

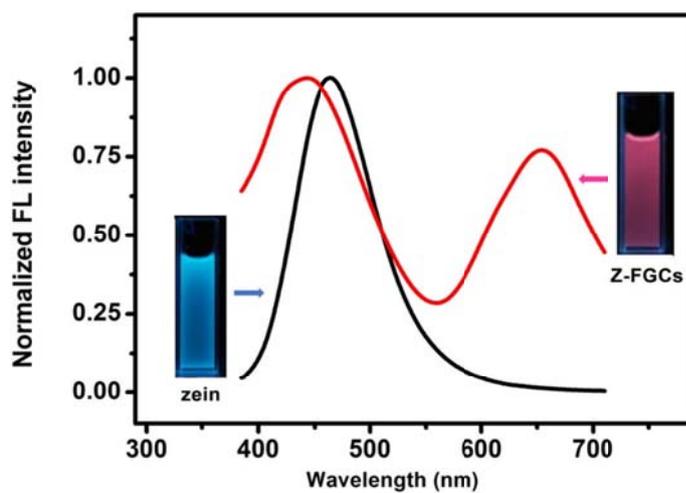
## Electronic Supporting Information

# Dual-emitting zein-protected gold nanoclusters for ratiometric fluorescent detection of $\text{Hg}^{2+}$ / $\text{Ag}^+$ ions in both aqueous solution and self-assembled protein film

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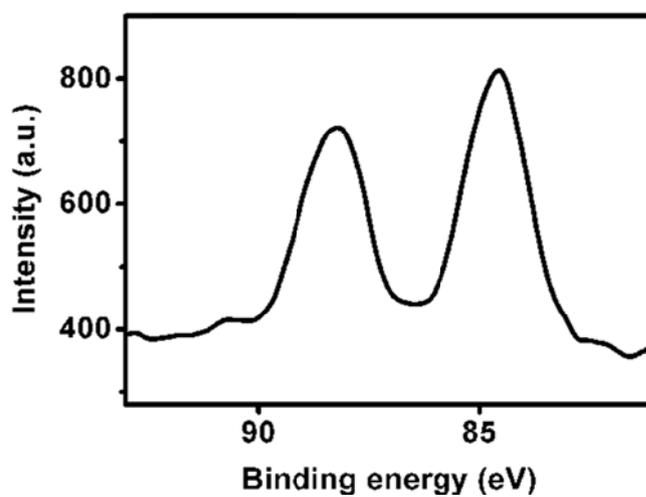
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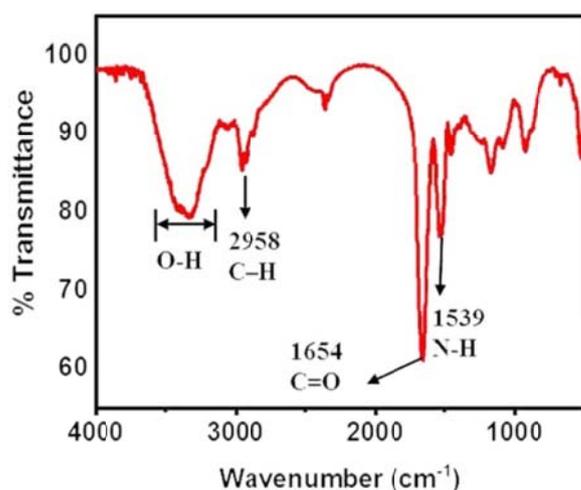


**Fig. S1.** Fluorescence emission spectra of zein (black) and Z-FGCs (red) under 365 nm excitation.

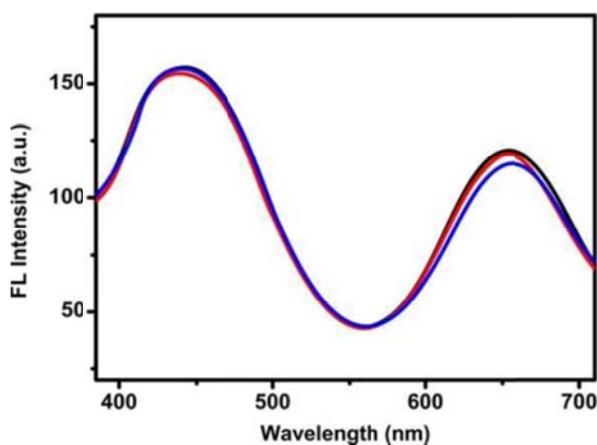
Inset: Photographs of zein and Z-FGCs under 365 nm UV lamp.



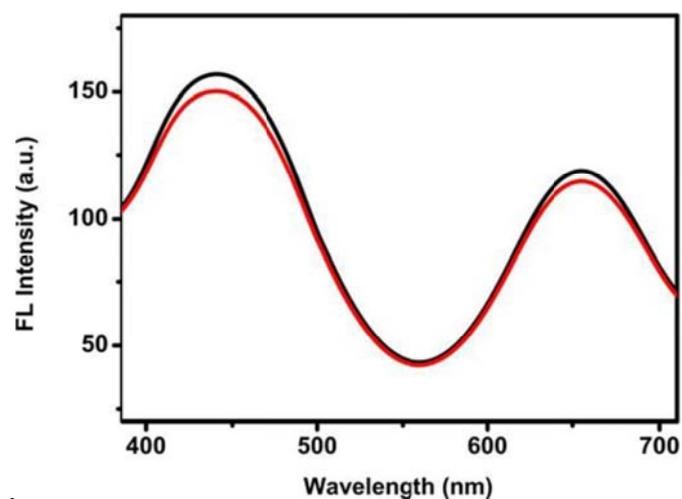
**Fig. S2.** Au 4f XPS spectrum of the Z-FGCs.



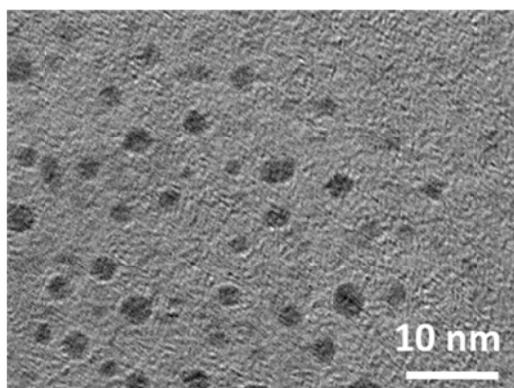
**Fig. S3.** FTIR spectrum of the Z-FGCs. The FTIR study of Z-FGCs exhibited the typical stretching vibration bands of C-H, C=O, N-H and O-H in protein, indicating the encapsulation of FGCs by zein. The bands at 1539 and 1654 cm<sup>-1</sup> can be attributed to the N-H and C=O stretching vibration, respectively. The band at 2958 cm<sup>-1</sup> can be assigned to C-H vibration. In addition, it appeared a broad band at 3100–3700 cm<sup>-1</sup>, which should be corresponded to the O-H stretches.



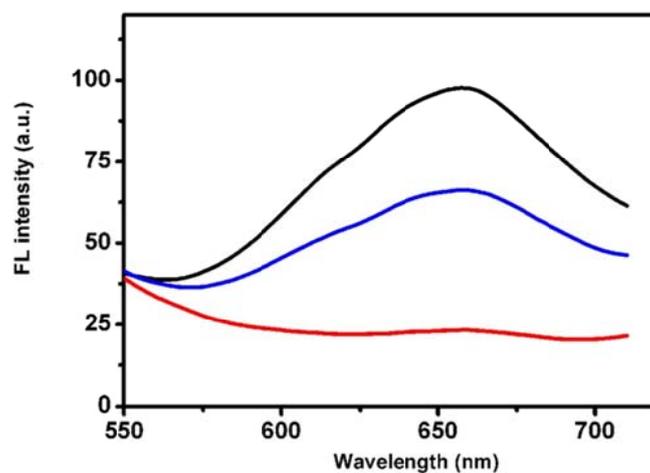
**Fig. S4.** Fluorescence emission spectra of the redispersed solution of Z-FGCs precipitate (black) and Z-FGCs powder acquired through freeze-drying for 12 h (red) or drying at 70 °C for 5 h (blue). No significant difference was observed in the fluorescence of the redispersed solution of the Z-FGCs precipitate/powder obtained through different ways.



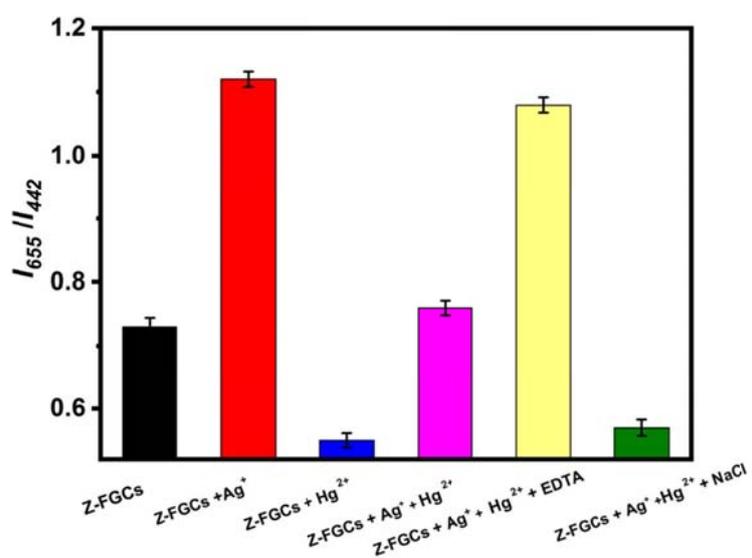
**Fig. S5.** Fluorescence emission spectra of the redispersed solution of Z-FGCs powder before (black) and after (red) the storage at 4 °C for 6 months. No significant loss in fluorescence intensity was observed, indicating that the Z-FGCs powder can be stored at 4 °C for at least 6 months.



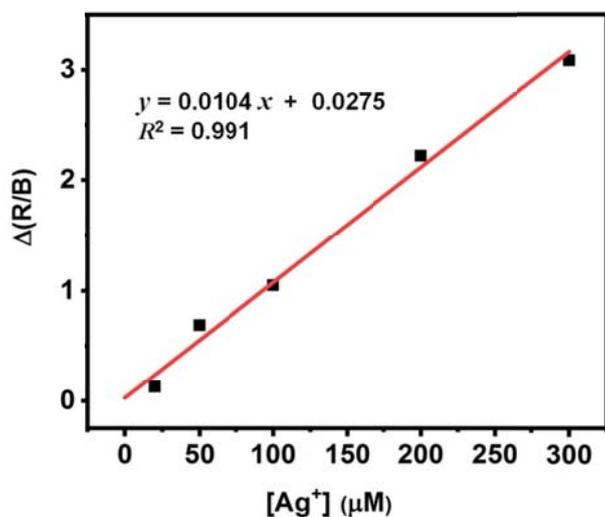
**Fig. S6.** TEM image of Z-FGCs after the addition of 50  $\mu\text{M}$   $\text{Ag}^+$ .



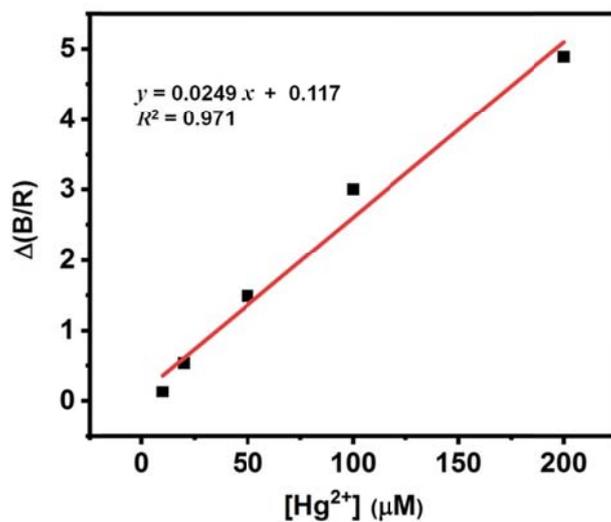
**Fig. S7.** Fluorescence emission spectra of Z-FGCs (black), Z-FGCs+ 50  $\mu\text{M}$   $\text{Hg}^{2+}$  ions (red) and Z-FGCs + 50  $\mu\text{M}$   $\text{Hg}^{2+}$  ions + 1.0 mM  $\text{NaBH}_4$  (blue).



**Fig. S8.** The change in  $I_{655}/I_{442}$  of Z-FGCs before (black) and after the addition of  $\text{Ag}^+$  (15  $\mu\text{M}$ , red),  $\text{Hg}^{2+}$  (15  $\mu\text{M}$ , blue),  $\text{Ag}^+$  (15  $\mu\text{M}$ ) +  $\text{Hg}^{2+}$  (15  $\mu\text{M}$ , magenta),  $\text{Ag}^+$  (15  $\mu\text{M}$ ) +  $\text{Hg}^{2+}$  (15  $\mu\text{M}$ ) + EDTA (1 mM, yellow) and  $\text{Ag}^+$  (15  $\mu\text{M}$ ) +  $\text{Hg}^{2+}$  (15  $\mu\text{M}$ ) + NaCl (10 mM, green).



**Fig. S9.** Change in the  $\Delta(R/B)$  of Z-FGCs-based film sensor for detection of  $Ag^+$  ions. Photographs of Z-FGCs film before and after the addition of different concentration of  $Ag^+$  were acquired by using Nikon D80 digital camera. The photographs were then processed by using a free RGB analysis software (Color cop) for recording the intensity ratio of red to blue (R/B) channel (in RGB chroma). The  $\Delta(R/B)$  was calculated by using intensity ratio of (R/B) of Z-FGCs film after the addition of different concentration of  $Ag^+$  minus that of (R/B) of Z-FGCs film alone. The detection limit was estimated to be  $\sim 6 \mu M$  by the IUPAC method.



**Fig. S10.** Change in the  $\Delta(\text{B/R})$  of Z-FGCs-based film sensor for detection of  $\text{Ag}^+$  ions. The  $\Delta(\text{B/R})$  was calculated by using intensity ratio of (B/R) of Z-FGCs film after the addition of different concentration of  $\text{Hg}^{2+}$  minus that of (B/R) of Z-FGCs film alone. The detection limit was estimated to be  $\sim 3 \mu\text{M}$  by the IUPAC method.

**Table S1.** List of FGCs-based method for detection of  $\text{Ag}^+$  and the relevant parameters of probe synthesis and purification

Probe	method	Time for probe synthesis	Purification	Linear range	LOD	refs
glutathione - FGCs	turn-on	24 h	dialysis for 24 h	0.5 nM–20 $\mu\text{M}$	0.2 nM	13
BSA- FGCs	turn-on	12 h	dialysis	0–40.0 $\mu\text{M}$	-	14
cytidine- FGCs	turn-on	1 h	-	0.01 – 6 mM	10 nM	40
11-mercaptoundecanoic acid-FGCs	turn-off	2 h	adding NaCl	25 nM – 3 mM	9 nM	41
BSA- FGCs	turn-on	6 h	dialysis for 24 h	0–20.0 mM	0.46 mM	42
zein- FGCs	ratiometric	10 min	mixing with acidic buffer for several minutes	1-15 $\mu\text{M}$	200 nM	this work

**Table S2.** List of FGCs-based method for detection of  $\text{Hg}^{2+}$  and the relevant parameters of probe synthesis and purification

Probe	method	Time for probe synthesis	Purification	Linear range	LOD	refs
aprotinin- FGCs	turn-off	12 h	dialysis for 24 h	0 - 330 $\mu\text{M}$	19.43 $\mu\text{M}$	7
esterase- FGCs	turn-off	18 h	dialysis for 24 h	1–30 nM	0.88 nM	8
BSA- FGCs	turn-off	12 h	dialysis for 24 h	1–20 nM	0.5 nM	12
keratin- FGCs	turn-off	12 h	size exclusion column	2.44–2500 nM	2.31 nM	16
egg-white-FNCs	turn-off	5 min	-	5–100.0 nM	0.89 nM	17
$\text{Tb}^{3+}$ /BSA-FGCs conjugates	ratiometric	12 h	dialysis for 10 h	0.005 - 7 $\mu\text{M}$	1 nM	18
N-acetyl-l-cysteine - FGCs	turn-off	24 h	dialysis for 72 h	2.0–3200 nM	0.2 nM	43
zein- FGCs	ratiometric	10 min	mixing with acidic buffer for several minutes	0.1-15 $\mu\text{M}$	25 nM	this work

**Table S3.** Detection of Ag<sup>+</sup> by Z-FGCs probe in tap water

Samples	Ag <sup>+</sup> added (μm)	Ag <sup>+</sup> found (μm) <sup>a</sup>	Recover (%)
Tap water	-	N.D. <sup>b</sup>	-
	2	2.21±0.06	107.5
	4	4.25±0.11	106.2
	6	6.28±0.14	104.7
	8	8.60±0.20	106.3

<sup>a</sup> Average of six replicate measurements ± standard deviation.

<sup>b</sup> N.D. : not detected.

**Table S4.** Detection of Hg<sup>2+</sup> by Z-FGCs probe in tap water

Samples	Hg <sup>2+</sup> added (μm)	Hg <sup>2+</sup> found (μm) <sup>a</sup>	Recover (%)
Tap water	-	N.D. <sup>b</sup>	-
	4	4.30±0.12	106.7
	6	6.35±0.14	105.8
	8	8.58±0.17	107.2
	10	10.60±0.22	106.0

<sup>a</sup> Average of six replicate measurements ± standard deviation.

<sup>b</sup> N.D. :not detected.