

## Facile synthesis of silver-rich Au/Ag bimetallic nanoparticles with highly active SERS property

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## **1. Experimental Sections**

### **Materials and Reagents**

Rabbit IgG, monoclonal Anti-rabbit IgG (Fc specific) antibody (designated as Ab1), atto-610 labeled polyclonal Anti-rabbit IgG (designated as Ab2), lipoic acid N-hydroxysuccinimide ester (LPA-NHS), sodium thiophenolate (TP), Poly(diallyldimethylammonium chloride) solution (PDDA, 20 wt.% in H<sub>2</sub>O), tetrachloroauric(III) acid trihydrate (HAuCl<sub>4</sub>·3H<sub>2</sub>O) and silver nitrate were purchased from Sigma-Aldrich. All other reagents were of analytical grade and used without further purification. Ultrapure water was obtained from a Millipore purification system ( $\geq 18$  M $\Omega$ , Milli-Q, Millipore). Phosphate-buffered saline (PBS; 0.01 M, pH 7.4) was used to dissolve Rabbit IgG and anti-Rabbit IgG, the washing buffer was the addition of 0.05% Tween-20 and 0.1% NaN<sub>3</sub> to PBS. The blocking buffer was an ethanolamine-tris buffer, which was prepared by dissolving 1.2 g of ethanolamine and 0.121 g of tris base in 20 mL of water and further adjusted pH to 8.4 using 1 M HCl solution.

### **Preparation of silver nanoparticles**

Briefly, 36 mg of AgNO<sub>3</sub> was dissolved in 200 mL water, and the solution was heated to a slight boiling state ( $\sim 95^\circ\text{C}$ ) in a flask equipped with stirring and reflux condensation device. Then, 4 mL 2% trisodium citrate solution (80 mg) was added rapidly, and the whole system was kept for 1 h at 90°C under vigorous magnetic stirring to obtain a gray-green transparent solution. The concentration of the silver nanoparticles was estimated to be  $5.5 \times 10^{-11}$  M (given that the silver nanoparticles had a spherical-like shape with  $\sim 50$  nm diameter, estimated from the TEM image)

### **Preparation of Au/Ag island films**

Au/Ag island films were fabricated by electrodepositing silver islands on gold slides and then magnetron sputtering a gold layer with 10 nm thickness. Electrochemical deposition was performed in a thermostated cell at room temperature, using a conventional three-electrode system. The gold slide was the working electrode, a platinum and an Hg/Hg<sub>2</sub>SO<sub>4</sub> electrode were used as the auxiliary and reference electrodes, respectively. Silver was deposited from a cyanide free plating bath (0.1 M AgNO<sub>3</sub> + 0.1 M EDTA + 0.05 M NH<sub>4</sub>NO<sub>3</sub> and several milliliters of concentrated ammonia were added to ensure the pH value was 9–10). The silver islands were prepared using multi-current pulse plating with the first pulse to a current density of 30 mA cm<sup>-2</sup> for 100 ms followed by a train of pulses of 5 mA cm<sup>-2</sup> for 10 ms separated by a rest time of 1 s (zero current) [1].

### **Characterization and SERS measurement**

Extinction spectra of the as-synthesized bimetallic nanoparticles were detected at a Shimadzu UV-3600 UV-vis-NIR spectrometer. TEM images were taken with a FEI-Tecni G20 transmission electron microscope (FEI). HRTEM and high angle annular dark-field scanning TEM (HAADF-STEM) investigations were carried out using a FEI-Tecni F20 (FEI) with 200 kV accelerating voltage. X-ray diffraction (XRD) patterns were recorded at a Bruker AXSD8 ADVANCE X-ray diffractometer (Cu K $\alpha$  radiation,  $\lambda = 0.15406$  nm) at room temperature. SERS spectra were measured on a Jobin Yvon/HORIBA HR500 Raman spectrometer equipped with an integral Olympus BX 41 confocal microscope with a 50 $\times$  objective lens. The excitation sources included an air-cooled argon ion laser (532 nm, 20 mW) and a Diode Pumped Solid State Laser (785 nm, 100 mW), both were equipped with a holographic notch filter with a grating of 1200 lines/mm. Raman scattering was detected with 180 geometry using a multichannel air-cooled (-70 $^{\circ}$ C) charge-coupled device (CCD) camera. For the detection of protein in this work, the laser power at the sample was  $\sim$ 1.5 mW, and the accumulation time was 30 s. For the SERS detection of TP, the accumulation time was 10 s.

## 2. Figures and Descriptions

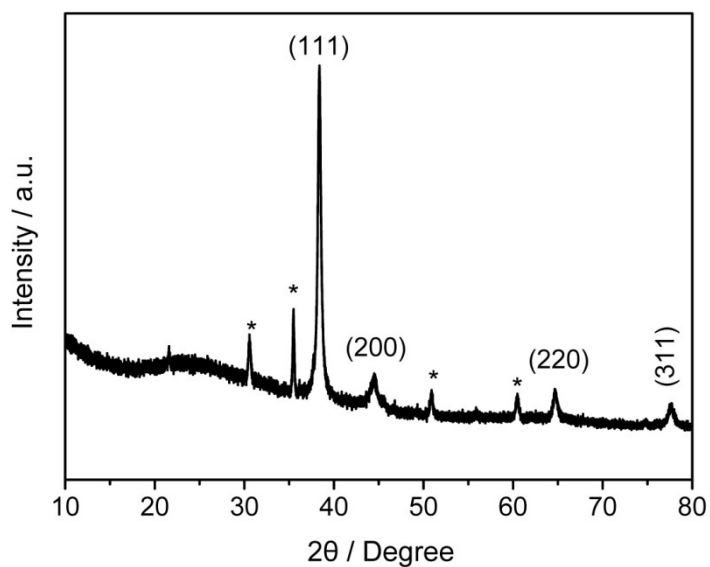


Figure S1 XRD pattern of the obtained BNPs, the peaks for ITO are marked with \*

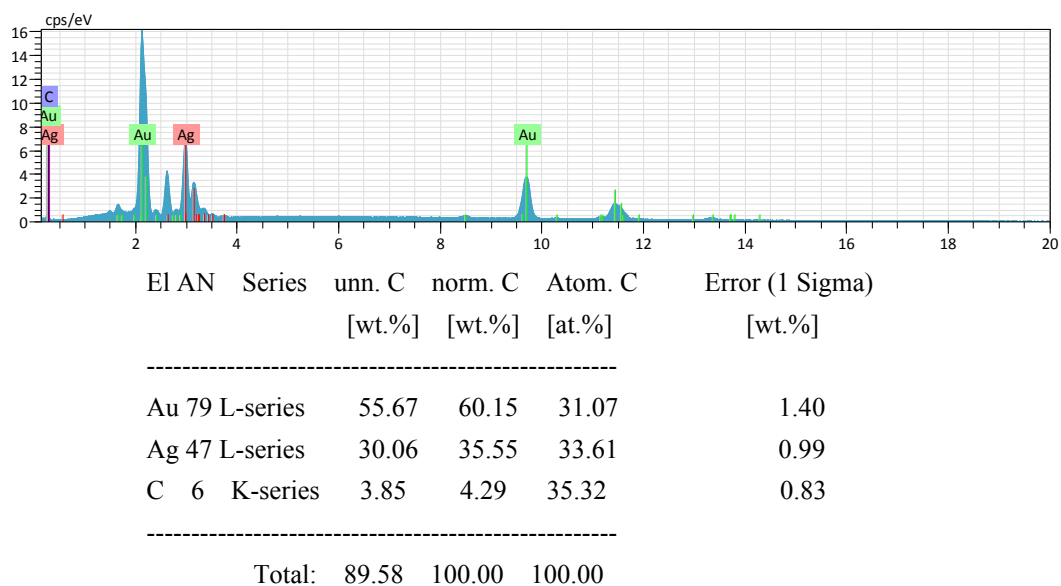
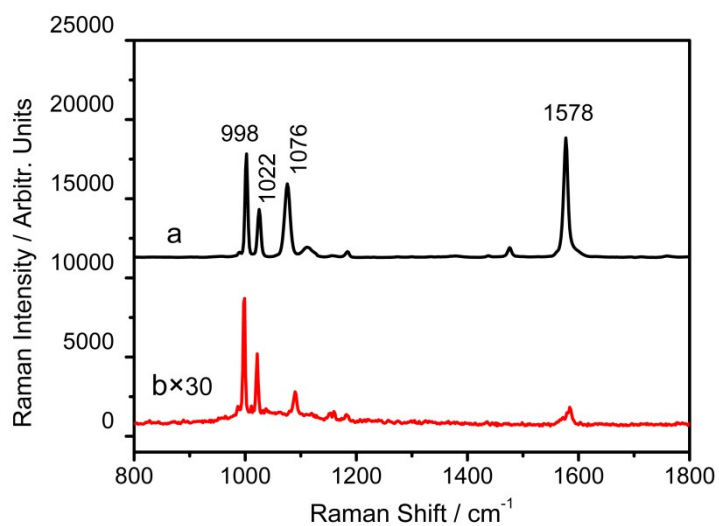


Figure S2 EDS analysis of the Au/Ag BNPs

Table S1 EDS analysis of the Au/Ag BNPs with different H<sub>AuCl<sub>4</sub></sub> added

Volume of H <sub>AuCl<sub>4</sub></sub> solution (mL)	Normalized atom percentage (at.%) In the presence of citrate		Normalized atom percentage (at.%) In the absence of citrate	
	Ag	Au	Ag	Au
	20	89.6	10.4	76.5
40	74.3	25.7	58.4	42.6
60	65.8	34.2	10.7	89.3
80	54.2	45.8	2.5	97.5
100	49.6	50.4	/	/
120	32.8	67.2	/	/
140	10.2	89.8	/	/
180	5.1	94.9	/	/
200	1.7	98.3	/	/

## EF calculation



**Fig. S3** (a) SERS spectrum of TP obtained on the BNPs thin film, (b) normal Raman spectrum of solid TP

### Calculation of ASEF values for the BNPs thin film

Enhancement factor (EF) of the BNPs thin film can be calculated using the following equation:[2]

$$EF = \frac{N_{bulk} I_{surf}}{N_{surf} I_{bulk}} \quad (1)$$

where  $I_{surf}$  and  $I_{bulk}$  denote the intensities for the strongest band of the TP adsorbed on the surface of BNPs and the solid TP, respectively.  $N_{surf}$  and  $N_{bulk}$  represent the numbers of the corresponding surface and solid molecules effectively excited by the laser beam, respectively.

### Calculation of $N_{bulk}$

The number of TP molecules excited in the bulk solid,  $N_{bulk}$ , can be calculated as following equation (2):

$$N_{bulk} = \frac{\pi \left( \frac{d_{spot}}{2} \right)^2 D \rho_{TP} N_A}{M_{r,TP}} \quad (2)$$

where  $d_{spot}$  is the diameter of circular laser spot,  $D$  is the depth of the incident laser beneath the surface of TP solid,  $\rho_{TP}$  and  $M_{r,TP}$  are the density and molecular weight of TP, respectively,  $N_A$  represents the Avogadro constant. In this study, the laser spot was a circle with diameter of  $\sim 4 \mu\text{m}$ , and the depth the laser could reach was about  $19 \mu\text{m}$ , the density ( $1.079 \text{ g/cm}^3$ ) and molecular weight ( $132.16 \text{ g/mol}$ ) of solid TP. The calculated value of  $N_{bulk}$  equals to  $1.9 \times 10^{12}$ .

### Calculation of $N_{surf}$

For the TP molecules adsorbed on the surface of BSSC thin films, assuming that the TP molecules are fully adsorbed and adopt a standing-up orientation on Ag surface, then the area occupied by one TP molecule is considered to be equal to the cross-sectional area of the molecule. The numbers of the TP molecules effectively excited by the laser beam on the surface of the BSSC thin films,  $N_{surf}$ , could be calculated as following equation (3):

$$N_{surf} = \frac{\pi \left( \frac{d_{spot}}{2} \right)^2}{A_{cs,TP}} \quad (3)$$

where  $d_{spot}$  has the same definition as equation (2),  $A_{cs,TP}$  is the cross-sectional area of the TP molecule diameter of circular laser spot, which is  $\sim 0.20 \text{ nm}^2$ . The calculated

value of  $N_{bulk}$  equals to  $1.71 \times 10^8$ .

### EF calculation

A baseline correction was conducted for EF value calculation at each spectrum, and Raman intensity at  $1578 \text{ cm}^{-1}$  was used to calculate the EF. The  $I_{surf}$  and  $I_{bulk}$  at  $1573 \text{ cm}^{-1}$  were 7572 and 42. Thus, the EF was calculated to be  $2.3 \times 10^6$ .

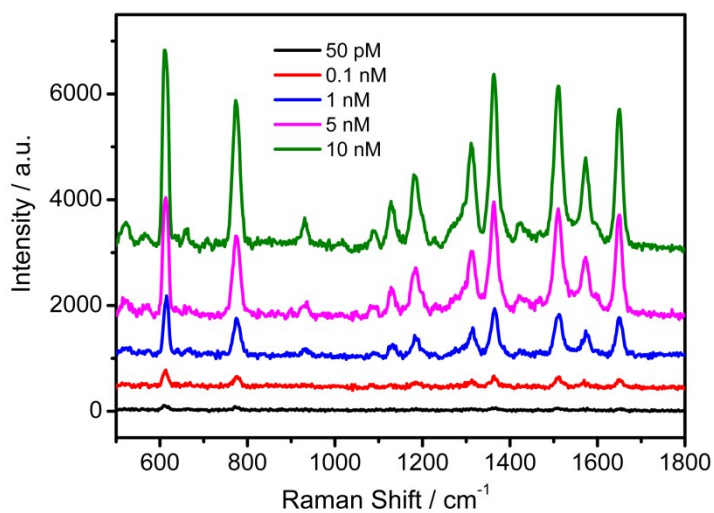


Figure S4 Raman Spectra of increasing concentrations of R6G on the BNPs (100 mL  $\text{HAuCl}_4$  added) at 785 nm laser excitation

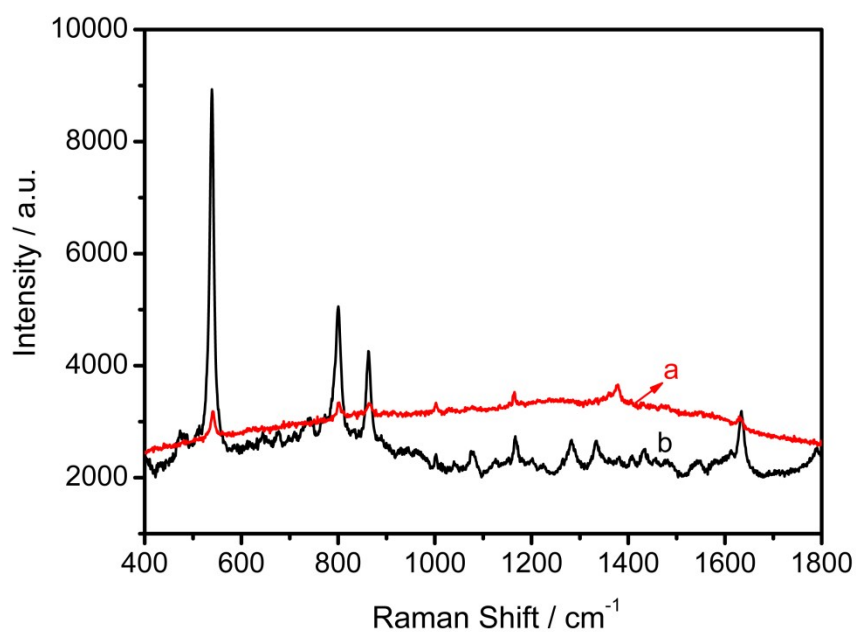


Figure S5 SERS spectra of Atto-610 labeled immunocomplex on Au thin film substrate (a) with and (b) without gold enhancement. The concentration of rabbit IgG is  $10 \text{ ng}\cdot\text{L}^{-1}$

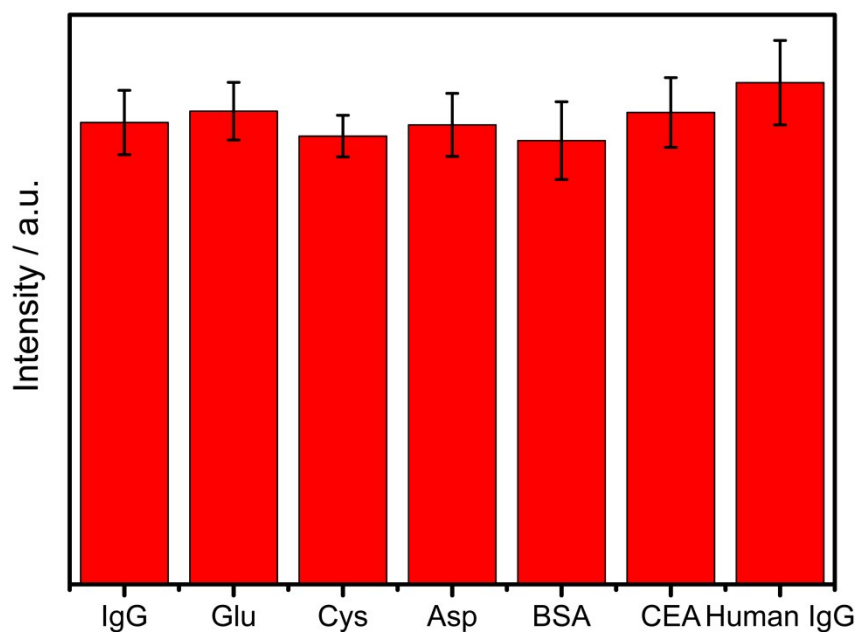


Figure S6 Specificity of the SERS-based immunosensor examined by addition of several interferents to  $10 \text{ ng}\cdot\text{L}^{-1}$  rabbit IgG. Each error bar represents the standard



deviation with  $n=5$ .

## References

- [1] X. Gu, Y. Yan, G. Jiang, J. Adkins, J. Shi, G. Jiang, S. Tian, Using a silver-enhanced microarray sandwich structure to improve SERS sensitivity for protein detection, *Anal. Bioanal. Chem.*, 406 (2014) 1885-1894.
- [2] X.-M. Lin, Y. Cui, Y.-H. Xu, B. Ren, Z.-Q. Tian, Surface-enhanced Raman spectroscopy: substrate-related issues, *Anal. Bioanal. Chem.*, 394 (2009) 1729-1745.