Electronic Supplementary Material (ESI) for RSC Medicinal Chemistry. This journal is © The Royal Society of Chemistry 2020

Electronic Supplementary information

to accompany the manuscript entitled:

Synthesis and in vivo evaluation of a radiofluorinated ketone body derivative

Stephanie J. Mattingly,^a Melinda Wuest,^{a,e} Eugene J. Fine,^b Ralf Schirrmacher,^{a,c} Frank Wuesta,^{c,d,e*}

^aDepartment of Oncology, University of Alberta, Edmonton, Canada
^bDepartment of Nuclear Medicine, Albert Einstein College of Medicine, NY, NY, USA
^cDepartment of Chemistry, University of Alberta, Edmonton, Canada
^dFaculty of Pharmacy and Pharmaceutical Sciences, University of Alberta, Edmonton, Canada
^eCancer Research Institute of Northern Alberta, University of Alberta, Edmonton, Canada

Contents

Figure S1 PET images in normal BALB/c mice	2
Figure S2 Time-activity curves of select organs in normal BALB/c mice	3
Figure S3 PET images of MDA-MB231 xenograft mice with nitrile intermediates	4
Figure S4 Changes in blood glucose and blood ketone body concentrations	5
Preparation of compound 3a	5
Preparation of compound 3b	5
Preparation of compound 3c	5
Preparation of compound 4	5
Preparation of nitrile reference standard FCH ₂ CHOHCH ₂ CN	6
Notes: nitrilase conversion step	6
NMR spectra: compound 3a	1
NMR spectra: compound 3b	3
NMR spectra: compound 3c	5
NMR spectra: compound 4	7
NMR spectra: nitrile reference standard FCH ₂ CHOHCH ₂ CN	8

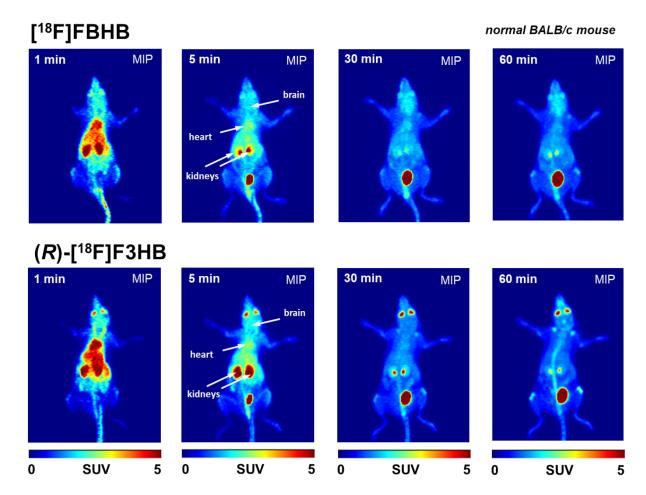


Figure S1 PET images in normal BALB/c mice post tail vein injection of [¹⁸F]FBHB (upper) and (*R*)-[¹⁸F]F3HB (lower) over 60 min. MIP, maximum intensity projection; SUV, standard uptake value.

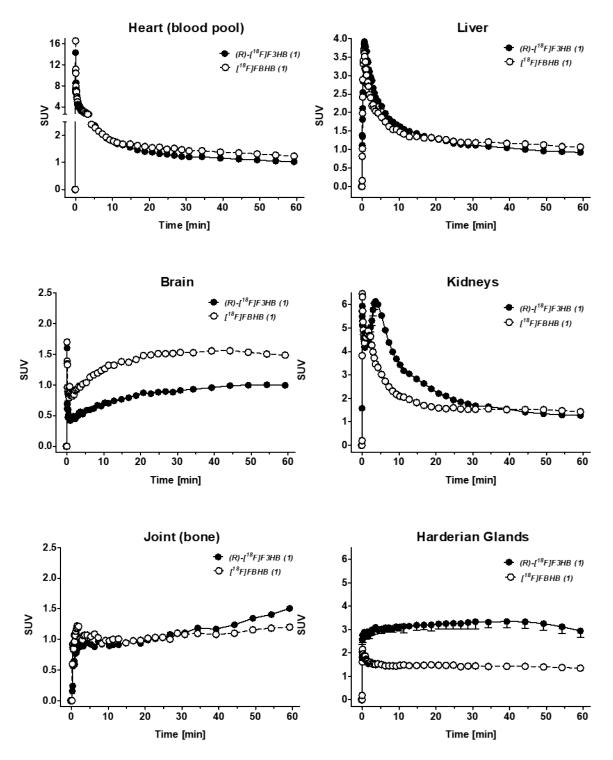


Figure S2 Time-activity curves of select organs in normal BALB/c mice post tail vein injection of $[^{18}F]FBHB$ or (*R*)- $[^{18}F]F3HB$ over 60 min.

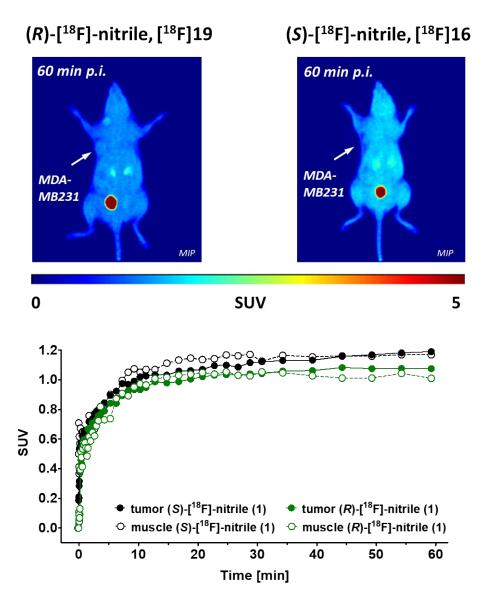


Figure S3 PET images of MDA-MB231 xenograft mice with nitrile intermediates compounds $[^{18}F]19((R)-[^{18}F]-nitrile)$ or $[^{18}F]16((S)-[^{18}F]-nitrile)$ 1h post injection (*upper*); corresponding time-activity curves of tumor versus muscle uptake (*lower*)

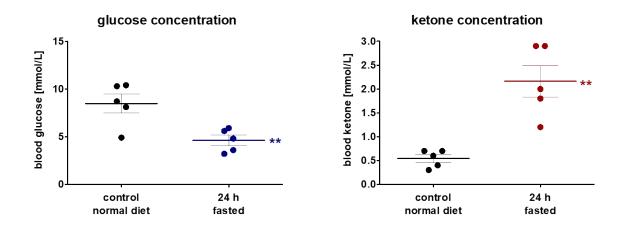
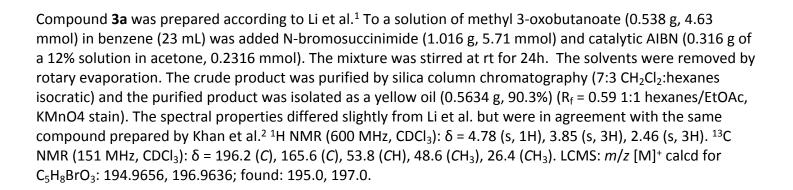


Figure S4 Changes in blood glucose and blood ketone body concentrations prior to and post a 24 h fast

OMe

Preparation of compound 3a



Preparation of compound 3b



Compound **3b** was prepared using the same methodology as for **3a**. Spectral properties were in agreement with the compound as prepared by Wu et al.³ ¹H NMR (600 MHz, CDCl₃): δ = 4.76 (s, 1H), 4.29 (q, *J* = 6.40 Hz, 2H), 2.45 (s, 3H), 1.32 (t, *J* = 7.15, 3H). ¹³C NMR (151 MHz, CDCl₃): δ = 196.4 (*C*), 165.1 (*C*), 63.2 (*C*H₂), 49.1 (*C*H), 26.4 (*C*H₃), 13.9 (*C*H₃).



Compound **3c** was prepared using the same methodology as for **3a**. Spectral properties were in agreement with the same compound prepared by Khan et al.² ¹H NMR (600 MHz, CDCl₃): δ = 4.68 (s, 1H), 2.42 (s, 3H), 1.50 (s, 9H). ¹³C NMR (151 MHz, CDCl₃): δ = 196.7 (*C*), 164.0 (*C*), 84.5 (*C*), 50.7 (*C*H), 27.7 (*C*H₃), 26.3 (*C*H₃).

Preparation of compound 4



Compound **4** was prepared according to Coats et al.⁴ To *tert*-butyl acetoacetate (0.477 g, 3.01 mmol) in CH₃CN (15 mL) was added Koser's reagent ([hydroxy(tosyloxy)iodo]benzene) (1.42 g, 3.62 mmol). The slurry was heated to 70 °C until a colorless homogenous solution was obtained (<2h). The crude product was purified by silica column chromatography, gradient elution 1:0 to 7:3 hexanes:EtOAc (R_f = 0.31 4:1 hexanes/EtOAc). Compound **4** was isolated as a white solid (0.7418 g, 99%). Spectral properties compare well with those reported for the methyl ester derivative by Yamamoto et al.⁵ ¹H NMR (600 MHz, CDCl₃): δ = 7.84 (d, *J* = 8.28 Hz, 2H), 7.37 (d, *J* = 8.66 Hz, 2H), 5.31 (s, 1H), 2.46 (s, 3H), 2.29 (s, 3H), 1.40 (s, 9H).

Preparation of nitrile reference standard FCH₂CHOHCH₂CN

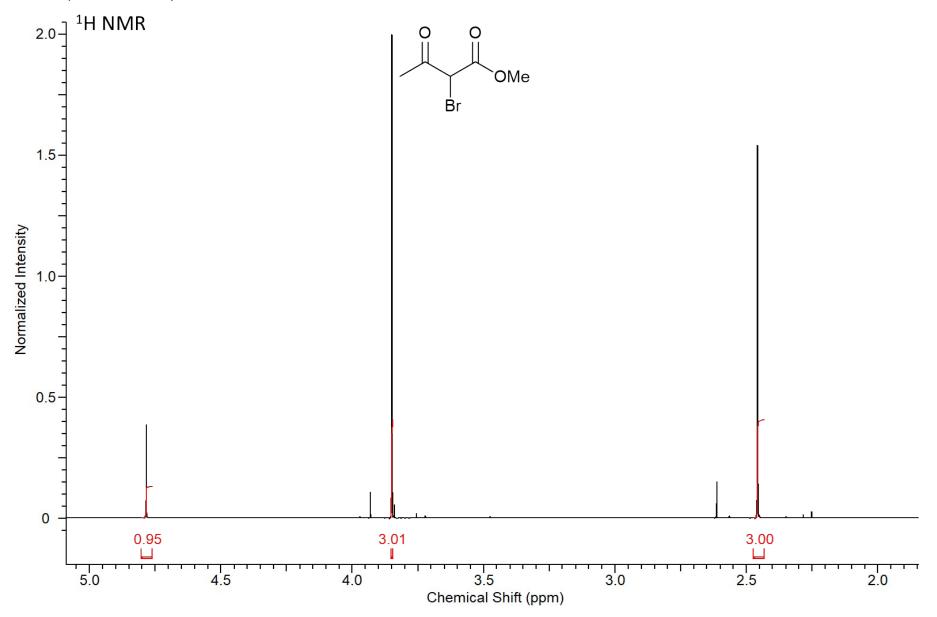


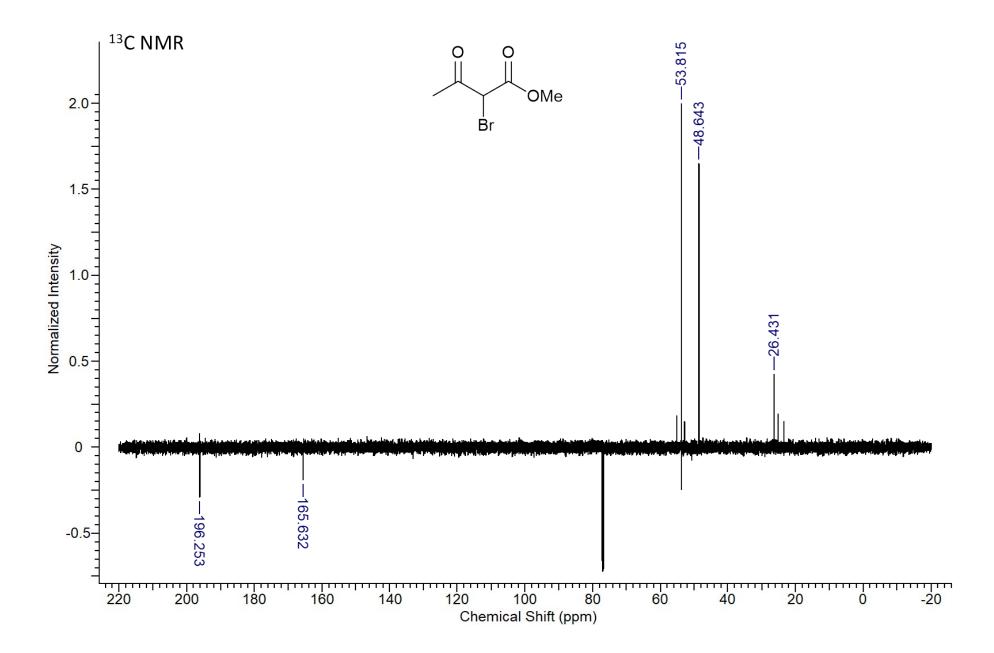
Nitrile reference standard FCH₂CHOHCH₂CN was prepared according to Liu et al.⁶ CAUTION: HCN (b.p. 25.6 °C) and epifluorohydrin (b.p. 85-86 °C) are toxic. H₂SO₄ (0.967 g, 9.86 mmol) was dissolved in water (33 mL) and chilled in an ice bath. To this solution was added epifluorohydrin (0.250 g, 3.29 mmol) followed by KCN (1.712 g, 26.3 mmol). The reaction mixture was allowed to gradually come to rt and was stirred for 16 h. The reaction mixture was extracted with EtOAc (3x30 mL), washed with saturated aqueous NaHCO₃ (20 mL) and brine (20 mL), dried over Na₂SO₄, and the solvent evaporated by rotary evaporation. The crude product was purified by silica column chromatography eluting with hexanes/EtOAc (R_f = 0.28 1:1 hexanes/EtOAc, KMnO₄ stain). The solvents were evaporated yielding FCH₂CHOHCH₂CN (0.3097 g, 91.3%) as a bright yellow oil. ¹⁹F NMR (565 MHz, CDCl₃): δ = -230.94 (td, *J* = 47.0, 17.8 Hz). ¹H NMR (600 MHz, CDCl₃): δ = 4.47 (dd, ²*J*_{H-F} = 47.1 Hz, ³*J*_{H-H} = 4.89 Hz, 2H), 4.17-4.27 (m, 1H),3.08 (brs, 1H), 2.71-2.61 (m, 2H). ¹³C NMR (151 MHz, CDCl₃): δ = 116.8 (*C*), 85.6 (d, ¹*J*_{C-F} = 172.6 Hz, *C*H₂F), 66.1 (d, ²*J*_{C-F} = 22.1 Hz, *C*H), 21.7 (d, ³*J*_{C-F} = 7.7 Hz, *C*H₂).

Notes: nitrilase conversion step

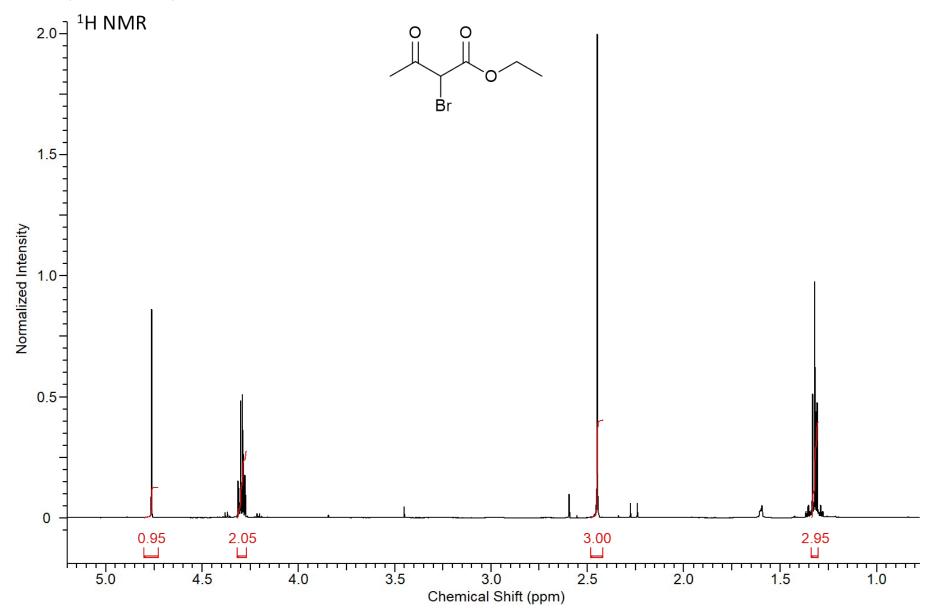
The enzymatic conversion step resulted in lower yields at temperatures above 30 °C and at pH below 9. Yields improved when PBS buffer was added; however, in this instance the presence of buffer salts hindered QMA cartridge capture of the final product acids. Note that for HPLC-based verifications of acids [¹⁸F]FBHB and (R)-[¹⁸F]F3HB, elution times vary depending on solvent pH and ionic strength (i.e. the presence of buffer salts).

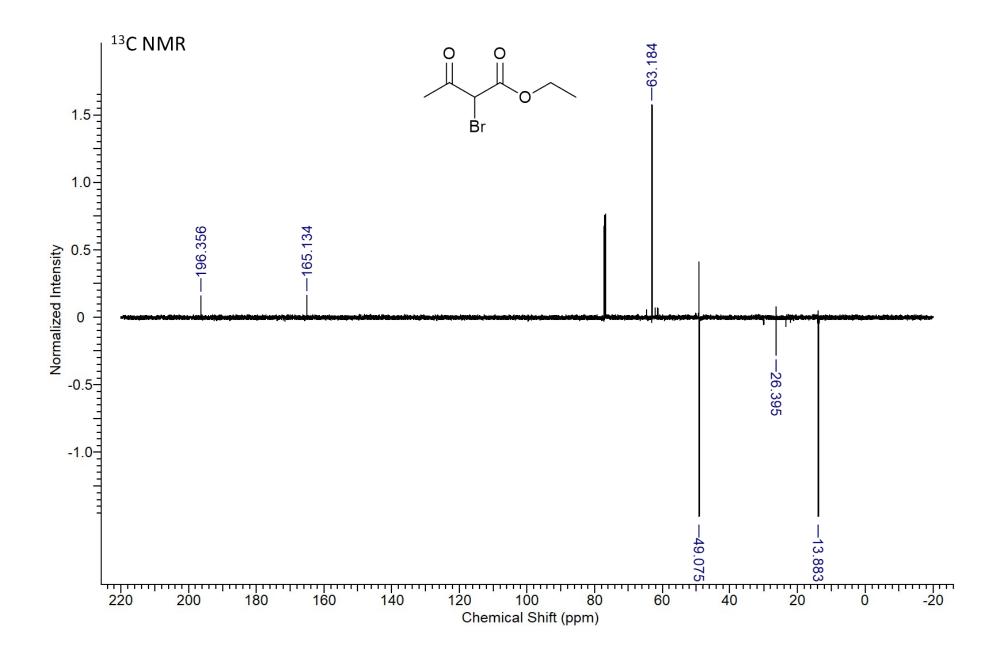
NMR spectra: compound 3a



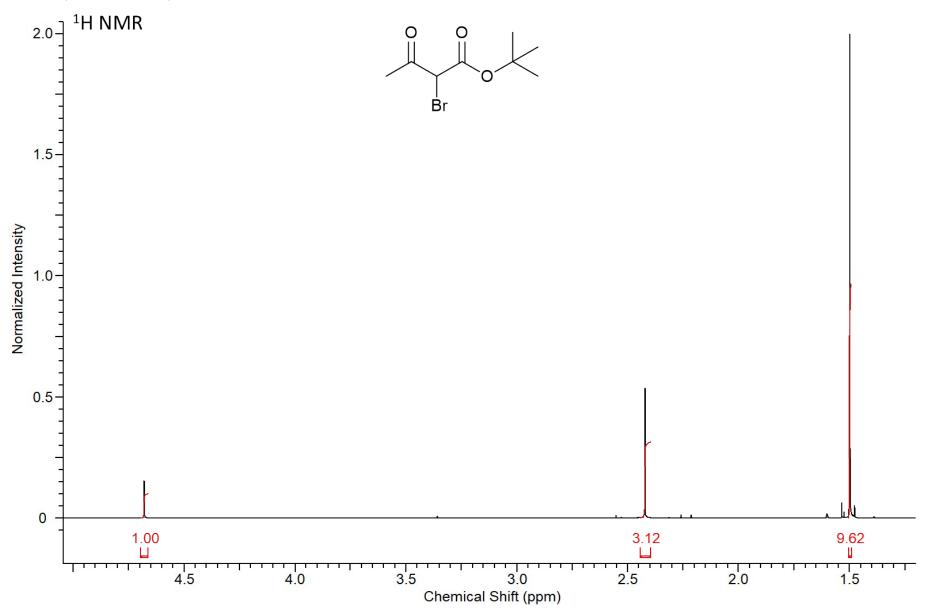


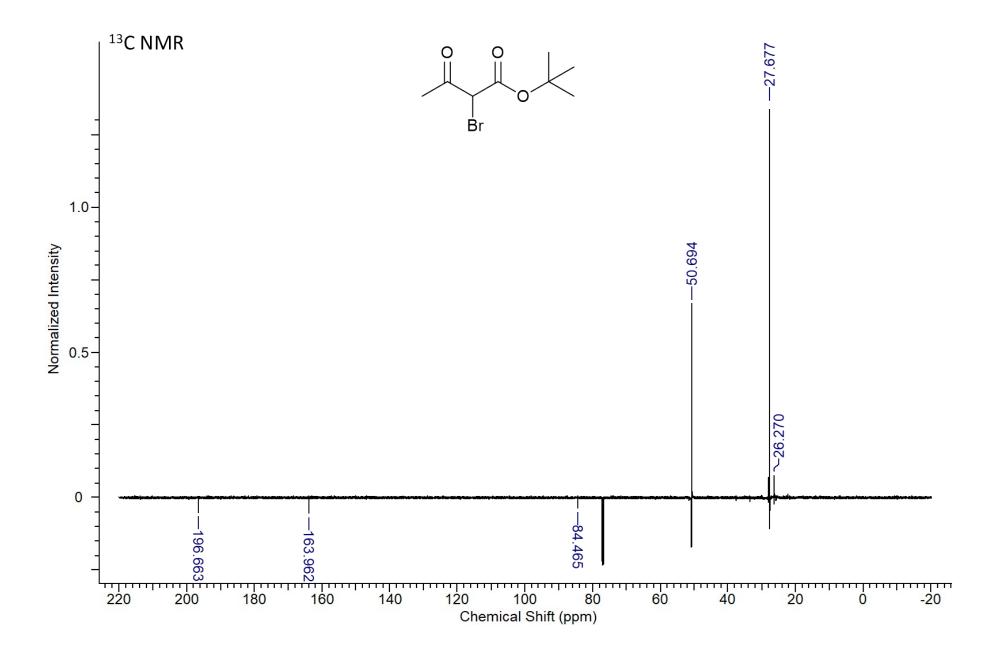
NMR spectra: compound **3b**



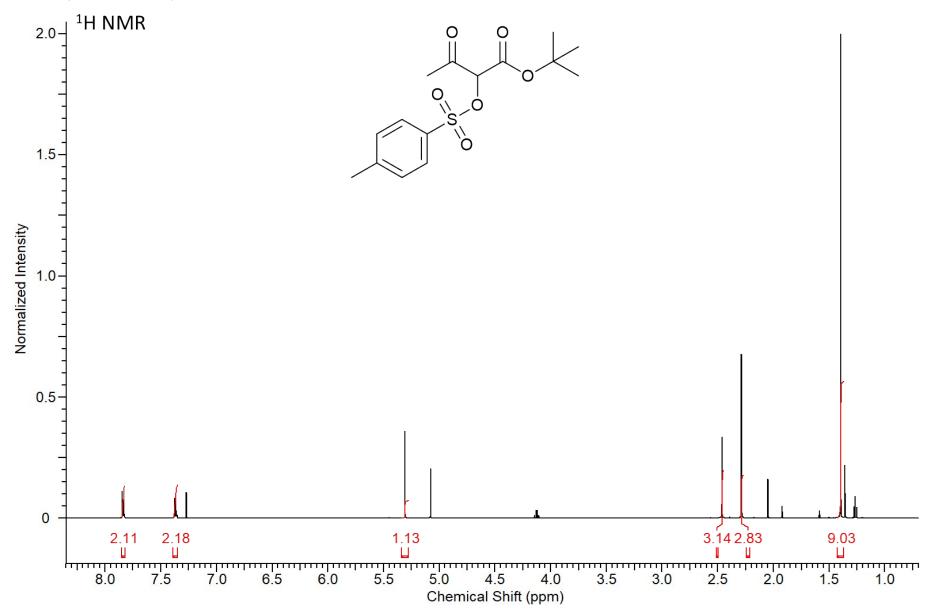


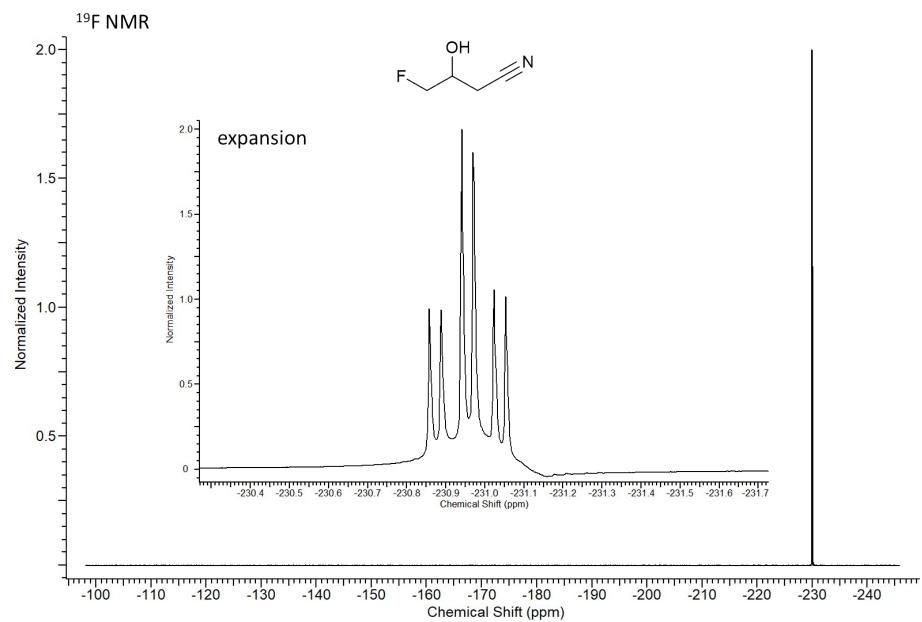
NMR spectra: compound **3c**



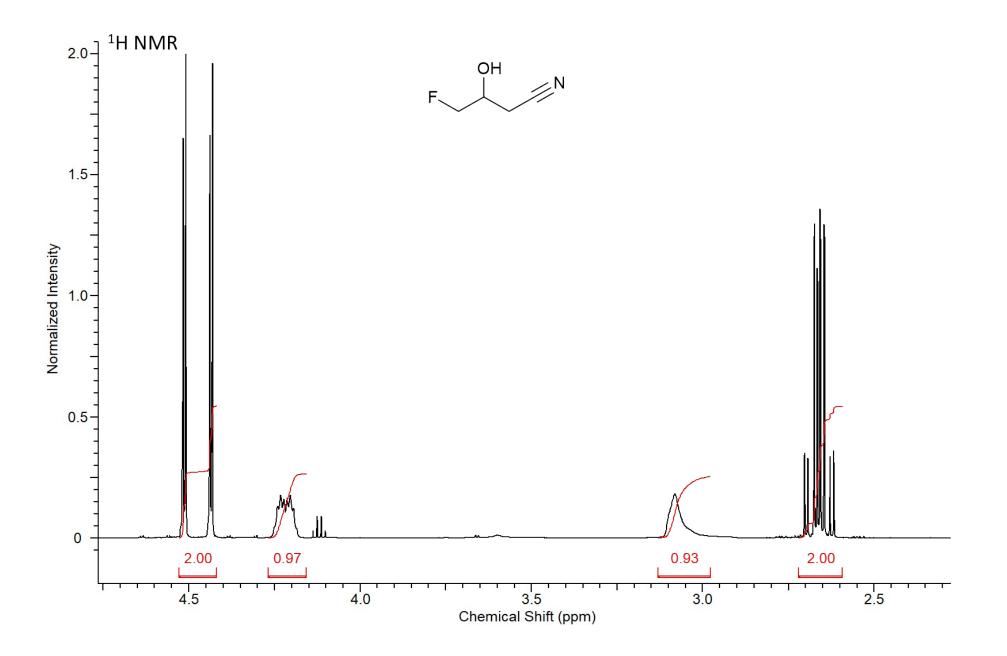


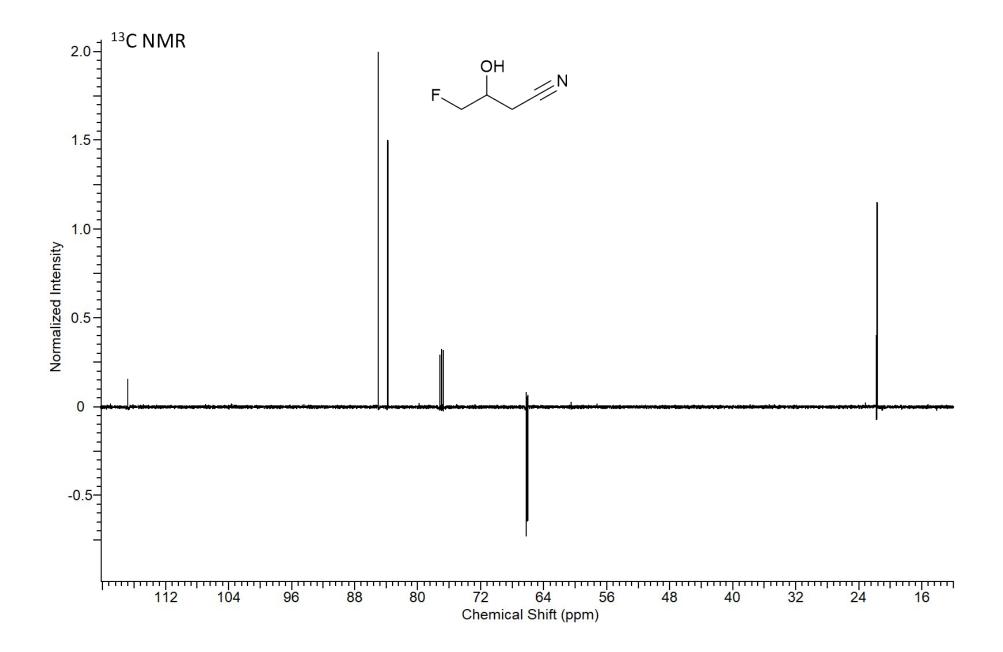
NMR spectra: compound 4





NMR spectra: nitrile reference standard FCH₂CHOHCH₂CN





- 1. Z. Li, M. Khaliq, Z. Zhou, C. B. Post, R. J. Kuhn and M. Cushman, *J. Med. Chem.*, 2008, **51**, 4660-4671.
- 2. A. T. Khan, M. A. Ali, P. Goswami and L. H. Choudhury, *J. Org. Chem.*, 2006, **71**, 8961-8963.
- 3. L. Wu and Z. Yin, *European Journal of Inorganic Chemistry*, 2013, **2013**, 6156-6163.
- 4. S. J. Coats and H. H. Wasserman, *Tetrahedron Lett.*, 1995, **36**, 7735-7738.
- 5. Y. Yamamoto, Y. Kawano, P. H. Toy and H. Togo, *Tetrahedron*, 2007, **63**, 4680-4687.
- 6. Y. Liu, C. Hazzard, A. S. Eustaquio, K. A. Reynolds and B. S. Moore, *J. Am. Chem. Soc.*, 2009, **131**, 10376-10377.