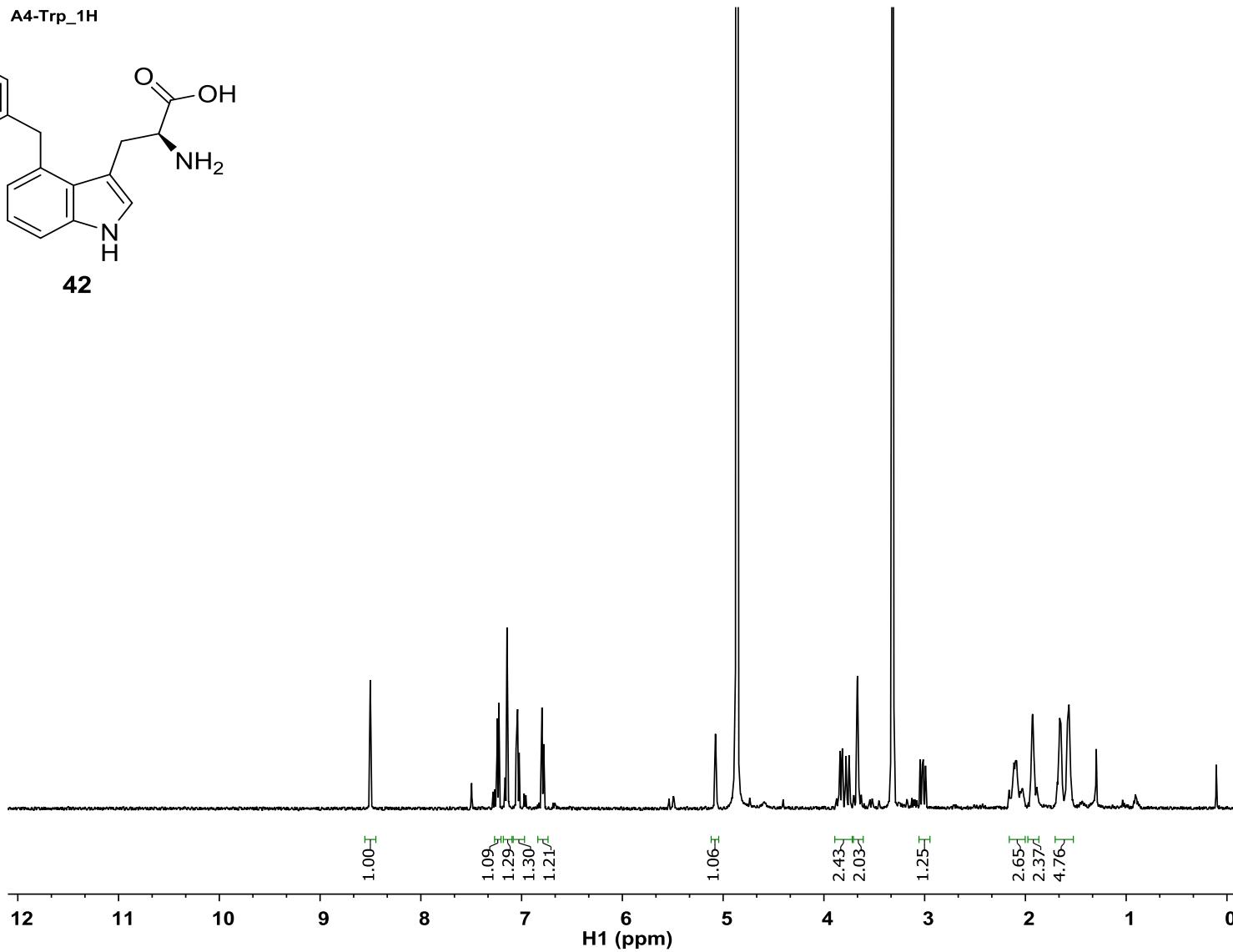
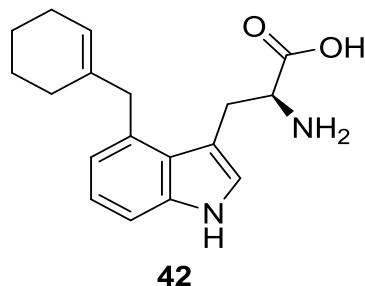
2D  $^1\text{H}$ - $^{13}\text{C}$  HSQC NMR spectrum (500 MHz) of **37** in  $\text{DMSO-d}_6$

Sample Name	TRP-25	Position	P1-C7	Instrument Name	Instrument 1	User Name	
Inj Vol	5	InjPosition		SampleType	Sample	IRM Calibration Status	Success
Data Filename	112816SS079.d	ACQ Method	drugC18_pos_ms.m	Comment		Acquired Time	11/29/2016 2:48:13 AM

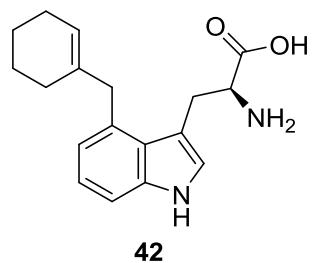


HRMS spectrum of 37

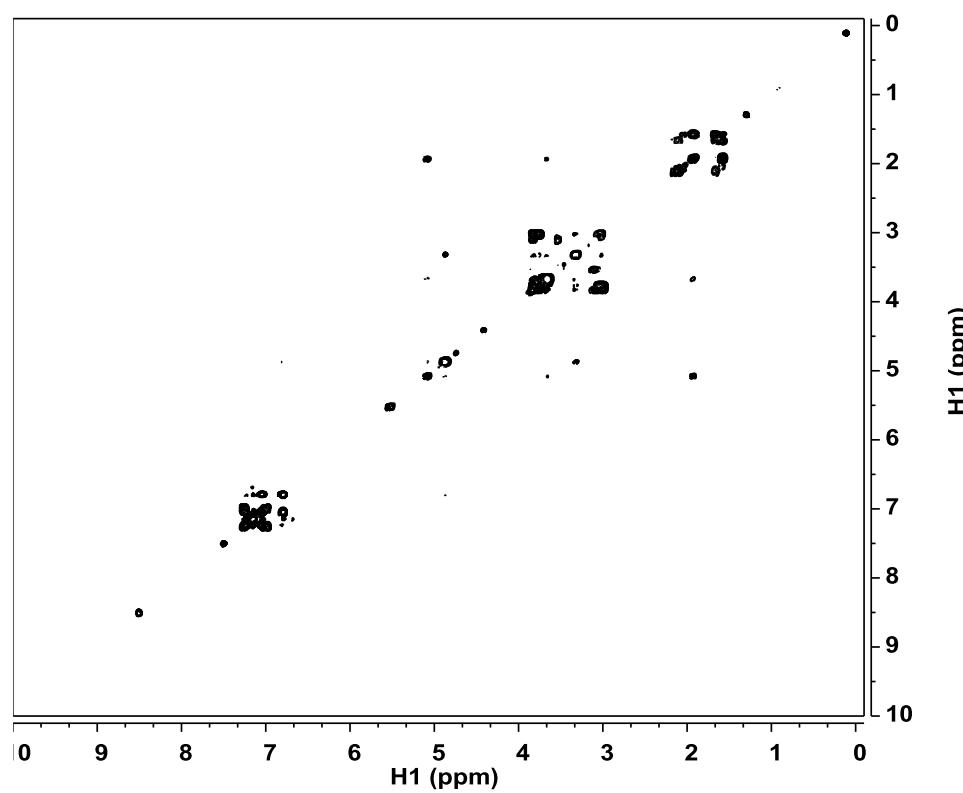
A4-Trp\_1H



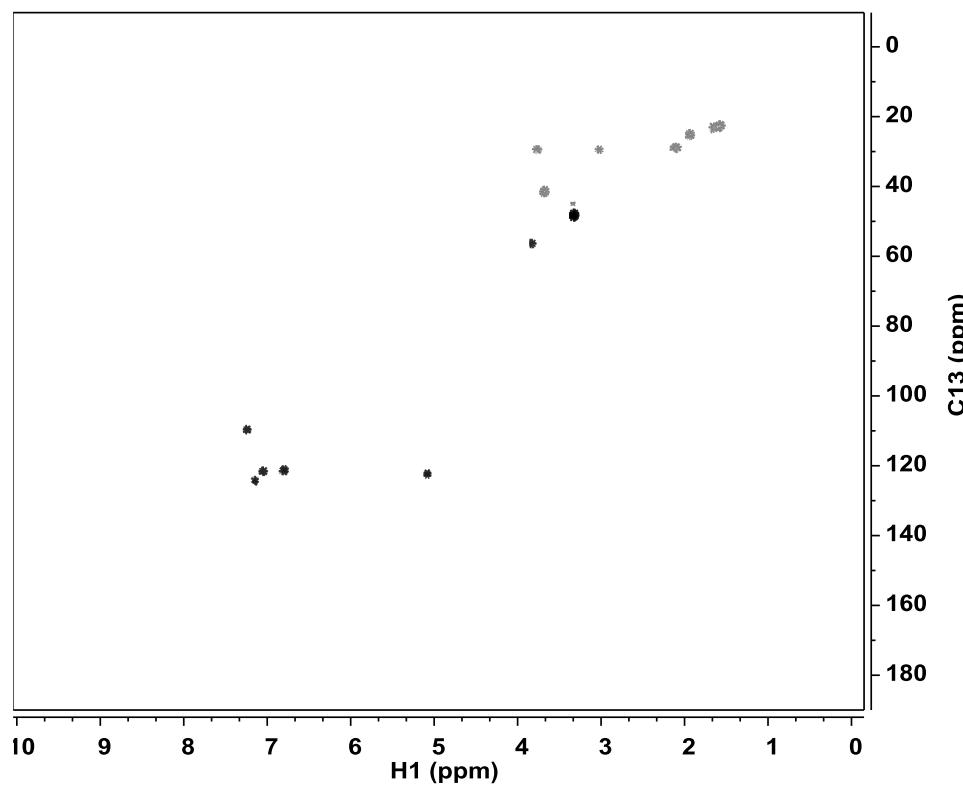
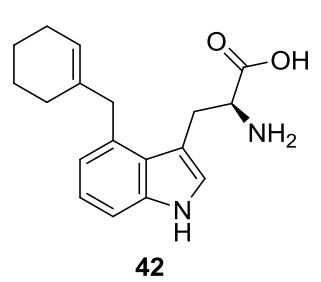
<sup>1</sup>H NMR Spectrum (500 MHz) of compound **42** in MeOH-d<sub>4</sub>



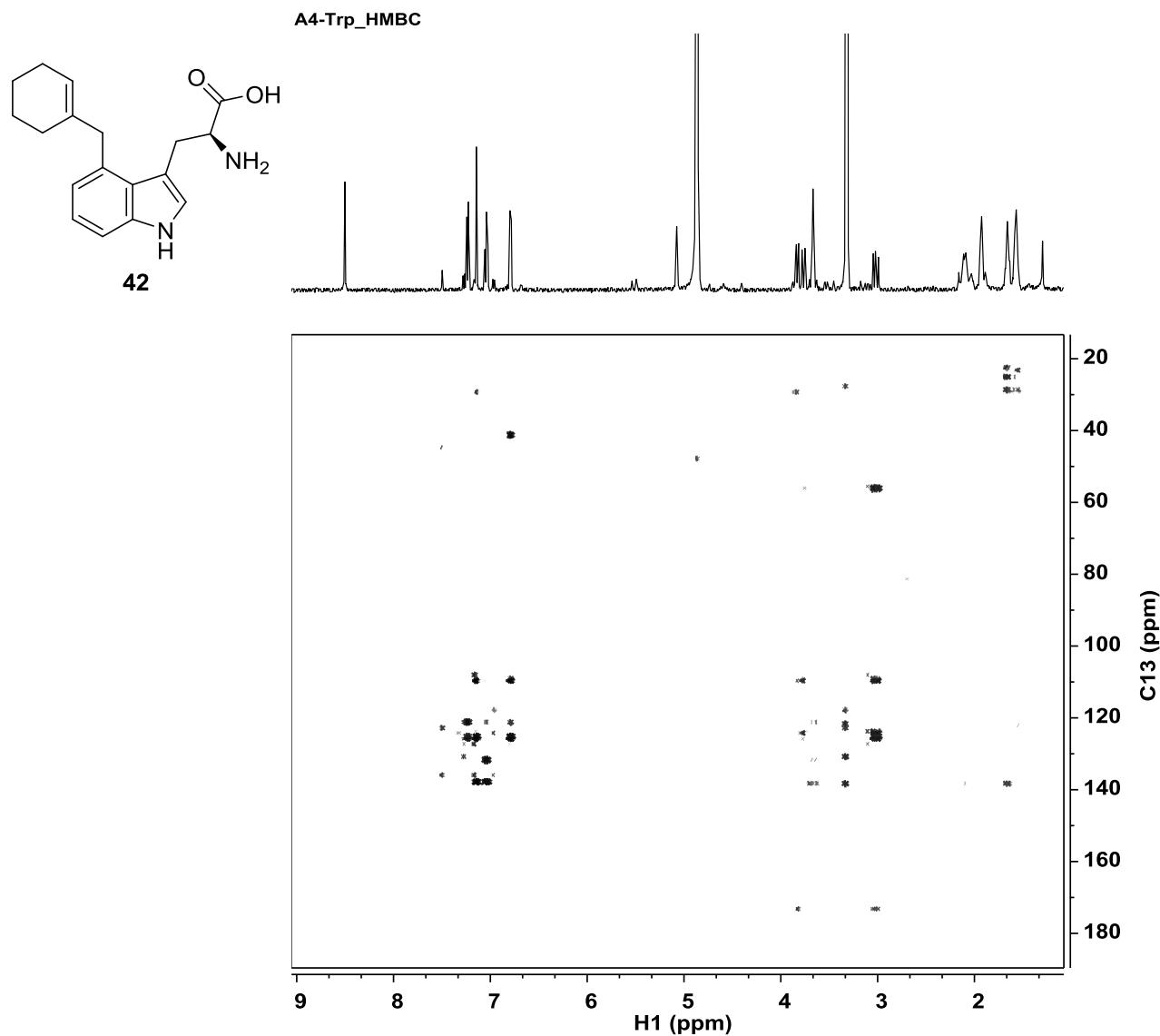
A4-Trp\_COSY



2D  $^1\text{H}$ - $^1\text{H}$  COSY NMR spectrum (500 MHz) of **42** in  $\text{MeOH-d}_4$

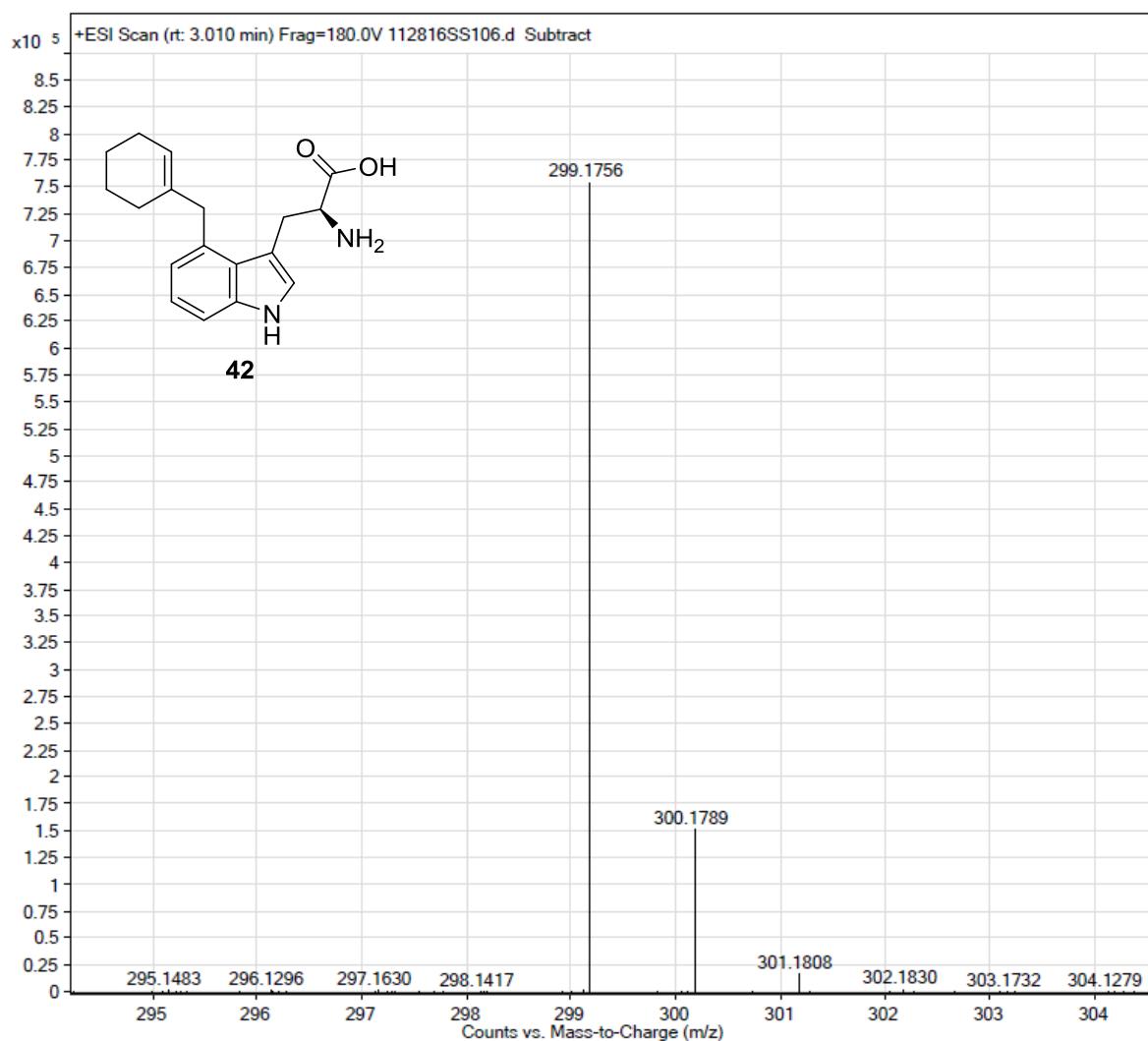


2D  $^1\text{H}$ - $^{13}\text{C}$  HSQC NMR spectrum (500 MHz) of **42** in  $\text{MeOH-d}_4$



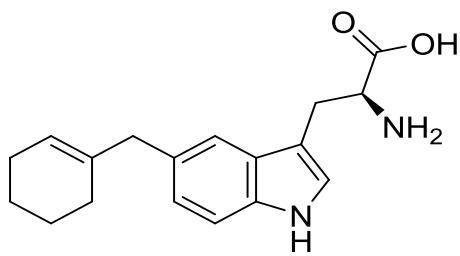
2D  $^1\text{H}$ - $^{13}\text{C}$  HMBC NMR spectrum (500 MHz) of **42** in  $\text{MeOH-d}_4$

<b>Sample Name</b>	TRP-34	<b>Position</b>	P1-D7	<b>Instrument Name</b>	Instrument 1
<b>User Name</b>		<b>Inj Vol</b>	5	<b>InjPosition</b>	
<b>Sample Type</b>	Sample	<b>IRM Calibration Status</b>	Success	<b>Data Filename</b>	11281655106.d
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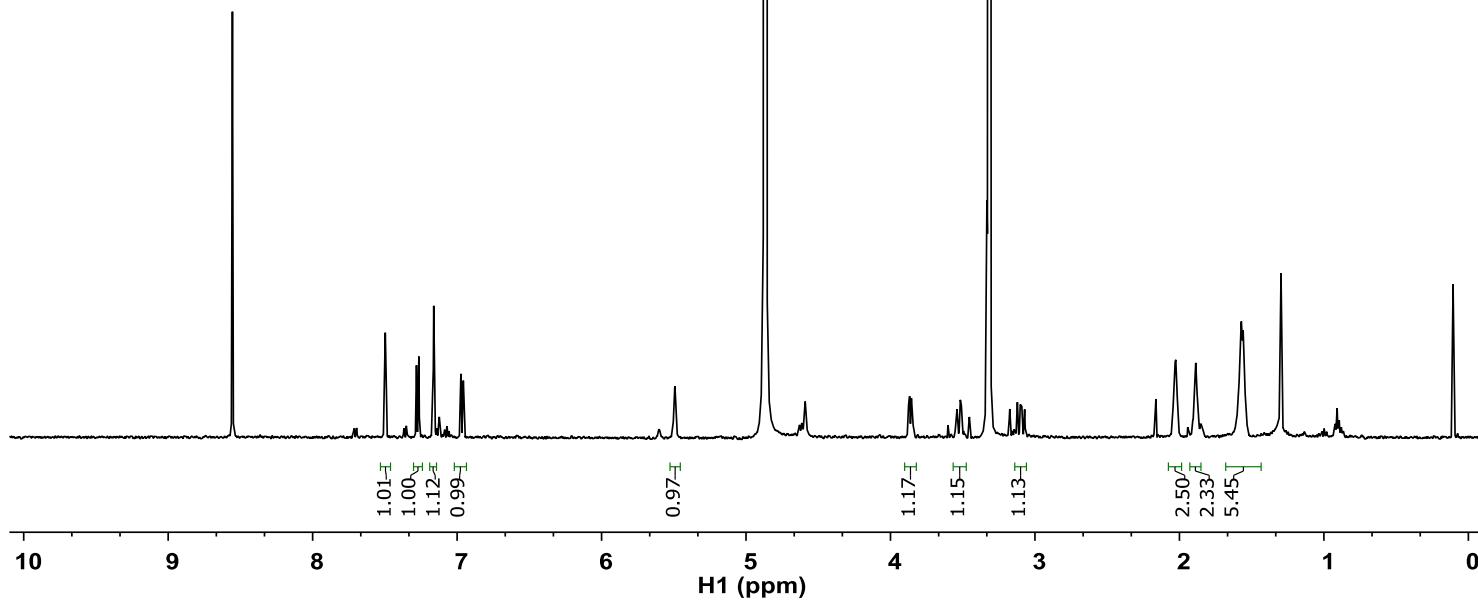


### HRMS spectrum of **42**

A4-Trp2\_1H

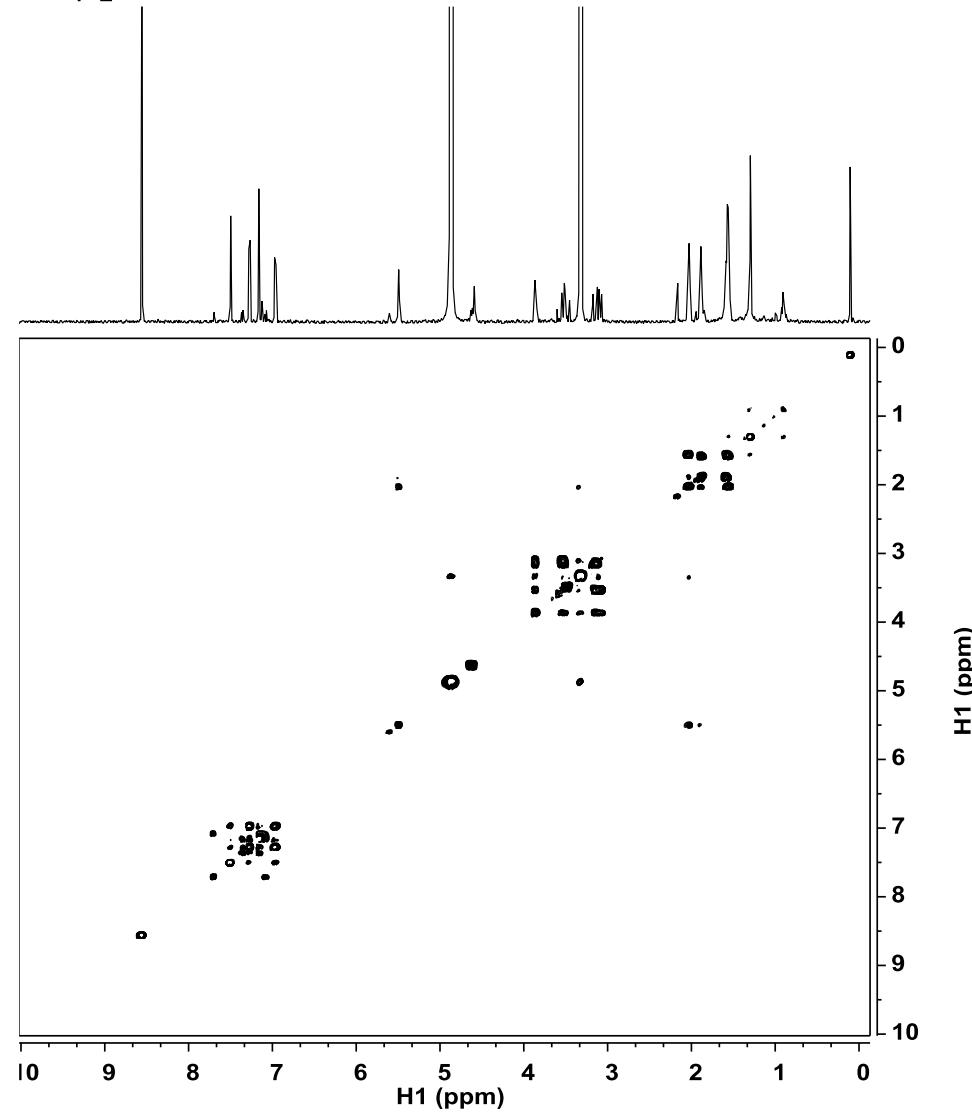


**43**



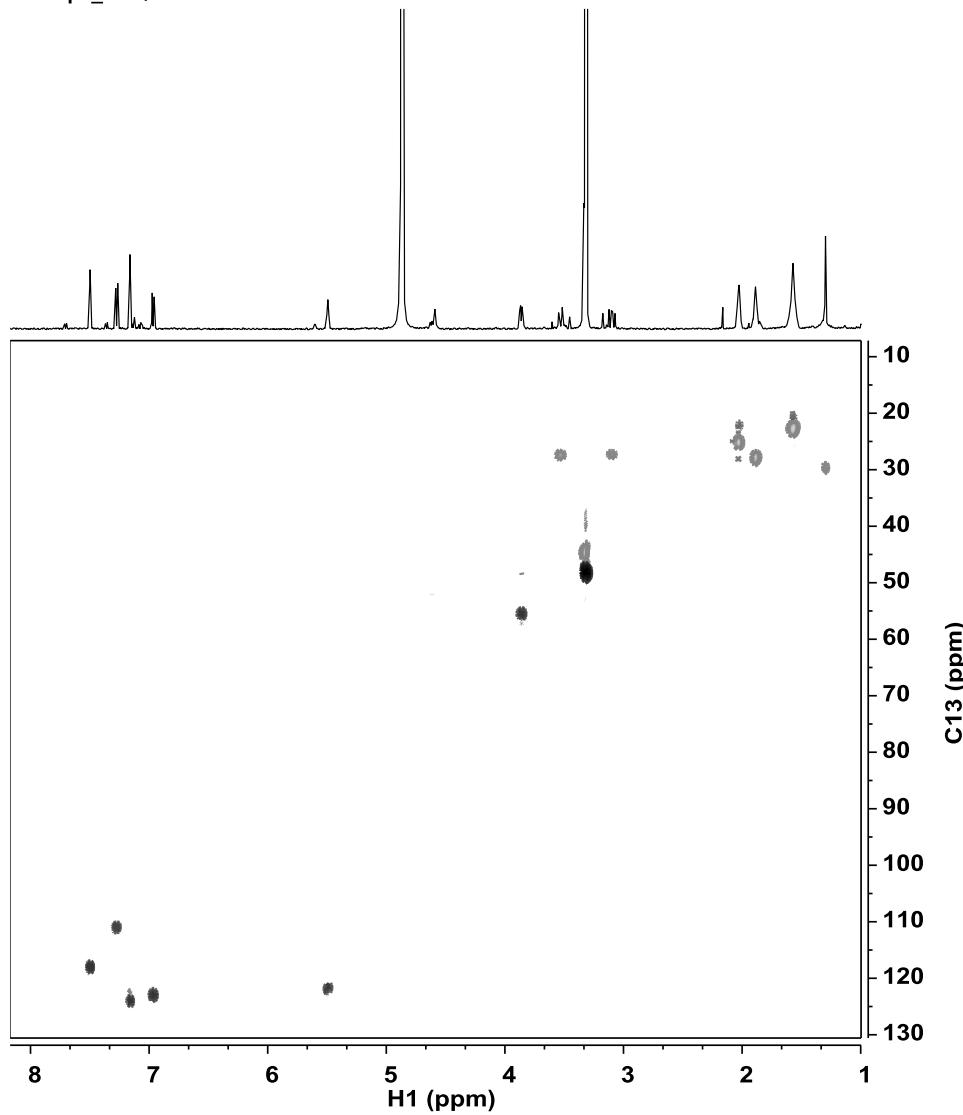
<sup>1</sup>H NMR spectrum (500 MHz) of **43** in MeOH-d<sub>4</sub>

A4-Trp2\_COSY



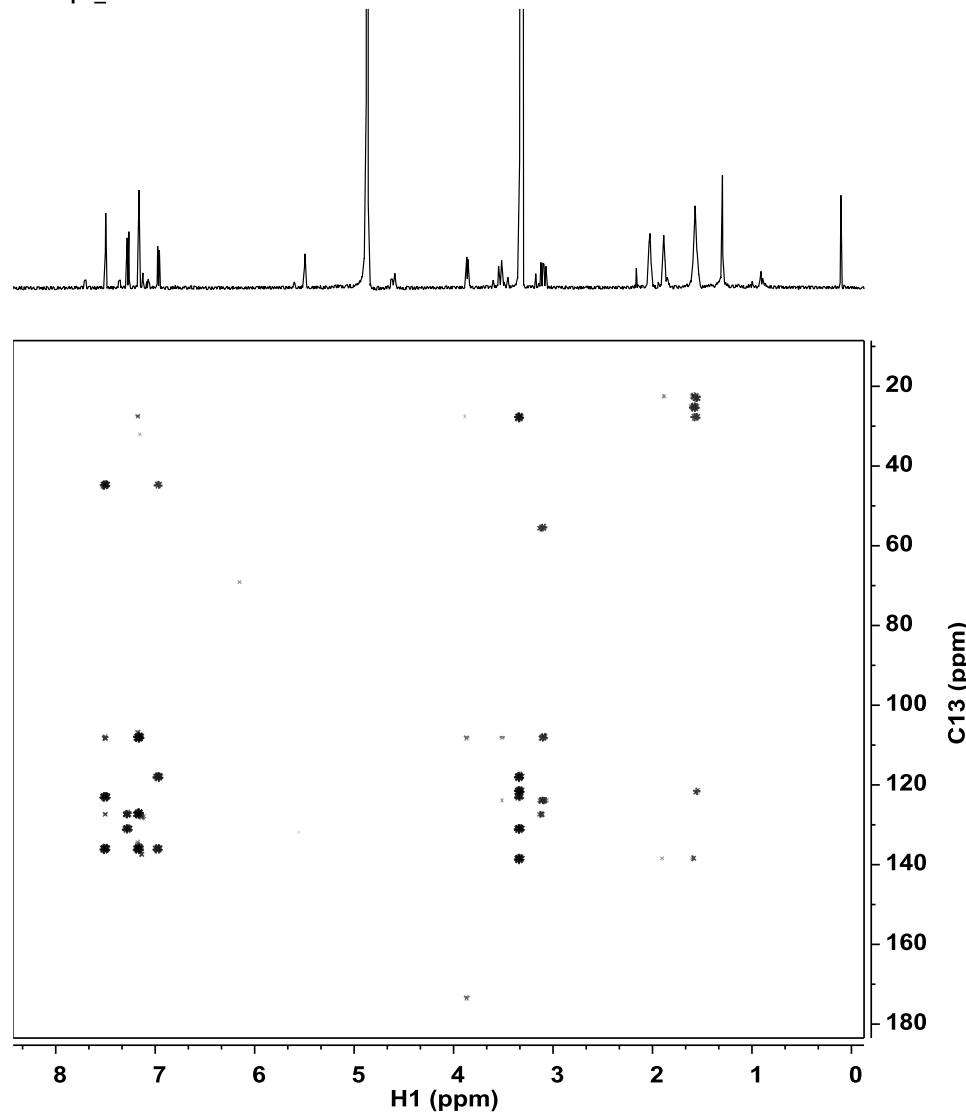
2D  $^1\text{H}$ - $^1\text{H}$  COSY NMR spectrum (500 MHz) of **43** in  $\text{MeOH-d}_4$

A4-Trp2\_HSQC



2D  $^1\text{H}$ - $^{13}\text{C}$  HSQC NMR spectrum (500 MHz) of **43** in  $\text{MeOH-d}_4$

A4-Trp2\_HMBC

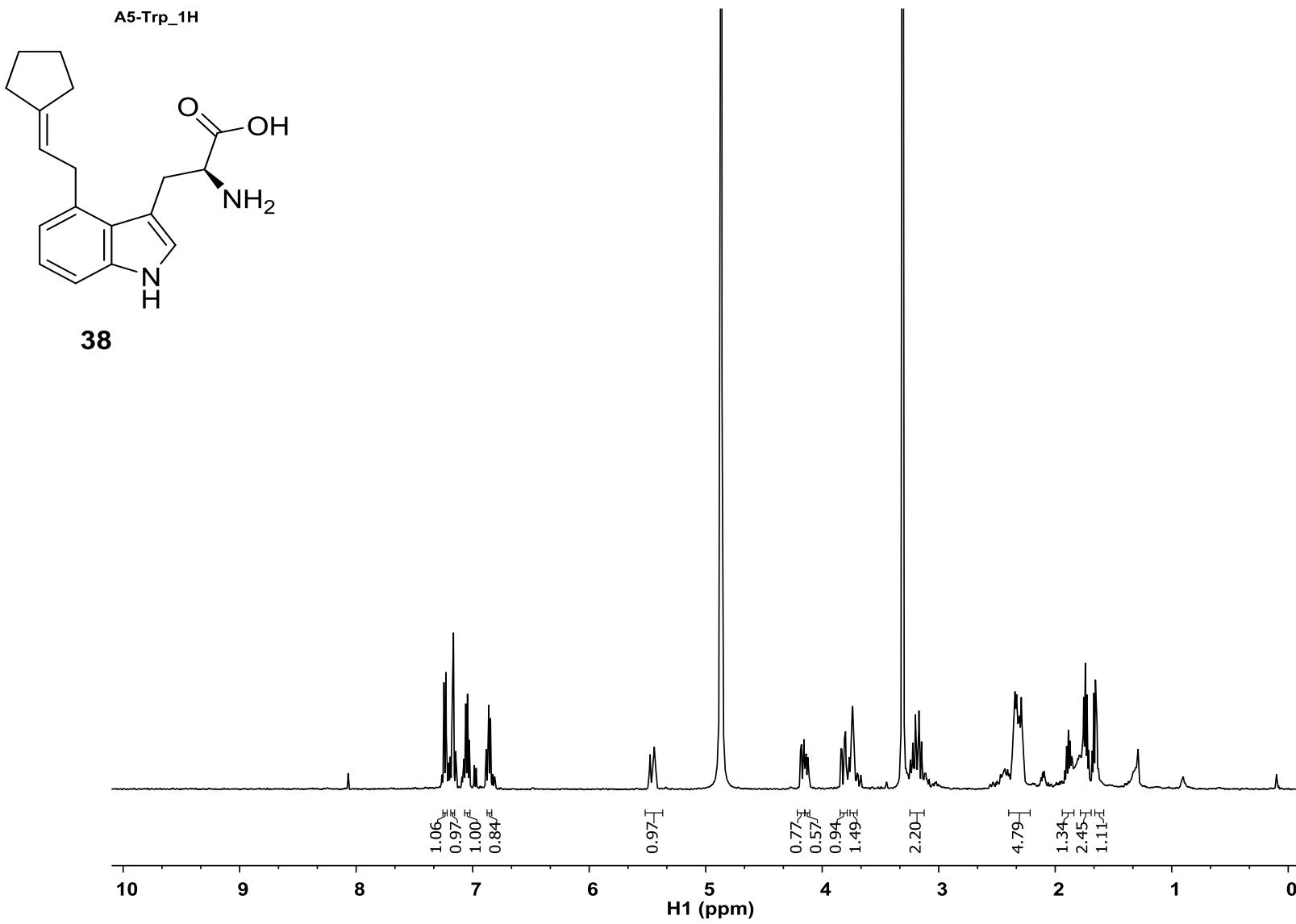


2D  $^1\text{H}$ - $^{13}\text{C}$  HMBC NMR spectrum (500 MHz) of **43** in  $\text{MeOH-d}_4$

Sample Name	TRP-34	Position	P1-D7	Instrument Name	Instrument 1	User Name	
Inj Vol	5	InjPosition		SampleType	Sample	IRM Calibration Status	Success
Data Filename	112816SS106.d	ACQ Method	drugC18_pos_ms.m	Comment		Acquired Time	11/29/2016 7:42:31 AM

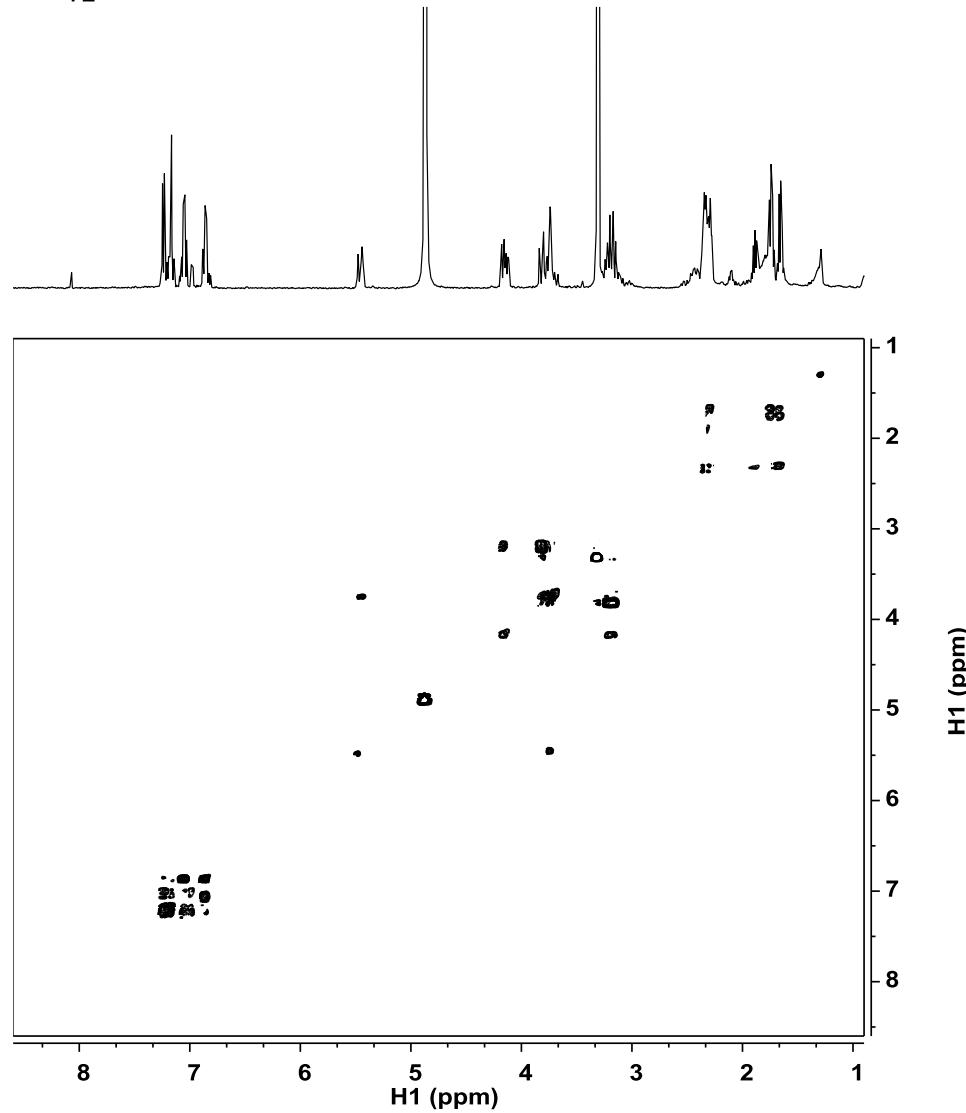


HRMS spectrum of 43

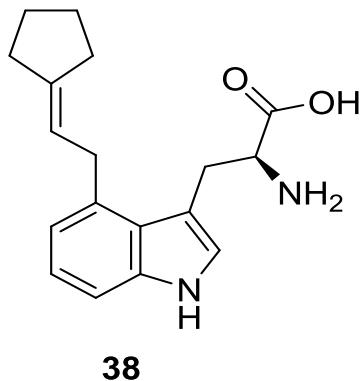


<sup>1</sup>H NMR Spectrum (500 MHz) of compound **38** in MeOH-d<sub>4</sub> (*C4<sup>N</sup>* isomer + 20% unknown isomer)

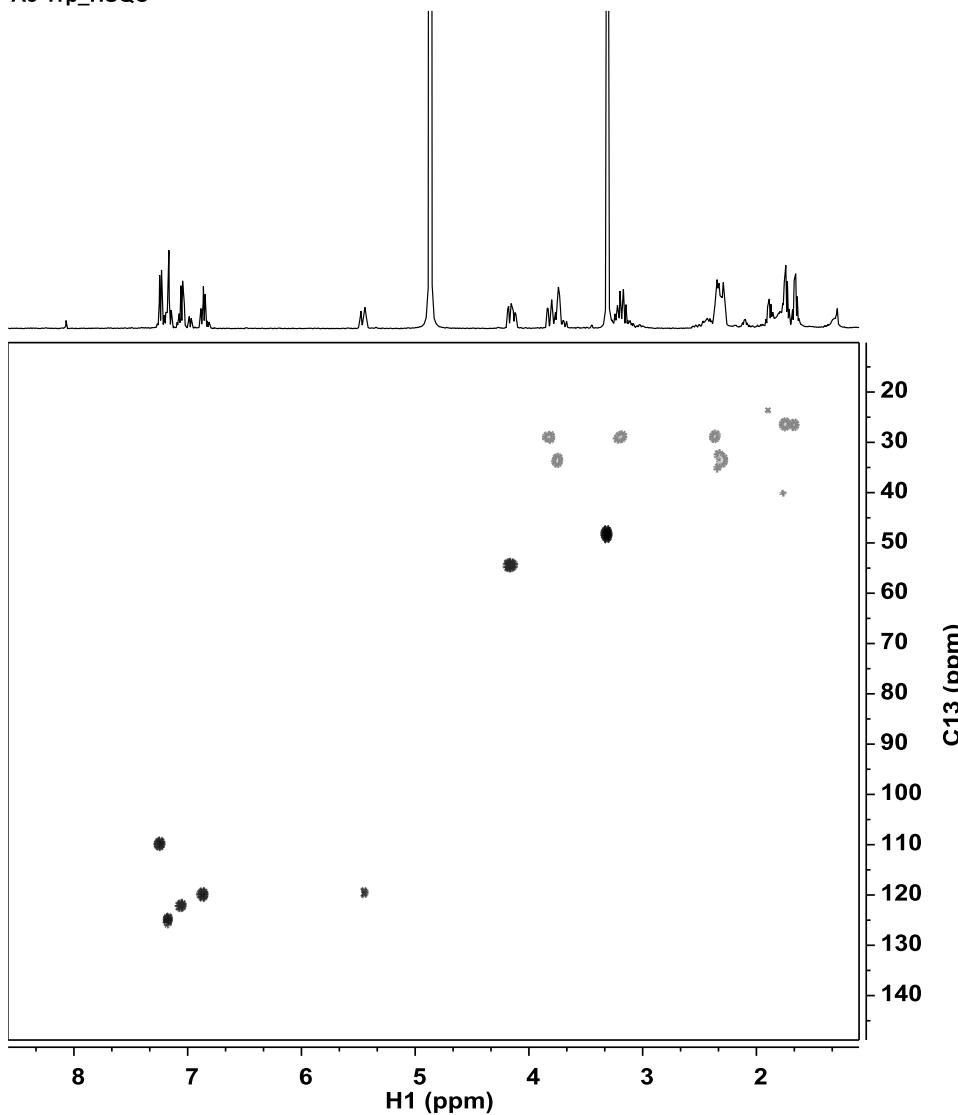
A5-Trp\_COSY



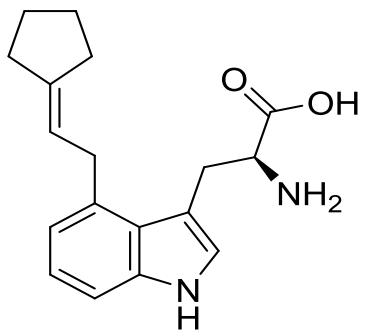
2D  $^1\text{H}$ - $^1\text{H}$  COSY NMR spectrum (500 MHz) of compound **38** in MeOH-d<sub>4</sub> ( $\text{C}4^N$  isomer + 20% unknown isomer)



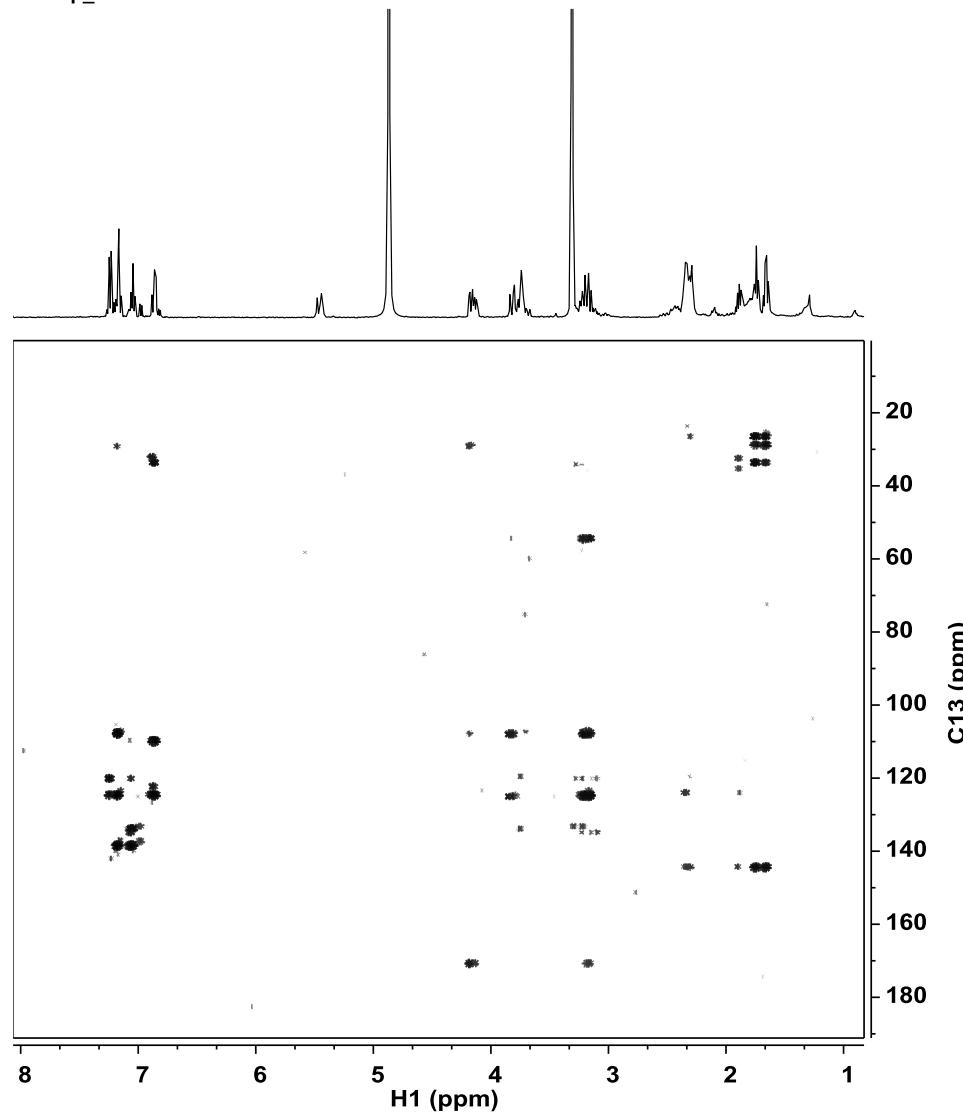
A5-Trp\_HSQC



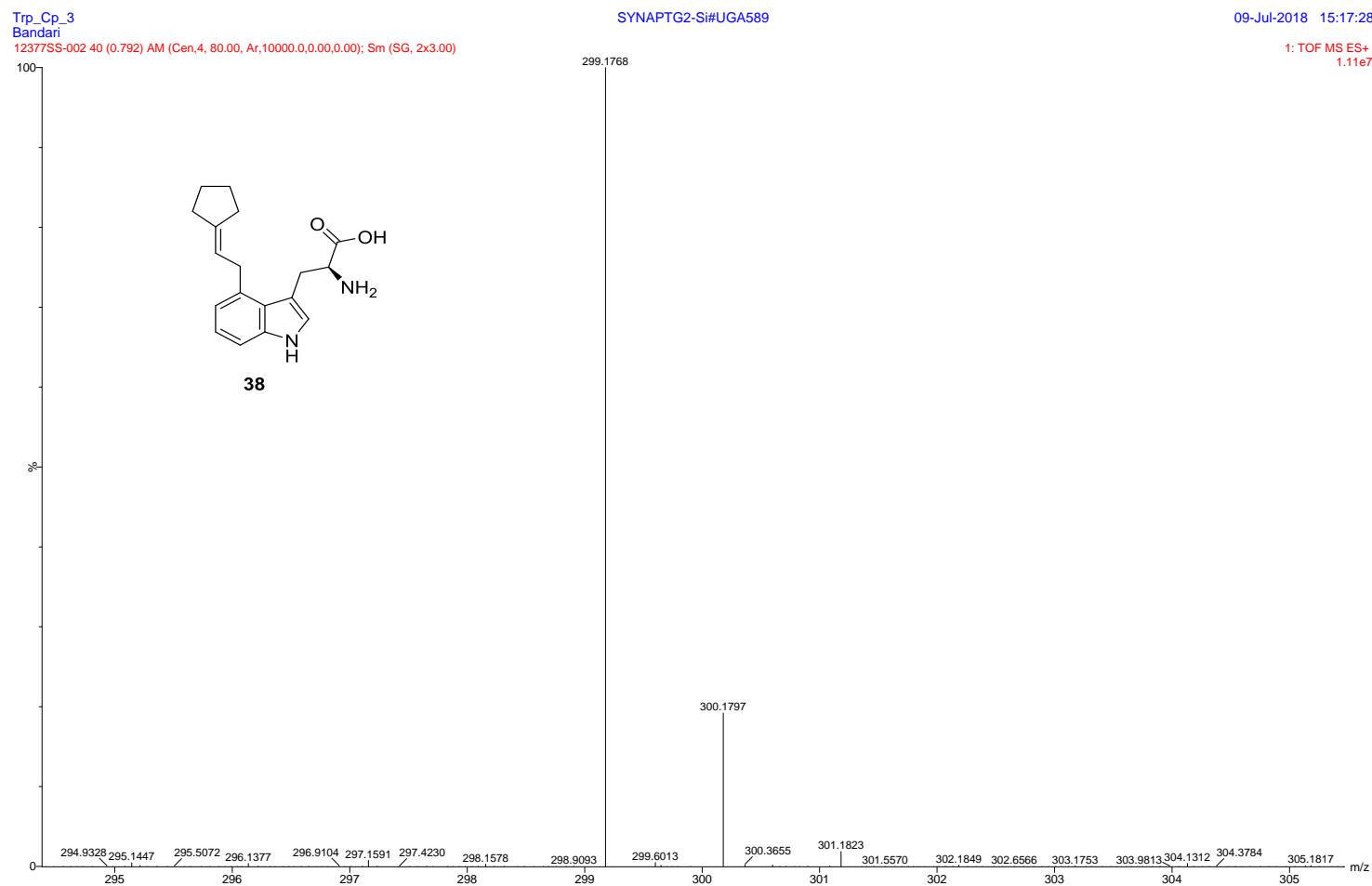
2D  $^1\text{H}$ - $^{13}\text{C}$  HSQC NMR spectrum (500 MHz) of **38** in  $\text{MeOH-d}_4$



A5-Trp\_HMBC

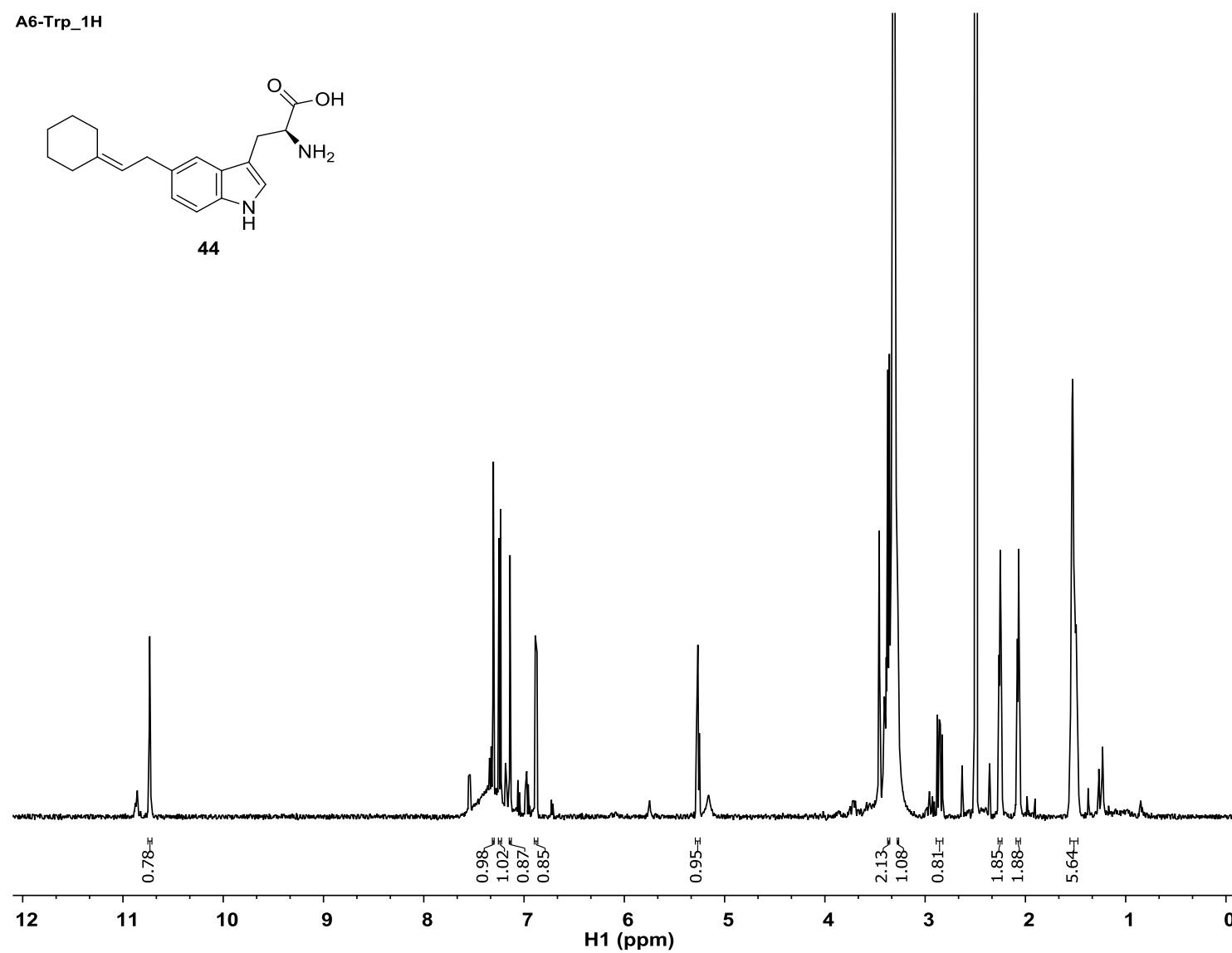


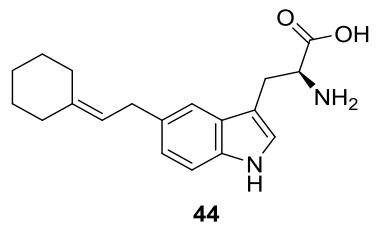
2D  $^1\text{H}$ - $^{13}\text{C}$  HMBC NMR spectrum (500 MHz) of **38** in  $\text{MeOH-d}_4$



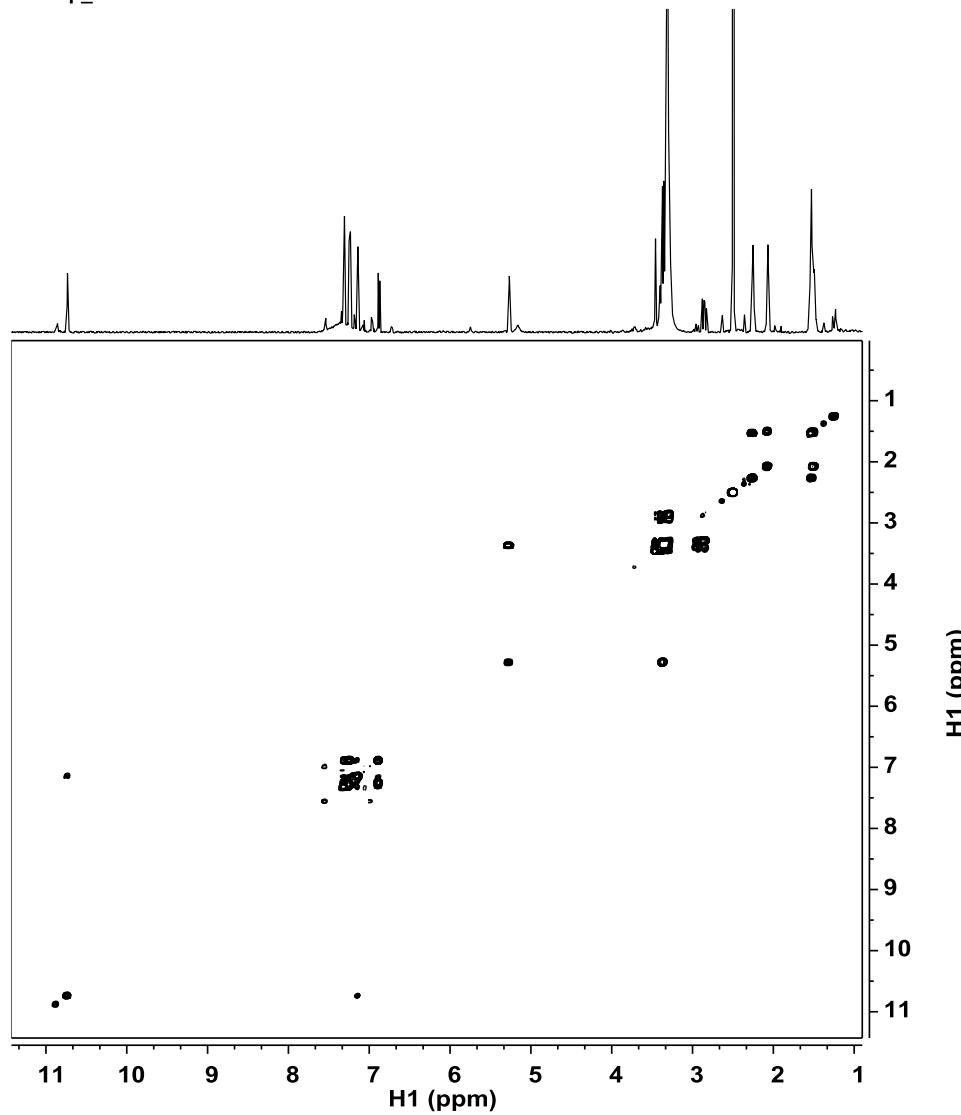
HRMS spectrum of **38**

A6-Trp\_1H

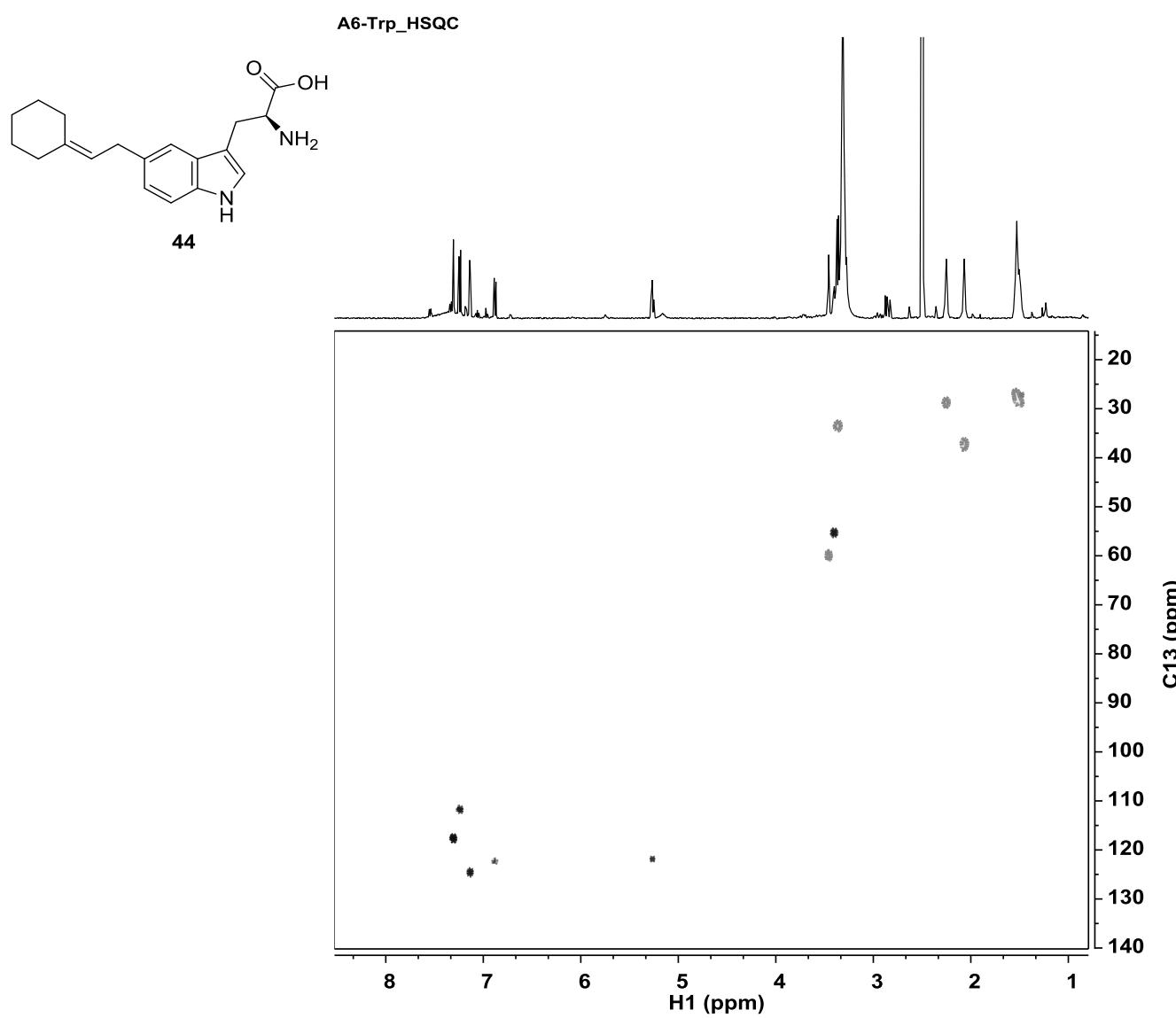




A6-Trp\_COSY

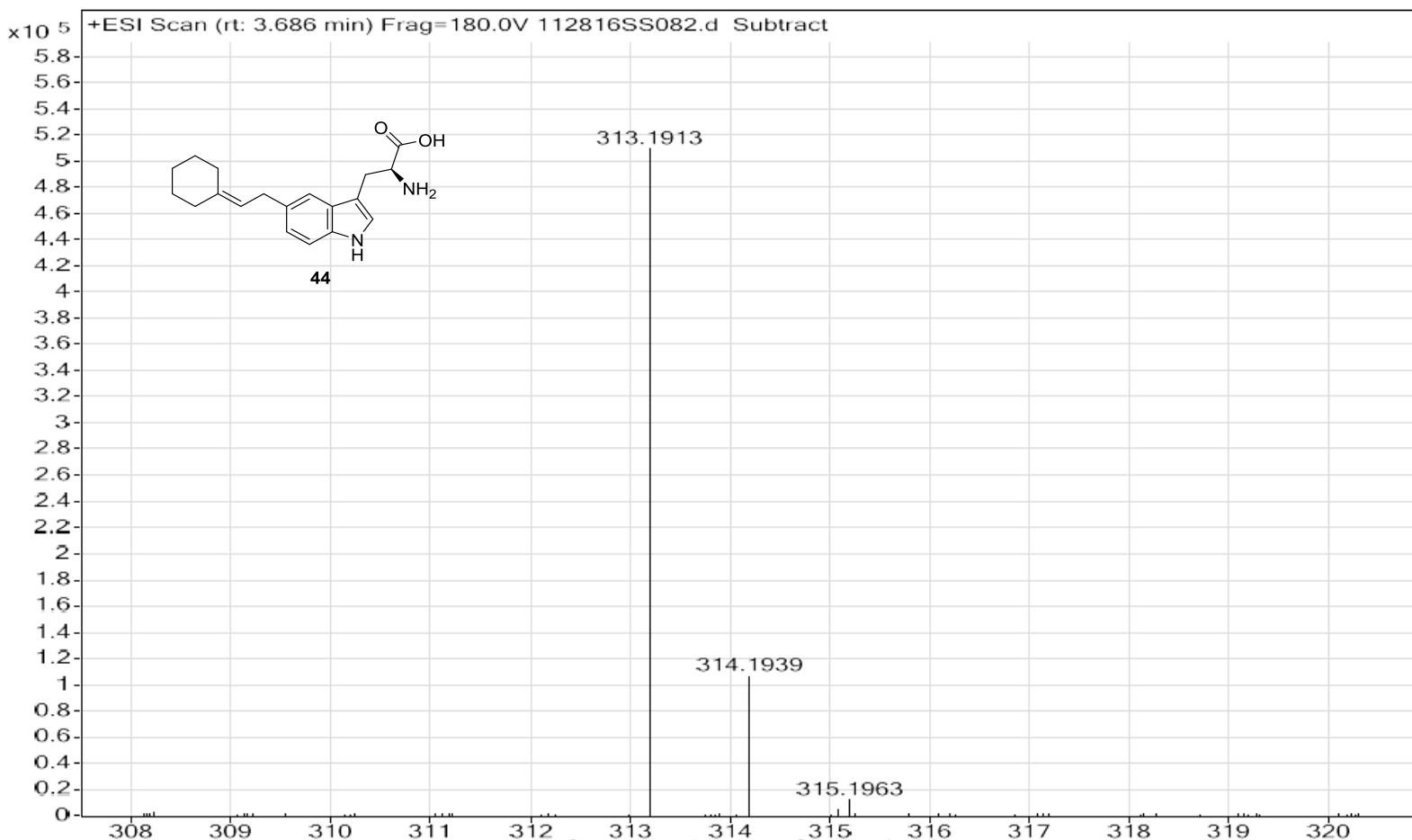


2D  $^1\text{H}$ - $^1\text{H}$  COSY NMR spectrum (500 MHz) of **44** in DMSO-d<sub>6</sub>



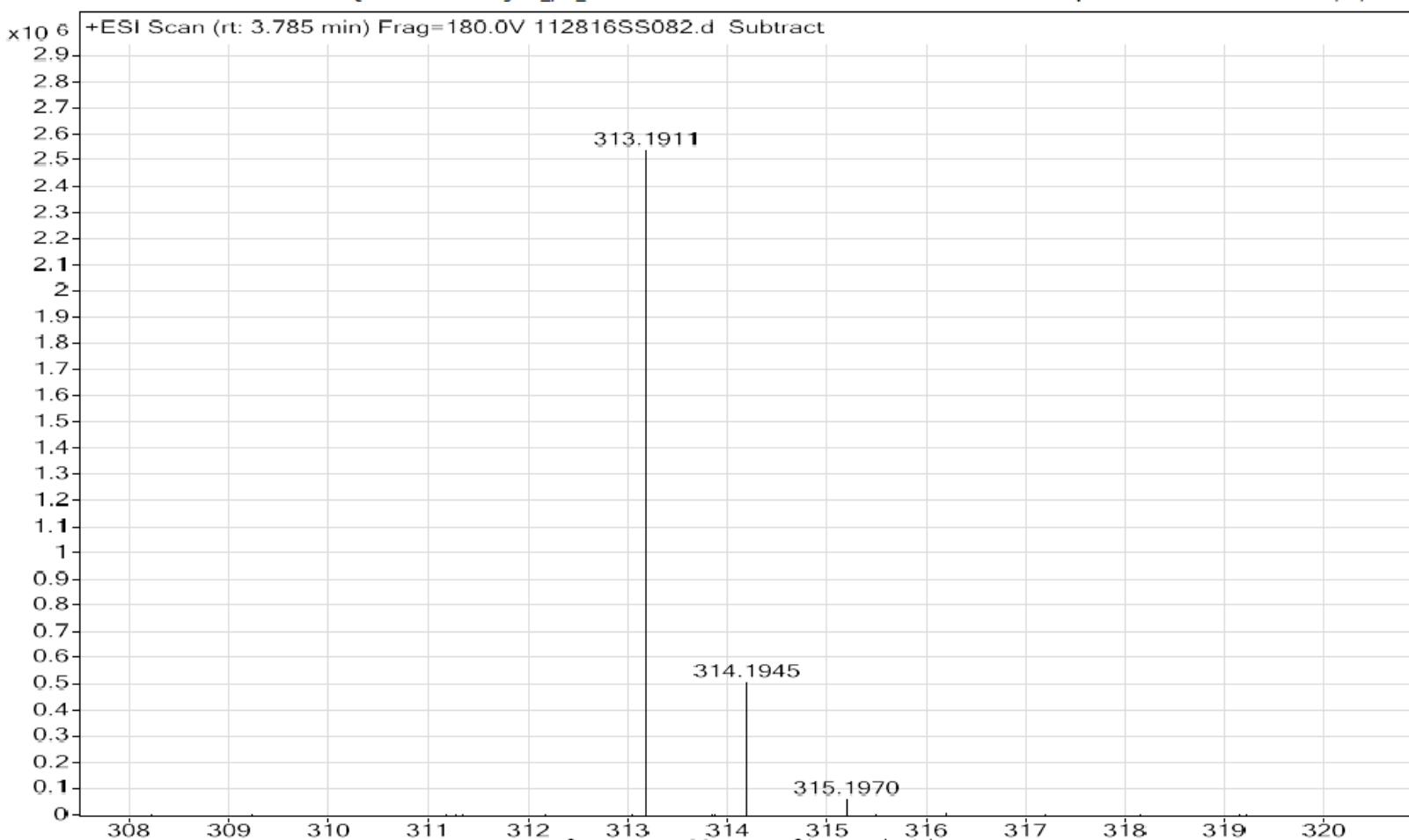
2D <sup>1</sup>H-<sup>13</sup>C HSQC NMR spectrum (500 MHz) of **44** in DMSO-d<sub>6</sub>

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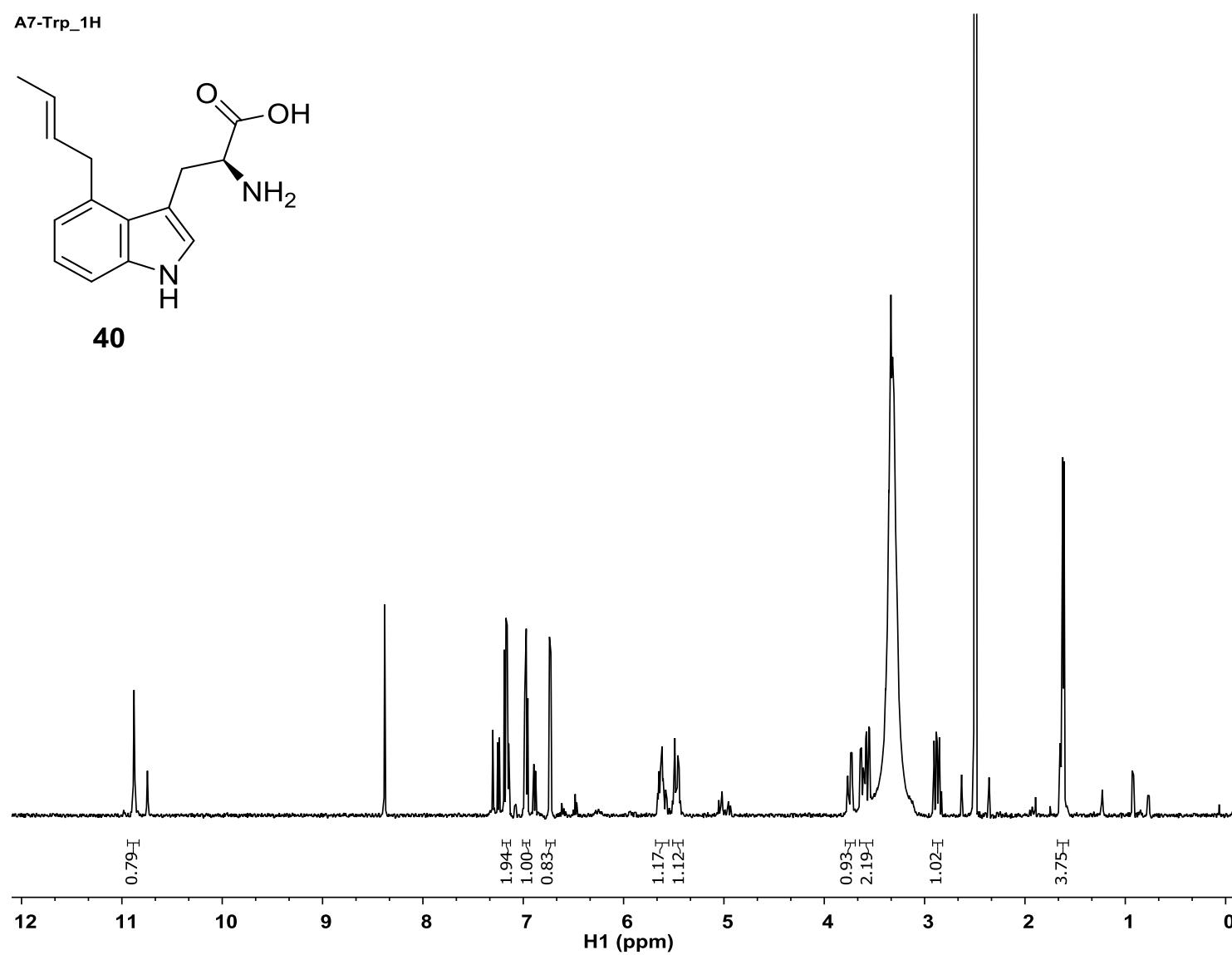
HRMS spectrum of 44

Sample Name	TRP-26	Position	P1-C8	Instrument Name	Instrument 1	User Name	
Inj Vol	5	InjPosition		SampleType	Sample	IRM Calibration Status	Success
Data Filename	112816SS082.d	ACQ Method	drugC18_pos_ms.m	Comment		Acquired Time	11/29/2016 3:20:56 AM

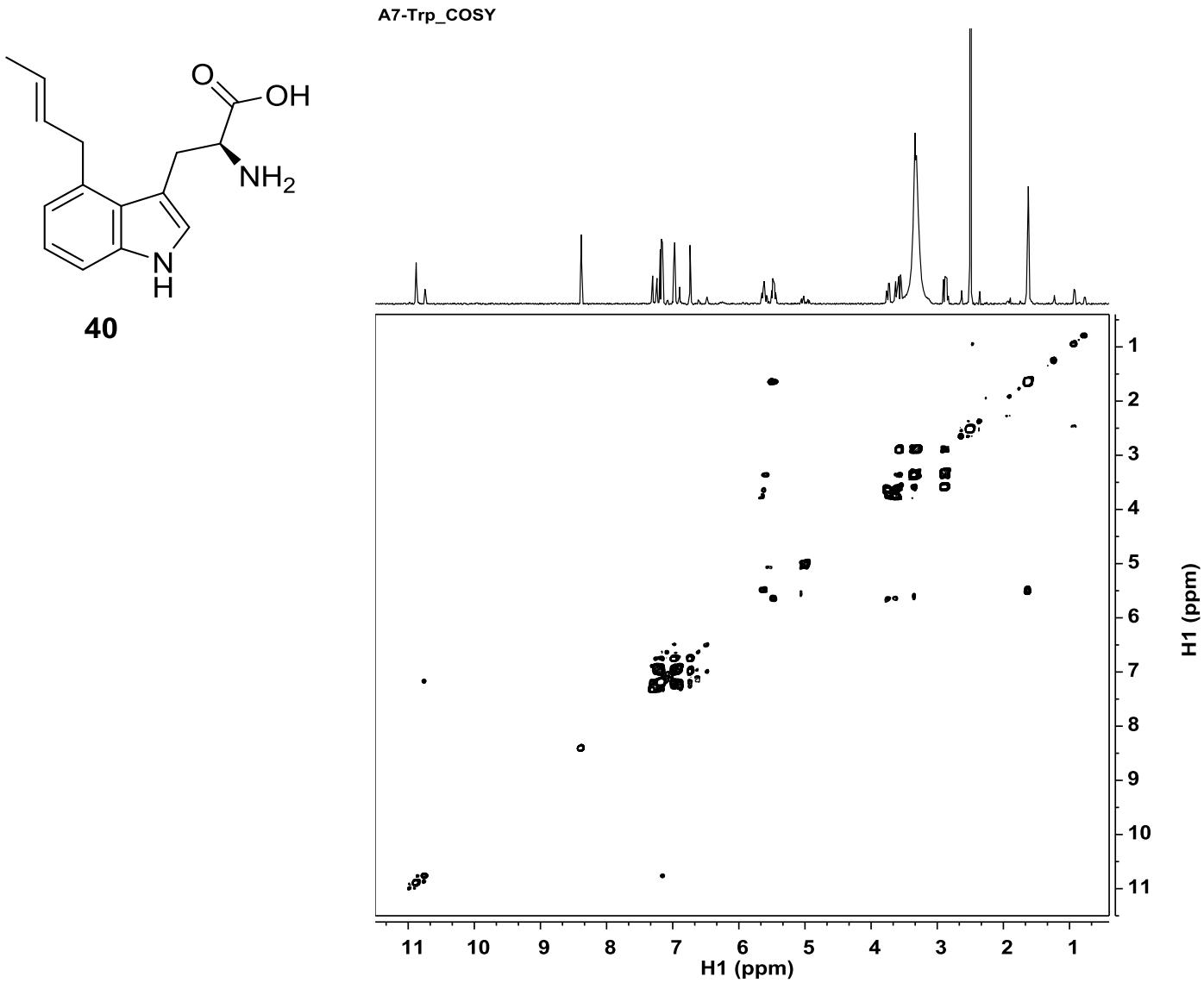


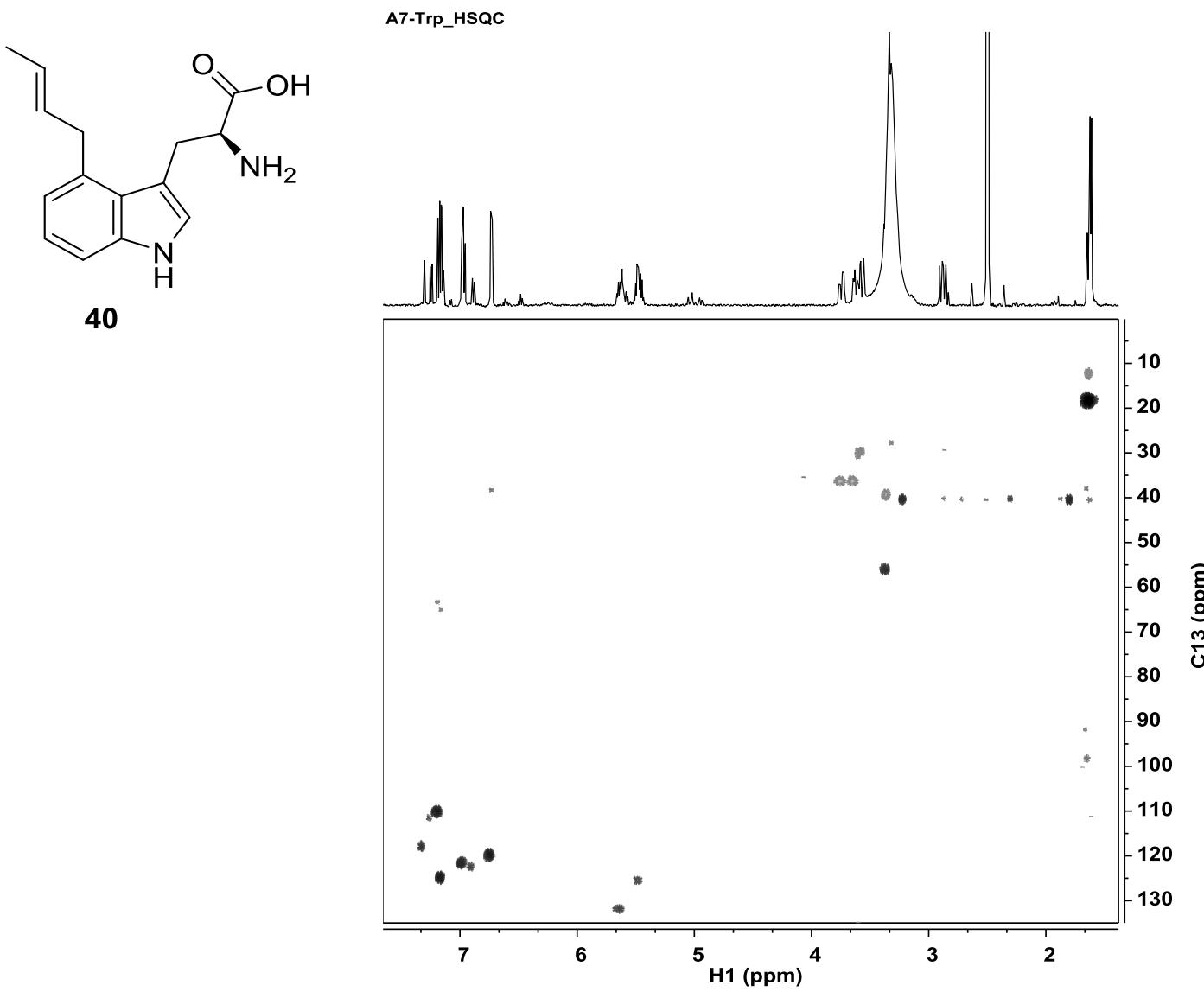
### HRMS of 6-Trp<sub>2</sub>

A7-Trp\_1H



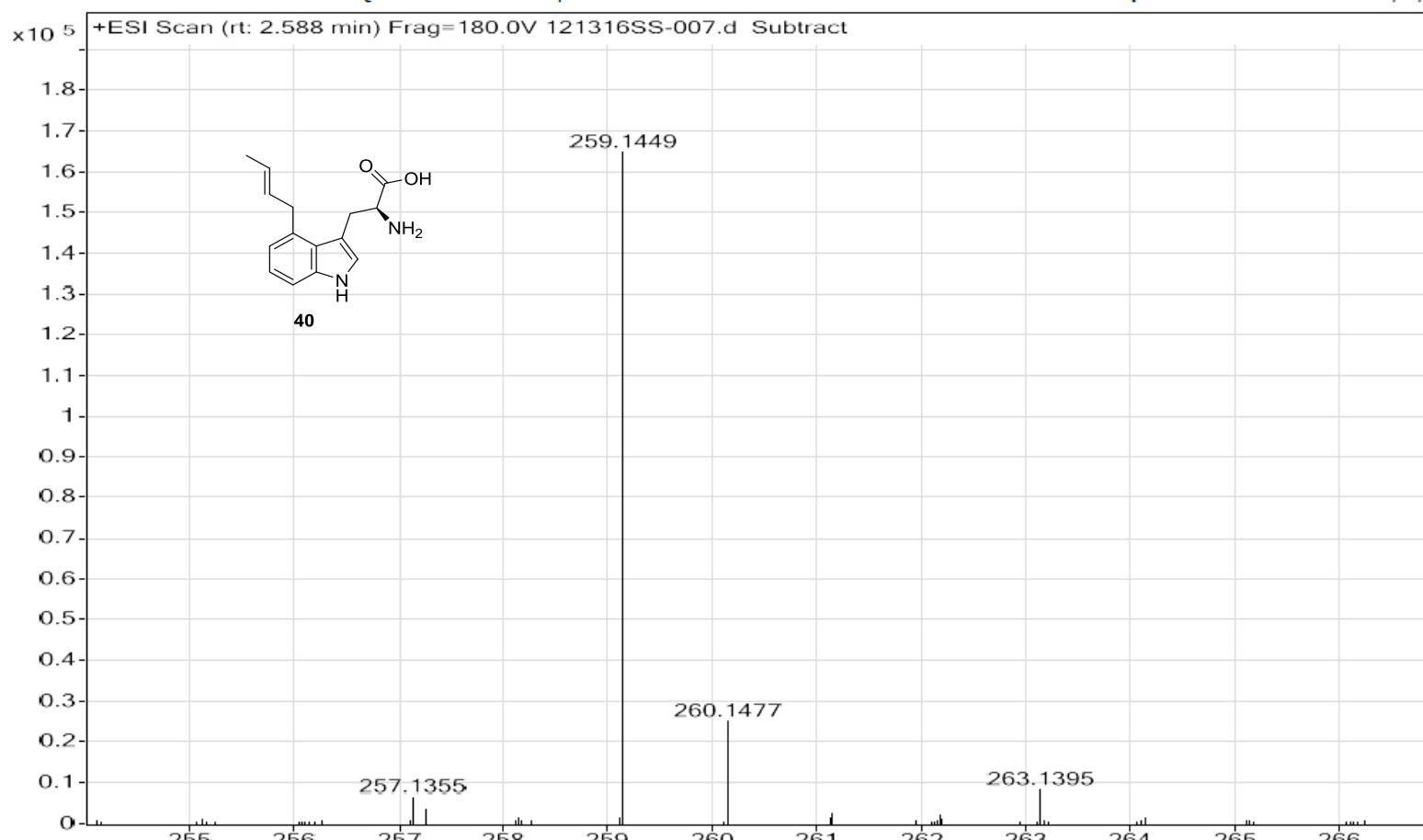
<sup>1</sup>H NMR Spectrum (500 MHz) of compound **40** in DMSO-d<sub>6</sub> ( $C4^N$  isomer + 30%  $C5^N$  isomer)





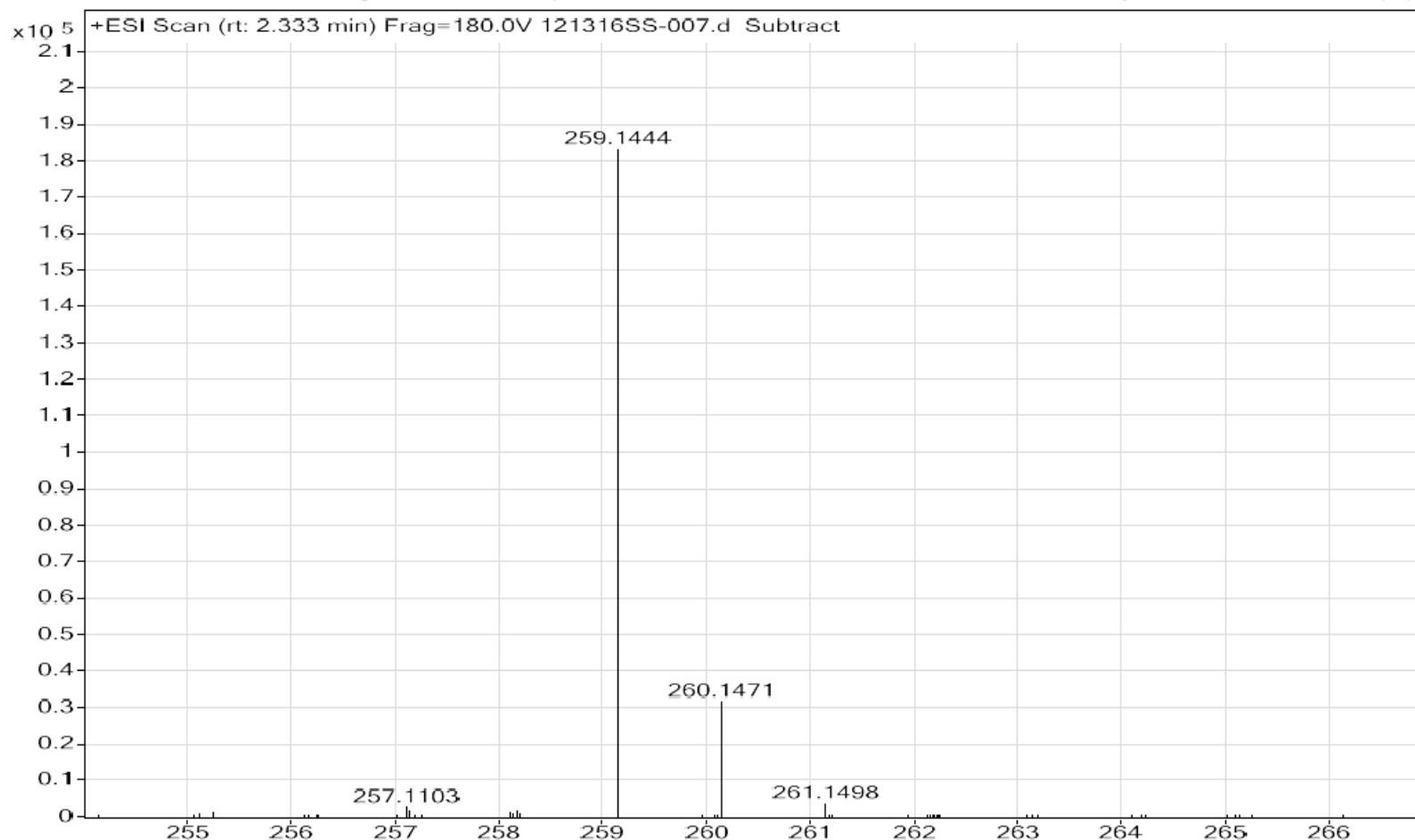
2D  $^1\text{H}$ - $^{13}\text{C}$  HSQC NMR spectrum (500 MHz) of **40** in DMSO-d6 ( $\text{C}4^N$  isomer + 30%  $\text{C}5^N$  isomer)

Sample Name	Vial-34	Position	P1-A2	Instrument Name	Instrument 1	User Name	
Inj Vol	5	InjPosition		SampleType	Sample	IRM Calibration Status	Success
Data Filename	121316SS-007.d	ACQ Method	LCMSUV-pos-short-120	Comment		Acquired Time	12/13/2016 6:02:52 PM



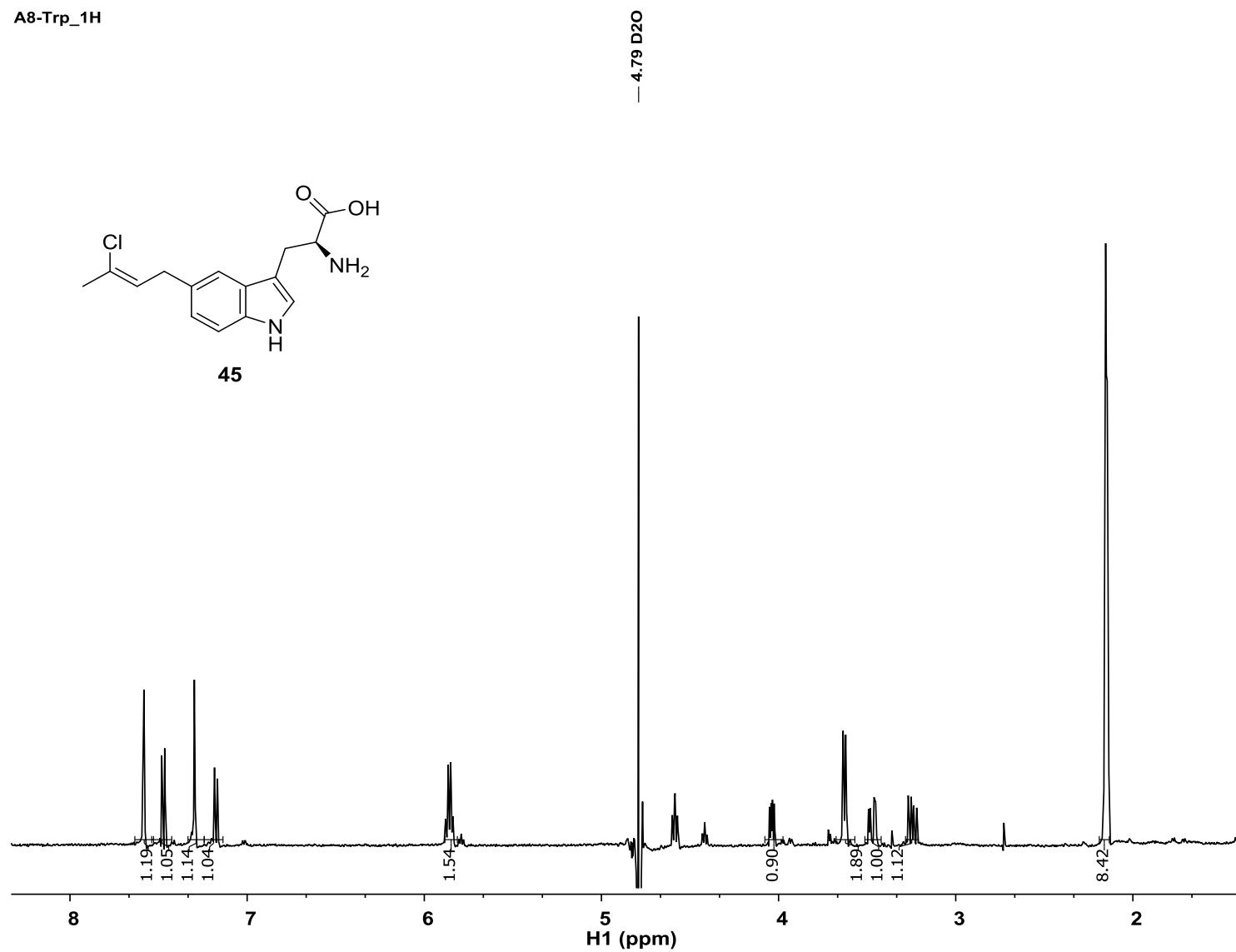
HRMS spectrum of **40**

Sample Name	Vial-34	Position	P1-A2	Instrument Name	Instrument 1	User Name	
Inj Vol	5	InjPosition		SampleType	Sample	IRM Calibration Status	Success
Data Filename	121316SS-007.d	ACQ Method	LCMSUV-pos-short-120	Comment		Acquired Time	12/13/2016 6:02:52 PM



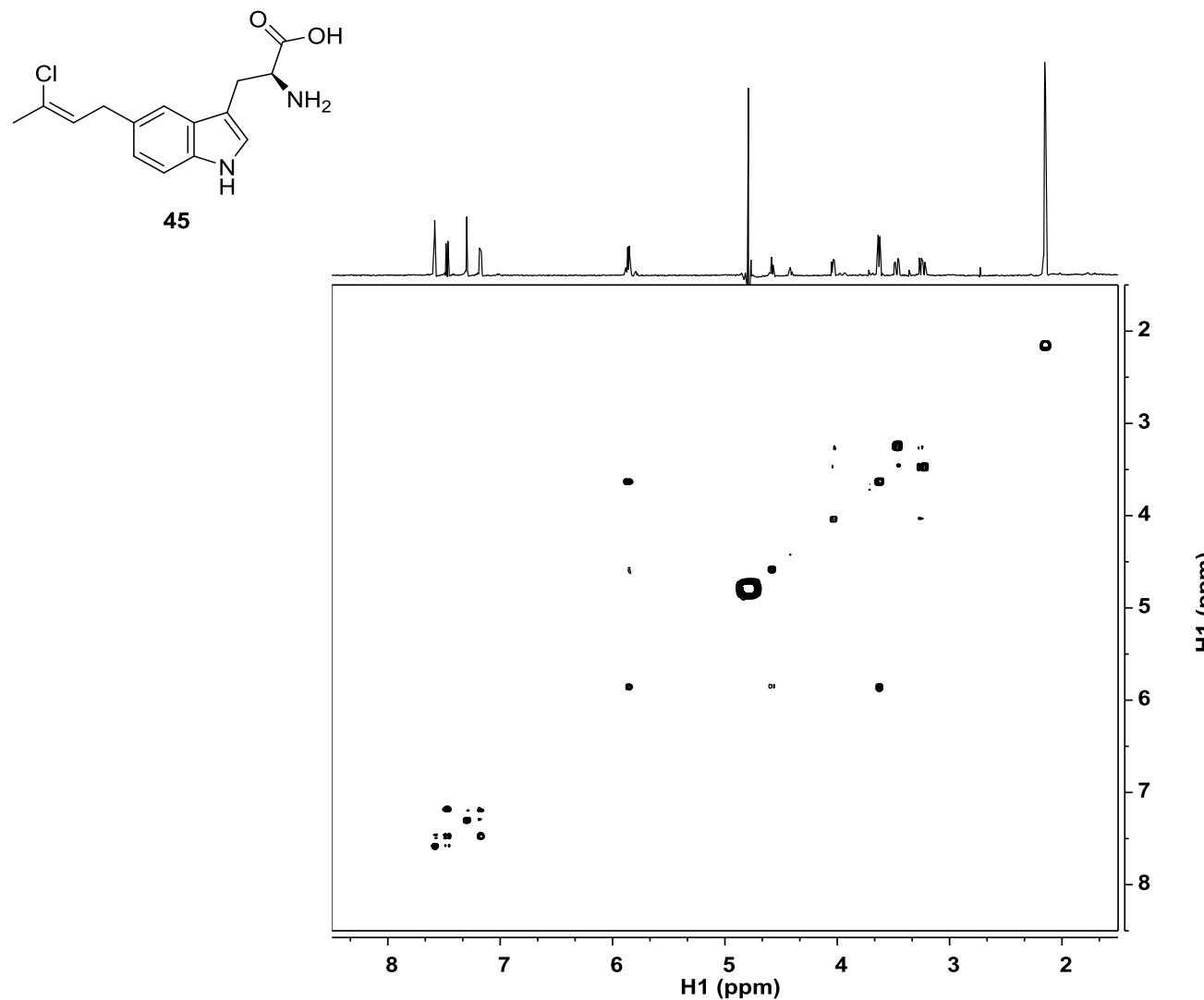
HRMS of 41

A8-Trp\_1H



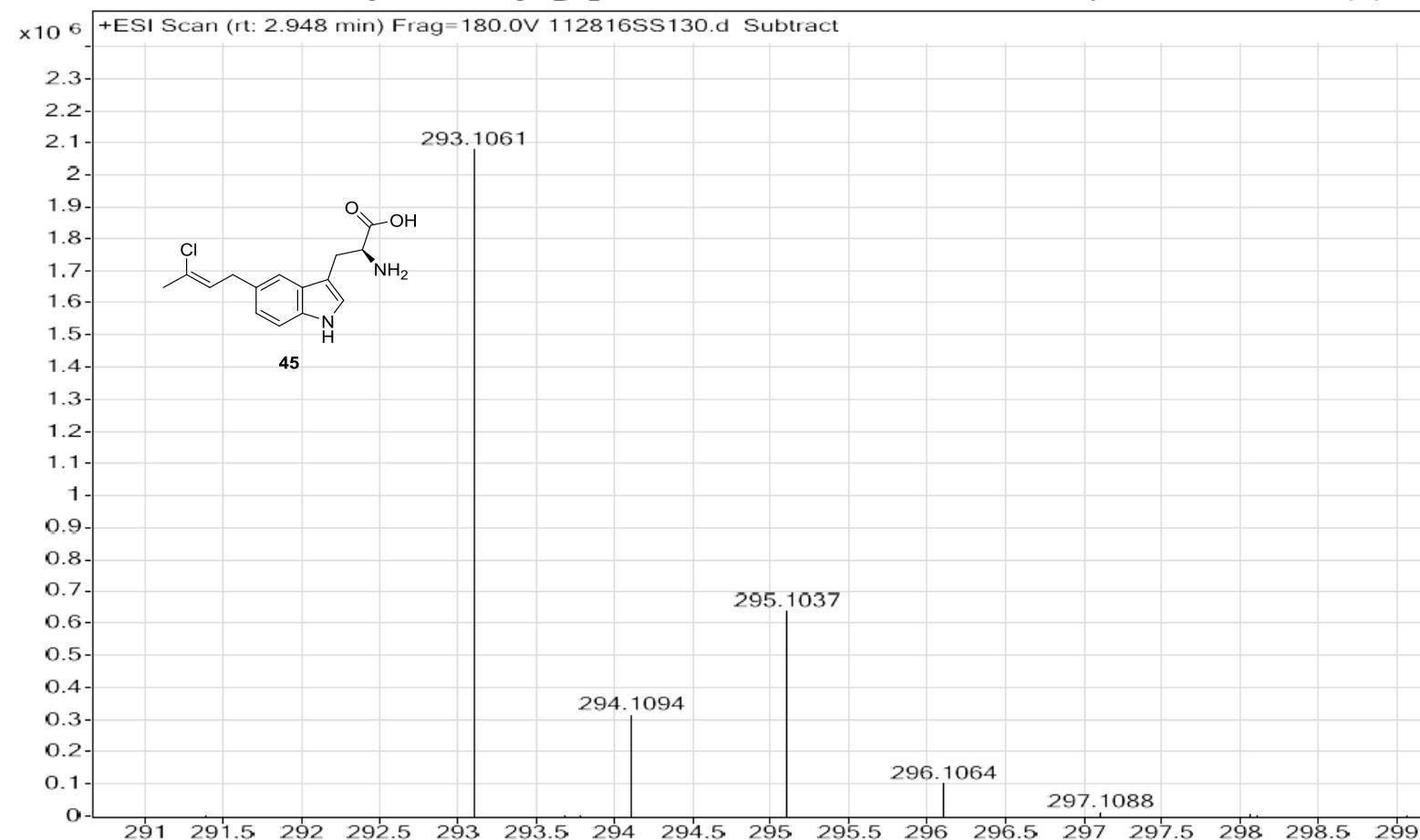
<sup>1</sup>H NMR Spectrum (500 MHz) of compound **45** in D<sub>2</sub>O

A8-Trp\_COSY



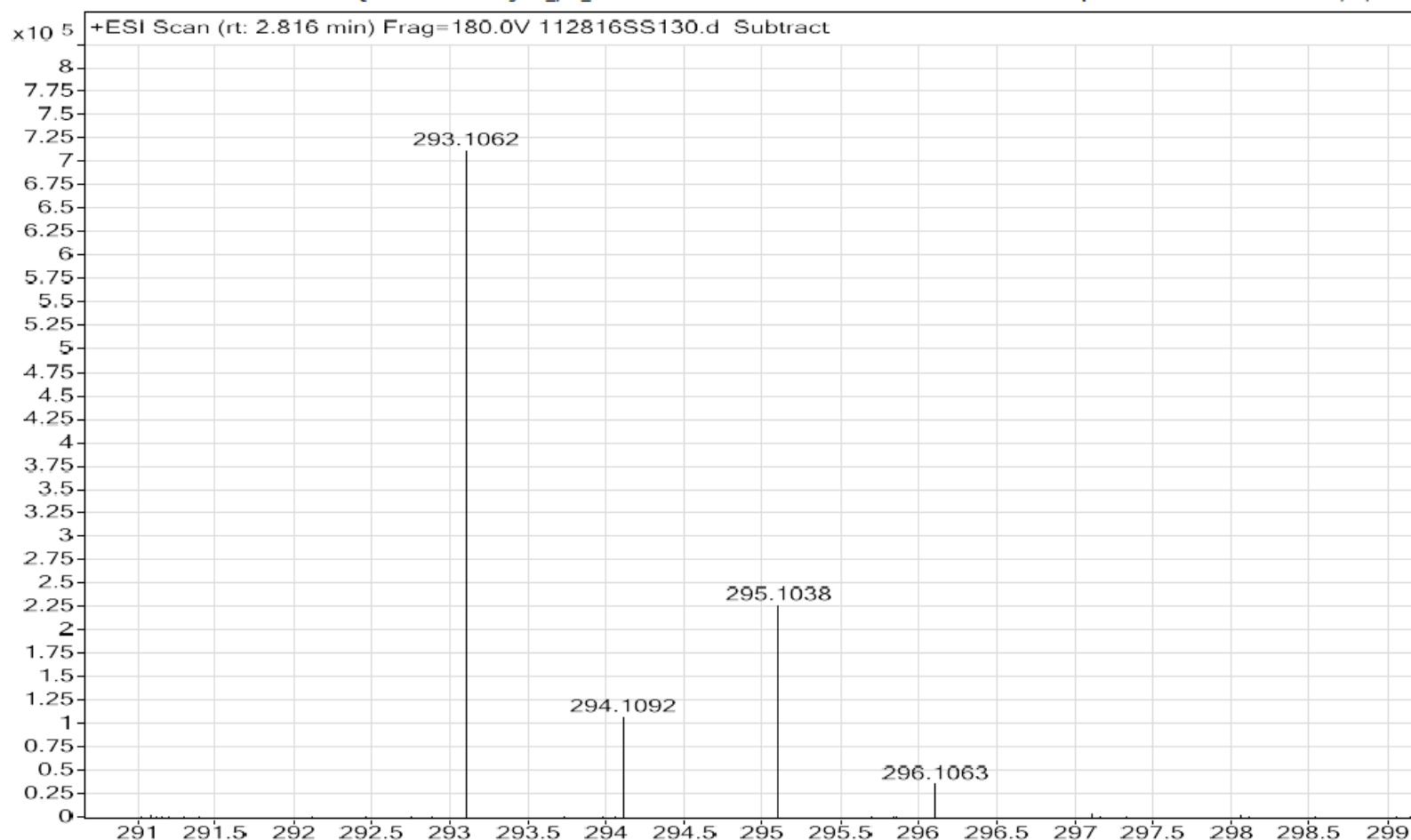
2D  $^1\text{H}$ - $^1\text{H}$  COSY NMR spectrum (500 MHz) of **45** in  $\text{D}_2\text{O}$

Sample Name	TRP-42	Position	P1-E6	Instrument Name	Instrument 1	User Name	
Inj Vol	5	InjPosition		SampleType	Sample	IRM Calibration Status	Success
Data Filename	112816SS130.d	ACQ Method	drugC18_pos_ms.m	Comment		Acquired Time	11/29/2016 12:04:09 PM



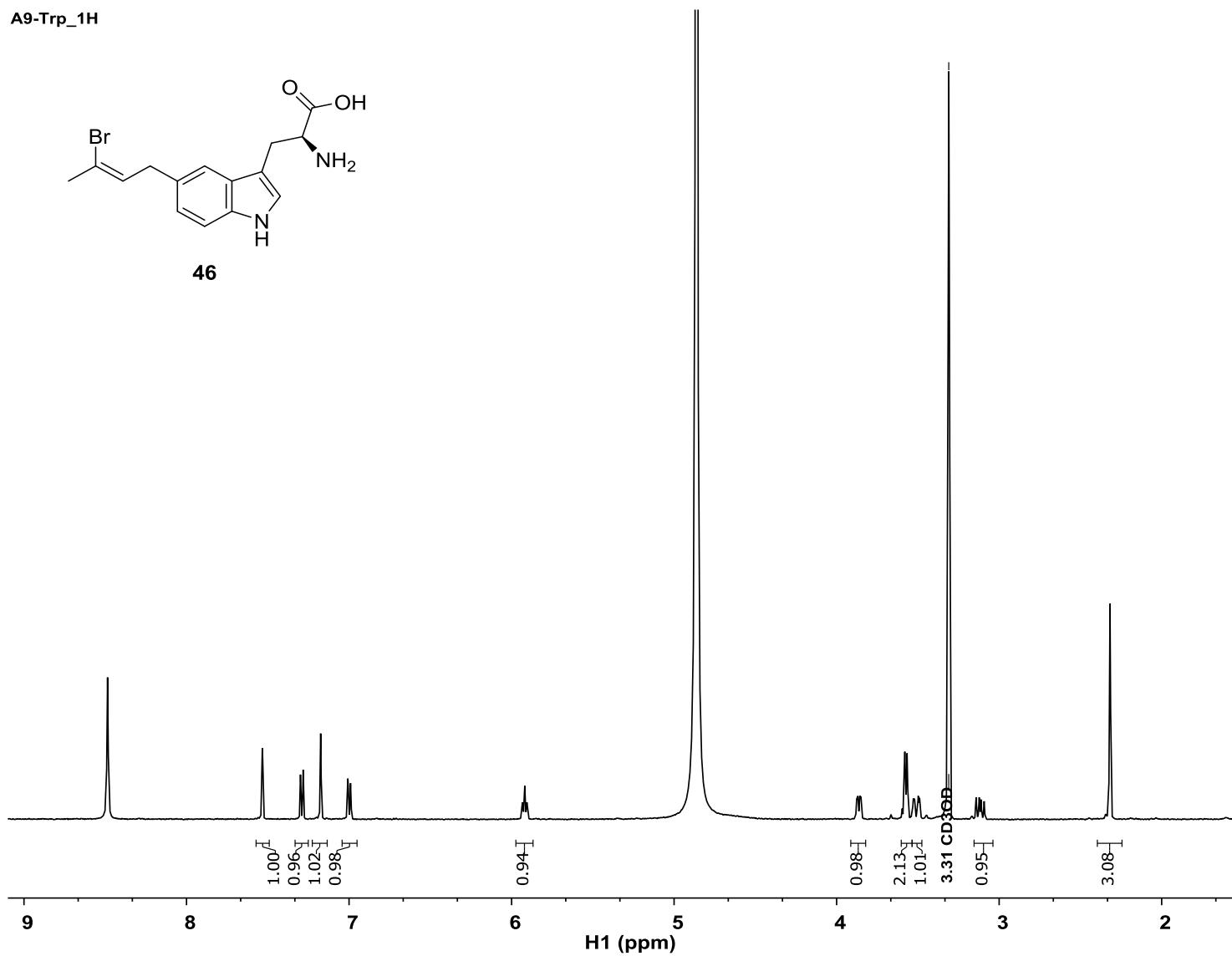
HRMS spectrum of **45**

Sample Name	TRP-42	Position	P1-E6	Instrument Name	Instrument 1	User Name	
Inj Vol	5	InjPosition		SampleType	Sample	IRM Calibration Status	Success
Data Filename	112816SS130.d	ACQ Method	drugC18_pos_ms.m	Comment		Acquired Time	11/29/2016 12:04:09 PM

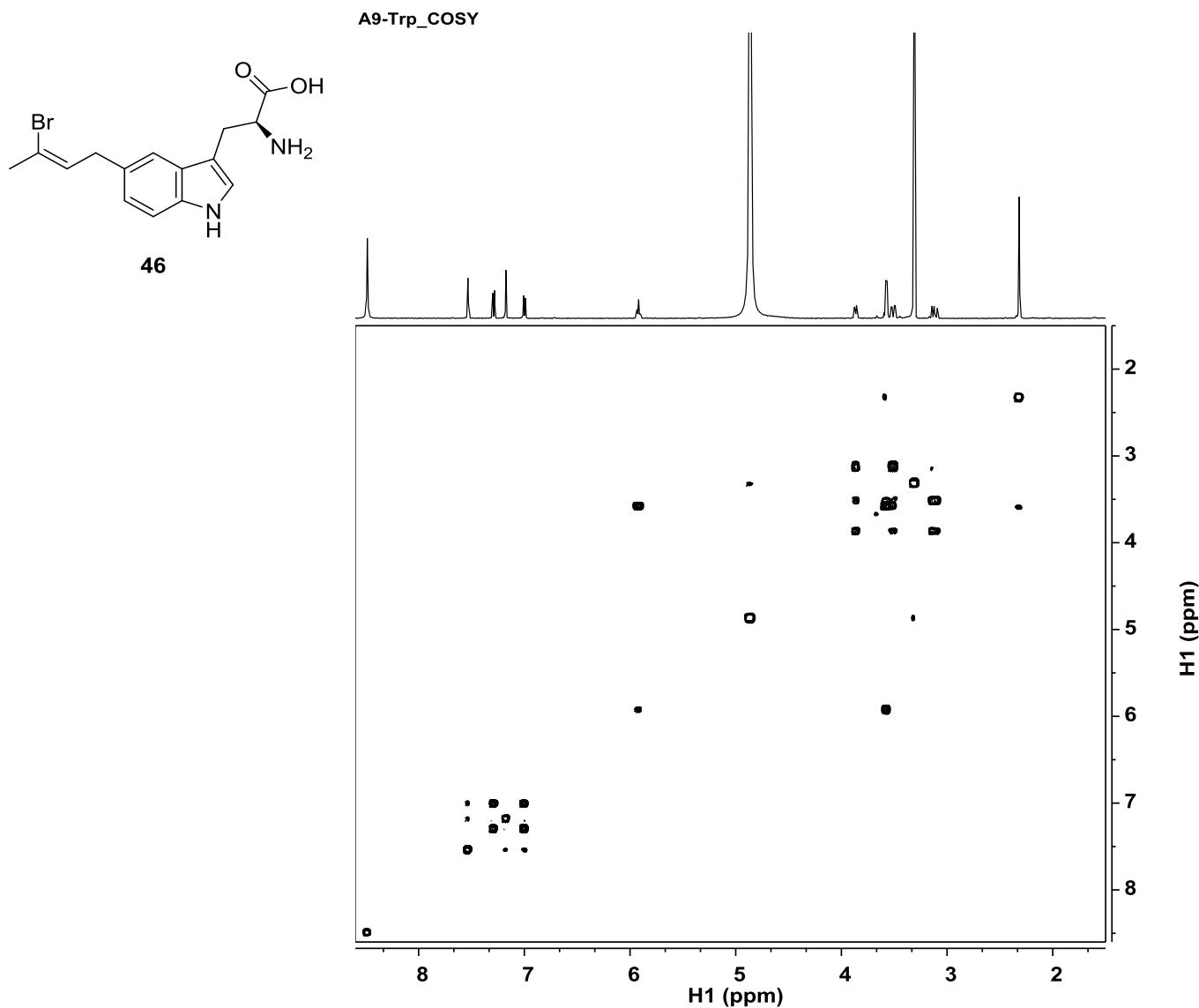


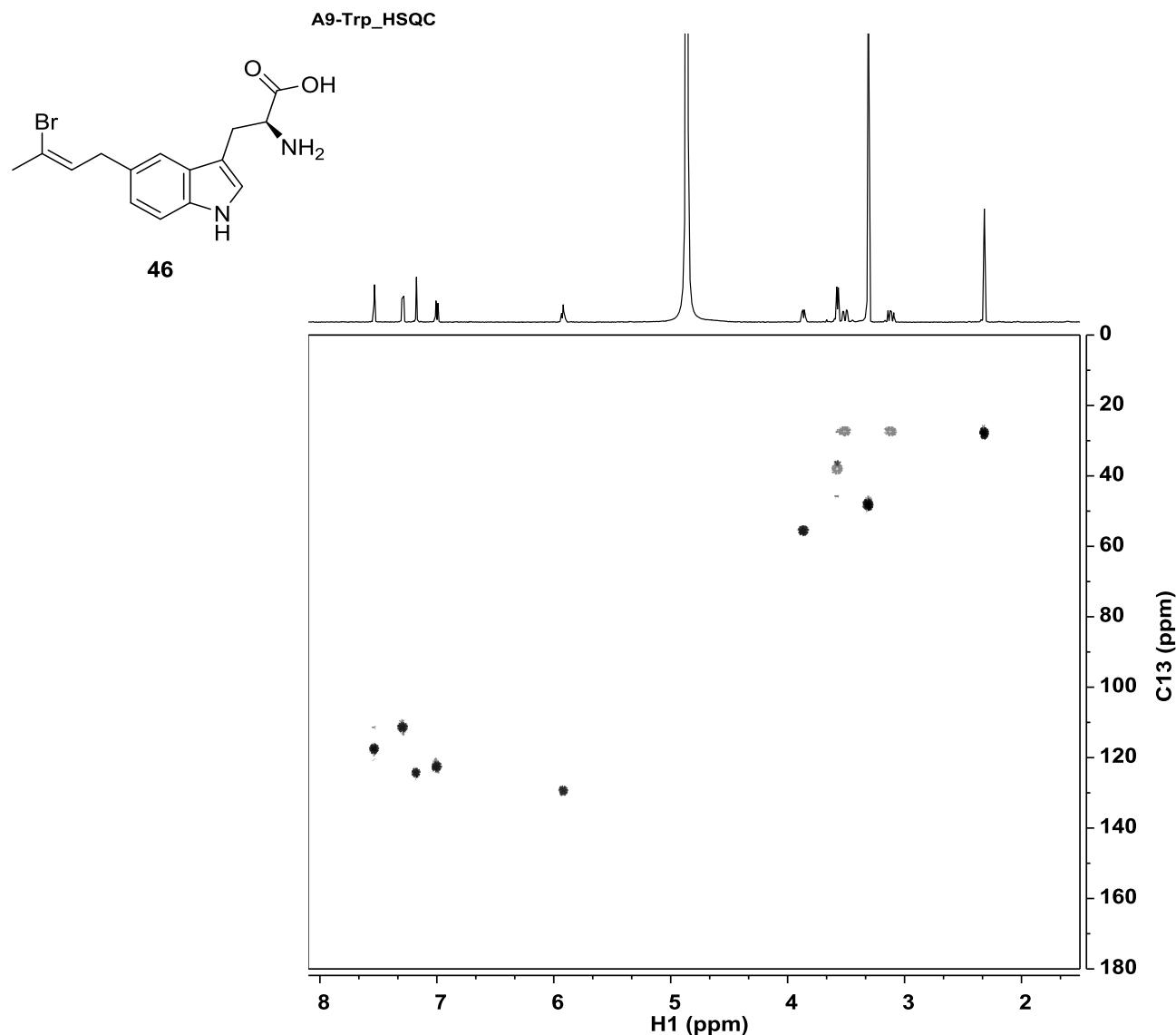
### HRMS of 8-Trp<sub>2</sub>

A9-Trp\_1H

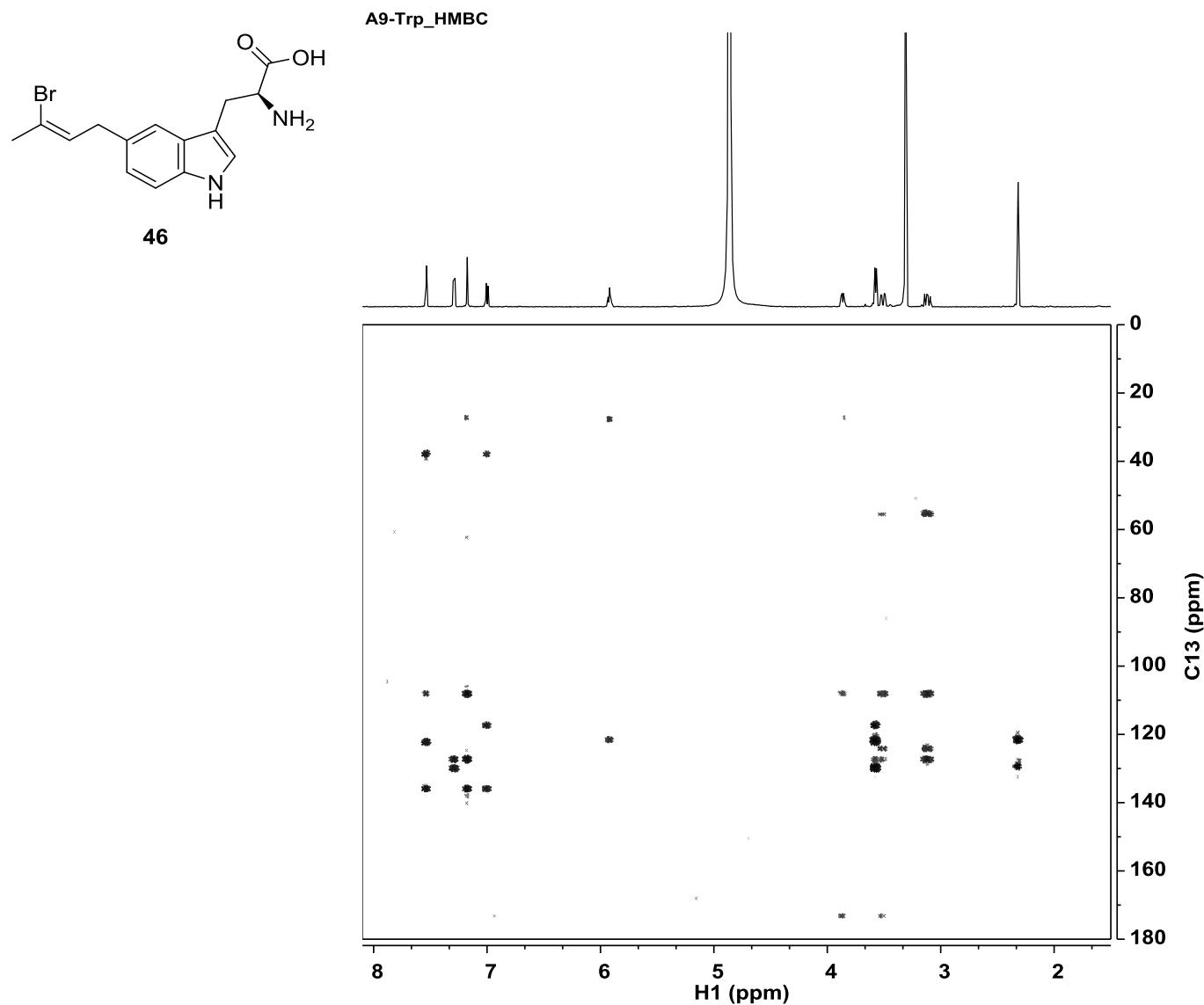


<sup>1</sup>H NMR Spectrum (500 MHz) of compound **46** in MeOH d4.

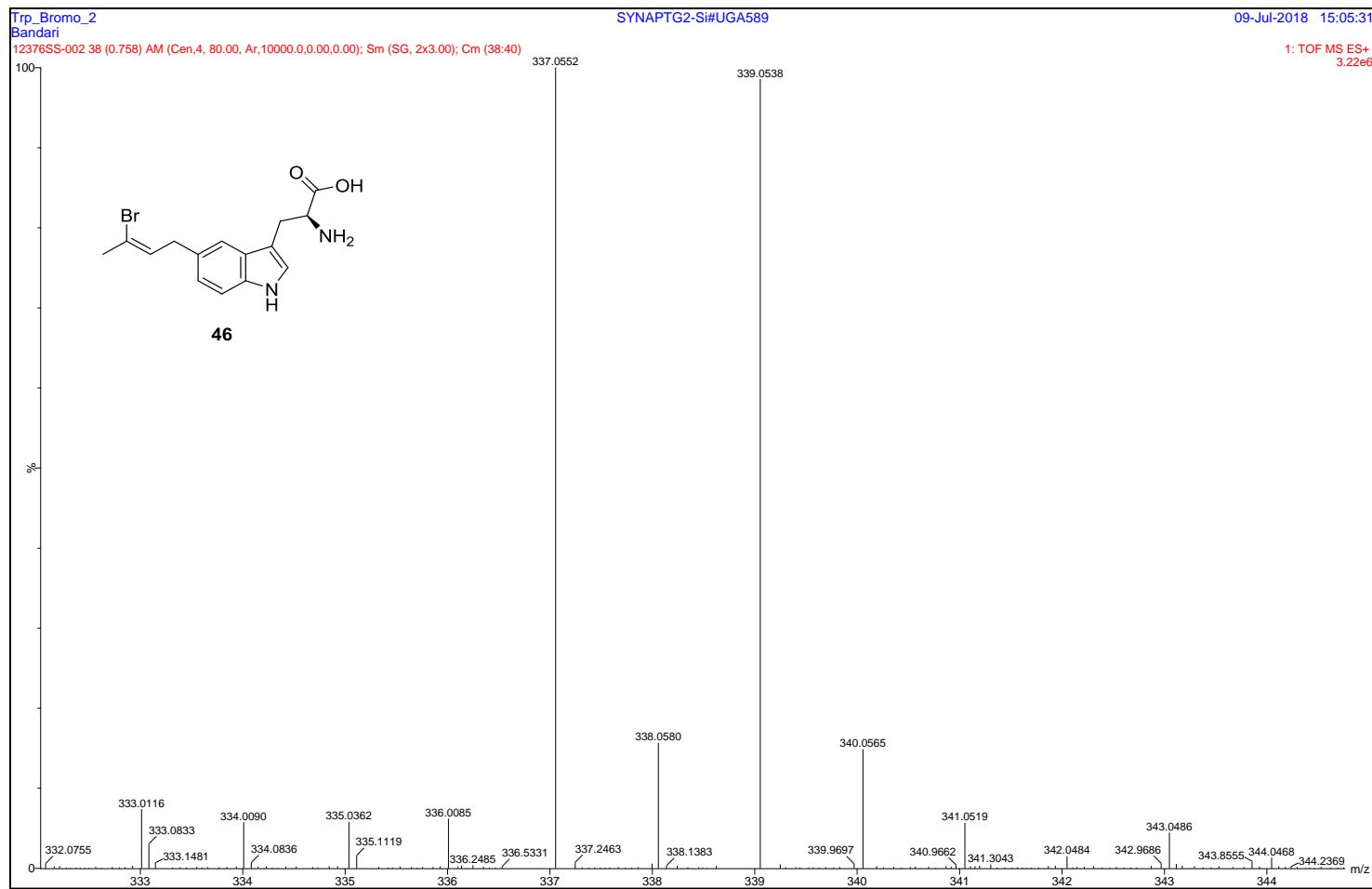




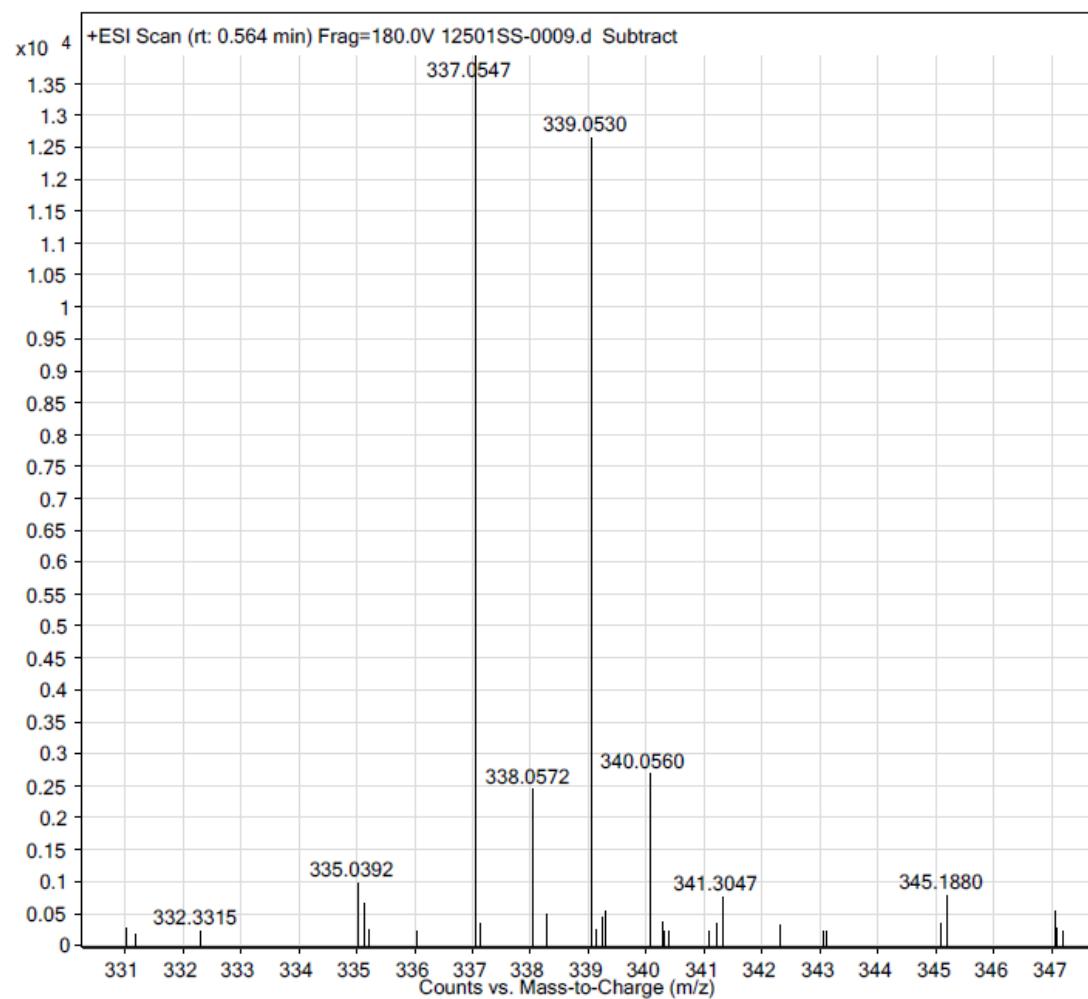
2D  $^1\text{H}$ - $^{13}\text{C}$  HSQC NMR spectrum (500 MHz) of **46** in  $\text{MeOH-d}_4$



2D  $^1\text{H}$ - $^{13}\text{C}$  HSQC NMR spectrum (500 MHz) of **46**- in  $\text{MeOH-d}_4$

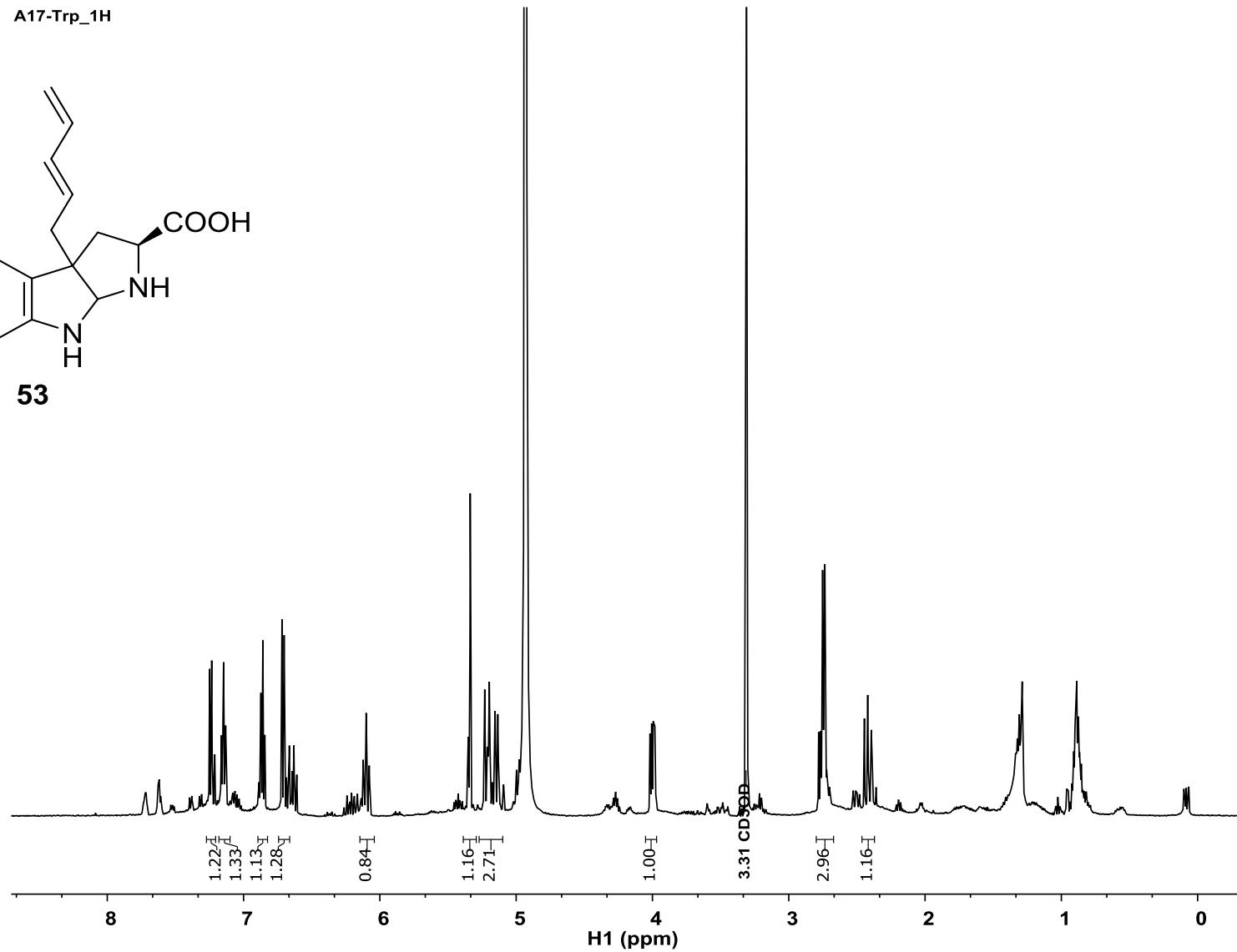
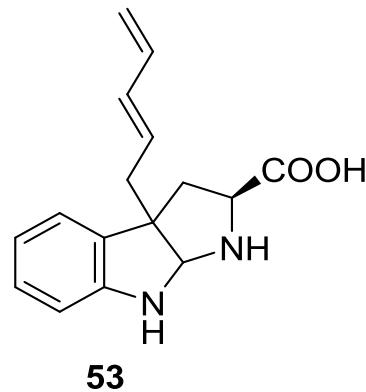


HRMS spectrum of **46**

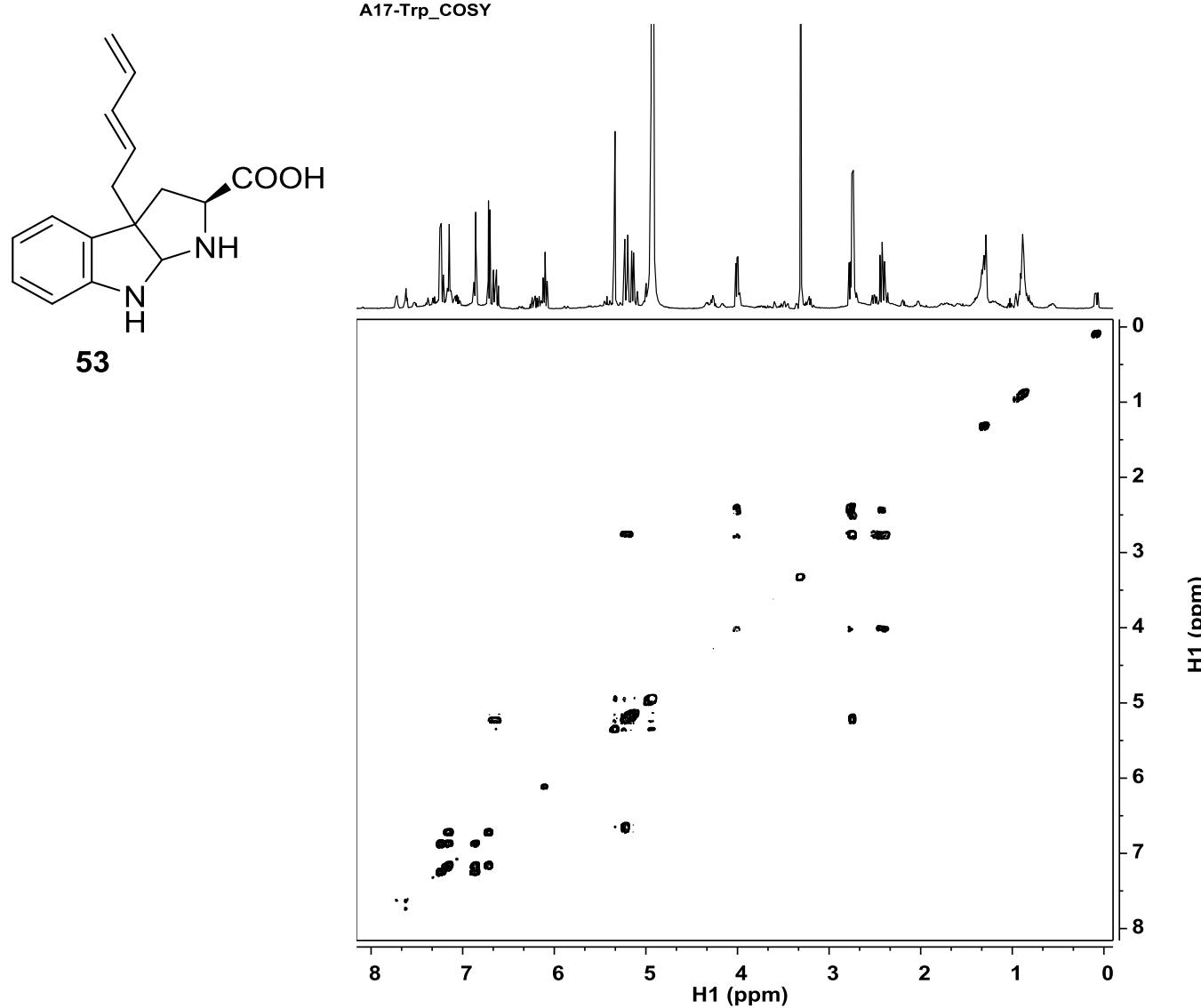


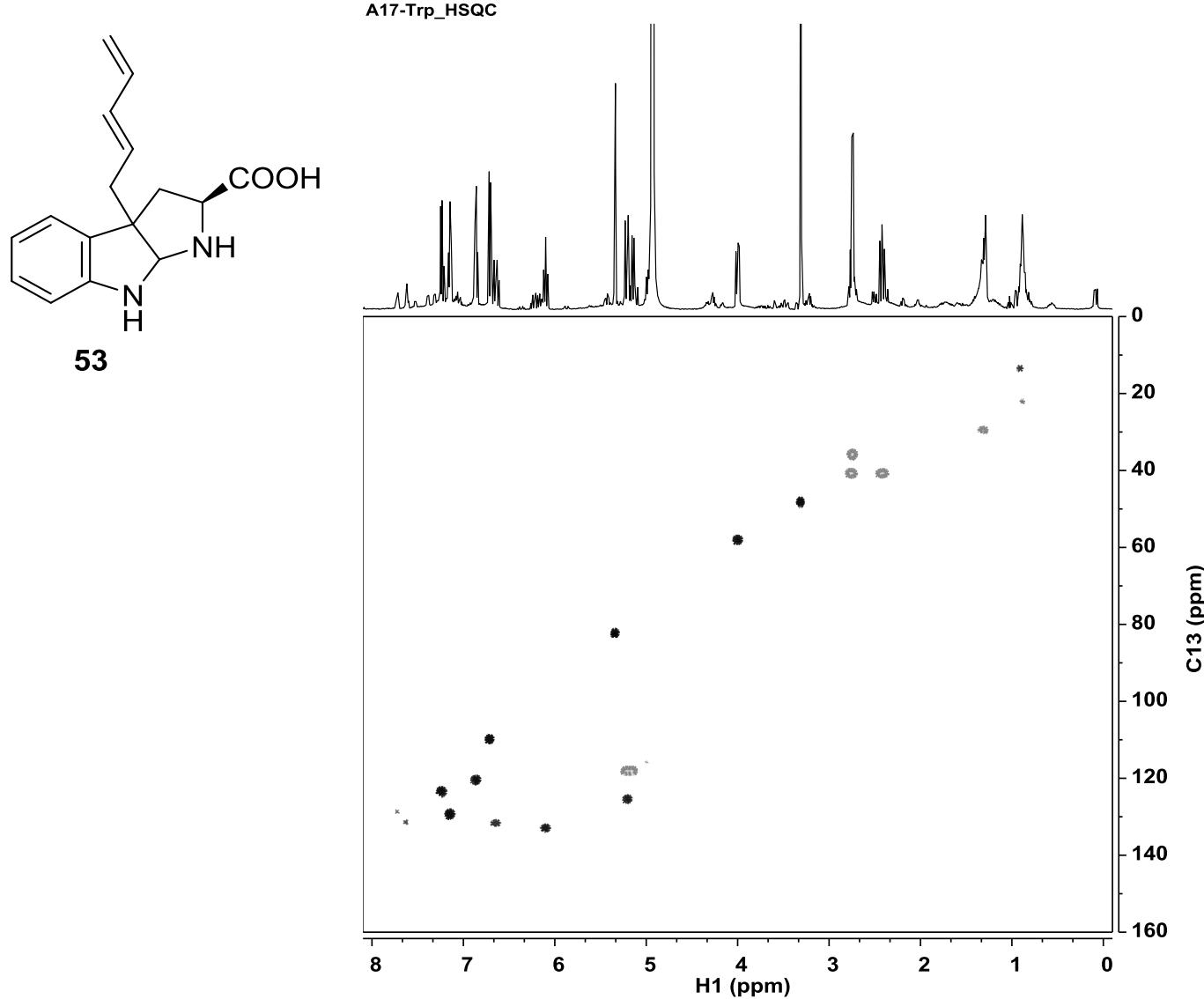
HRMS spectrum of **9-Trp<sub>2</sub>**

A17-Trp\_1H



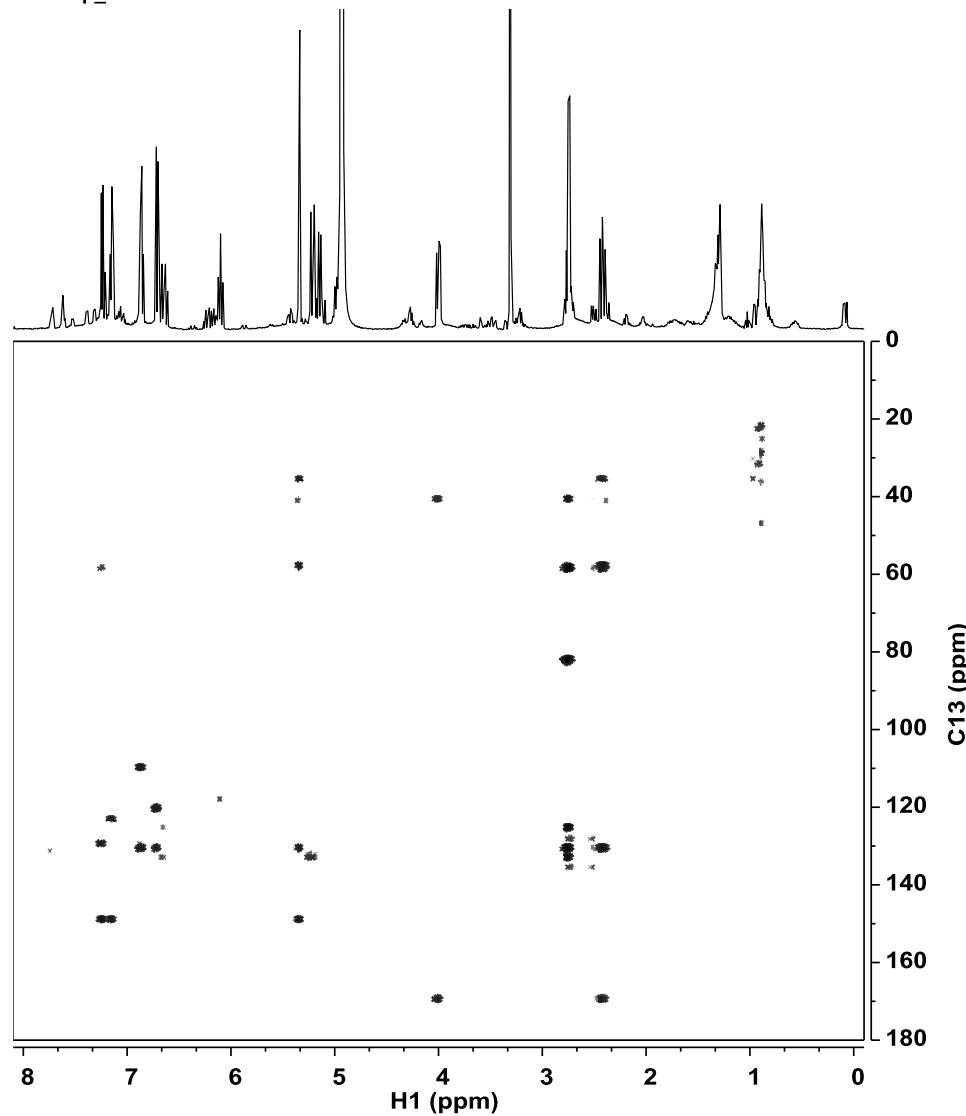
<sup>1</sup>H NMR Spectrum (500 MHz) of compound **53** in MeOH-d<sub>4</sub>





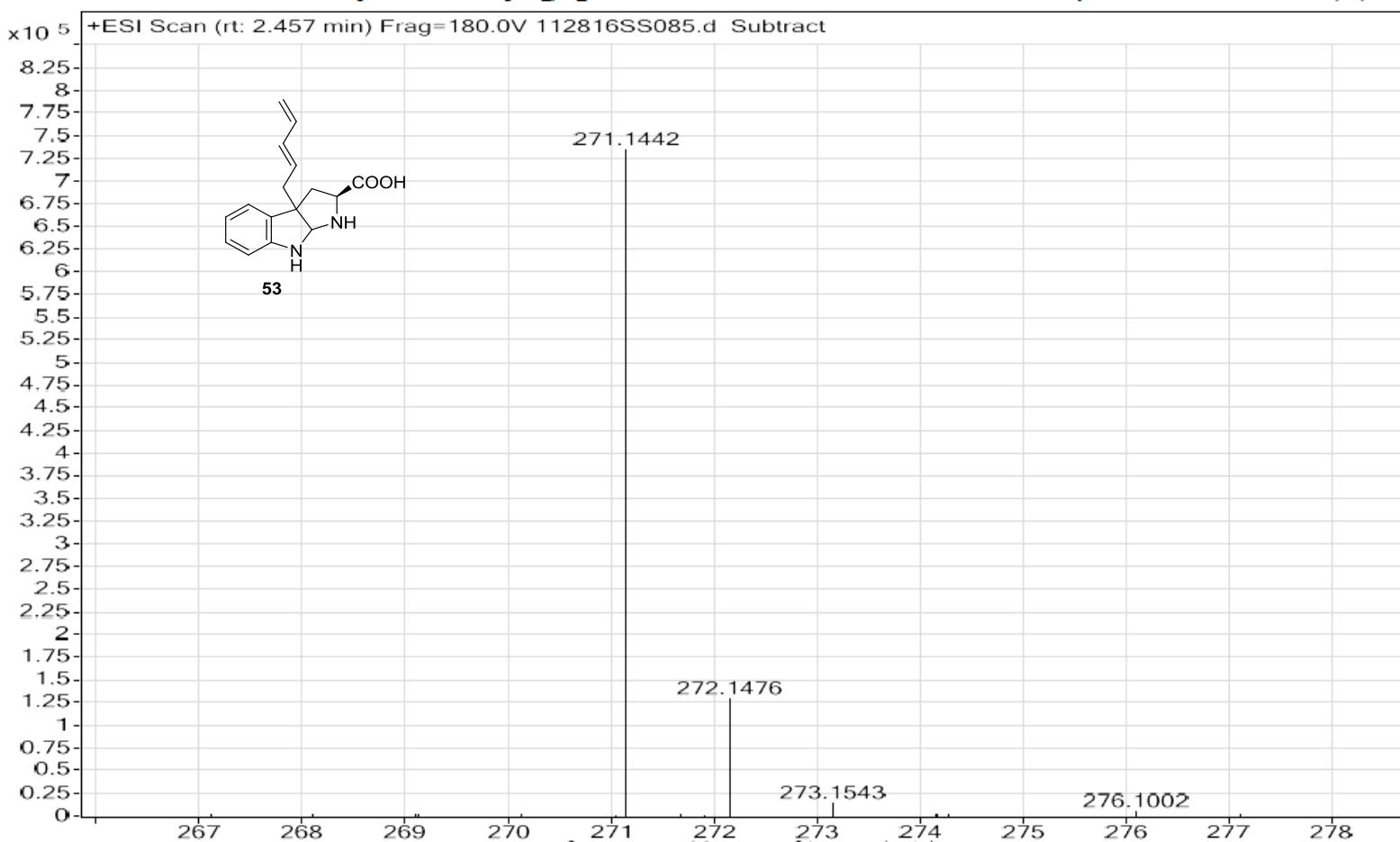
2D  $^1\text{H}$ - $^{13}\text{C}$  HSQC NMR spectrum (500 MHz) of **53** in  $\text{MeOH-d}_4$

A17-Trp\_HMBC



2D  $^1\text{H}$ - $^{13}\text{C}$  HMBC NMR spectrum (500 MHz) of **53** in MeOH-d4

Sample Name	TRP-27	Position	P1-C9	Instrument Name	Instrument 1	User Name	
Inj Vol	5	InjPosition		SampleType	Sample	IRM Calibration Status	Success
Data Filename	112816SS085.d	ACQ Method	drugC18_pos_ms.m	Comment		Acquired Time	11/29/2016 3:53:36 AM

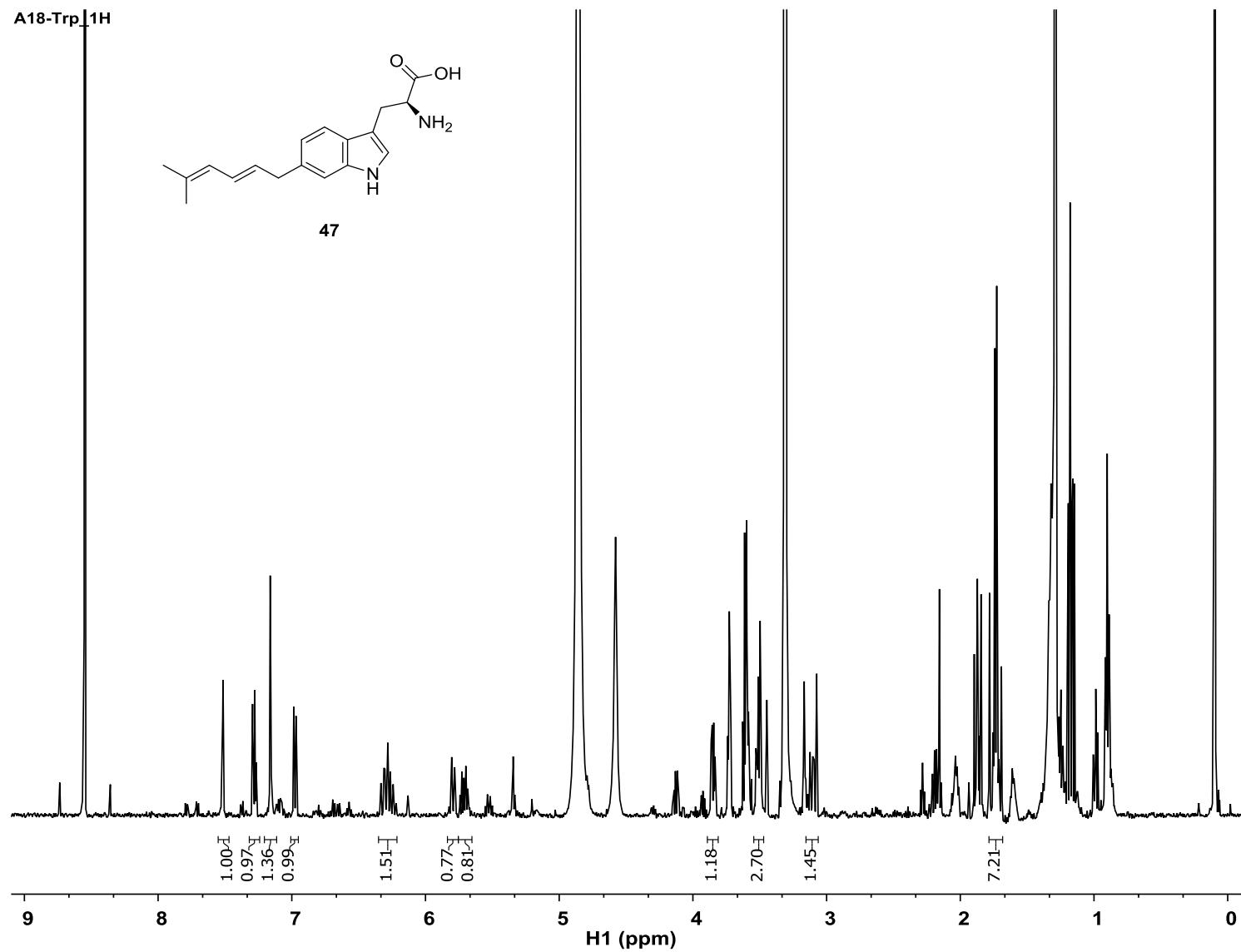


## HRMS spectrum of 53

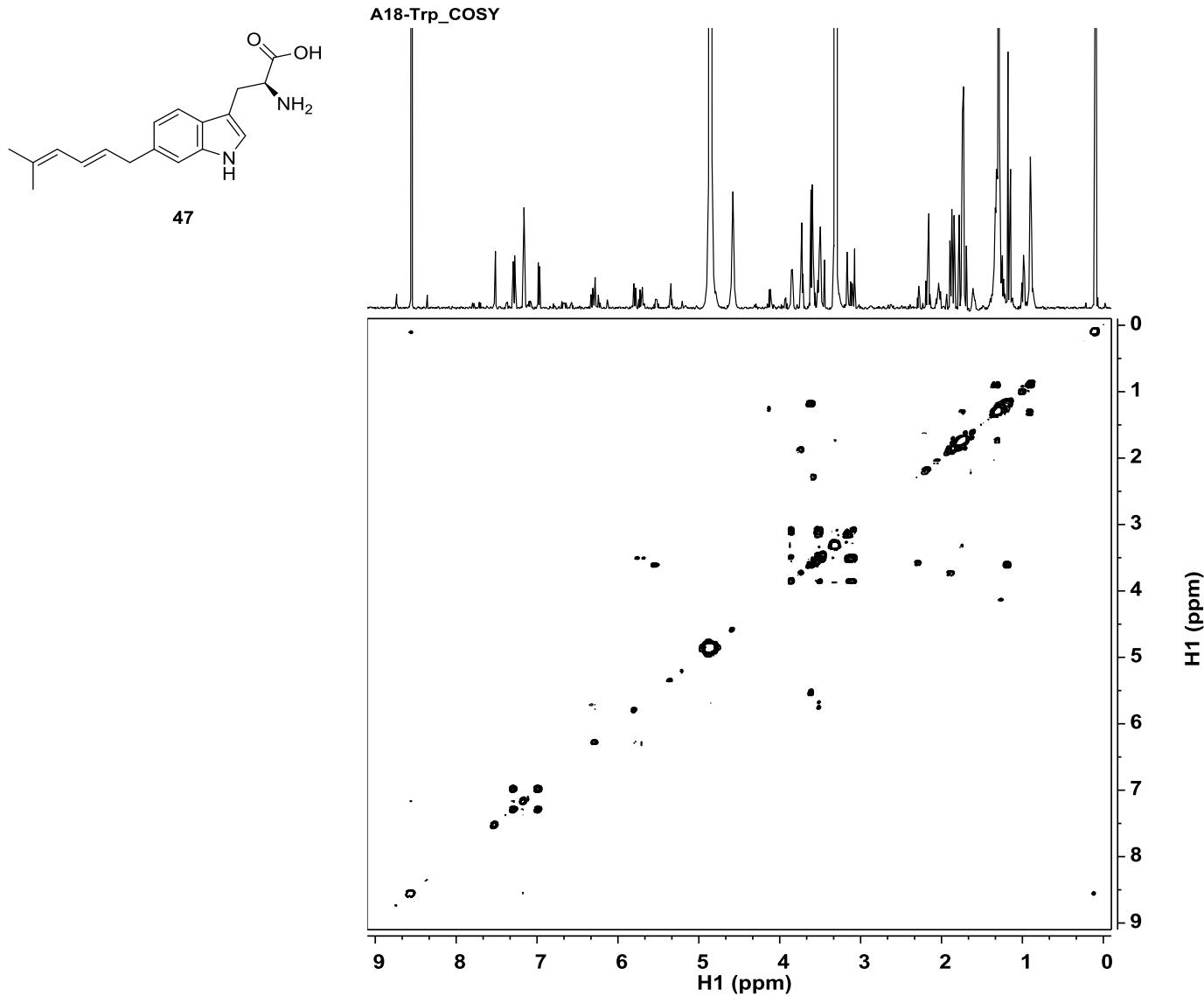
Sample Name	TRP-27	Position	P1-C9	Instrument Name	Instrument 1	User Name	
Inj Vol	5	InjPosition		SampleType	Sample	IRM Calibration Status	Success
Data Filename	112816SS085.d	ACQ Method	drugC18_pos_ms.m	Comment		Acquired Time	11/29/2016 3:53:36 AM



HRMS of 17-Trp<sub>2</sub>

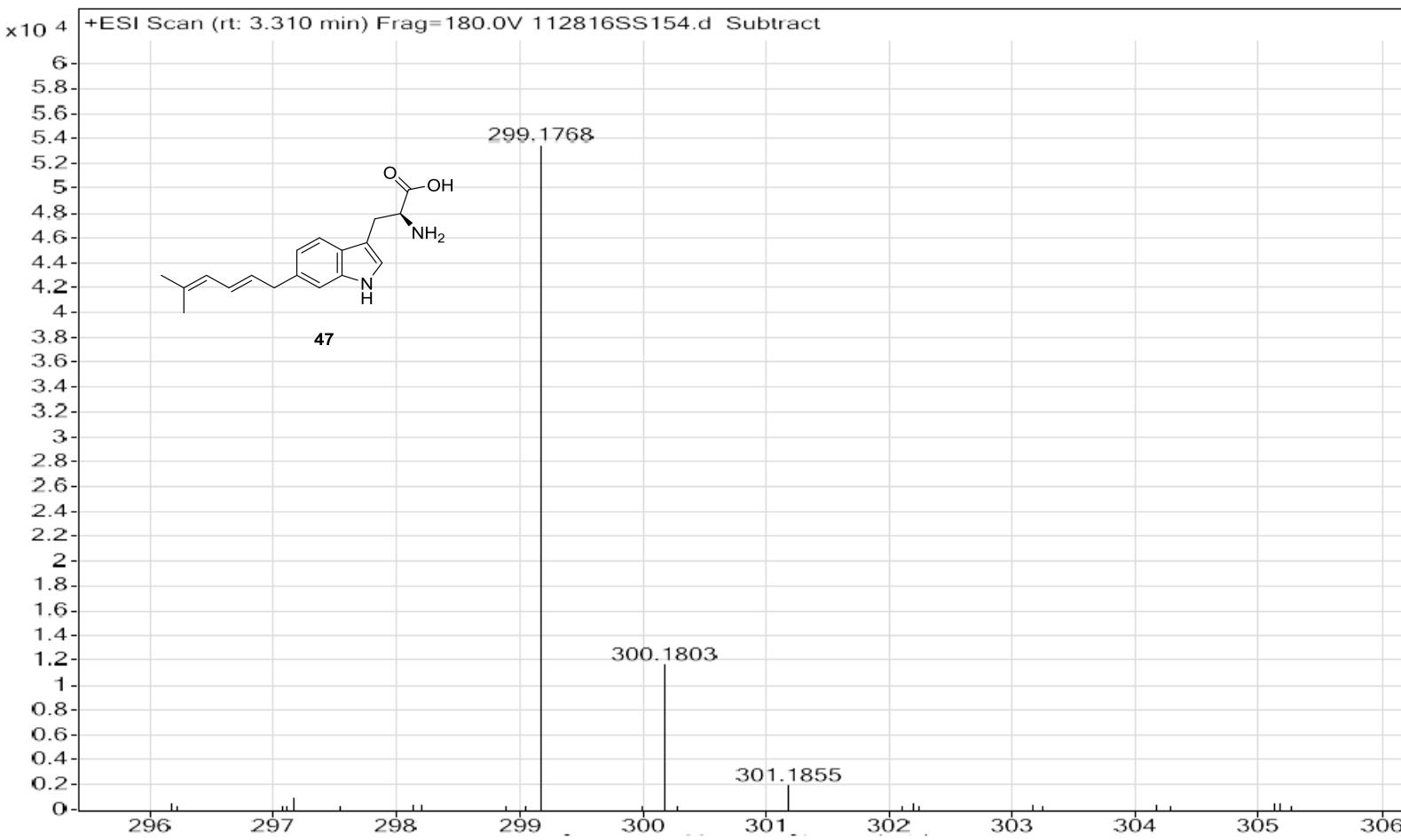


<sup>1</sup>H NMR Spectrum (500 MHz) of compound **47** in MeOH-d4



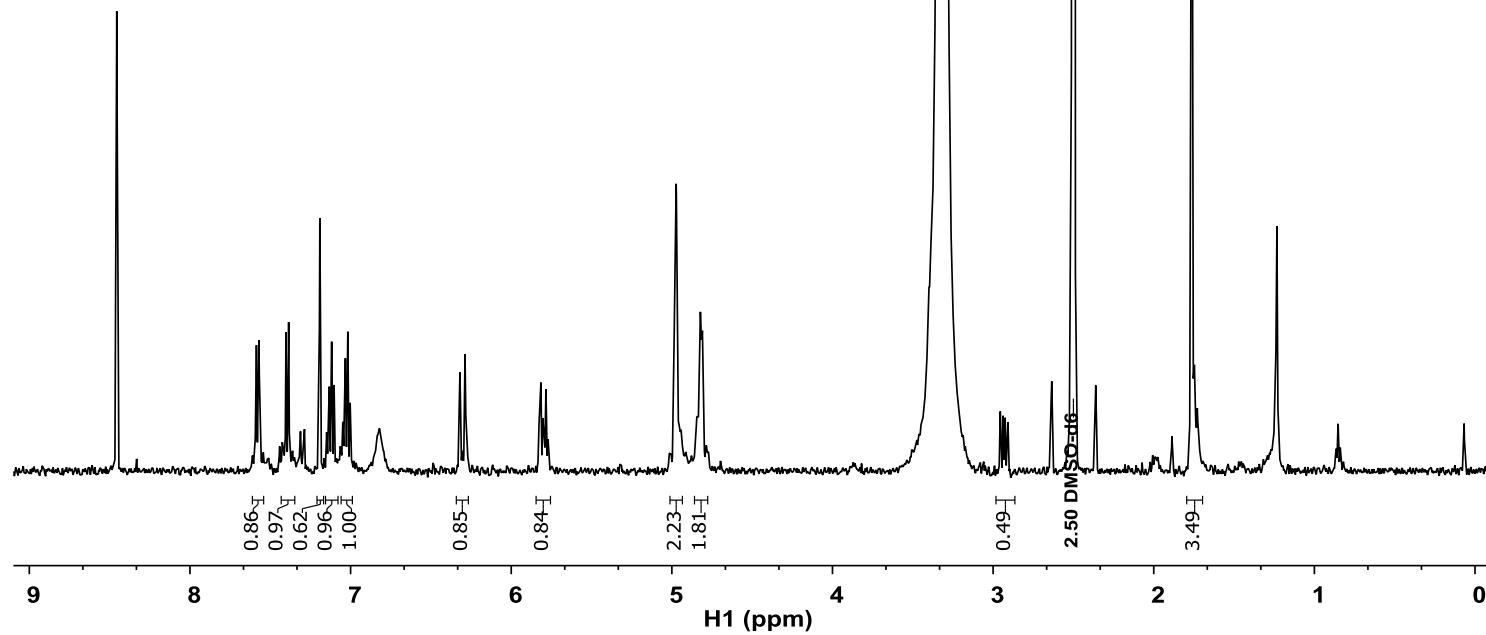
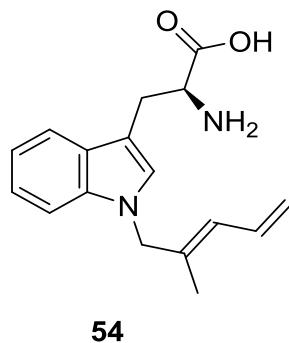
2D  $^1\text{H}$ - $^1\text{H}$  COSY NMR spectrum (500 MHz) of **47** in MeOH-d4

Sample Name	TRP-50	Position	P1-F5	Instrument Name	Instrument 1	User Name	
Inj Vol	5	Inj Position		SampleType	Sample	IRM Calibration Status	Success
Data Filename	112816SS154.d	ACQ Method	drugC18_pos_ms.m	Comment		Acquired Time	11/29/2016 4:25:41 PM

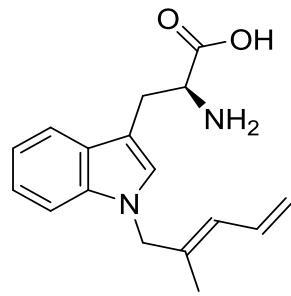


HRMS spectrum of 47

A19-Trp\_1H

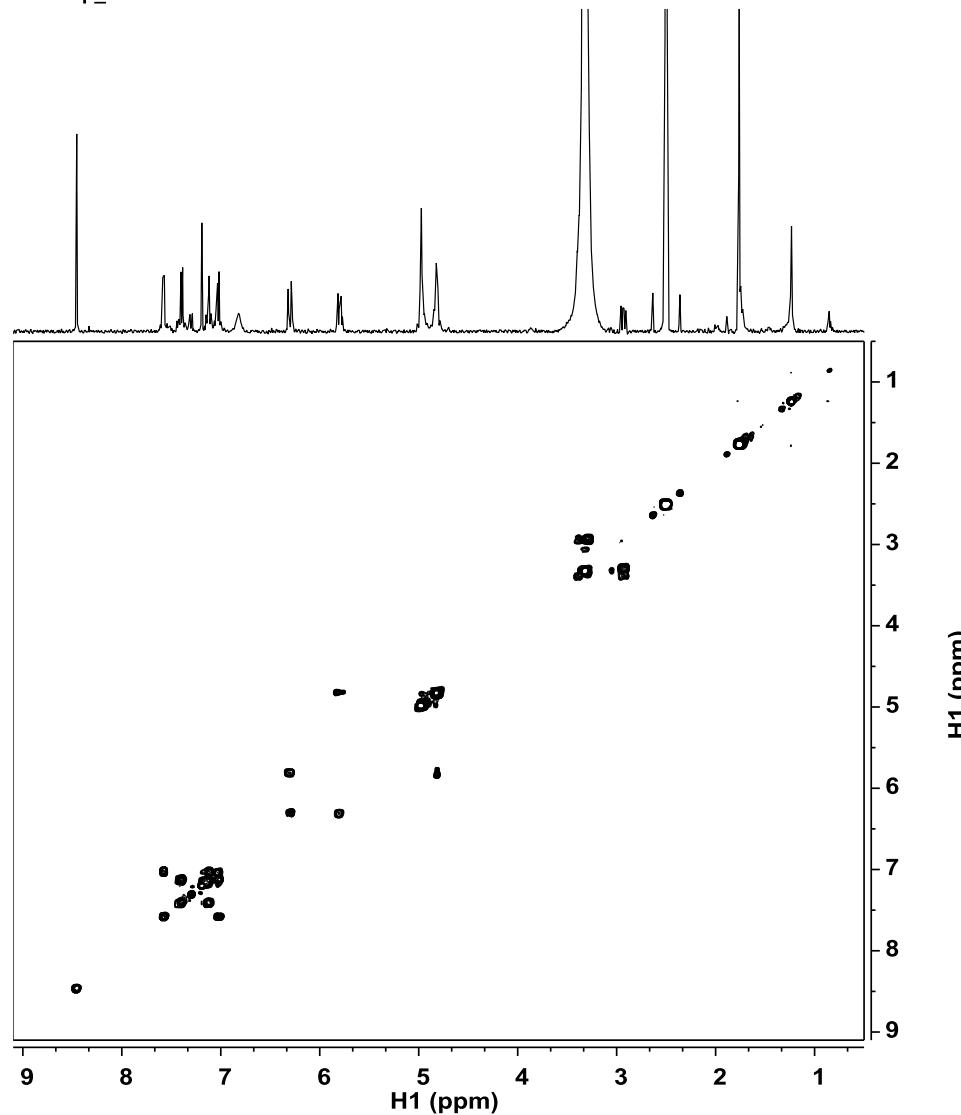


$^1\text{H}$  NMR Spectrum (500 MHz) of compound **54** in  $\text{DMSO-d}_6$



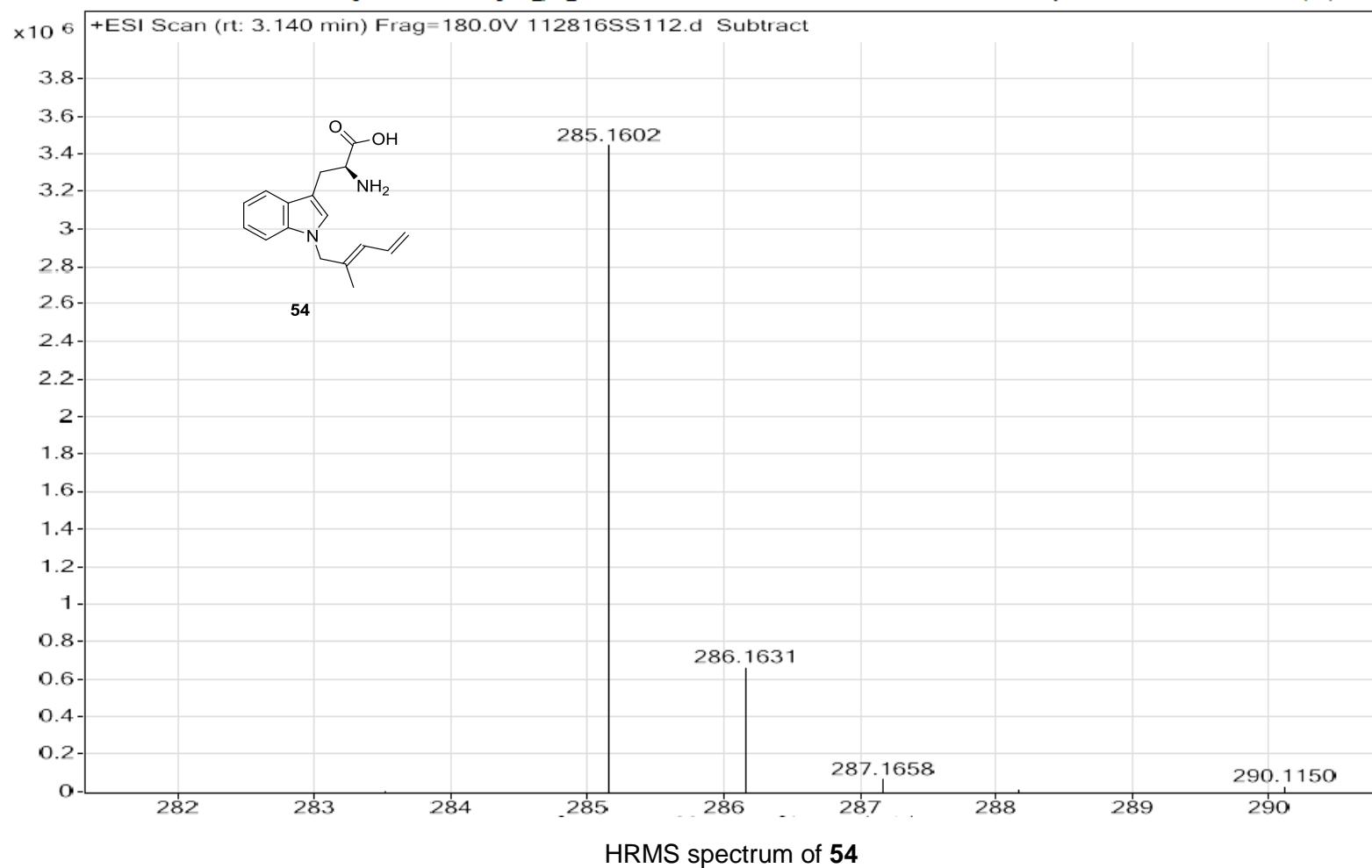
54

A19-Trp\_COSY

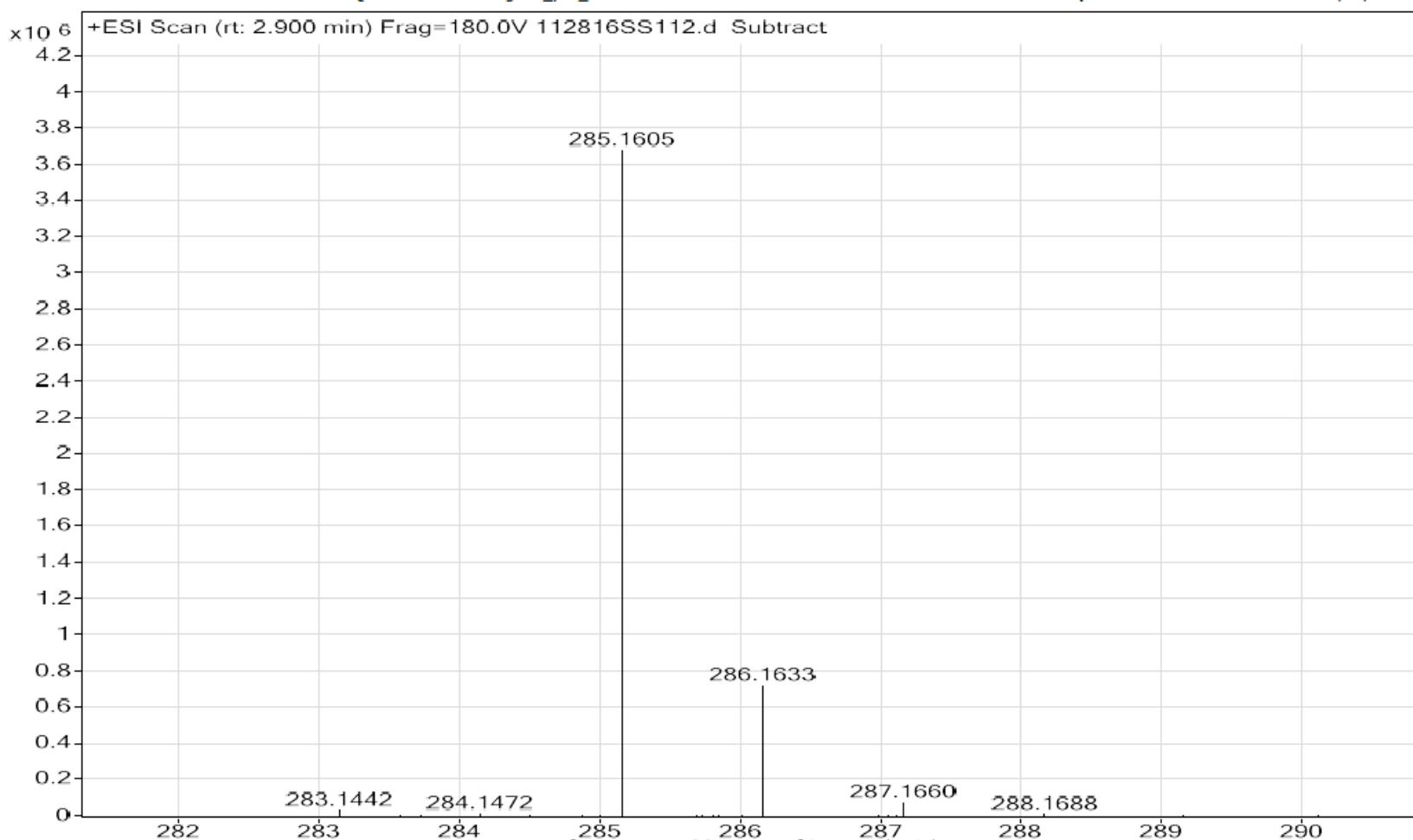


2D  $^1\text{H}$ - $^1\text{H}$  COSY NMR spectrum (500 MHz) of 54 in DMSO-d6

Sample Name	TRP-36	Position	P1-D9	Instrument Name	Instrument 1	User Name	
Inj Vol	5	Inj Position		Sample Type	Sample	IRM Calibration Status	Success
Data Filename	112816SS112.d	ACQ Method	drugC18_pos_ms.m	Comment		Acquired Time	11/29/2016 8:47:58 AM

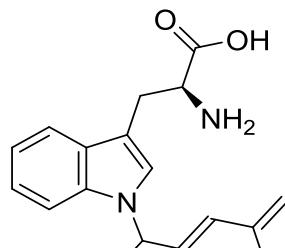


Sample Name	TRP-36	Position	P1-D9	Instrument Name	Instrument 1	User Name	
Inj Vol	5	InjPosition		SampleType	Sample	IRM Calibration Status	Success
Data Filename	112816SS112.d	ACQ Method	drugC18_pos_ms.m	Comment		Acquired Time	11/29/2016 8:47:58 AM

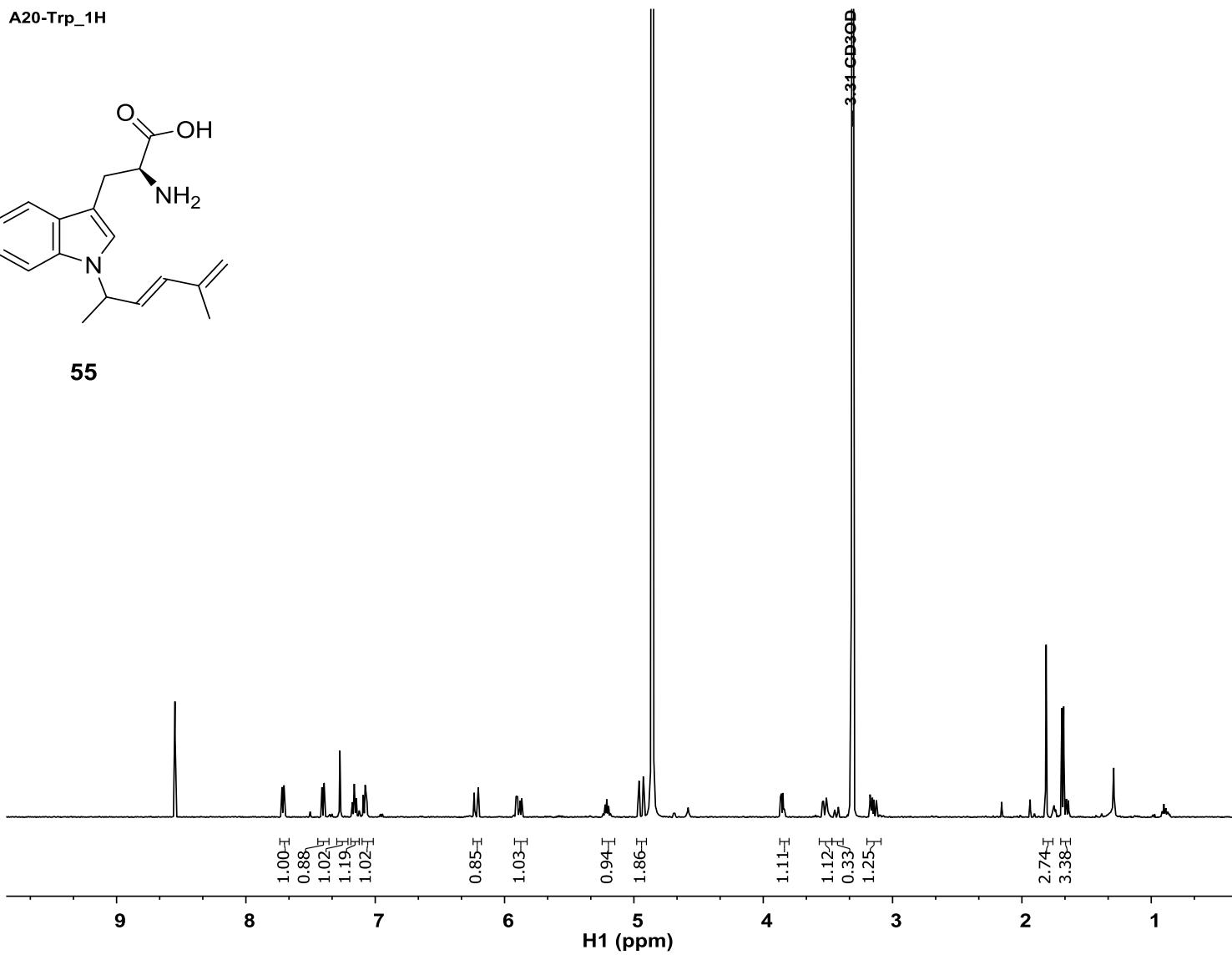


HRMS of 19-Trp<sub>2</sub>

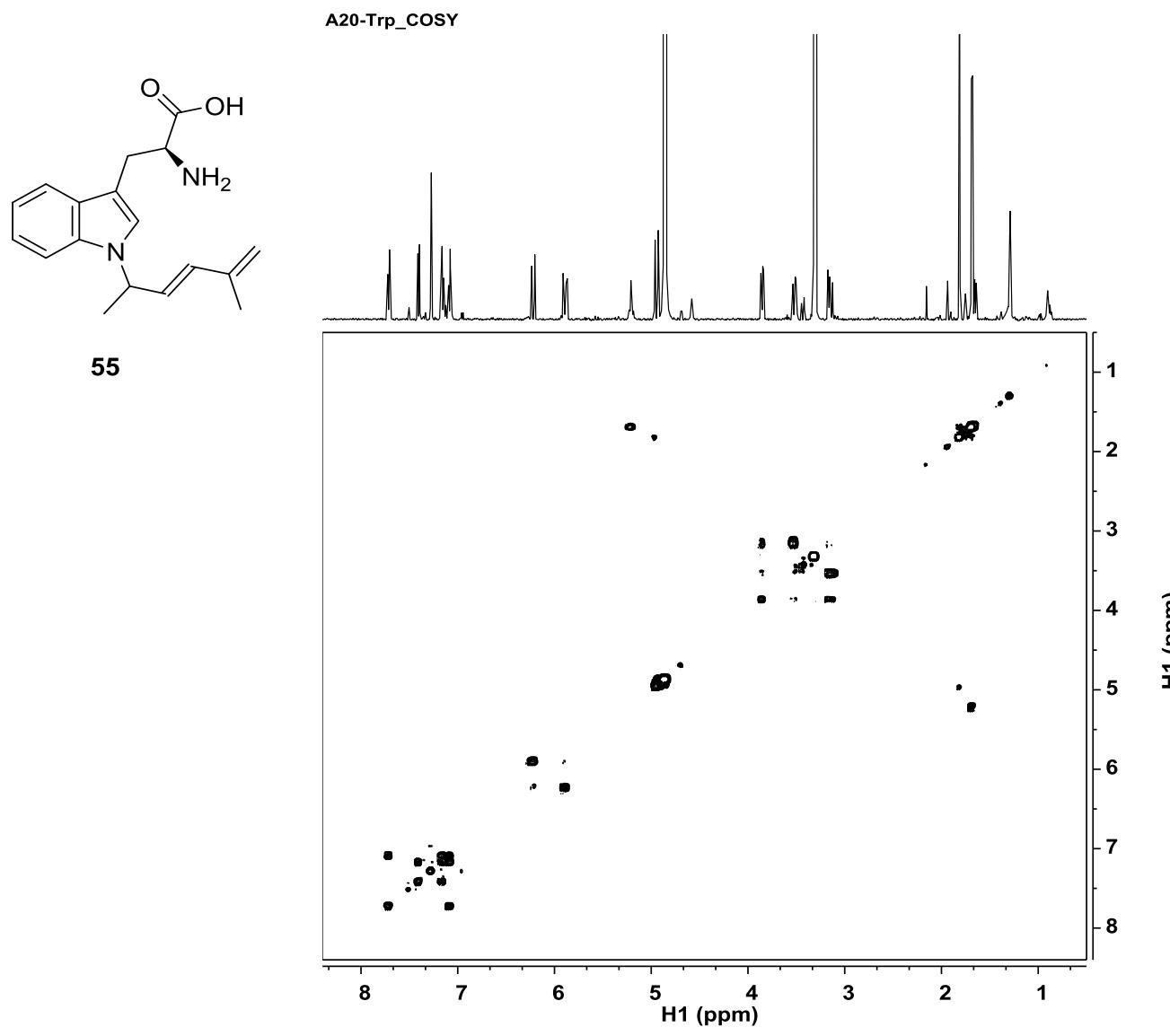
A20-Trp\_1H



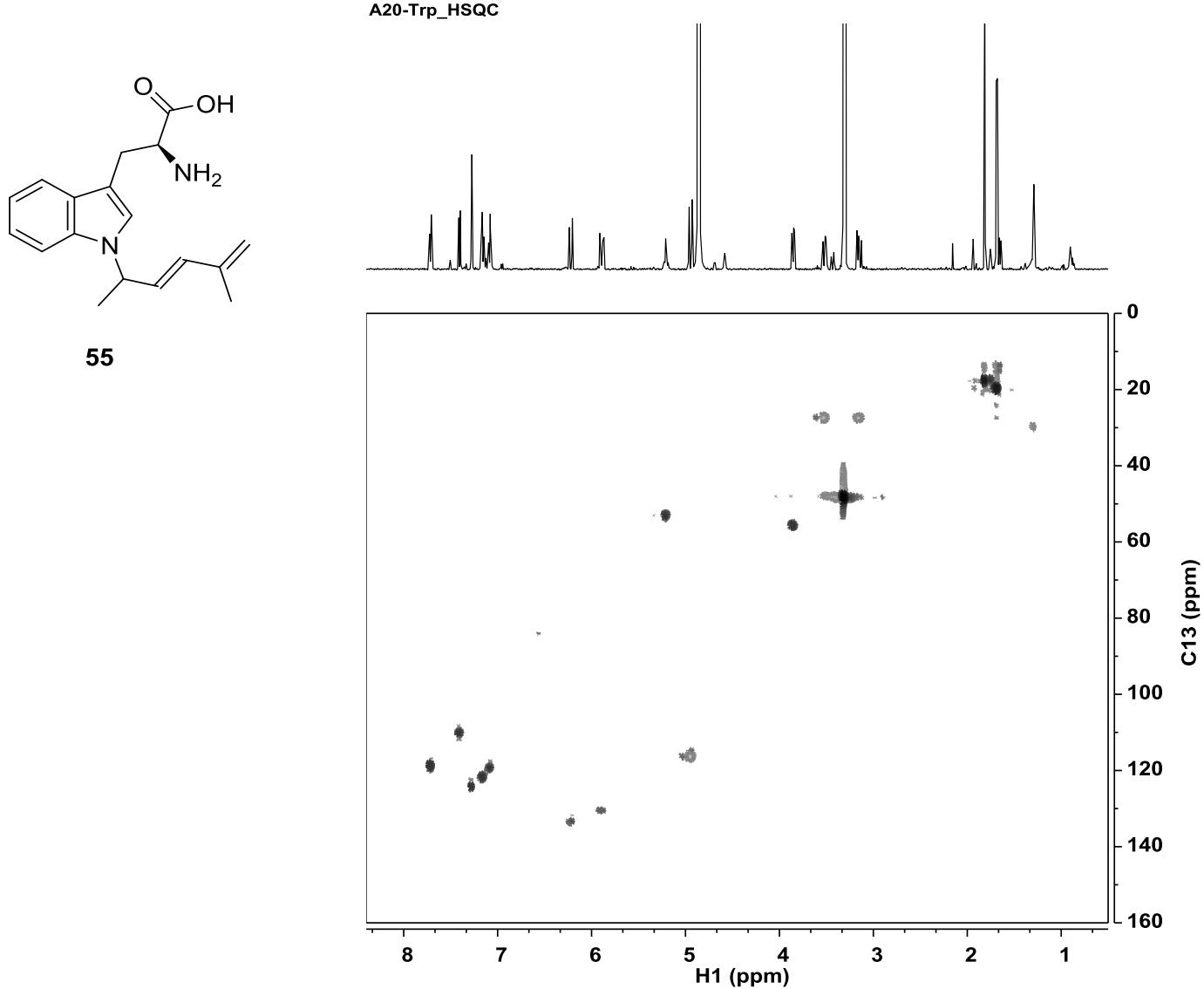
55



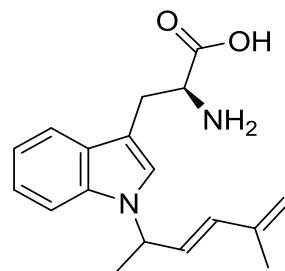
<sup>1</sup>H NMR Spectrum (500 MHz) of compound 55 in MeOH-d4



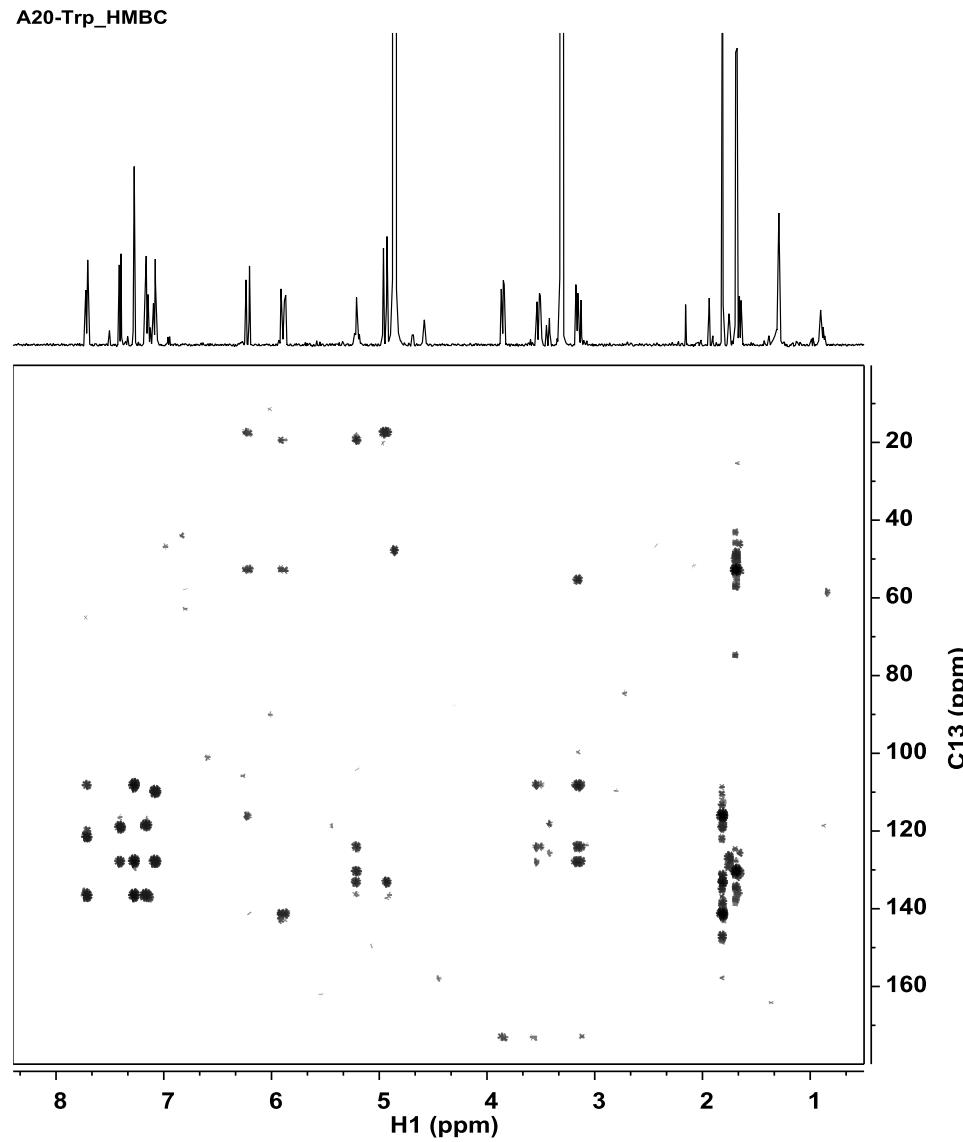
2D  $^1\text{H}$ - $^1\text{H}$  COSY NMR spectrum (500 MHz) of **55** in  $\text{MeOH-d}_4$



2D  $^1\text{H}$ - $^{13}\text{C}$  HSQC NMR spectrum (500 MHz) of **55** in MeOH-d4

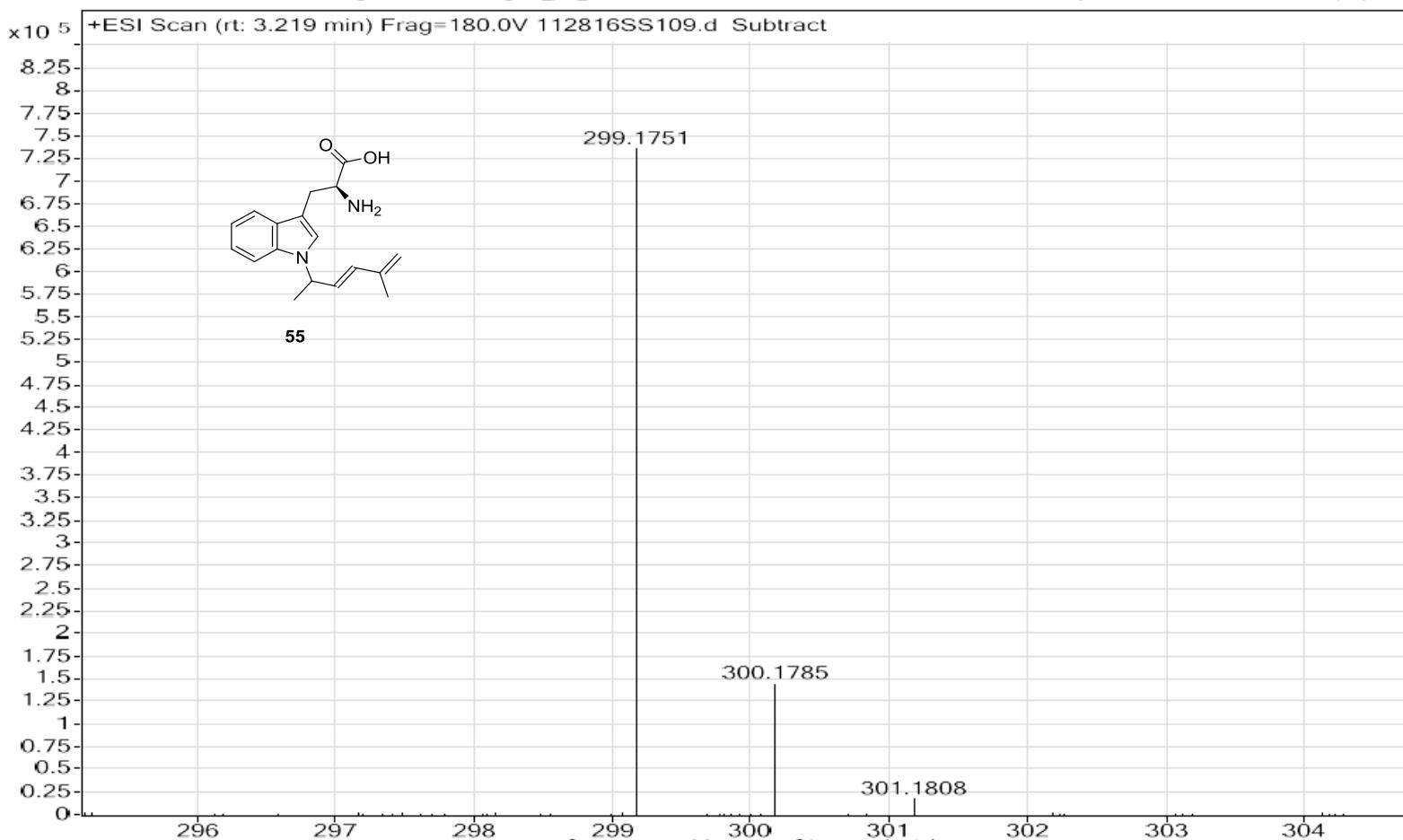


55



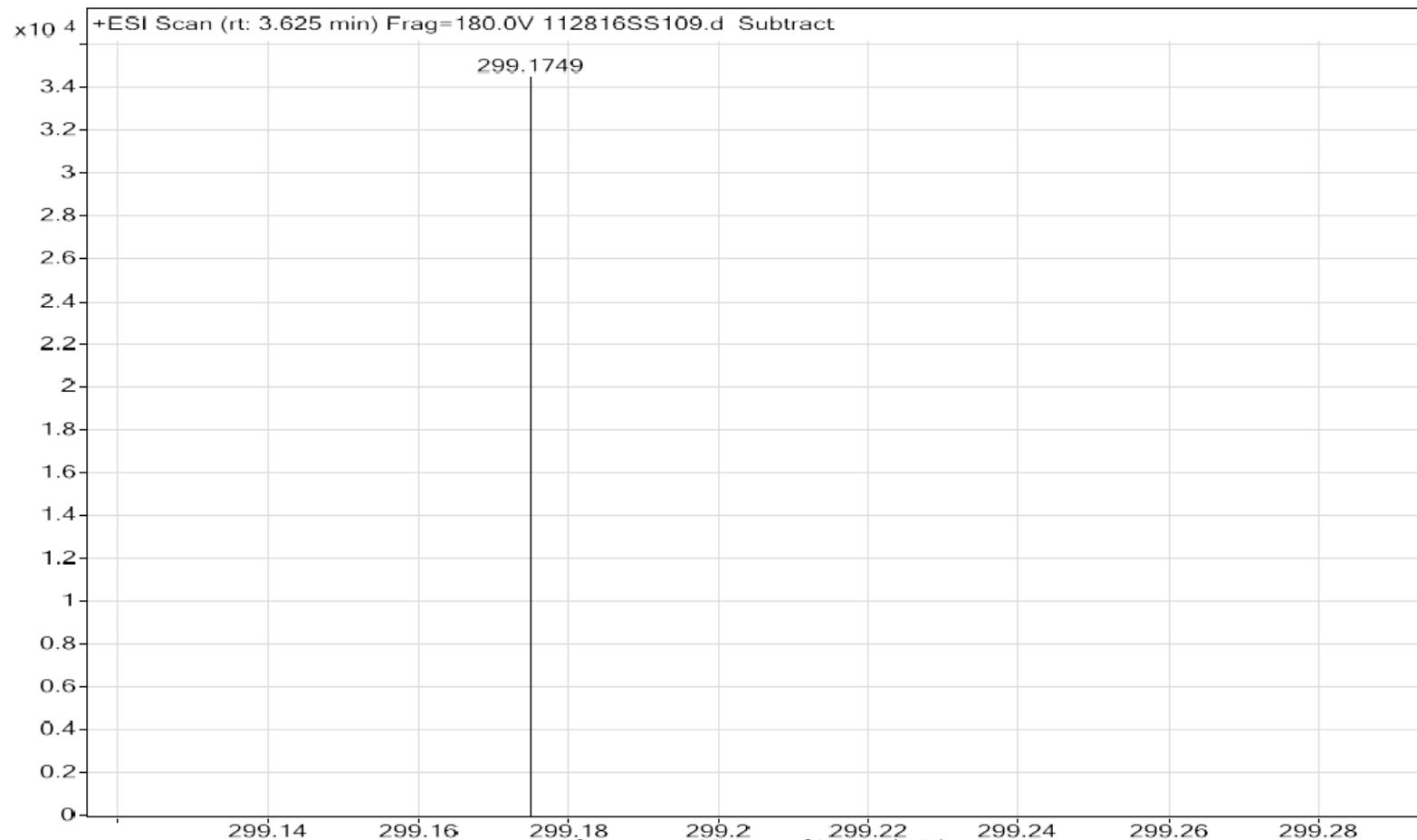
2D  $^1\text{H}$ - $^{13}\text{C}$  HMBC NMR spectrum (500 MHz) of **55** in  $\text{MeOH-d}_4$

<b>Sample Name</b>	TRP-35	<b>Position</b>	P1-D8	<b>Instrument Name</b>	Instrument 1	<b>User Name</b>	
<b>Inj Vol</b>	5	<b>InjPosition</b>		<b>SampleType</b>	Sample	<b>IRM Calibration Status</b>	Success
<b>Data Filename</b>	112816SS109.d	<b>ACQ Method</b>	drugC18_pos_ms.m	<b>Comment</b>		<b>Acquired Time</b>	11/29/2016 8:15:14 AM



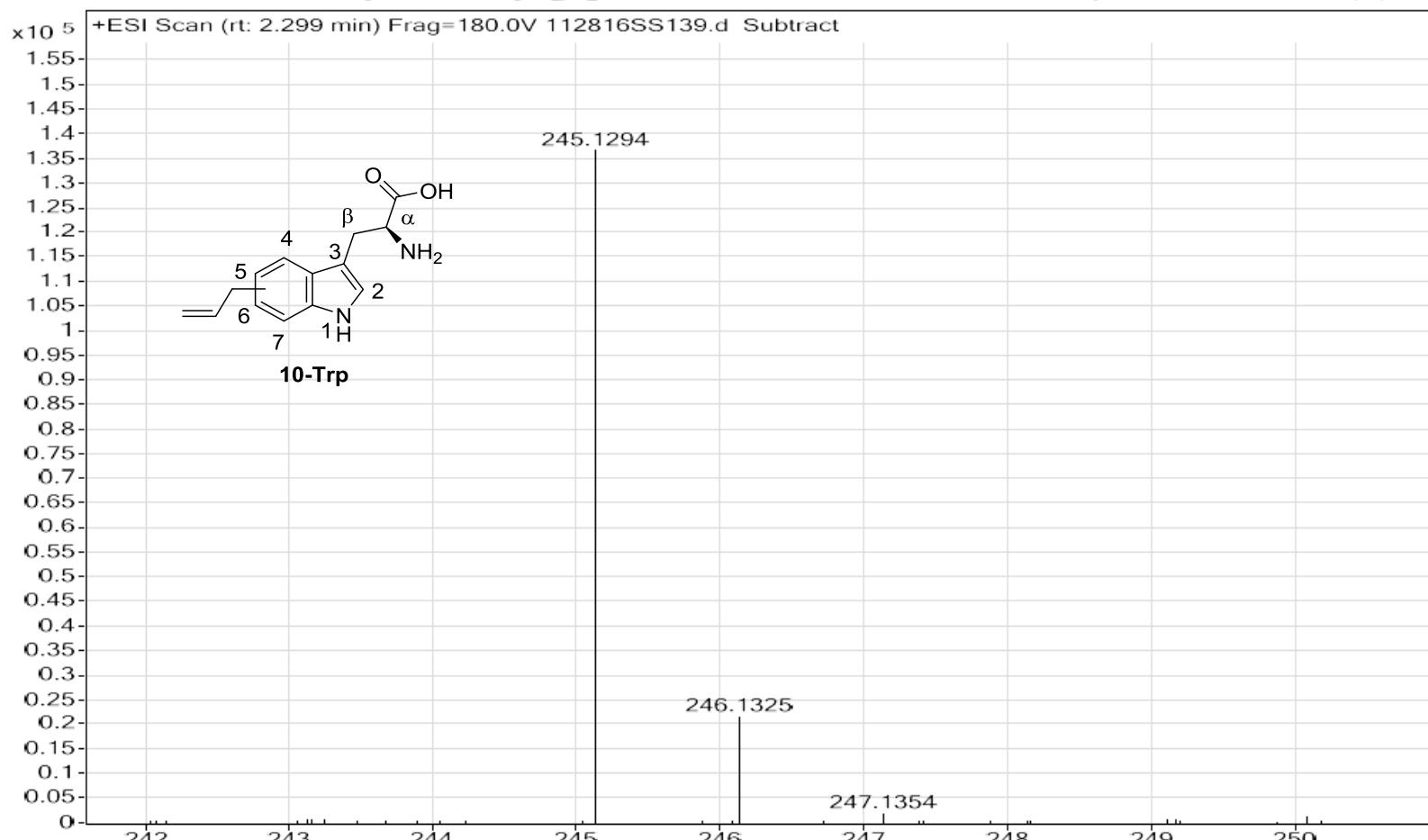
## HRMS spectrum of 55

Sample Name	TRP-35	Position	P1-D8	Instrument Name	Instrument 1	User Name	
Inj Vol	5	InjPosition		SampleType	Sample	IRM Calibration Status	Success
Data Filename	112816SS109.d	ACQ Method	drugC18_pos_ms.m	Comment		Acquired Time	11/29/2016 8:15:14 AM



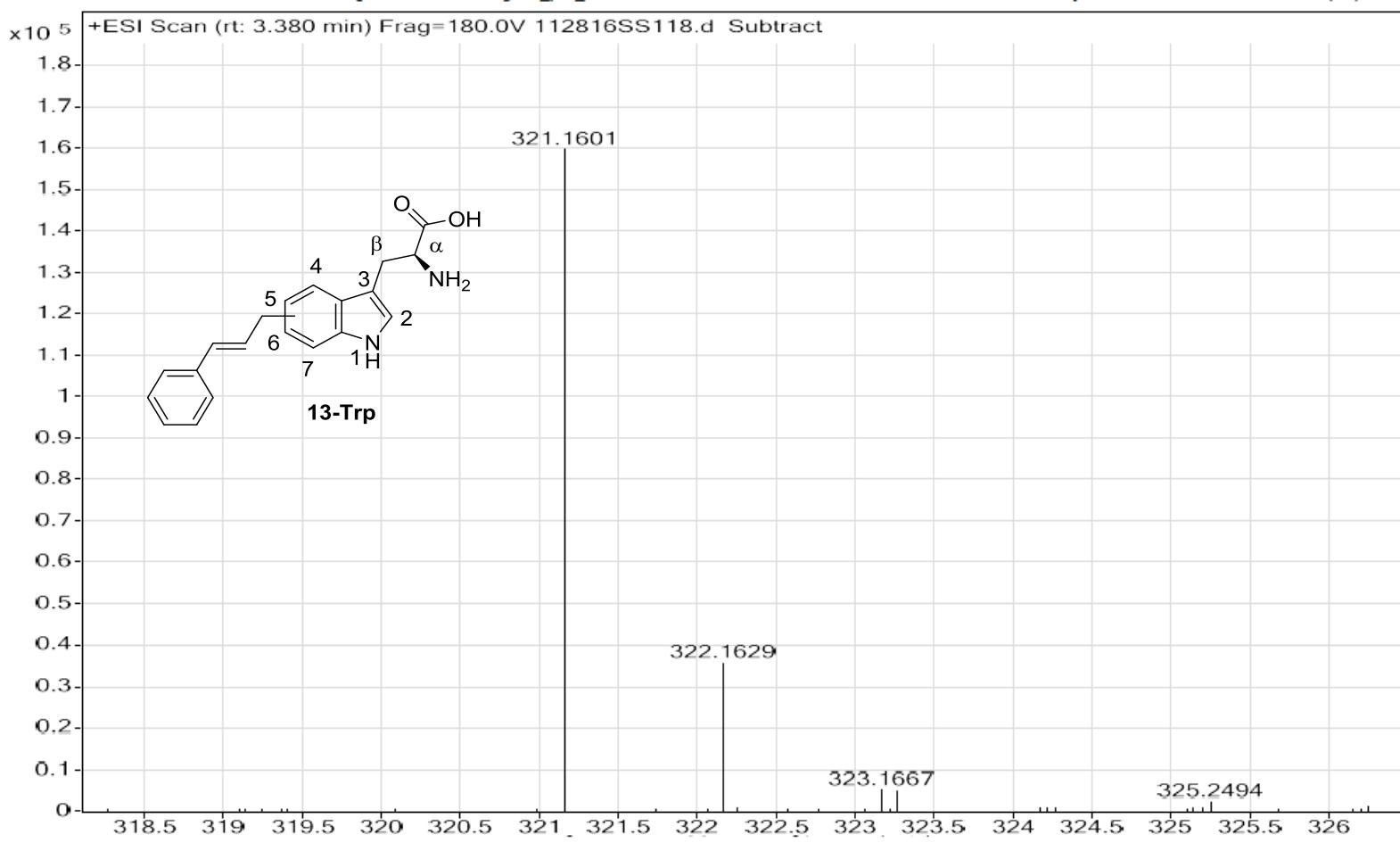
**HRMS of 20-Trp<sub>2</sub>**

Sample Name	TRP-45	Position	P1-E9	Instrument Name	Instrument 1	User Name	
Inj Vol	5	InjPosition		SampleType	Sample	IRM Calibration Status	Success
Data Filename	112816SS139.d	ACQ Method	drugC18_pos_ms.m	Comment		Acquired Time	11/29/2016 1:42:16 PM



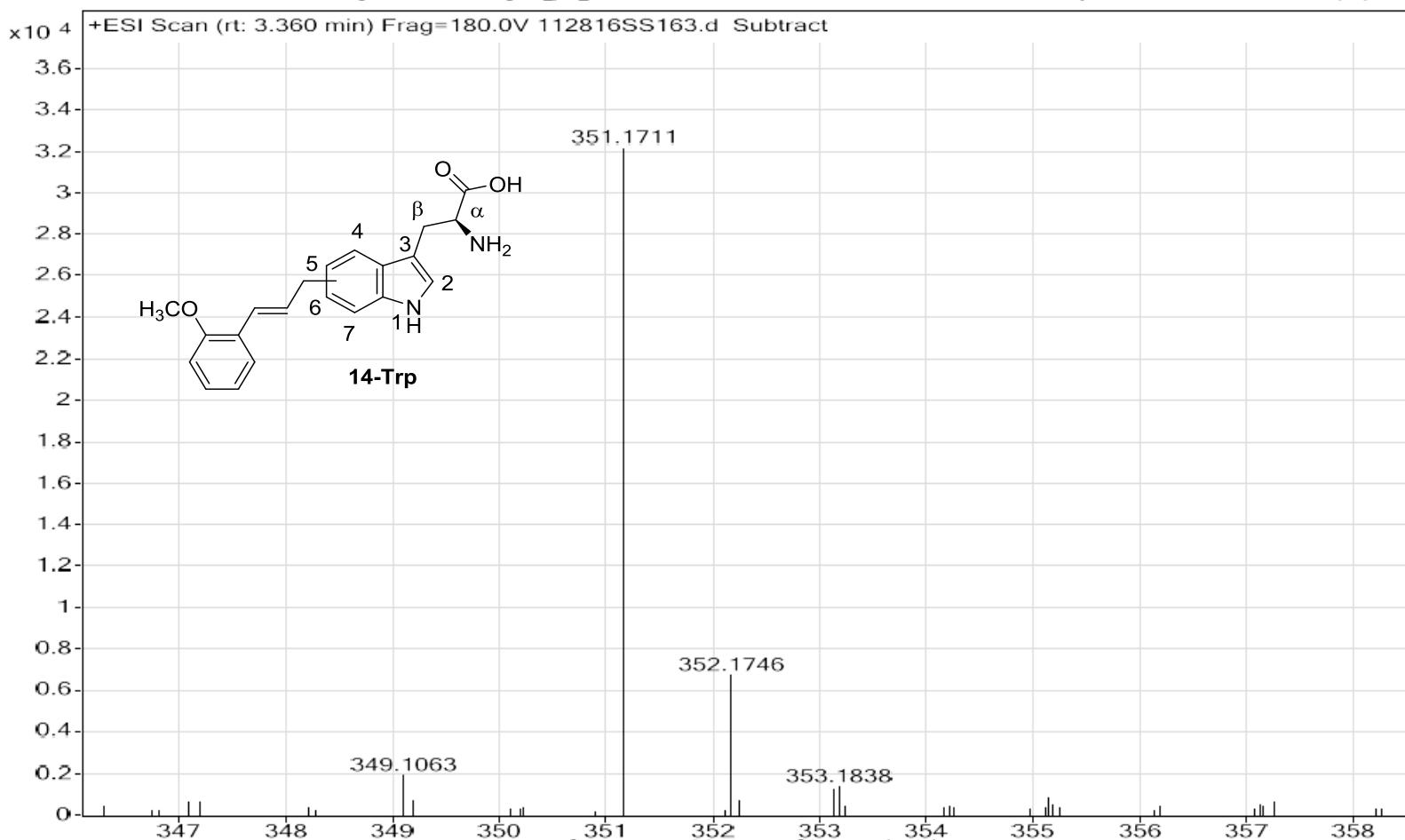
HRMS spectrum of 10-Trp

Sample Name	TRP-38	Position	P1-E2	Instrument Name	Instrument 1	User Name	
Inj Vol	5	Inj Position		SampleType	Sample	IRM Calibration Status	Success
Data Filename	112816SS118.d	ACQ Method	drugC18_pos_ms.m	Comment		Acquired Time	11/29/2016 9:53:21 AM



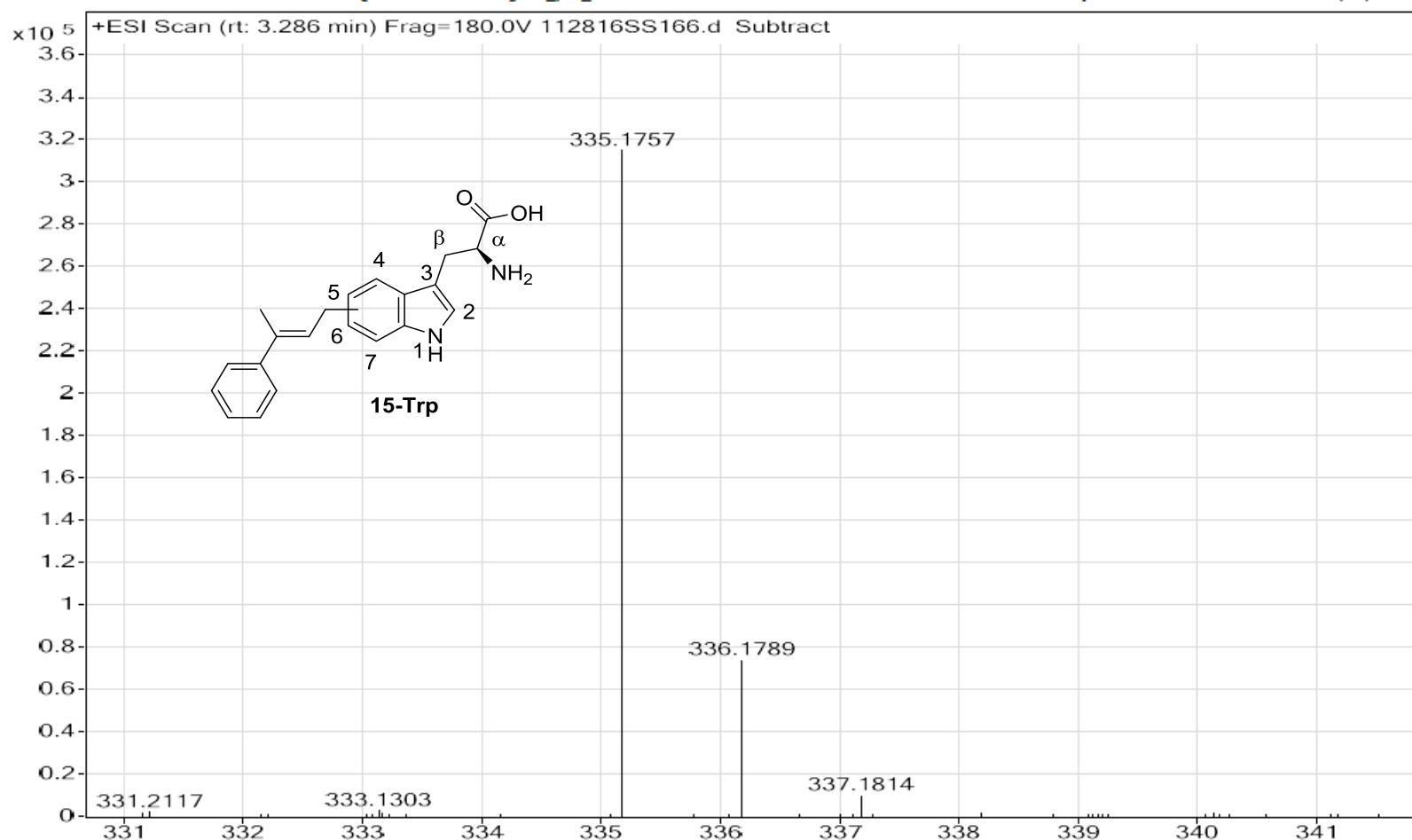
HRMS spectrum of 13-Trp

Sample Name	TRP-53	Position	P1-F8	Instrument Name	Instrument 1	User Name	
Inj Vol	5	InjPosition		SampleType	Sample	IRM Calibration Status	Success
Data Filename	112816SS163.d	ACQ Method	drugC18_pos_ms.m	Comment		Acquired Time	11/29/2016 6:03:53 PM



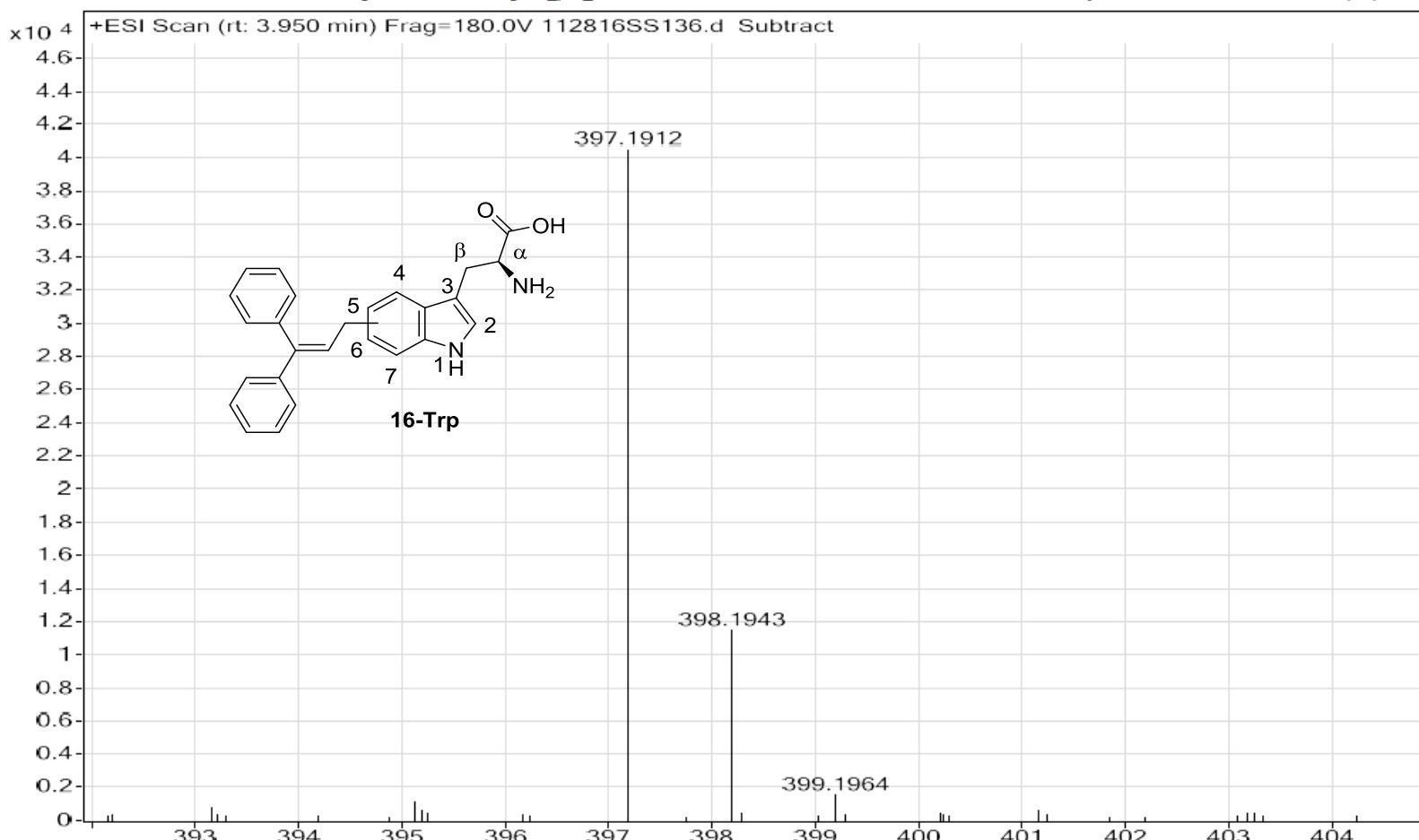
HRMS spectrum of 14-Trp

Sample Name	TRP-54	Position	P1-F9	Instrument Name	Instrument 1	User Name	
Inj Vol	5	InjPosition		SampleType	Sample	IRM Calibration Status	Success
Data Filename	112816SS166.d	ACQ Method	drugC18_pos_ms.m	Comment		Acquired Time	11/29/2016 6:36:34 PM



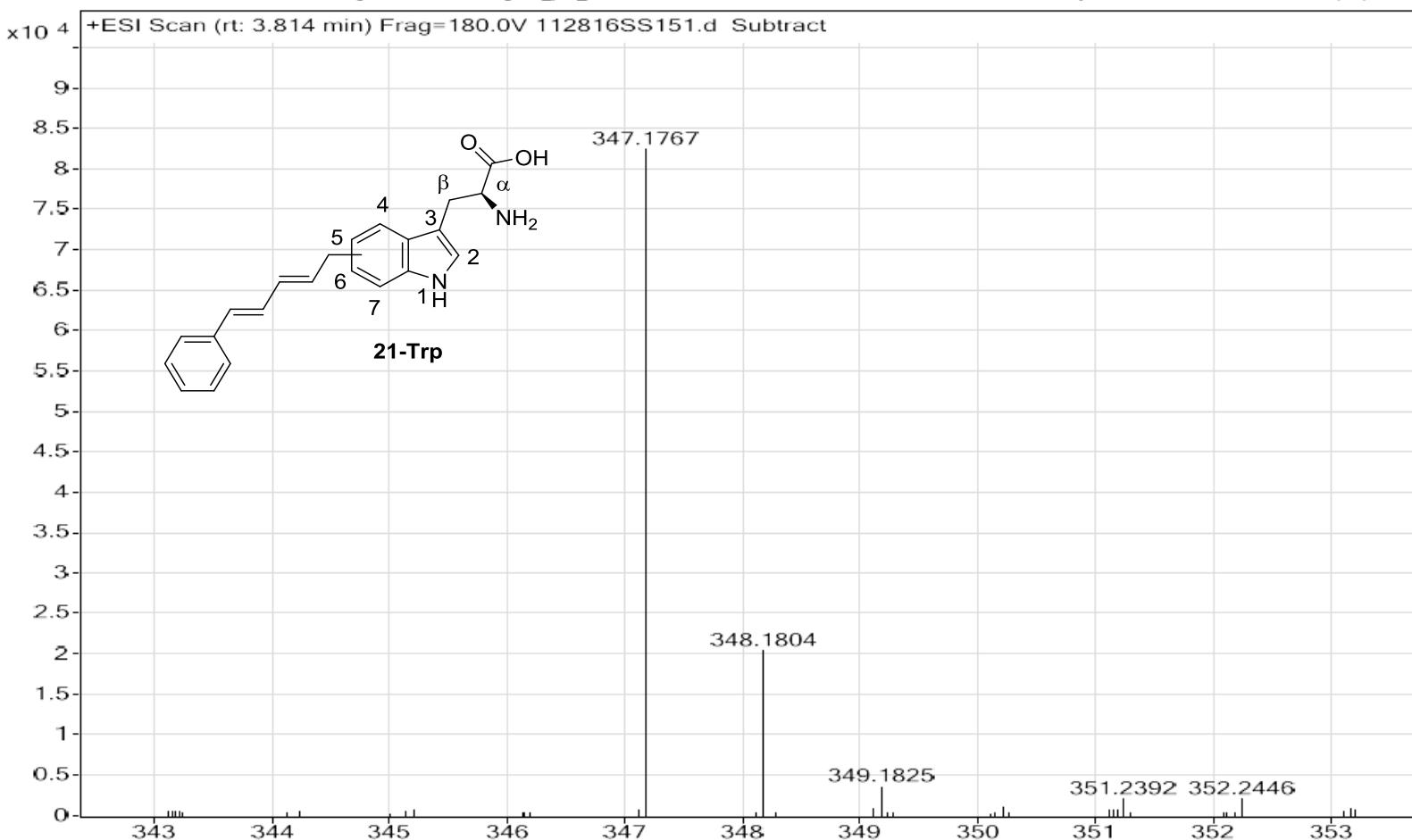
HRMS spectrum of 15-Trp

Sample Name	TRP-44	Position	P1-E8	Instrument Name	Instrument 1	User Name	
Inj Vol	5	InjPosition		SampleType	Sample	IRM Calibration Status	Success
Data Filename	112816SS136.d	ACQ Method	drugC18_pos_ms.m	Comment		Acquired Time	11/29/2016 1:09:33 PM



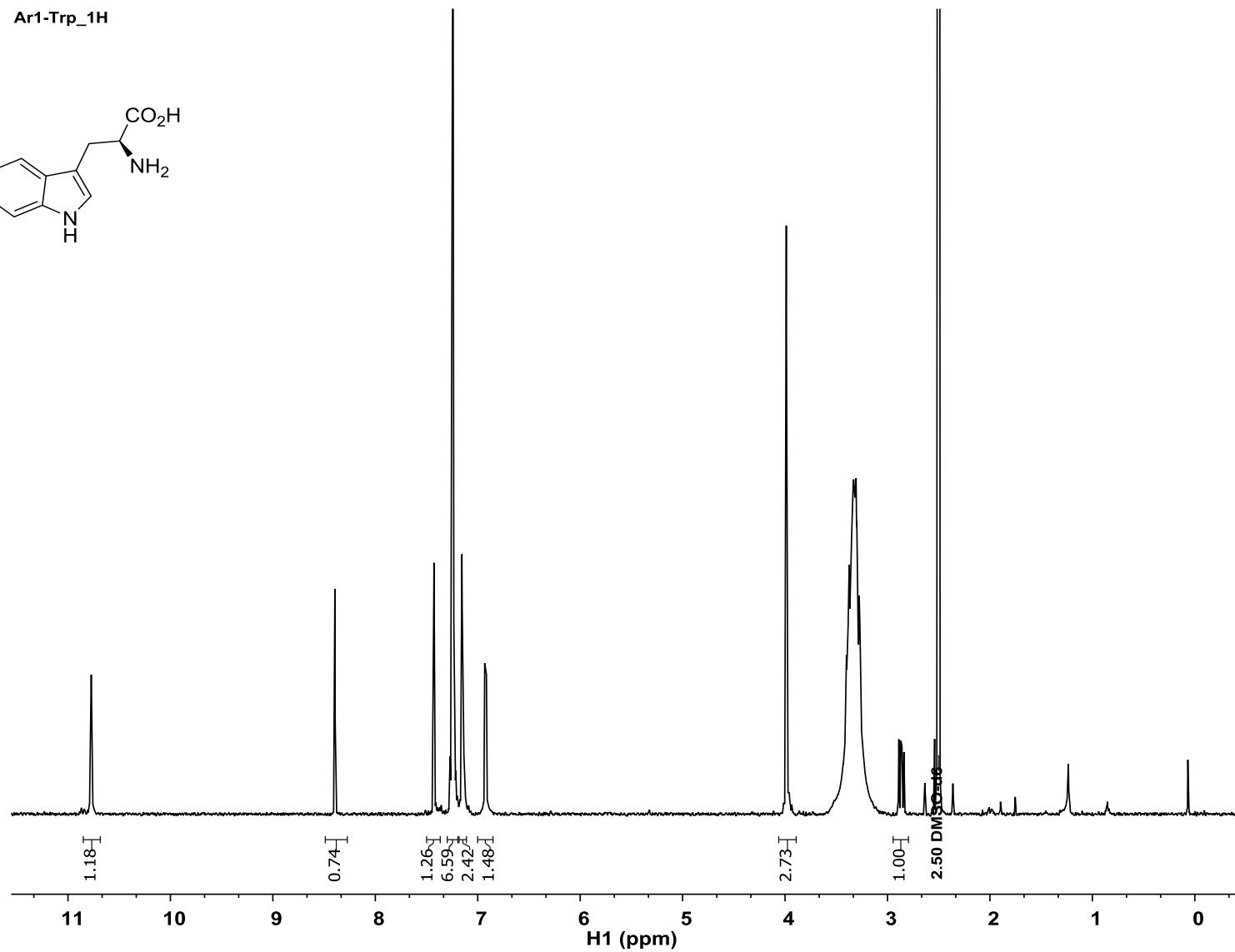
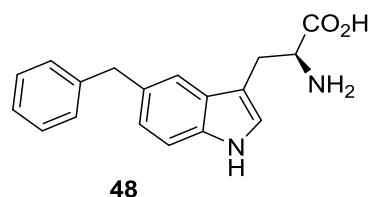
HRMS spectrum of 16-Trp

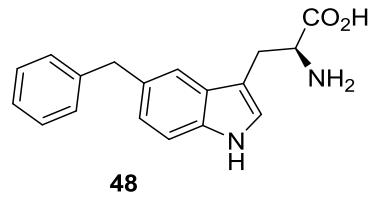
Sample Name	TRP-49	Position	P1-F4	Instrument Name	Instrument 1	User Name	
Inj Vol	5	InjPosition		SampleType	Sample	IRM Calibration Status	Success
Data Filename	112816SS151.d	ACQ Method	drugC18_pos_ms.m	Comment		Acquired Time	11/29/2016 3:53:00 PM



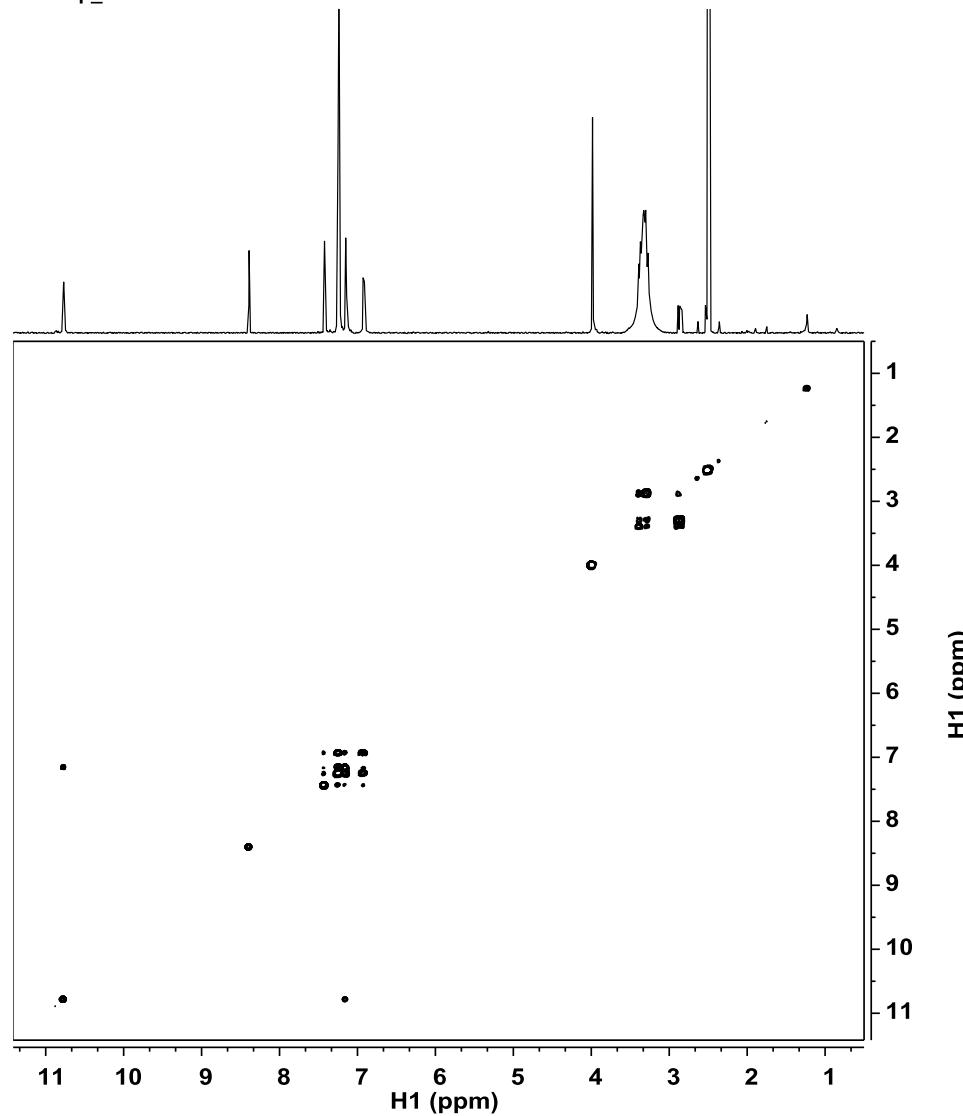
HRMS spectrum of 21-Trp

Ar1-Trp\_1H

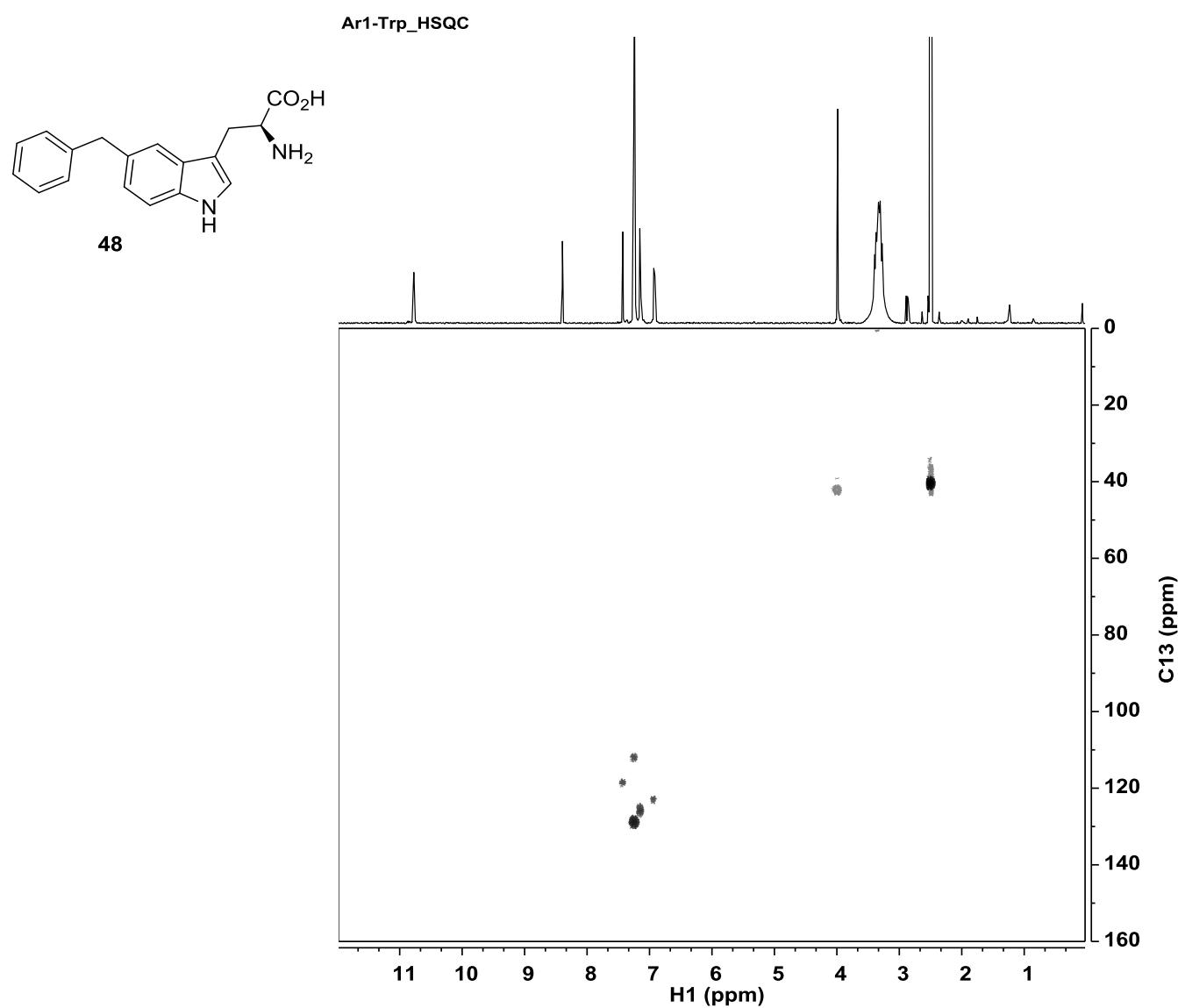




Ar1-Trp\_COSY



2D <sup>1</sup>H-<sup>1</sup>H COSY NMR spectrum (500 MHz) of **48** in DMSO-d<sub>6</sub>



2D  $^1\text{H}$ - $^{13}\text{C}$  COSY NMR spectrum (500 MHz) of **48** in DMSO-d6

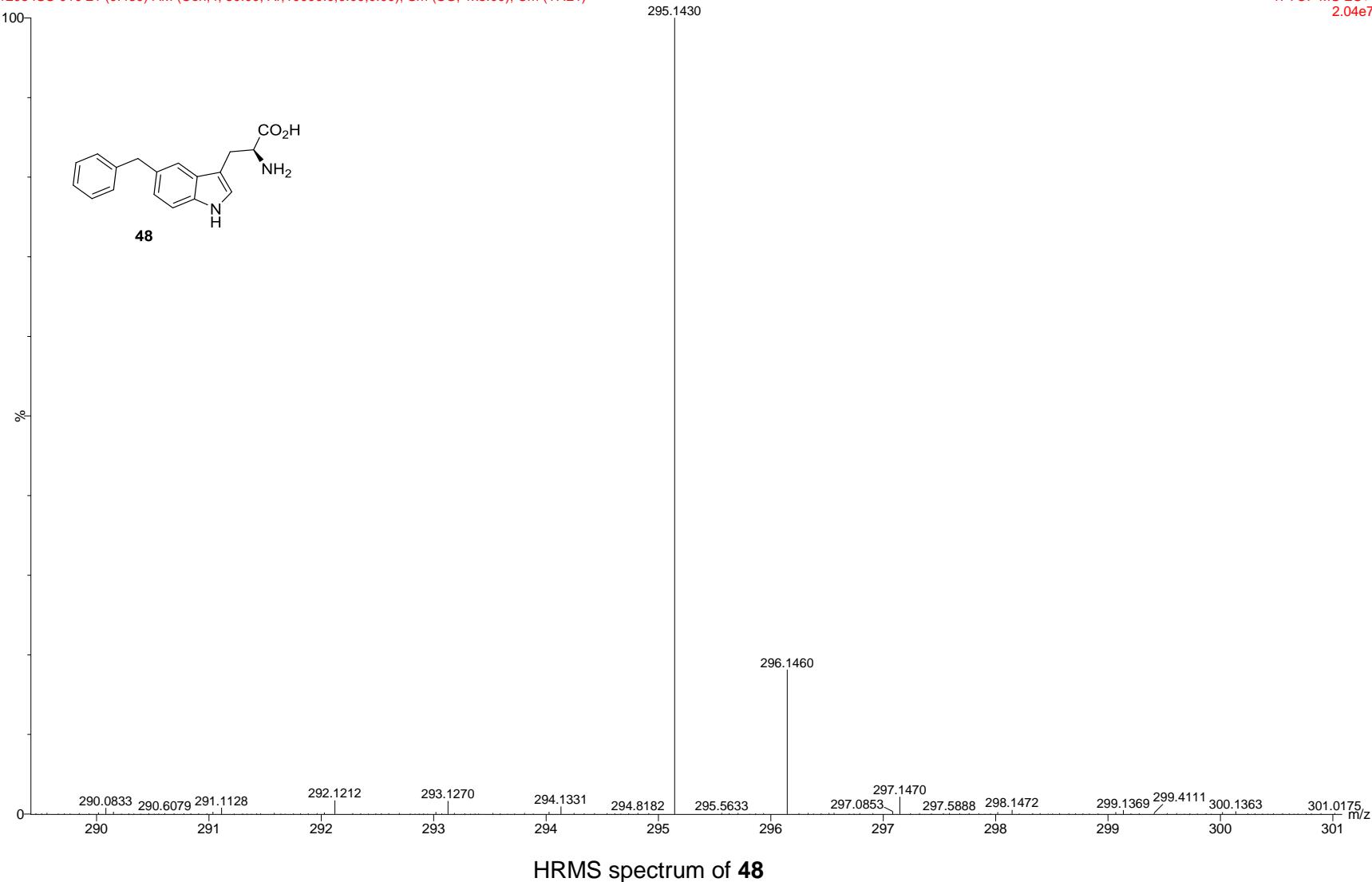
Trp-Ar1  
Bandari

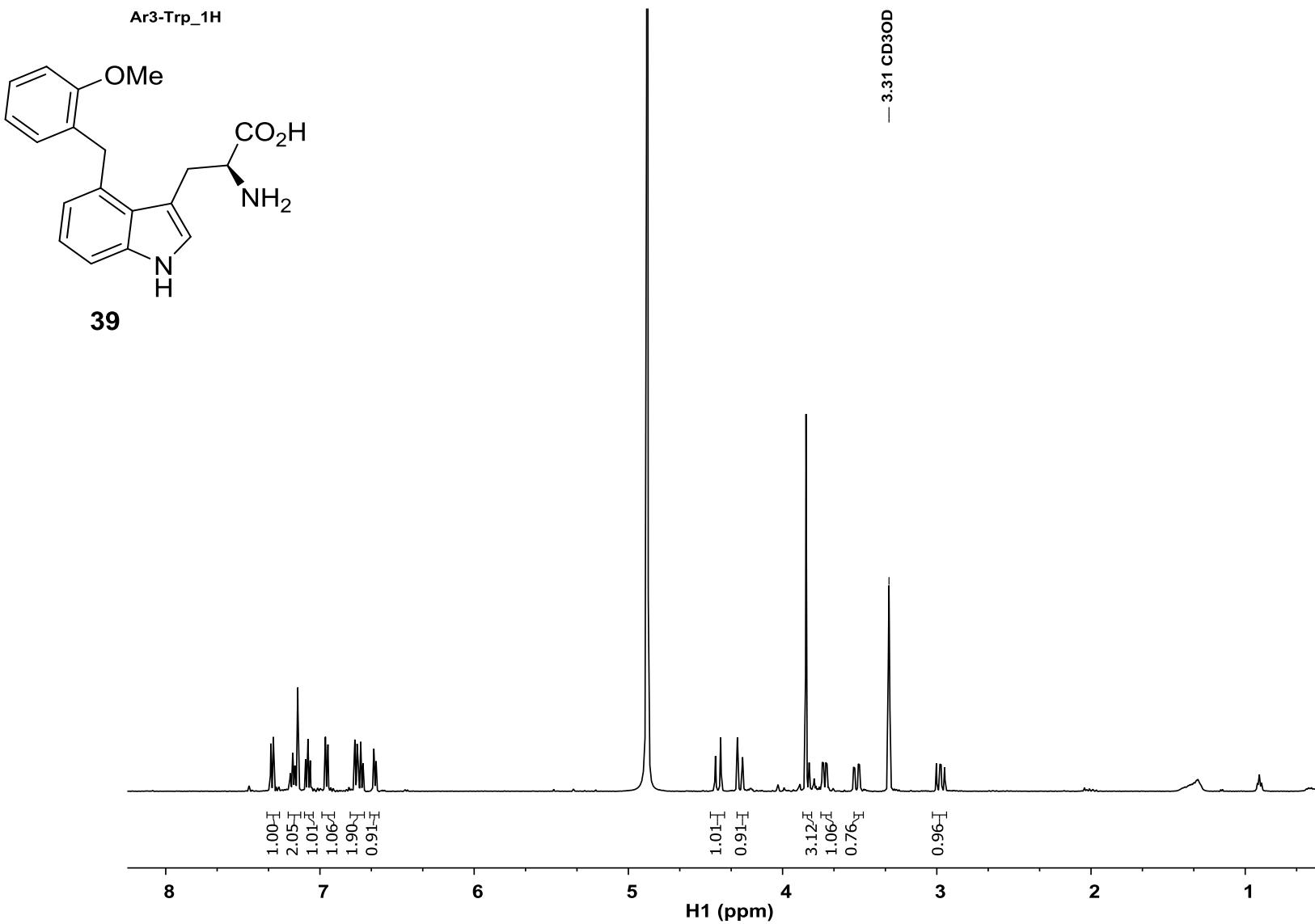
12054SS-010 21 (0.430) AM (Cen,4, 80.00, Ar,10000.0,0.00,0.00); Sm (SG, 1x5.00); Cm (17:21)

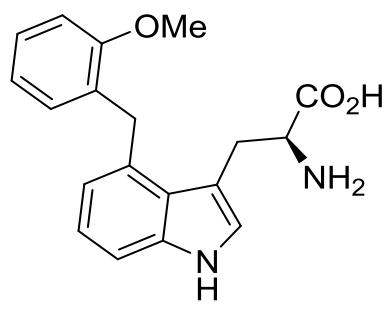
SYNAPTG2-Si#UGA589

29-Aug-2017 17:01:44

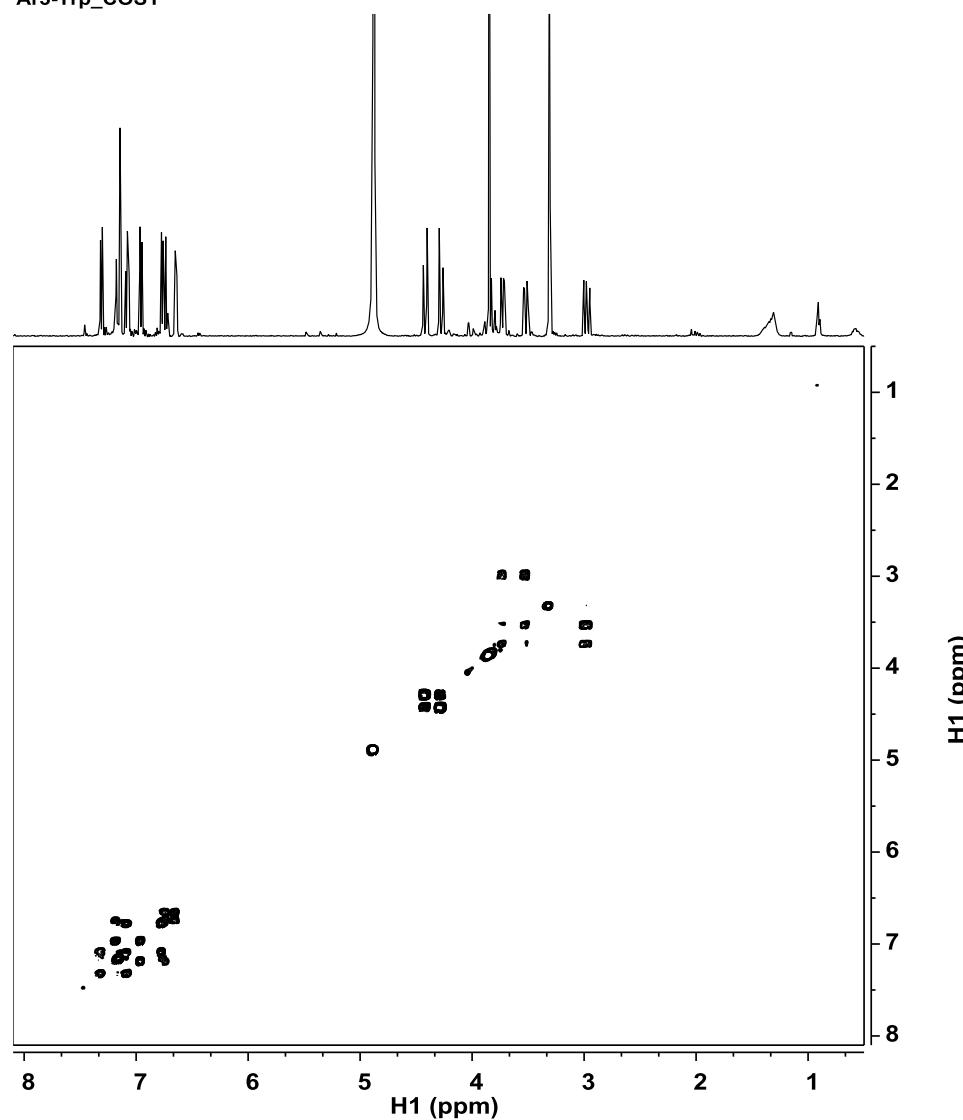
1: TOF MS ES+  
2.04e7



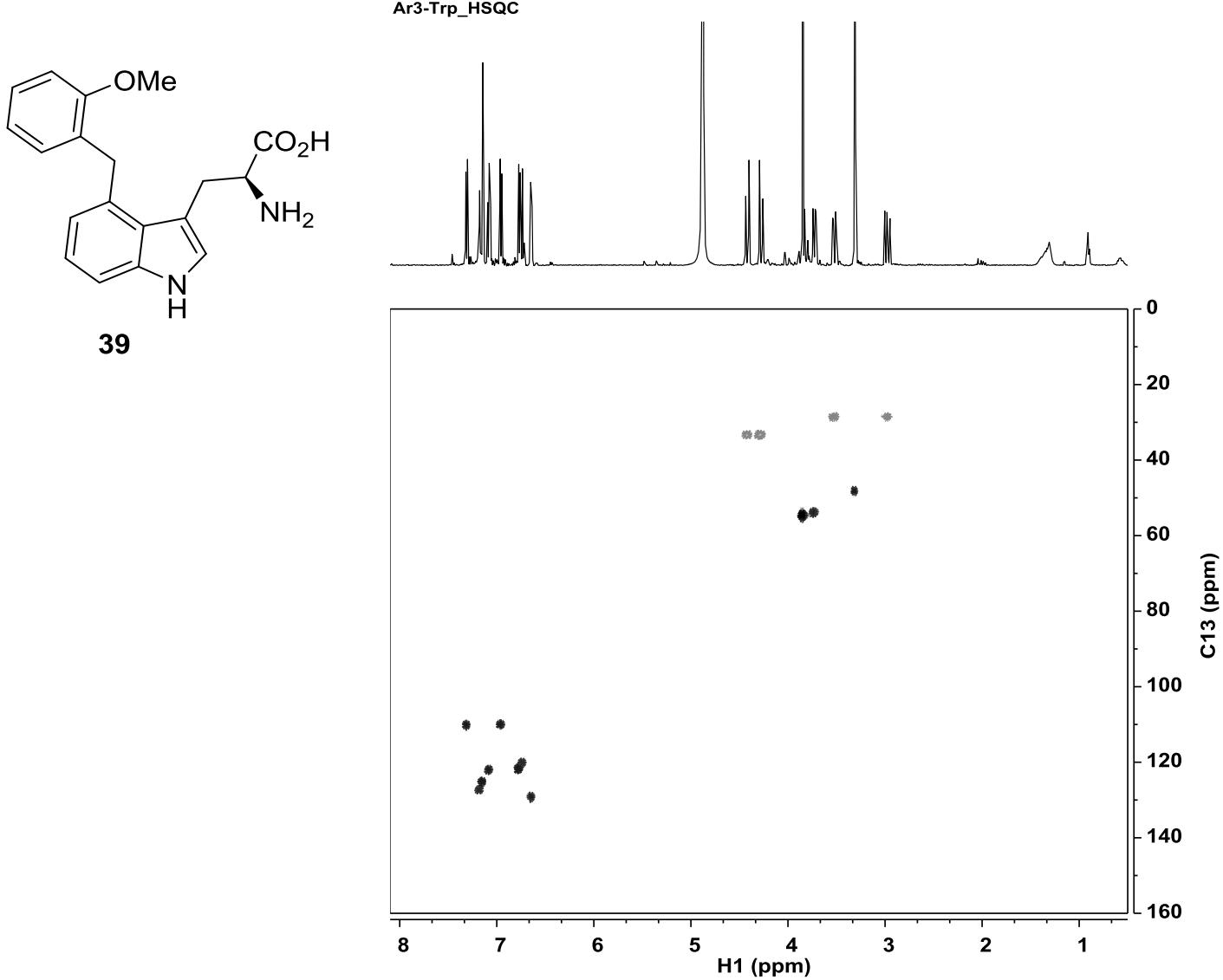




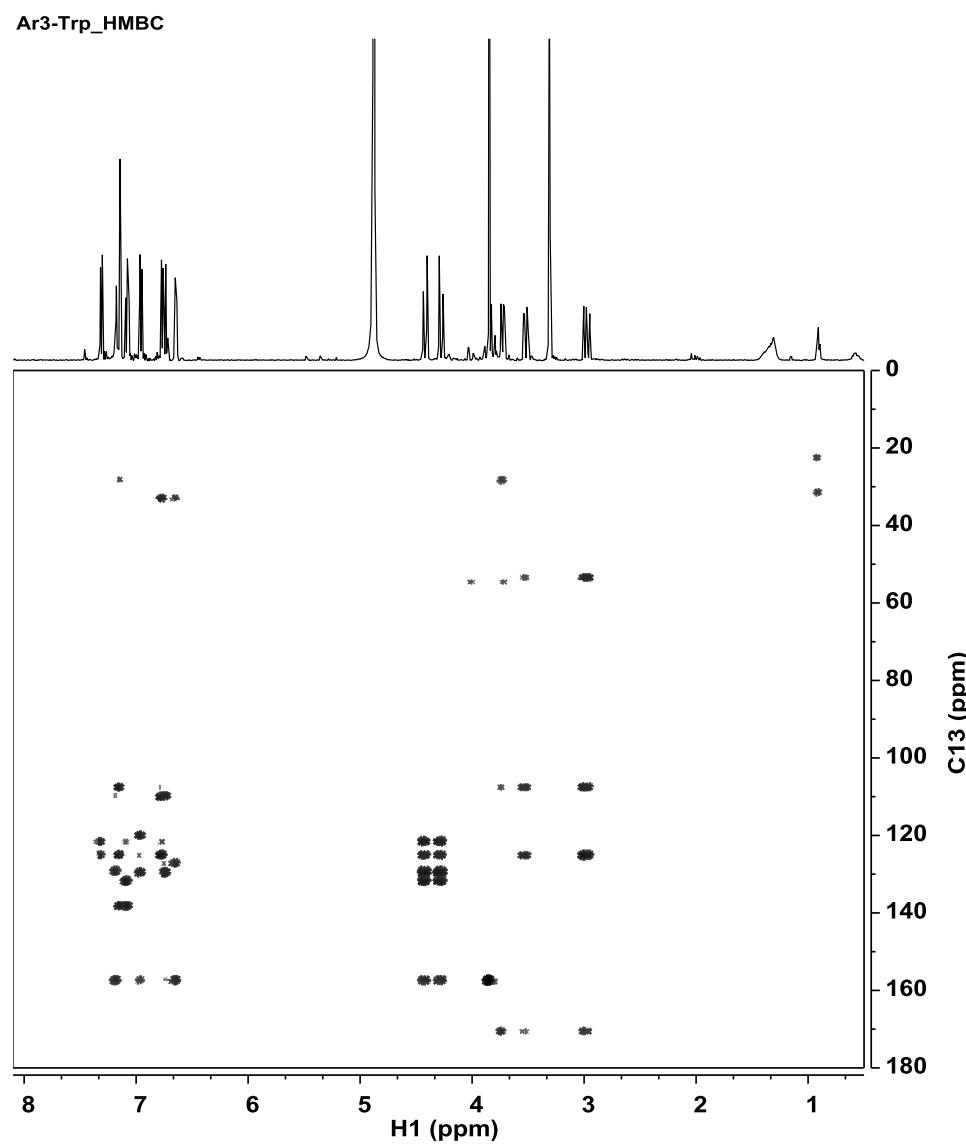
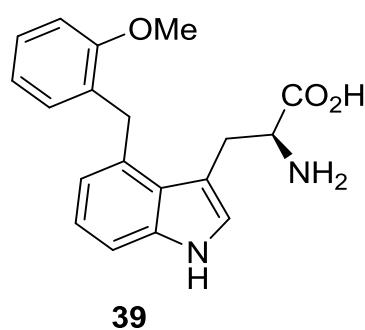
Ar3-Trp\_COSY



2D  $^1\text{H}$ - $^1\text{H}$  COSY NMR spectrum (500 MHz) of 39 in  $\text{MeOH-d}_4$



2D  $^1\text{H}$ - $^{13}\text{C}$  HSQC NMR spectrum (500 MHz) of **39** in MeOH-d4



2D  $^1\text{H}$ - $^{13}\text{C}$  HMBC NMR spectrum (500 MHz) of **39** in MeOH-d4

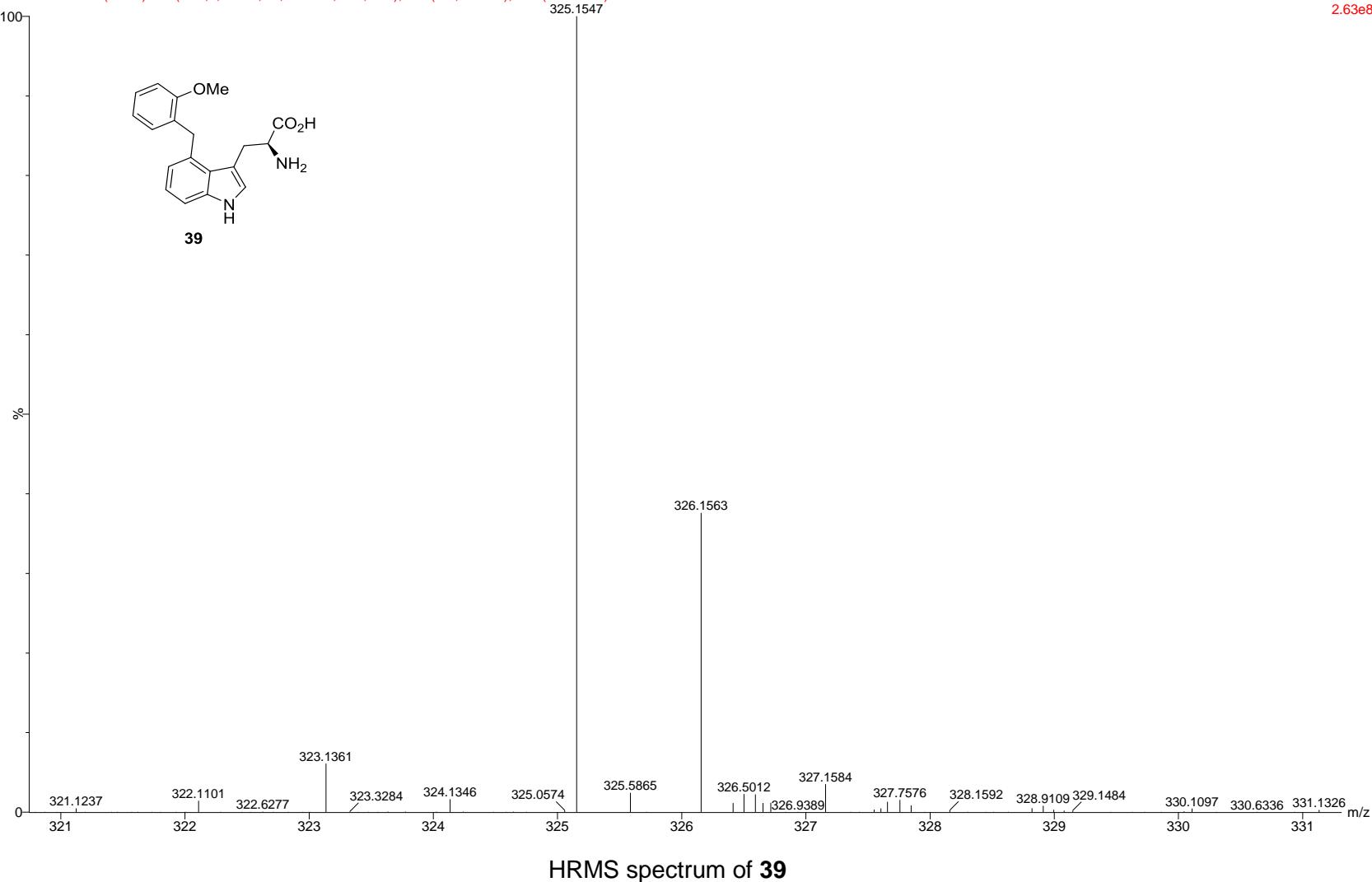
Trp-Ar3  
Bandari

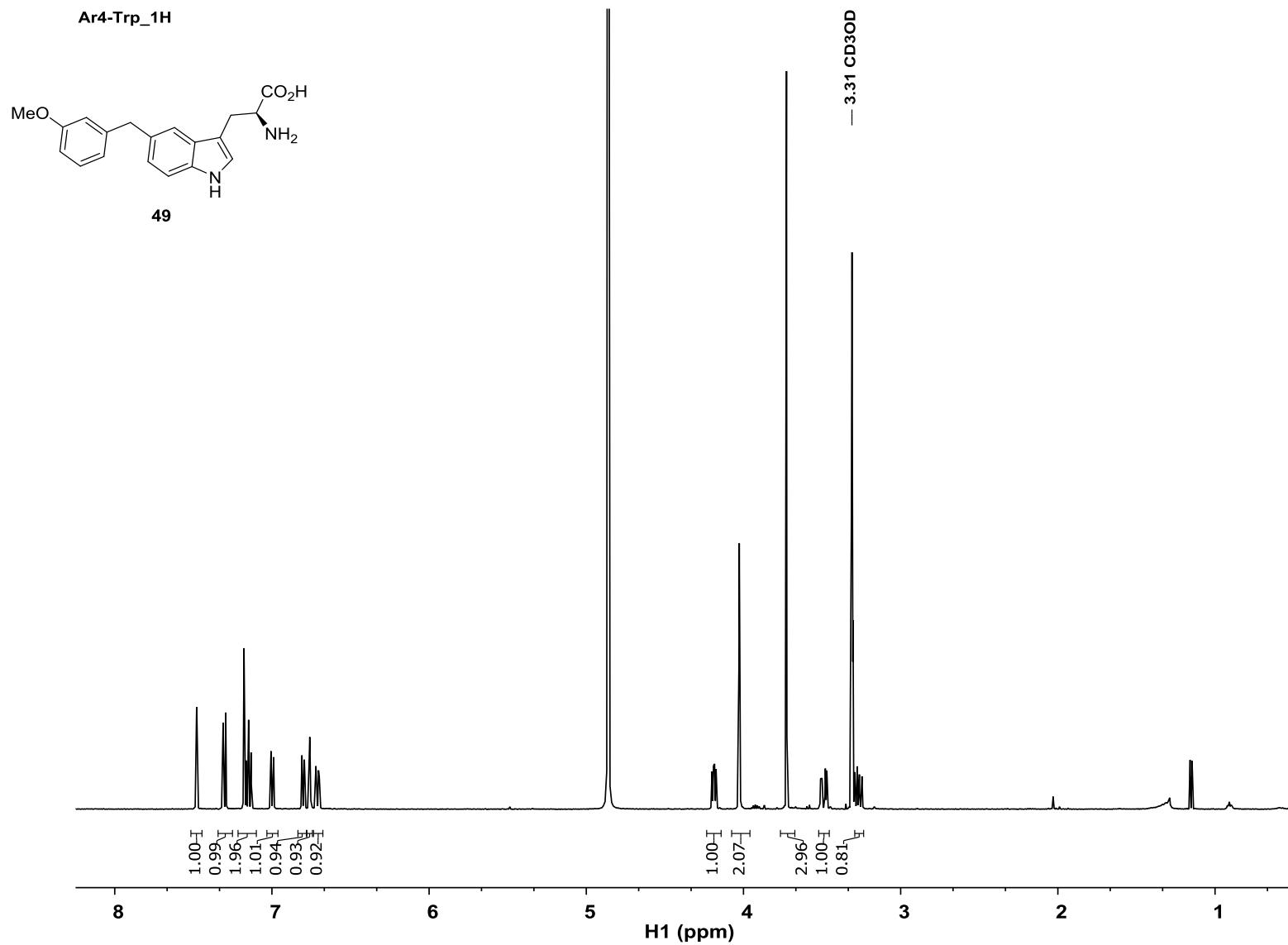
12055SS-004 24 (0.482) AM (Cen,4, 80.00, Ar,10000.0,0.00,0.00); Sm (SG, 1x5.00); Cm (19:29-2:9)

SYNAPTG2-Si#UGA589

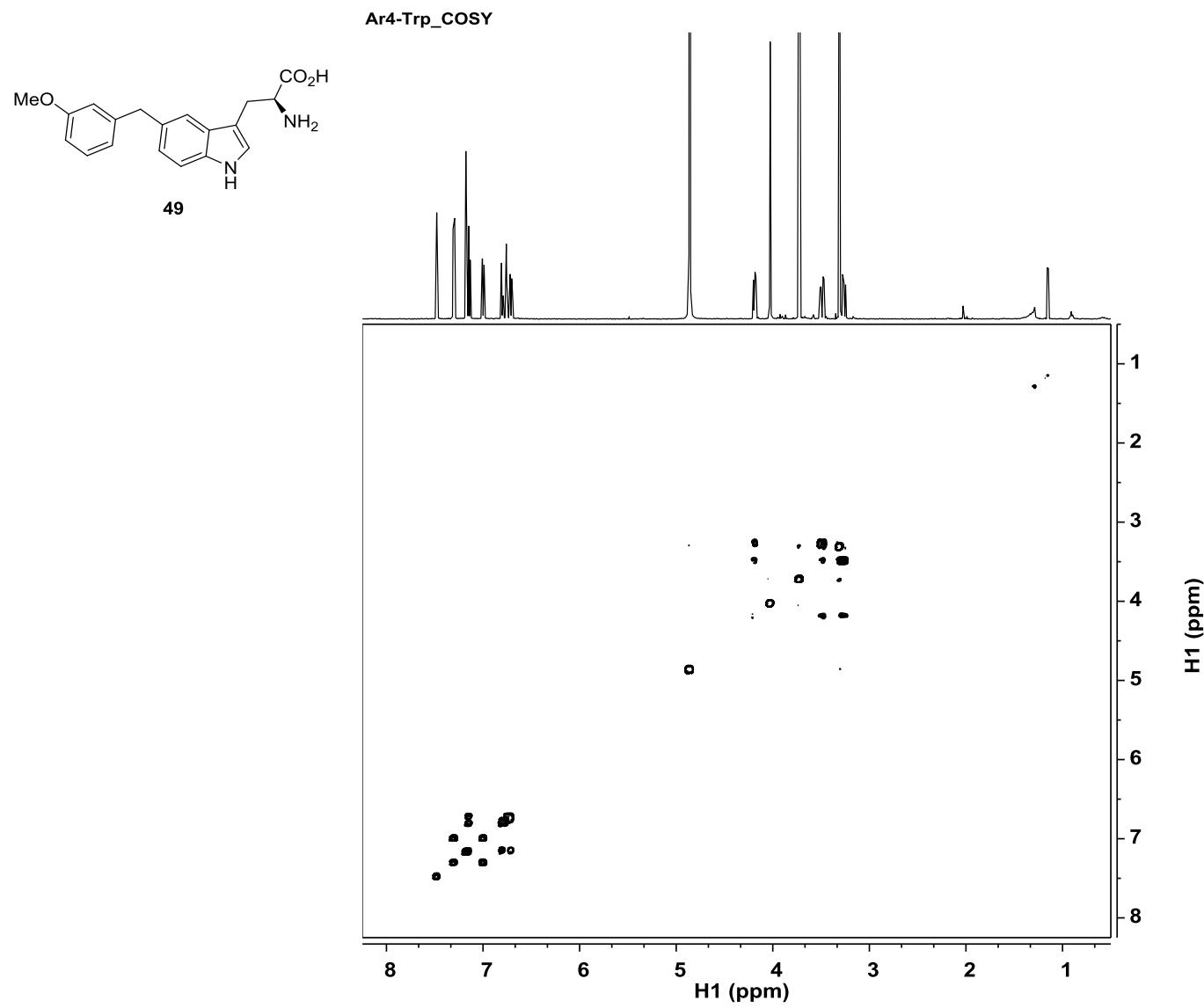
29-Aug-2017 16:04:56

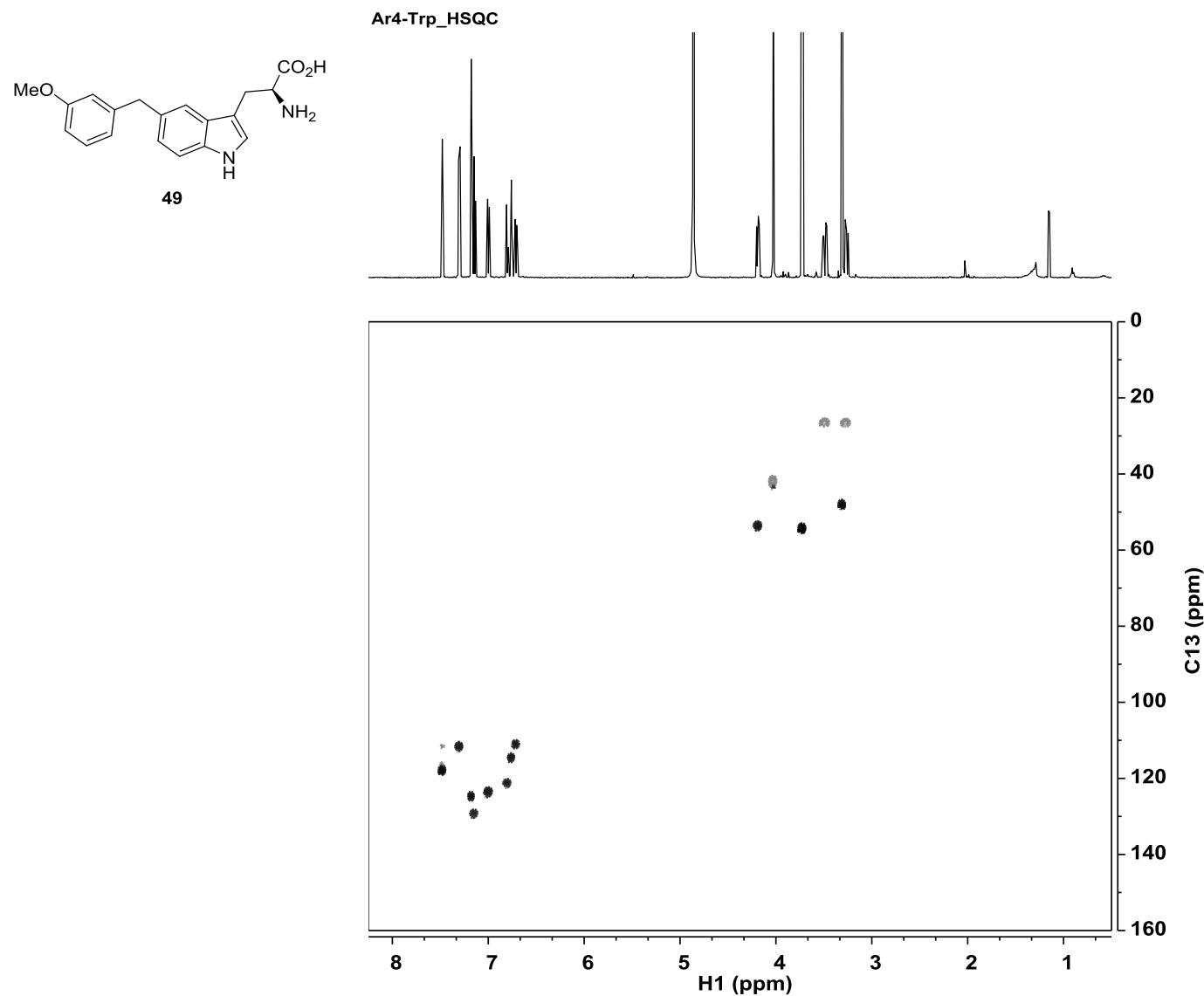
1: TOF MS ES+  
2.63e8



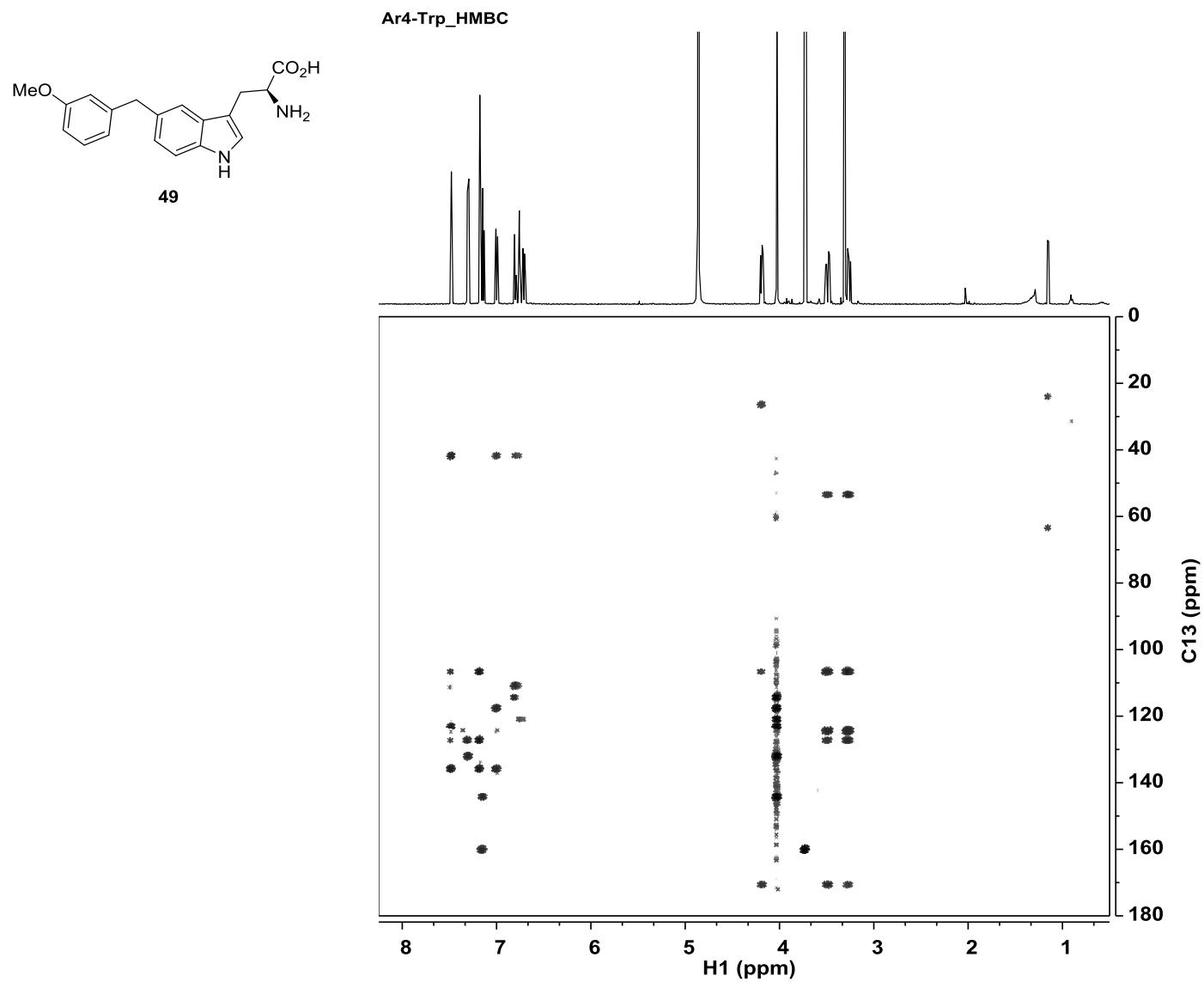


1D <sup>1</sup>H-NMR spectrum (500 MHz) of **49** in MeOH-d4





2D  $^1\text{H}$ - $^{13}\text{C}$  HSQC NMR spectrum (500 MHz) of **49** in MeOH-d4



2D  $^1\text{H}$ - $^{13}\text{C}$  HMBC NMR spectrum (500 MHz) of **49** in MeOH-d4

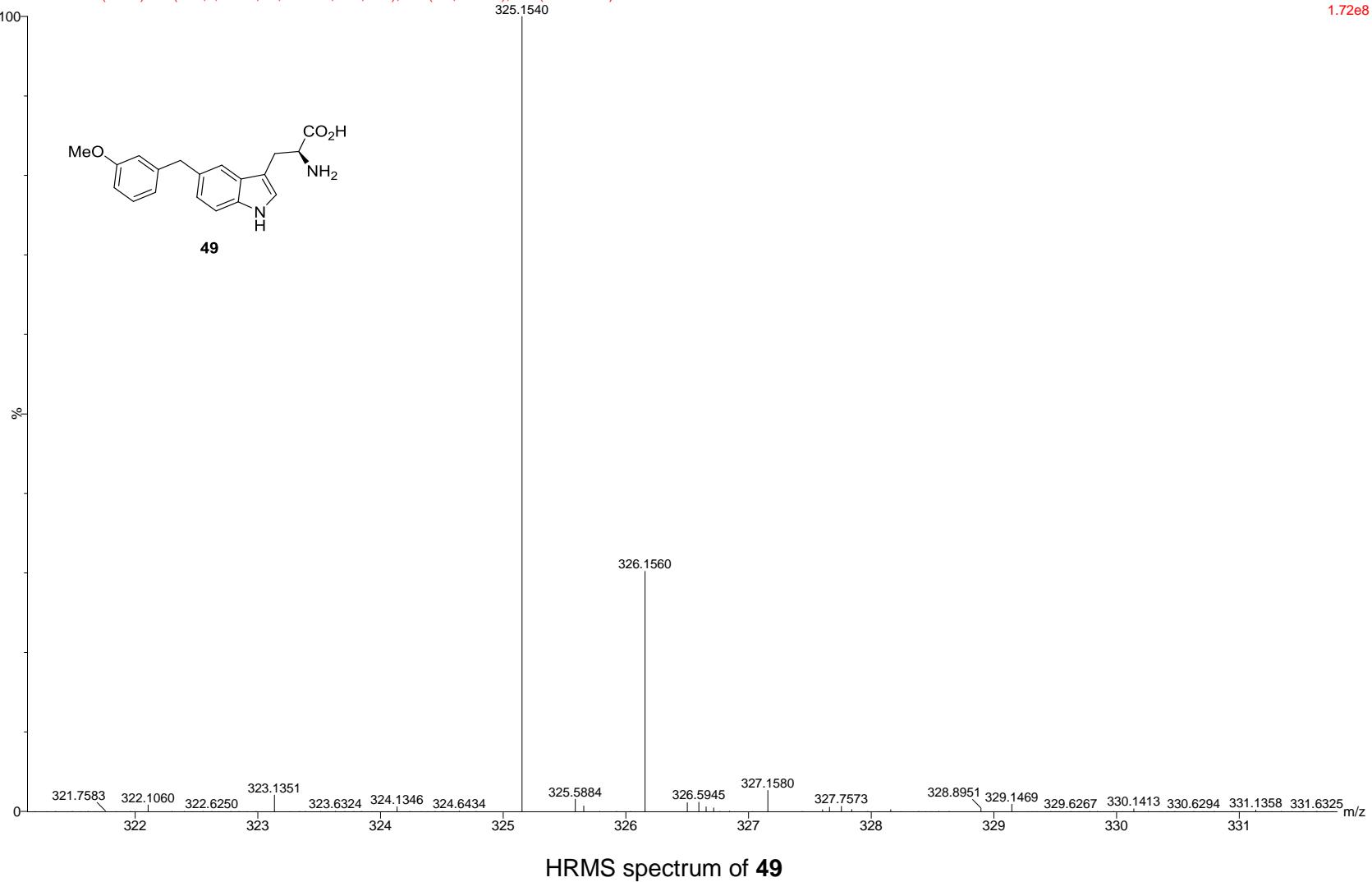
Trp-Ar4  
Bandari

12056SS-002 24 (0.482) AM (Cen,4, 80.00, Ar,10000.0,0.00,0.00); Sm (SG, 1x5.00); Crm (18:29-2:14)

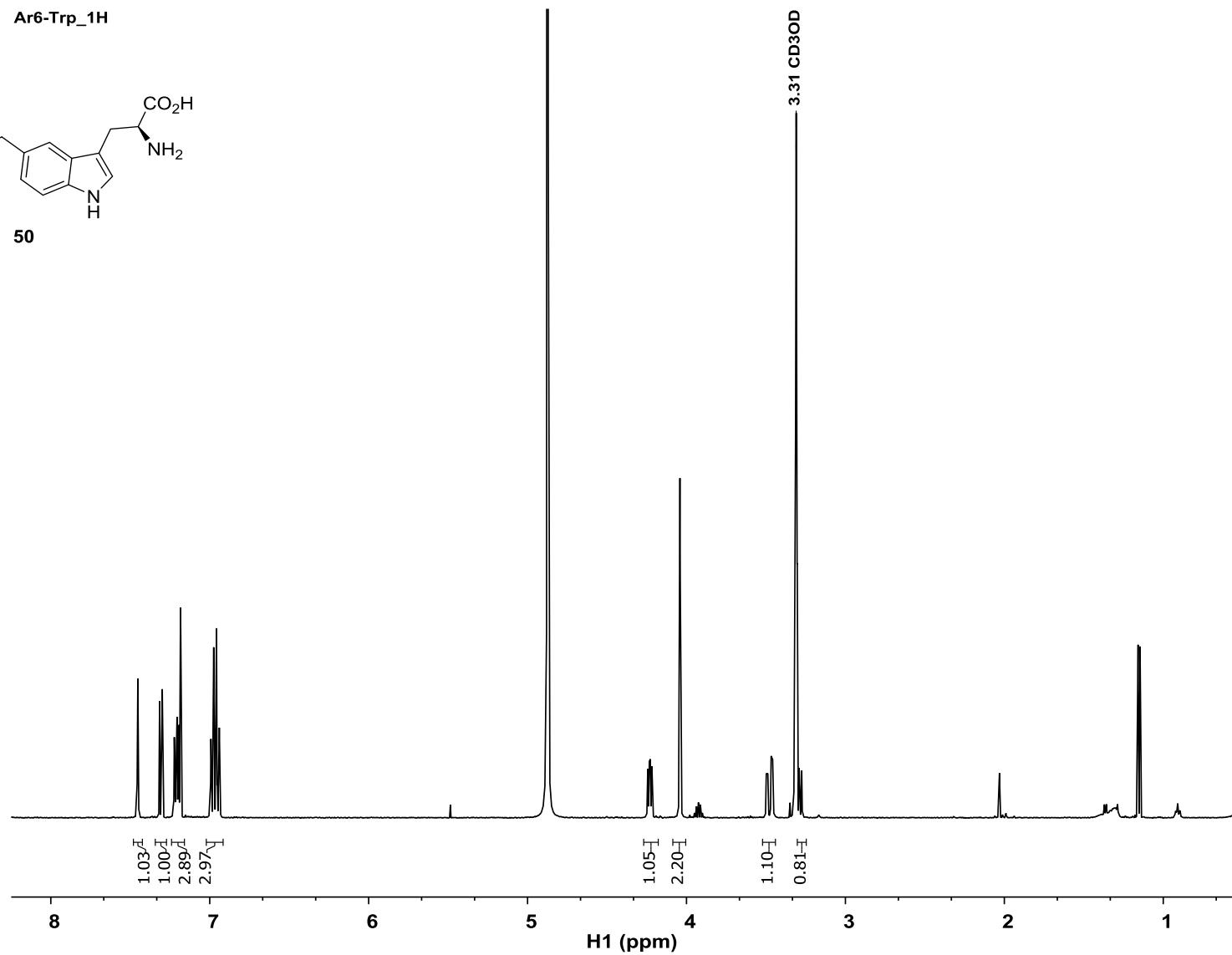
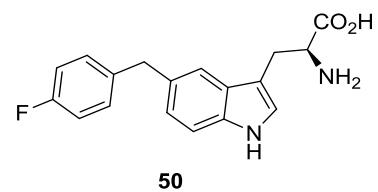
SYNAPTG2-Si#UGA589

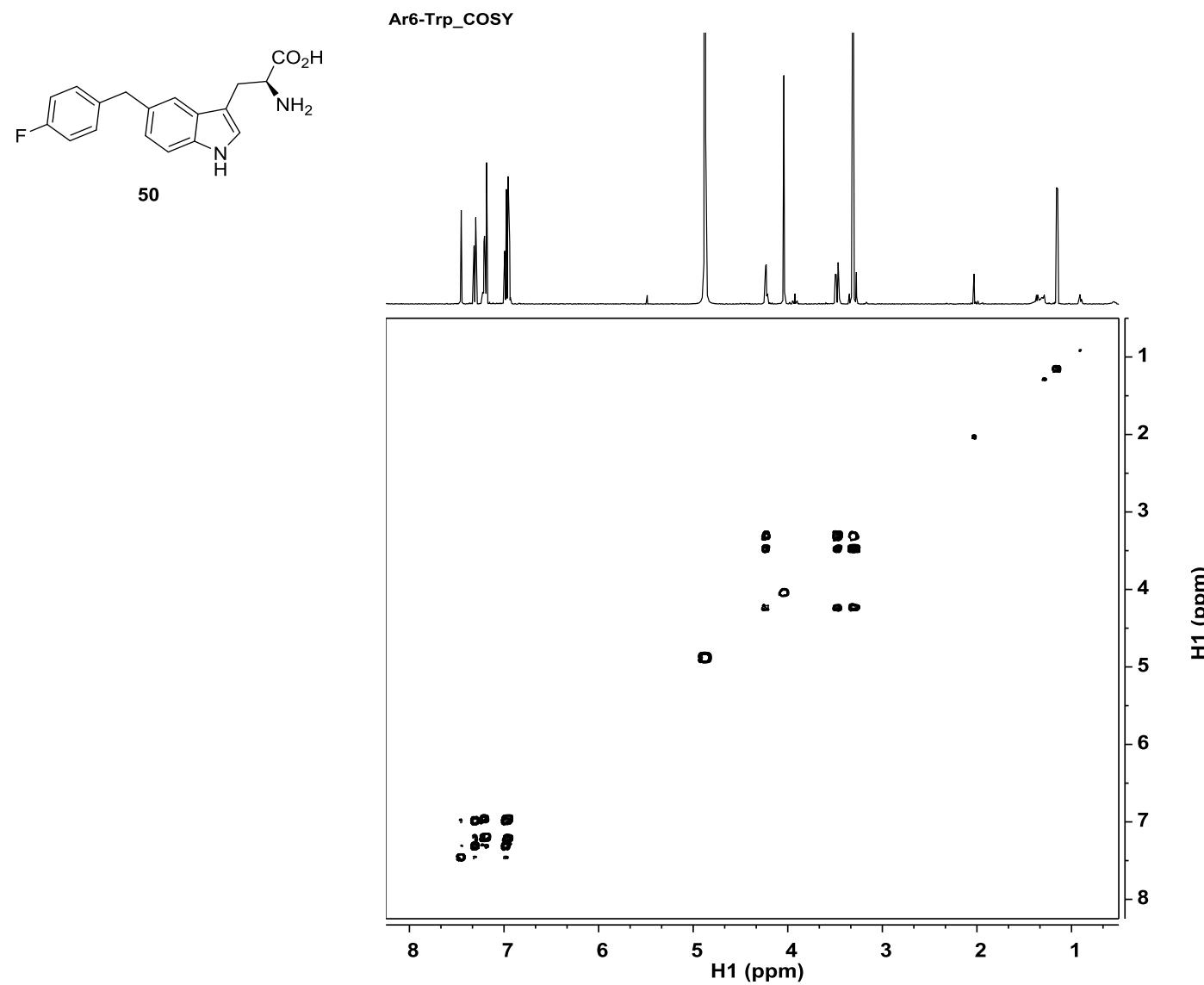
29-Aug-2017 16:16:41

1: TOF MS ES+  
1.72e8

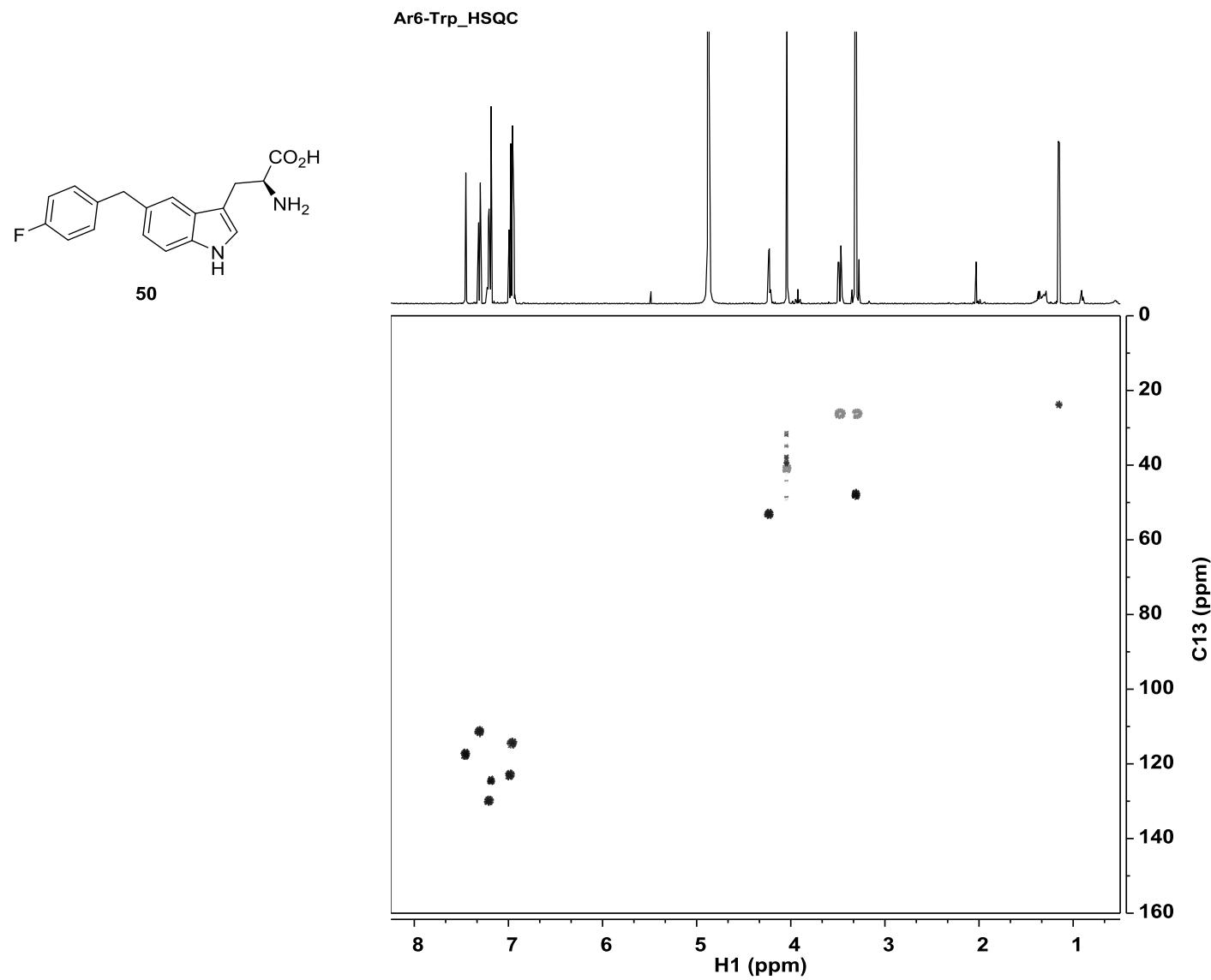


Ar6-Trp\_1H

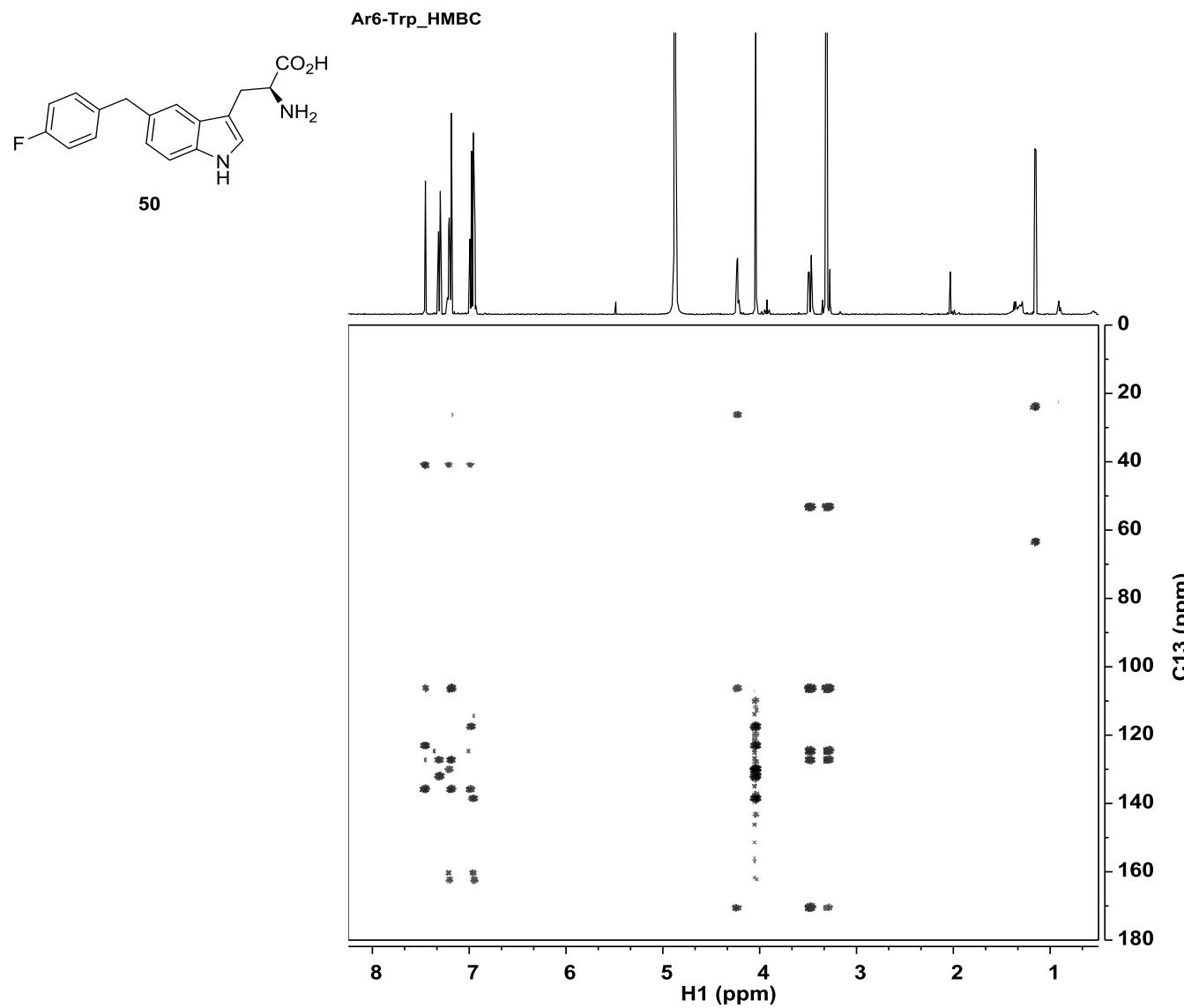




2D  $^1\text{H}$ - $^1\text{H}$  COSY NMR spectrum (500 MHz) of 50 in MeOH-d4



2D  $^1\text{H}$ - $^{13}\text{C}$  HSQC NMR spectrum (500 MHz) of **50** in MeOH-d4



2D  $^1\text{H}$ - $^{13}\text{C}$  HMBC NMR spectrum (500 MHz) of **50** in  $\text{MeOH-d}_4$

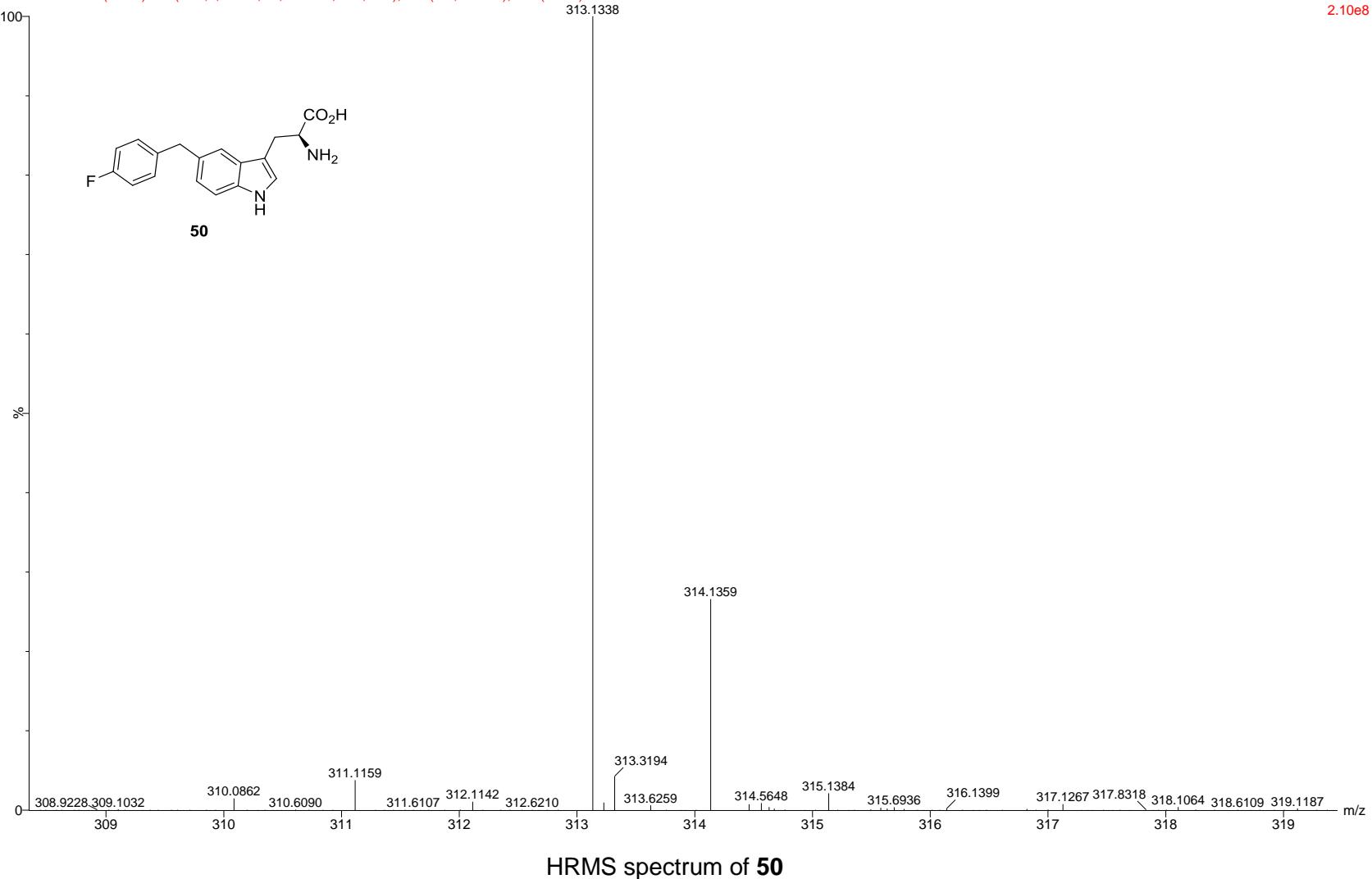
Trp-Ar6  
Bandari

12057SS-002 24 (0.482) AM (Cen,4, 80.00, Ar,10000.0,0.00,0.00); Sm (SG, 1x5.00); Cm (18:28)

SYNAPTG2-Si#UGA589

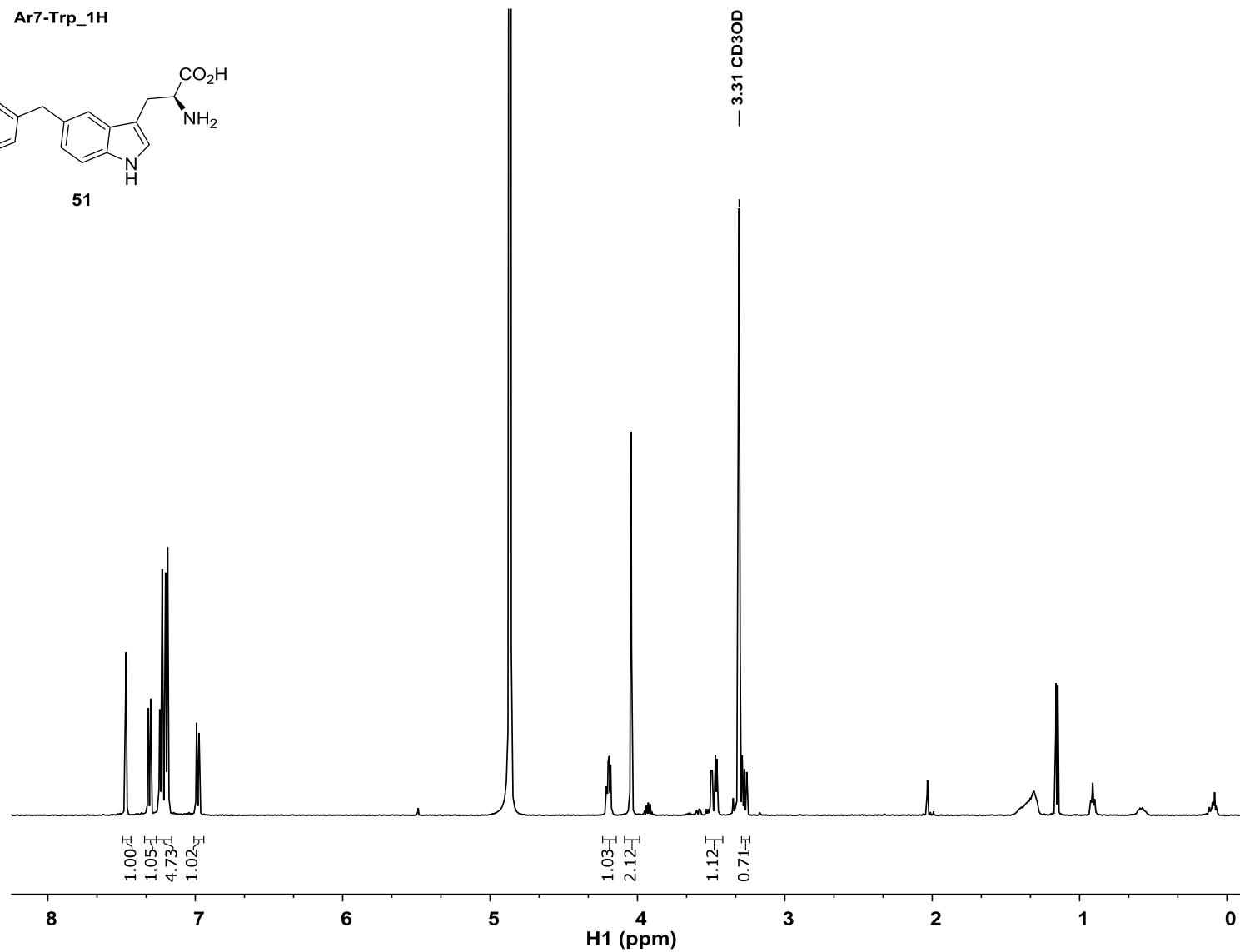
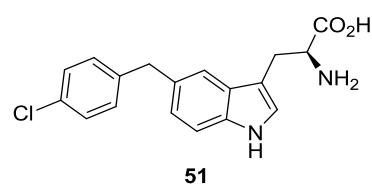
29-Aug-2017 16:31:41

1: TOF MS ES+  
2.10e8

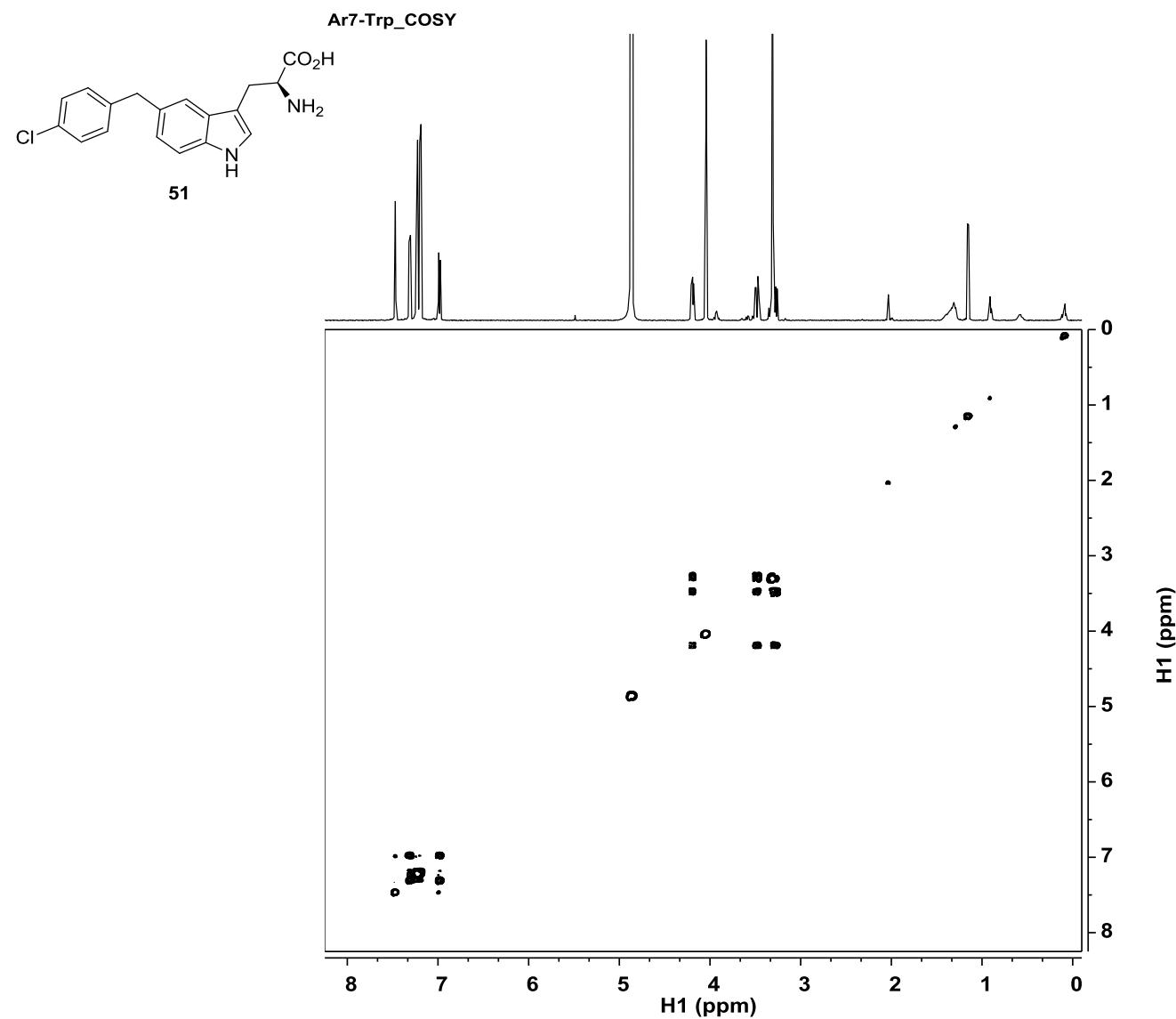


HRMS spectrum of **50**

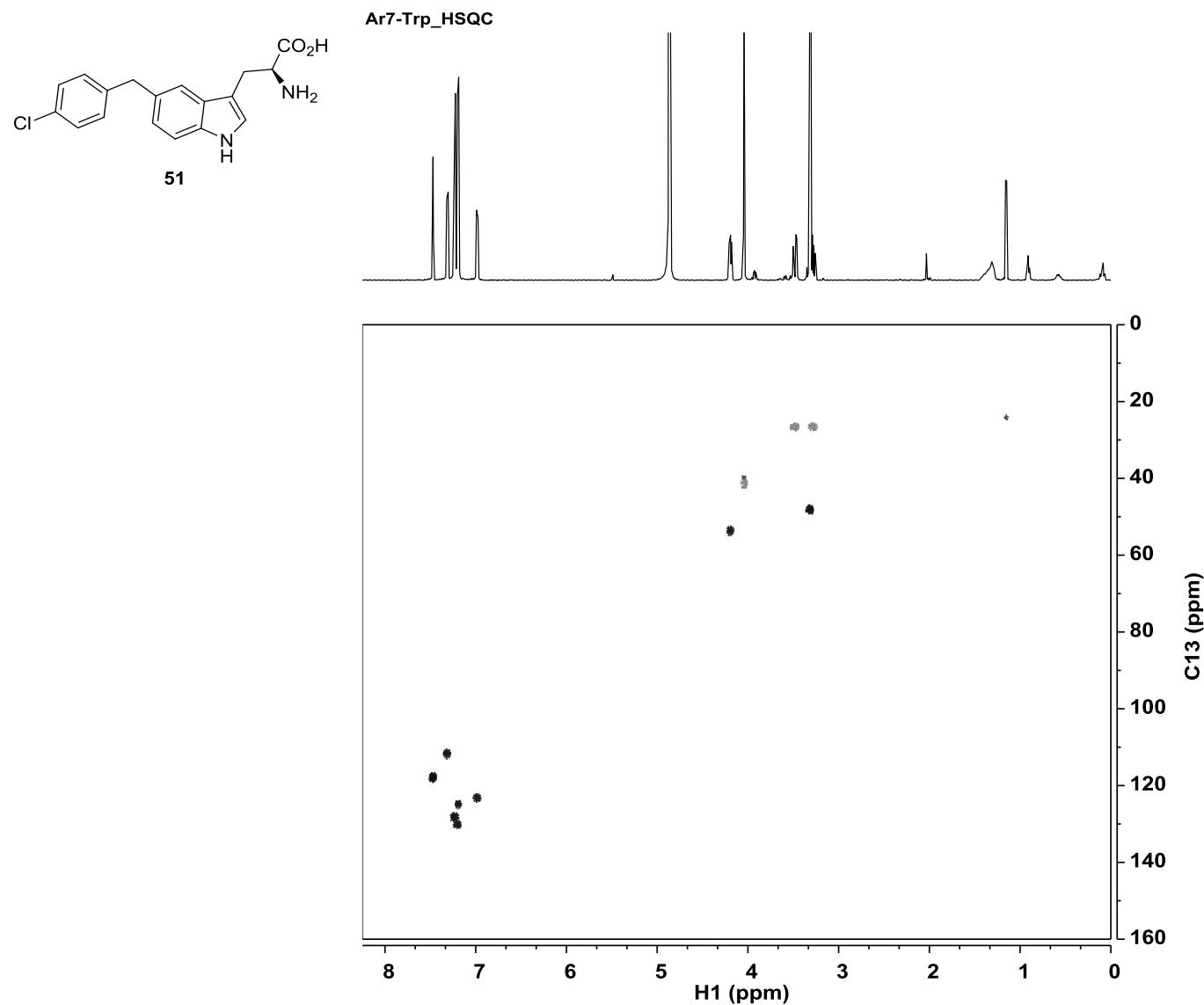
Ar7-Trp\_1H



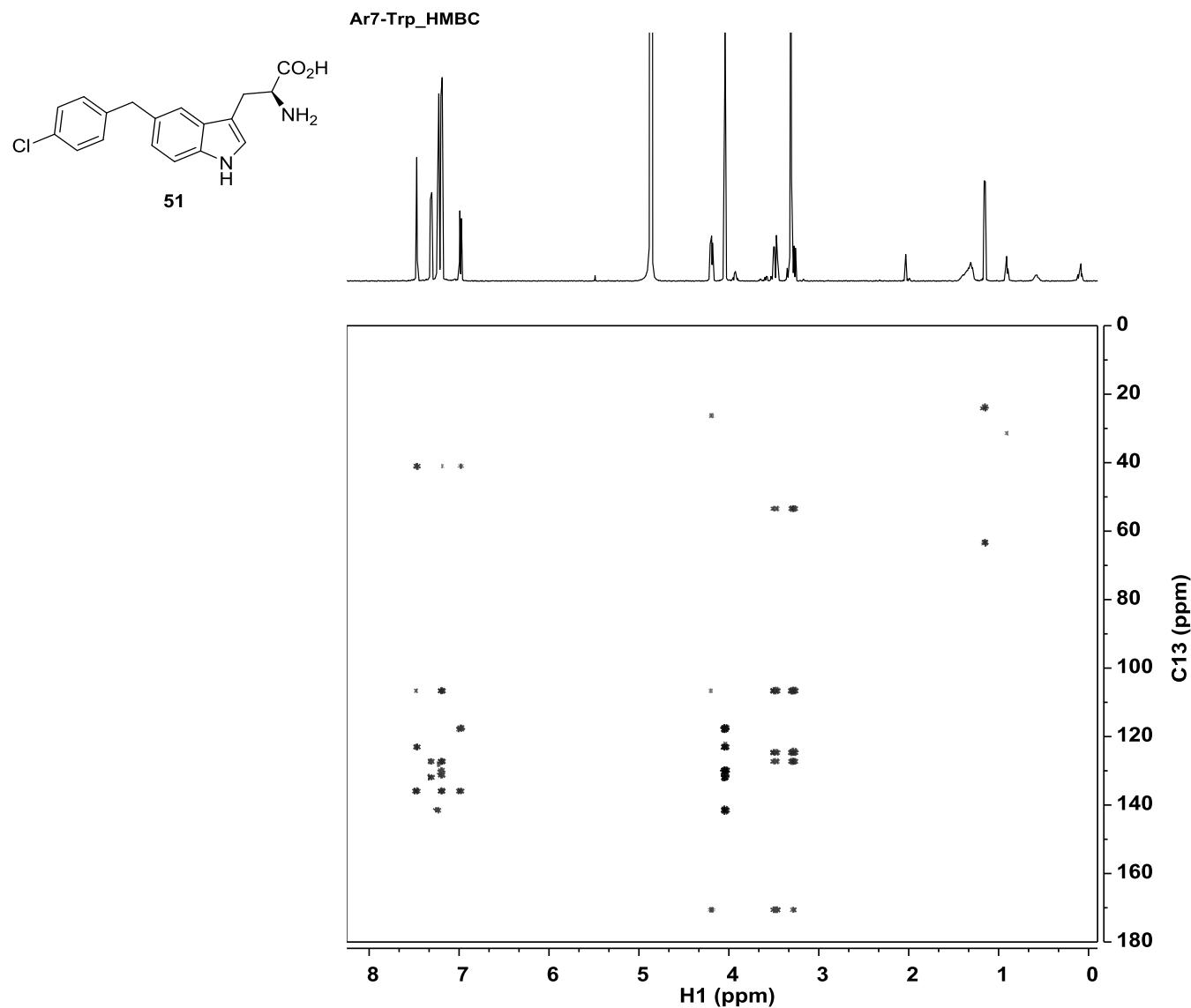
1D  $^1\text{H}$ -NMR spectrum (500 MHz) of **51** in  $\text{MeOH-d}_4$



2D  $^1\text{H}$ - $^1\text{H}$  COSY NMR spectrum (500 MHz) of **51** in  $\text{MeOH-d}_4$



2D  $^1\text{H}$ - $^{13}\text{C}$  HSQC NMR spectrum (500 MHz) of **51** in MeOH-d4



2D  $^1\text{H}$ - $^{13}\text{C}$  HMBC NMR spectrum (500 MHz) of **51** in MeOH-d4

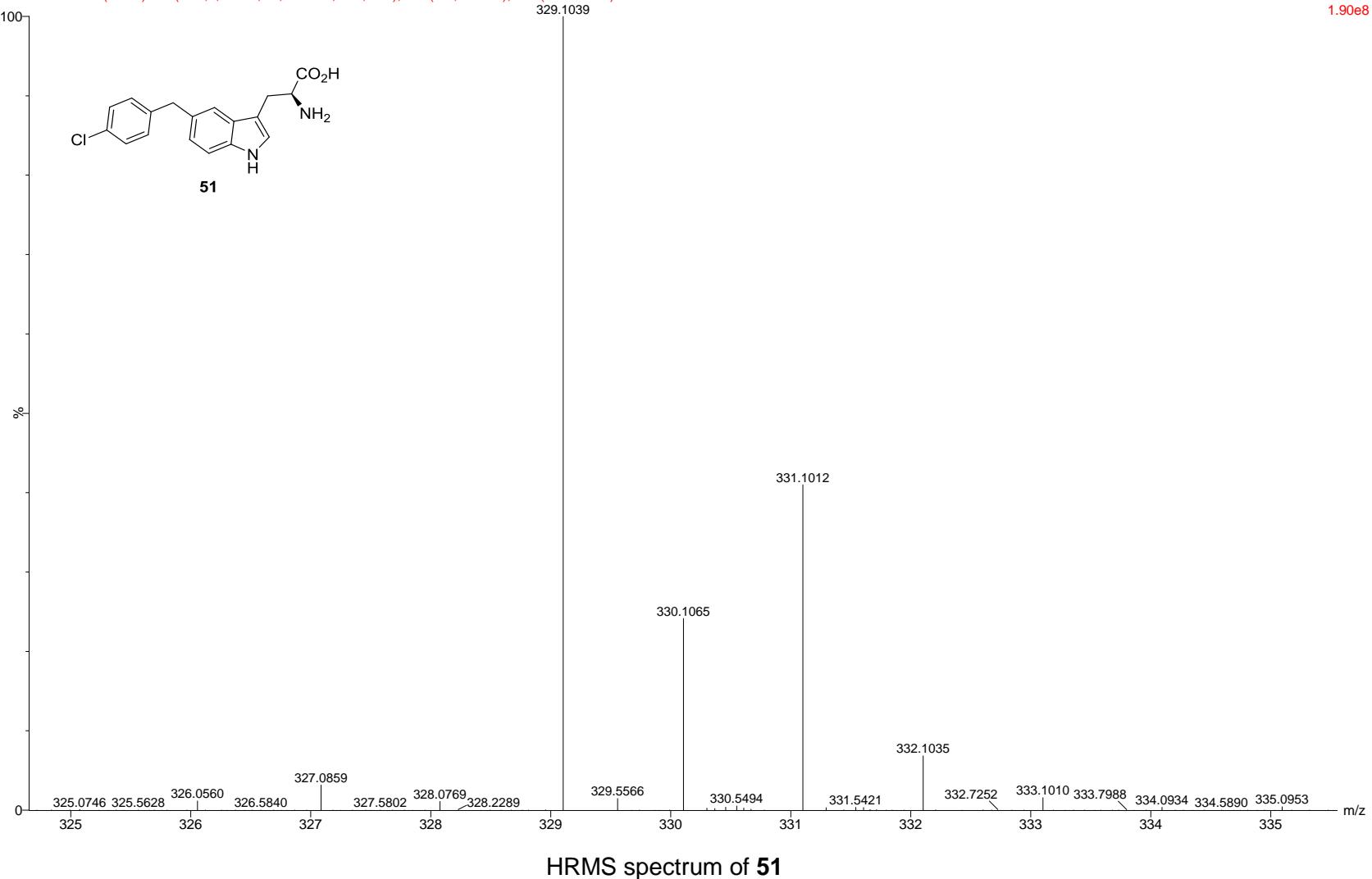
Trp-Ar7  
Bandari

12058SS-002 24 (0.482) AM (Cen,4, 80.00, Ar,10000.0,0.00,0.00); Sm (SG, 1x5.00); Cm (18:30-3:14)

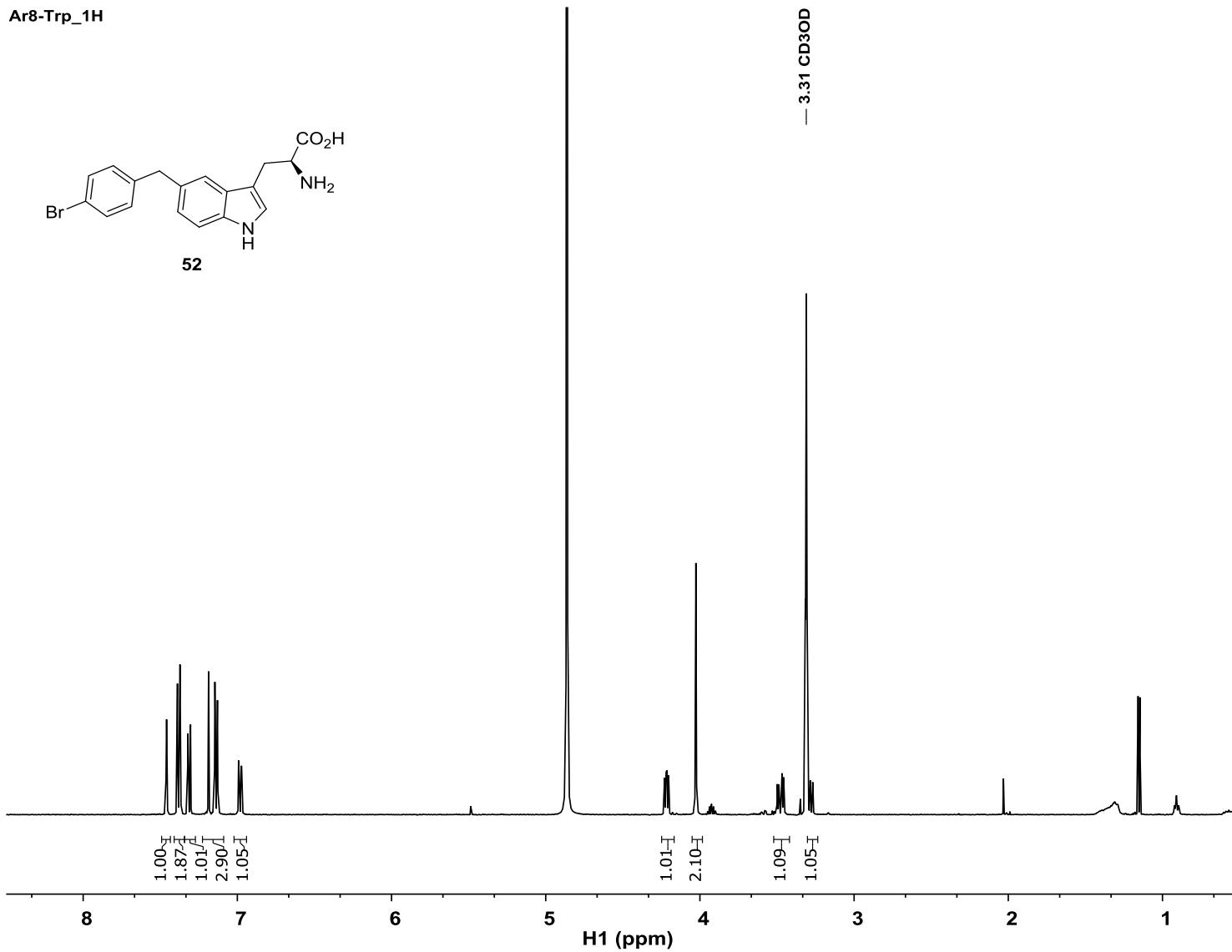
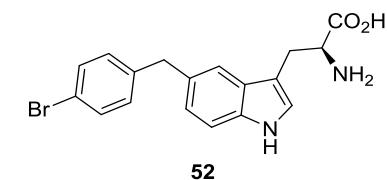
SYNAPTG2-Si#UGA589

29-Aug-2017 16:41:25

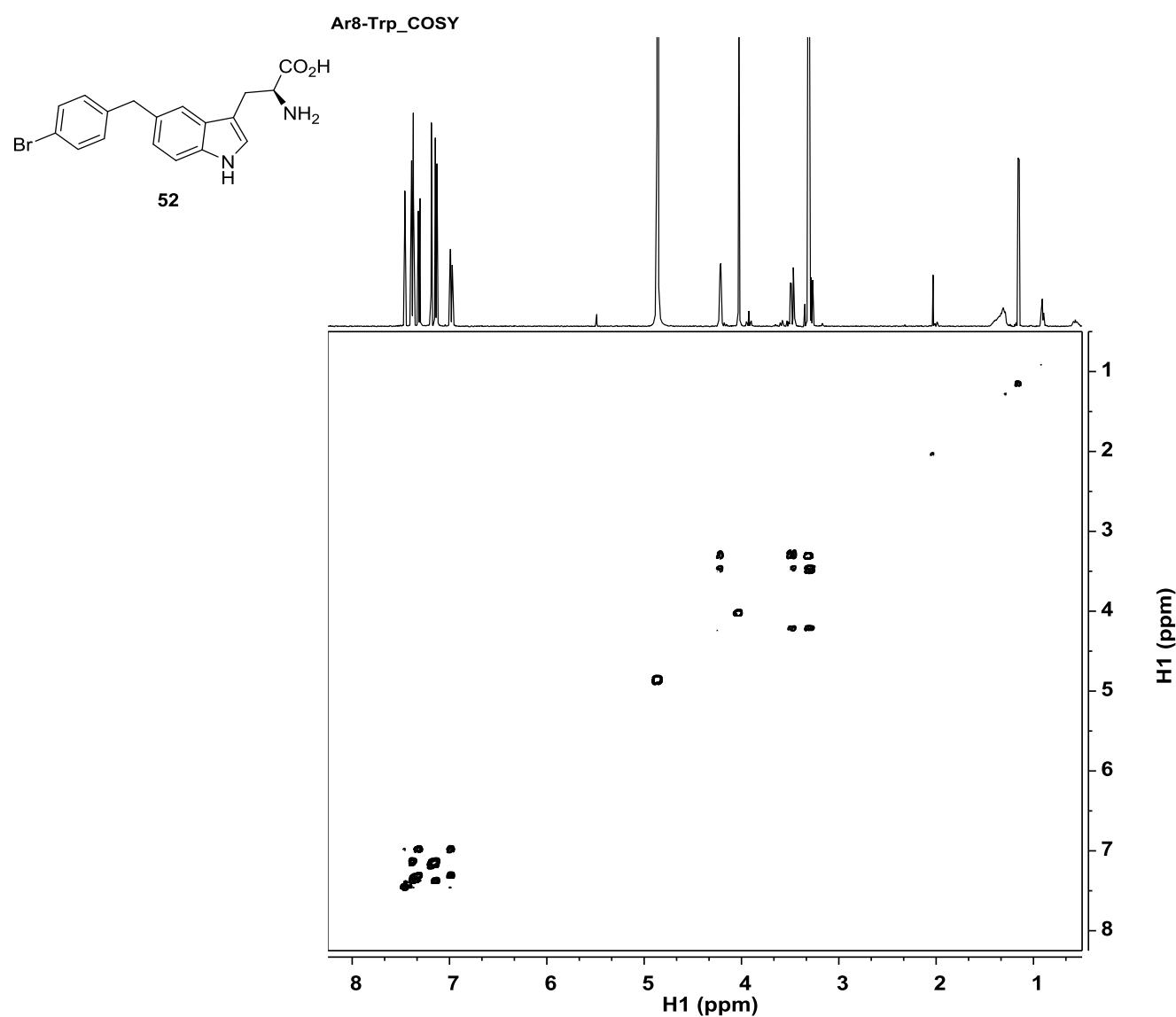
1: TOF MS ES+  
1.90e8



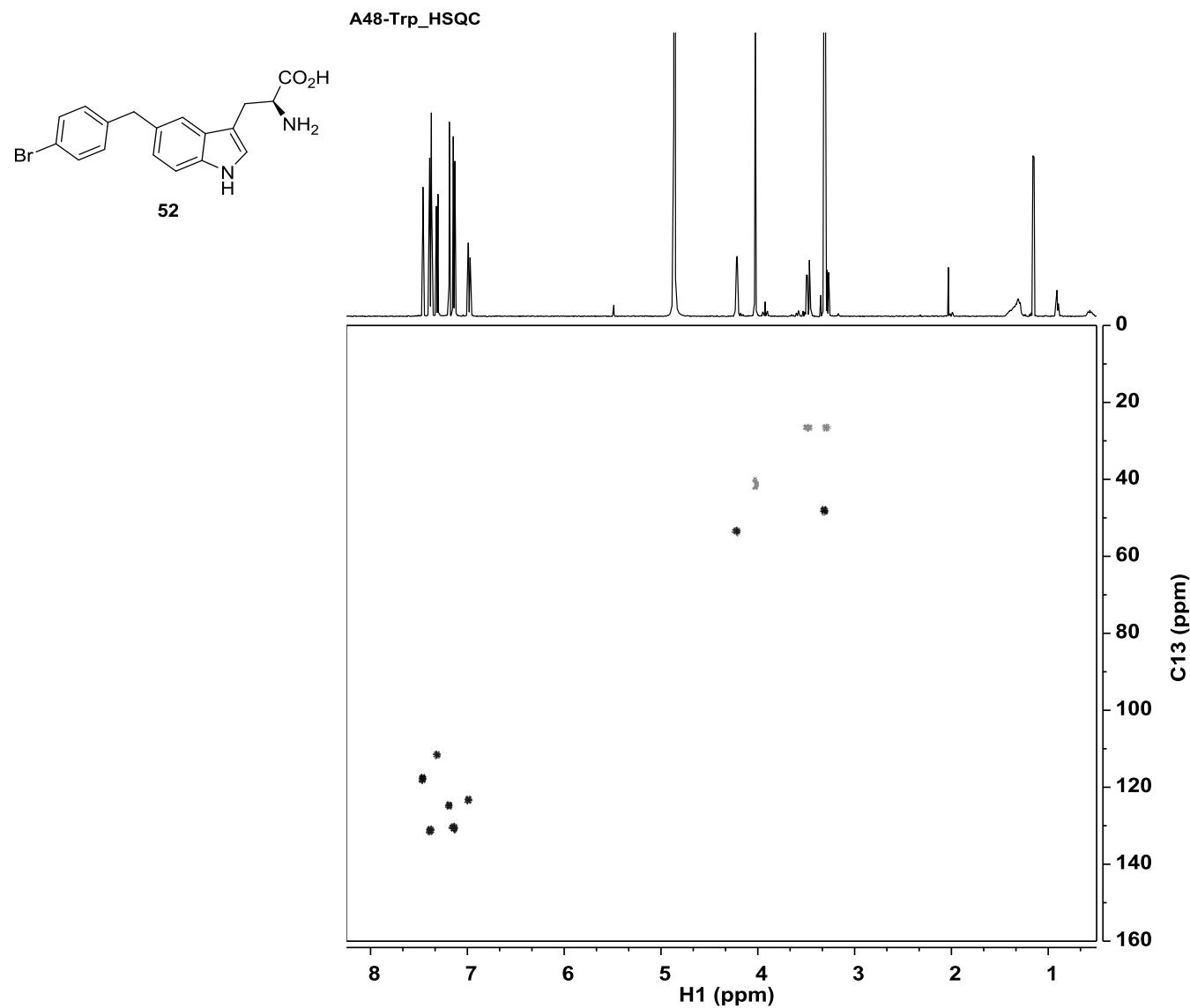
Ar8-Trp\_1H



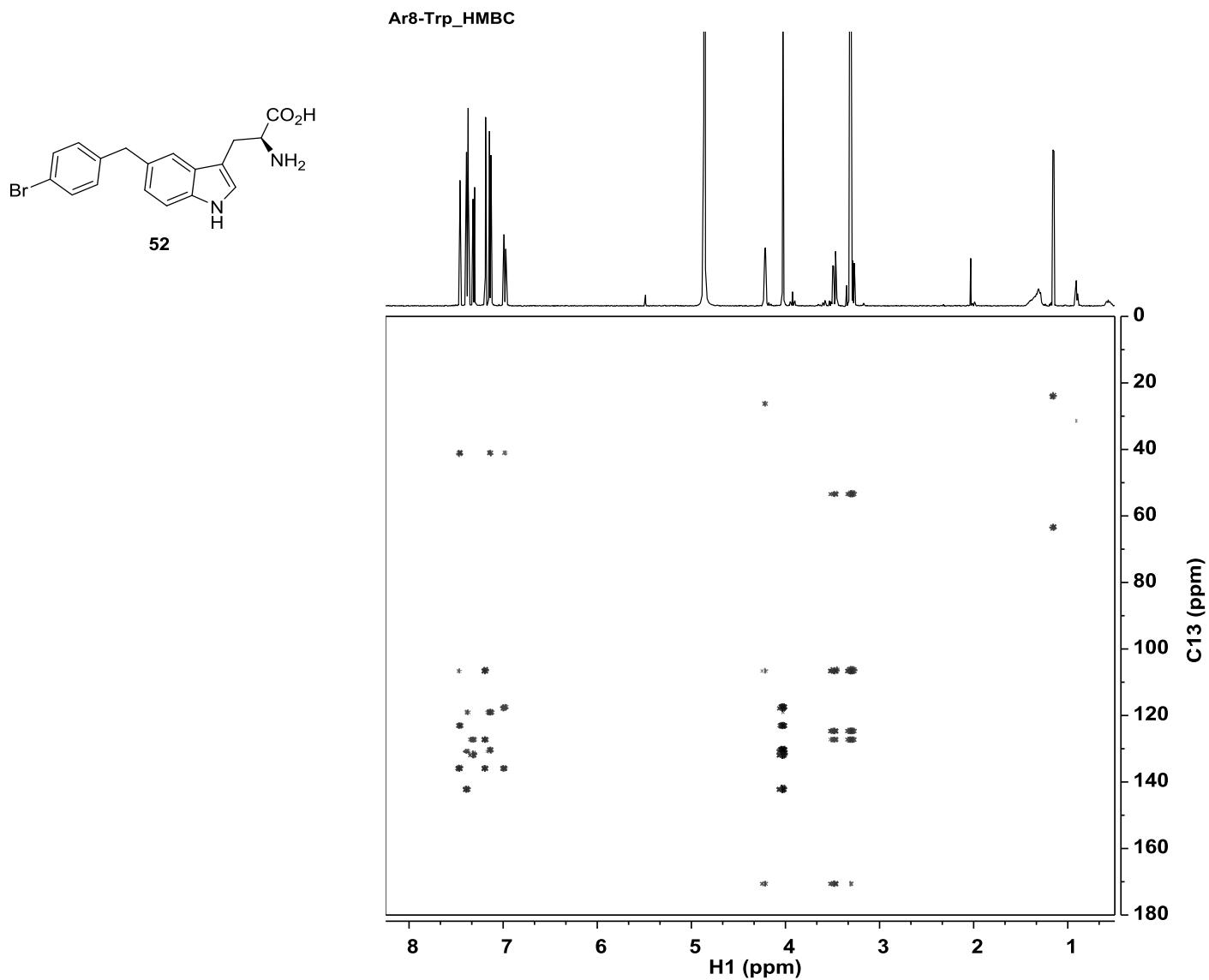
1D <sup>1</sup>H-NMR spectrum (500 MHz) of **52** in MeOH-d4



2D  $^1\text{H}$ - $^1\text{H}$  COSY NMR spectrum (500 MHz) of **52** in MeOH-d4



2D  $^1\text{H}$ - $^{13}\text{C}$  HSQC NMR spectrum (500 MHz) of **52** in  $\text{MeOH-d}_4$



2D  $^1\text{H}$ - $^{13}\text{C}$  HMBC NMR spectrum (500 MHz) of **52** in MeOH-d4

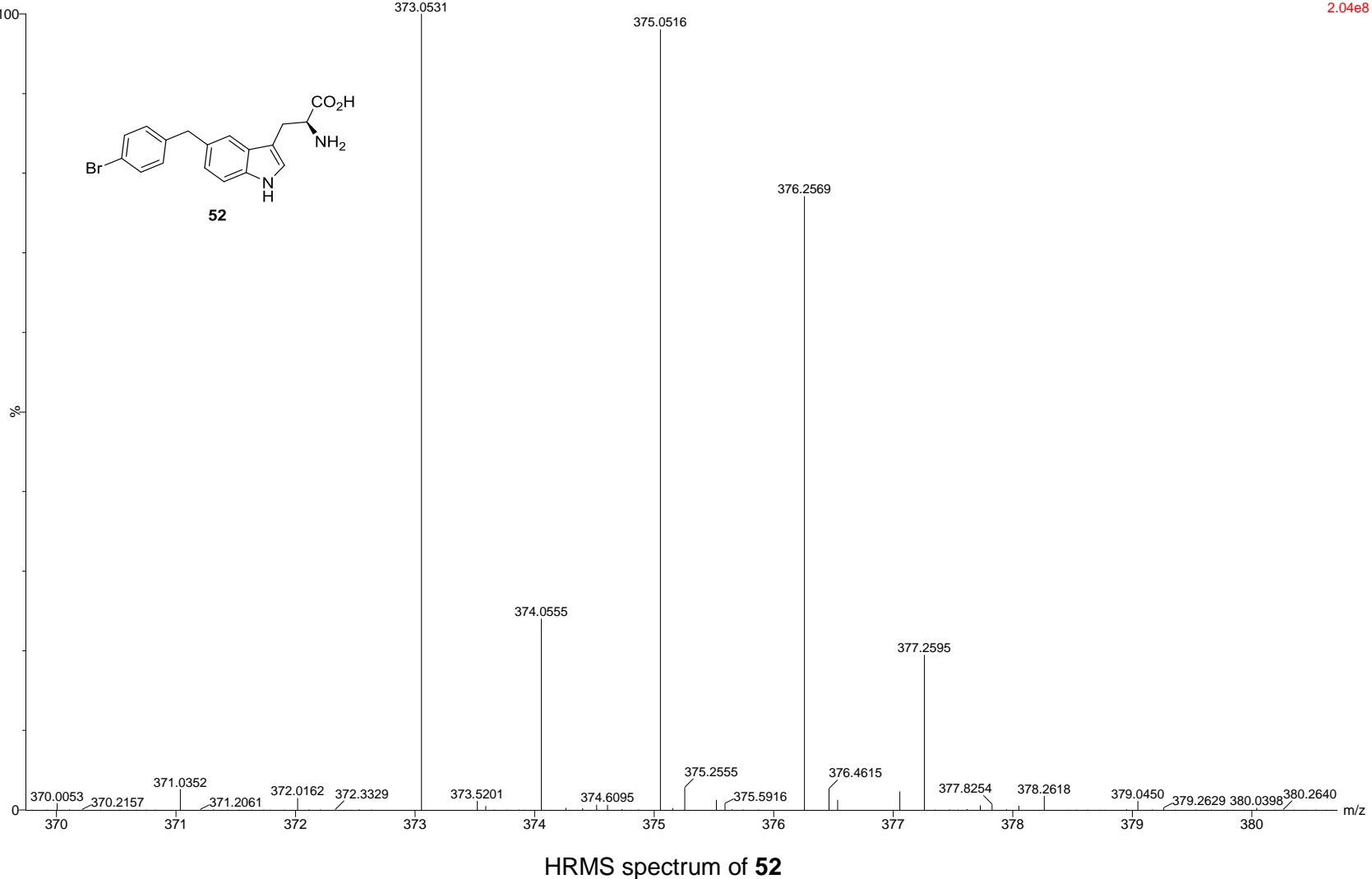
Trp-Ar8  
Bandari

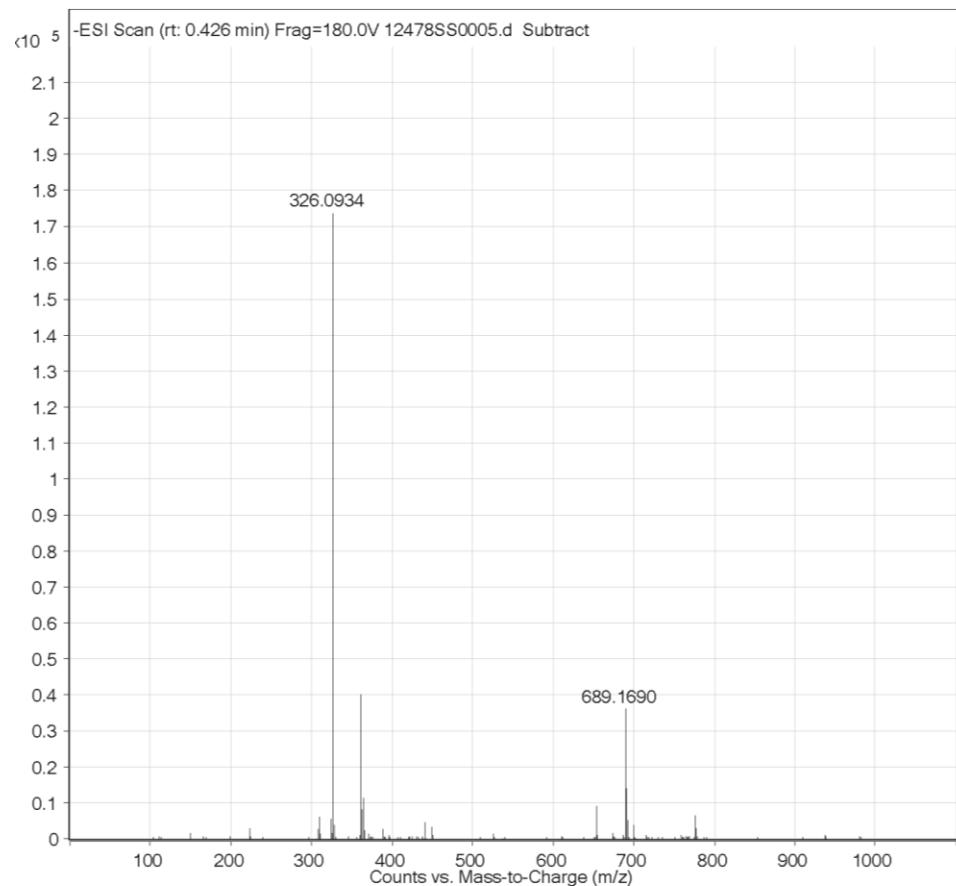
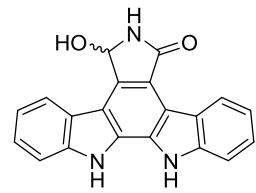
12059SS-002 23 (0.465) AM (Cen,4, 80.00, Ar,10000.0,0.00,0.00); Sm (SG, 1x5.00); Cm (19:31)

SYNAPTG2-Si#UGA589

29-Aug-2017 16:51:03

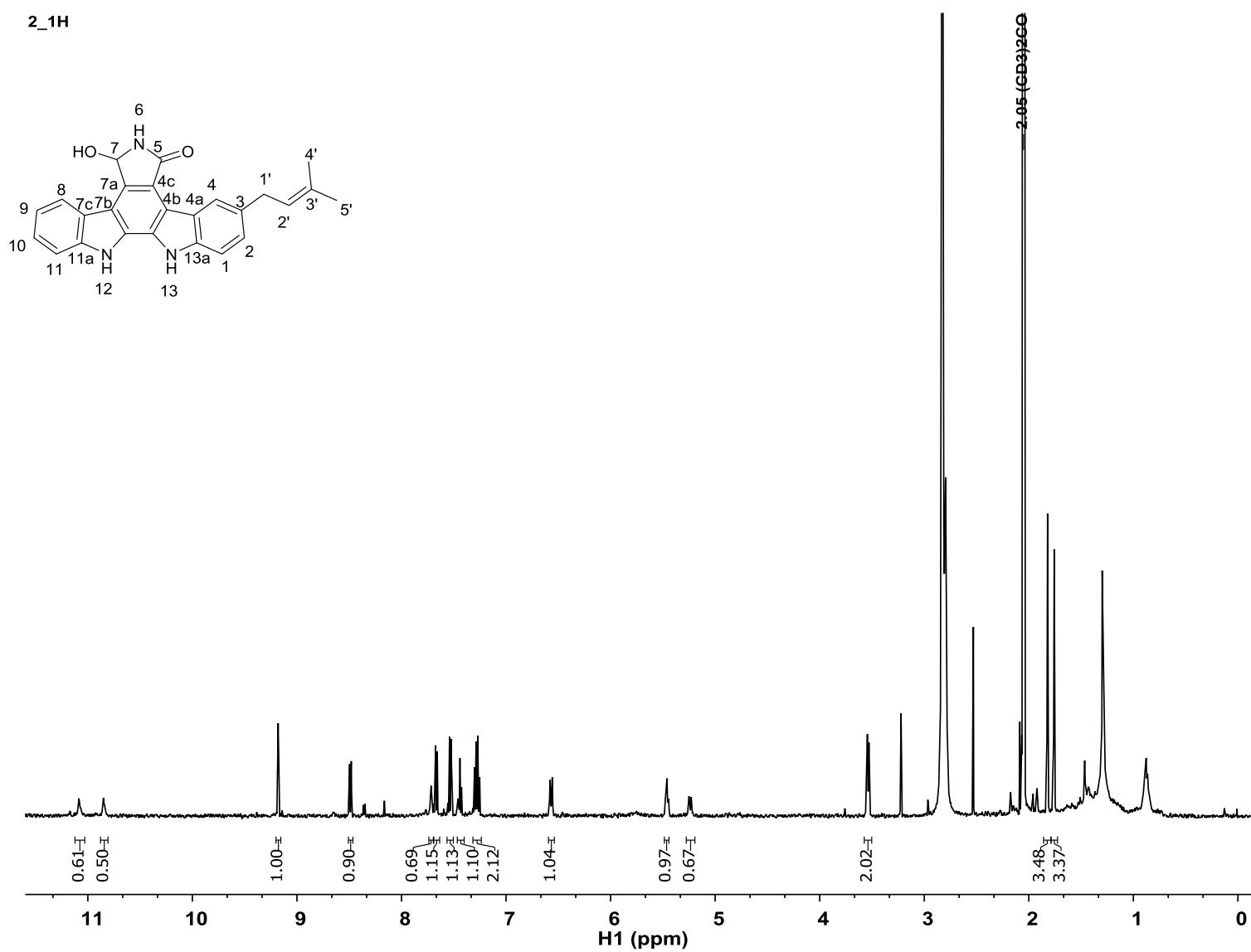
1: TOF MS ES+  
2.04e8



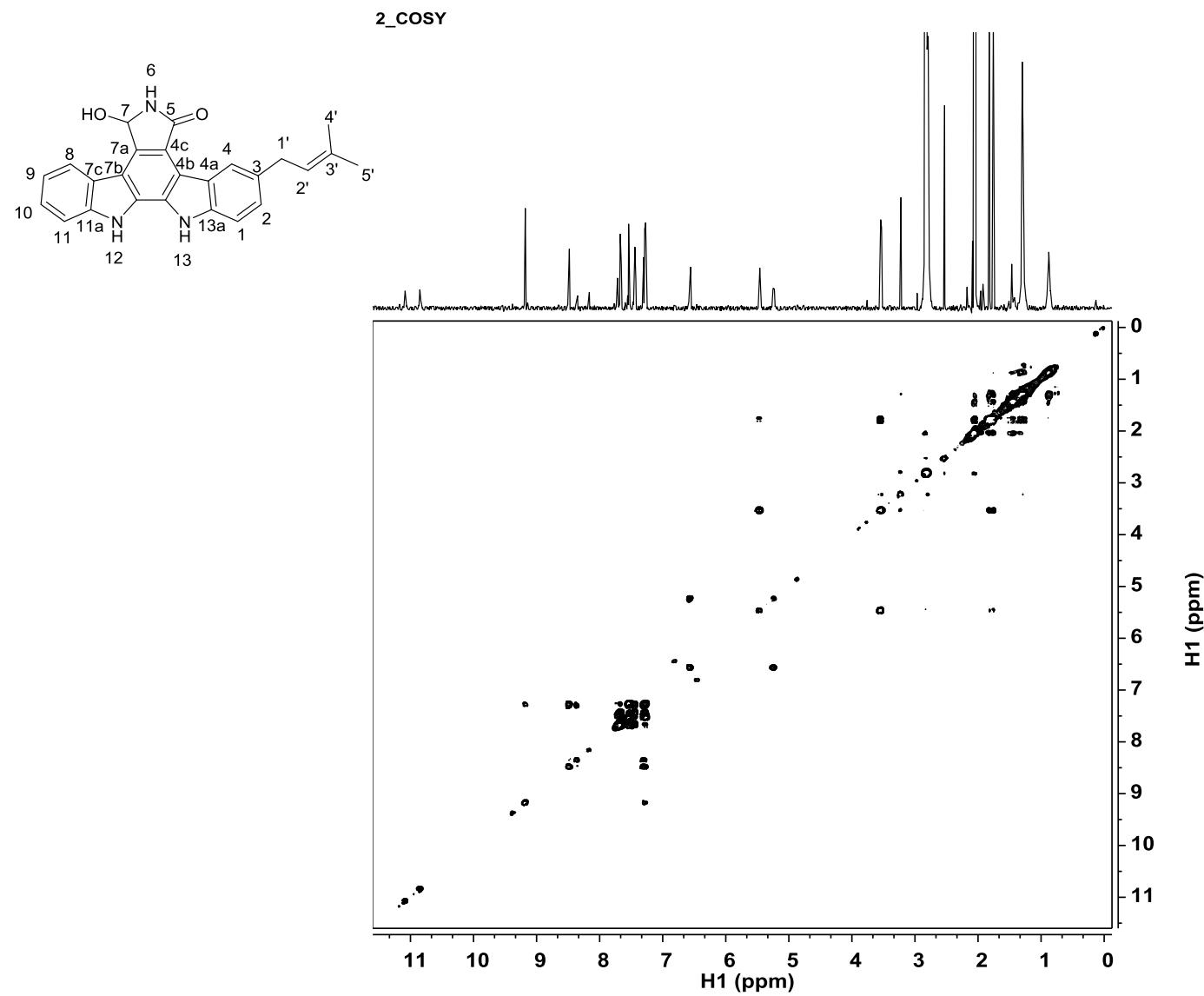


HRMS spectrum of **56**

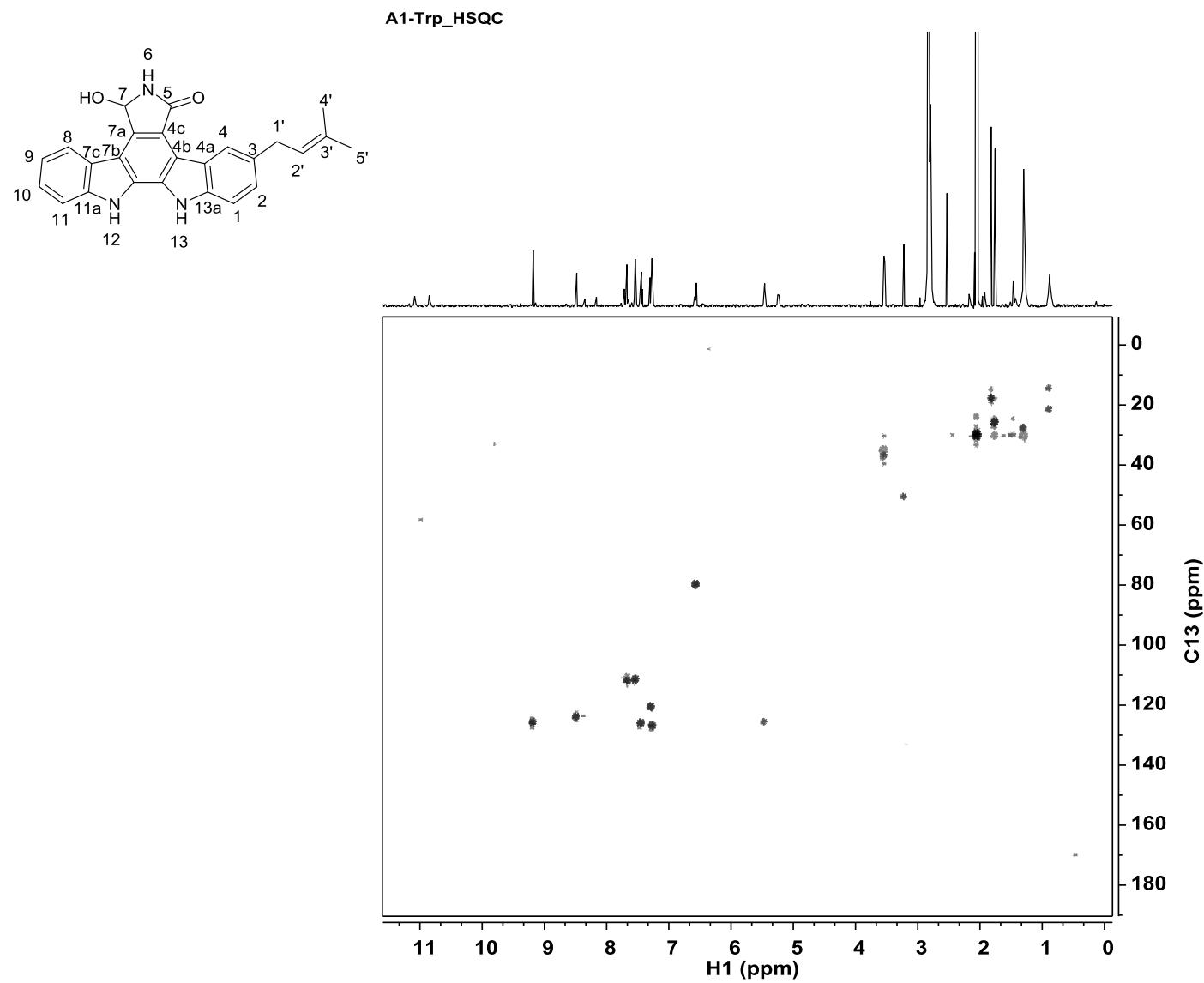
**2\_1H**



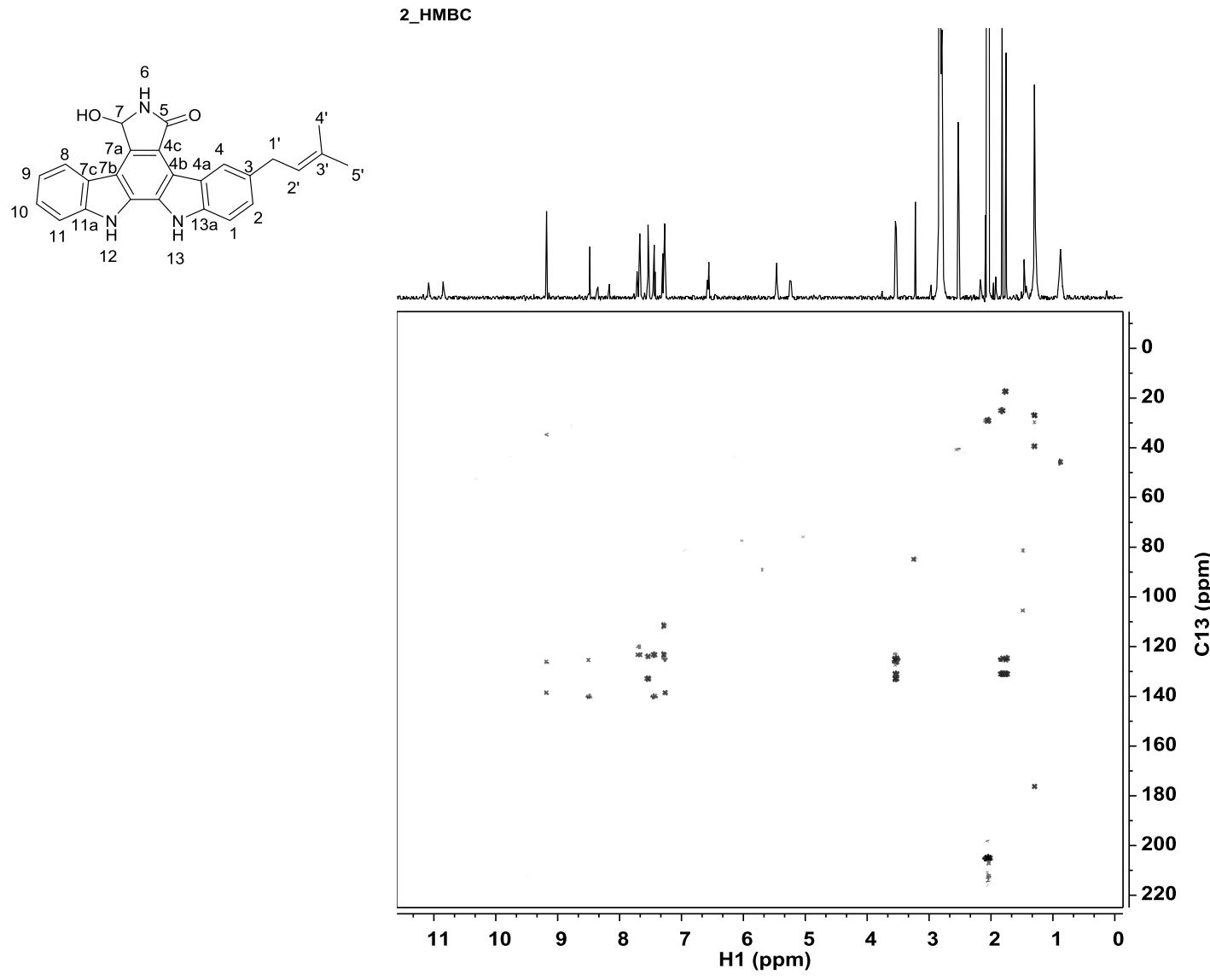
<sup>1</sup>H NMR Spectrum (500 MHz) of compound **57** in Acetone-d<sub>6</sub>



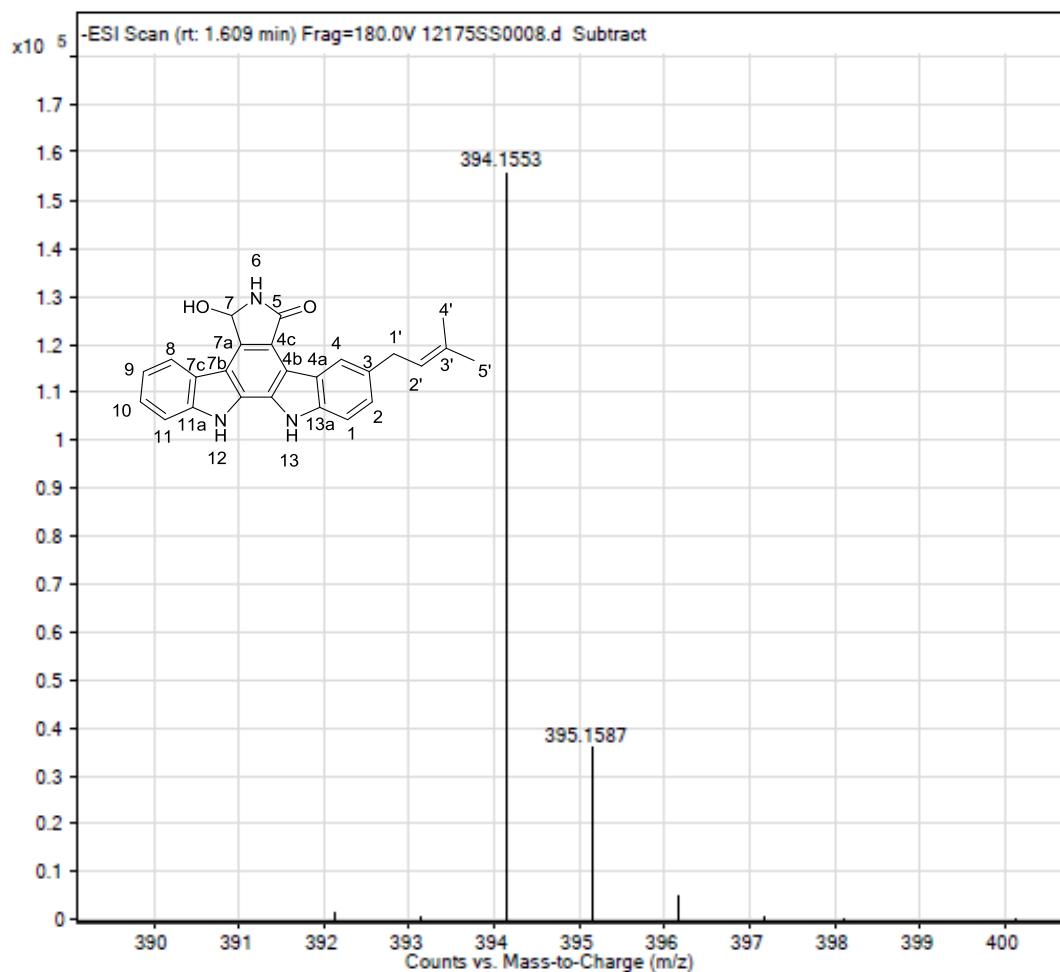
2D  $^1\text{H}$ - $^1\text{H}$  COSY NMR spectrum (500 MHz) of **57** in Acetone-d6



2D  $^1\text{H}$ - $^{13}\text{C}$  HSQC NMR spectrum (500 MHz) of **57** in Acetone-d<sub>6</sub>

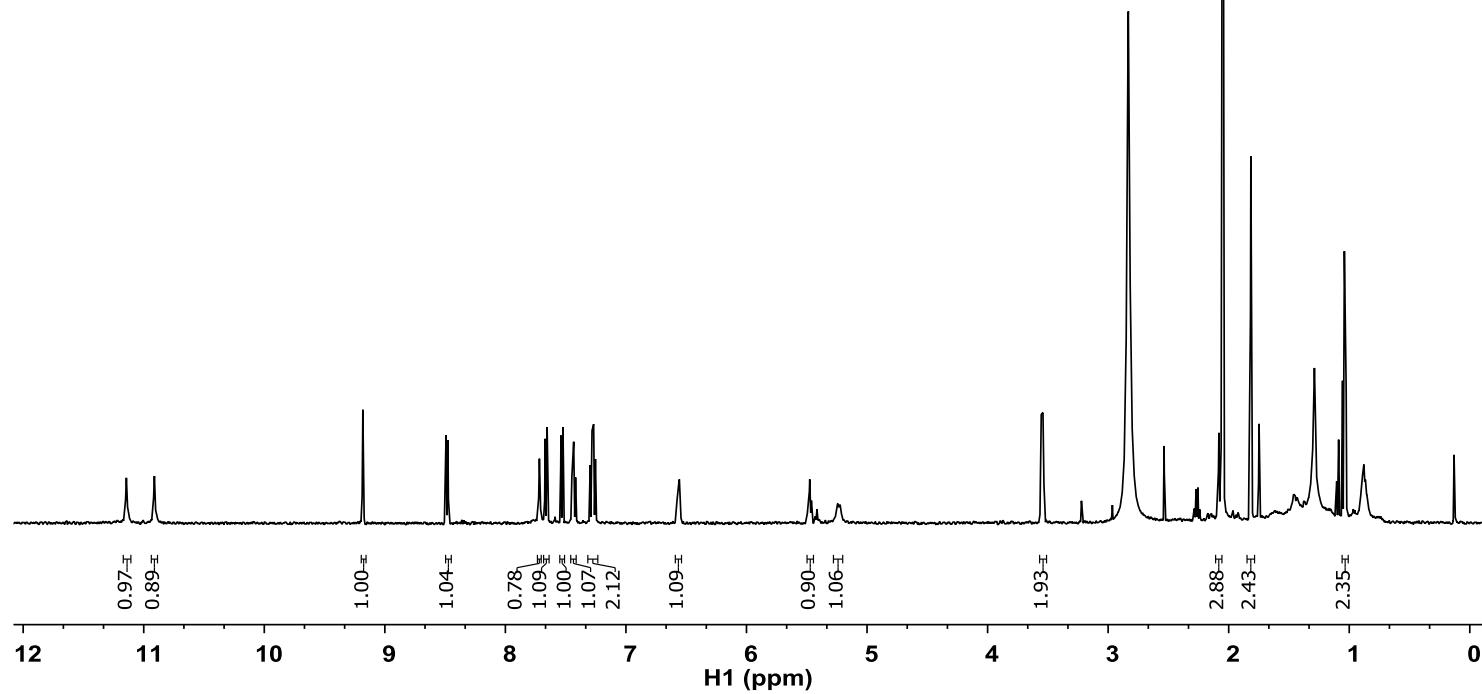
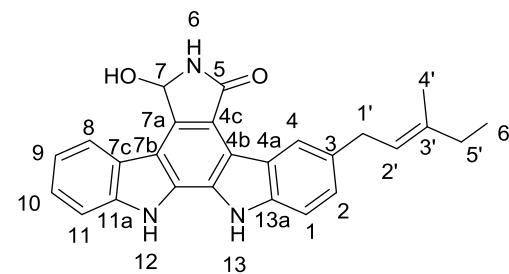


### 2D $^1\text{H}$ - $^{13}\text{C}$ HMBC NMR spectrum (500 MHz) of **57** in Acetone-d6

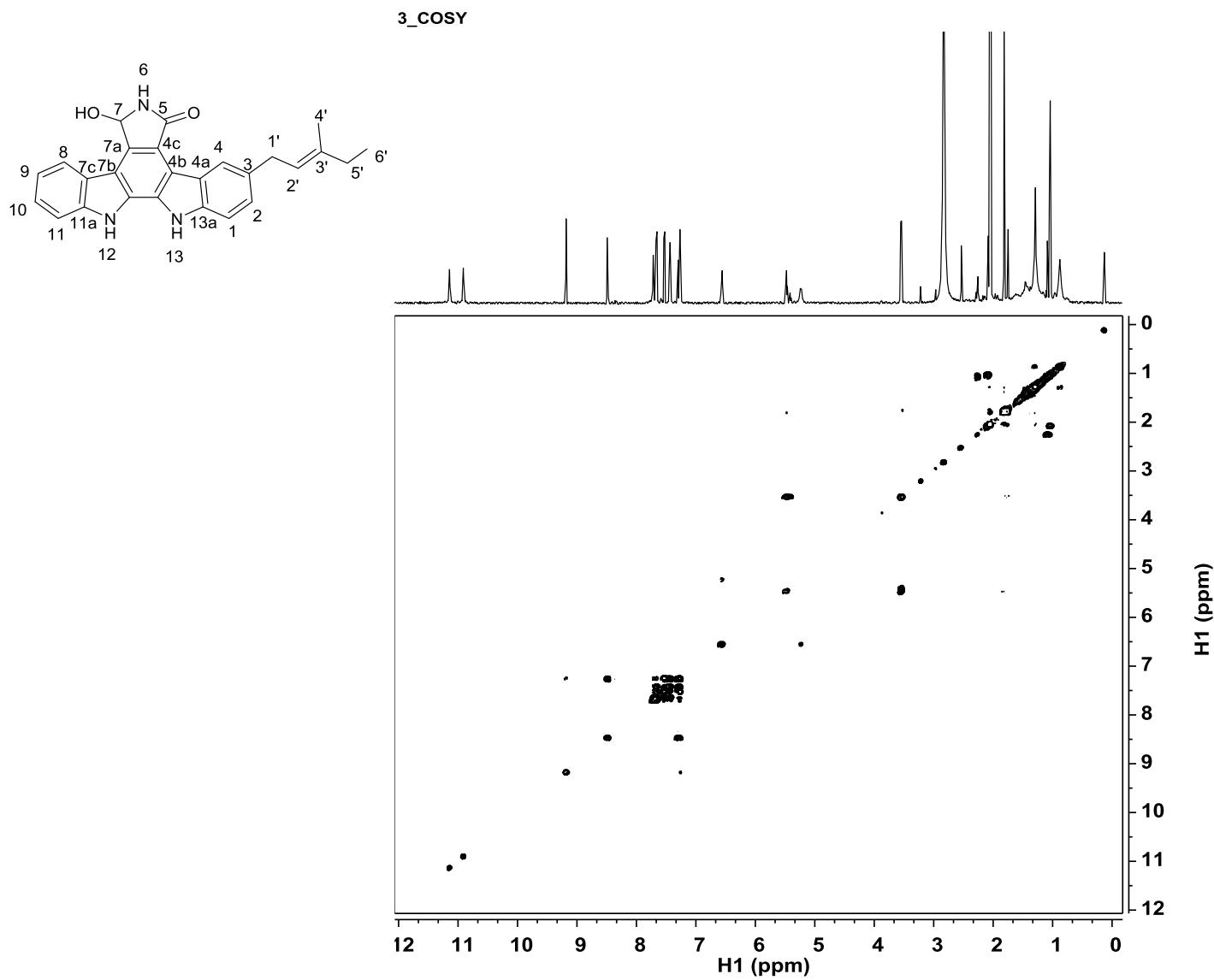


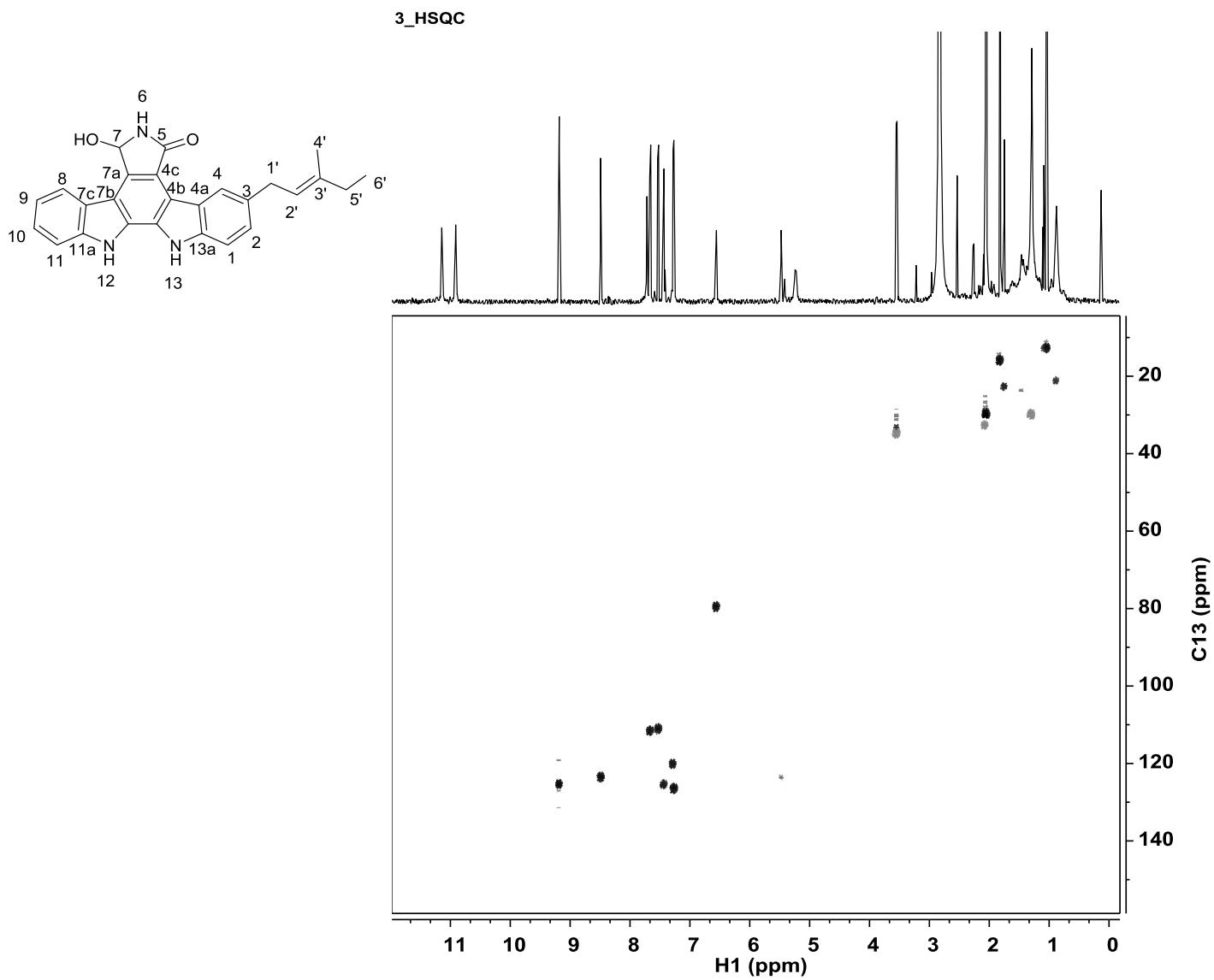
HRMS spectrum of **57**

3\_1H



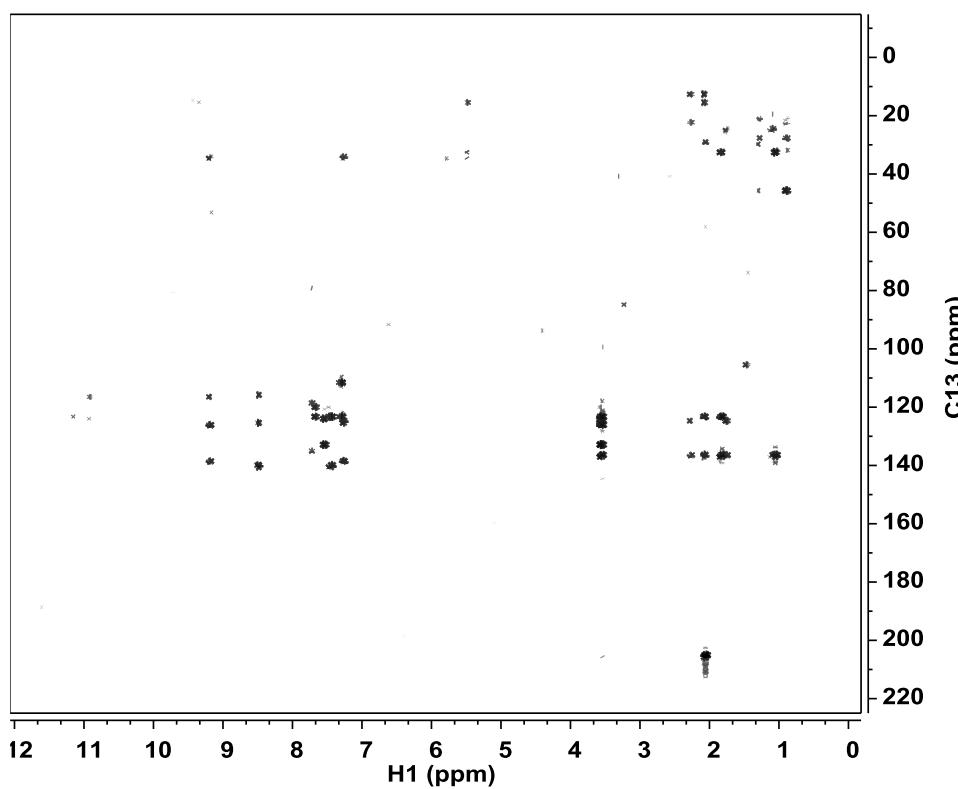
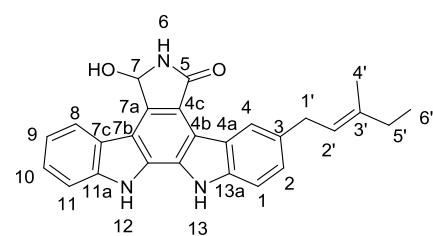
<sup>1</sup>H NMR Spectrum (500 MHz) of compound **58** in Acetone-d6



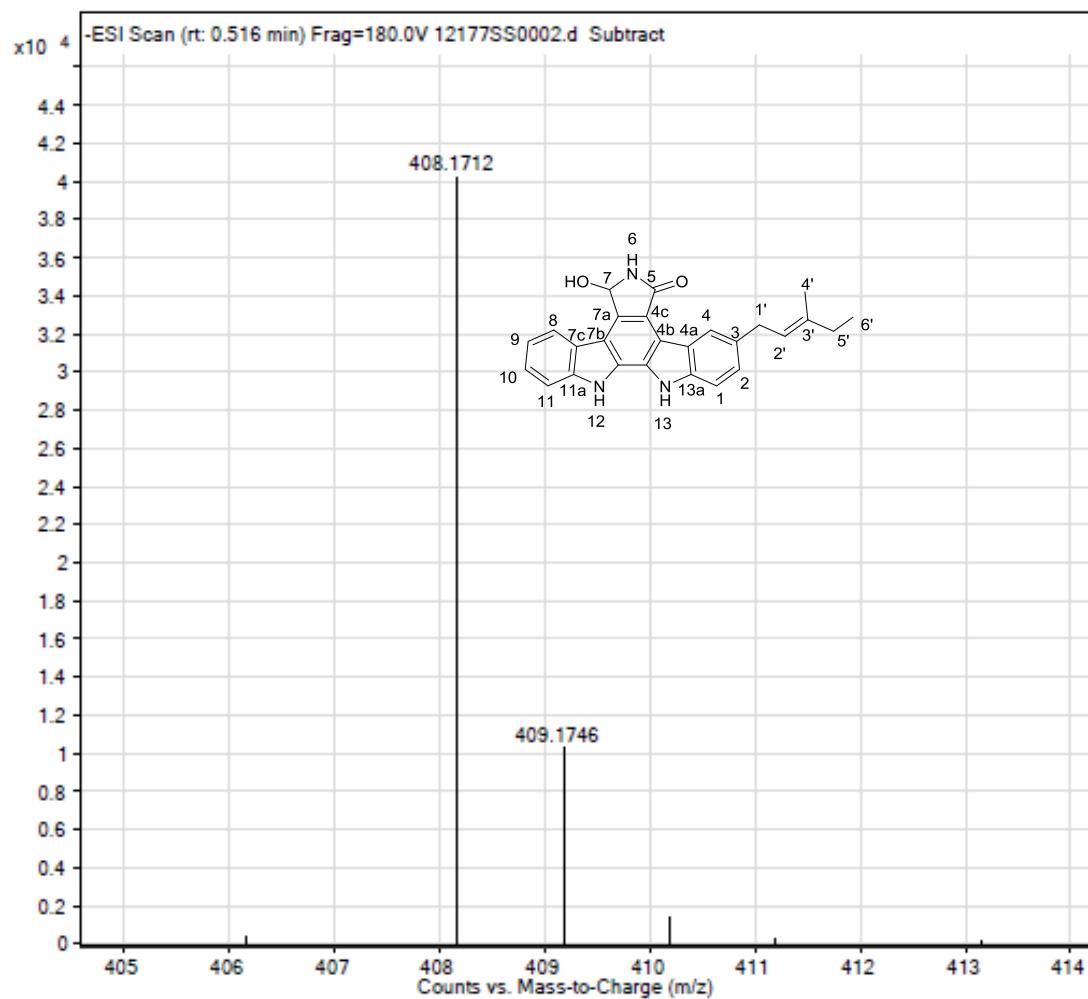


2D  $^1\text{H}$ - $^{13}\text{C}$  HSQC NMR spectrum (500 MHz) of **58** in Acetone-d6

**3\_HMBC**

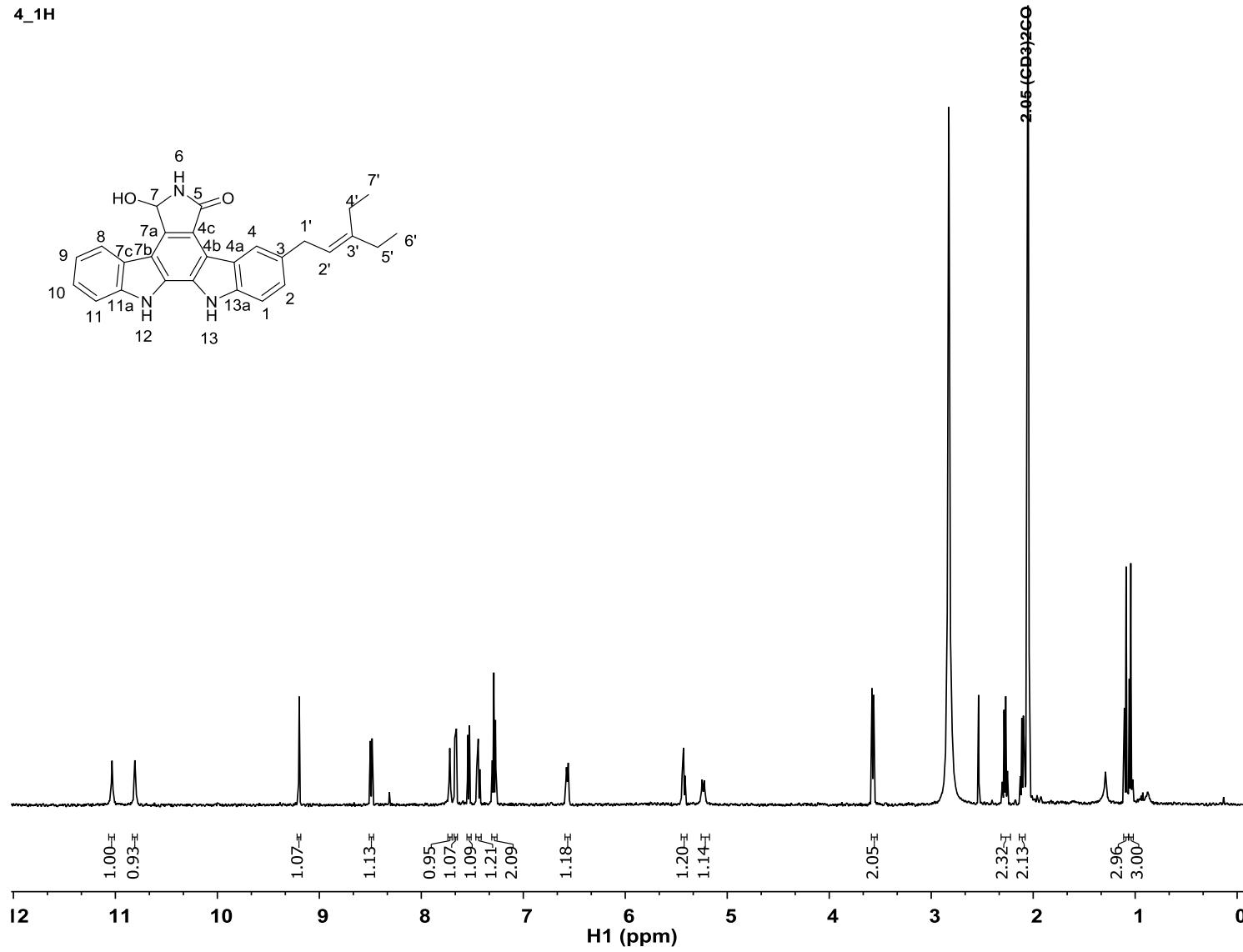
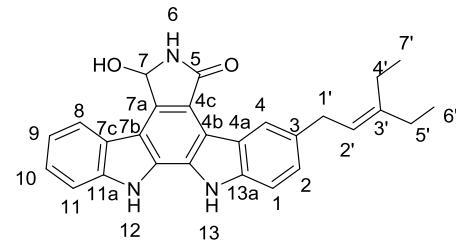


2D <sup>1</sup>H-<sup>13</sup>C HMBC NMR spectrum (500 MHz) of **58** in Acetone-d<sub>6</sub>



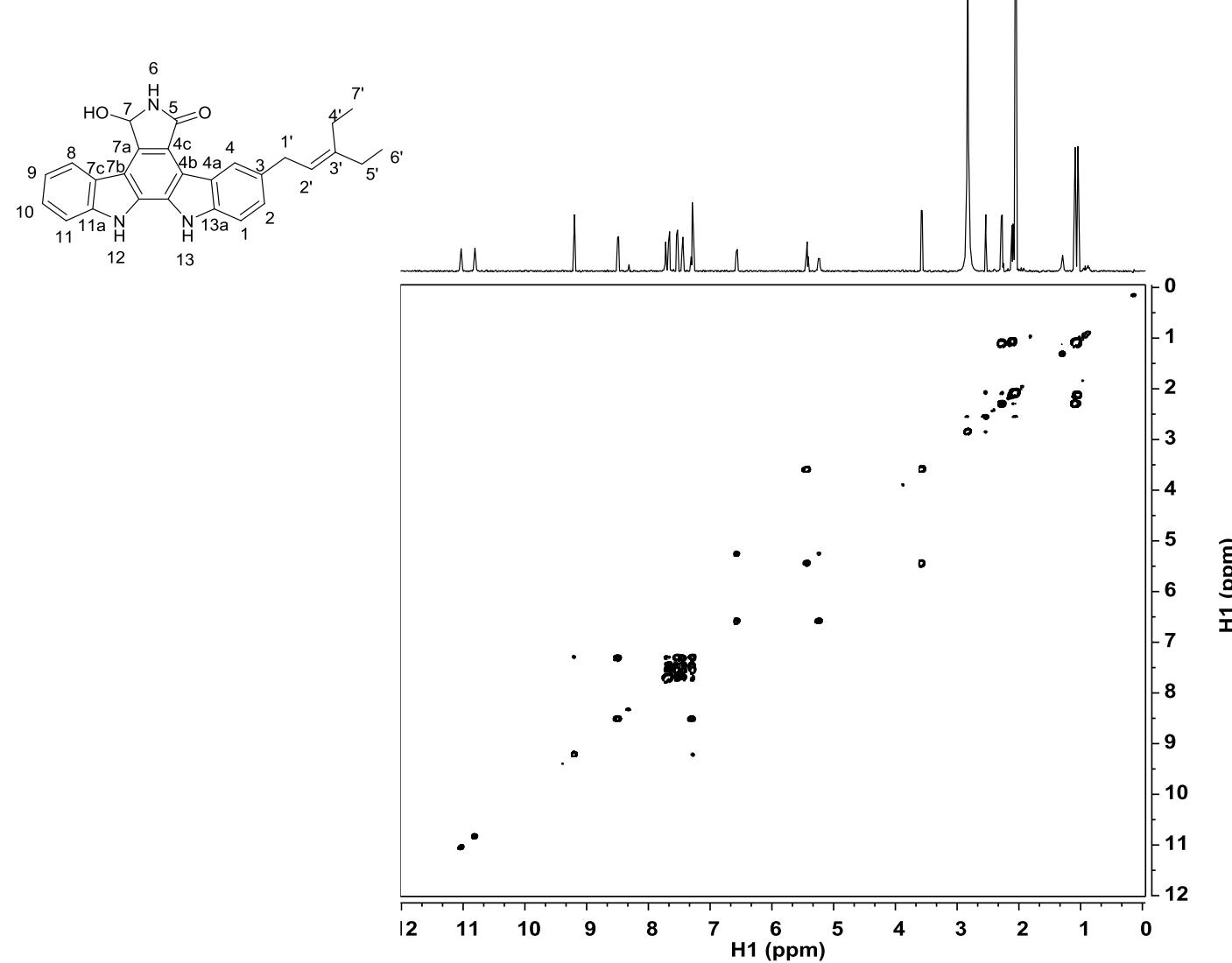
HRMS spectrum of **58**

4\_1H

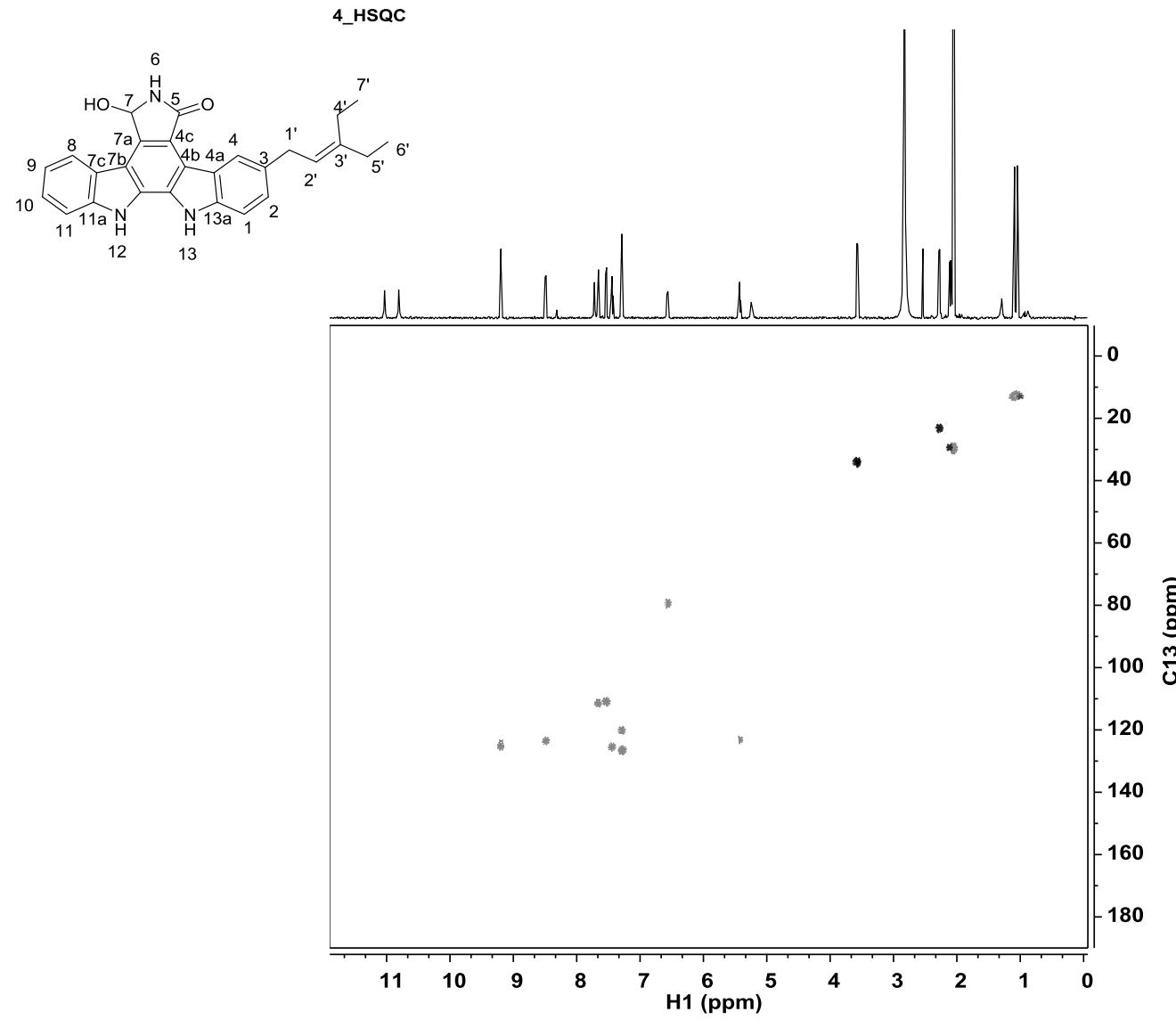


<sup>1</sup>H NMR Spectrum (500 MHz) of compound **59** in Acetone-d<sub>6</sub>

4\_COSY

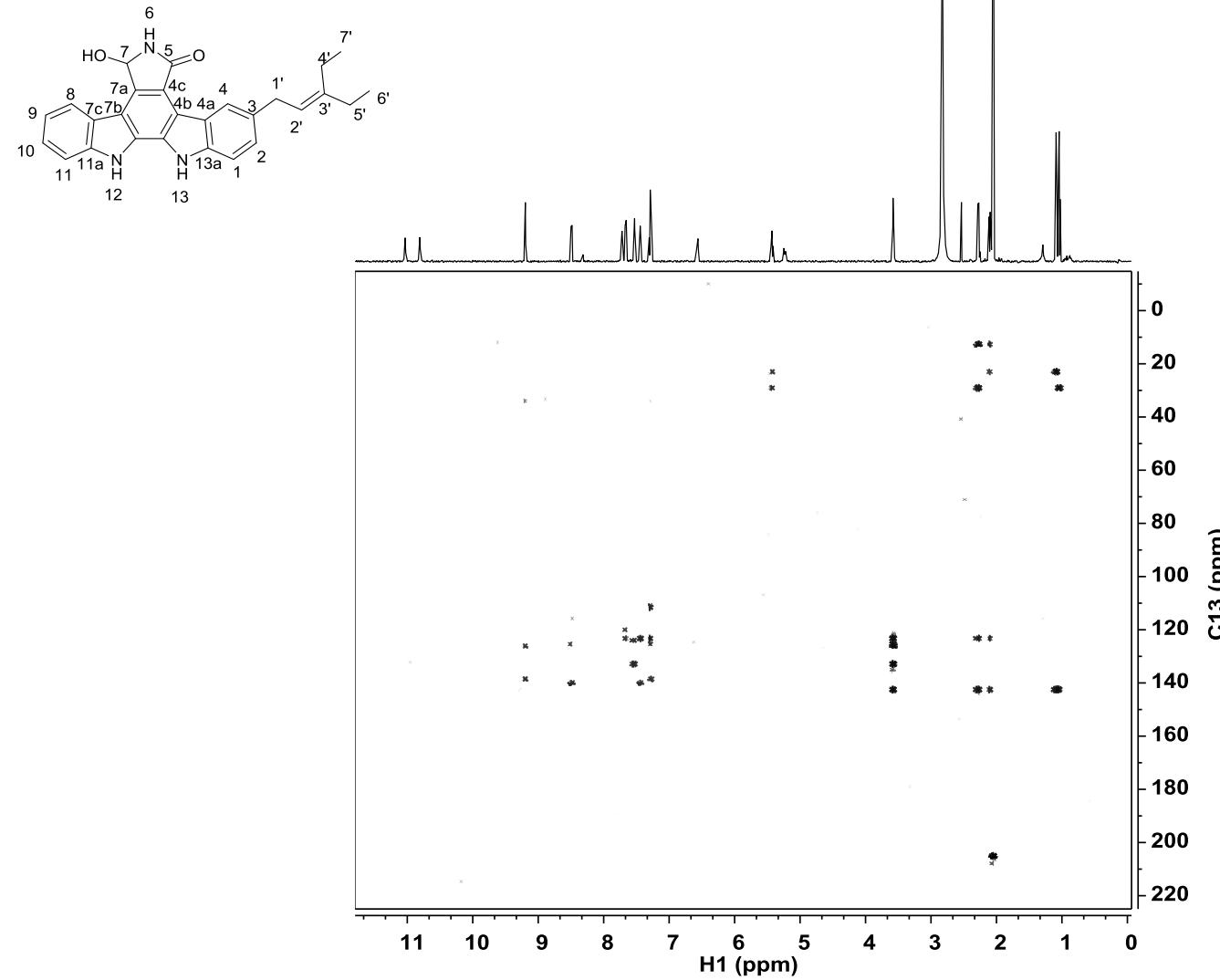


2D  $^1\text{H}$ - $^1\text{H}$  COSY NMR spectrum (500 MHz) of **59** in Acetone-d6

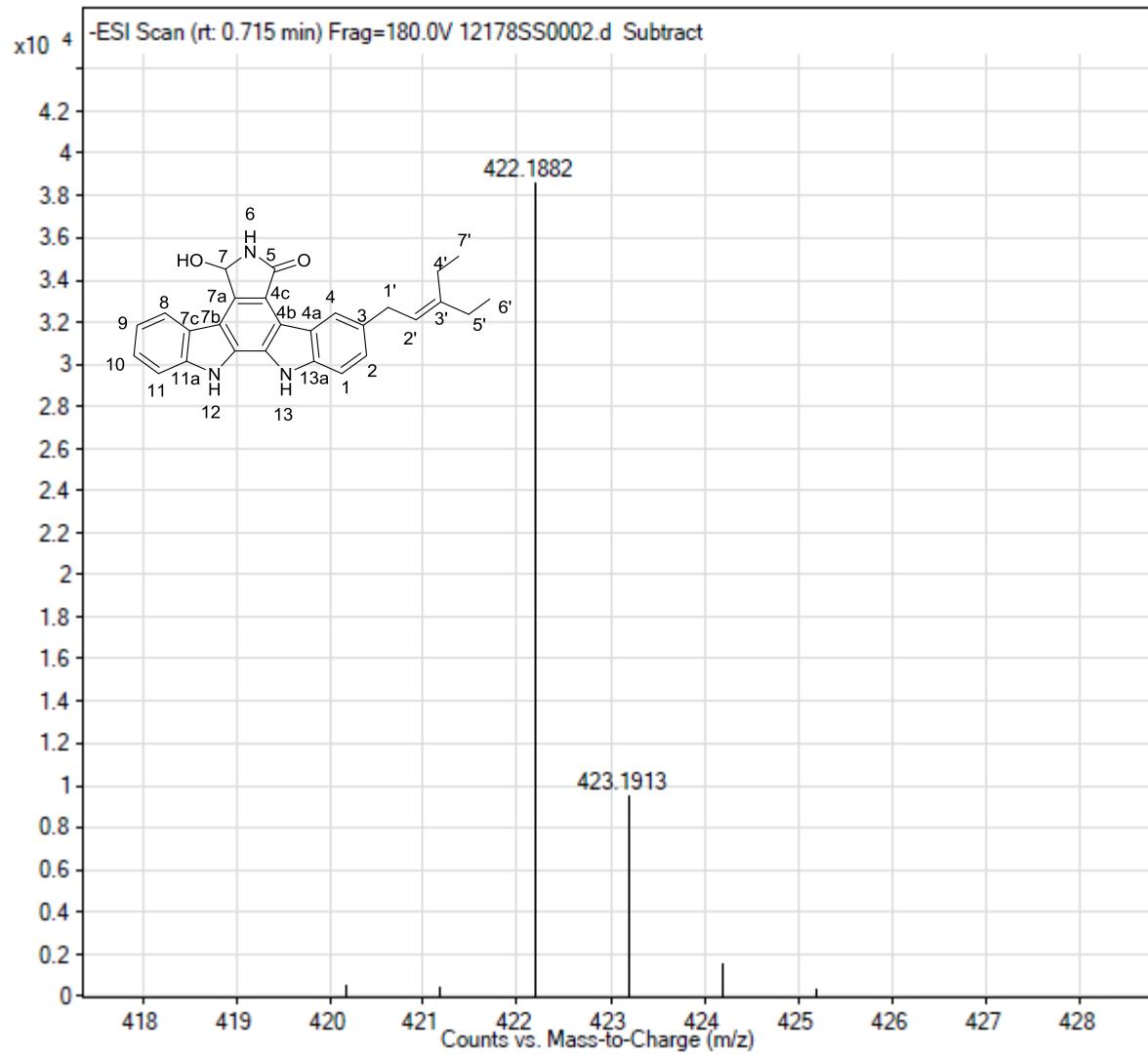


### 2D $^1\text{H}$ - $^{13}\text{C}$ HSQC NMR spectrum (500 MHz) of **59** in Acetone-d6

**4\_HMBC**

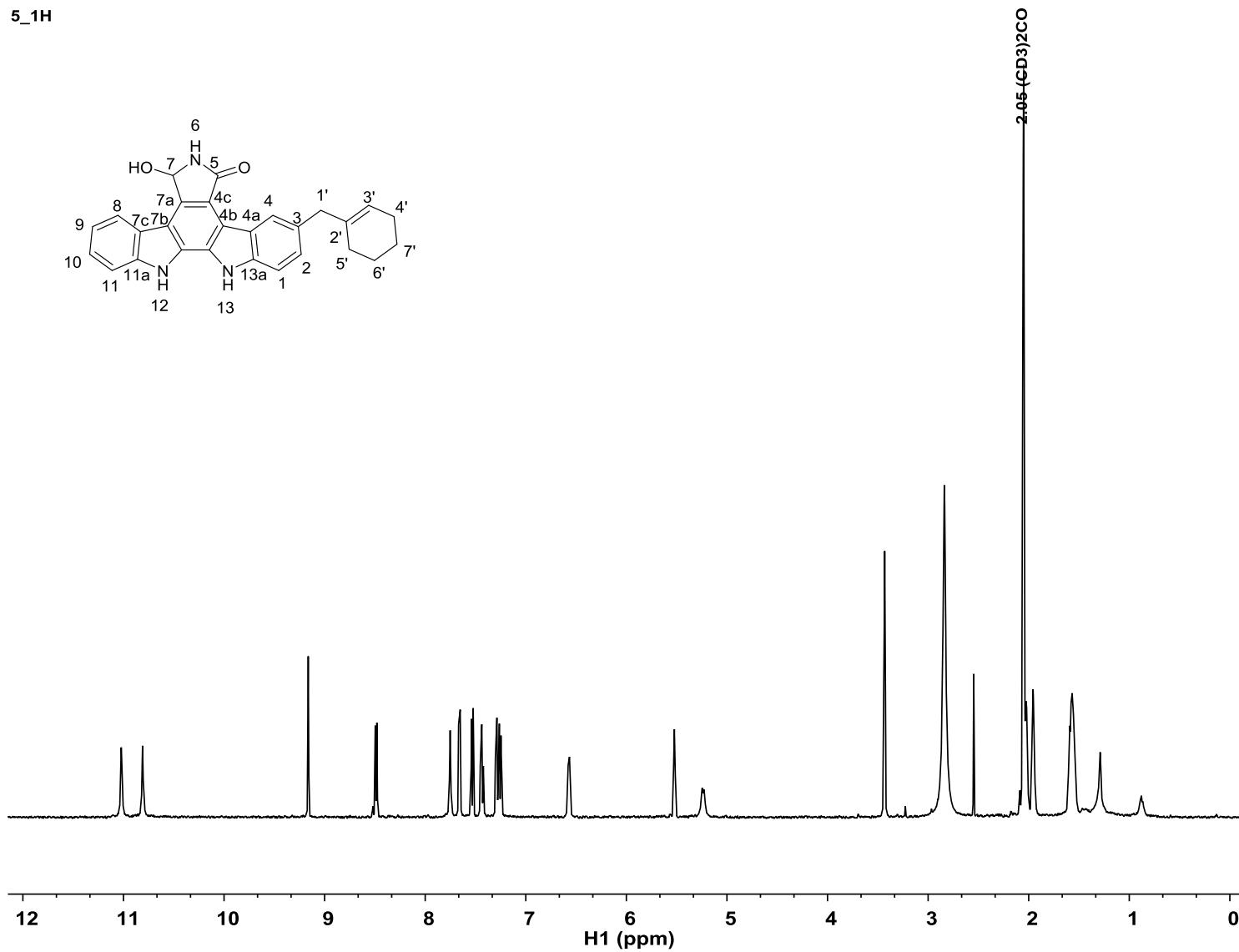


2D  $^1\text{H}$ - $^{13}\text{C}$  HMBC NMR spectrum (500 MHz) of **59** in Acetone-d6

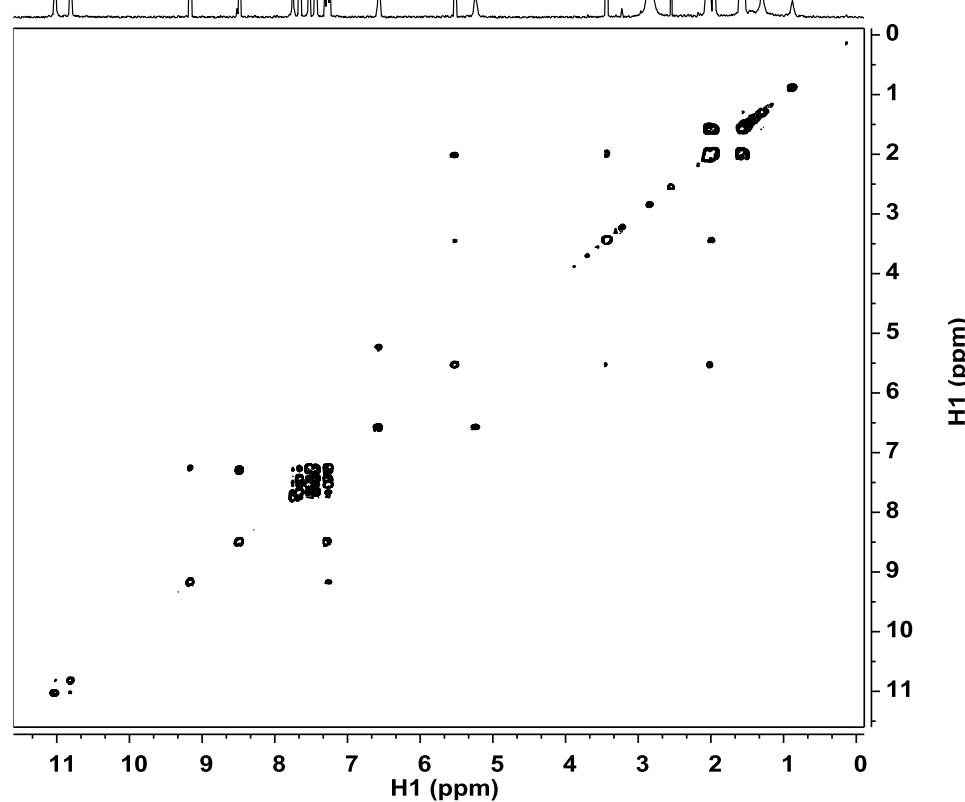
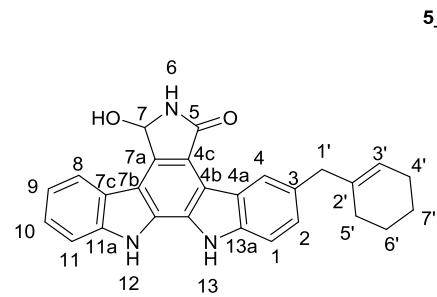


HRMS spectrum of **59**

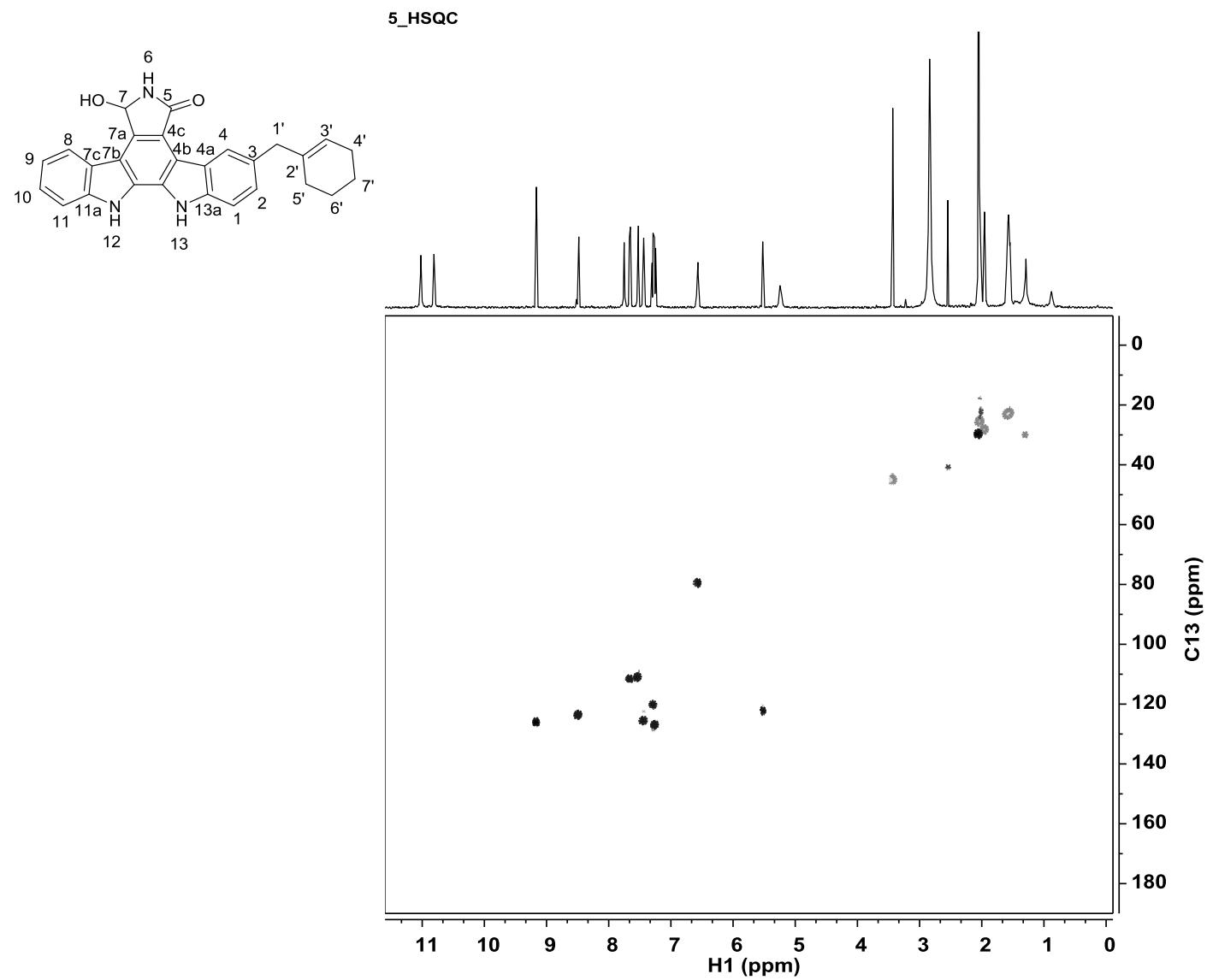
5\_1H



$^1\text{H}$  NMR Spectrum (500 MHz) of compound **60** in Acetone-d<sub>6</sub>

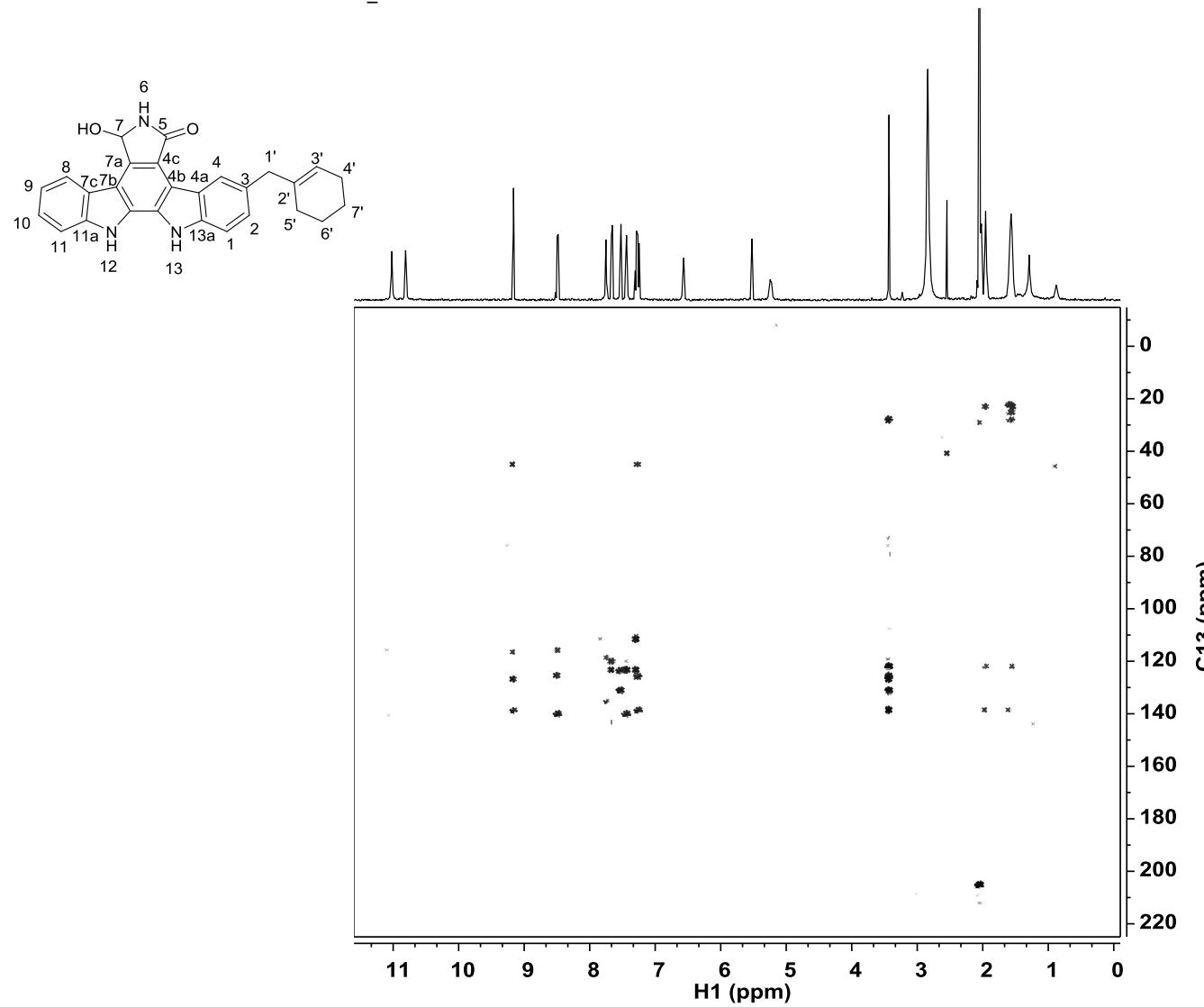


2D  $^1\text{H}$ - $^1\text{H}$  COSY NMR spectrum (500 MHz) of **60** in Acetone-d<sub>6</sub>

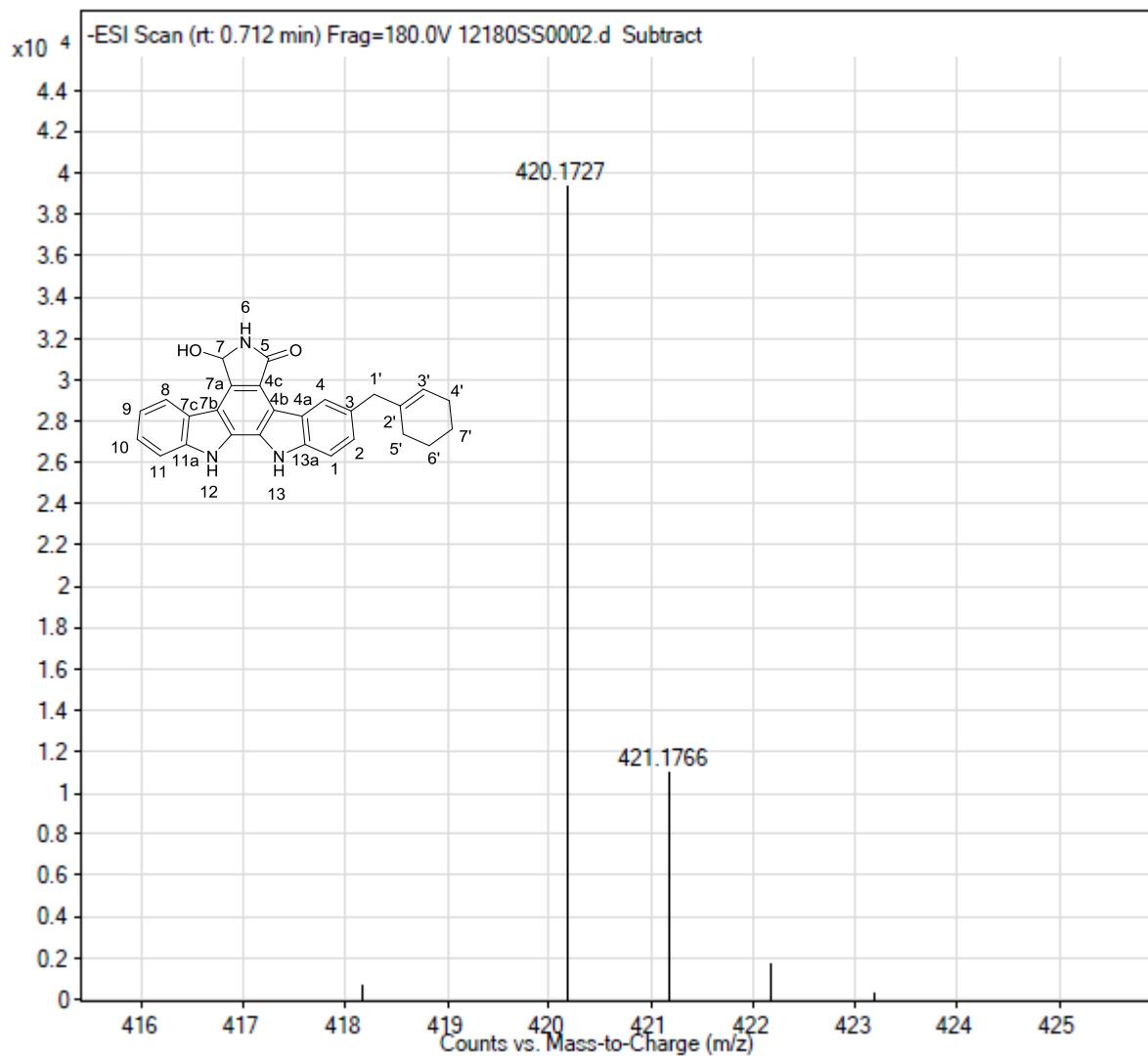


2D  $^1\text{H}$ - $^{13}\text{C}$  HSQC NMR spectrum (500 MHz) of **60** in Acetone-d6

**5\_HMBC**

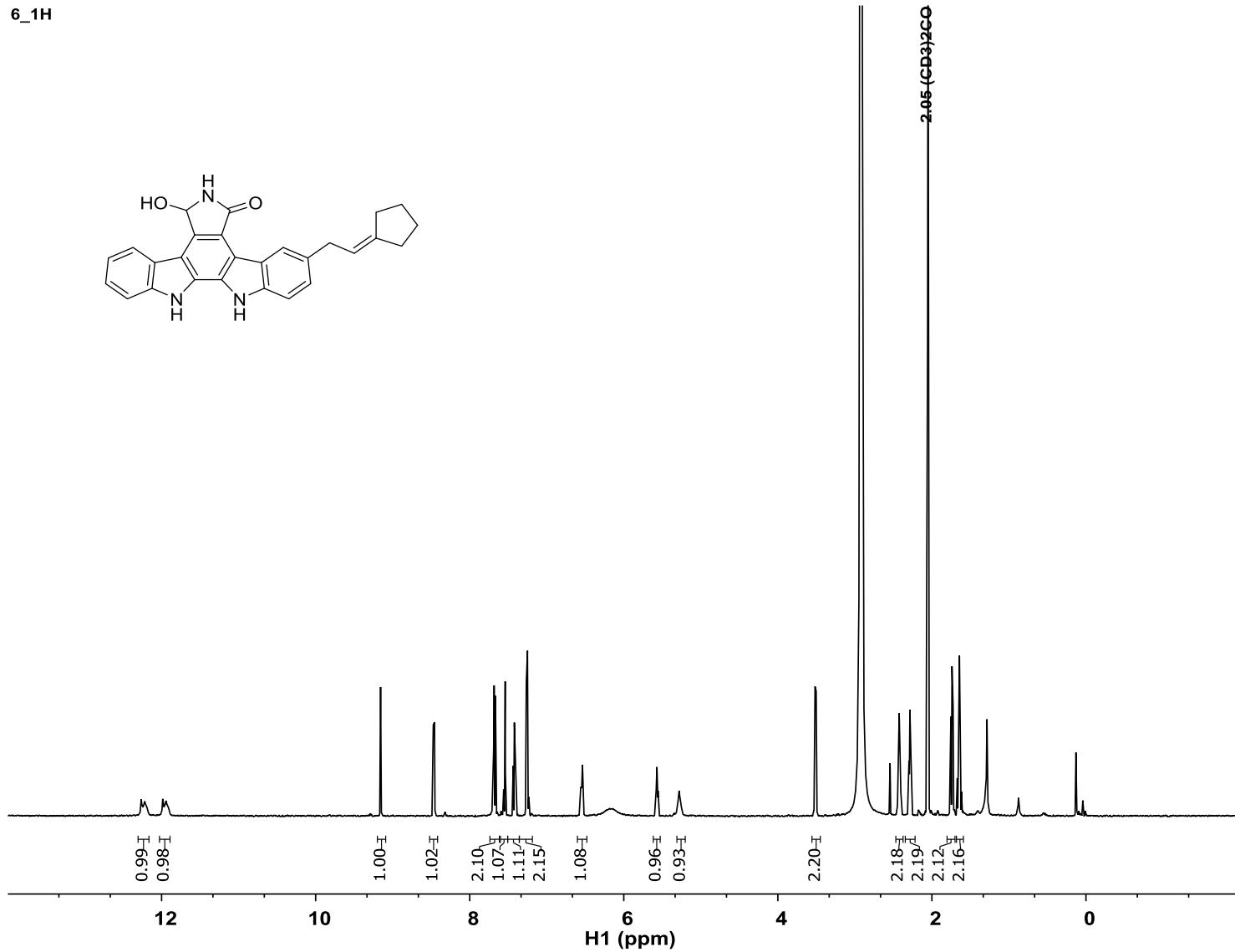


2D  $^1\text{H}$ - $^{13}\text{C}$  HMBC NMR spectrum (500 MHz) of **60** in Acetone-d6



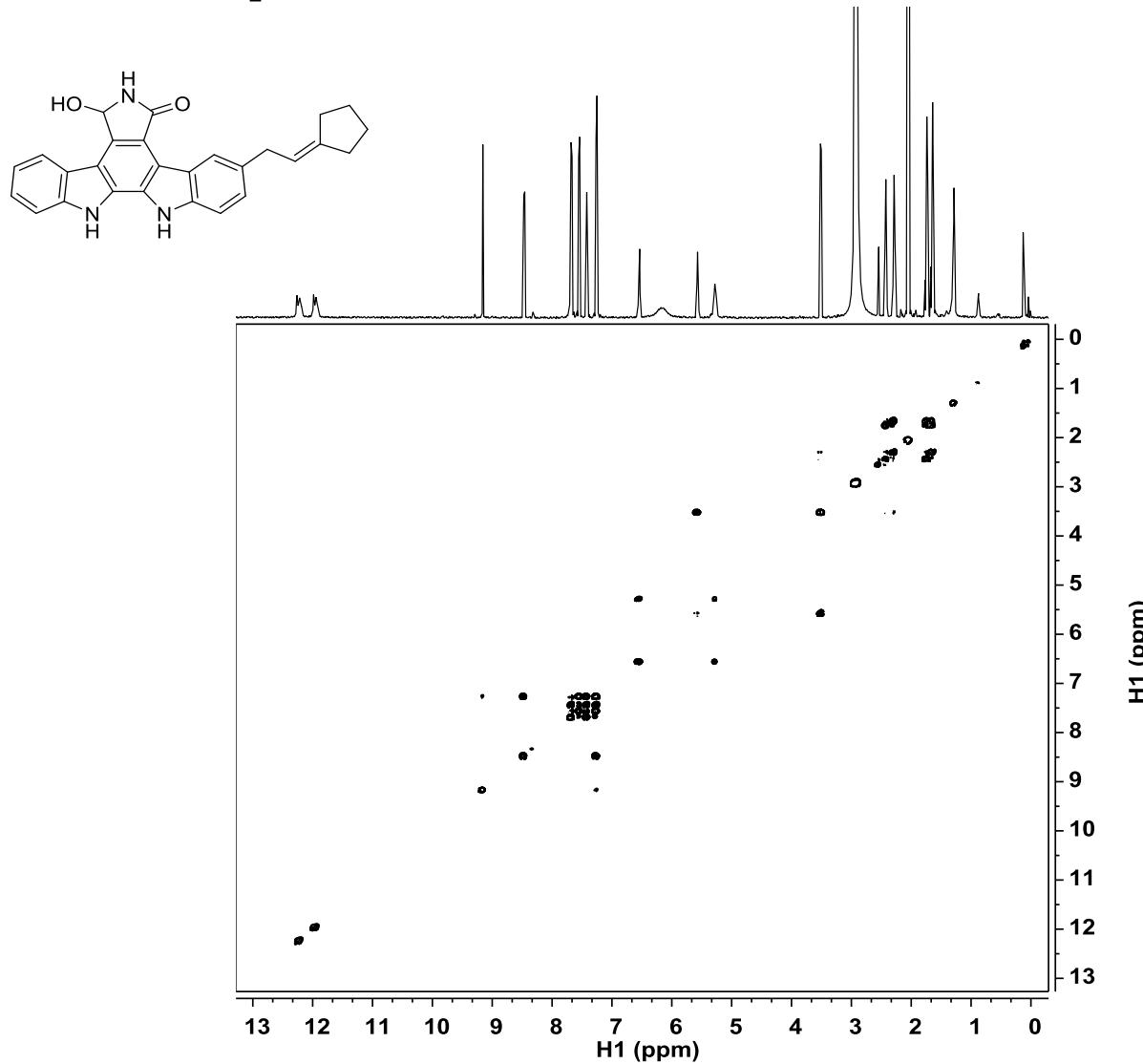
HRMS spectrum of **60**

**6\_1H**

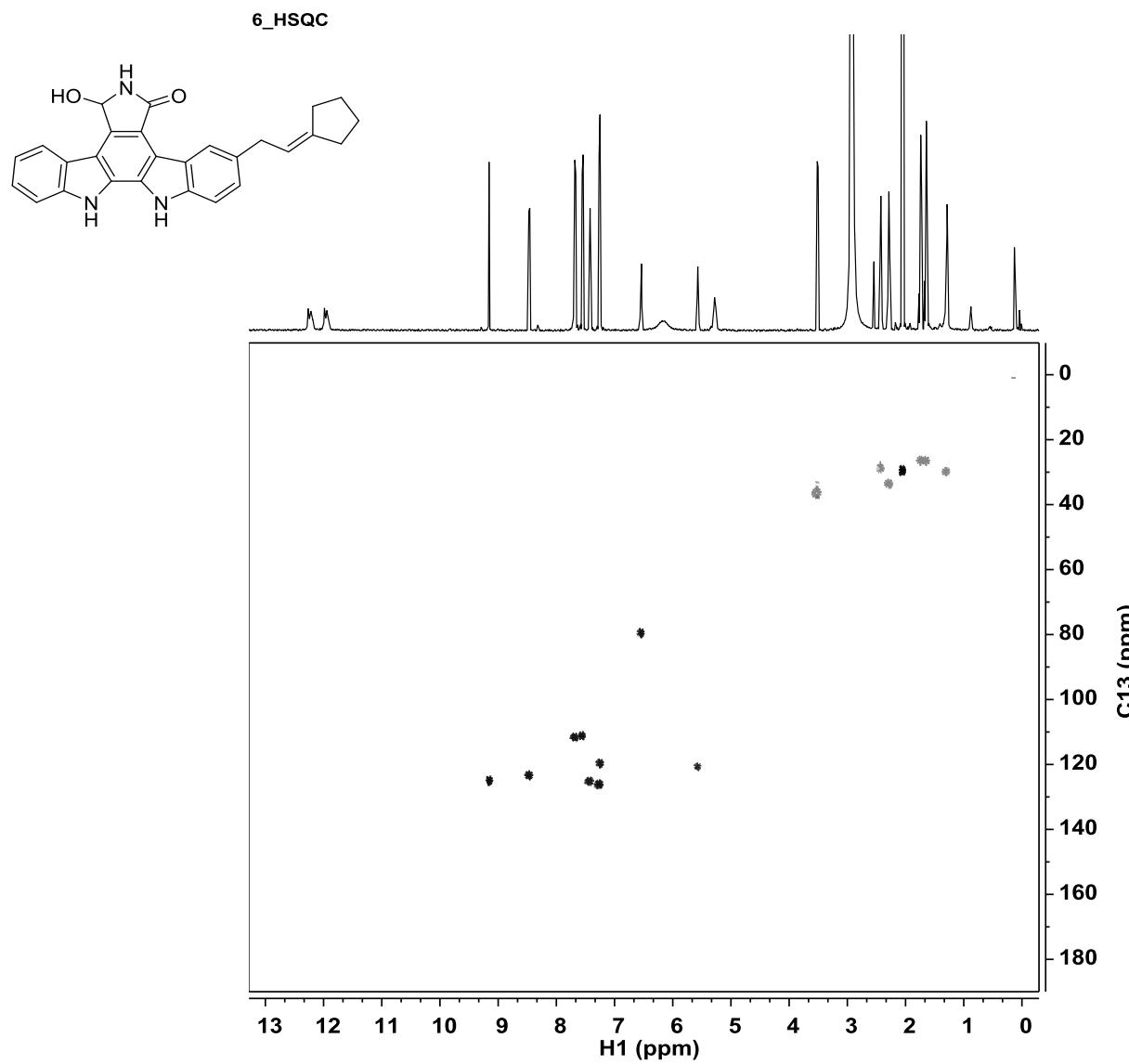


<sup>1</sup>H NMR Spectrum (500 MHz) of compound **61** in Acetone-d6

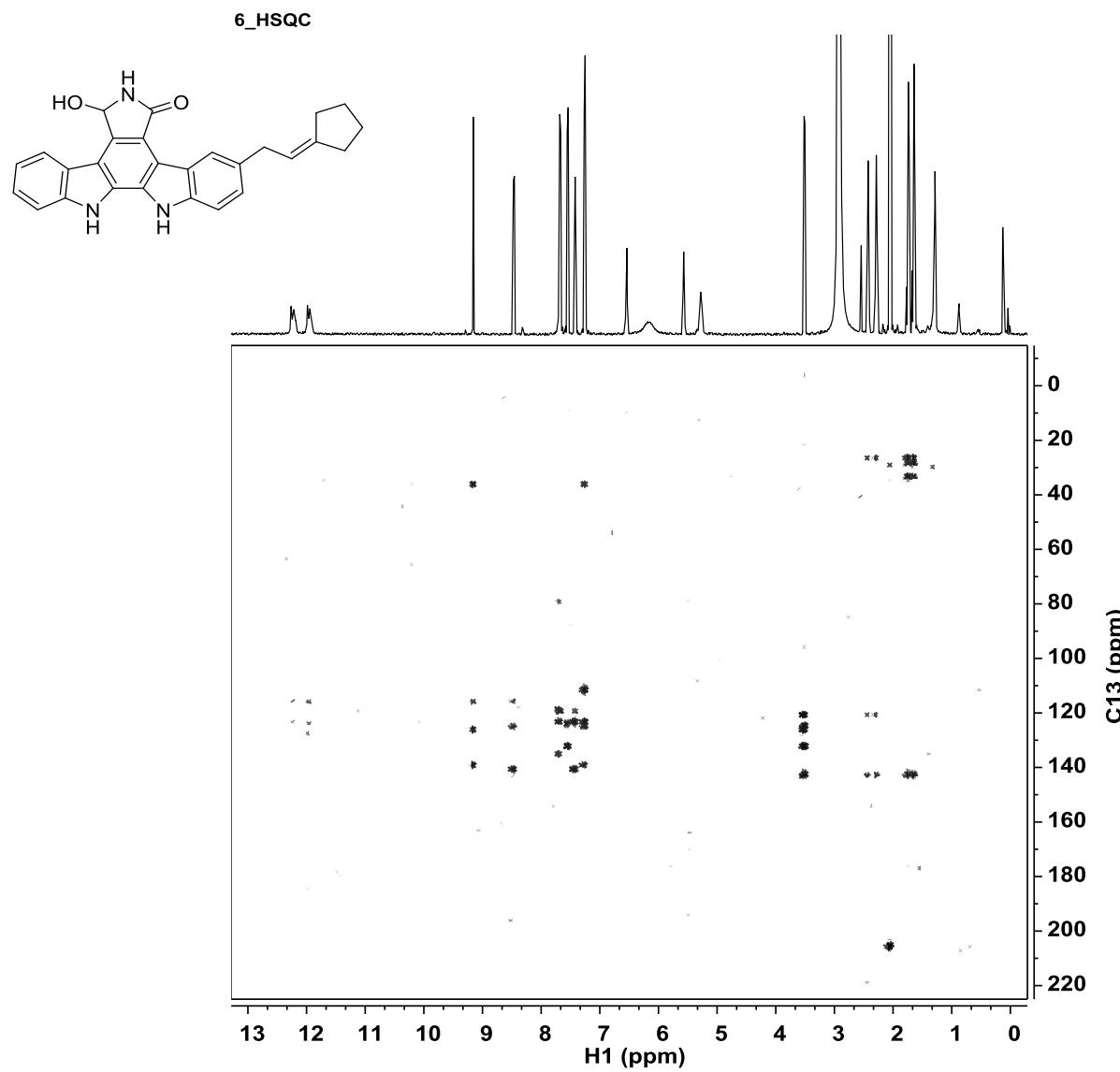
**6\_COSY**



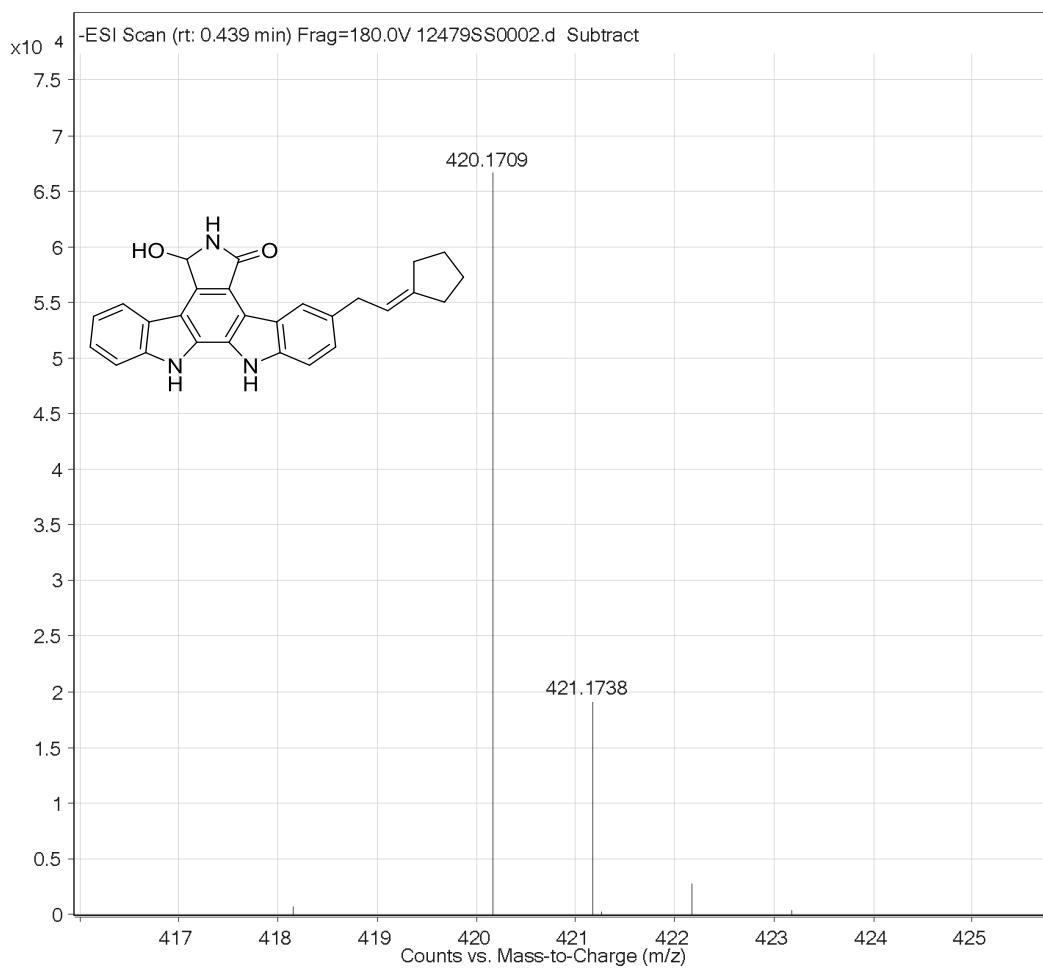
2D  $^1\text{H}$ - $^1\text{H}$  COSY NMR spectrum (500 MHz) of **61** in Acetone- $d_6$



2D  $^1\text{H}$ - $^{13}\text{C}$  HSQC NMR spectrum (500 MHz) of **61** in Acetone-d<sub>6</sub>

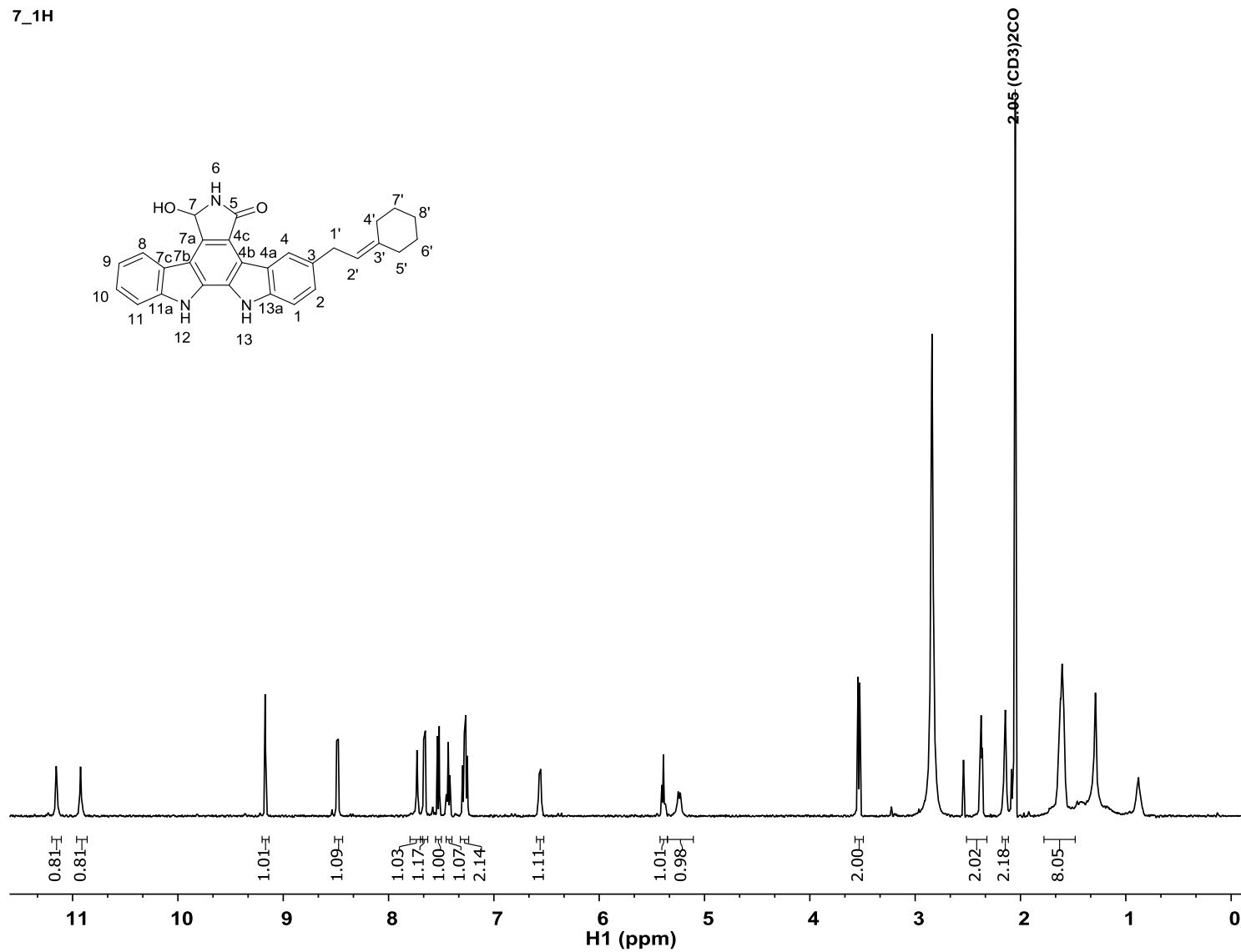


2D  $^1\text{H}$ - $^{13}\text{C}$  HMBC NMR spectrum (500 MHz) of **61** in Acetone-d6



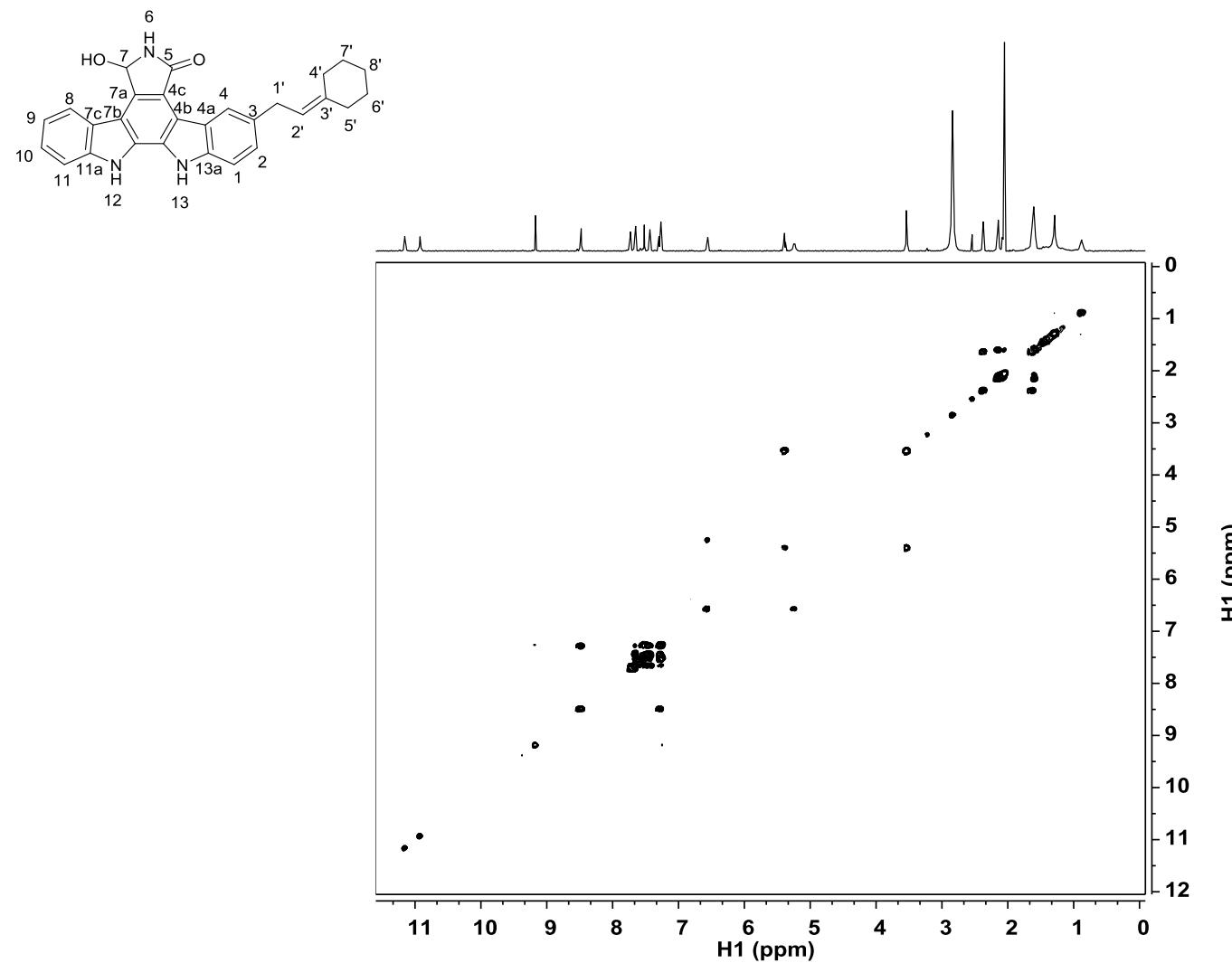
HRMS spectrum of **61**

7\_1H

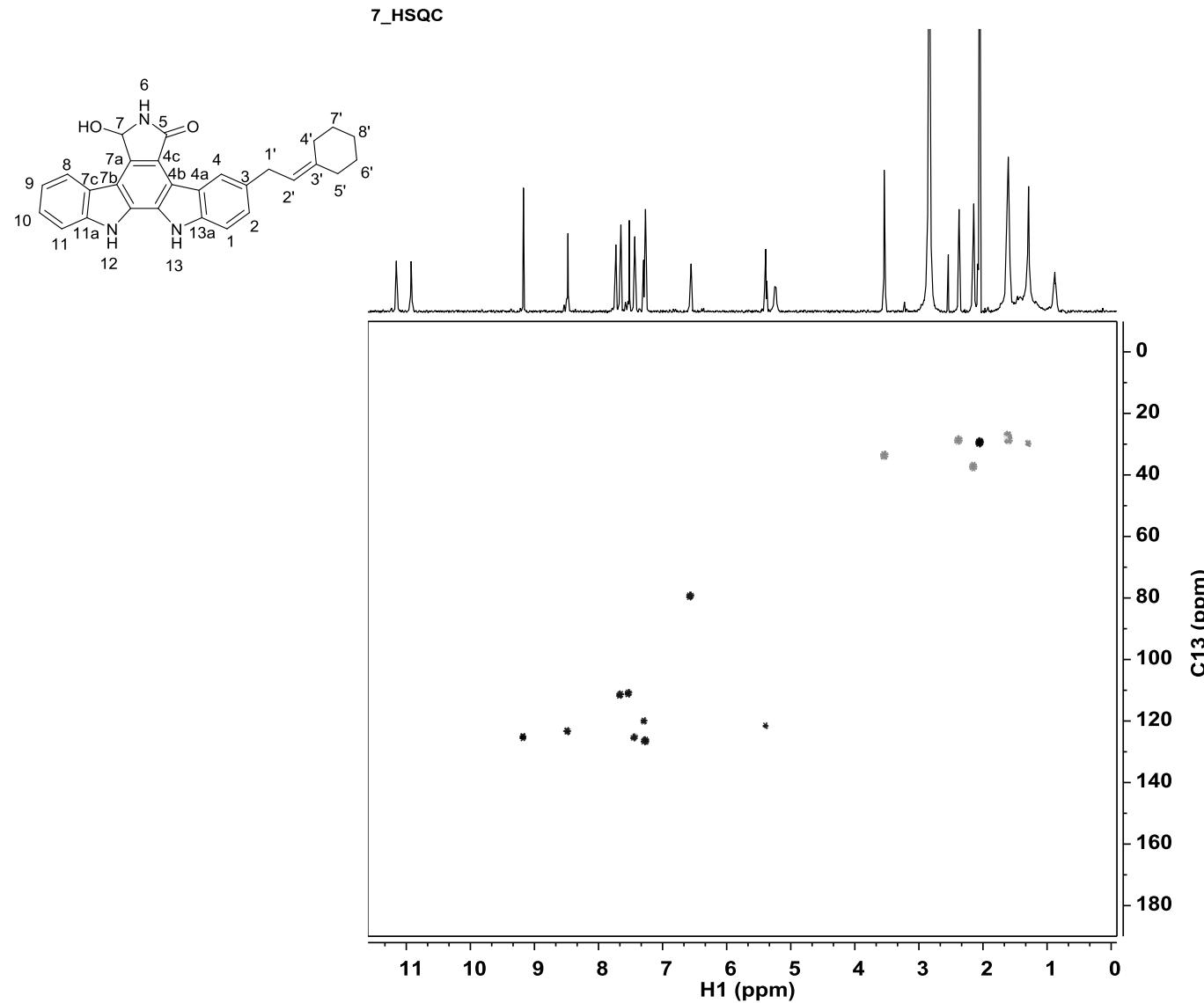


<sup>1</sup>H NMR Spectrum (500 MHz) of compound **62** in Acetone-d<sub>6</sub>

7\_COSY

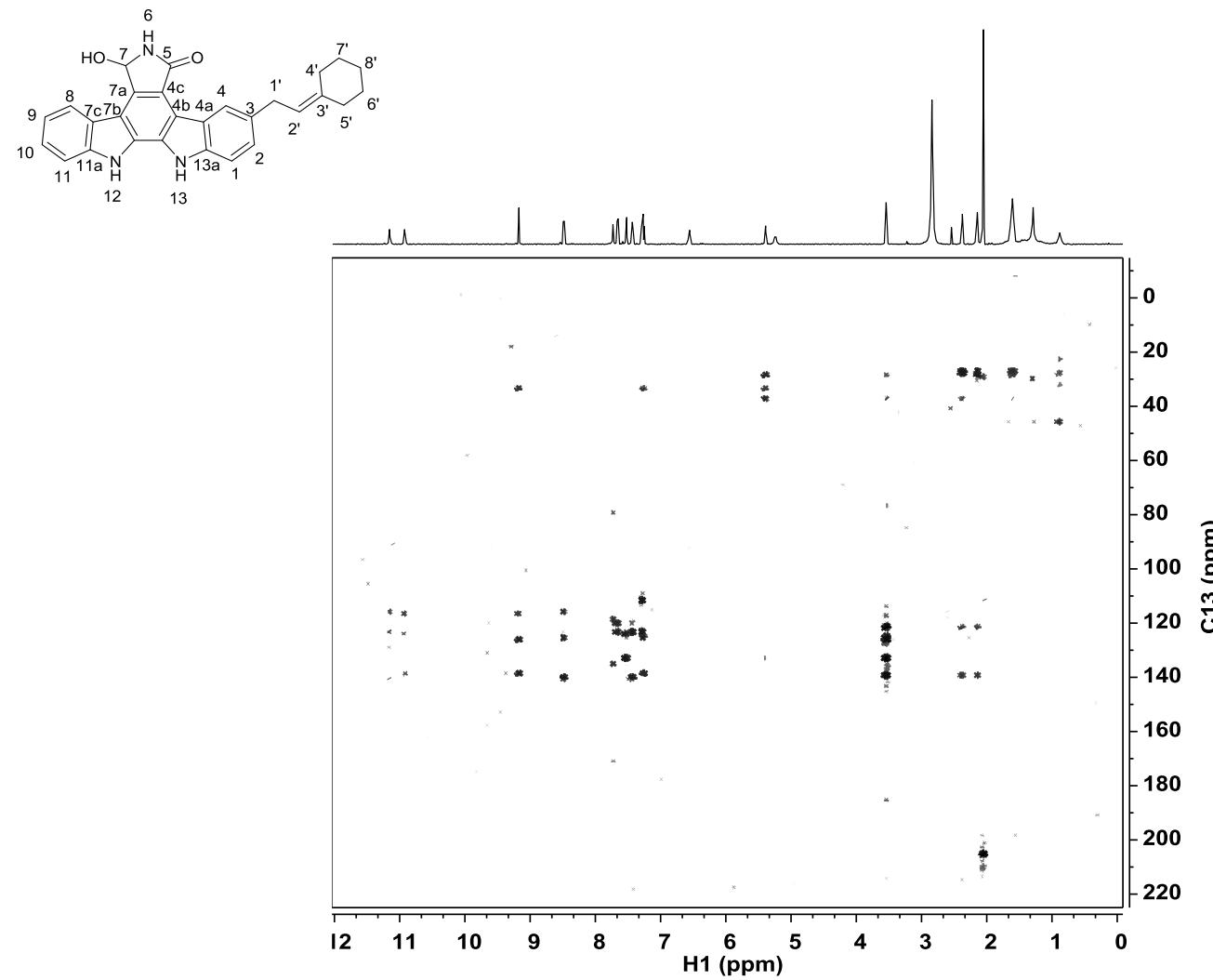


2D  $^1\text{H}$ - $^1\text{H}$  COSY NMR spectrum (500 MHz) of **62** in Acetone- $d_6$

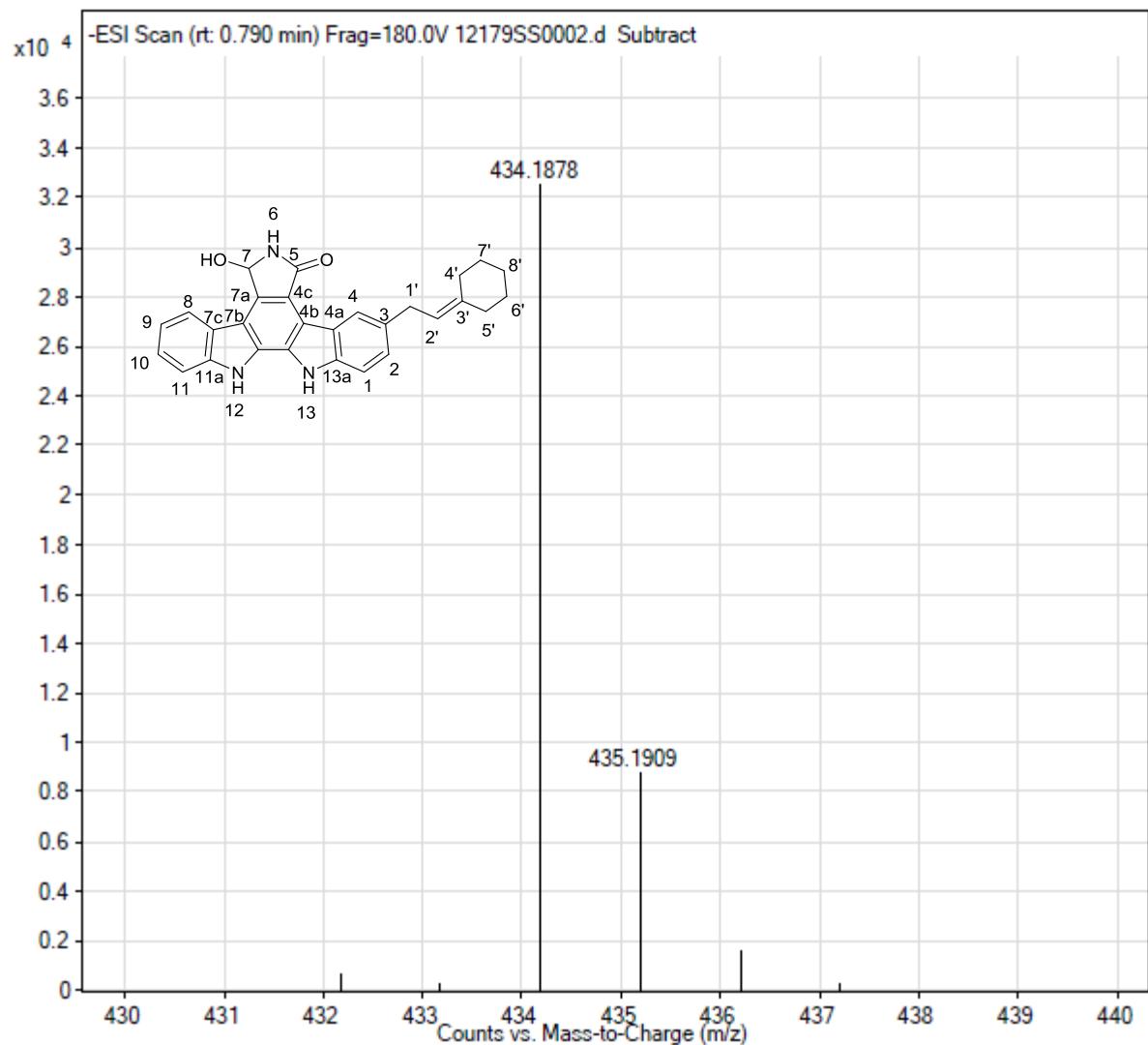


2D  $^1\text{H}$ - $^{13}\text{C}$  HSQC NMR spectrum (500 MHz) of **62** in Acetone-d6

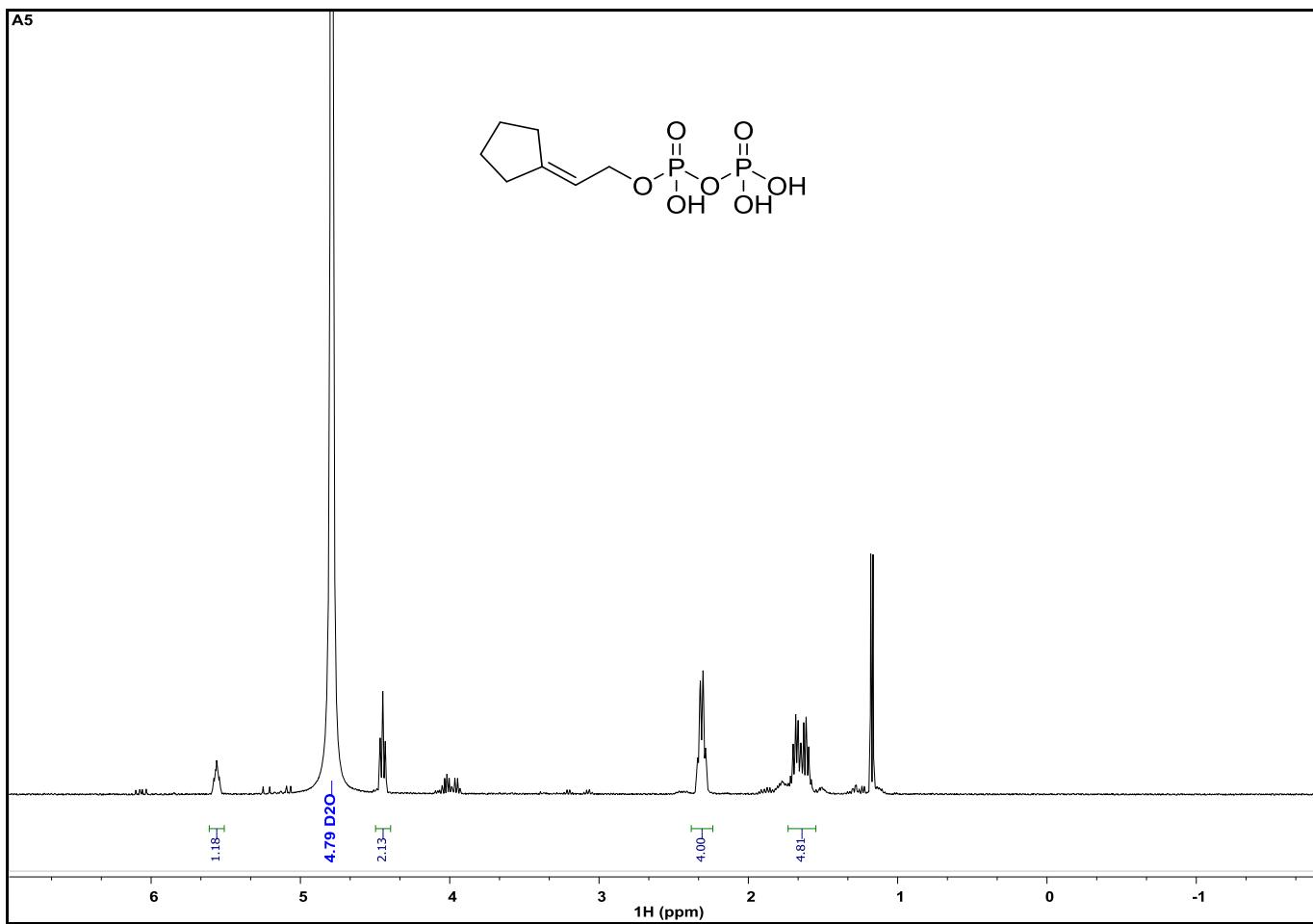
7\_HSQC



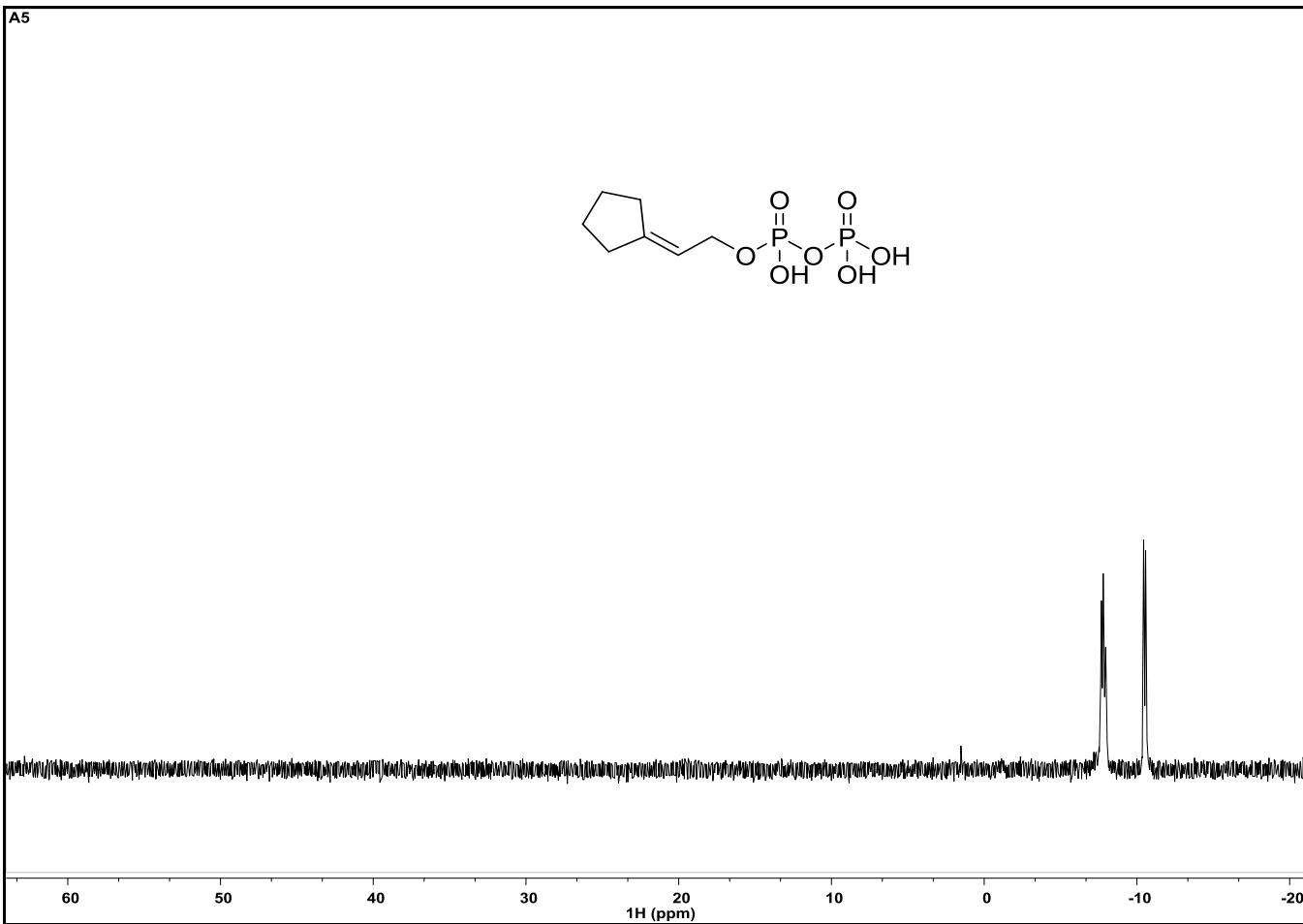
2D <sup>1</sup>H-<sup>13</sup>C HMBC NMR spectrum (500 MHz) of **62** in Acetone-d6



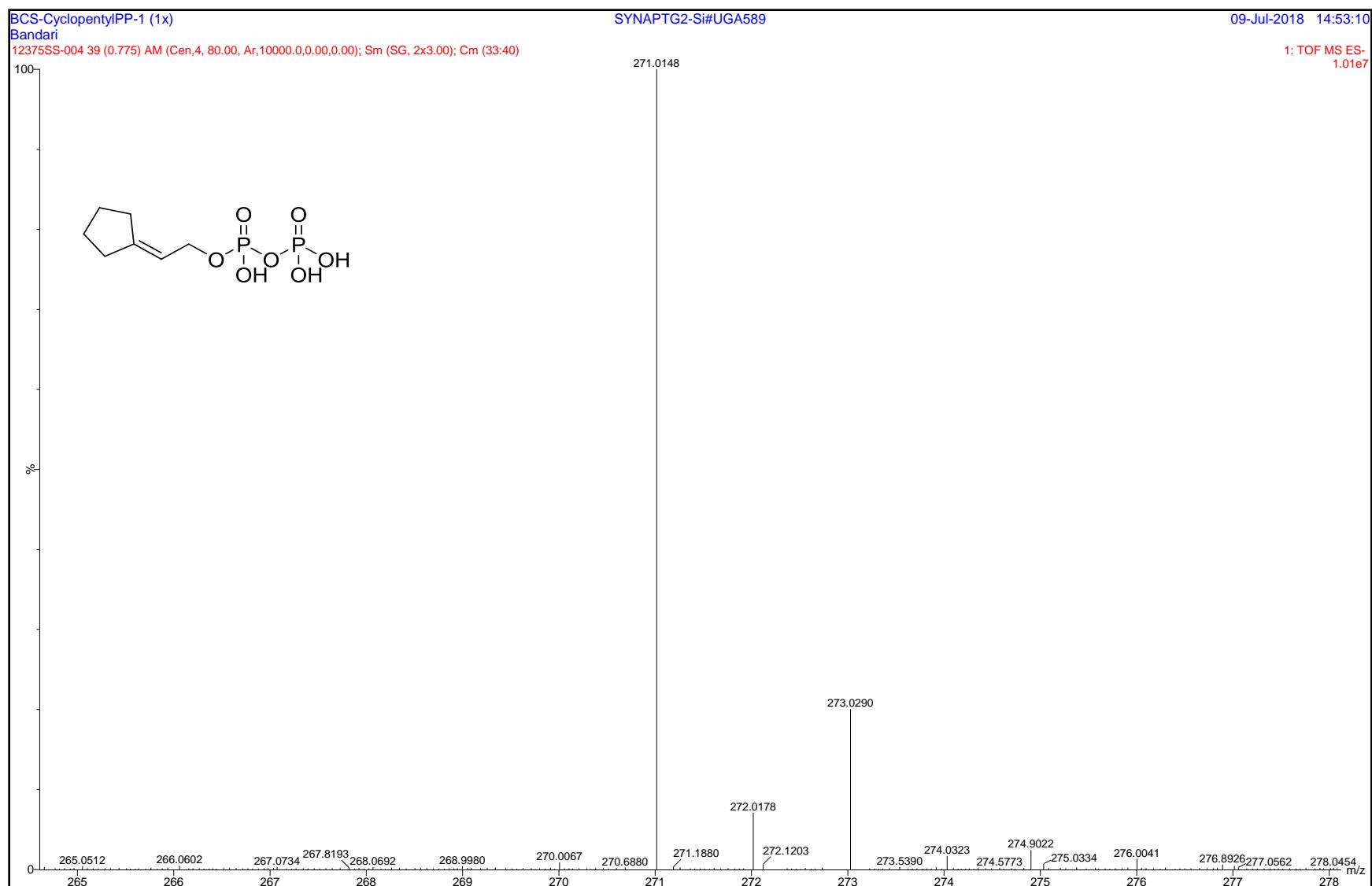
HRMS spectrum of **62**

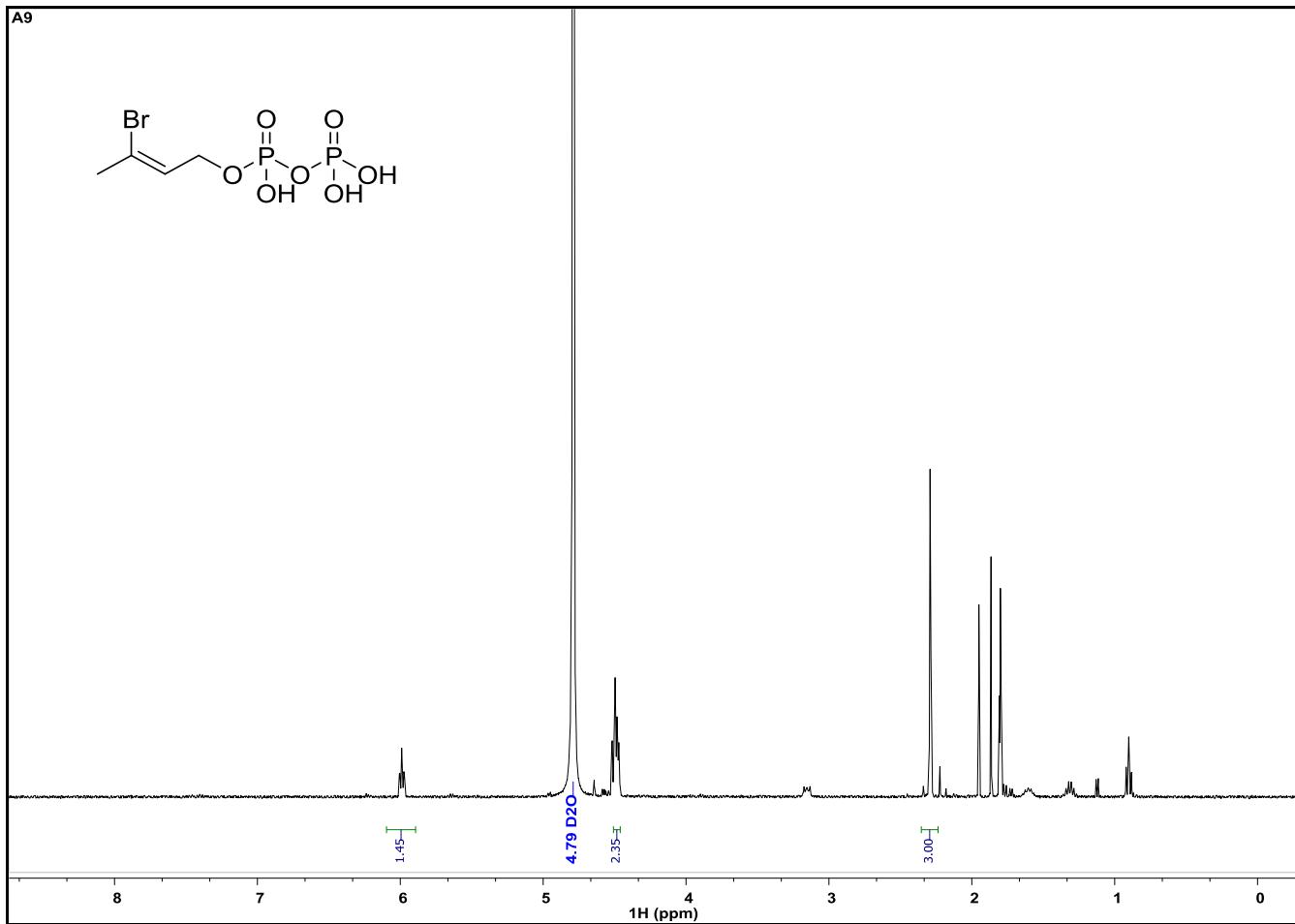


$^1\text{H}$  NMR Spectrum (400 MHz) of compound 5 in  $\text{D}_2\text{O}$

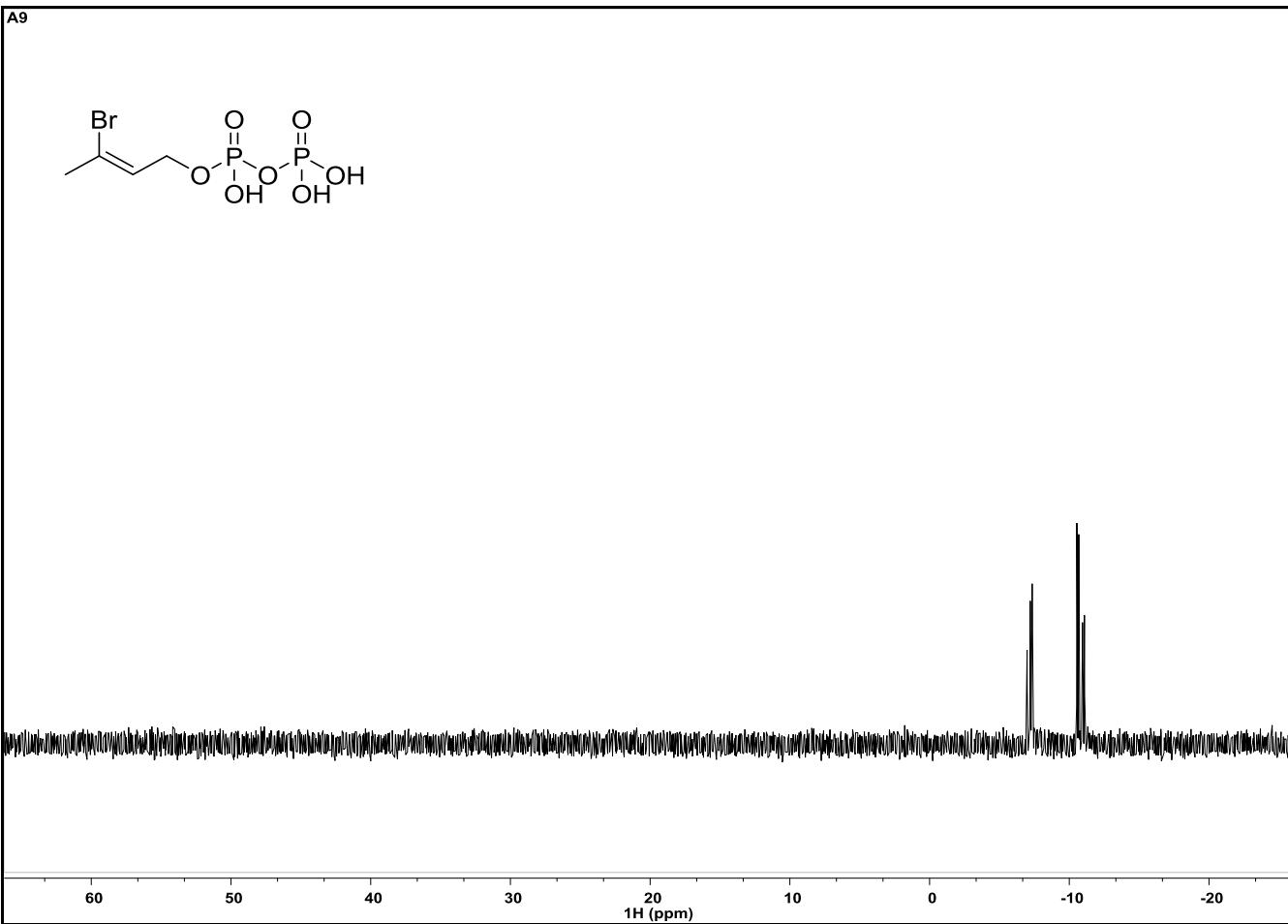


$^{31}\text{P}$  NMR Spectrum (162 MHz) of compound **5** in  $\text{D}_2\text{O}$

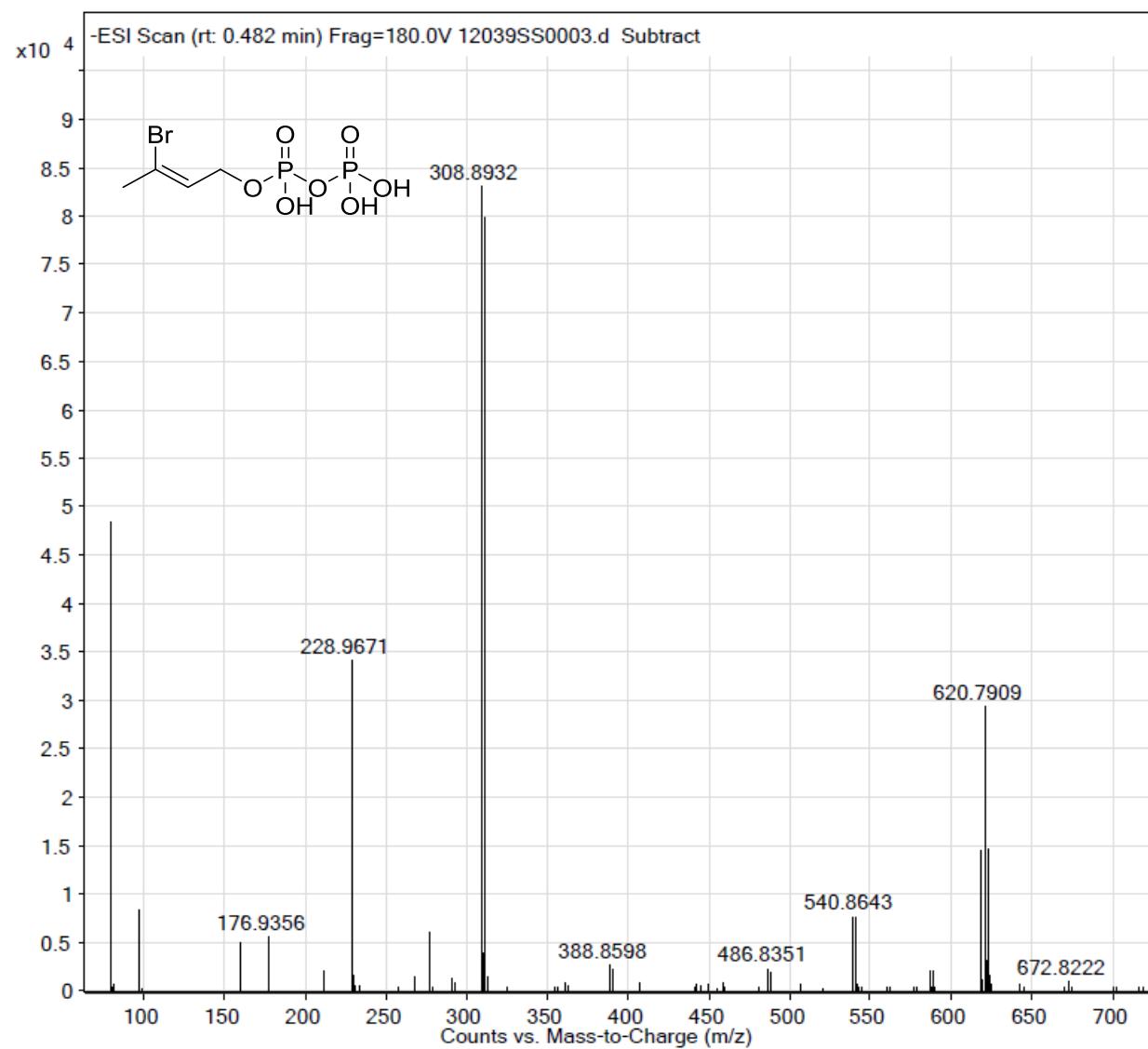




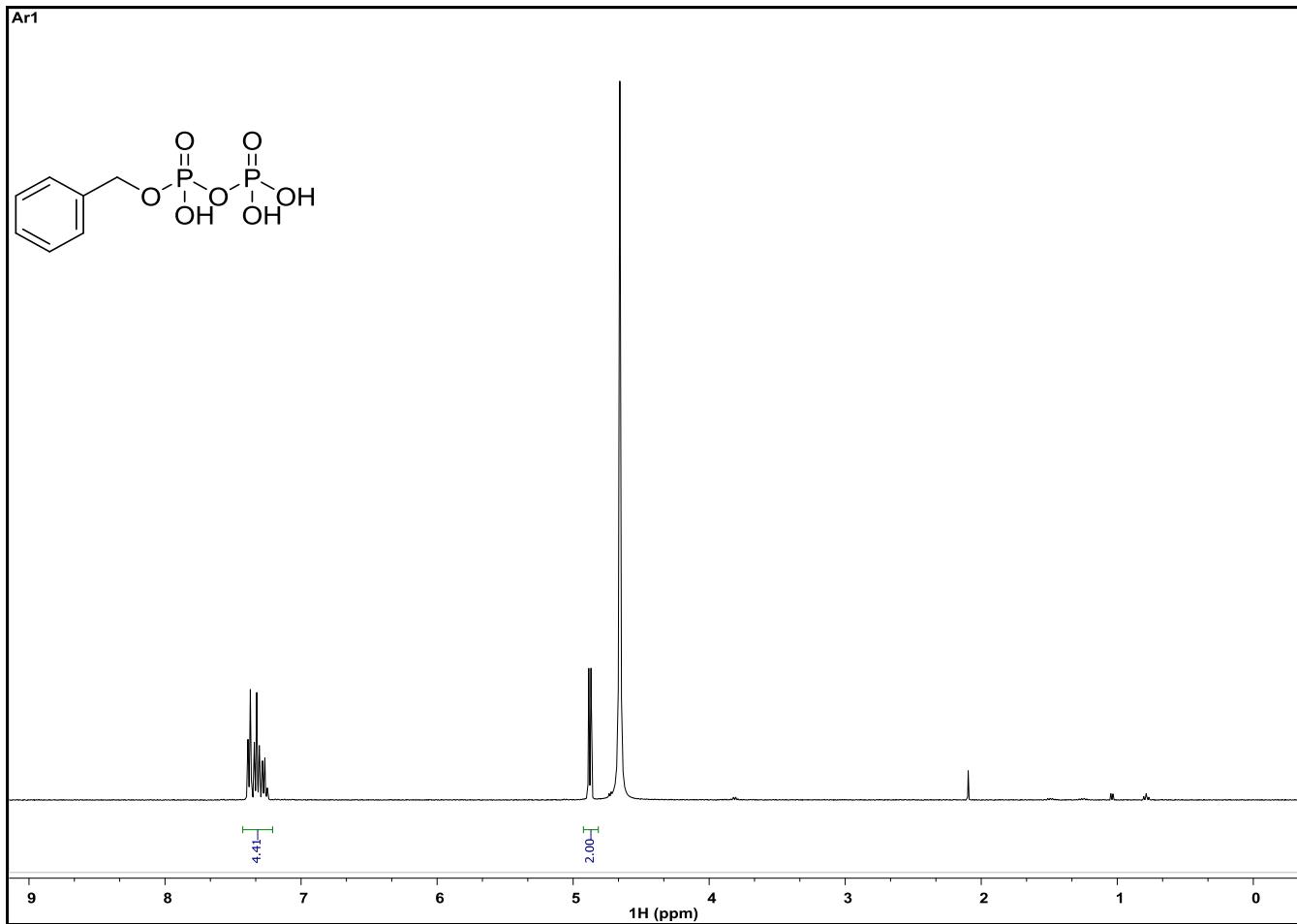
<sup>1</sup>H NMR Spectrum (400 MHz) of compound **9** in D<sub>2</sub>O



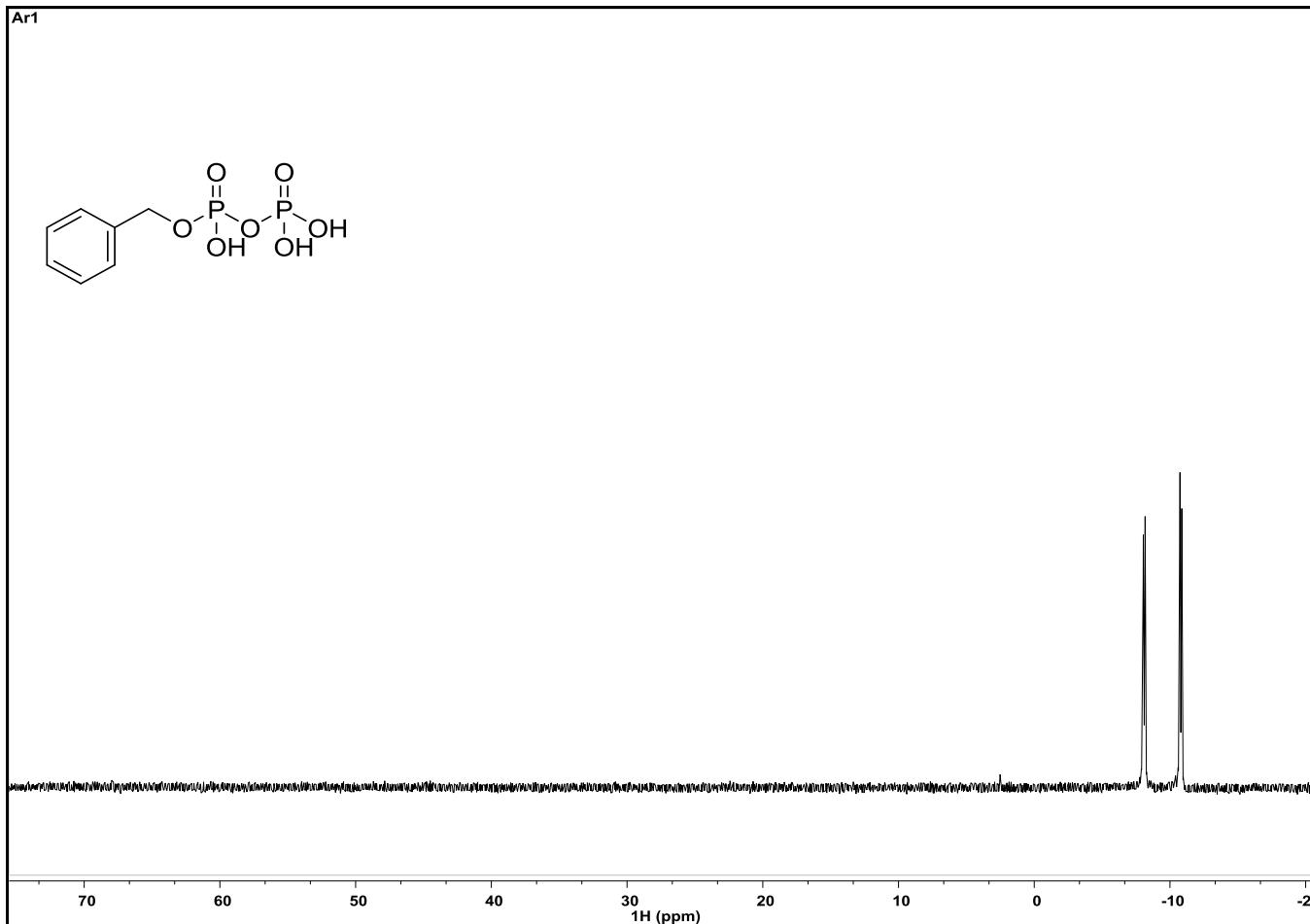
$^{31}\text{P}$  NMR Spectrum (162 MHz) of compound **9** in  $\text{D}_2\text{O}$



HRMS spectrum of **9**

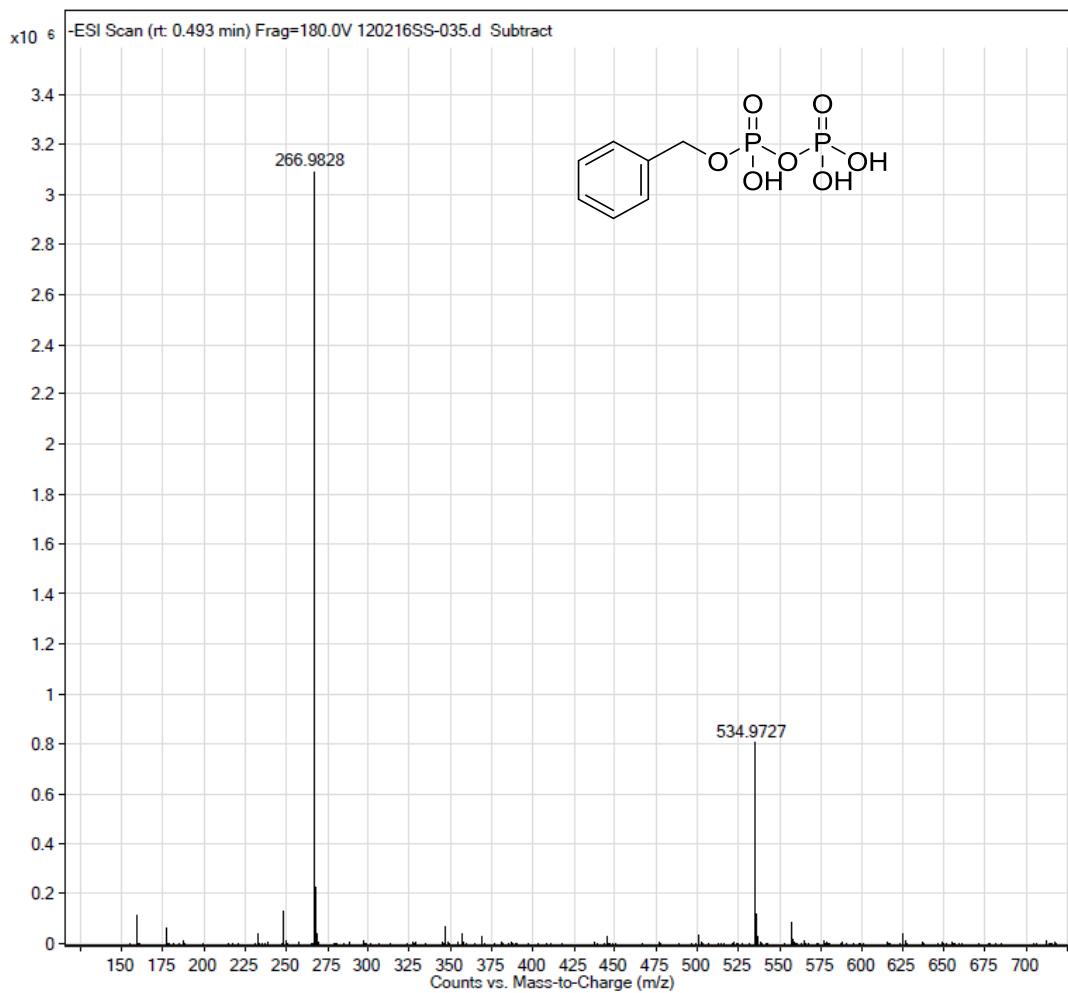


<sup>1</sup>H NMR Spectrum (400 MHz) of compound **22** in D<sub>2</sub>O

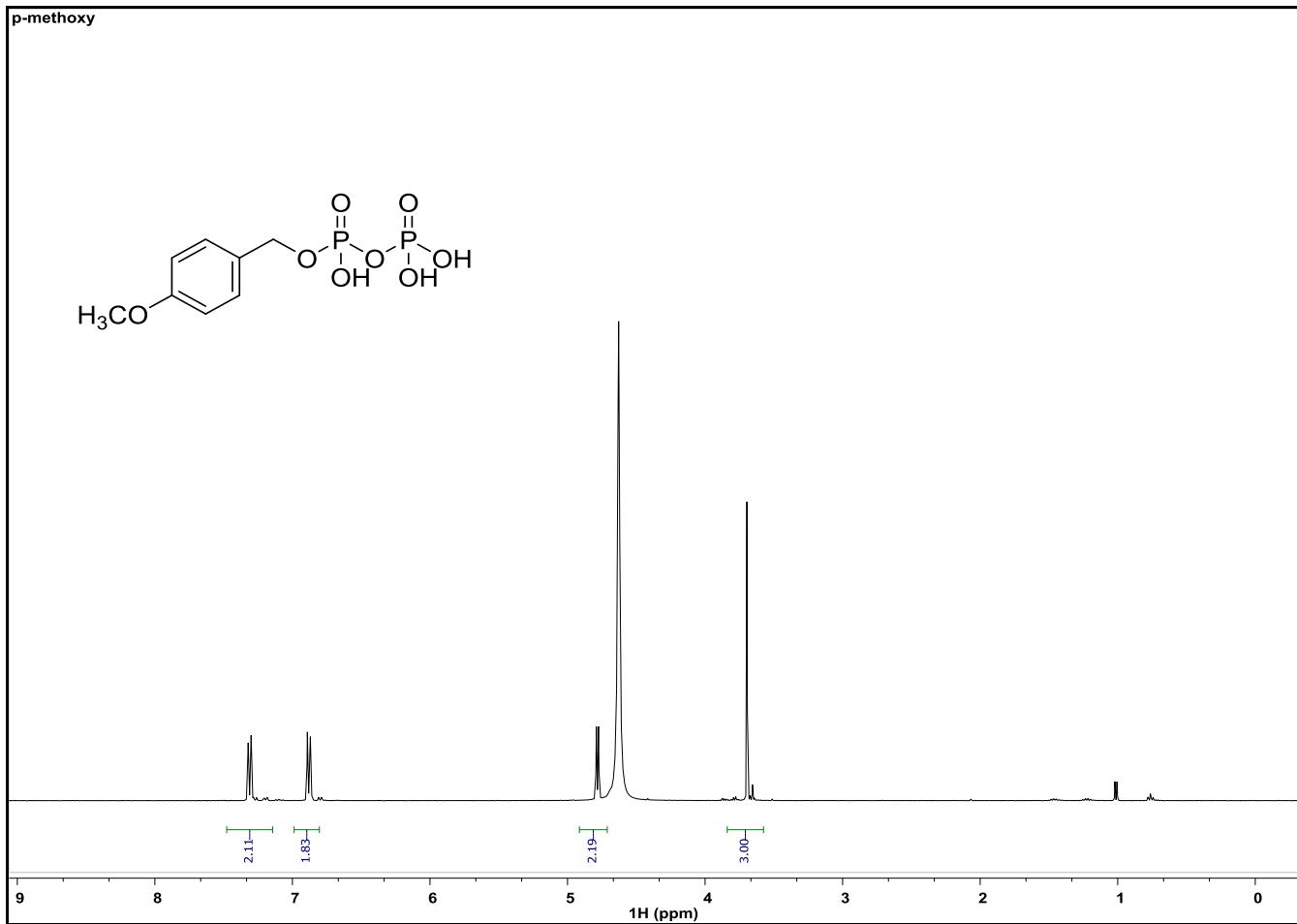


$^{31}\text{P}$  NMR Spectrum (162 MHz) of compound **22** in  $\text{D}_2\text{O}$

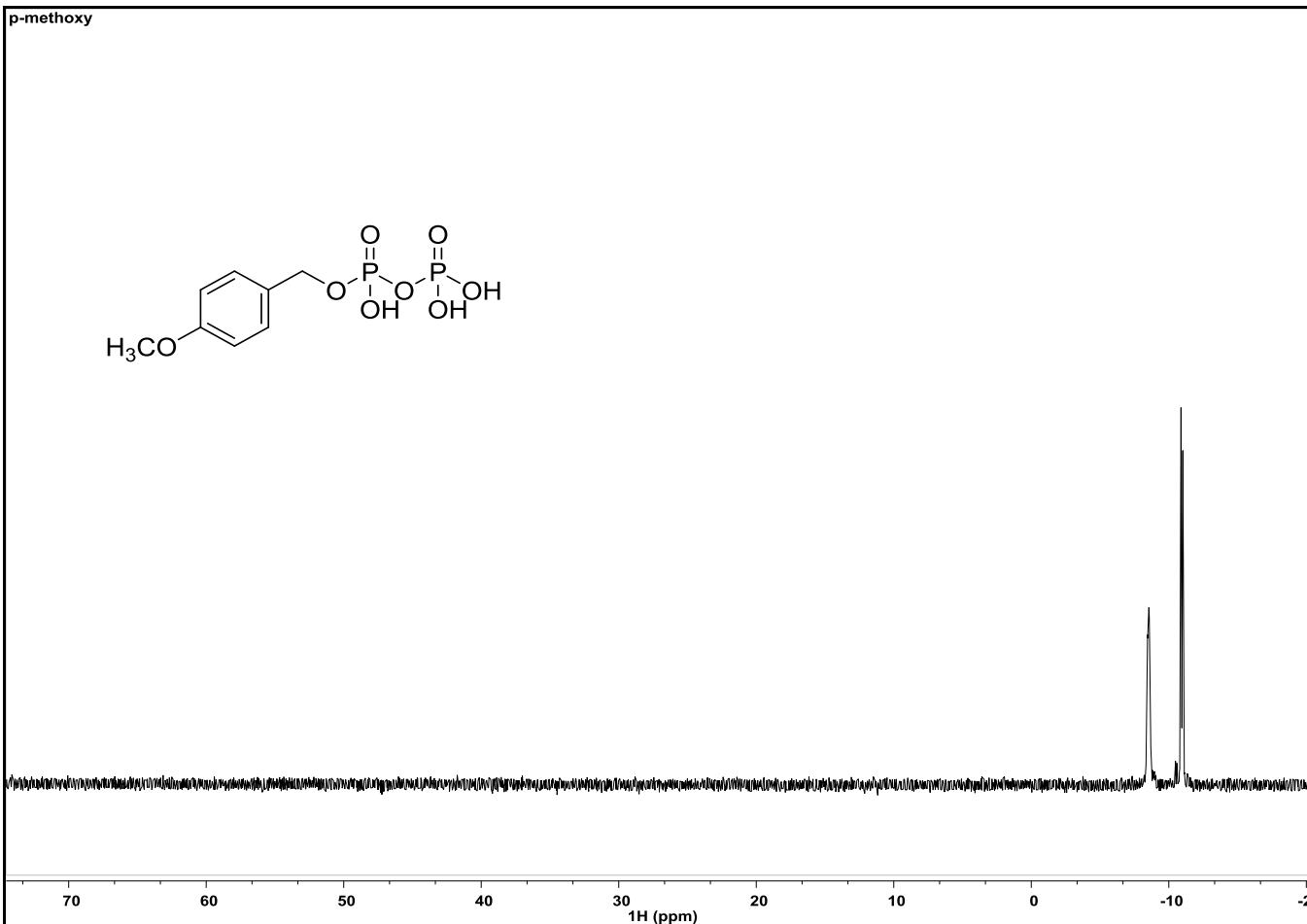
Sample Name	BCS-33	Position	P1-A9	Instrument Name	Instrument 1
User Name		Inj Vol	5	InjPosition	
Sample Type	Sample	IRM Calibration Status	Success	Data Filename	120216SS-035.d
ACQ Method	Neg-Loop-MS.m	Comment		Acquired Time	12/2/2016 6:51:09 PM



HRMS spectrum of **22**

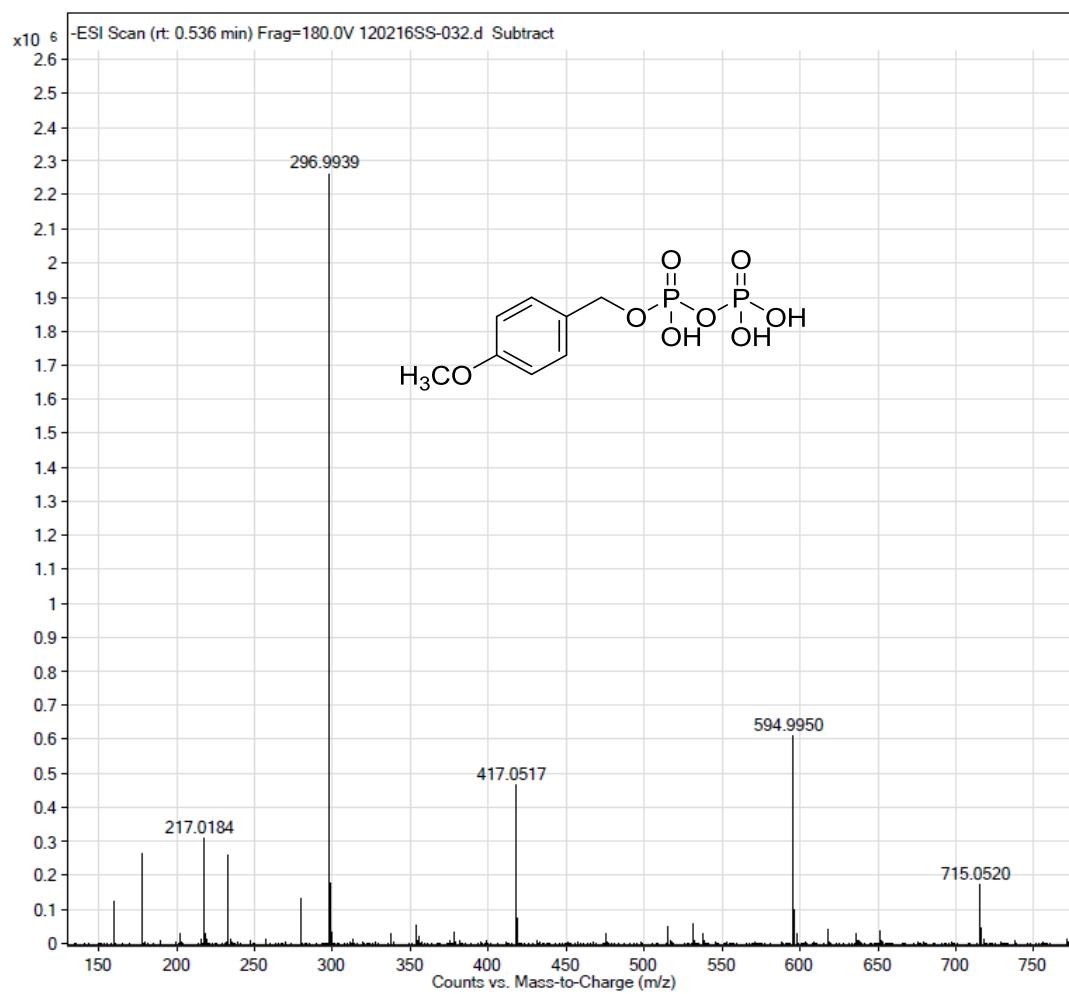


$^1\text{H}$  NMR Spectrum (400 MHz) of compound **23** in  $\text{D}_2\text{O}$

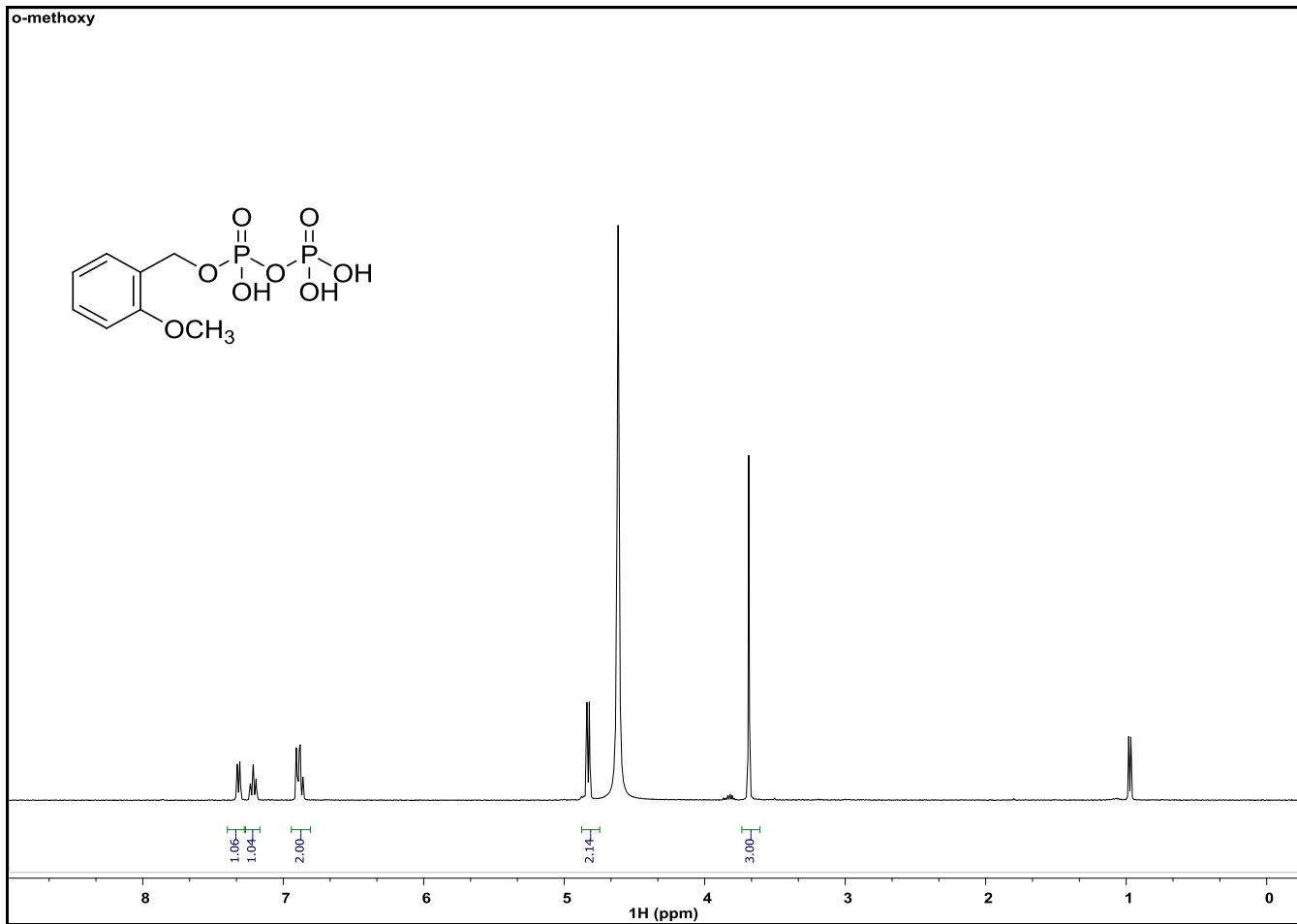


$^{31}\text{P}$  NMR Spectrum (162 MHz) of compound **23** in  $\text{D}_2\text{O}$

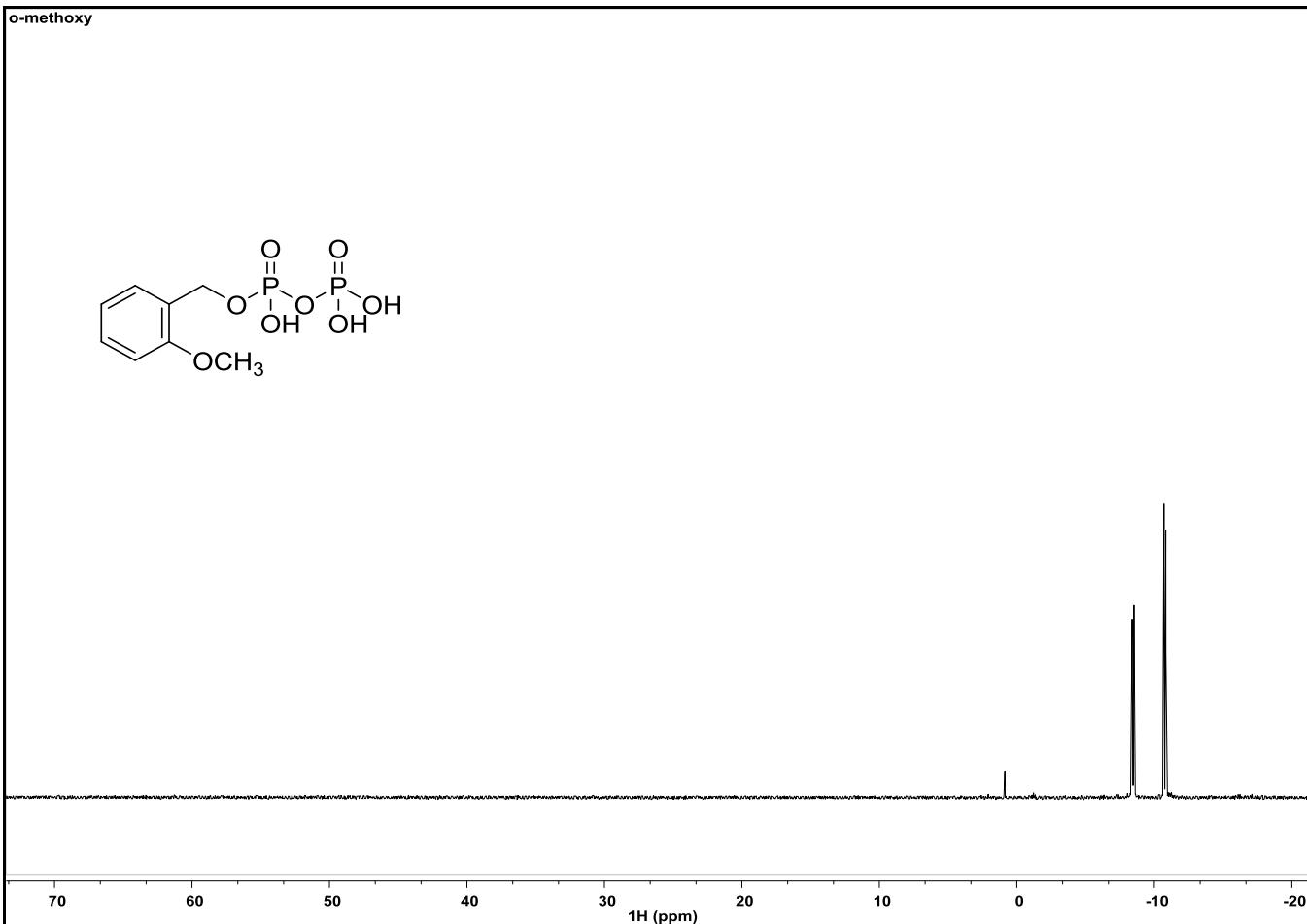
Sample Name	BCS-41	Position	P1-A8	Instrument Name	Instrument 1
User Name		Inj Vol	5	InjPosition	
Sample Type	Sample	IRM Calibration Status	Success	Data Filename	120216SS-032.d
ACQ Method	Neg-Loop-MS.m	Comment		Acquired Time	12/2/2016 6:39:13 PM



HRMS spectrum of **23**



$^1\text{H}$  NMR Spectrum (400 MHz) of compound **24** in  $\text{D}_2\text{O}$



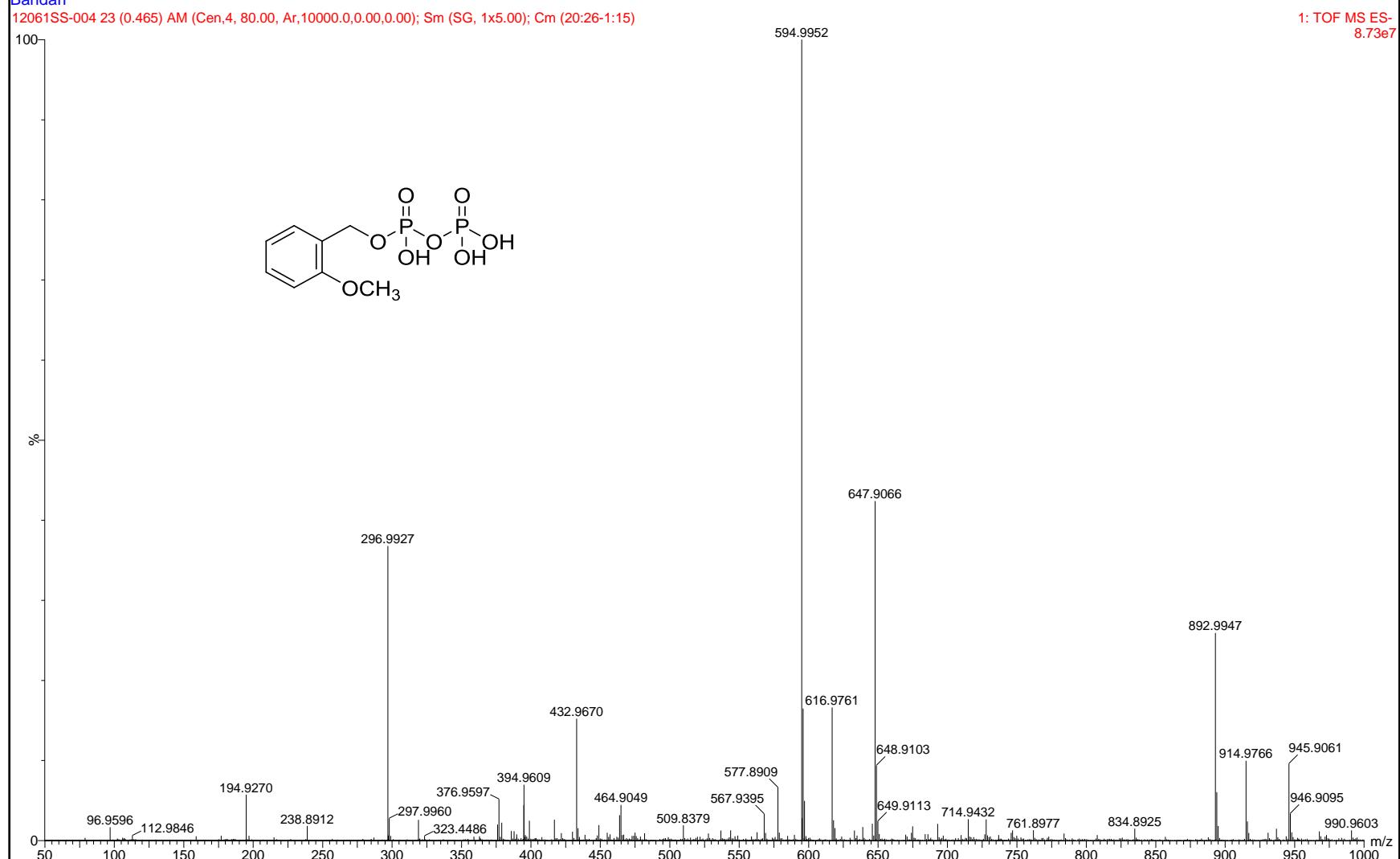
$^{31}\text{P}$  NMR Spectrum (162 MHz) of compound 24 in  $\text{D}_2\text{O}$

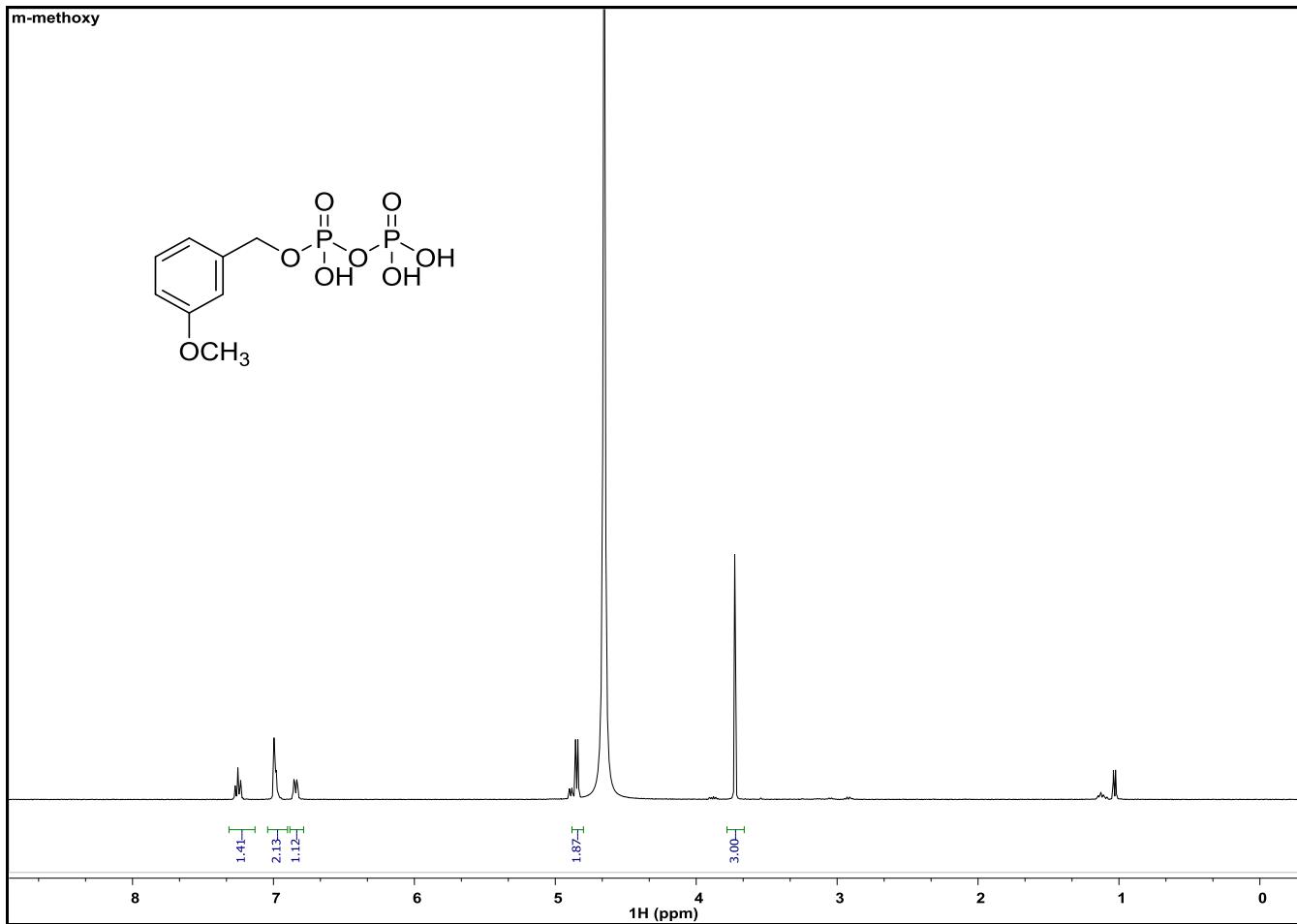
Ar3  
Bandari

SYNAPTG2-Si#UGA589

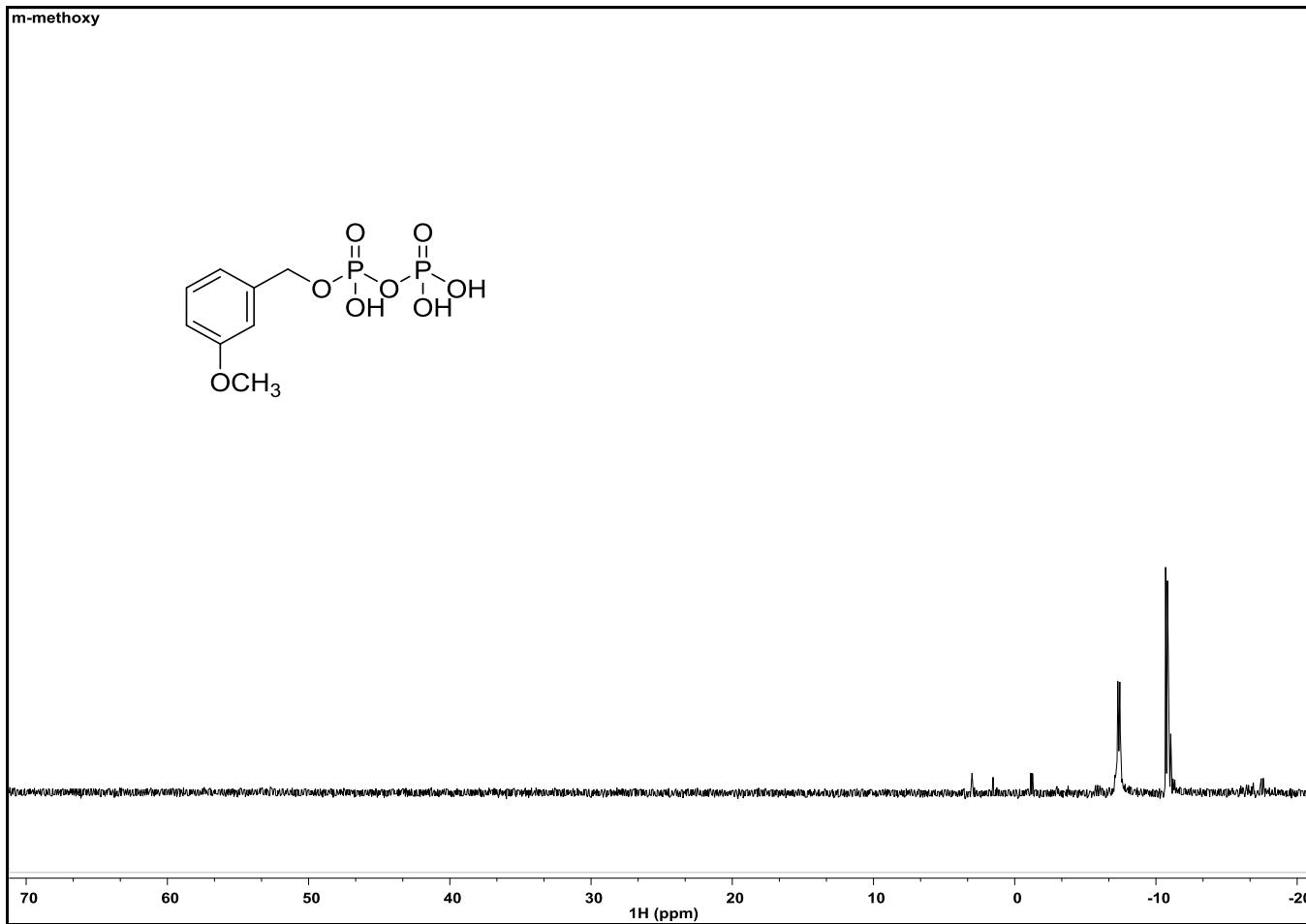
06-Sep-2017 09:45:34

1: TOF MS ES-  
8.73e7





$^1\text{H}$  NMR Spectrum (400 MHz) of compound **25** in  $\text{D}_2\text{O}$



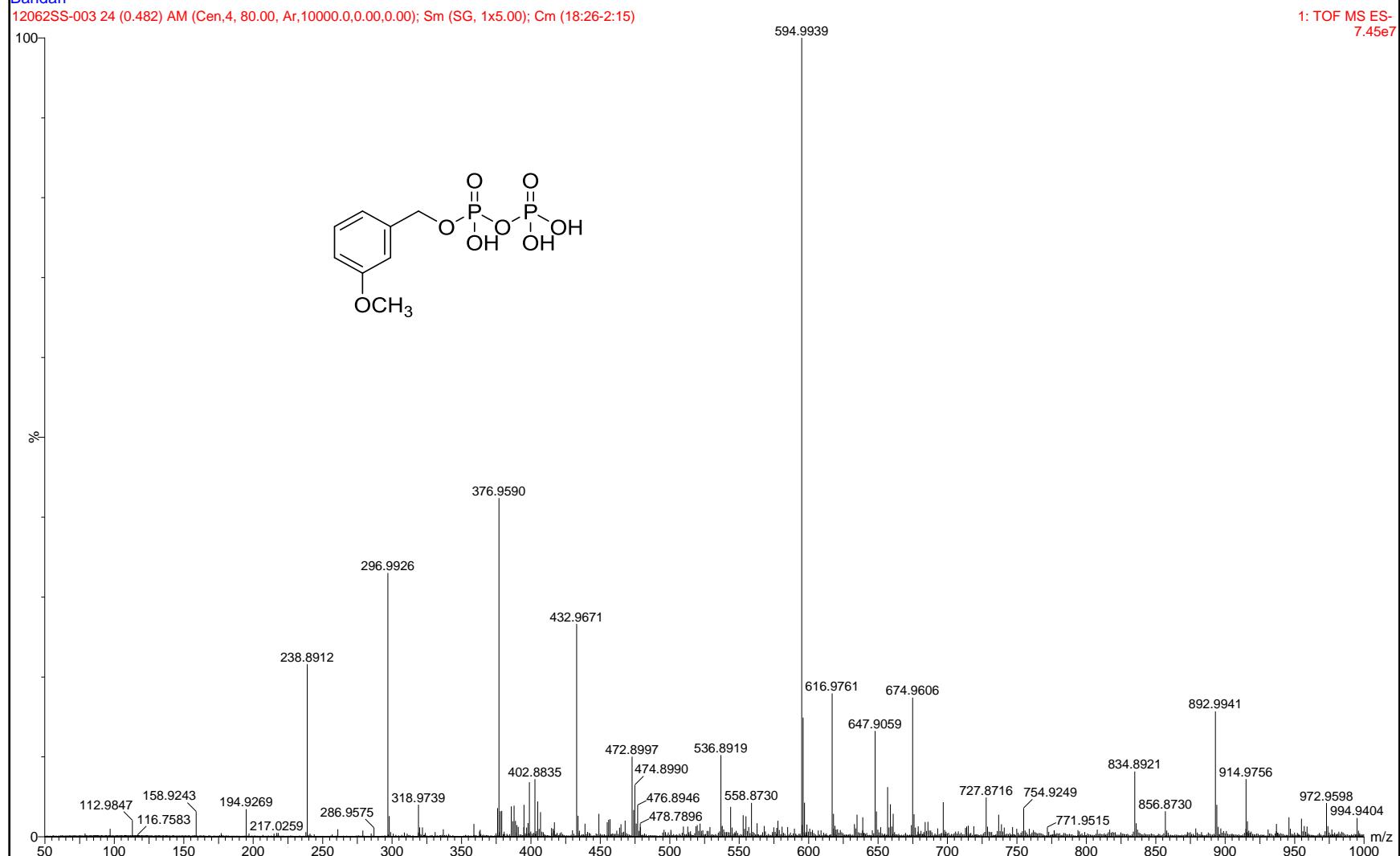
<sup>31</sup>P NMR Spectrum (162 MHz) of compound **25** in D<sub>2</sub>O

Ar4  
Bandari

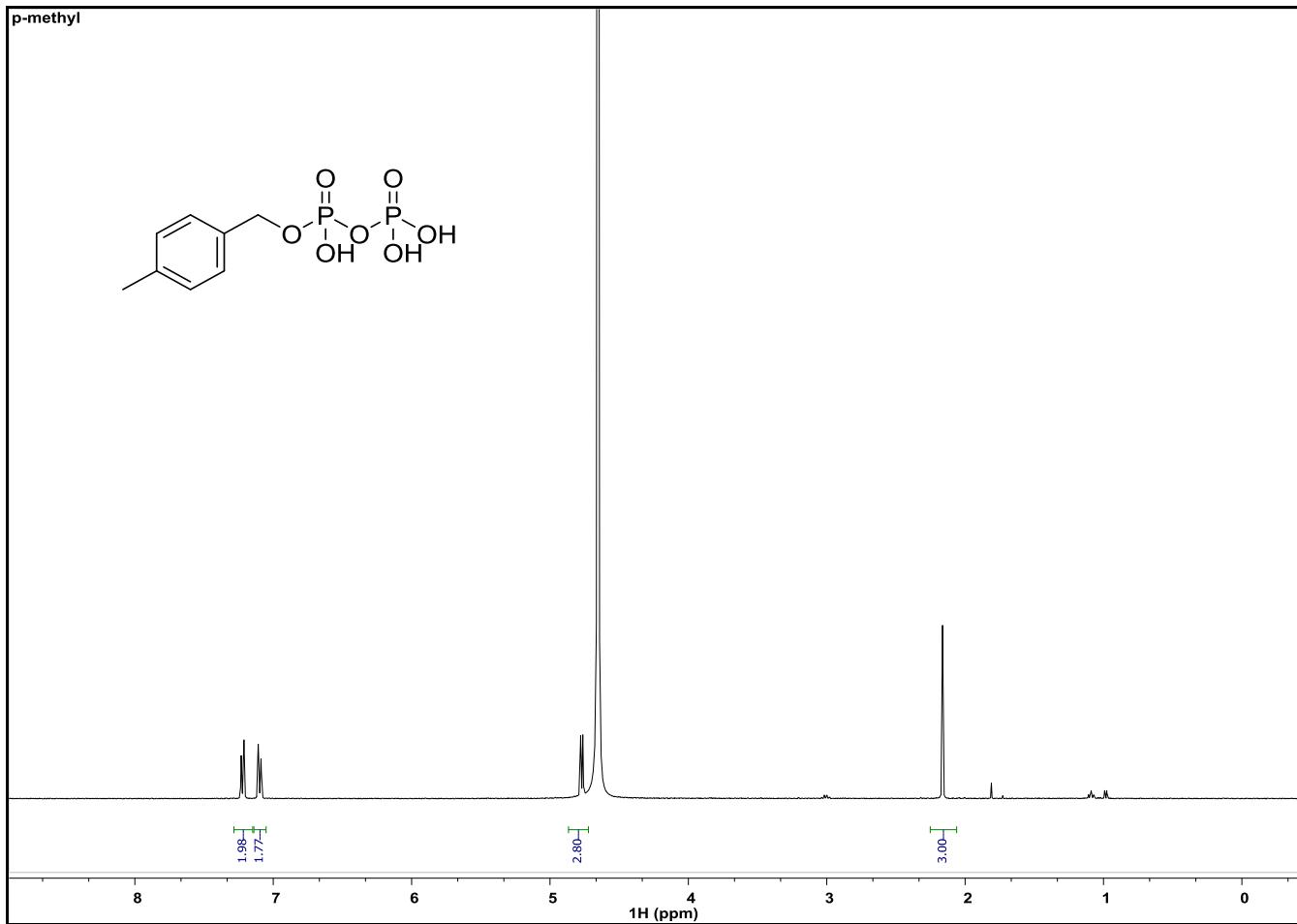
SYNAPTG2-Si#UGA589

06-Sep-2017 10:01:43

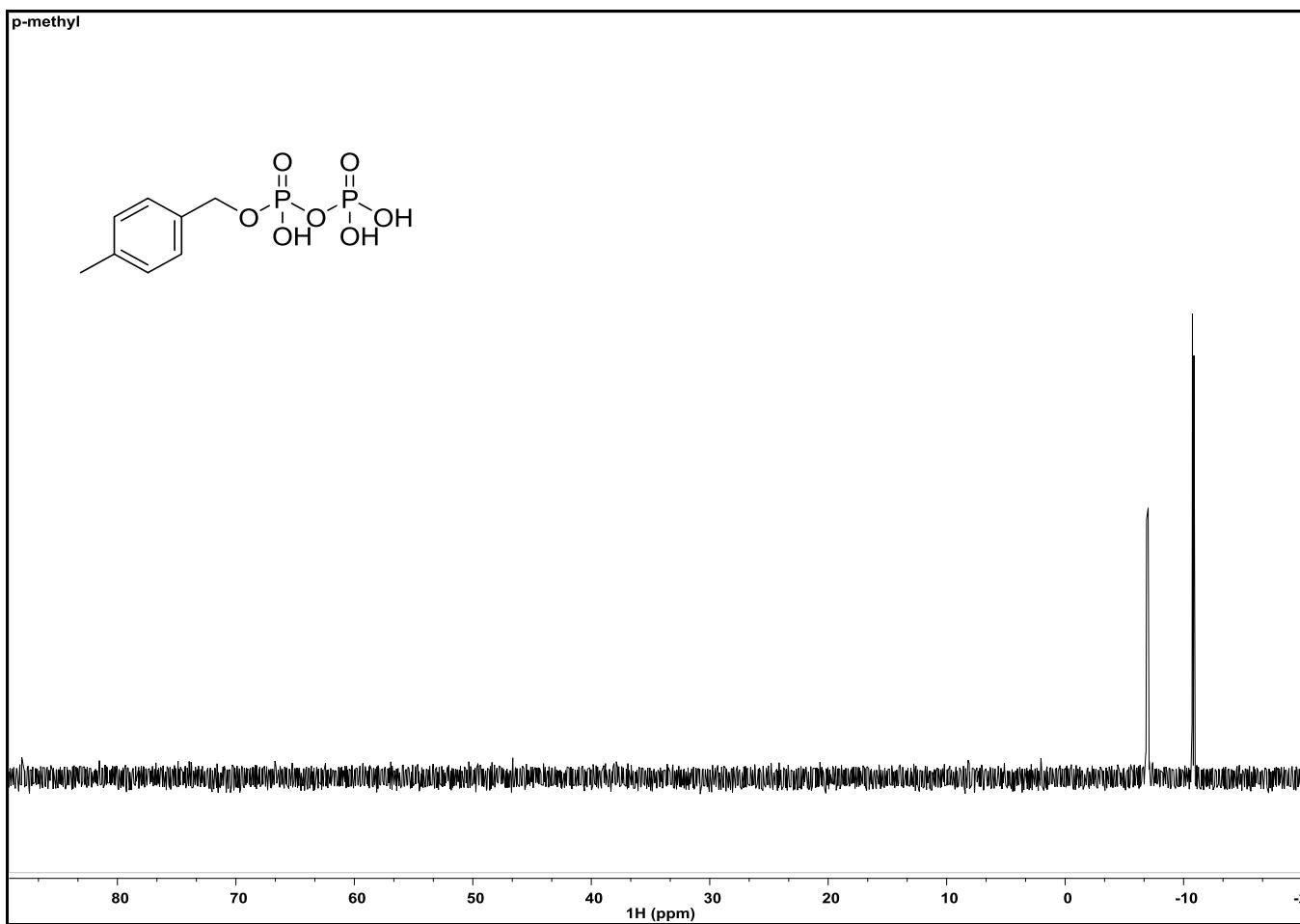
1: TOF MS ES-  
7.45e7



HRMS spectrum of 25



<sup>1</sup>H NMR Spectrum (400 MHz) of compound **26** in D<sub>2</sub>O



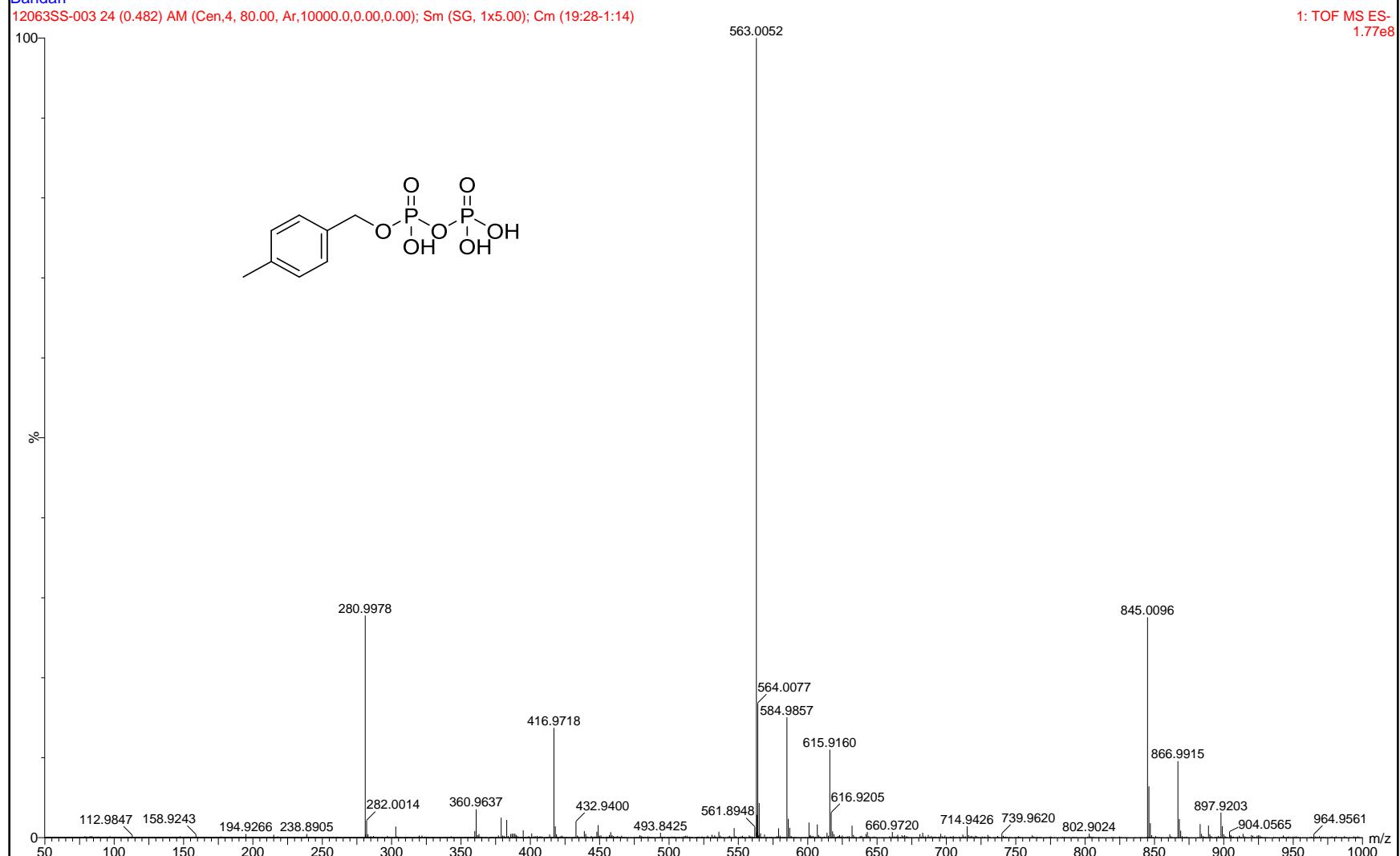
$^{31}\text{P}$  NMR Spectrum (162 MHz) of compound **26** in  $\text{D}_2\text{O}$

Ar5  
Bandari

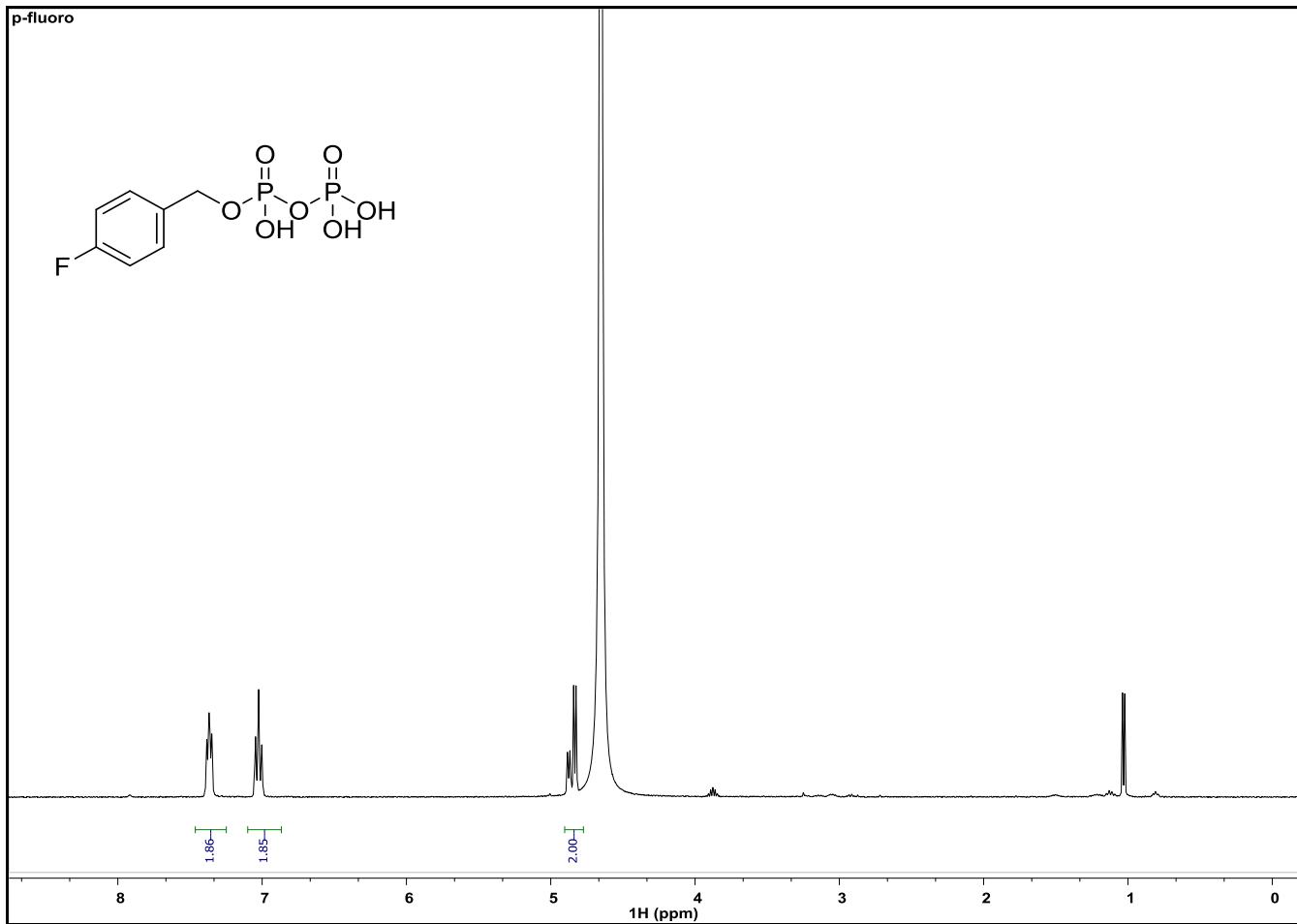
SYNAPTG2-Si#UGA589

06-Sep-2017 10:17:54

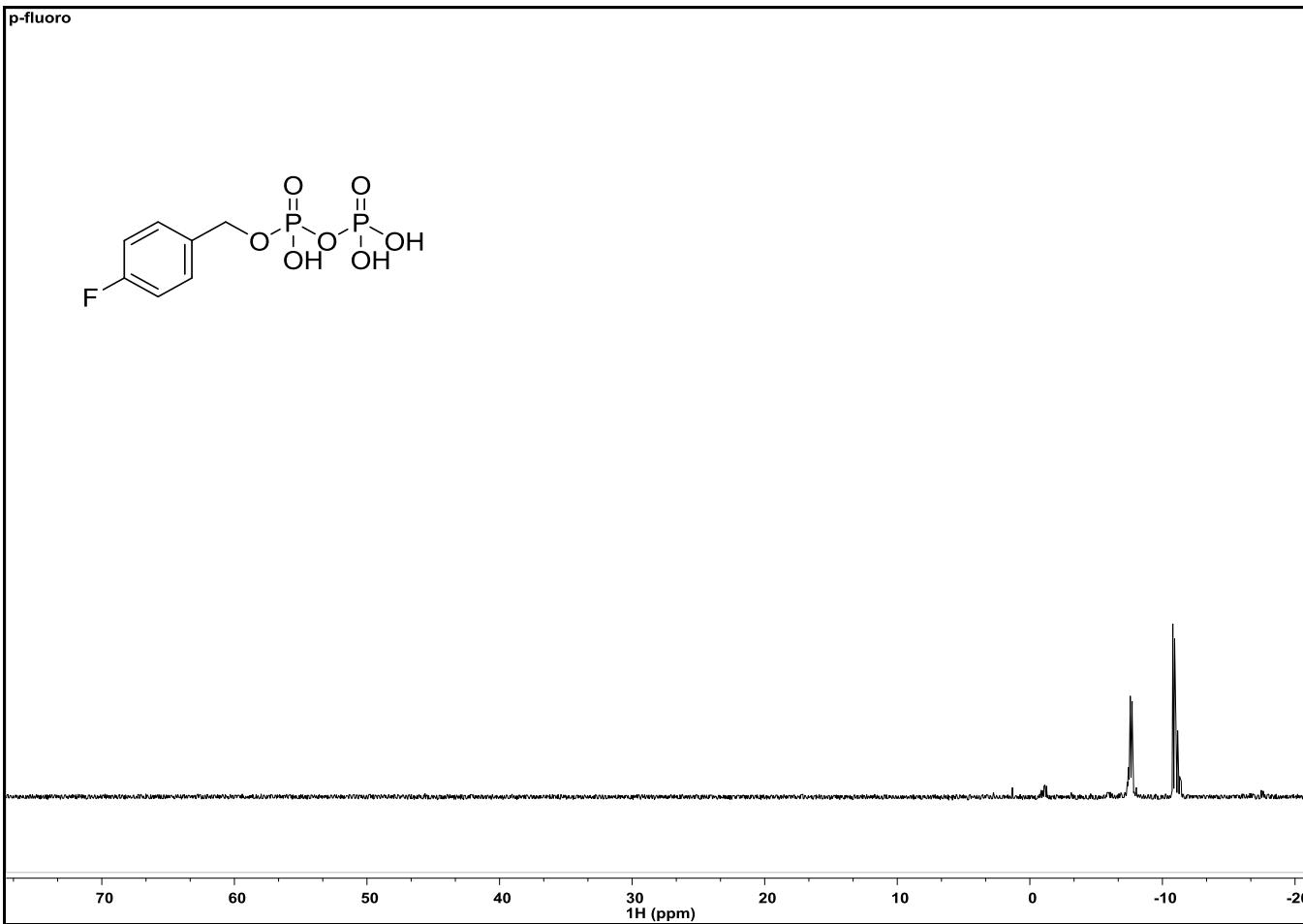
1: TOF MS ES-  
1.77e8



HRMS spectrum of **26**



$^1\text{H}$  NMR Spectrum (400 MHz) of compound **27** in  $\text{D}_2\text{O}$



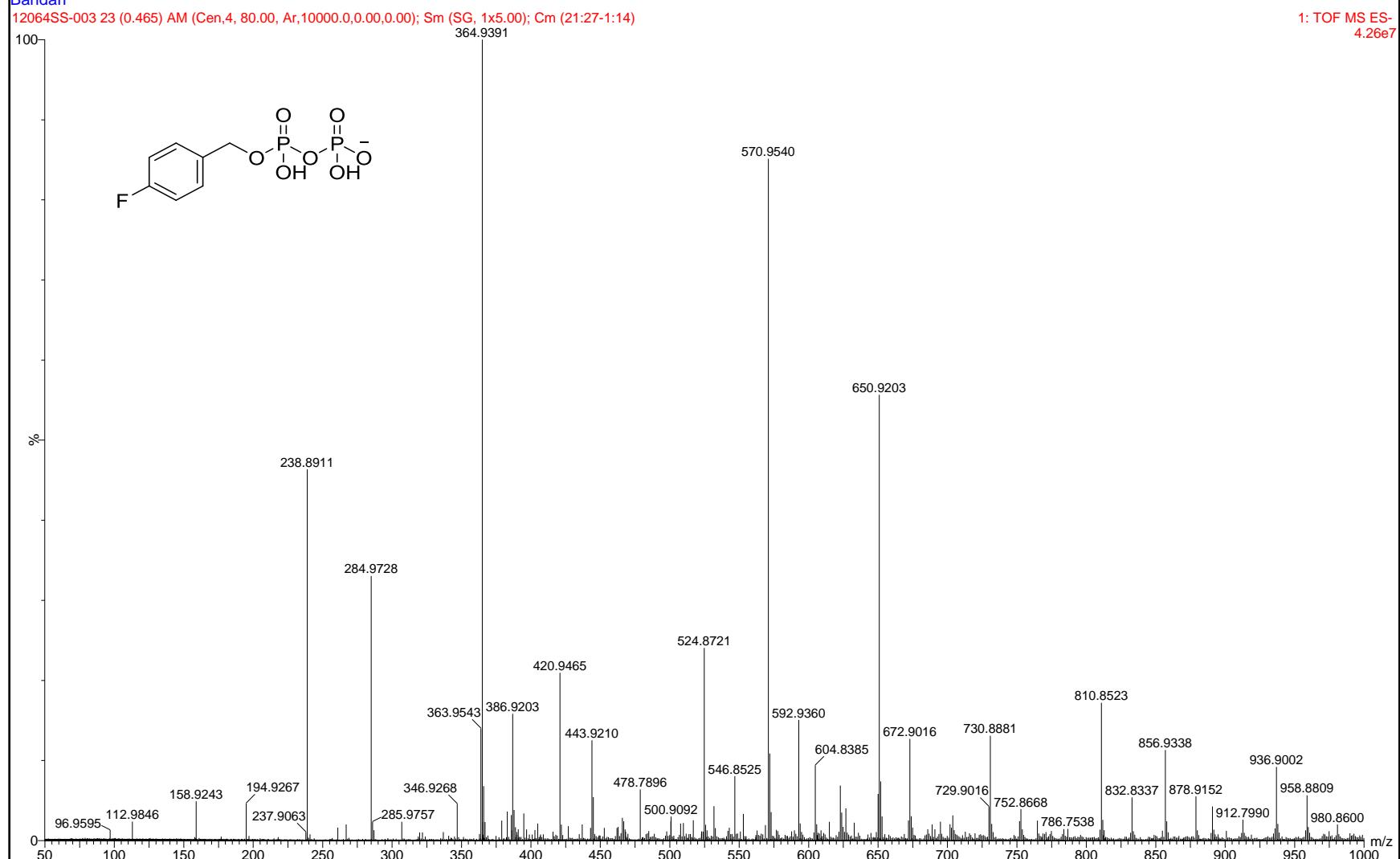
$^{31}\text{P}$  NMR Spectrum (162 MHz) of compound **27** in  $\text{D}_2\text{O}$

Ar6  
Bandari

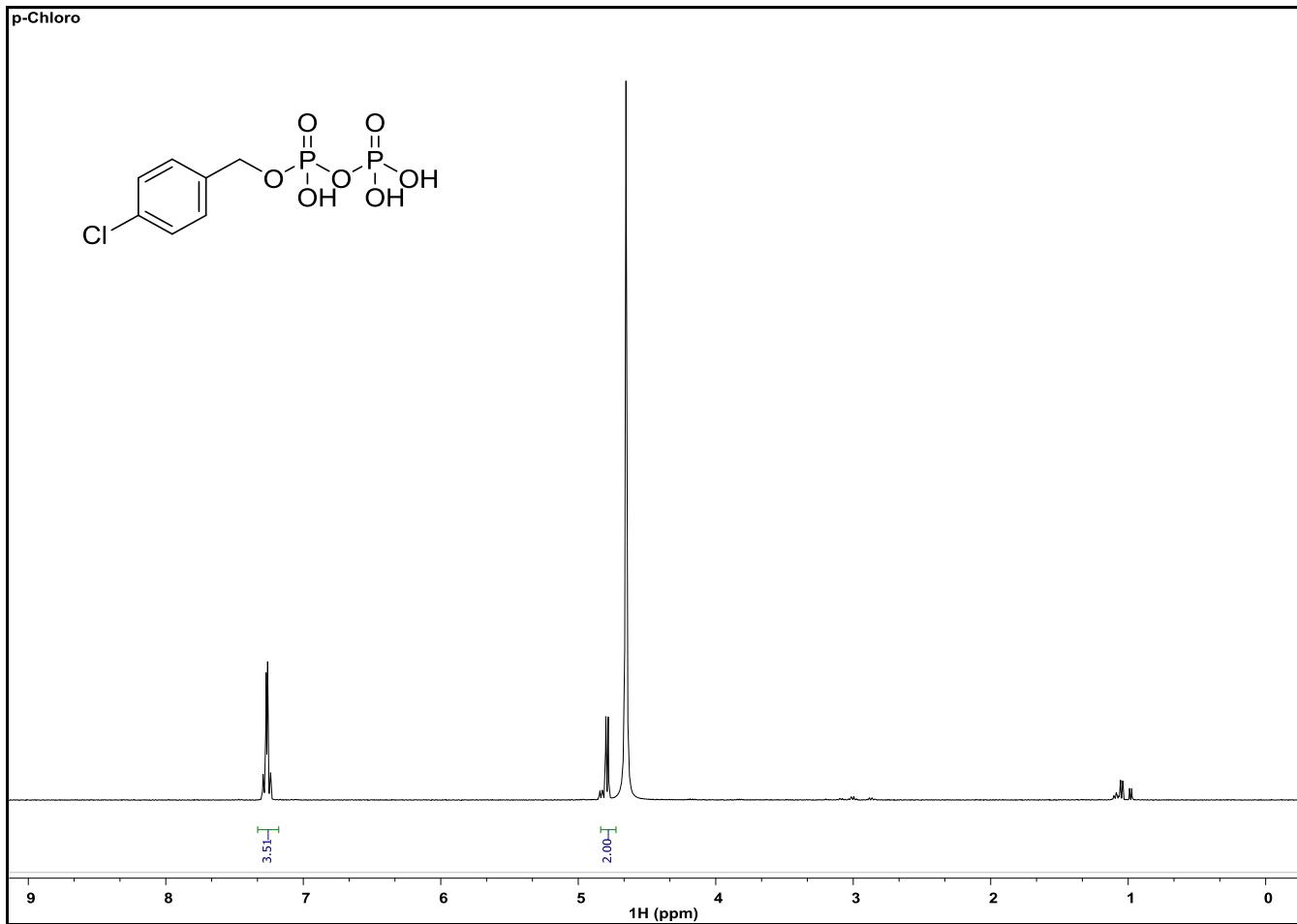
SYNAPTG2-Si#UGA589

06-Sep-2017 10:34:00

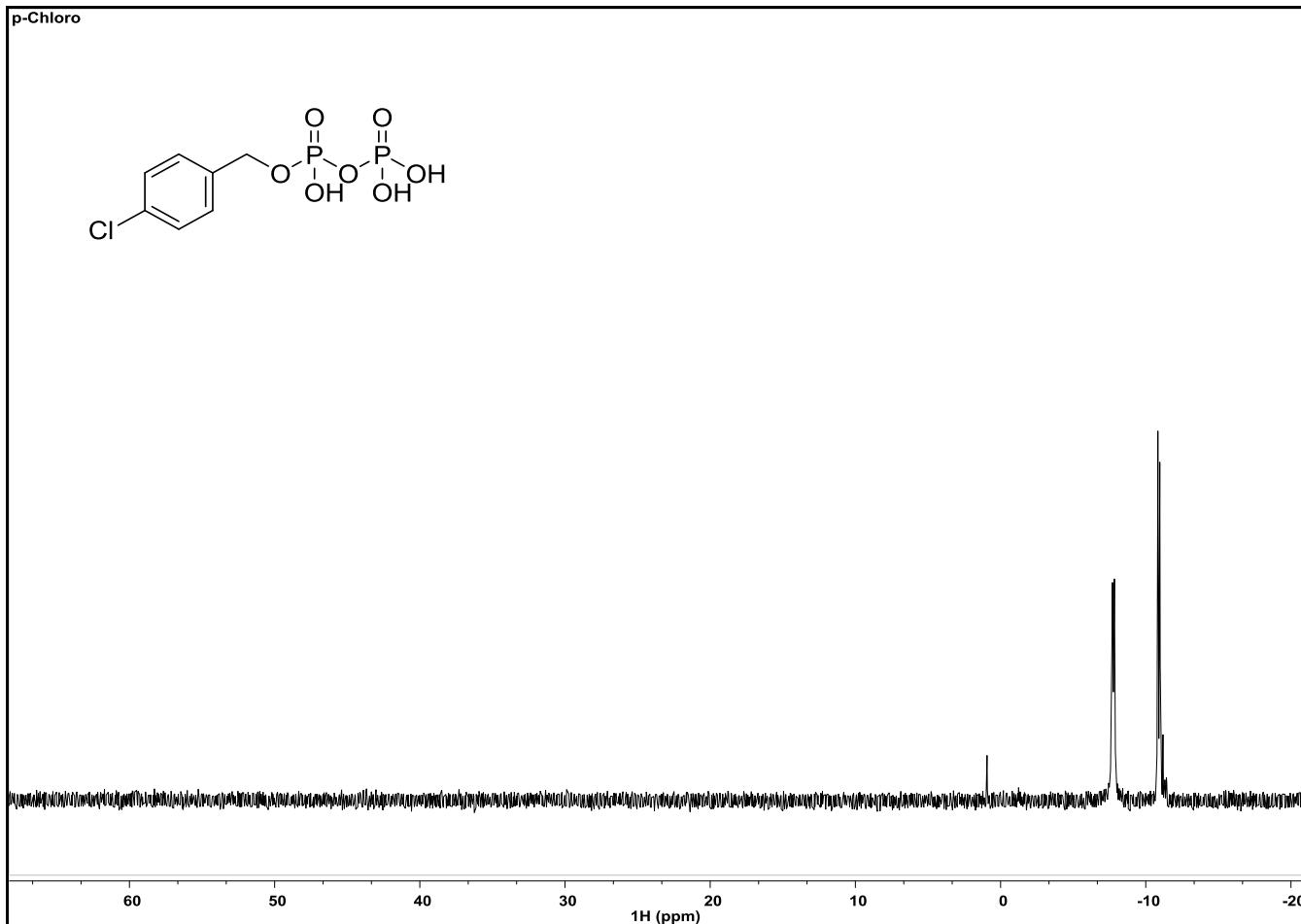
1: TOF MS ES-  
4.26e7



HRMS spectrum of 27



$^1\text{H}$  NMR Spectrum (400 MHz) of compound **28** in  $\text{D}_2\text{O}$

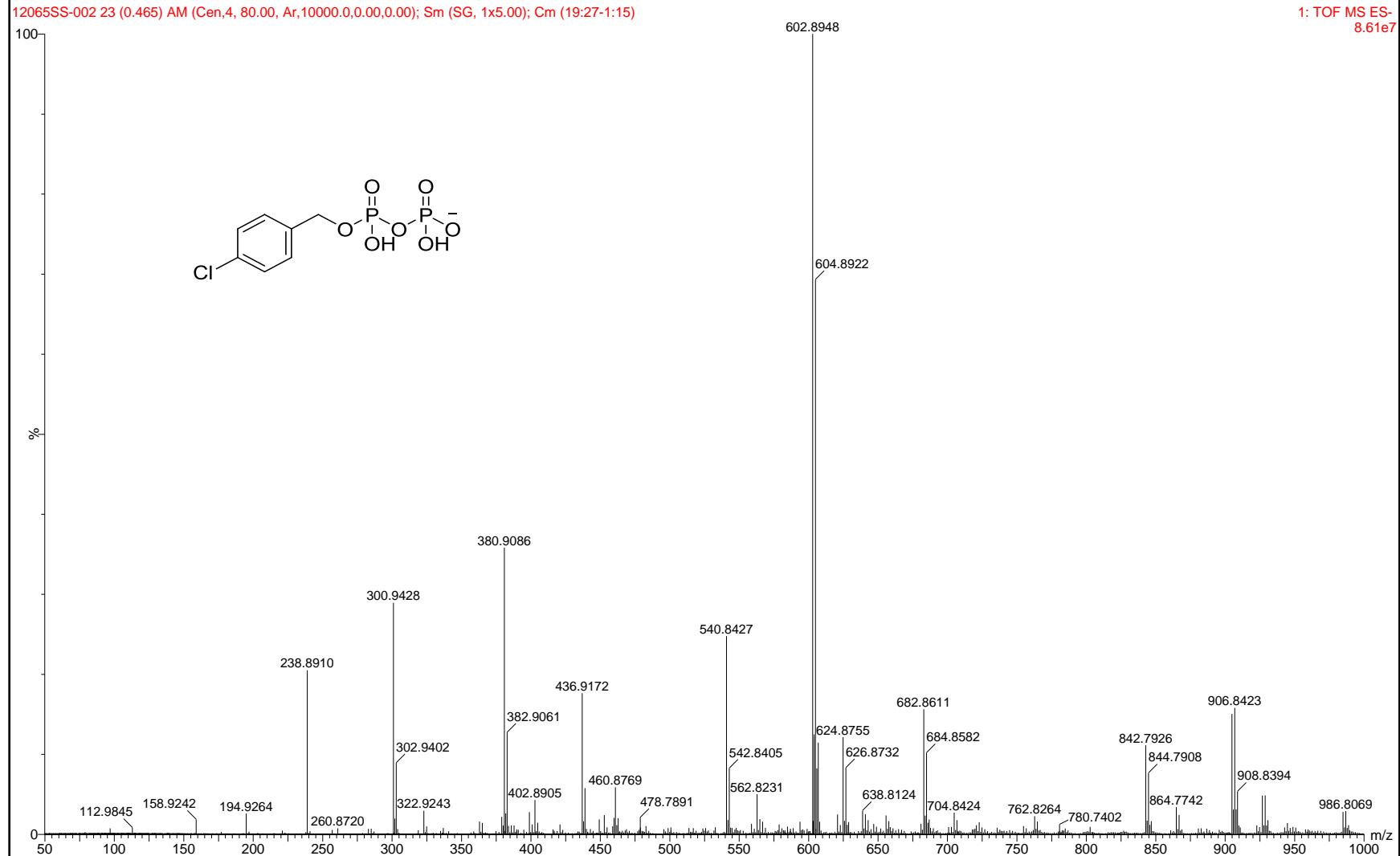


Ar7  
Bandari

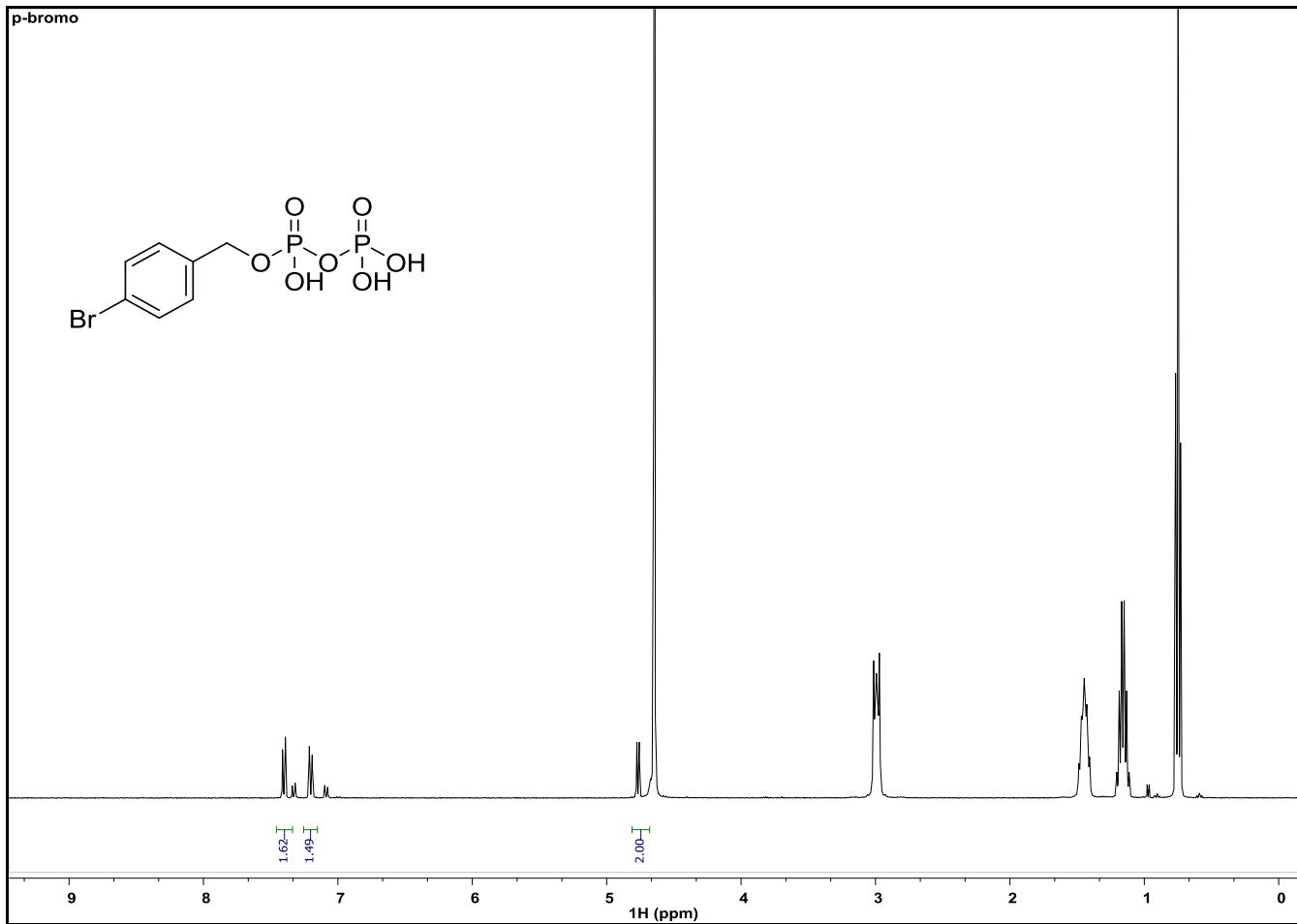
SYNAPTG2-Si#UGA589

06-Sep-2017 10:46:05

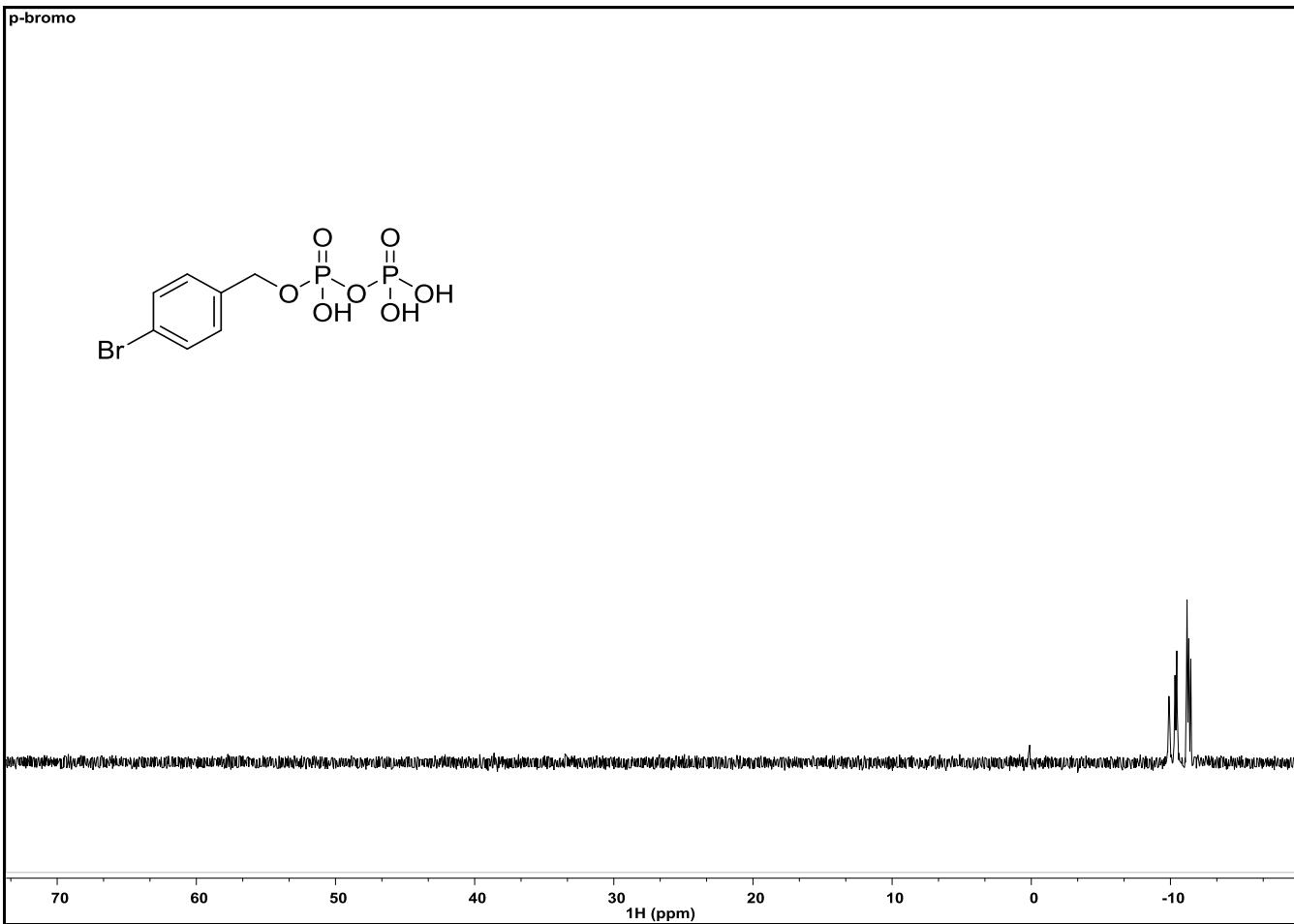
1: TOF MS ES-  
8.61e7



HRMS spectrum of **28**



$^1\text{H}$  NMR Spectrum (400 MHz) of compound **29** in  $\text{D}_2\text{O}$



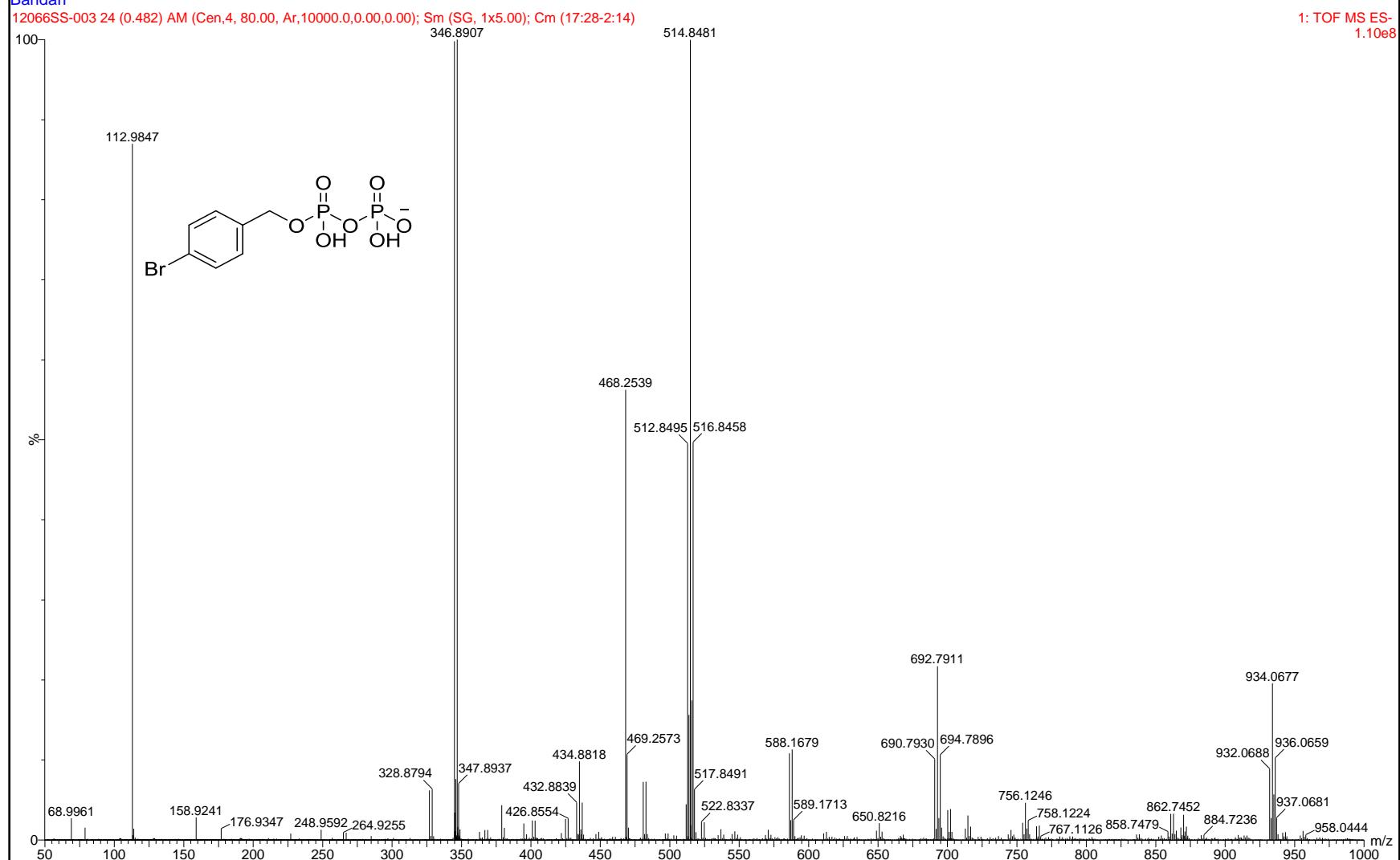
<sup>31</sup>P NMR Spectrum (162 MHz) of compound **29** in D<sub>2</sub>O

Ar8  
Bandari

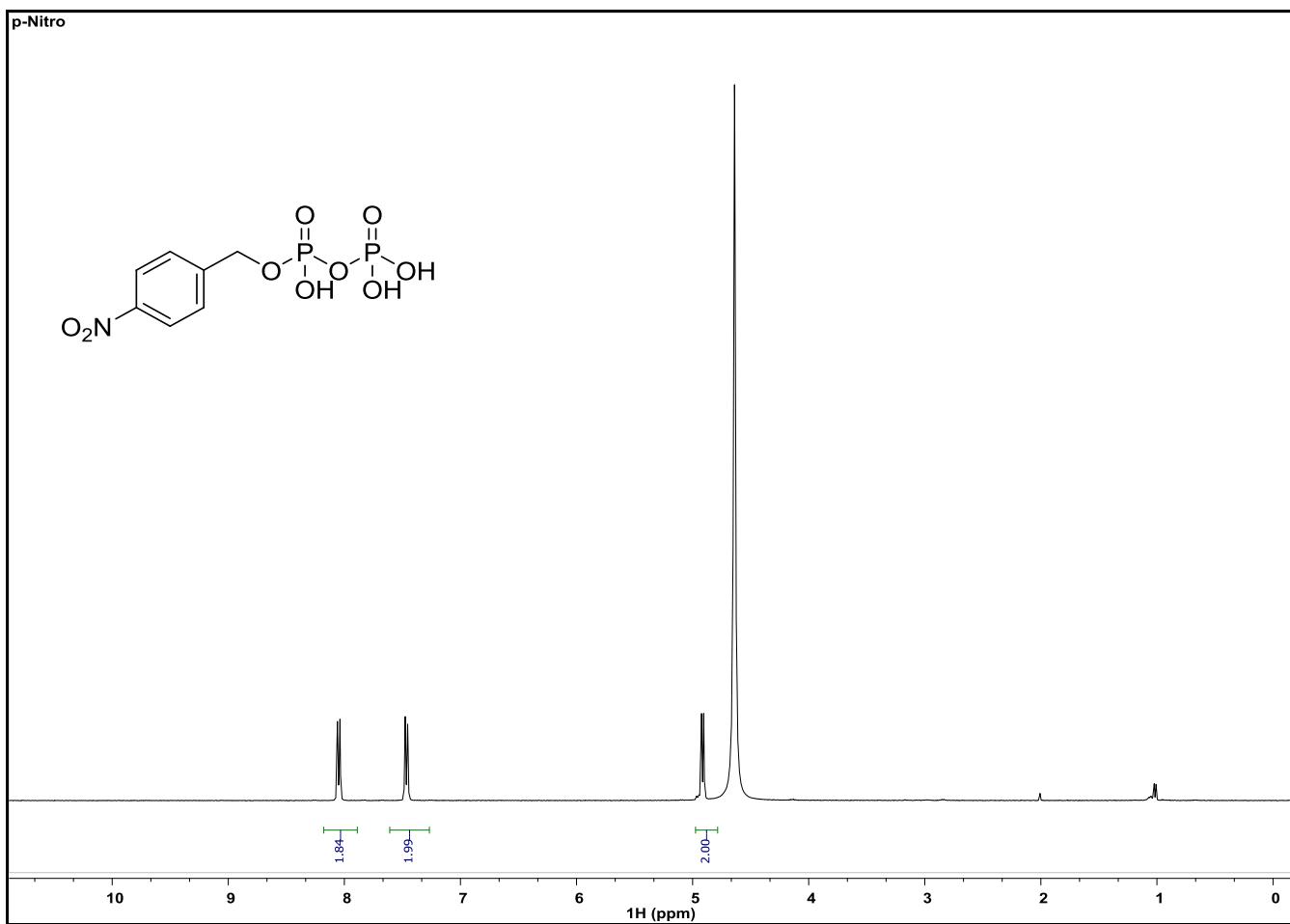
SYNAPTG2-Si#UGA589

06-Sep-2017 11:06:07

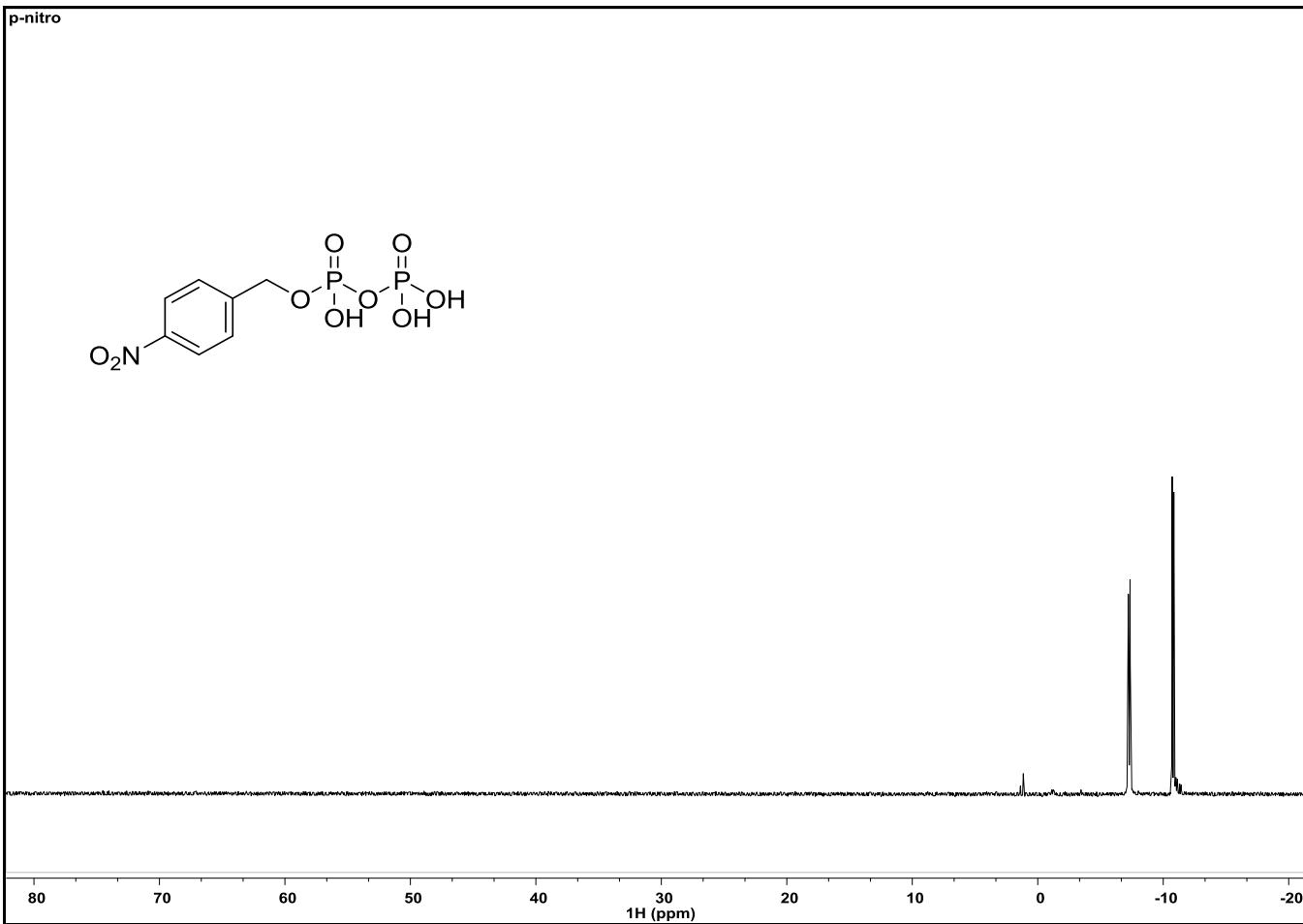
1: TOF MS ES-  
1.10e8



HRMS spectrum of 29



$^1\text{H}$  NMR Spectrum (400 MHz) of compound **30** in  $\text{D}_2\text{O}$



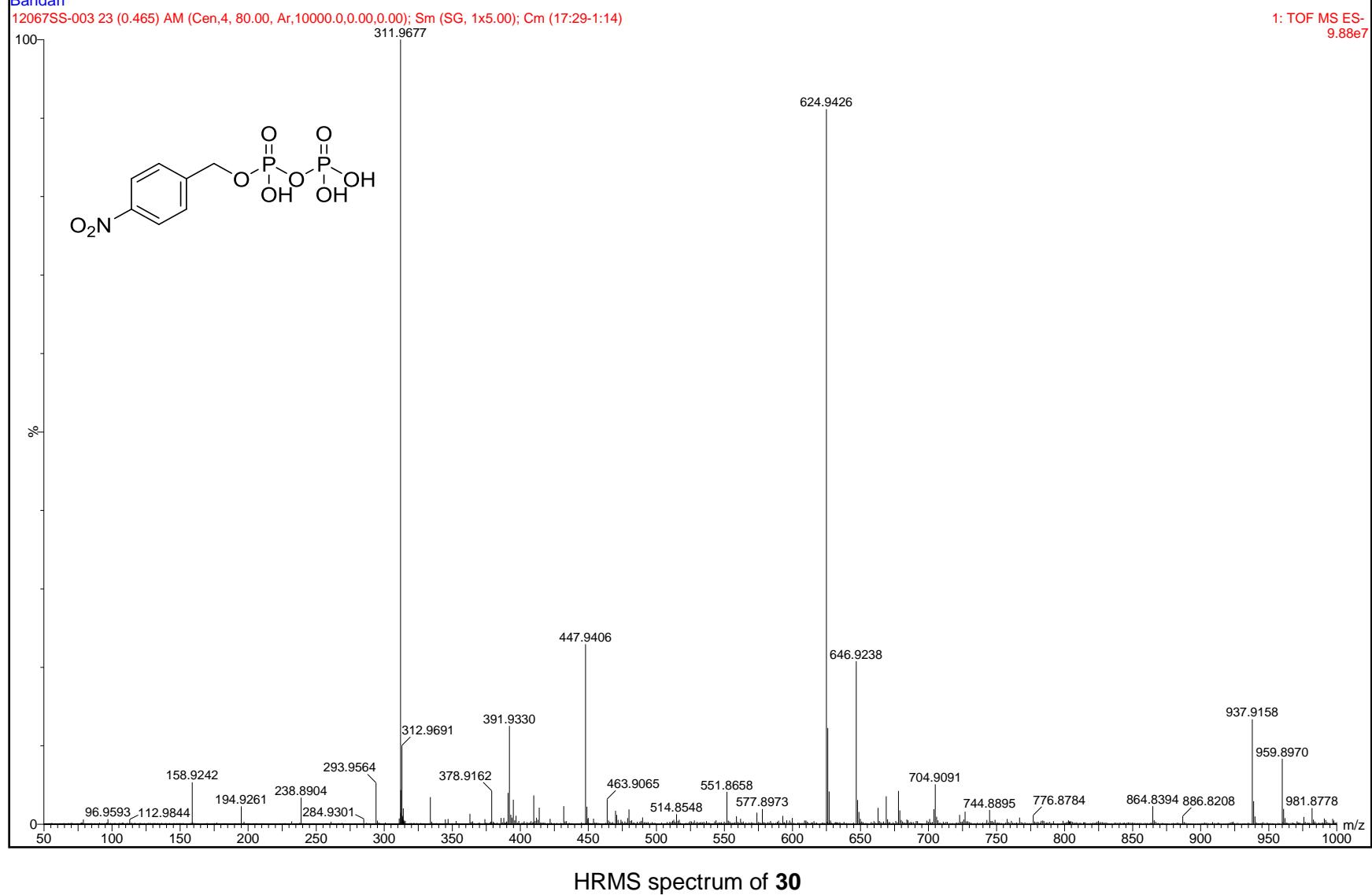
$^{31}\text{P}$  NMR Spectrum (162 MHz) of compound **30** in  $\text{D}_2\text{O}$

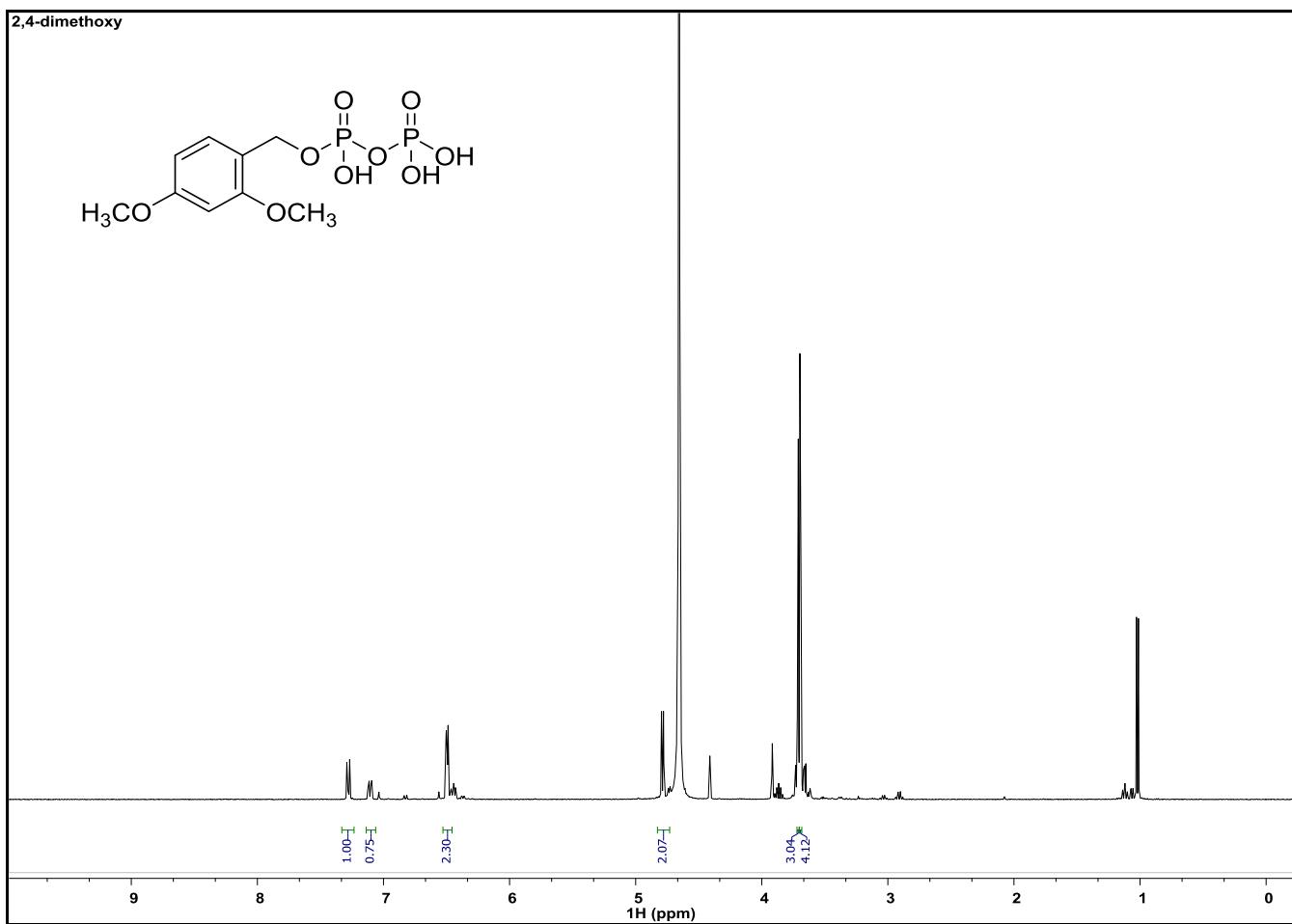
Ar9  
Bandari

SYNAPTG2-Si#UGA589

06-Sep-2017 11:22:11

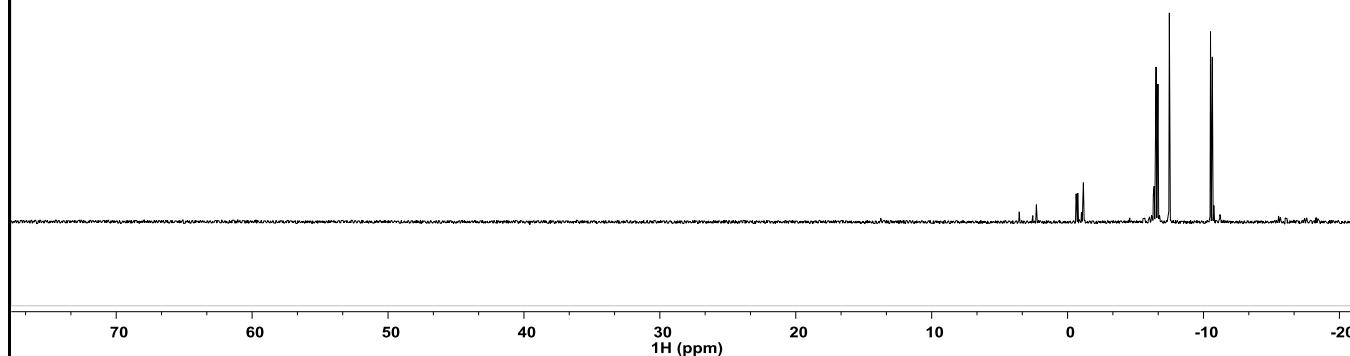
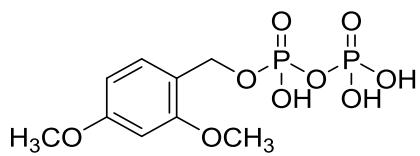
1: TOF MS ES-  
9.88e7





<sup>1</sup>H NMR Spectrum (400 MHz) of compound **31** in  $D_2O$

2,4-dimethoxy



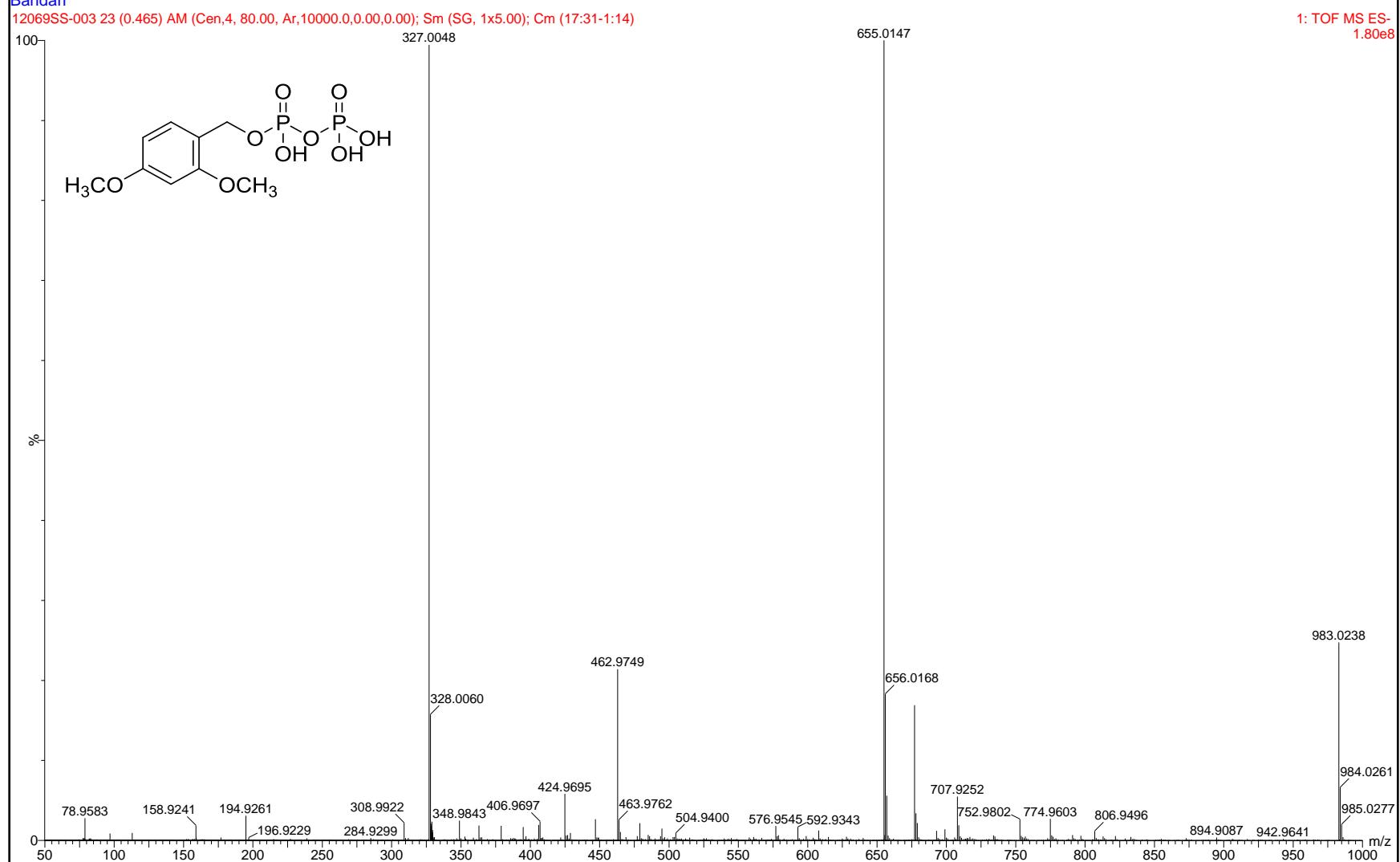
$^{31}\text{P}$  NMR Spectrum (162 MHz) of compound **31** in  $\text{D}_2\text{O}$

Ar10  
Bandari

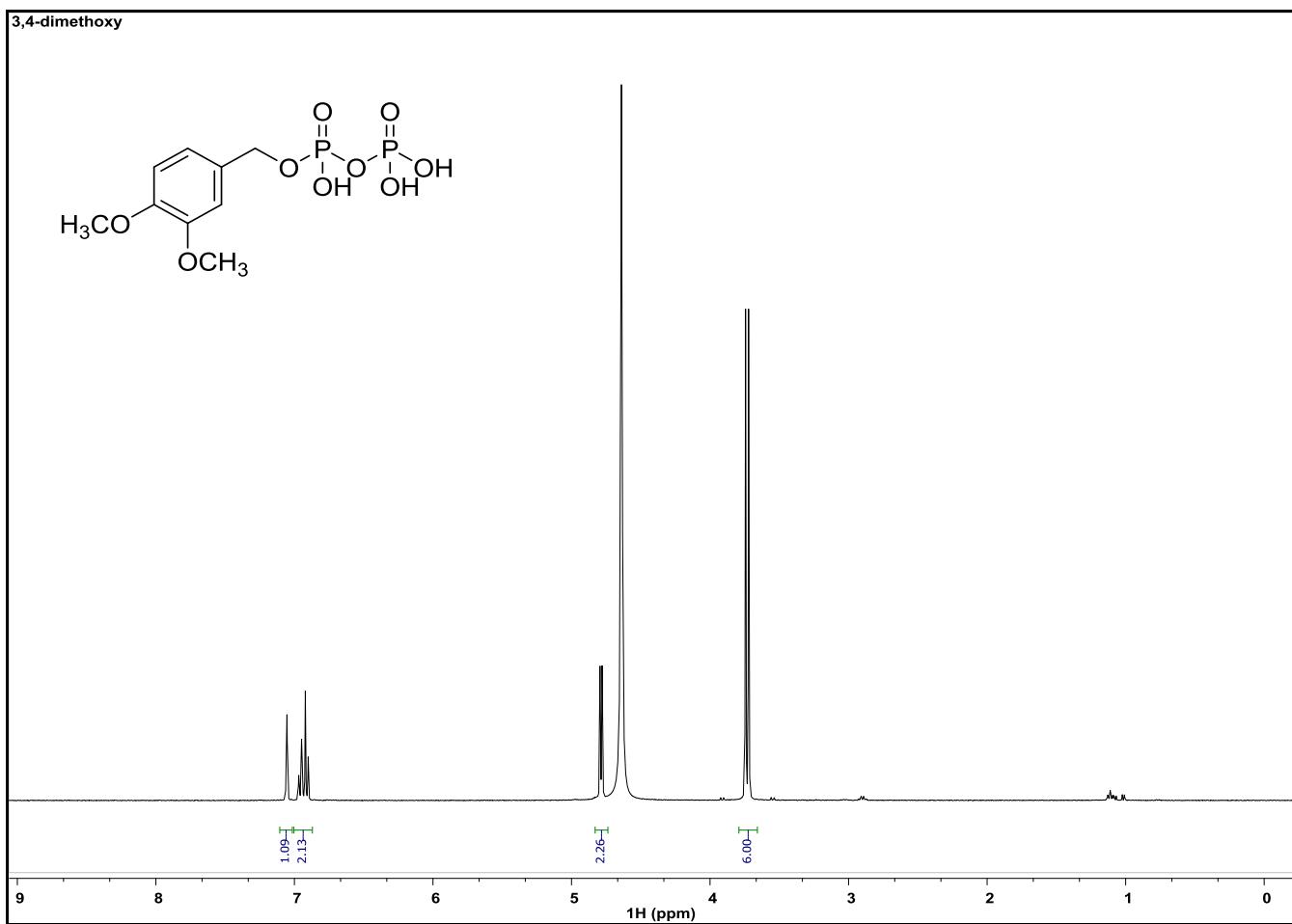
SYNAPTG2-Si#UGA589

06-Sep-2017 11:38:20

1: TOF MS ES-  
1.80e8

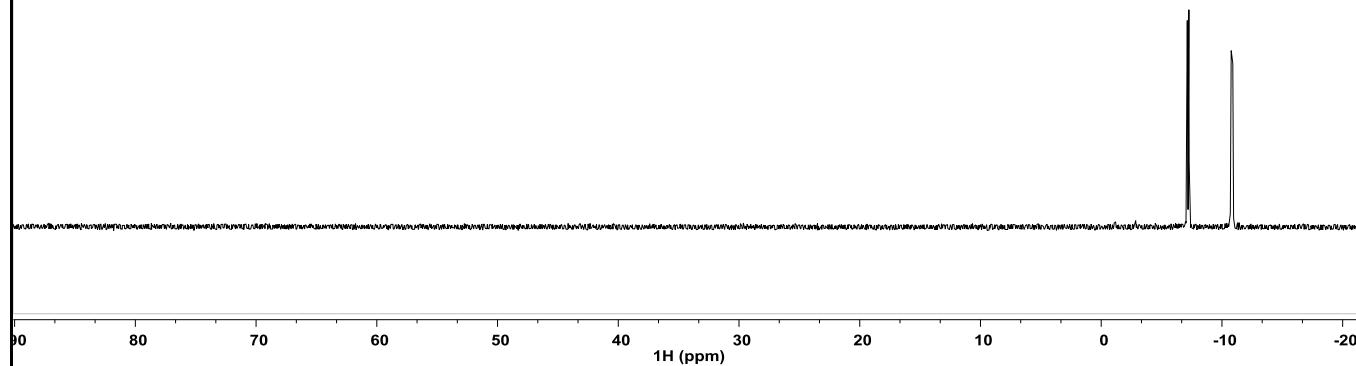
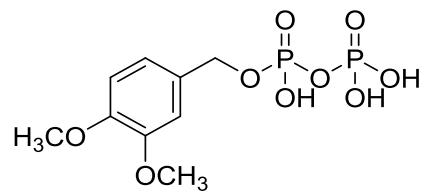


HRMS spectrum of 31



$^1\text{H}$  NMR Spectrum (400 MHz) of compound **32** in  $\text{D}_2\text{O}$

3,4-dimethoxy



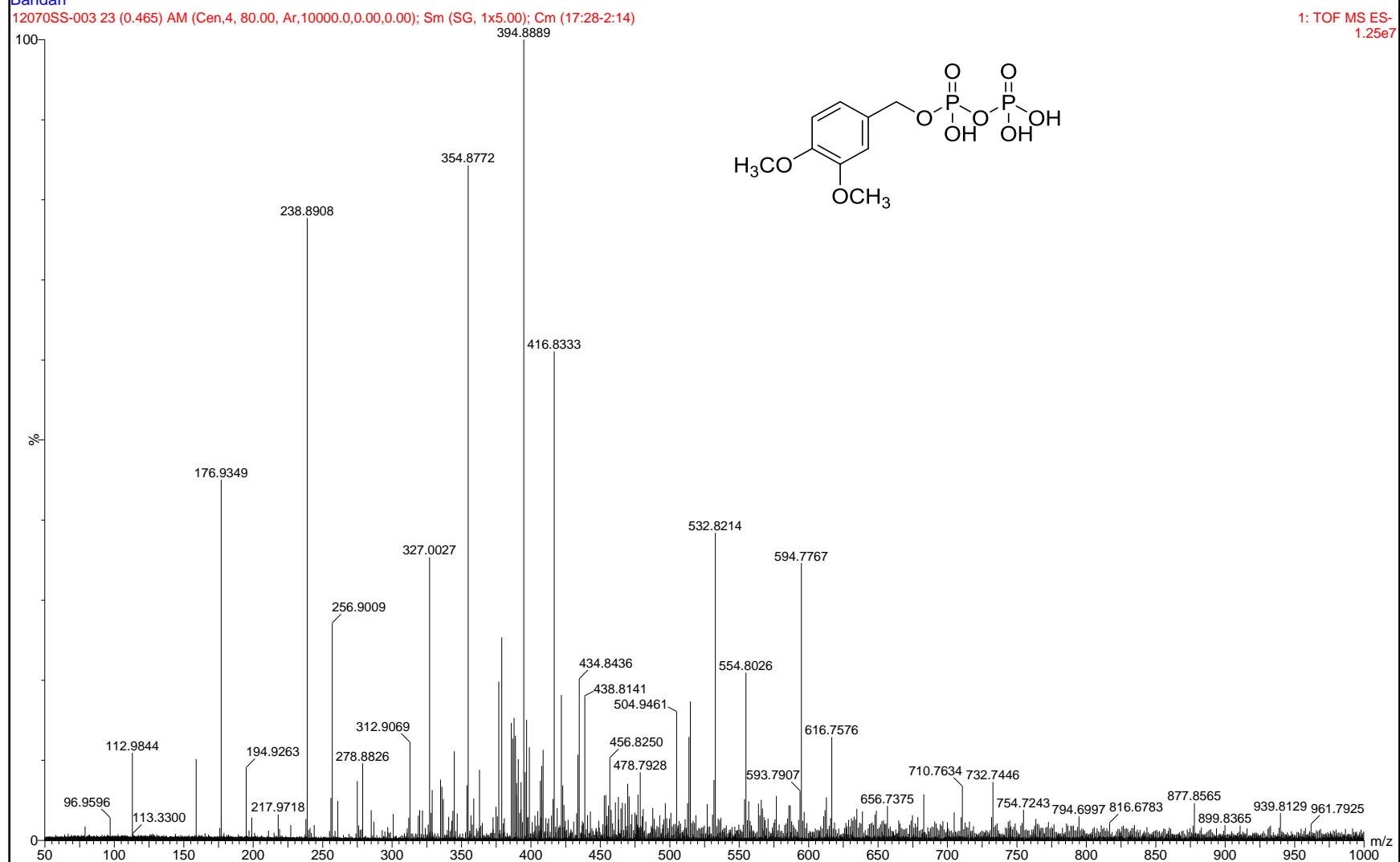
<sup>31</sup>P NMR Spectrum (162 MHz) of compound 32 in D<sub>2</sub>O

Ar11  
Bandari

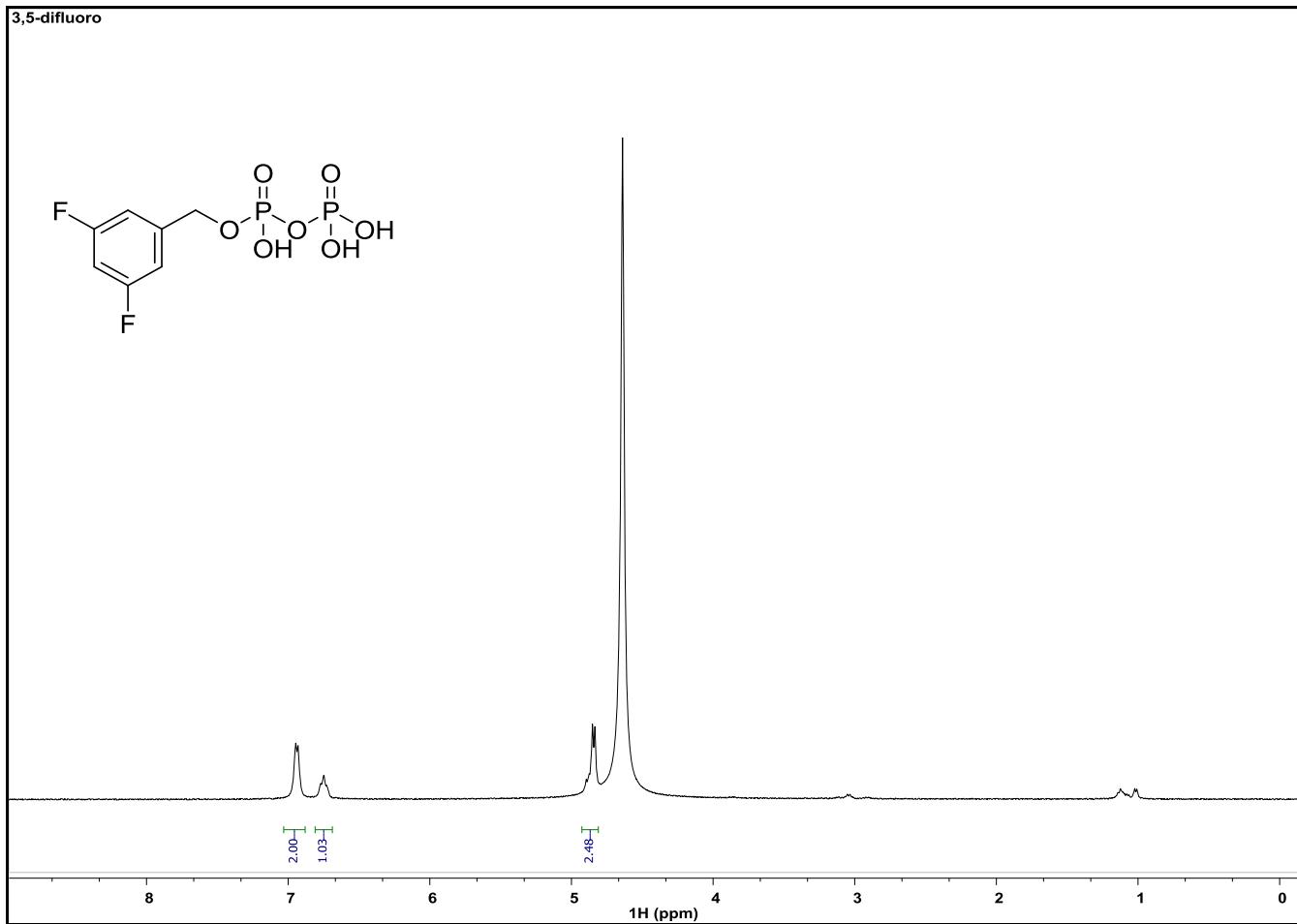
SYNAPTG2-Si#UGA589

06-Sep-2017 11:54:27

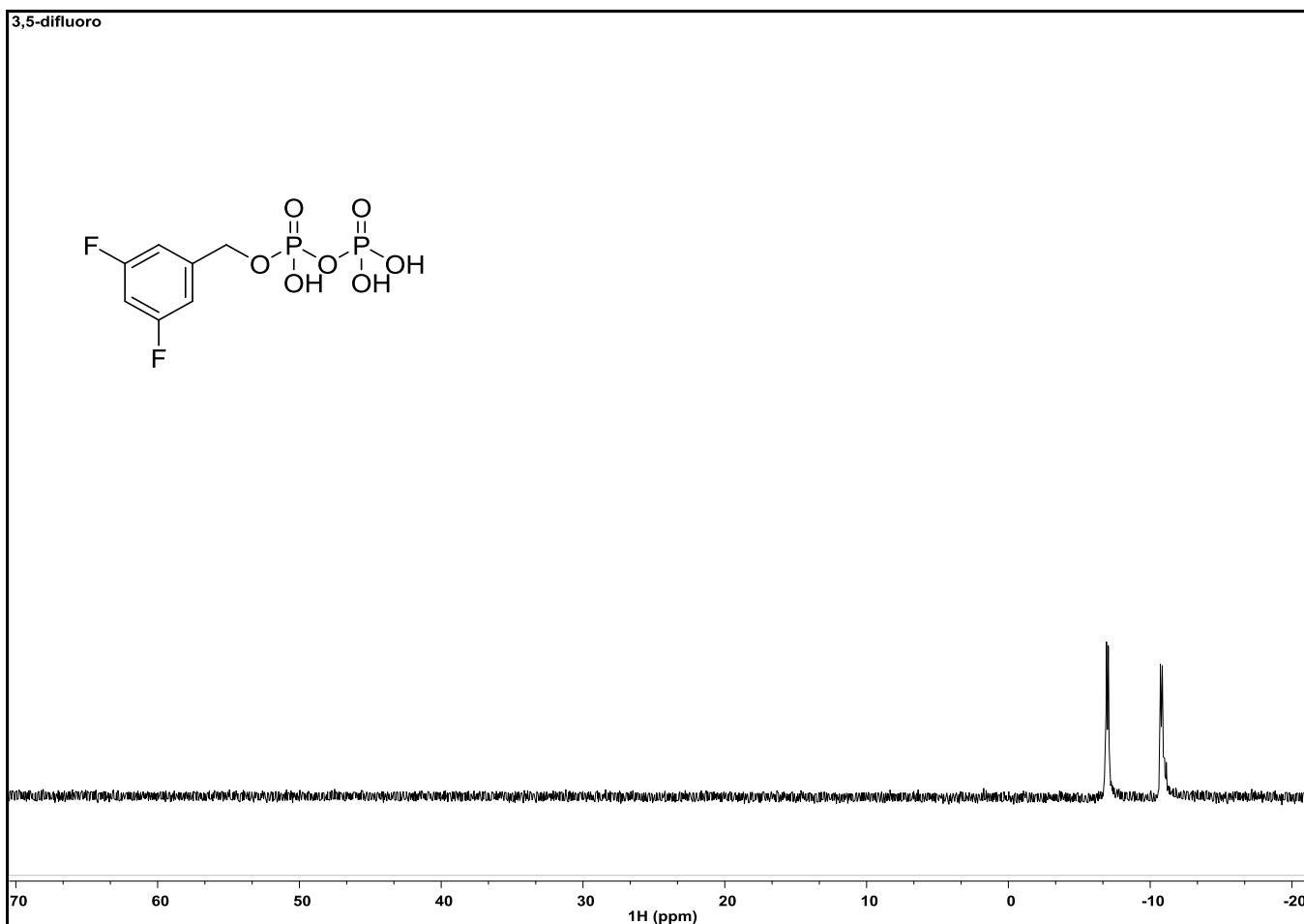
1: TOF MS ES-  
1.25e7



HRMS spectrum of 32



$^1\text{H}$  NMR Spectrum (400 MHz) of compound **33** in  $\text{D}_2\text{O}$



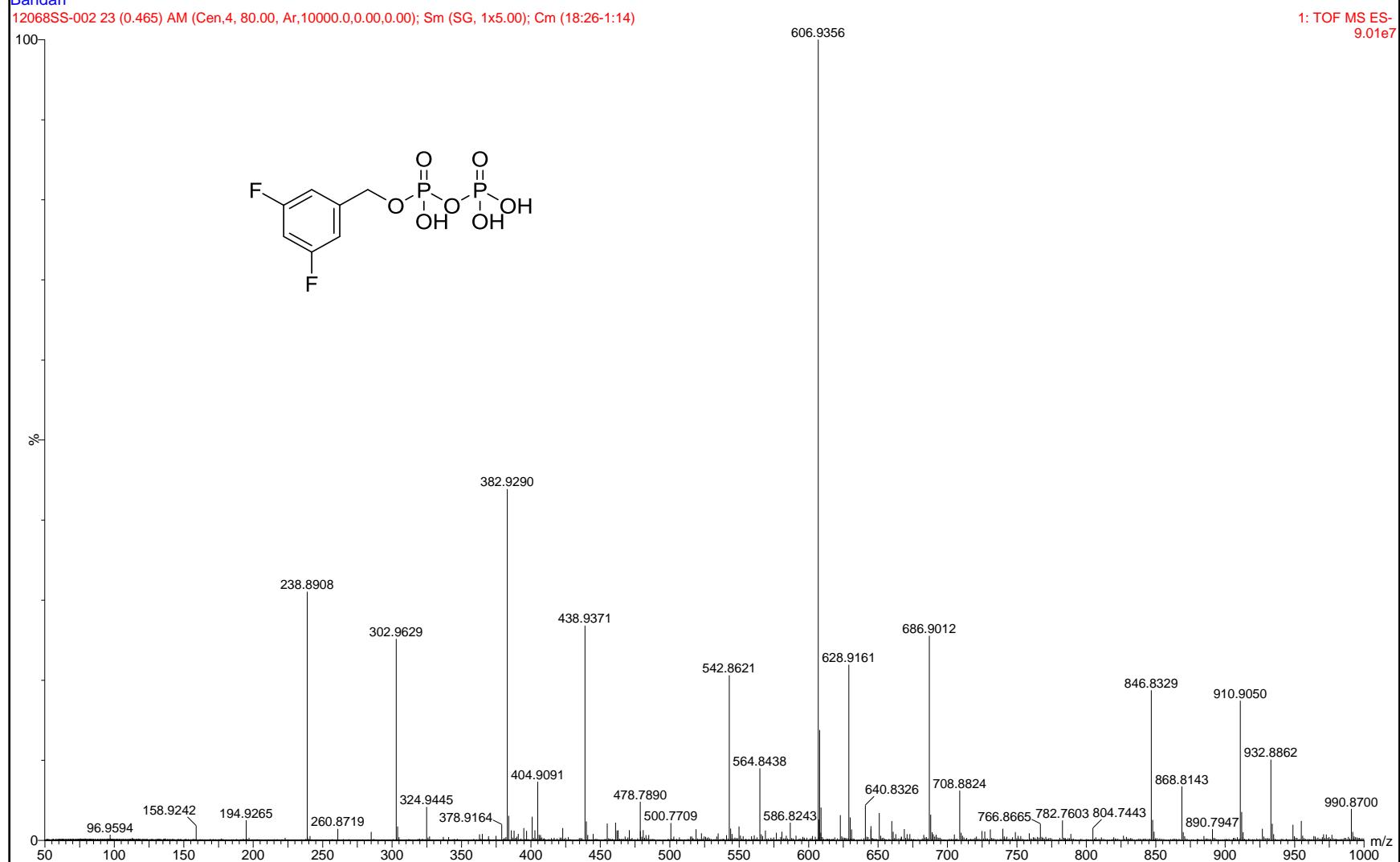
$^{31}\text{P}$  NMR Spectrum (162 MHz) of compound 33 in  $\text{D}_2\text{O}$

Ar12  
Bandari

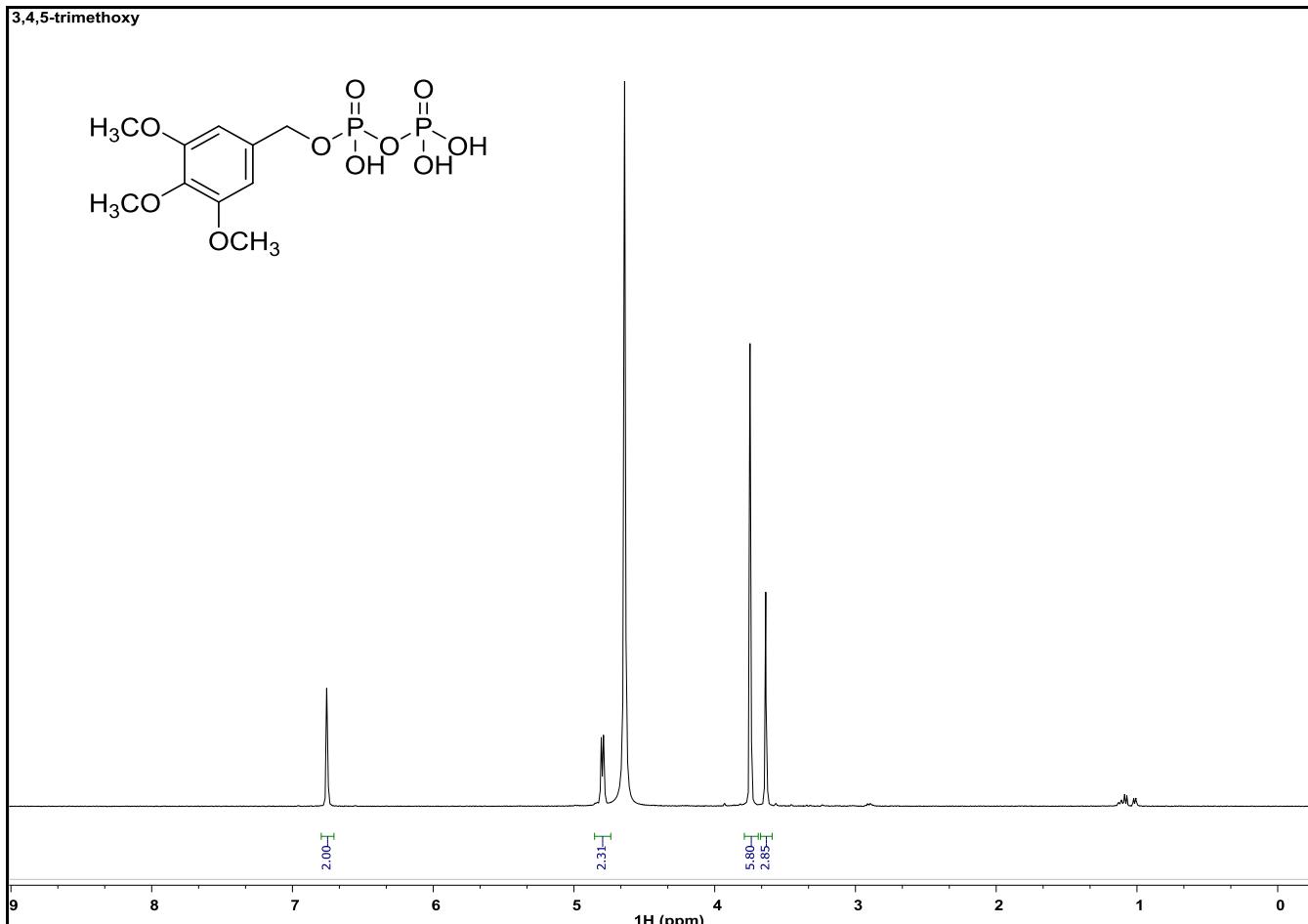
SYNAPTG2-Si#UGA589

06-Sep-2017 12:06:32

1: TOF MS ES-  
9.01e7

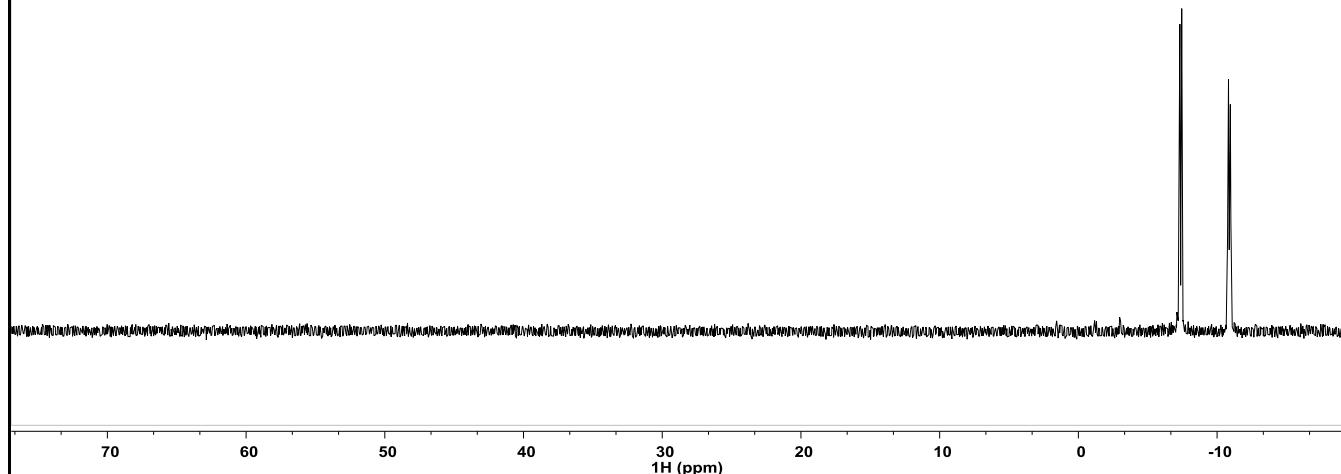
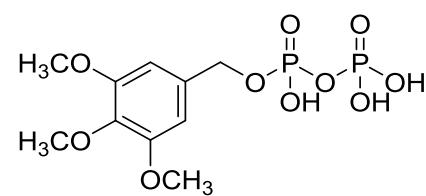


HRMS spectrum of 33



<sup>1</sup>H NMR Spectrum (400 MHz) of compound **34** in D<sub>2</sub>O

3,4,5-trimethoxy



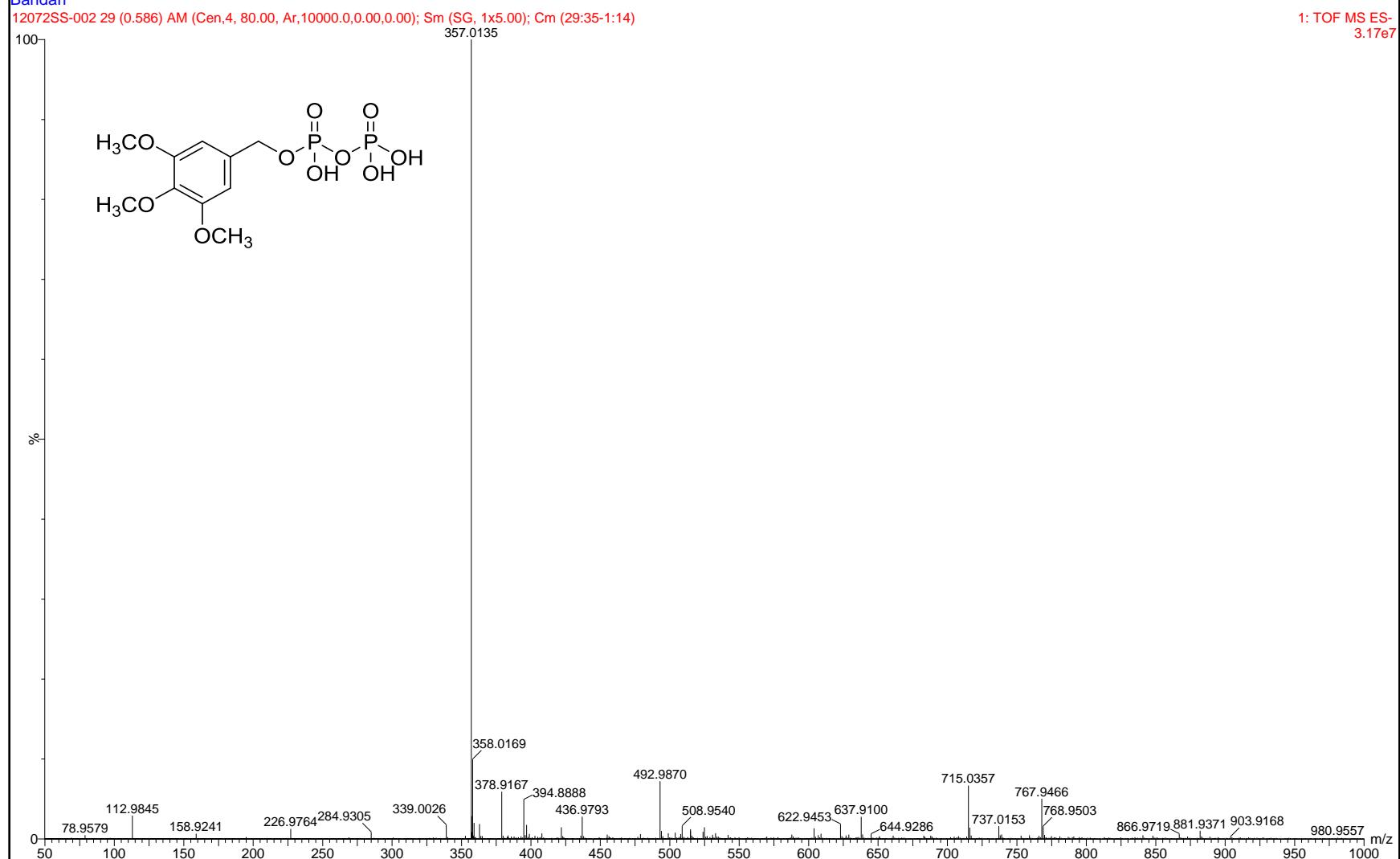
$^{31}\text{P}$  NMR Spectrum (162 MHz) of compound **34** in  $\text{D}_2\text{O}$

Ar14  
Bandari

SYNAPTG2-Si#UGA589

06-Sep-2017 12:38:42

1: TOF MS ES-  
3.17e7



HRMS spectrum of **34**