

Electronic supplementary information

## **Dimers of Glutaredoxin 2 as Mitochondrial Redox Sensors in Selenite-induced Oxidative Stress**

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**Fig. S 1. Determination of protein-bound glutathione**

**Fig. S 2. Determination of Grx2 monomerization in cells after oxidative stress induction**

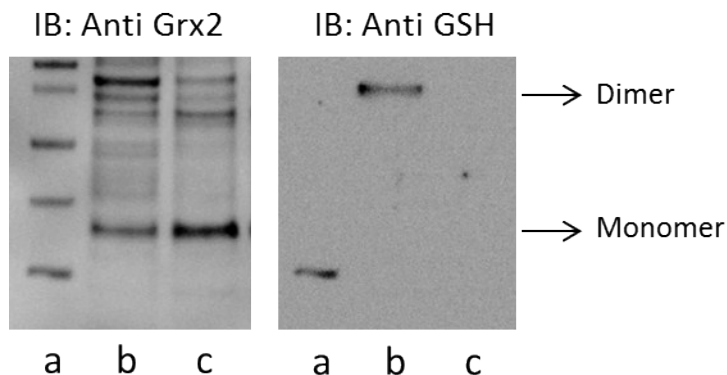
**Fig. S 3. Specific activity of TrxR1 and TrxR2 and of cytosolic and mitochondrial Grx in Sel treated cells**

**Fig. S 4. Iron distribution in the two mitochondrial fractions obtained after treatment with *meta*-phosphoric acid**

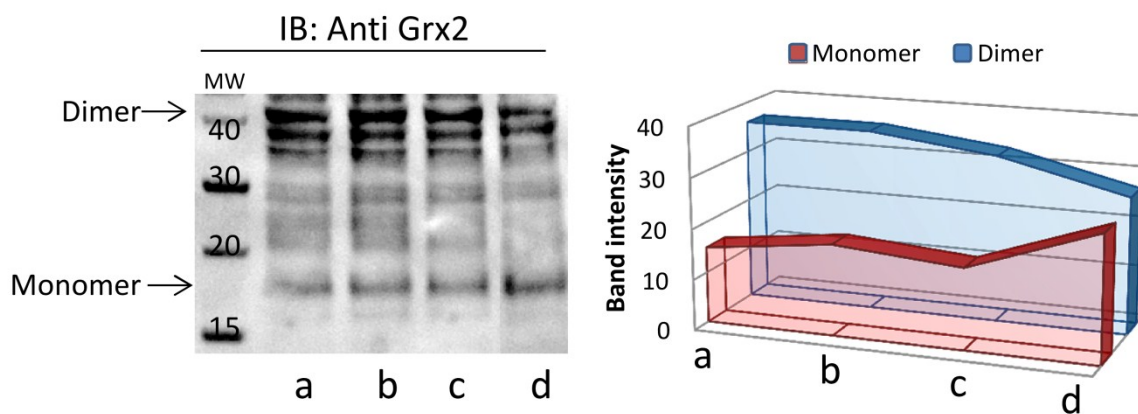
**Fig. S 5. FACS analysis of lipid peroxidation in HeLa cells treated with Sel +/- BHT using C11-BODIPY dye**

**Fig. S 6. Release of Cyt C after Sel treatment in HeLa cells.**

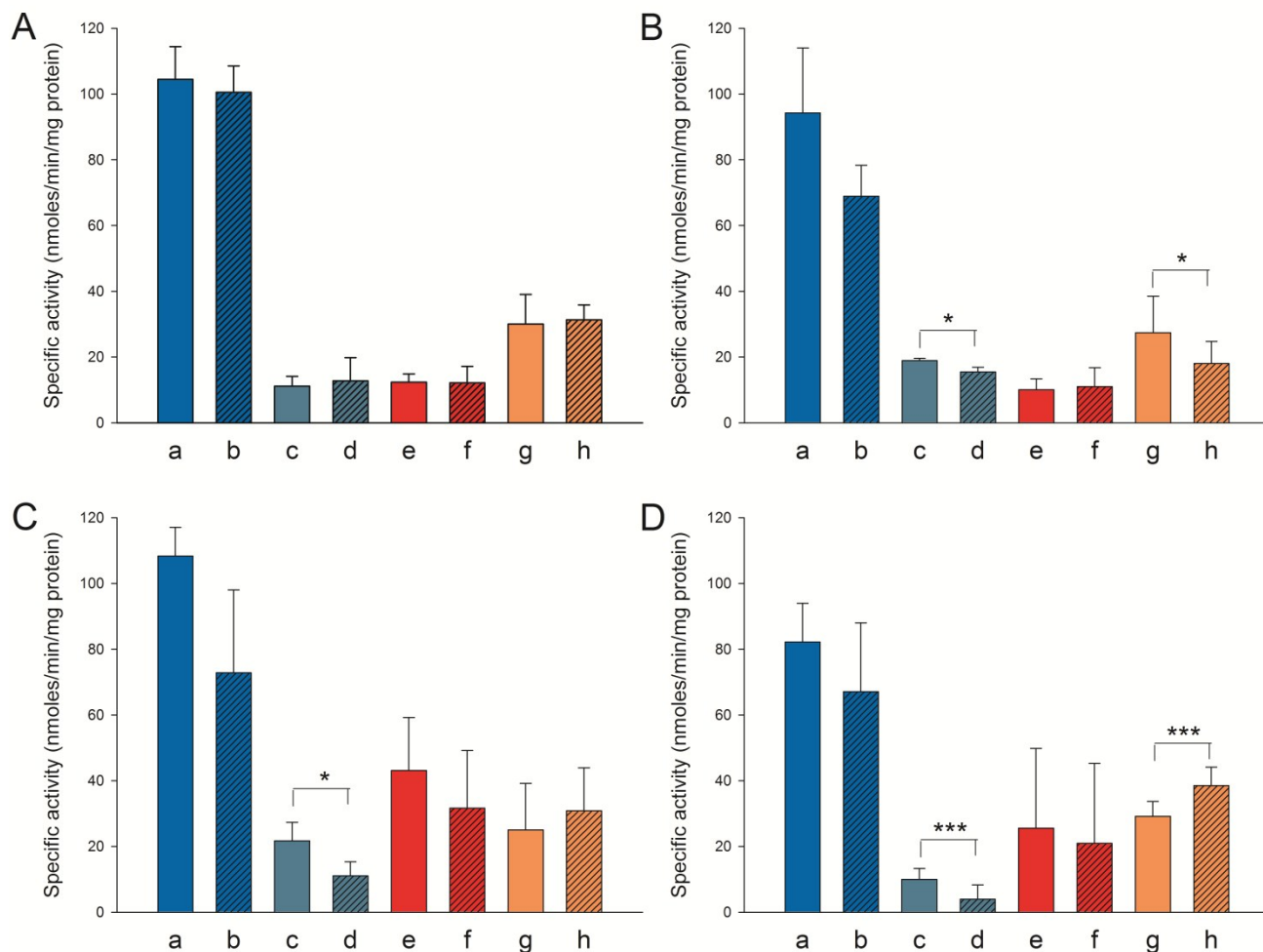
**Fig. S 7. Western blot analysis of GPx4 in mitochondria samples in the presence of 15  $\mu$ M Sel.**



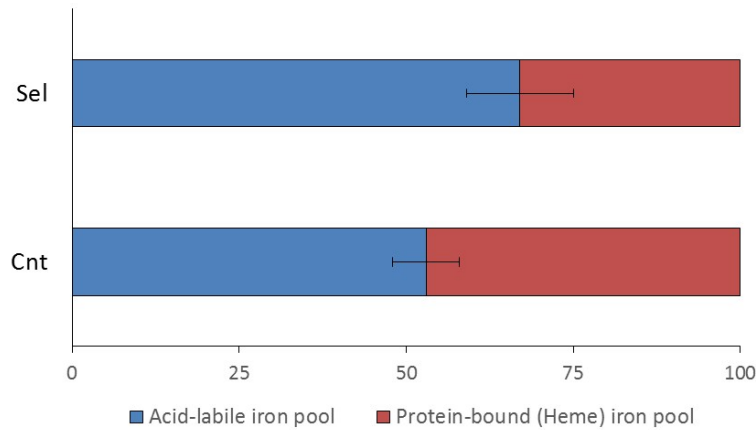
**Figure S 1. Determination of protein-bound glutathione.** HeLa cells were treated for 18 h with 15  $\mu$ M Sel. To analyze the monomerization of Grx2, mitochondrial cell fractions derivatized with 10 mM AIS, were subjected to Nu-PAGE in non-reducing conditions and WB analysis. The membrane was probed with an anti-Grx2 and an anti-GSH Ab. (a) Markers; (b) Cnt; (c) 15  $\mu$ M Sel. (IB: immunoblot).



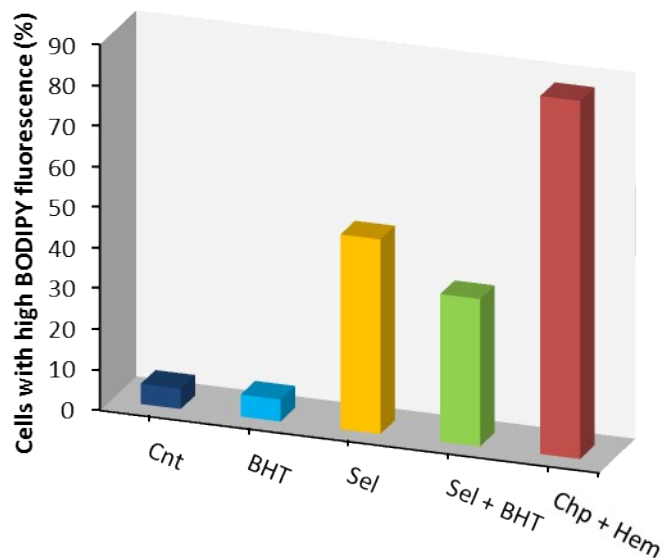
**Figure S 2. Determination of Grx2 monomerization in cells after oxidative stress induction.** HeLa cells were treated with **a:** Cnt; **b:** 1 mM BSO 48 h; **c:** 1  $\mu$ M AF 3 h, **d:** 1 mM BSO 48 h + 1  $\mu$ M AF 3 h. To analyze the monomerization of Grx2, mitochondrial cell fractions, derivatized with 10 mM AIS, were subjected to SDS-PAGE in non-reducing conditions and WB analysis. The densitometric analysis of the bands performed with NineAlliance software is reported in the right panel.



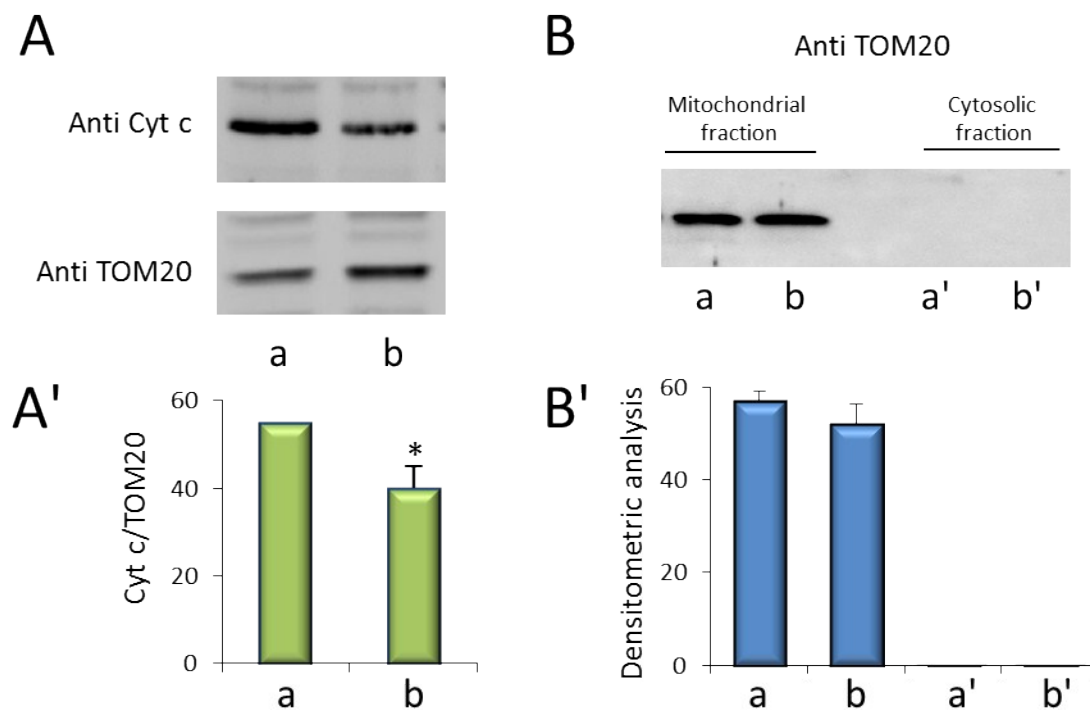
**Figure S 3. Specific activity of TrxR1 and TrxR2 and of cytosolic and mitochondrial Grx in Sel treated cells.** HeLa cells ( $3 \times 10^7$ ) were incubated for 3 (A), 6 (B), 12 (C) and 18 (D) h with 15  $\mu$ M Sel or in control conditions and then subjected to cell sub-fractionation in order to obtain the cytosol and mitochondria enriched cell fractions. The cytosolic fraction was checked for TrxR1 (blue bars) and total glutaredoxin (red bars) activities; the mitochondrial fraction was probed for TrxR2 (gray bars) and Grx2 (orange bars) activities. The graphs reports the specific activity of each enzyme, expressed as nmoles/min/mg protein, in Sel treated or control cells. **a:** TrxR1 Cnt; **b:** TrxR1 15  $\mu$ M Sel; **c:** TrxR2 Cnt; **d:** TrxR2 15  $\mu$ M Sel; **e:** cytosolic Grx Cnt; **f:** cytosolic Grx 15  $\mu$ M Sel; **g:** mitochondrial Grx Cnt; **h:** mitochondrial Grx 15  $\mu$ M Sel. The graph shows the mean  $\pm$  SD of 3 experiments (\* $p < 0.05$ , \*\*\* $p < 0.001$ ).



**Figure S 4. Iron distribution in the two mitochondrial fractions obtained after treatment with *meta*-phosphoric acid.** HeLa cells ( $3 \times 10^7$ ) were treated with 15  $\mu$ M Sel for 18 h and then processed to obtain mitochondria following the protocol of Clayton and Shadel (see Experimental section). Mitochondria were then treated with 600  $\mu$ L of 6% *meta*-phosphoric acid for 20 min at 4°C in order to extract the labile iron pool. At the end of incubation, samples were centrifuged at 15800g for 10 min at 4°C. Both the supernatants (Acid-labile iron pool) and the pellets (Protein-bound (Heme) iron pool) were subjected to mineralization and analysis of the Fe content. The percentage of iron measured in the two fractions (mean  $\pm$  SD of 4 experiments) is reported.

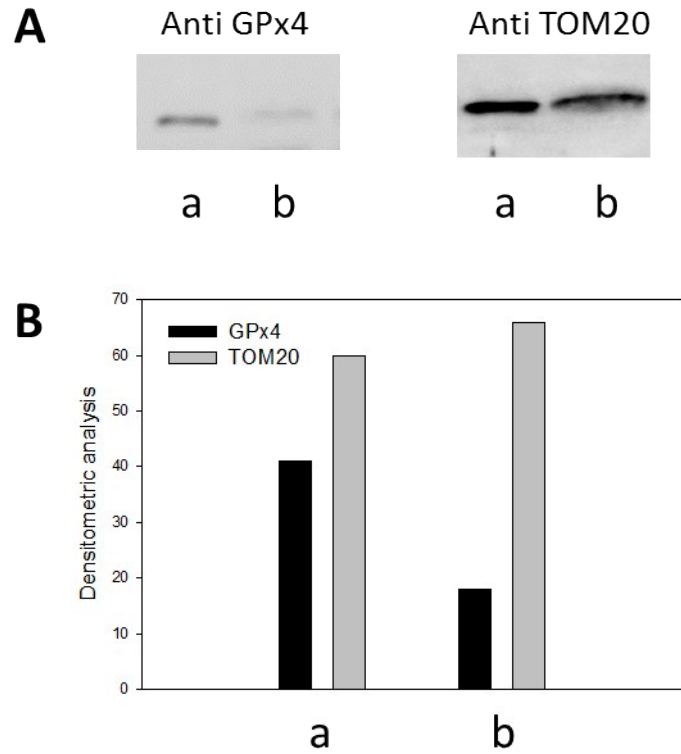


**Figure S 5. FACS analysis of lipid peroxidation in HeLa cells treated with Sel +/- BHT using C11-BODIPY dye.** HeLa cells ( $4.5 \times 10^5$ ) were treated with  $15 \mu\text{M}$  Sel for 18 h with or without pre-incubation with  $0.1 \text{ mM}$  butylated hydroxytoluene (BHT) for 2 h and then subjected to the analysis of lipid peroxidation using C11-BODIPY dye as described in Experimental. Treatment of cells with  $2.5 \text{ mM}$  cumene hydroperoxide +  $5 \mu\text{M}$  hemin (Chp + Hem) for 15 min is displayed as a positive control of lipid peroxidation induction.



**Figure S 6. Release of Cyt C after Sel treatment in HeLa cells**

**(A, A')** Detection of Cyt c remaining in the mitochondrial fraction. Cells ( $4.5 \times 10^5$ ) were treated with 15  $\mu$ M Sel for 18 h and then processed as reported in the Experimental section. **(A)** Aliquots of 10  $\mu$ g protein of the mitochondrial fractions were subjected to SDS-PAGE (15%) followed by Western blot using a Cyt c monoclonal antibody. TOM20 content was determined as a loading control. **(A')** Relative amount of Cyt c in the mitochondrial fraction of control (a) or Sel-treated (b) HeLa cells showing a decrease of Cyt c in Sel-treated mitochondria. The graph reports the mean  $\pm$  SD of 3 experiments (\* $p < 0.05$ ). **(B)** Estimation of the purity of the cytosolic cellular fraction (absence of mitochondrial residues) through the analysis of the amount of the mitochondrial marker TOM20. **(B')** Densitometric analysis of the Western blot bands reported in **B** utilizing the Nine-Alliance software. Control (**a, a'**); Sel-treated (**b, b'**).



**Figure S 7. Western blot analysis of GPx4 in mitochondria samples in the presence of 15  $\mu$ M Sel. (A)** Level of GPx4 decreases in mitochondria of HeLa cells treated with 15  $\mu$ M Sel for 18 h. In the same, the detection of TOM20 was performed as loading control. **(B)** Densitometric analysis of the Western blot bands reported in **A** utilizing the ImageJ software. **a:** Cnt; **b:** 15  $\mu$ M Sel.