Supporting information

A naphthalimide-indole fused chromophore-based fluorescent probe for detection of biothiol with a red emission and a large Stokes shift

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Fig. S1. Left: The normalized absorption and emission spectra of Compound 4 in PBS buffer (10.0 mM, 1.0 mM CTAB, pH = 7.4). Right: The absorption and emission spectra of probe 1 in the absence and presence of Cys, Hcy and GSH in PBS buffer (10.0 mM, 1.0 mM CTAB, pH = 7.4).



Fig. S2. (a) Fluorescence spectra of Probe **1** (10.0 μ M) after reacting with Cys (0.0-3.5 equiv.) in PBS buffer (10.0 mM, 1.0 mM CTAB, pH = 7.4) for 4 min at 25 °C (λ_{ex} = 488 nm). (b) Fluorescence intensity at 590 nm of Probe **1** (10.0 μ M) against Cys concentration. Inset: the linear correlation between fluorescence intensity at 590 nm and the concentration of Cys (0-2.5 μ M).



Fig. S3. (a) Fluorescence spectra of Probe **1** (10.0 μ M) and after reacting with Hcy (0.0-3.5 equiv.) in PBS buffer (10.0 mM, 1.0 mM CTAB, pH = 7.4) for 4 min at 25 °C (λ_{ex} = 488 nm). (b) Fluorescence intensity at 590 nm of Probe **1** (10.0 μ M) against Hcy concentration. Inset: the linear correlation between fluorescence intensity at 590 nm and the concentration of Hcy (0-3.5 μ M).



Fig. S4. Fluorescence spectra of Probe **1** (10.0 μ M) towards GSH (3.5 equiv.) with other relevant species (3.5 equiv.) in PBS buffer (10.0 mM, 1.0 mM CTAB, pH = 7.4). The spectra were recorded after reacting 4 min at 25 °C. λ_{em} = 590 nm, λ_{ex} = 488 nm. (1. Arg, 2. Ile, 3. Ser, 4. His, 5. Ala, 6. Phe, 7. Glu, 8. Trp, 9. Thr, 10. Val, 11. Lys, 12. Asp, 13. Tyr, 14. Leu, 15. Gly, 16. Pro, 17. Met)



Fig. S5. Fluorescence spectra of Probe **1** (10.0 μ M) towards (a) Cys and (b) Hcy (3.5 equiv). with some relevant species (3.5 equiv.) in PBS buffer (10.0 mM, 1.0 mM CTAB, pH = 7.4). The spectra were recorded after reacting 4 min at 25 °C. $\lambda_{em} = 590$ nm, $\lambda_{ex} = 488$ nm. (1. Arg, 2. Ile, 3. Ser, 4. His, 5. Ala, 6. Phe, 7. Glu, 8. Trp, 9. Thr, 10. Val, 11. Lys, 12. Asp, 13. Tyr, 14. Leu, 15. Gly, 16. Pro, 17. Met)



Fig. S6. The ¹H NMR spectrum of reaction product between Probe 1 and Cys in DMSO-d₆.



Fig. S7. The HRMS spectrum of reaction product between Probe 1 and Cys.



Fig. S8. The cytotoxicity of Probe 1 at different concentrations (0 μ M, 2 μ M, 5 μ M, 10 μ M, 15 μ M and 20 μ M) in MCF - 7 cells for 24 hours.



Fig. S9. The ¹H NMR spectrum of Probe 1 in CDCl₃.



Fig. S10. The ¹³C NMR spectrum of Probe 1 in CDCl₃.



Fig. S11. The HRMS of Probe 1.



Fig. S12. The emission spectra of probe 1 in response to biothiols at different pH values.

| Probe | $\lambda_{ex}/\lambda_{em}$ | Stokes shift | Response time | Literature |
|--|-----------------------------|-----------------|------------------|--|
| $(\mathcal{A}_{\mathcal{A}})^{\mathcal{O}}$ | 646 nm/656 nm | 10 nm | 90 min | Dyes and Pigments, 2018, 152, 85-92 |
| $\begin{array}{c} & & & \\ & & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & &$ | 365 nm/558 nm | 193 nm | 40 min | Dyes Pigments, 2014, 108 24-31 |
| $(1) \qquad (1) $ | 490 nm/580 nm | 90 nm | 60 min | Biomaterials, 2014, 35 6078- 6085 |
| | 510 nm/615 nm | 105 nm | 30 min | Sensor. Actuat. B-Chem., 2015, 211, 275-282. |
| | 365 nm/440 nm | 75 nm | / | Sensor. Actuat. B-Chem., 2016, 233, 307- 313. |
| NO2 NFF OF OF OF NO2 | 527 nm/570 nm | 43 nm | 60 min | Org. Biomol. Chem., 2010, 8, 3627-3630. |
| $(\mathcal{A}_{N^+}^{O}, \mathcal{A}_{N^-}^{O}, \mathcal{A}_{N^-}^{O}) \xrightarrow{\mathcal{O}_{N^-}}_{\mathcal{O}_2N^-} \xrightarrow{\mathcal{O}_{N^-}}_{\mathcal{O}_2N^-} \xrightarrow{\mathcal{O}_{N^-}}_{\mathcal{N}_2N^-}$ | 490 nm/553 nm | 63 nm | / | Org. Biomol. Chem., 2009, 7 4017-4020 |
| | 498 nm/613 nm | 115 nm | 120 min | Chinese Chem. Lett., 2019, 30, 563-565. |
| | 392 nm/583 nm | 191 nm | 2 min | Analyst, 2019, 144, 439-447. |
| | 560 nm/625 nm | 65 nm | 15 min | Chem. Commun. 2018, 54, 4786-4789 |

 Table. S1 Fluorescent probes for biothiols.

| | 450nm/525nm | 75 nm | 20 min | Talanta, 2018, 179, 326-330. |
|---|--------------------------------|-----------------|---------|---|
| | 485nm/619nm | 134 nm | 100 min | Tetrahedron, 2016, 72, 6909- 6913 |
| | 320nm/482nm | 162 nm | 20 min | Tetrahedron, 2017, 73, 589- 593. |
| $ \begin{pmatrix} N \\ N$ | 309nm/510nm | 201 nm | 10 min | Tetrahedron Lett., 2017, 58, 2654-2657 |
| $(\begin{array}{c} & & & & \\ & & & & \\ & & & & \\ & & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & & \\ & & & \\ & & & & \\ & & & & \\ & & & & \\ & & & \\ & & & & \\ & & & & \\ & & & & \\ & & & & \\ & & &$ | 365 nm/437 nm 500 nm/674 nm | 72 nm 174 nm | 5 h | Biomaterials,20 17, 139, 139- 150 |
| $ \begin{array}{c} OHC \\ OS \\ O2N \\ O$ | 346 nm/500 nm | 155 nm | 30 min | Dyes Pigments, 2018, 156, 338- 347. |
| | 420 nm/550 nm | 130 nm | 25 min | Anal. Chem., 2017, 89 8097- 8103. |
| | 580 nm/660 nm | 80 nm | 10 min | J. Mater. Chem. B, 2017, 5, 3836-3841. |
| | 586 nm/635 nm | 49 nm | 30 min | Res. Chem. Intermediat., 2017, 43, 7387- 7398. |
| O_2N O_2N O_2N O_2 O_2N O_2 O_2N O_2 O_2N O_2 O_2N | 453 nm/493 nm | 40 nm | 30 min | Dyes Pigments, 2018, 149 475- 480 |
| $O_{C}N$ Q Q Q Q Q Q Q Q | 504 nm/517 nm | 13 nm | 10 min | Dyes Pigments, 2018, 152, 29- 35. |

| | | | Food Chem., |
|---------------|-------|--------|----------------|
| 337 nm/347 nm | 10 nm | 27 min | 2018, 262, 67- |
| | | | 71 |