

# First Aromatic Amine Organocatalysed Activation of $\alpha,\beta$ - Unsaturated Ketones

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## Supporting Information

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## 1. General Information

Starting materials **1a** and **2a-d**, as well as the amine catalysts **I-IX**, are commercially available and were employed as received without further treatment or purification. In case of solvent, commercial tetrahydrofuran (THF, HPLC grade) was transferred to a new recipient and molecular sieves (4 Å) were added.

All reactions were performed at room temperature under ambient conditions. The reactions were monitored by thin-layer chromatography (TLC) using aluminum sheets recoated with silica gel and a fluorescent indicator (60 F<sub>254</sub>, 0.2 mm). The compounds were visualized at 254 nm by employment of UV light. The products **3** were isolated by flash chromatography using silica gel (0.06-0.2 mm) as stationary phase and mixtures of commercial dichloromethane/ethyl acetate as eluent.

The chiral HPLC analysis of products **3** was performed in a Waters 600 equipment, using a Daicel ChiralPak IC column as stationary phase and mixtures of commercial *n*-hexane/isopropyl alcohol as eluent. The specific rotation of products **3** was determined using a Jasco P-1020 polarimeter, in acetonitrile or tetrahydrofuran (both HPLC grade) as solvent. The absolute configuration of products **3a**<sup>1</sup> and **3d**<sup>2</sup> was assigned comparing their specific rotation with those reported in the literature. The same absolute configuration is assumed for the rest of products **3**.

The <sup>1</sup>H-NMR and <sup>13</sup>C-NMR (APT) spectra of reagents and products were recorded at 300 MHz (Bruker ARX300 spectrometer) or 400 MHz (Bruker AV400 spectrometer), in chloroform-*d* (CDCl<sub>3</sub>) or dimethyl sulfoxide-*d*<sub>6</sub> ((CD<sub>3</sub>)<sub>2</sub>SO) as deuterated solvent. Infrared spectra of starting materials and products were obtained employing an attenuated total reflection infrared (ATR-FTIR) in a PerkinElmer FTIR spectrometer equipped with a universal ATR sampling accessory. The HRMS analysis of reagents and products was performed using a MicroTof-Q mass spectrometer and electrospray (ESI) as ionization method. Melting point of reagents and products was determined employing a Gallenkamp MPD 350 BM 2.5 device.

The spectroscopic data recorded for synthesized starting materials **1b**,<sup>3</sup> **1c**,<sup>3</sup> **1d**,<sup>3</sup> **1e**,<sup>3</sup> and **1f**,<sup>3</sup> as well as products obtained **3a**,<sup>4</sup> **3b**,<sup>4</sup> **3c**,<sup>1b</sup> **3e**,<sup>1b</sup> **3f**,<sup>5</sup> **3g**,<sup>6</sup> **3h**,<sup>7</sup> **3i**<sup>6</sup> and **3j**<sup>6</sup> are in agreement with values previously reported by other authors.

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<sup>1</sup> a) West, B. D.; Preis, S.; Schroeder, C. H.; Link, K. P. *J. Am. Chem. Soc.* **1961**, *83*, 2676-2679; b) Shi, T.; Guo, Z.; Yu, H.; Xie, J.; Zhong, Y.; Zhu, W. *Adv. Synth. Catal.* **2013**, *355*, 2538-2543.

<sup>2</sup> Dong, Z.; Wang, L.; Chen, X.; Liu, X.; Lin, L.; Feng, X. *Eur. J. Org. Chem.* **2009**, 5192-5197.

<sup>3</sup> Stern, T.; Rückbrod, S.; Czekelius C.; Donner, C.; Brunner, H. *Adv. Synth. Catal.* **2010**, *352*, 1983-1992.

<sup>4</sup> Halland, N.; Hansen, T.; Jørgensen, K. A. *Angew. Chem. Int. Ed.* **2003**, *42*, 4955-4957.

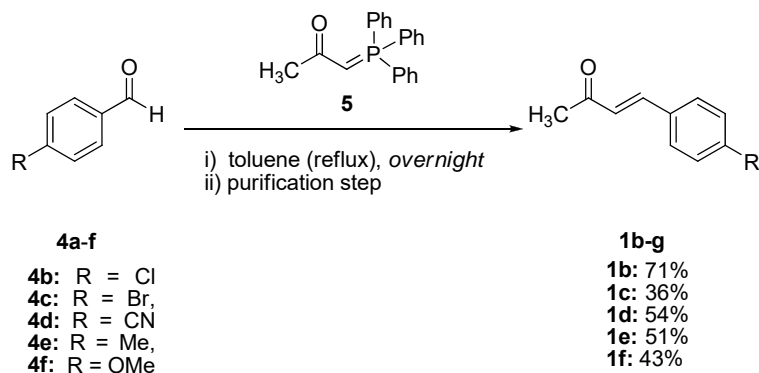
<sup>5</sup> Yang, H.-M.; Li, L.; Jiang, K.-Z.; Jiang, J.-X.; Lai, G.-Q.; Xu, L.-W. *Tetrahedron* **2010**, *66*, 9708-9713.

<sup>6</sup> Modrocká, V.; Veverková, E.; Mečiarová, M.; Šebesta, R. *J. Org. Chem.* **2018**, *83*, 13111-13120.

<sup>7</sup> Xie, J.-W.; Yue, L.; Chen, W.; Du, W.; Zhu, J.; Deng, J.-G.; Chen, Y.-C. *Org. Lett.* **2007**, *9*, 413-415.

## 2. Synthesis of benzylideneacetone derivatives **1b-f**

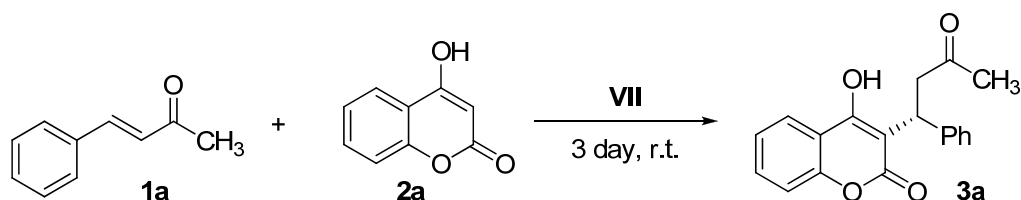
The substrates **1b-f** were synthesized from the corresponding commercial aldehydes **4b-f** and the phosphonium ylide **5**, which was previously prepared following the procedure reported in the literature using commercially available reagents.<sup>8</sup> The respective yields after purification are collected in the Scheme S1. The ratio (E)/(Z) of products **1b-f** was determined by <sup>1</sup>H-RMN spectroscopy, using DMSO-*d*<sub>6</sub> as solvent.



**Scheme S1.** Synthesis of electrophiles **1b-f**.

## 3. Screening of the reaction conditions

**Table S1.** Screening of the reaction conditions using catalyst VII.<sup>a</sup>



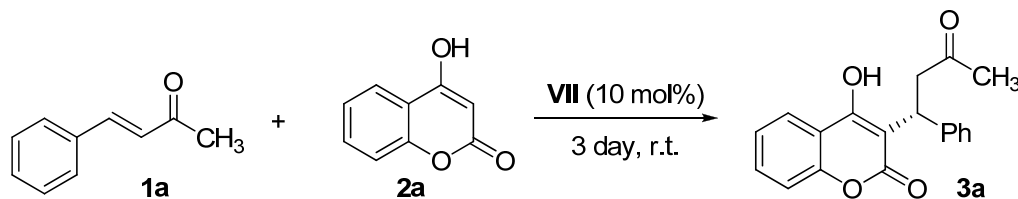
Entry	THF (μL)	Equiv. <b>1a</b>	Equiv. <b>2a</b>	<b>VII</b>	Yield (%) <sup>b</sup>	ee (%) <sup>c</sup>
1	200	1.2	1	20	19	68
2	200	1.2	1	10	10	68
3	200	1	1	10	13	70
4	200	1	1.2	10	15	68
5	200	1	1.5	10	20	68
6	200	1	2	10	22	67
7	200	1	1.5	20	58	66
8	200	1	2	20	30	68
9	100	1.2	1	20	12	66
10	100	1	1.5	20	66	63

<sup>8</sup> Vicente, J.; Chicote, M. T.; Saura-Llamas, I. *J. Chem. Ed.* **1993**, *70*, 163-164.

11	100	1	2	20	68	64
12	100	1	1.2	10	42	64

<sup>a</sup> To a mixture of catalyst **VII** (10-20 mol%) and coumarin **2a** (0.1-0.2 mmol) in THF (100-200  $\mu$ L), benzylideneacetone **1a** (0.1-0.12 mmol) was added at room temperature. <sup>b</sup> After isolation by column chromatography. <sup>c</sup> Determined by chiral HPLC analysis (Chiralpak IC, Hex:*i*PrOH 80:20, 1ml/min).

**Table S2. Screening of solvents.**



Entry	Equiv. <b>1a</b>	Equiv. <b>2a</b>	Solvent (200 $\mu$ L)	Yield (%) <sup>a</sup>	ee (%) <sup>b</sup>
1	1.0	1.2	Toluene	n.r.	n.d.
2	1.0	1.2	Acetonitrile	n.r.	n.d.
3	1.0	1.2	H <sub>2</sub> O	n.r.	n.d.
4	1.0	1.2	Hexane	$\geq$ 23	40
5	1.0	1.2	CH <sub>2</sub> Cl <sub>2</sub>	traces	n.d.
6	1.0	1.2	1,2-DCE	traces	n.d.
7	1.0	1.2	CHCl <sub>3</sub>	13	26
8	1.0	1.2	C1Ph	traces	n.d.
9	1.0	1.2	1,2-(Cl) <sub>2</sub> Ph	traces	n.d.
10	1.0	1.2	Diethyl ether	32	38
11	1.0	1.2	1,4-dioxane	traces	n.d.
12	1.0	1.2	AcOEt	traces	n.d.
13	1.0	1.2	DMF	$\geq$ 5	44
14	1.0	1.2	<i>i</i> PrOH	36	44

<sup>a</sup> After isolation by column chromatography. <sup>b</sup> Determined by chiral HPLC analysis (Chiralpak IC, Hex:*i*PrOH 80:20, 1ml/min).

#### 4. <sup>1</sup>H-NMR spectra of starting materials 1b-f

Figure S1. <sup>1</sup>H-NMR spectrum of compound **1b** (400 MHz, DMSO-d<sub>6</sub>)

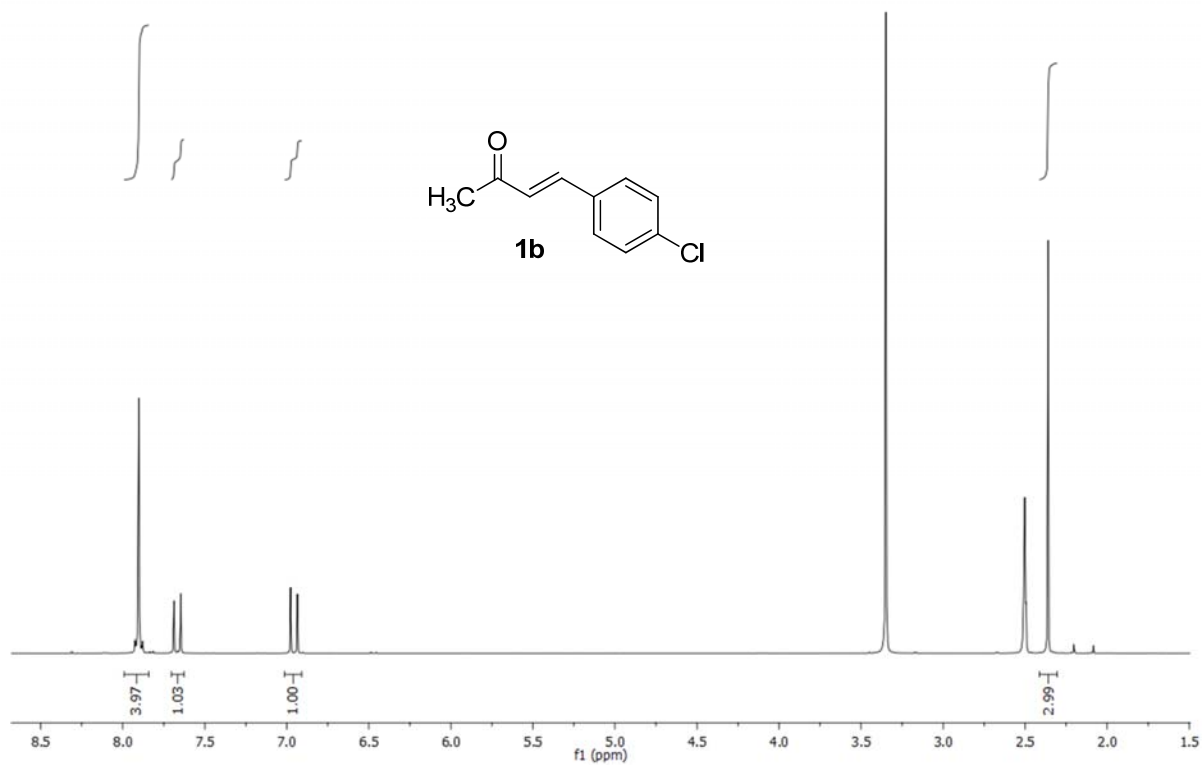
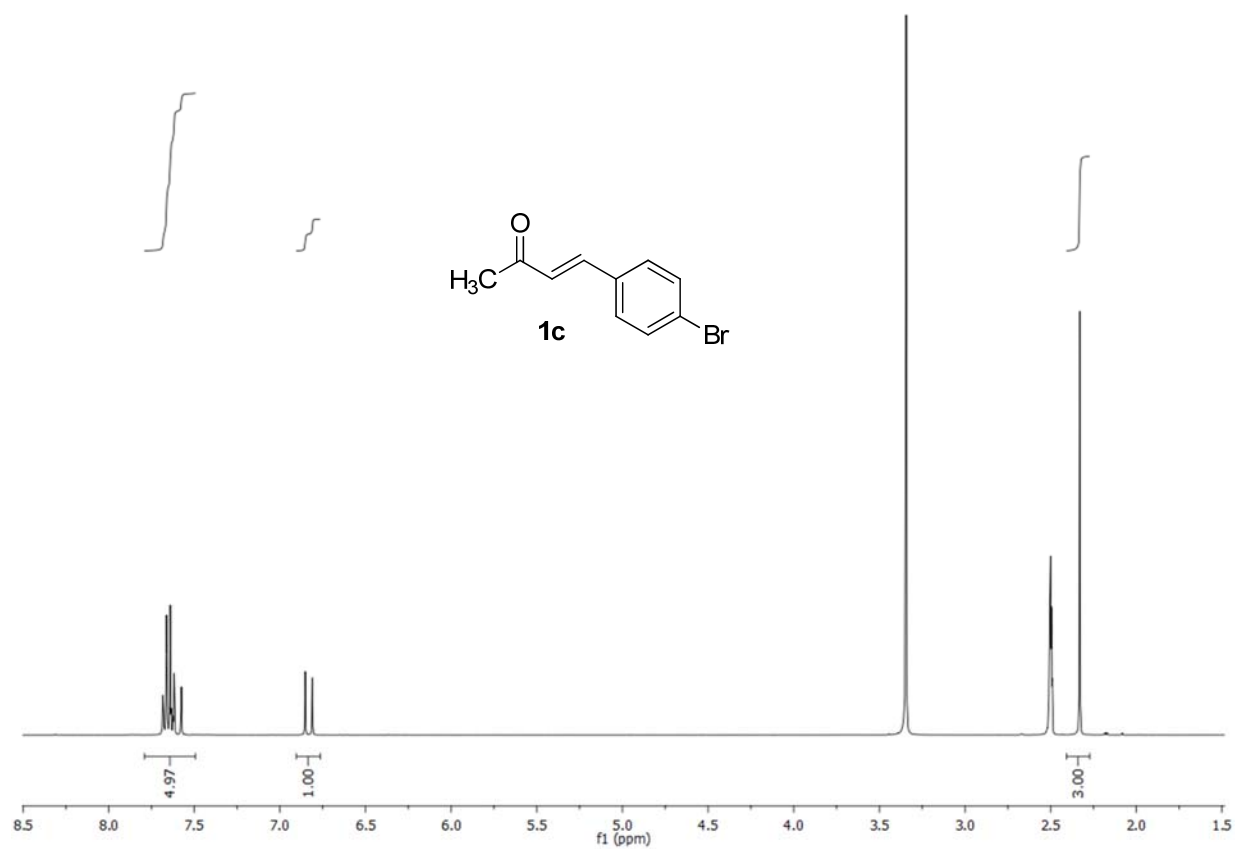
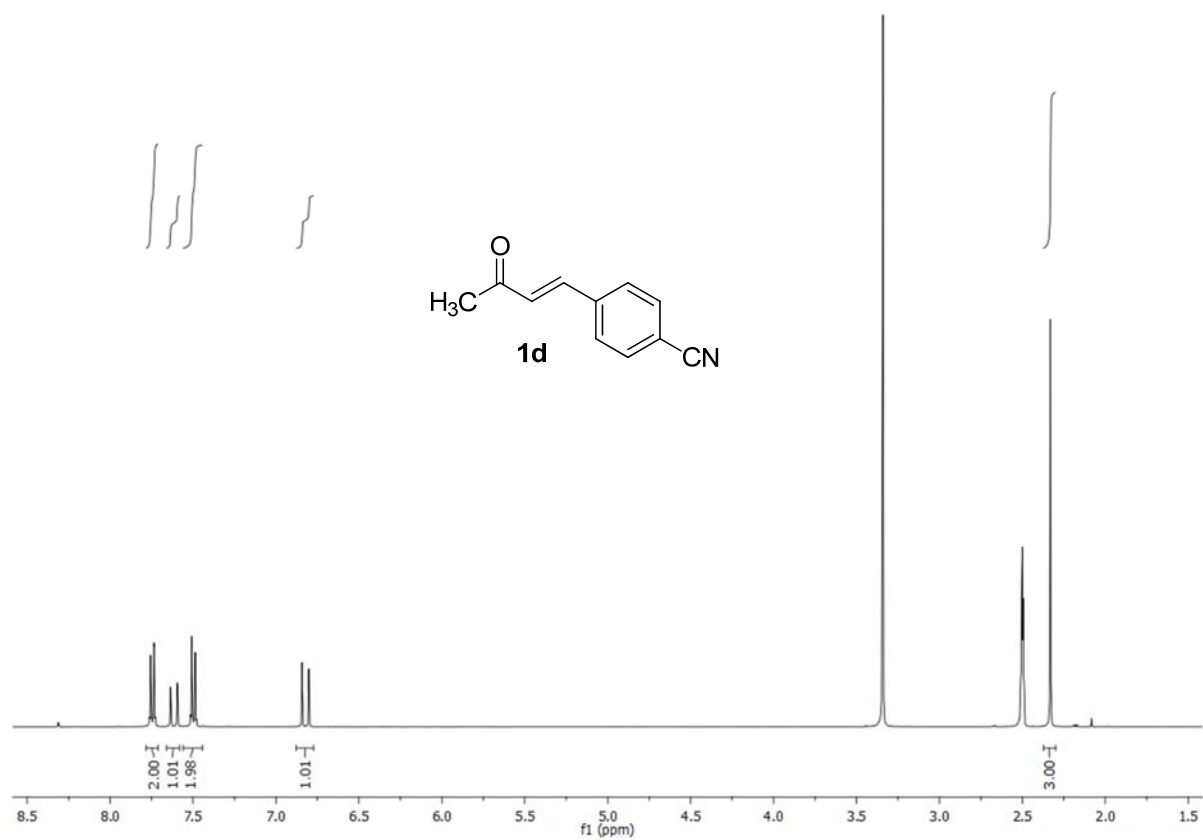


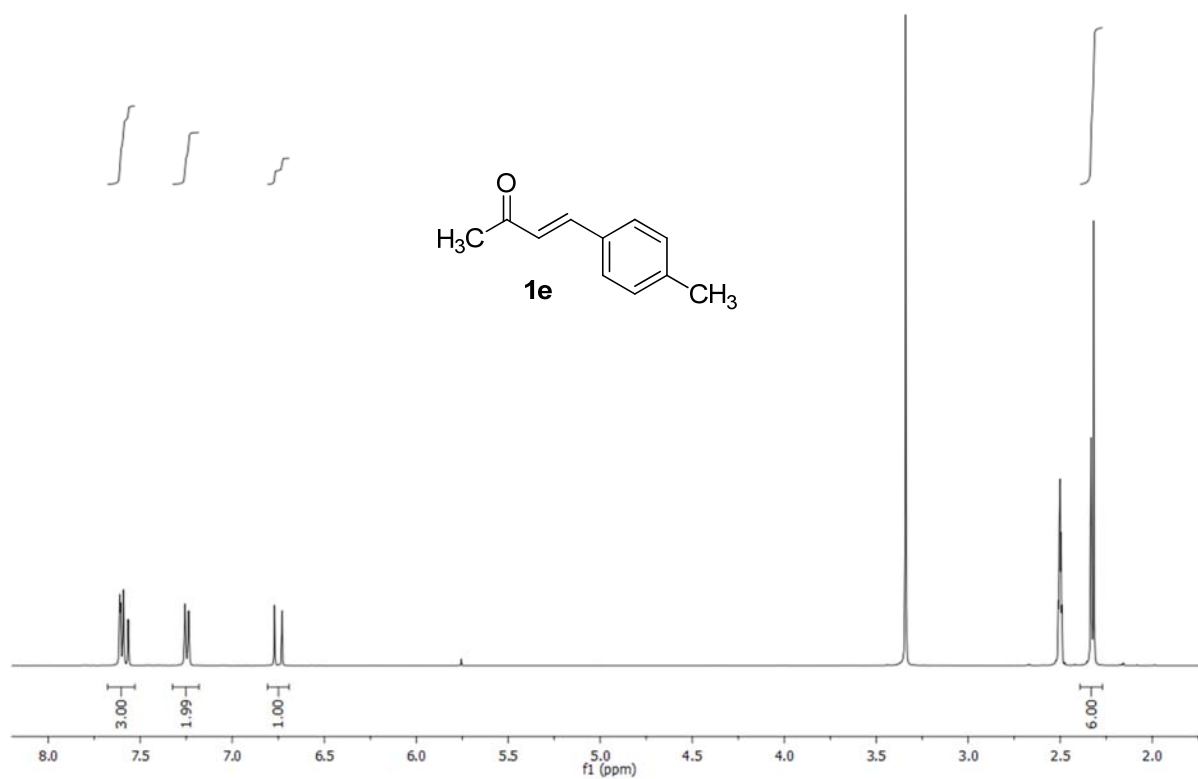
Figure S2. <sup>1</sup>H-NMR spectrum of compound **1c** (400 MHz, DMSO-d<sub>6</sub>)



**Figure S3.**  $^1\text{H-NMR}$  spectrum of compound **1d** (400 MHz,  $\text{DMSO-d}_6$ )

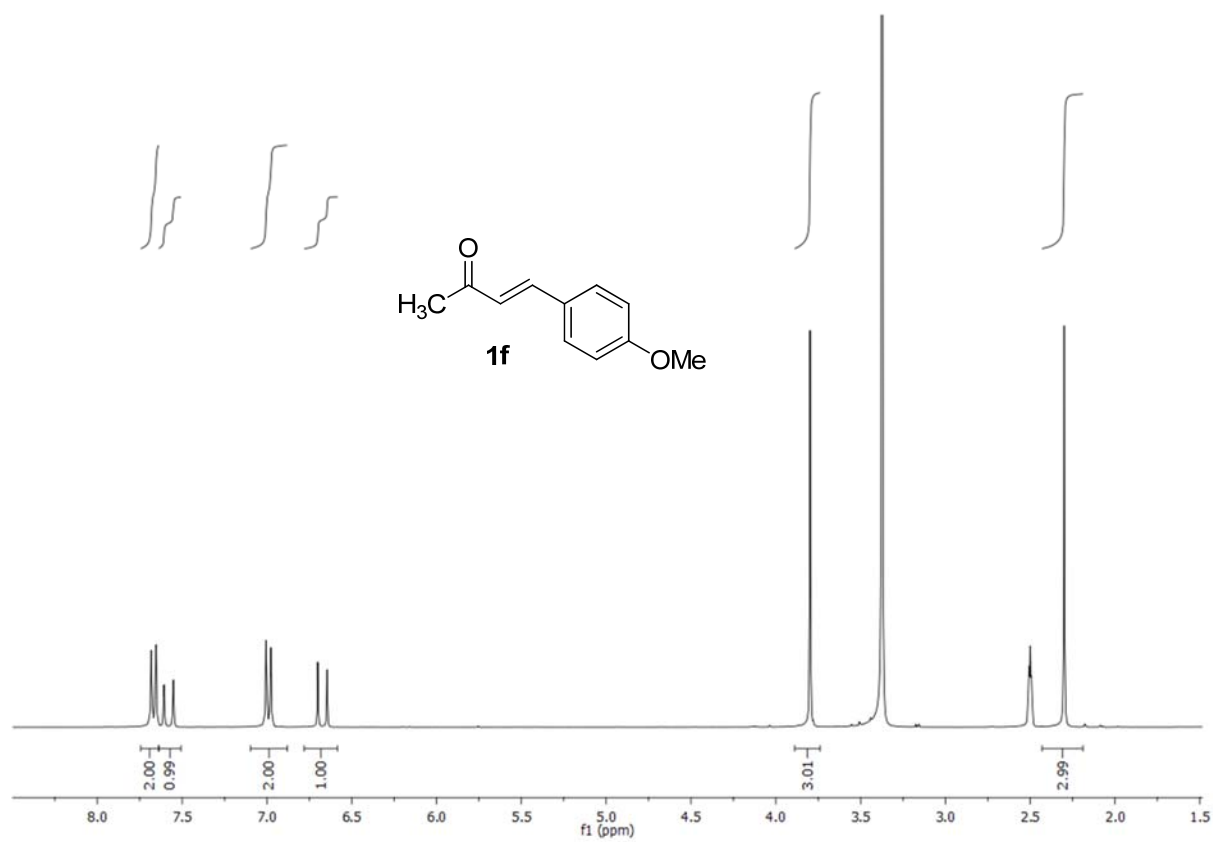


**Figure S4.**  $^1\text{H-NMR}$  spectrum of compound **1e** (400 MHz,  $\text{DMSO-d}_6$ )



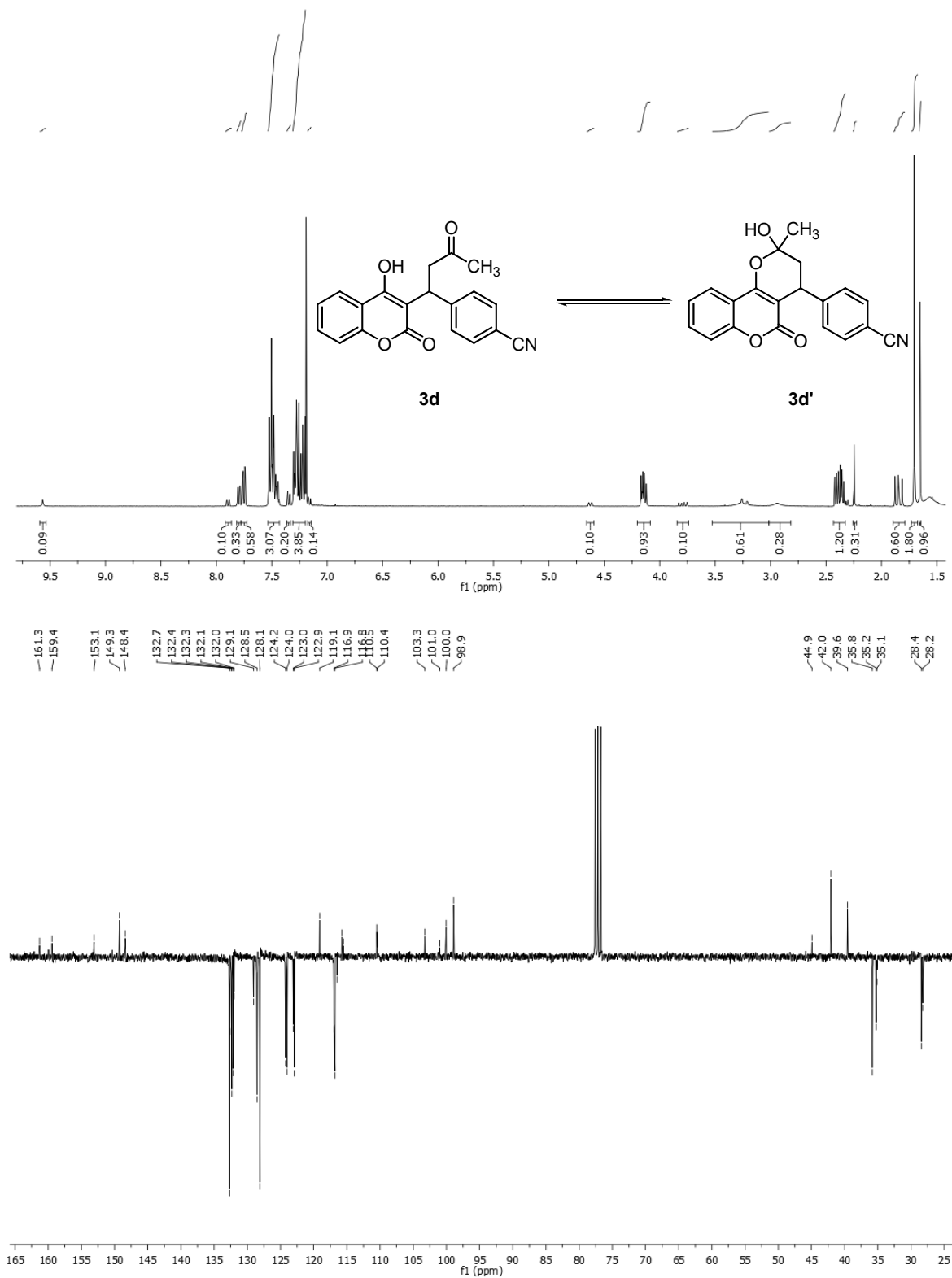


**Figure S5.**  $^1\text{H-NMR}$  spectrum of compound **1f** (400 MHz,  $\text{DMSO-d}_6$ ).

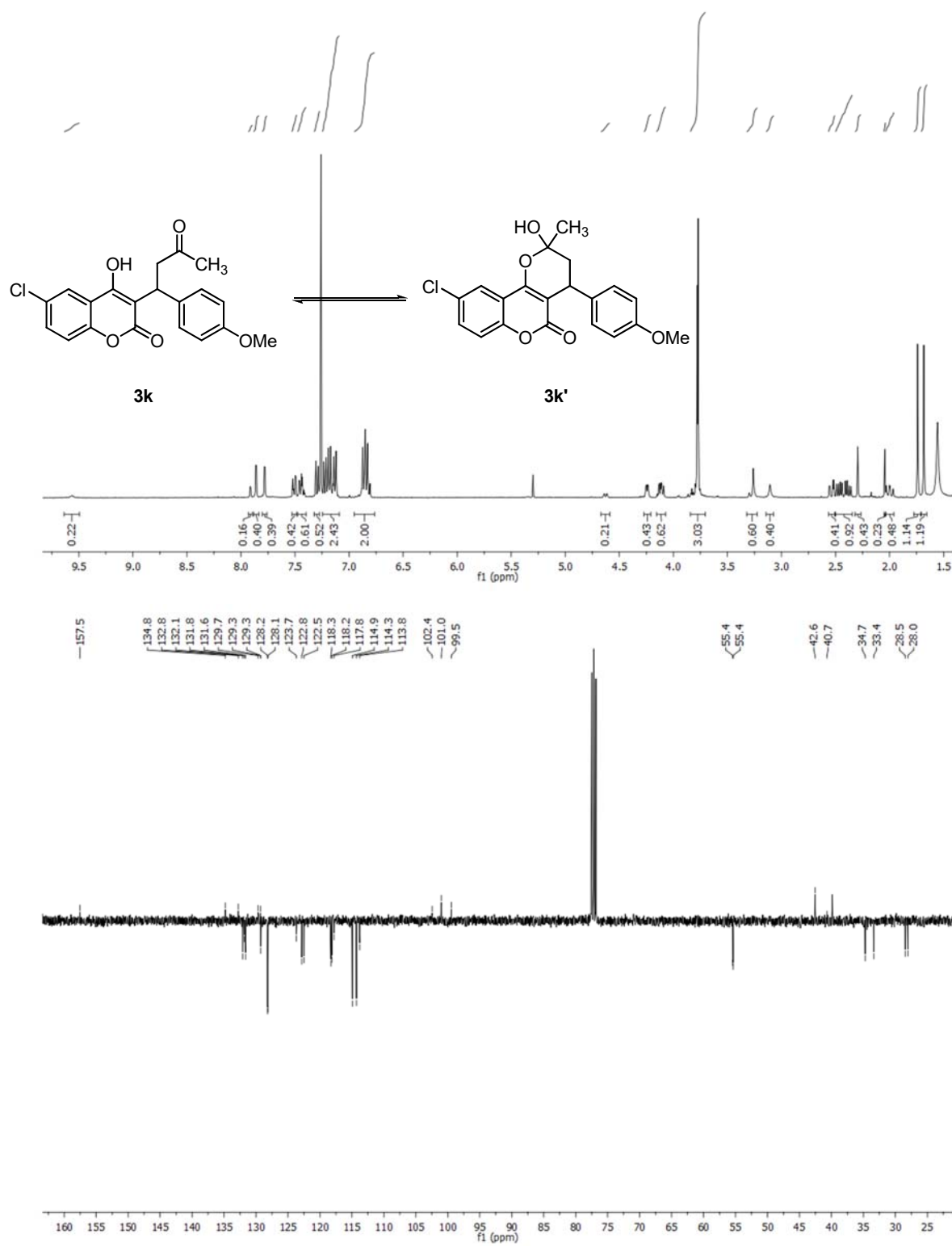


5.  $^1\text{H-NMR}$  and  $^{13}\text{C-NMR}$  (APT) spectra of products 3d,k

Figure S6.  $^1\text{H-NMR}$  and  $^{13}\text{C-NMR}$  (APT) spectra of compound 3d.

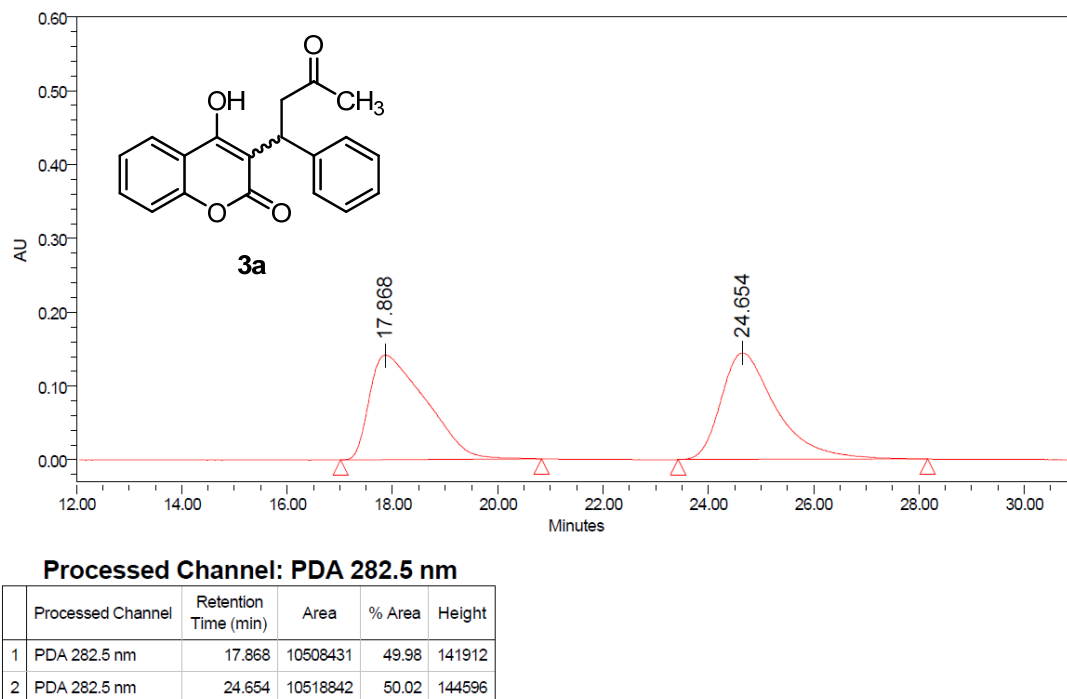


**Figure S7.**  $^1\text{H-NMR}$  and  $^{13}\text{C-NMR}$  (APT) spectra of compound **3k**.

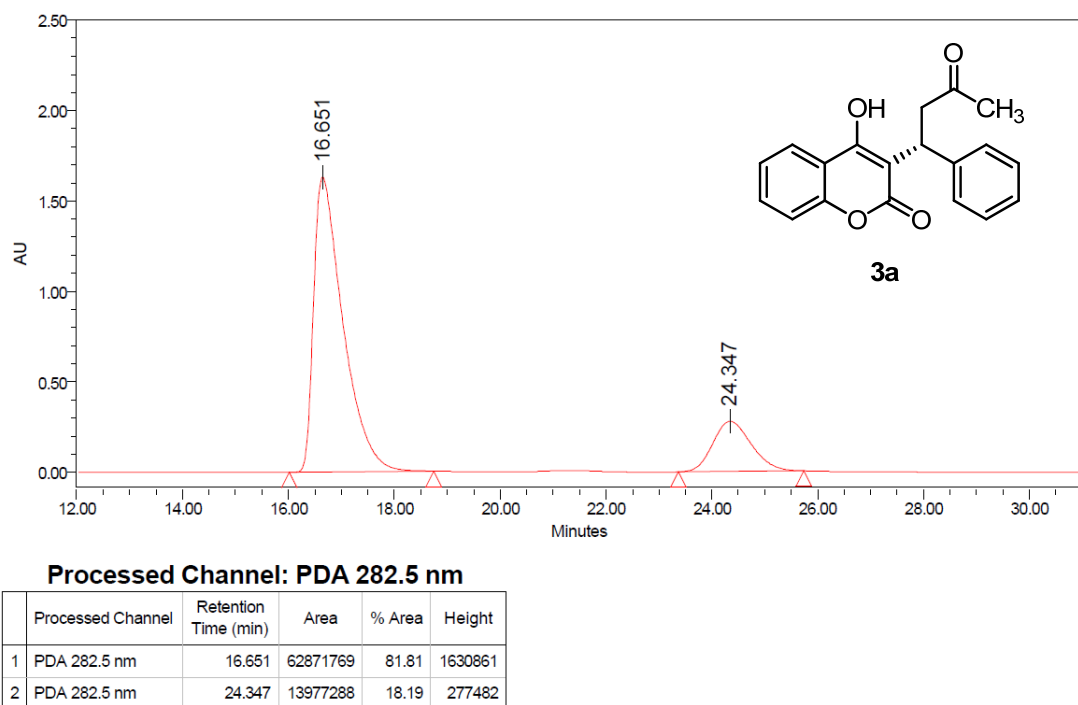


## 6. Chiral HPLC analysis of compounds 3a-l

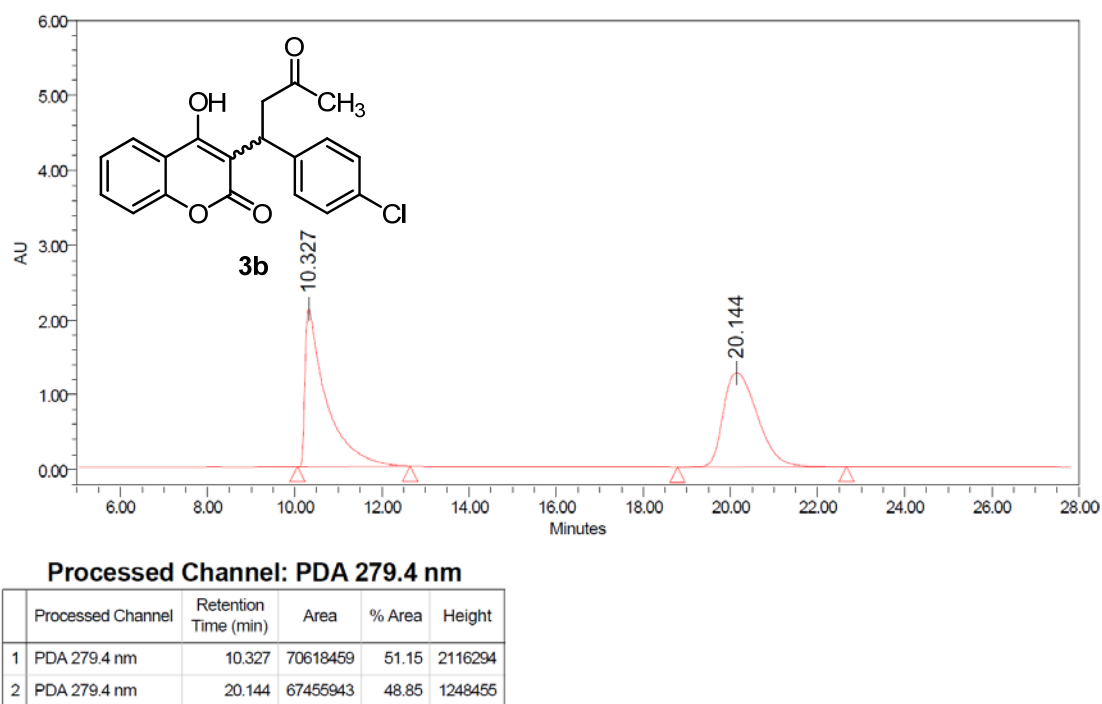
**Figure S8.** Racemic mixture of **3a**. Daicel ChiralPak IC column (*n*-hexane/isopropyl alcohol = 80:20, 1 mL min<sup>-1</sup>,  $\lambda$  = 282.5 nm).



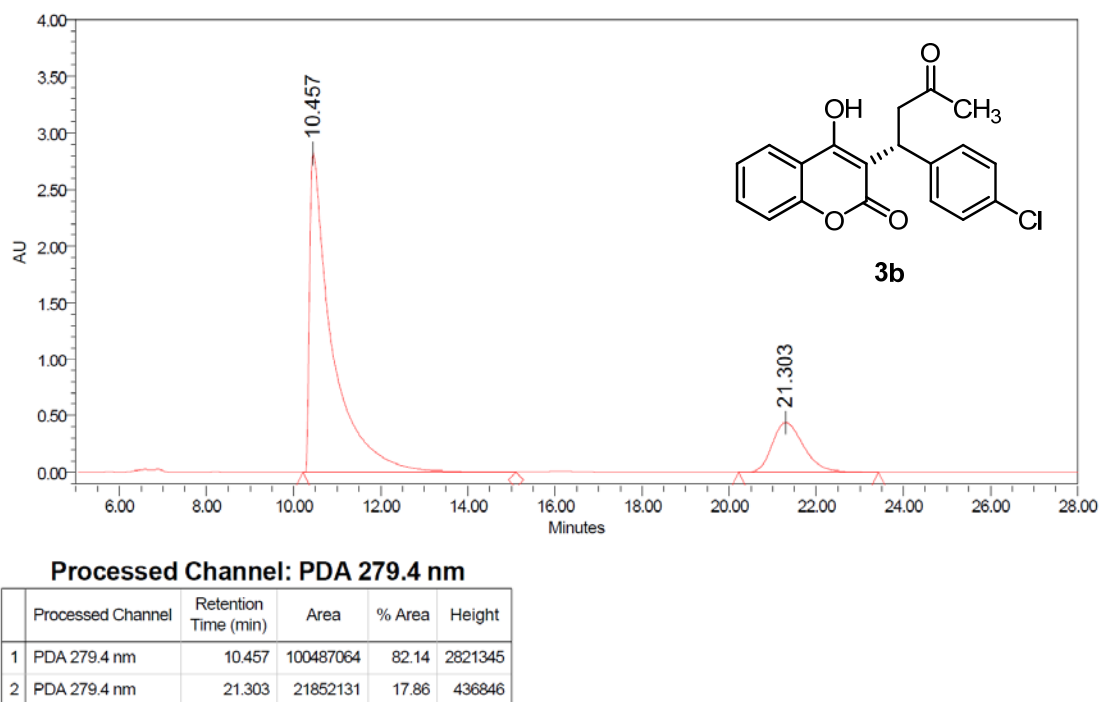
**Figure S9.** Enantioenriched mixture of **3a** (64% ee). Daicel ChiralPak IC column (*n*-hexane/isopropyl alcohol = 80:20, 1 mL min<sup>-1</sup>,  $\lambda$  = 282.5 nm).



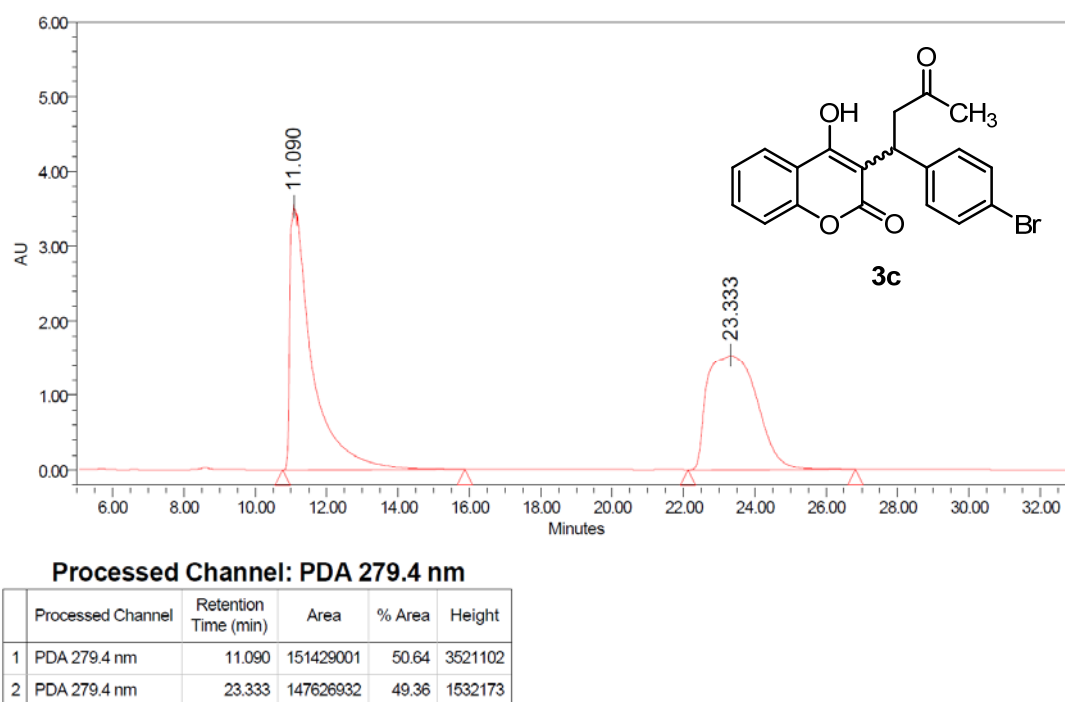
**Figure S10.** Racemic mixture of **3b**. Daicel ChiralPak IC column (*n*-hexane/isopropyl alcohol = 80:20, 1 mL min<sup>-1</sup>, λ = 279.4 nm).



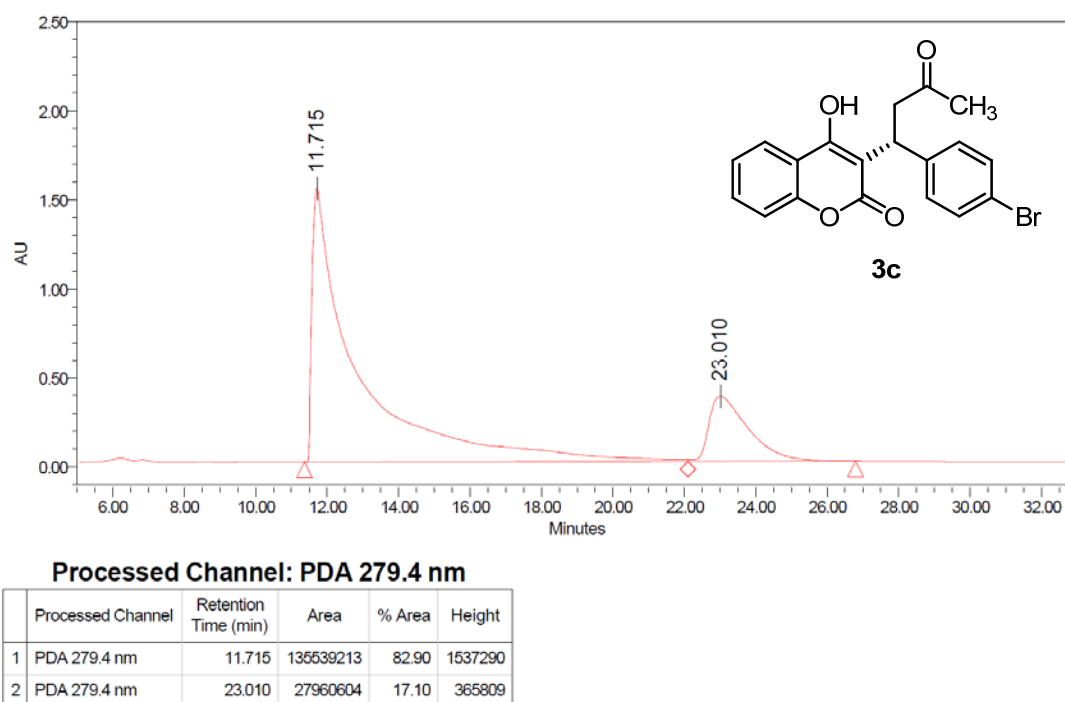
**Figure S11.** Enantioenriched mixture of **3b** (64% ee). Daicel ChiralPak IC column (*n*-hexane/isopropyl alcohol = 80:20, 1 mL min<sup>-1</sup>, λ = 279.4 nm).



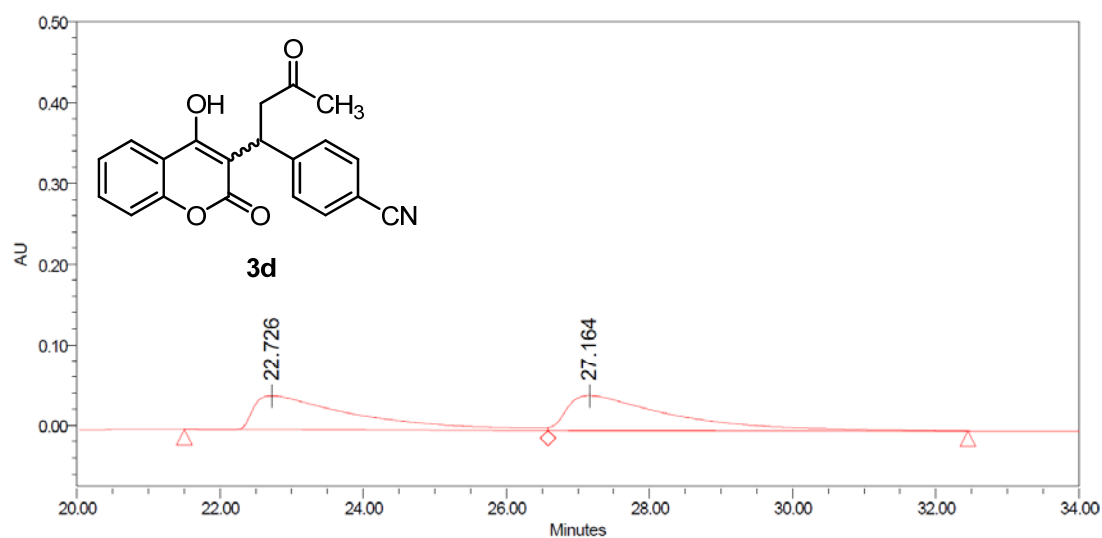
**Figure S12.** Racemic mixture of **3c**. Daicel ChiralPak IC column (*n*-hexane/isopropyl alcohol = 80:20, 1 mL min<sup>-1</sup>,  $\lambda$  = 279.4 nm).



**Figure S13.** Enantioenriched mixture of **3c** (66% ee). Daicel ChiralPak IC column (*n*-hexane/isopropyl alcohol = 80:20, 1 mL min<sup>-1</sup>,  $\lambda$  = 279.4 nm).



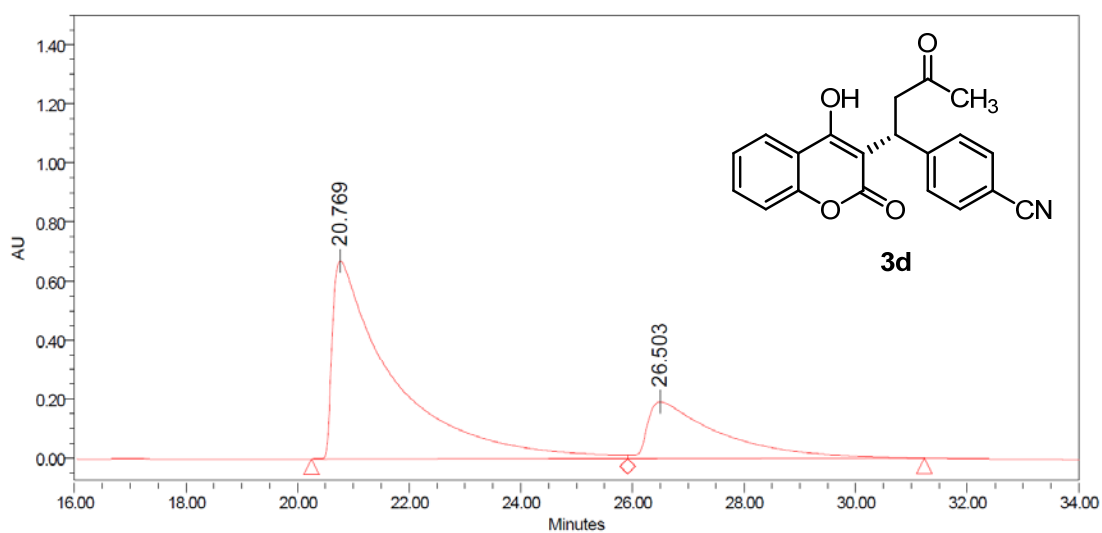
**Figure S14.** Racemic mixture of **3d**. Daicel ChiralPak IC column (*n*-hexane/isopropyl alcohol = 80:20, 1 mL min<sup>-1</sup>, λ = 250.0 nm).



**Processed Channel: PDA 250.0 nm**

	Processed Channel	Retention Time (min)	Area	% Area	Height
1	PDA 250.0 nm	22.726	4081700	48.63	41799
2	PDA 250.0 nm	27.164	4312163	51.37	42798

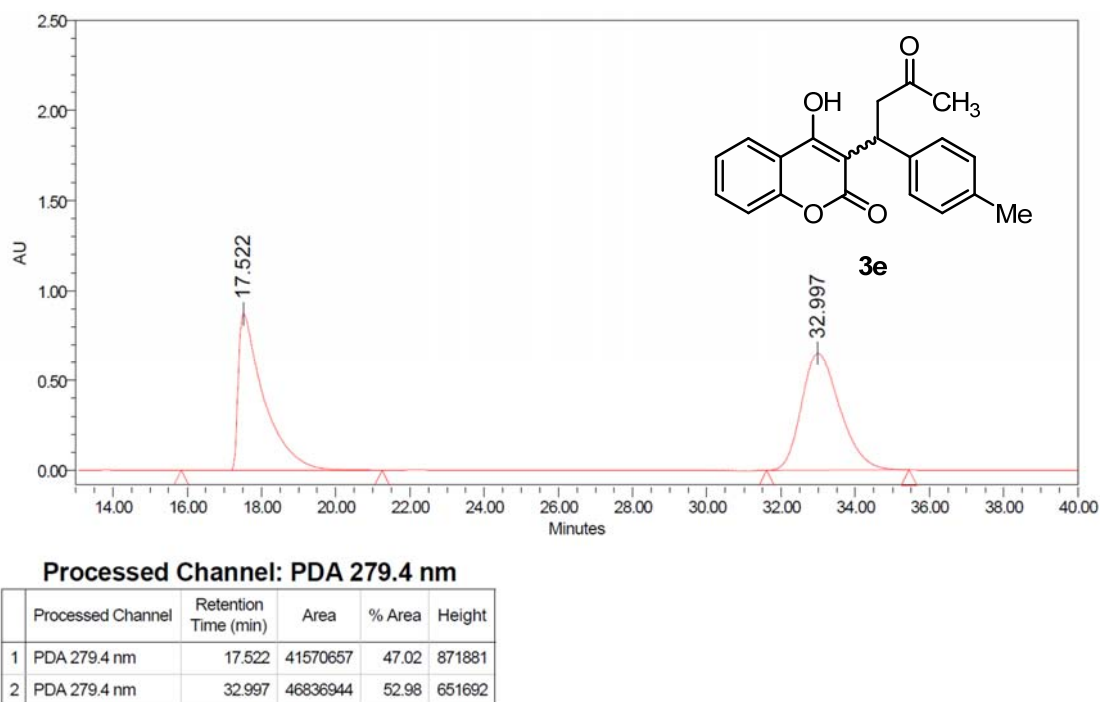
**Figure S15.** Enantioenriched mixture of **3d** (50% ee). Daicel ChiralPak IC column (*n*-hexane/isopropyl alcohol = 80:20, 1 mL min<sup>-1</sup>, λ = 250.0 nm).



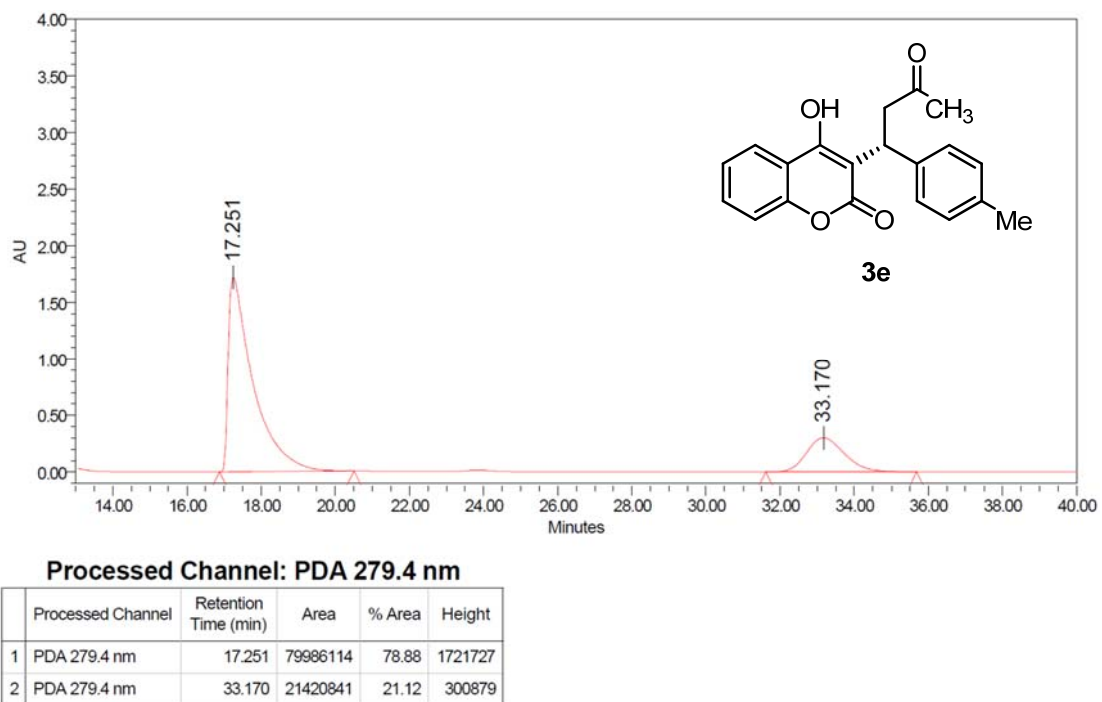
**Processed Channel: PDA 250.0 nm**

	Processed Channel	Retention Time (min)	Area	% Area	Height
1	PDA 250.0 nm	20.769	50779910	75.00	671849
2	PDA 250.0 nm	26.503	16926677	25.00	190616

**Figure S16.** Racemic mixture of **3e**. Daicel ChiralPak IC column (*n*-hexane/isopropyl alcohol = 80:20, 1 mL min<sup>-1</sup>, λ = 279.4 nm).

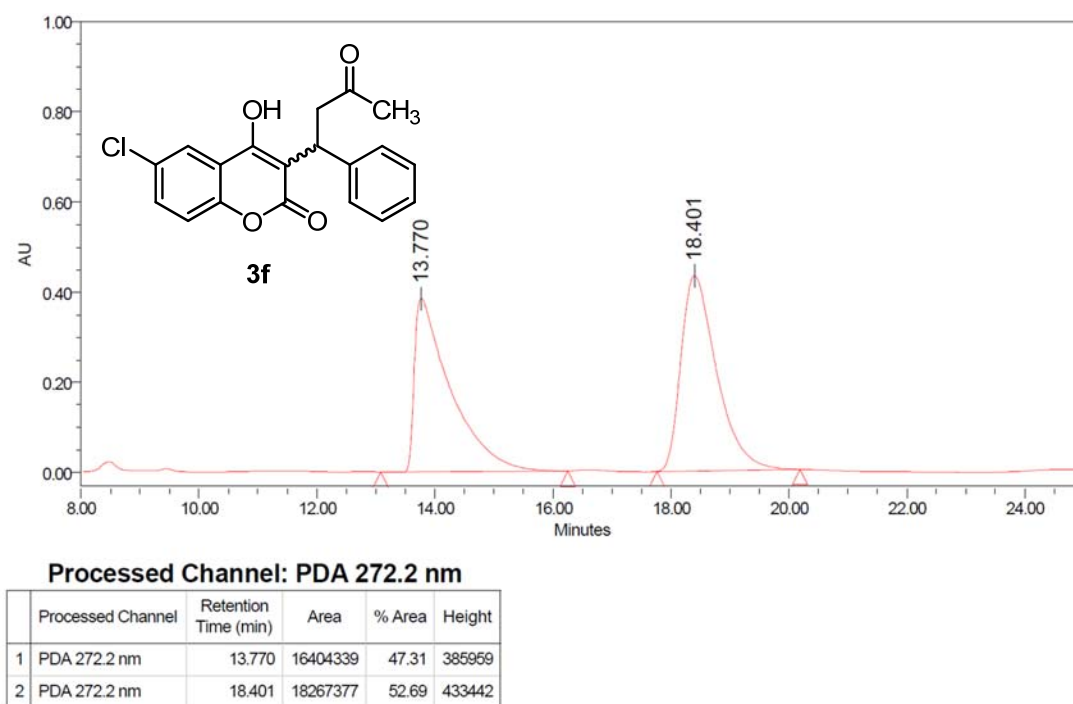


**Figure S17.** Enantioenriched mixture of **3e** (58% ee). Daicel ChiralPak IC column (*n*-hexane/isopropyl alcohol = 80:20, 1 mL min<sup>-1</sup>, λ = 279.4 nm).

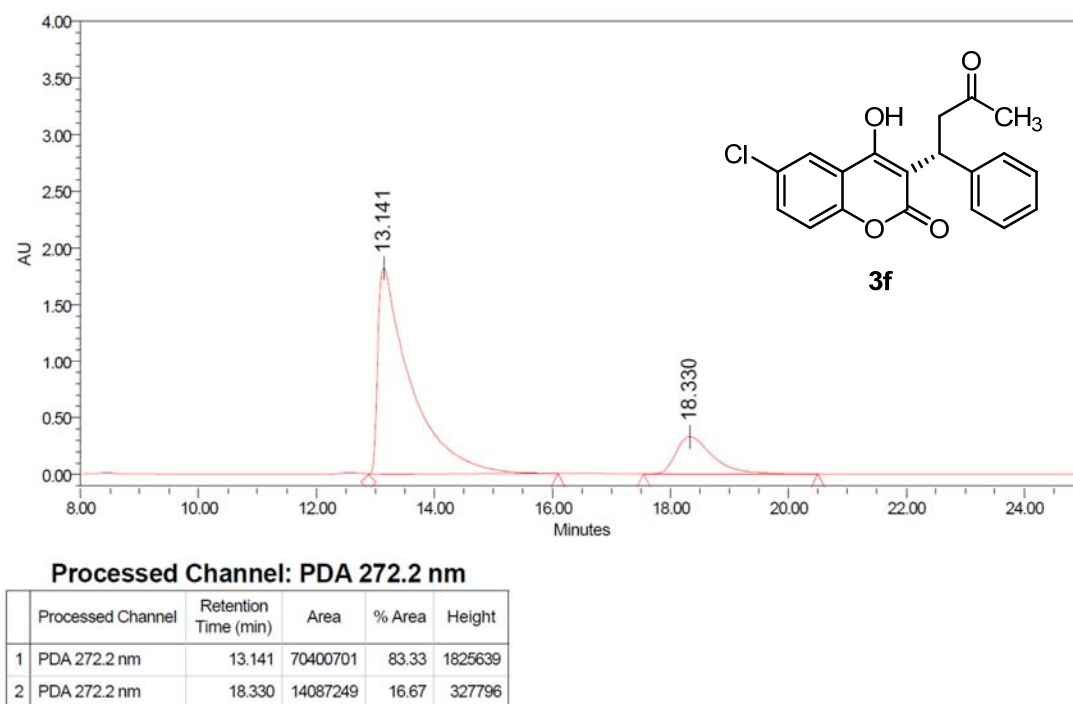




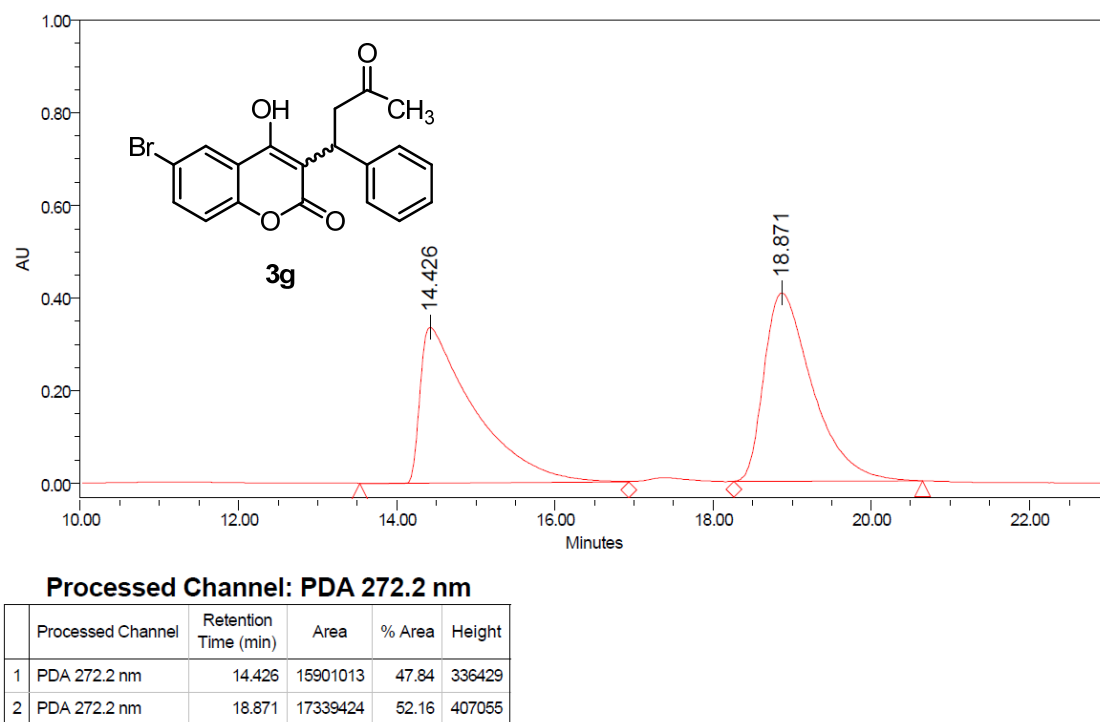
**Figure S18.** Racemic mixture of **3f**. Daicel ChiralPak IC column (*n*-hexane/isopropyl alcohol = 80:20, 1 mL min<sup>-1</sup>, λ = 272.2 nm).



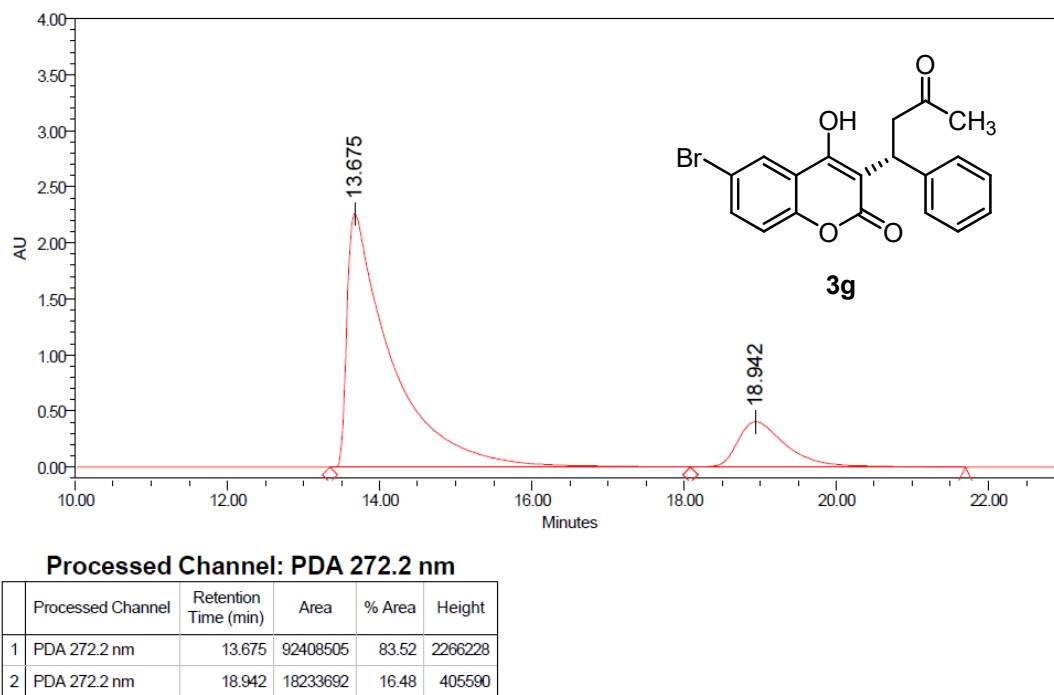
**Figure S19.** Enantioenriched mixture of **3f** (67% ee). Daicel ChiralPak IC column (*n*-hexane/isopropyl alcohol = 80:20, 1 mL min<sup>-1</sup>, λ = 272.2 nm).



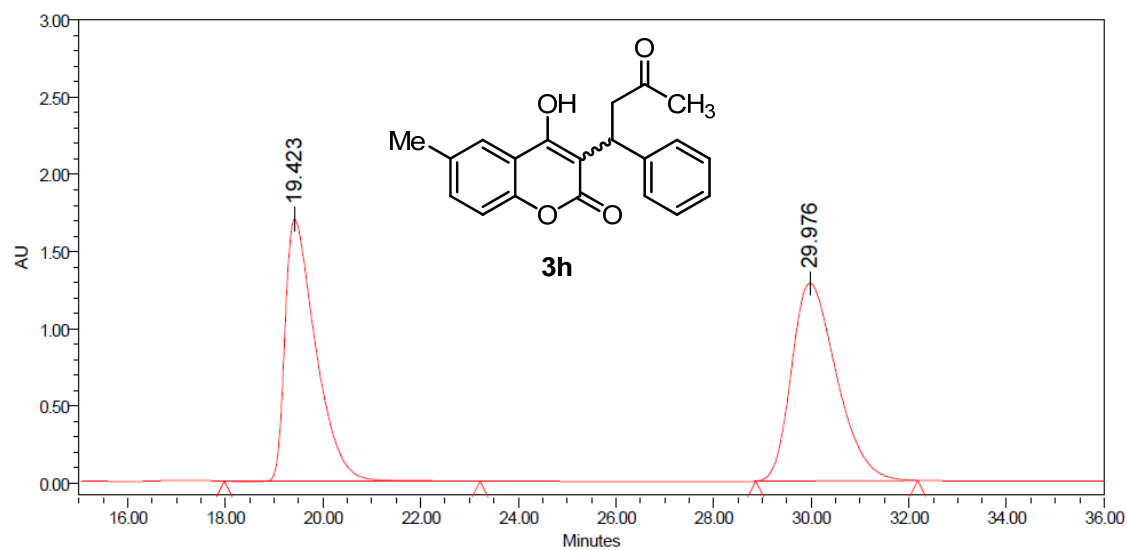
**Figure S20.** Racemic mixture of **3g**. Daicel ChiralPak IC column (*n*-hexane/isopropyl alcohol = 80:20, 1 mL min<sup>-1</sup>, λ = 272.2 nm).



**Figure S21.** Enantioenriched mixture of **3g** (67% ee). Daicel ChiralPak IC column (*n*-hexane/isopropyl alcohol = 80:20, 1 mL min<sup>-1</sup>, λ = 272.2 nm).



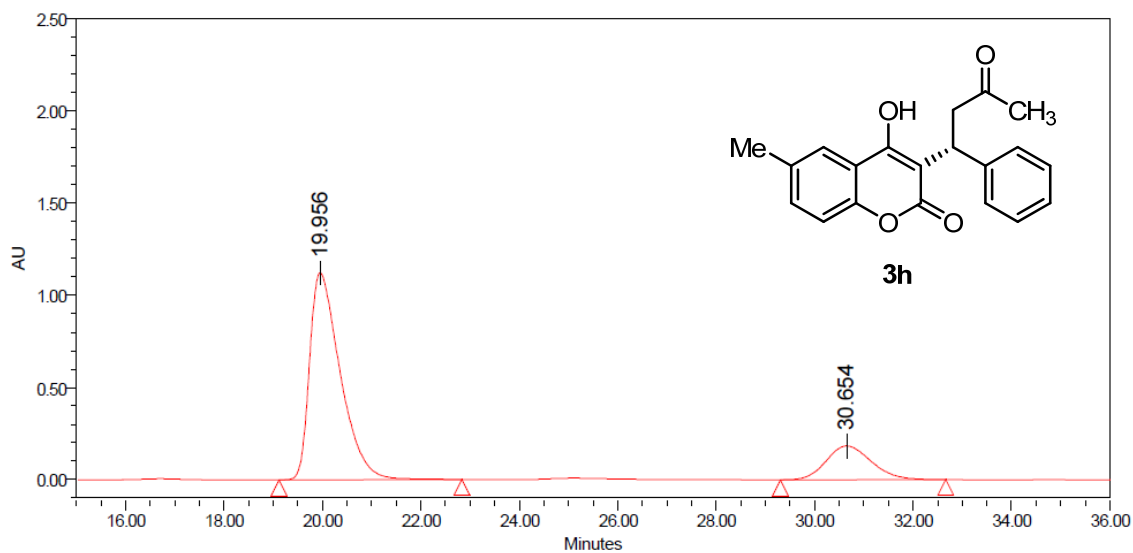
**Figure S22.** Racemic mixture of **3h**. Daicel ChiralPak IC column (*n*-hexane/isopropyl alcohol = 80:20, 1 mL min<sup>-1</sup>, λ = 271.0 nm).



Processed Channel: PDA 271.0 nm

	Processed Channel	Retention Time (min)	Area	% Area	Height
1	PDA 271.0 nm	19.423	74071932	47.42	1698641
2	PDA 271.0 nm	29.976	82145423	52.58	1280434

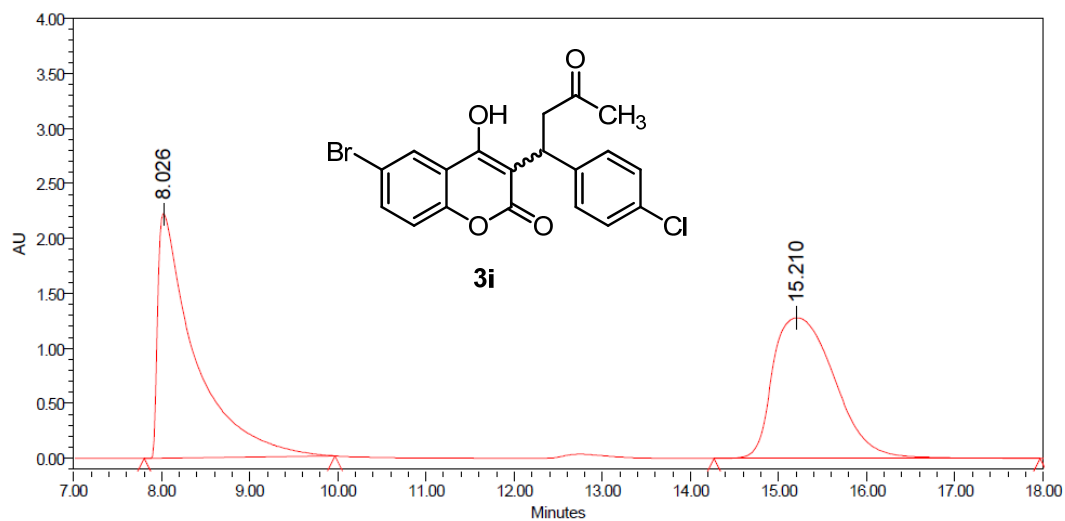
**Figure S23.** Enantioenriched mixture of **3h** (61% ee). Daicel ChiralPak IC column (*n*-hexane/isopropyl alcohol = 80:20, 1 mL min<sup>-1</sup>, λ = 271.0 nm).



Processed Channel: PDA 271.0 nm

	Processed Channel	Retention Time (min)	Area	% Area	Height
1	PDA 271.0 nm	19.956	48489860	80.62	1122777
2	PDA 271.0 nm	30.654	11655126	19.38	183148

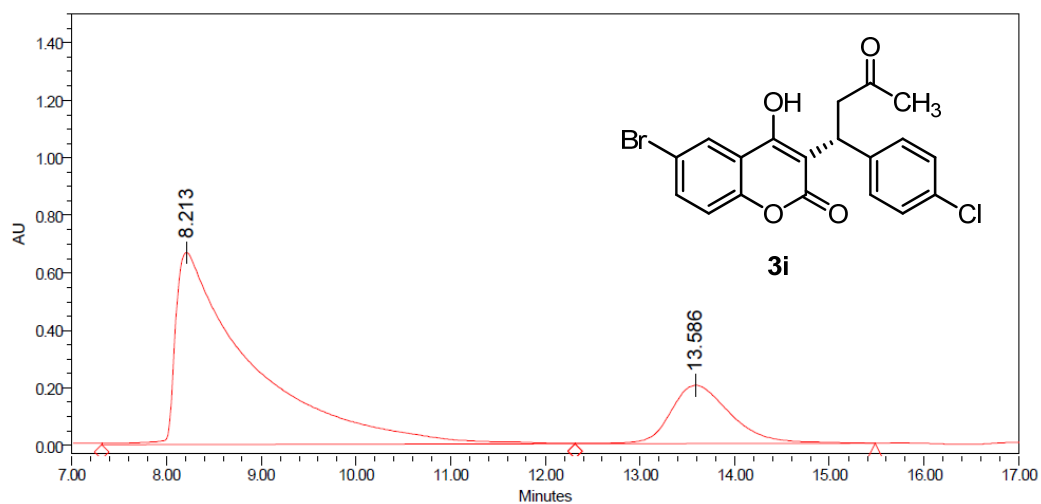
**Figure S24.** Racemic mixture of **3i**. Daicel ChiralPak IC column (*n*-hexane/isopropyl alcohol = 80:20, 1 mL min<sup>-1</sup>,  $\lambda$  = 272.2 nm).



**Processed Channel: PDA 272.2 nm**

	Processed Channel	Retention Time (min)	Area	% Area	Height
1	PDA 272.2 nm	8.026	64255438	50.75	2233890
2	PDA 272.2 nm	15.210	62364144	49.25	1277353

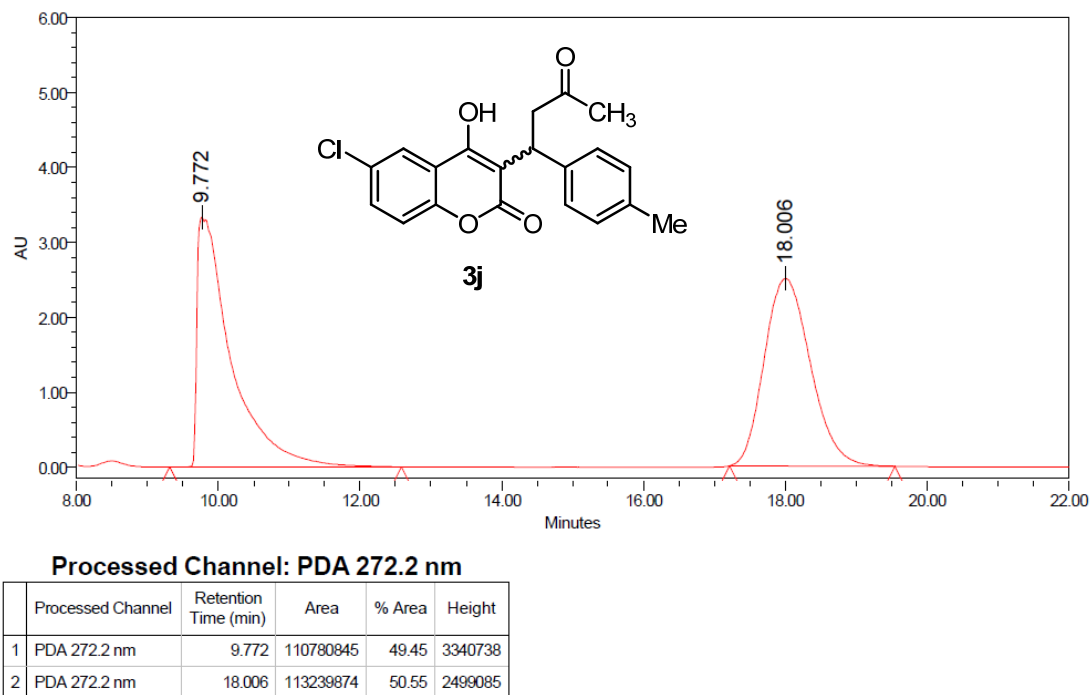
**Figure S25** Enantioenriched mixture of **3i** (62% ee). Daicel ChiralPak IC column (*n*-hexane/isopropyl alcohol = 80:20, 1 mL min<sup>-1</sup>,  $\lambda$  = 272.2 nm).



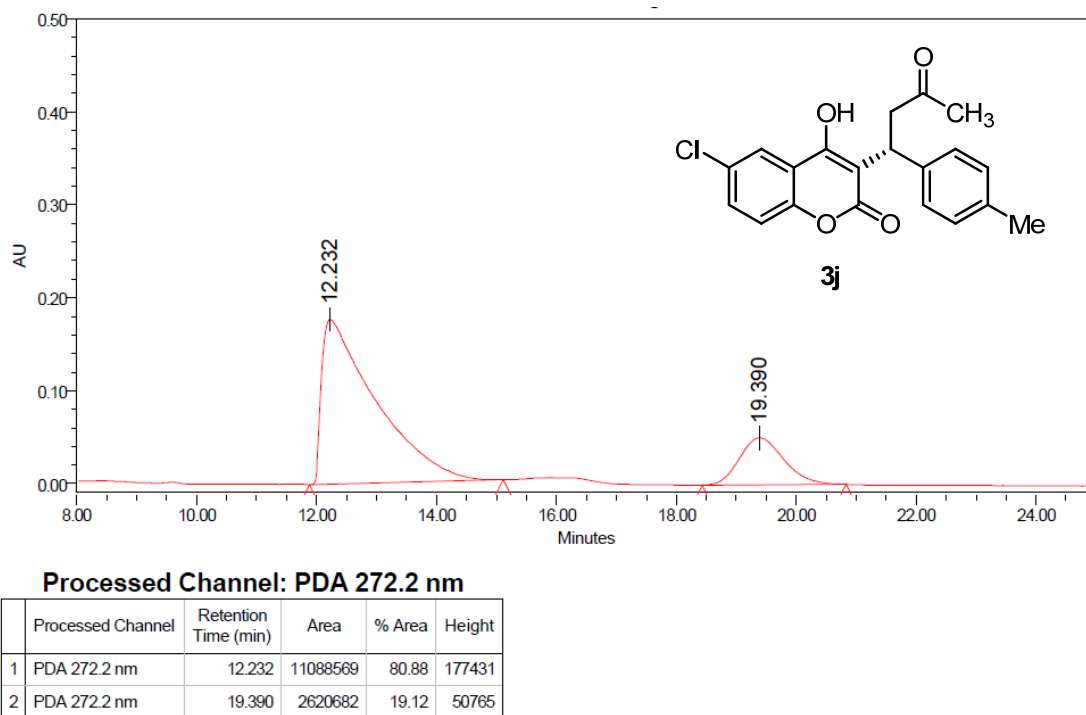
**Processed Channel: PDA 272.2 nm**

	Processed Channel	Retention Time (min)	Area	% Area	Height
1	PDA 272.2 nm	8.213	37159642	80.81	667403
2	PDA 272.2 nm	13.586	8824184	19.19	201837

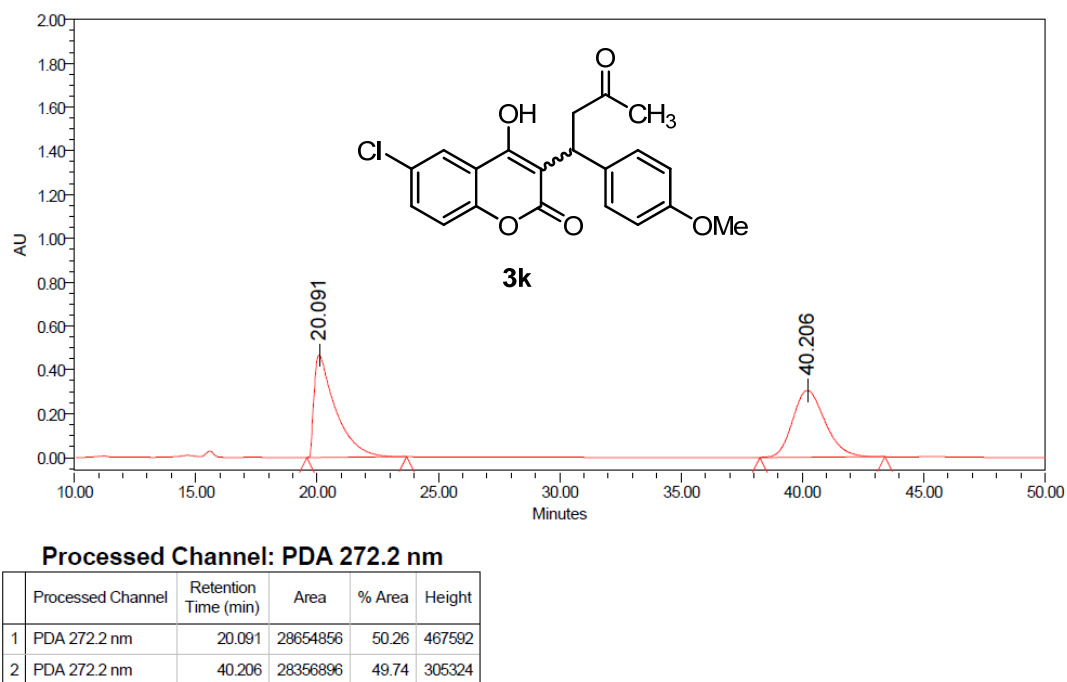
**Figure S26.** Racemic mixture of **3j**. Daicel ChiralPak IC column (*n*-hexane/isopropyl alcohol = 80:20, 1 mL min<sup>-1</sup>,  $\lambda$  = 272.2 nm).



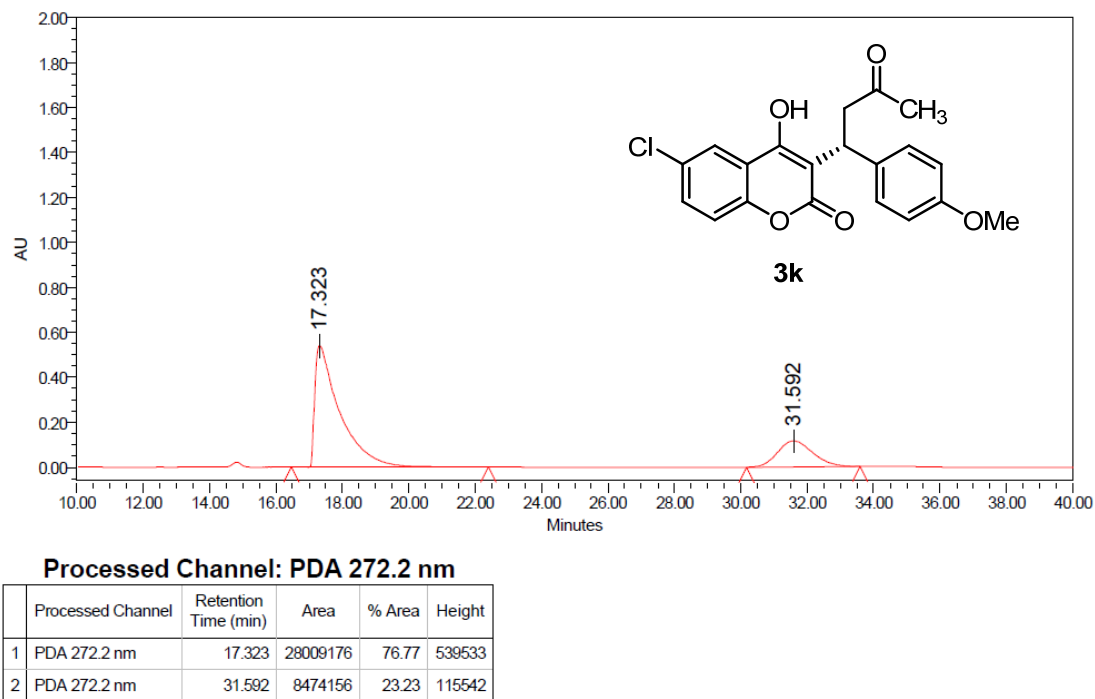
**Figure S27.** Enantioenriched mixture of **3j** (62% ee). Daicel ChiralPak IC column (*n*-hexane/isopropyl alcohol = 80:20, 1 mL min<sup>-1</sup>,  $\lambda$  = 272.2 nm).



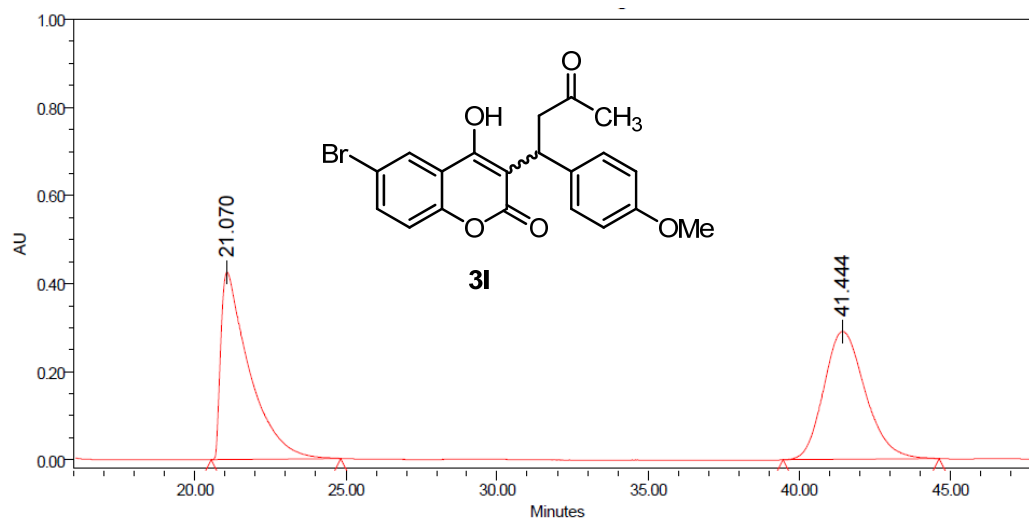
**Figure S28.** Racemic mixture of **3k**. Daicel ChiralPak IC column (*n*-hexane/isopropyl alcohol = 80:20, 1 mL min<sup>-1</sup>,  $\lambda$  = 272.2 nm).



**Figure S29.** Enantioenriched mixture of **3k** (54% ee). Daicel ChiralPak IC column (*n*-hexane/isopropyl alcohol = 80:20, 1 mL min<sup>-1</sup>,  $\lambda$  = 272.2 nm).



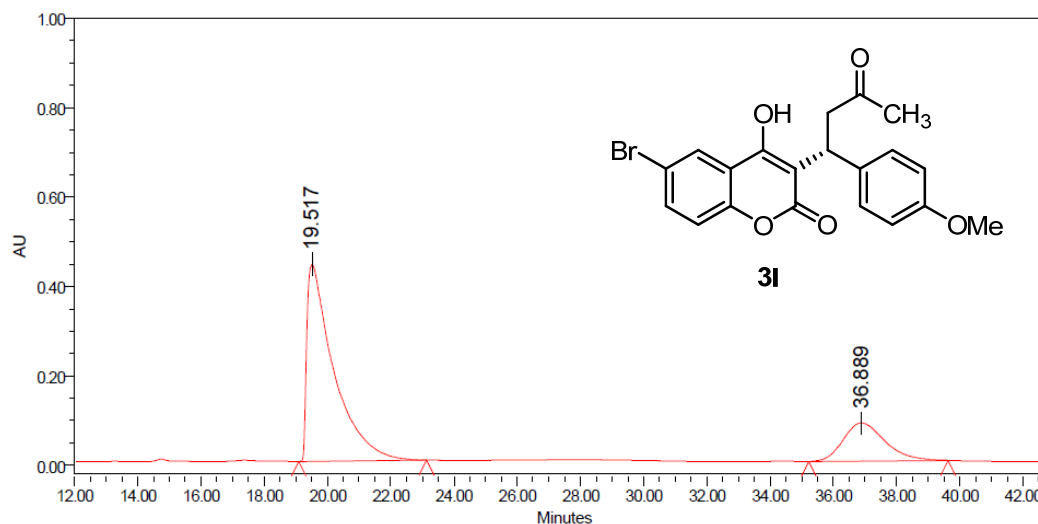
**Figure S30.** Racemic mixture of **3I**. Daicel ChiralPak IC column (*n*-hexane/isopropyl alcohol = 80:20, 1 mL min<sup>-1</sup>,  $\lambda$  = 272.2 nm).



Processed Channel: PDA 272.2 nm

	Processed Channel	Retention Time (min)	Area	% Area	Height
1	PDA 272.2 nm	21.070	28057165	50.25	425860
2	PDA 272.2 nm	41.444	27774925	49.75	290366

**Figure S31.** Enantioenriched mixture of **3I** (54% ee). Daicel ChiralPak IC column (*n*-hexane/isopropyl alcohol = 80:20, 1 mL min<sup>-1</sup>,  $\lambda$  = 272.2 nm).



Processed Channel: PDA 272.2 nm

	Processed Channel	Retention Time (min)	Area	% Area	Height
1	PDA 272.2 nm	19.517	25978463	76.86	440514
2	PDA 272.2 nm	36.889	7820214	23.14	85308