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Supplementary Information

Conjugated Polymer Dots for Biocompatible siRNA Delivery

Fei Wang,^{‡ac} Haobin Chen,^{‡a} Zhihe Liu,^a Feixue Mi,^a Xiaofeng Fang,^a Jie Liu,^a Mingxue Wang,^a

Pik Kwan Loc and Qiong Li*b

^a Department of Biomedical Engineering, Southern University of Science and Technology, Shenzhen, Guangdong 518055, China

^b Collaborative Innovation Center of Tumor Marker Detection Technology, Equipment and Diagnosis-Therapy Integration in Universities of Shandong, Shandong Provincial Key Laboratory of Detection Technology for Tumor Markers, College of Chemistry and Chemical Engineering, Linyi University, Linyi 276005, P. R. China.

^c Department of Chemistry, City University of Hong Kong, Tat Chee Avenue, Kowloon Tong, Hong Kong SAR, China

*Corresponding Author: Qiong Li

E-mail: qiongli7@163.com

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 Table S1. Detailed siRNA sequences used in this work

siRNA	Strand	Sequence (5'-end to 3'-end)
Control, siCtrl	Sense	AUGUAUUGGCCUGUAUUAGdTdT
	Anti-sense	CUAAUACAGGCCAAUACAUdTdT
Cy3-siRNA	Sense	Cy3-labeled UUCUCCGAACGUGUCACGUdTdT
	Anti-sense	ACGUGACACGUUCGGAGAAdTdT
siEG5	Sense	UCGAGAAUCUAAACUAACUdTdT
	Anti-sense	AGUUAGUUUAGAUUCUCGAdTdT



Fig. S1 Size distribution of G-PFO/siRNA. Hydrodynamic diameter of G-PFO/siRNA complexes measured by dynamic light scattering.



Fig. S2 Optical properties of and PFO and G-PFO Pdots. (a) Photographs of the PFO and G-PFO solutions in water under daylight (upper) or 365 nm irradiation (lower). (b) Absorption spectra and emission spectra of PFO dots and G-PFO Pdots.



Fig. S3 Stability of G-PFO Pdots in biological conditions. Hydrodynamic diameter of G-PFO Pdots after incubation in (a) PBS buffer or (b) DMEM medium for 24 hours measured by dynamic light scattering.



Fig. S4 siRNA release profile of G-PFO/Cy3-siRNA complexes in biological condition.

G-PFO/Cy3-siRNA complexes was prepared according to the method in the experimental section. Subsequently, 5 mL G-PFO/Cy3-siRNA complexes in PBS were transferred into a dialysis bag which was submerged in 20 mL of PBS solution later at 37 °C. At predetermined intervals (1, 3, 6, 12 and 24 h), 2 mL solution was taken out to determine the release siRNA amount by a fluorescence spectrometer.



Fig. S5 Cytotoxicity of G-PFO Pdots and Lipofectamine 2000. Relative viability of MCF-7 cells incubated with (a) G-PFO Pdots and (b) transfection agent Lipofectamine 2000 for 24 hours.



Fig. S6 Endosomal escape of G-PFO Pdots in MCF-7 cells (Blue: PFO Pdots; Red: LysoTracker Red).

MCF-7 were incubated with G-PFO Pdots for different time intervals (1, 3 and 6 h) and followed by LysoTracker Red staining. The time-different colocalization of G-PFO Pdots and endosome/lysosome was observed under a confocal laser scanning microscope.



Fig. S7 Bright field and merged images of MCF-7 cells corresponding to Fig. 6.



Fig. S8 Relative EG5 mRNA level of MCF-7 cells incubated with different treatments.

Quantitative Real-Time PCR (qRT-PCR) was conducted to evaluate the EG5 mRNA level. After different treatments, total RNA was isolated from MCF-7 cells using the RNeasy Mini Kit and 2 µg of total RNA was then used for cDNA synthesis. qRT-PCR was carried out by an initial denaturation at 94 °C for 30 s followed by 40 cycles, each consisting of 5 s at 94 °C, 30 s at 60 °C. Sequences for EG5 primers were 5'- GAACAATCATTAGCAGCAGAA-3' (Forward) and 5'- TCAGTATAGACACCACAGTTG-3' (Reverse). EG5 gene expression was normalized to Glyceraldehyde-3-phosphate dehydrogenase (GAPDH).